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**Influences of Kupffer cells and platelets on ischemia-reperfusion injury in mild steatotic liver**

Ogawa K *et al*. Ischemia-reperfusion injury in mild steatotic liver

Koichi Ogawa, Tadashi Kondo, Takafumi Tamura, Hideki Matsumura, Kiyoshi Fukunaga, Tatsuya Oda, Nobuhiro Ohkohchi

**Koichi Ogawa, Tadashi Kondo, Takafumi Tamura, Hideki Matsumura, Kiyoshi Fukunaga, Tatsuya Oda, Nobuhiro Ohkohchi**, Department of Surgery, Graduate School of Comprehensive Human Sciences, University of Tsukuba, Tsukuba 305-8575, Japan

**Author contributions:** Ogawa K and Kondo T contributed equally to this work; Ogawa K, Kondo T, Tamura T and Matsumura H designed the study; Ogawa K and Tamura T performed the experiments; Fukunaga K, Oda T and Ohkohchi N contributed the analysis of the data; Ogawa K, Kondo T, Fukunaga K and Ohkohchi N wrote the manuscript.

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**Correspondence to: Nobuhiro Ohkohchi, MD, PhD, Professor,** Department of Surgery, Doctoral Program in Clinical Science, Graduate School of Comprehensive Human Sciences, University of Tsukuba, 1-1-1 Tennodai, Tsukuba 305-8575, Japan. [nokochi3@md.tsukuba.ac.jp](mailto:nokochi3@md.tsukuba.ac.jp)

**Telephone:** +81-29-8533221  **Fax:** +81-29-8533222

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**Abstract**

**AIM:** To investigate the effect of mild steatotic liver on ischemia-reperfusion injury by focusing on Kupffer cells (KCs) and platelets.

**METHODS:** Previously, we reported that the interaction between KCs and platelets played a key role in hepatic ischemia-reperfusion (IR) injury as revealed by intravital microscopy (IVM) and electron microscopy. Wistar rats were divided into a normal liver group (N group) and a mild steatotic liver group (S group) induced by feeding a choline deficient diet (CDD) for 2 wk. Both groups were subjected to 20 min of warm ischemia followed by 120 min of reperfusion. The number of labeled KCs and platelets in sinusoids and the blood perfusion in sinusoids were observed by IVM, which was performed at 30, 60 and 120 min after reperfusion. To evaluate serum alanine aminotransferase as a marker of liver deterioration, blood samples were taken at the same time as IVM.

**RESULTS:** In the S group, the number of platelets adhering to KCs decreased significantly compared with the N group (120 after reperfusion; 2.94 ± 1.1 cells/acinus *vs* 4.82 ± 1.16 cells/acinus, *P* < 0.01). The number of KCs in sinusoids was significantly less in the S group than in the N group throughout the observation periods (before ischemia; 19.6 ± 3.28 cells/acinus *vs* 28.2 ± 4.1 cells/acinus, *P* < 0.01 and 120 min after reperfusion; 29.0 ± 4.27 cells/acinus *vs* 40.2 ± 3.33, *P* < 0.01). The blood perfusion of sinusoids 120 min after reperfusion was maintained in the S group more than in the N group. Furthermore, elevation of serum ALT was lower in the S group than in the N group 120 min after reperfusion (99.7 ± 19.8 IU/L *vs* 166.3 ± 61.1 IU/L, *P =* 0.041), and histological impairment of hepatocyte structure was prevented in the S group.

**CONCLUSION:** IR injury in mild steatotic liver was attenuated compared to normal liver due to the decreased number of KCs and the reduction of the KC-platelet interaction.

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**Key words:** Steatotic liver; Mild steatotic liver; Kupffer cell; Platelet; Ischemia-reperfusion; Intravital microscopy

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**INTRODUCTION**

It is widely accepted that the steatotic liver is more susceptible to hepatic ischemia-reperfusion (IR) injury. Increased microcirculatory deterioration is suggested as a reason for this increased liver damage in the steatotic liver[1-3]. Morphologically, the steatotic liver is characterized by a demonstrable deposition of large, macronodular fatty droplets in liver parenchymal cells[4]. In addition, narrow and distorted lumens of sinusoids, resulting from the swelling of hepatic parenchymal cells due to accumulated lipid, cause a decrease in sinusoidal perfusion[2]. These changes to microcirculation are exacerbated by leukocytes, either mechanically trapped in the narrowed sinusoids or adhering to activated Kupffer cells (KCs) with the release of cytokines, in addition to free radicals[2, 5].

Thrty percent of the population in Japan and Western countries suffers from steatotic liver, and the percentage is still increasing[3,6]. Most of these patients have been diagnosed by abdominal ultrasonography screening, even though elevation of serum liver enzyme levels was detected in a lesser percentage[6]. Steatosis of the liver is classified clinically into three grades according to the proportion of hepatocytes with fatty droplets: mild (< 30%), moderate (30%–60%), and massive (> 60%)[7]. Fishbein *et al* [8] reported that liver enzyme levels were not elevated in the steatotic liver in which the proportion of hepatocytes with fat deposition was less than 18%. Steatotic liver even in a mild degree is a risk factor for complication after liver resection[9]. Most of the reports describing IR injury of the steatotic liver were investigations of moderate and severe steatosis. It is unclear whether the intensity of the hepatic IR injury depends on the degree of fatty change.

Previously, we reported that liver ischemia induced the adhesion of platelets to KCs in the early period after reperfusion, and that interaction between KCs and platelets played a key role in reperfusion injury of the liver[10,11]. In this study, we have focused on the interaction between KCs and platelets in the mild steatotic liver with intravital microscopy (IVM). We hypothesized that tolerance to hepatic IR injury differs according to the degree of steatosis. The aim of this study is to clarify the hepatic dysfunction after IR in the mild steatotic liver compared with the normal liver.

**MATERIALS AND METHODS**

***Animals***

Male Wistar rats were obtained from CLEA Japan, Inc. (Tokyo, Japan). We prepared two model types, normal liver and mild steatotic liver. In the normal liver model, rats weighting 250 g to 300 g were used. In the steatotic model, the rats’ weights were adjusted to the same weight range after they were fed a choline-deficient diet (CDD) (Oriental Bio Service Kanto Inc, Ibaraki, Japan) for 2 wk. Animal experiments were carried out in a humane manner after receiving approval from the Institutional University Experiment Committee of the University of Tsukuba, and in accordance with the Regulation for Animal Experiments in our university and the Fundamental Guideline for Proper Conduct of Animal Experiment and Related Activities in Academic Research Institutions under the jurisdiction of the Japanese Ministry of Education, Culture, Sports, Science, and Technology.

***Experimental design***

Animals were divided into two groups as follows: (1) the normal liver group (N group; *n* = 6); and (2) the mild steatotic liver group (S group; *n* = 6). In both groups, total normothermic hepatic ischemia was induced for 20 min by clamping the portal triad. The hepatic microcirculation and dynamics of platelets and KCs were observed just before ischemia and at 30, 60, and 120 min after reperfusion (Figure 1).

***Surgical procedure***

Under anesthesia using isoflurane, the animals were tracheotomized. To reduce spontaneous breathing, animals were ventilated mechanically (MK-V100; Muromachi Kikai Co. Ltd, Tokyo, Japan). The animals were placed in a supine position on a heated pad to maintain the rectal temperature at 37°C. To monitor arterial blood pressure and allow continuous infusion of Ringer’s solution, polyethylene catheters (PE- 50, 0.58/0.96-mm internal/external diameter; Becton Dickinson, Sparks, MD) were inserted into the left carotid artery and left jugular vein, respectively. After performing laparotomy *via* a transverse incision, the ligaments around the liver were dissected to mobilize the left lobe. At the same time, the hepatoduodenal ligament was taped for clamping later. The left hepatic lobe was exteriorized on a plate specially designed to minimize movements caused by respiration and covered with cover glass. Surgical procedures were performed using sterile technique. After 60 min of normal saline continuous infusion, IVM was performed as a pre ischemia study. Then, hepatic ischemia was induced by clamping the portal triad (the hepatic artery, portal vein, and bile duct) with a microclip (B. Braun Aesculap Japan Co., Ltd, Tokyo, Japan) for 20 min. IVM was performed at 30, 60 and 120 min after reperfusion. Blood samples were taken for analysis of enzyme activities from a catheter placed in the left carotid artery at the same time as IVM. Alanine aminotransferase (ALT) was evaluated as one liver enzyme. At 120 min of reperfusion, total body blood was taken for euthanasia. At the end of the experiments, liver tissue was taken for histological examination. Finally, the experimental animals underwent euthanasia by the total blood collection method.

***Platelet preparation***

Platelets were isolated from the whole blood of syngeneic rats and labeled with rhodamine-6G (50 μl/mL whole blood: R-4127; Sigma, St. Louis, MO, United States), as described by Massberg *et al*[12]. Briefly, the collected blood was diluted with buffer after the addition of prostaglandin E1 and rhodamine 6G. After two-cycle centrifugation, fluorescent platelets were resuspended in phosphate buffered saline (PBS). In this study, a total of 1×108 fluorescence-labeled platelets, approximately 1% of all circulating platelets in the recipient rat, were injected through the left carotid artery at 5 min before IVM.

***Liposome entrapment method (Fluorescence labeling of KCs)***

Fluorescently labeled phosphatidylcholine (PC) was incorporated into liposomes, as described by Watanabe *et al*[13]. The fluorescent pigment used was 2-(12-(7-nitrobenz-2-oxa 1,3-diazol-4-yl) amino) dodecanoyl-1-hexadecanoly-*sn*-glycero-3-phosphocholine (NBD-C12-HPC; Molecular Probes; Eugene, United States). After intra-arterial injection, KCs in the rat livers were stained and were clearly delineated under the fluorescence image in the IVM. Phagocytic activity of KCs after the administration of liposomes was reported by measuring the amount of hepatic uptake of intravenously administered fluorescent microspheres; no detrimental influence of the liposomes on the phagocytic activity was observed. Additionally, no histopathological changes were found in the livers from liposome-treated rats[13]. Sixty minutes before hepatic ischemia, liposome-encapsulated fluorescent liposomes (4 mL/kg) were administered from the carotid artery catheter.

***IVM***

IVM was performed using a modified microscope (BX30 FLA-SP; Olympus Co., Tokyo, Japan) with a 100 W mercury lamp attached to a filter block. The hepatic microcirculation was recorded by means of a CCD camera (C5810; Hamamatsu Photonics, Hamamatsu, Japan) and a digital video recorder (GV-HD700/1; Sony, Tokyo, Japan) for offline analysis. Using objective lenses (10 × 0.3 to 20 × 0.7; Olympus Co., Tokyo, Japan), a final magnification from x325 to x650 was achieved on the video screen. To assess sinusoidal perfusion, sodium fluorescein (2 × 10-3 M/kg, F-6377; Sigma, St. Louis, MO) was injected *via* the jugular catheter. Rhodamine-6G labeled platelets were infused intra-arterially just before ischemia and at 30, 60, and 120 min after reperfusion, and 10 randomly chosen acini were visualized. Quantitative assessment of the microcirculatory parameters was performed offline using WinROOF imaging software (version 5.0; Mitani Shoji, Tokyo, Japan).

***Microcirculatory analysis***

The following parameters were analyzed: (1) the number of adherent platelets, i.e., platelets firmly attached to the sinusoidal endothelium for longer than 20 s [The number of adherent platelets in the scanned acini was counted and results were expressed as the number of adherent platelets per field (1 field = approximately 0.2 mm2)]; (2) the number of adherent platelets adhering to KCs; (3) the number of KCs; and (4) sinusoidal perfusion failure rate (%) as an index of microcirculatory disturbance, calculated as the ratio of non-perfused sinusoids among the sinusoids observed in one acinus after 120 min of reperfusion.

***Immunohistochemical study of KCs***

We immunohistochemically assessed the number of KCs in the acini. To compare the differences between the normal liver group and the mild steatotic liver group before ischemia and after reperfusion, liver tissues were obtained from each group both before ischemia and at the end of surgical procedure, and from another animal before ischemia. The tissues were fixed in 10% formalin and embedded in paraffin and cut into 4 μm-thick sections. It was immersed in 0.03% hydrogen peroxidase to block endogenous peroxidase activity, and then blocked with 2% bovine serum albumin to reduce background staining. To specifically recognize KCs, mouse anti-rat ED2 antibody (Santa Cruz Biotechnology, Santa Cruz, CA, United States) was used as the primary antibody. The sections were incubated with primary diluted antibodies (1:50) at room temperature for 60 min. Primary antibody reactions were enhanced using horseradish peroxidase EnVision (Dako Japan, Tokyo, Japan). The immunoreaction was visualized with 0.05% 3,3-diaminobenzidine (DAB) solution. After washing in distilled water, specimens were counterstained with hematoxylin then mounted. The number of ED2-positive cells per acinus was counted in five randomly chosen acini.

***Biochemical assays***

As a marker of liver deterioration, serum ALT levels were measured using a Drychem 7000V autoanalyzer (Fuji Film, Tokyo, Japan). Serum was stored at -80°C until use for cytokine determination. Levels of interleukin (IL)-6 were measured using commercial enzyme-linked immunosorbent assay (ELISA) kits (R and D Systems, Minneapolis, MN, United States).

***Histological analysis***

Liver tissue was obtained before ischemia from the S group to assess the degree of steatosis and after 120 min of reperfusion from each group to assess the histological changes due to ischemia reperfusion. The samples were fixed with 10% formalin, and embedded in paraffin. Thin sections (4 μm) were prepared and stained with hematoxylin-eosin (HE). Tissue damage was evaluated in 5 randomly selected high-power fields (× 200).

***Statistical analysis***

All data are expressed as mean ± SD. The Mann-Whitney test and analysis of variance (ANOVA) were used, followed by Scheffe’s test. *P* values < 0.05 were considered statistically significant.

**RESULTS**

***Mild steatotic liver model***

Mild steatosis of the liver was induced by feeding CDD. The CDD-induced steatotic liver is an established experimental model, in which morphological and functional features are very similar to those of the clinical steatotic liver[14]. The rats fed on CDD for 2 wk developed mild steatotic livers. They were characterized by microvesicular lipid droplet filtration in 10% to 20% of hepatocytes (HE stain) (Figure 2). These findings were identified as a mild degree of steatosis of the liver. We preoperatively confirmed similar findings by liver biopsy in several animals.

***Number of adherent platelets in acini***

The number of adherent platelets in sinusoids increased along with the reperfusion time both in the N group and in the S group (Figure 3A). In the S group, the number of adherent platelets adhering to KCs was significantly less than in the N group at 30 min after reperfusion and concomitant with the reperfusion period (Figure 3B, C). In addition, there was no significant difference between the two groups in the number of blood platelets (data not shown).

***Number of KCs in acini***

The mild steatotic change in the liver significantly decreased the number of KCs in sinusoids compared to the normal liver at any point in time before and after ischemia reperfusion with IVM study (Figure 4A). In the IVM observation, the counted KCs represented only KCs labeled by liposome entrapment methods. In addition, the number of KCs was verified immunohistochemically. In immunohistochemical staining, the numbers of ED2-positive cells were lower in the S group than in the N group already at the time before ischemia and after 120 min of reperfusion (Figure 4B).

***Sinusoidal perfusion failure rate***

Vollmar *et al* [15] reported that the sinusoidal perfusion rate was one of the indexes of reperfusion injury. After 120 min of reperfusion, the rate of sinusoidal perfusion failure was significantly higher in the N group than in the S group (Figure 5). The perfusion of sinusoids in which KCs and platelets adhered had a failure rate of about 50% in both groups (data not shown). In contrast, the perfusion failure rate of sinusoids in which KCs adhering to platelets was not observed remained at approximately 30% in both groups (data not shown). This indicated that the interaction between KCs and platelets was associated with sinusoidal perfusion failure.

***Serum ALT and interleukin-6 levels***

Serum ALT level as a measure of hepatic parenchymal impairment was significantly lower in the S group compared with the N group after 120 min of reperfusion (Figure 6A). The concentration of serum interleukin-6 (IL-6) had a tendency to be lower in the S group than in the N group, but there was no significant difference (Figure 6B).

***Histological findings***

In the S group, histological damage to the liver, such as disturbance of the hepatocyte structure and sinusoidal narrowing, was slightly greater than in the N group. Necrotic changes were not observed in either group (Figure 7).

**DISCUSSION**

Steatosis of the liver is a common disorder with several different etiologies. This disorder is one of the most common obstacles in liver surgery, since steatotic livers are susceptible to some stressful loads, especially to IR injury[1,16]. With the increase in steatotic liver patients in the world, the number of mild steatotic liver case is also increasing[17]. A recent analysis of liver transplants, in which the graft liver suffered from cold ischemia, reported that the degree of liver graft steatosis is an important determinant of IR injury correlated with the rate of postoperative complications and mortality[18,19]. In warm ischemia, however, the influence of the degree of hepatic steatosis on liver dysfunction is unclear. In this study, we demonstrated that IR injury in mild setatotic liver was attenuated compared with that in normal liver, and that it resulted from the reduction of the interaction between KCs and platelets due to the decreased number of KCs.

It was reported that there are two distinct periods of liver injury after warm IR[20,21]. The early period of IR injury, which occurs within 120 min after reperfusion, is characterized by KC-induced [reactive oxygen species](javascript:void(0);) (ROS)[21,22]. KC production and the release of ROS result in acute hepatocellular injury. In addition, in response to the exposure to activated KCs, neutrophils accumulate in the post-ischemic liver. In the late period of IR injury, inflammatory responses from accumulating neutrophils induce hepatocyte injury, which appears more than 6 h after reperfusion[22,23]. Elevated liver enzymes and apoptosis of hepatocytes and sinusoidal endothelial cells can already be observed in the early period[10,24]. Reduction of the early period of IR injury, for instance KCs depletion, leads also to inhibition of injury in the late period[25]. Recently, some studies have focused on the function of platelets in hepatic IR injury[26,27]. Sindram *et al*[24] reported that platelets caused sinusoidal endothelial cell (SEC) apoptosis and significantly contributed to IR injury. Khandoga *et al*[28] have shown that warm hepatic IR induced rolling and adhesion of platelets to SECs as well as accumulation of platelets in sinusoids. We previously reported that platelet-SEC interactions occur earlier than leukocyte responses after reperfusion, and that adhesion of platelets requires the presence of activated KCs[11]. In addition, based on our study using the IVM system and electron microscopy, we also reported that platelet-KC interaction as well as platelet-SEC interaction contributes to early-period hepatic IR injury[10,24,28]. Most of the events that determine the extent of IR injury, such as KC activation, platelet adhesion to KCs or SECs, and neutrophil accumulation, have occurr in the early period of IR injury. Therefore, we focused on observation until 120 min after reperfusion. In this study, we demonstrated that the adherence of platelets to KCs decreased and reperfusion injury was reduced in the mild steatotic liver. The number of adherent platelets in sinusoids increased along with the reperfusion time both in the normal liver group and in the mild steatotic liver group. This suggests that the adherent ability of platelets was not reduced in the mild steatotic liver group. Therefore, we considered that the reduction in KC-platelet interaction was a result of the decreased number of KCs in the mild steatotic liver.

KCs are more likely to be activated in the steatotic liver[5,29]. In addition, hepatic IR activates KCs[30]. However, it is unknown whether KC activation after IR increases in steatotic liver more than normal liver. After IR, KCs secrete pro-inflammatory cytokines including IL-6[31,32]. Moreover, as described above, activated KCs cause adhesion of platelets to KCs, and lead to later leukocyte accumulation[11]. We consider that serum IL-6 levels reflect the degree of the interaction between KCs and platelets according to the activity of KCs. In the present study, elevation of serum IL-6 levels after IR was less in mild steatotic liver than normal liver. Our results indicated that in mild steatotic liver, IL-6 secretion was suppressed because of the decreased number of KCs, even if KCs were activated after IR.

In our present study, we demonstrated that there were fewer KCs in the sinusoids in our mild steatotic liver than in the normal liver. The results were confirmed by both the IVM study and immunohistochemical examination. Several studies reported on change in the number of KCs in steatotic liver models. Shono *et al*[33] reported that the number of KCs in a particular subgroup could change, for example, the proportion of CD68 positive KCs decreased in their steatotic model induced by a high-fat diet and a high-cholesterol diet (HCD). Vetelainen *et al*[34] investigated the differences between a methionine-choline deficient diet (MCD) model and a CDD model in their effect on the number of KCs, and reported that the number of KCs increased in the MCD model, but did not change in the CDD model. The degree of steatosis was moderate in their CDD model, and severe in their MCD model. These investigators indicated that the difference in the method of inducing steatosis of the liver resulted in change in the number of KCs in the sinusoids. In addition, Guo *et al*[35]reported that the number of KCs was reduced in their steatotic liver model induced by palmitoleate, a monounsaturated fatty acid. Thus, differences in nutrient factors may influence the number of KCs in the steatotic liver. We supposed that change in nutrient conditions induced by CDD might lead to decrease of KCs as well as mild steatotic liver. The relationship between KCs and steatotic liver has not been well established yet. Further research using various steatotic liver models and various degrees of steatosis will be necessary to elucidate the impact of steatotic liver on KCs.

Steatotic liver patients in the clinical setting of hepatic surgery have tended to increase[17]. IR injury is involved closely with complications after hepatic resection in steatotic liver[36]. It is known that steatotic liver is a risk factor for postoperative complications[9,36]. Some investigators reported that patients with steatotic liver who received major hepatectomy were more likely to suffer from infective, wound-related, hepatobiliary and gastrointestinal postoperative complications[37]. The decreased tolerance of steatotic liver to IR injury is a result of impaired microcirculation due to hepatocytes with fat deposition[38]. Between mild and severe steatotic liver, there are differences in microcirculation disturbances due to differences in the degree of fat deposition. Recent meta-analysis revealed a significant association between degree of steatosis and increased risk of postoperative complications and mortality[9]. Several investigators reported that postoperative complications, especially infectious complications, and mortality increased in patients with severe steatotic liver compared with those with mild setatotic liver[9,37]. It will be necessary to evaluate IR injury in the moderate to severe steatotic liver with a similar experimental model in the future.

On the other hand, there is a report that postoperative liver failure was slight in mild steatotic liver, and so mild steatotic liver can be an indication for hepatic surgery[39]. Moreover, mild to moderate seteatotic livers have been accepted as a marginal graft in transplantation[18,19]. Our study suggested that IR injury does not depend on the degree of fat deposition. In addition, our results provide some evidence that postoperative outcomes after liver resection or transplantation are not aggravated in mild steatotic liver.

In conclusion, in the mild steatotic liver induced by CDD, hepatic IR injury was attenuated compared with the normal liver. The small number of KCs in the sinusoids decreased the number of KCs adhering to platelets, and resulted in decreased interaction between KCs and platelets.

**COMMENTS**

***Background***

The steatotic liver is well known to be sensitive to ischemia-reperfusion (IR) injury. Increased microcirculatory deterioration is suggested as a reason for this increased liver damage in the steatotic liver. However, it is unclear whether IR injury increases in the mild steatotic liver. Previously, authors reported that the interaction between Kupffer cells (KCs) and platelets played a key role in hepatic IR injury. Authors investigated the effect of mild steatotic liver on IR injury with focusing on Kupffer cells and platelets.

***Research frontiers***

KCs activation plays a pivotal role in hepatic IR injury. Upon activation by hepatic ischemia, KCs release [reactive oxygen species](javascript:void(0);) and inflammatory mediators, such as cytokines and chemokines. In addition, KC-activation leads to platelet and neutrophil accumulation in hepatic sinusoids. On the other hand, KCs are more likely to be activated in the steatotic liver.

***Innovations and breakthroughs***

This is the first study focusing on KCs and platelets in IR injury in the mild steatotic liver. The authors demonstrated that hepatic IR injury in mild steatotic liver was attenuated compared with normal liver. The small number of KCs in the sinusoids decreased the number of KCs adhering to platelets, and resulted in decreased interaction between KCs and platelets.

***Applications***

The authors suggested that IR injury in steatotic liver does not depend on the degree of fat deposition. The results of this study indicate some evidences that the postoperative outcomes after liver resection or transplantation do not aggravate in mild steatotic liver.

***Terminology***

Intravital fluorescence microscopy (IVM) of the rat liver is a high-resolution real-time technique that allows authors visualize hepatic sinusoidal perfusion. The availability of an enormous number of different fluorescent markers for *ex vivo* and *in vivo* staining has extended the possibilities of IVM from purely morphological analysis to the study of complex physiological and pathological events.

***Peer review***

The manuscript demonstrated the decrease in IR injury in mild steatotic liver in Wistar rats by feeding in choline-deficient diet compared to normal rats. In addition, the results also showed that the attenuation of IR injury was associated with the decrease of KC number and platelet-KC adhesion in acini. The study is well designed and the results are interesting and conceivable.

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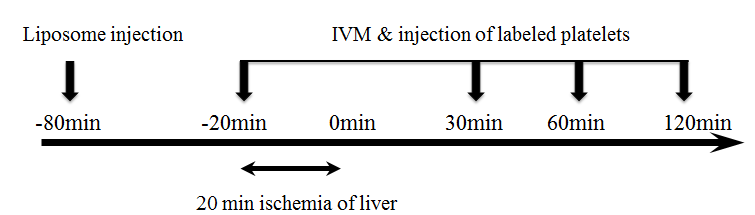
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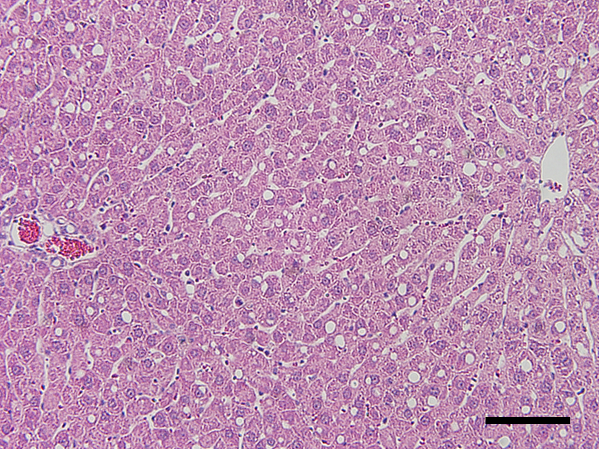
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**P-Reviewers** Acquaviva R, Han JY **S-Editor** Wen LL  **L-Editor**  **E-Editor**



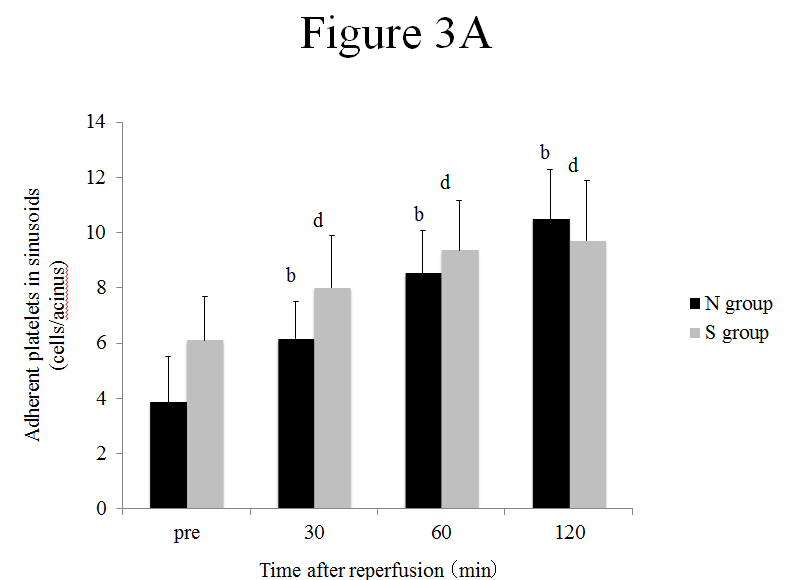
IVM and injection of labeled platelets

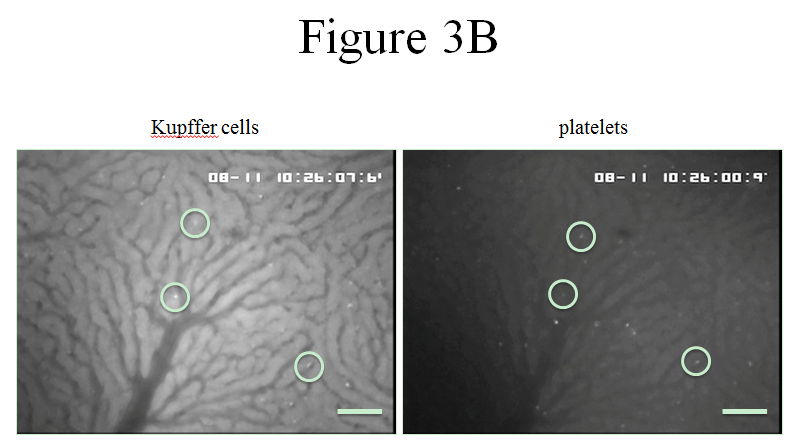
**Figure 1 Experimental design.** In all of groups, total warm hepatic ischemia was induced for 20 min by clamping the portal triad. A total of 1 × 108 fluorescence-labeled platelets, approximately 1% of all circulating platelets in the recipient rat, were injected *via* the left carotid artery 5 min before intravital microscopy (IVM).



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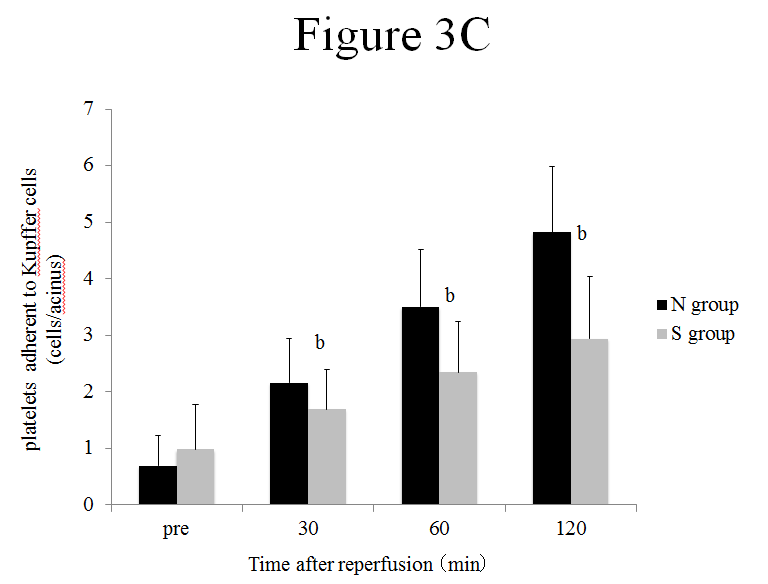
**Figure 2 Mild steatotic liver model.** Hematoxylin and eosin staining of liver specimens at 2 wk of choline deficient diet demonstrated that between 10% and 20% of hepatocytes had depositions of microvesicular lipid droplets.



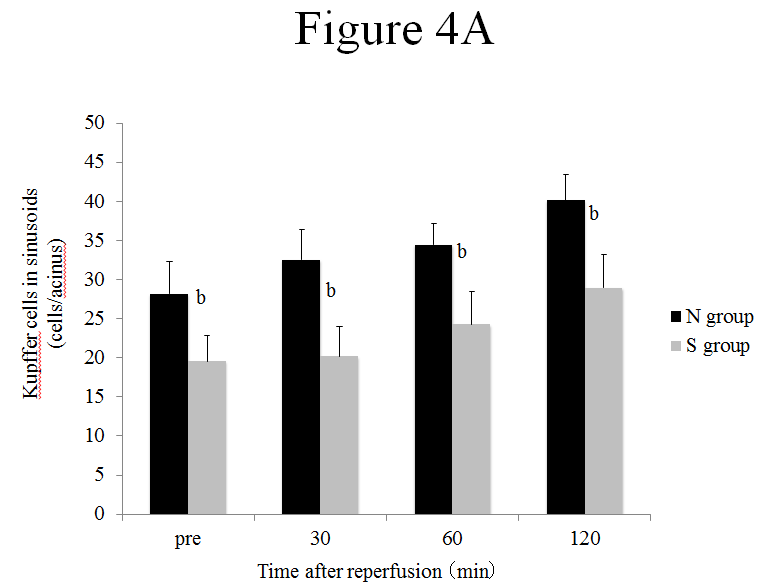


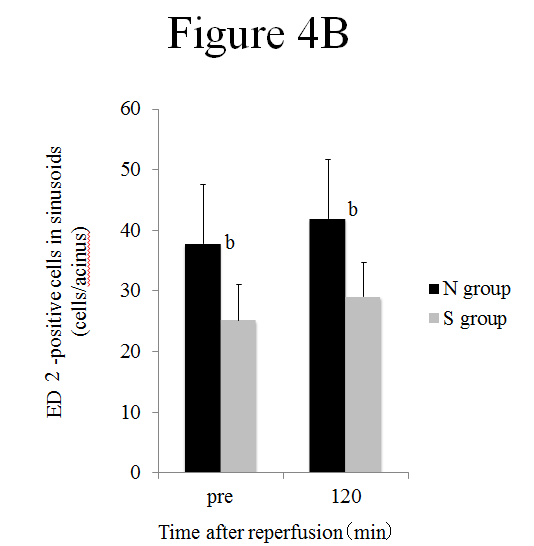
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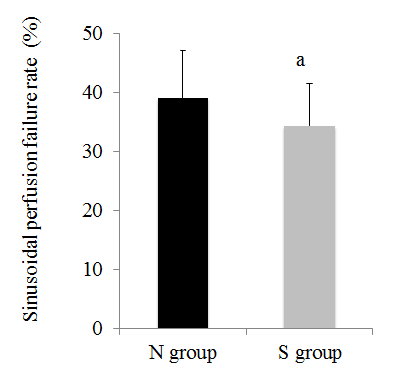


**Figure 3 Platelets and Kupffer cells dynamics in sinusoids.** A: The number of adherent platelets in sinusoids increased along with the reperfusion time in both groups. mean ± SD; *n* = 6. b*P* < 0.01 *vs* pre-ischemia in the N group. d*P* < 0.01 *vs* pre-ischemia in the S group; B: Video images of Kupffer cells (KCs) and platelets in acini 30 min after reperfusion. The field is approximately 0.2 mm2. The left figure shows the acini of fluorescently stained KCs, and in the right figure, the acini of fluorescently stained platelets. White circles indicate adhesion to KCs and platelets in the same place. C: In the S group, the number of adherent platelets adhering to KCs was significantly less than in the N group at 30 min after reperfusion and concomitant with the duration of reperfusion. mean ± SD; *n* = 6. b*P* < 0.01 *vs* the N group. N group means normal liver group, S group means mild steatotic liver group.

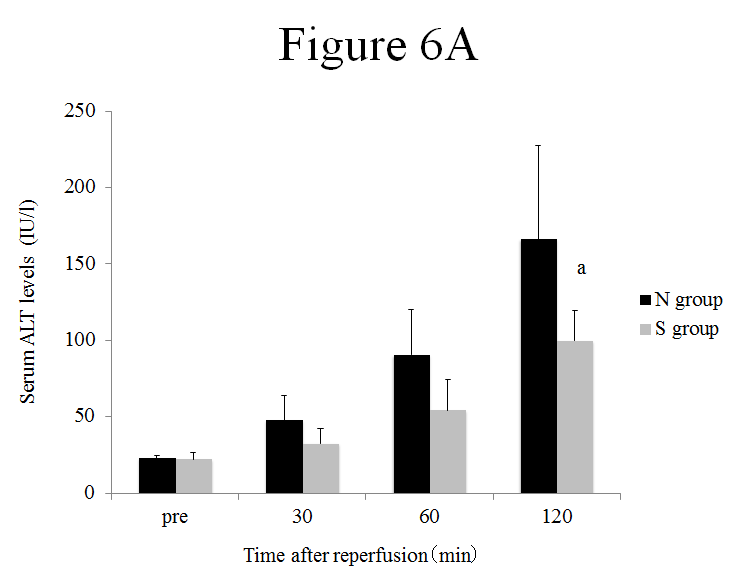


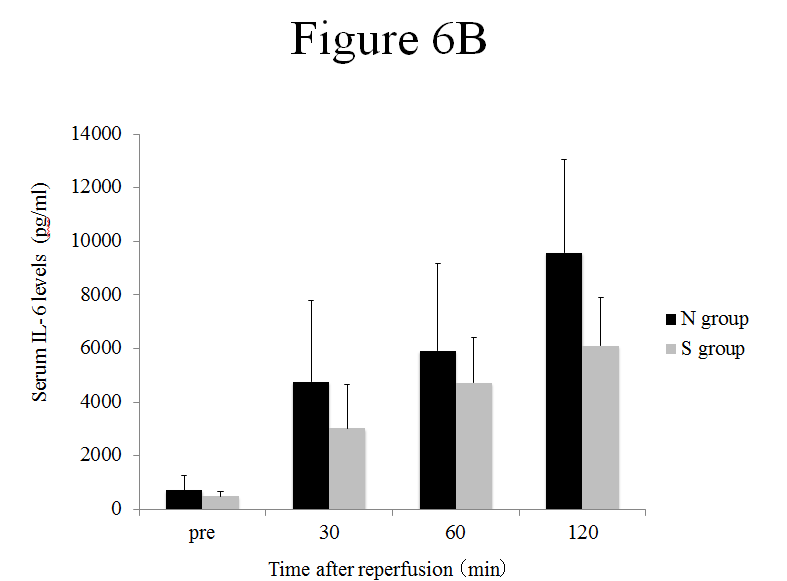


**Figure 4 The number of Kupffer cells in sinusoids (mean ± SD).** A: The number of Kupffer cells in sinusoids observed in the intravital microscopy system was decreased significantly in the S group compared to the N group at any point in time before and after ischemia reperfusion.; *n* = 6. b*P* < 0.01 *vs* the N group; B: In immunohistochemical staining, the number of ED2-positive cells was significantly less in the S group than in the N group. *n* = 6. b*P* < 0.01 *vs* the N group. N group means normal liver group, S group means mild steatotic liver group.

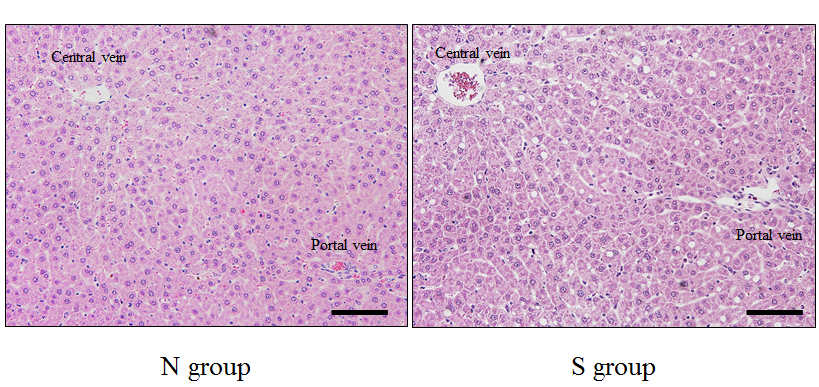


**Figure 5 Sinusoidal perfusion failure rate** **(mean ± SD).** The sinusoidal perfusion failure rate after 120 min of reperfusion was lower in the S group than in the N group.; *n* = 6. a*P* < 0.05 *vs* the N group. N group means normal liver group, S group means mild steatotic liver group.





**Figure 6 Serum alanine aminotransferase and interleukin-6 levels (mean ± SD).** A: Serum alanine aminotransferase (ALT), levels were significantly lower in the S group than in the N group at 120 min after reperfusion. *n* = 6. a*P* < 0.05 *vs* the N group; B: The concentration of serum interleukin-6 tended to be lower in the S group compared with in the N group, however, there was no significant difference in them (after 120 min of reperfusion, *P* = 0.09 *vs* the N group). *n* = 6. N group means normal liver group, S group means mild steatotic liver group.



100 μm

100 μm

**Figure 7 Histological findings after 120 min of reperfusion.** In the N group, hepatocyte structure was strongly impaired compared to the S group, and sinusoidal narrowing was observed. N group means normal liver group, S group means mild steatotic liver group.