

Caldecrin: A pancreas-derived hypocalcemic factor, regulates osteoclast formation and function

Mineko Tomomura, Akito Tomomura

Mineko Tomomura, Meikai Pharmaco-Medical Laboratory, MPL and Division of Biochemistry, Department of Oral Biology and Tissue Engineering, Meikai University School of Dentistry, Sakado, Saitama 350-0283, Japan

Akito Tomomura, Division of Biochemistry, Department of Oral Biology and Tissue Engineering, Meikai University School of Dentistry, Sakado, Saitama 350-0283, Japan

Author contributions: Tomomura M and Tomomura A contributed equally to the writing of the manuscript.

Conflict-of-interest statement: The authors declare no conflict of interest associated with this manuscript.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Mineko Tomomura, PhD, Associate Professor, Meikai Pharmaco-Medical Laboratory, MPL and Division of Biochemistry, Department of Oral Biology and Tissue Engineering, Meikai University School of Dentistry, 1-1 Keyakidai, Sakado, Saitama 350-0283, Japan. mineko-t@dent.meikai.ac.jp
Telephone: +81-49-2792766
Fax: +81-49-2712503

Received: May 29, 2015
Peer-review started: May 30, 2015
First decision: August 7, 2015
Revised: September 1, 2015
Accepted: September 29, 2015
Article in press: September 30, 2015
Published online: November 26, 2015

Abstract

Caldecrin was originally isolated from the pancreas as a

factor that reduced serum calcium levels. This secreted serine protease has chymotrypsin-like activity and is also known as chymotrypsin C; it belongs to the elastase family. Although intravenous administration of caldecrin decreases the serum calcium concentration even when its protease activity is blocked, this effect does require cleavage of caldecrin's pro-peptide by trypsin, converting it to the mature enzyme. Ectopic intramuscular expression of caldecrin prevented bone resorption in ovariectomized mice. Caldecrin inhibited parathyroid hormone-stimulated calcium release from fetal mouse long bone organ cultures. Furthermore, caldecrin suppressed the formation of osteoclasts from bone marrow cells by inhibiting the receptor activator of nuclear factor- κ B ligand (RANKL)-stimulated phospholipase C γ -calcium oscillation-calcineurin-nuclear factor of activated T-cells, cytoplasmic 1 pathway. Caldecrin also suppressed the bone resorption activity of mature osteoclasts by preventing RANKL-stimulated Src activation, calcium entry, and actin ring formation. *In vivo* and *in vitro* studies have indicated that caldecrin is a unique multifunctional protease with anti-osteoclastogenic activities that are distinct from its protease activity. Caldecrin might be a potential therapeutic target for the treatment of osteolytic diseases such as osteoporosis and osteoarthritis. This mini-review describes caldecrin's historical background and its mechanisms of action.

Key words: Serine protease; Osteoclasts; Hypocalcemia; Chymotrypsin; Bone resorption; Calcium signaling

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Caldecrin (also known as chymotrypsin C) reduces serum calcium levels. This activity is distinct from its protease activity but also requires trypsin-mediated cleavage of the pro-peptide, converting caldecrin to its active form. Ectopic intramuscular expression of caldecrin prevented bone resorption in ovariectomized mice. Caldecrin inhibited parathyroid hormone-stimulated calcium release from fetal mouse long bones. Furthermore, caldecrin suppressed receptor activator of nuclear factor-

kappa B ligand-induced activation of intracellular calcium signaling, thereby reducing osteoclast formation and bone resorption. Caldecrin is a unique multifunctional protease that possesses anti-osteoclastogenic activity, resulting in reduced serum calcium levels.

Tomomura M, Tomomura A. Caldecrin: A pancreas-derived hypocalcemic factor, regulates osteoclast formation and function. *World J Biol Chem* 2015; 6(4): 358-365 Available from: URL: <http://www.wjgnet.com/1949-8454/full/v6/i4/358.htm> DOI: <http://dx.doi.org/10.4331/wjbc.v6.i4.358>

INTRODUCTION

Calcium homeostasis is controlled by intestinal calcium absorption and calcium resorption in the kidney, as well as by bone formation and resorption. Clinical and experimental observations have also linked the pancreas to calcium homeostasis. Pancreas-derived glucagon^[1,2], amylin^[3,4], and calcitonin gene-related peptide^[5,6] have been shown to regulate calcium homeostasis, while acute and chronic pancreatitis have been shown to associate with hypocalcemia^[7].

In the 1960's, the pioneering work of Takaoka *et al.*^[8,9] demonstrated that a porcine pancreatic extract had hypocalcemic activity. In 1992, we first successfully purified a hypocalcemic factor named caldecrin from a pancreatic extract using chromatographic separation techniques including ion exchange, gel filtration chromatography, and high-performance liquid chromatography^[10]. To identify caldecrin, each fraction was intravenously administered to overnight-fasted mice and serum calcium concentrations were measured 4 h post-injection. In addition, the samples were assayed for their inhibition of parathyroid hormone-stimulated calcium release from fetal mouse long bone organ cultures. Caldecrin is an anionic protein (pI: 4.5) with a molecular weight of about 28 kDa; it was found to be a serine protease with chymotryptic activity.

In 1995, we isolated rat caldecrin cDNA from pancreatic cDNA expression library by immunoscreening with an anti-caldecrin antibody^[11]. A partial amino acid sequence of caldecrin purified from rat pancreas was completely matched with that encoded by the cDNA. The nucleotide sequence was almost identical (except for three nucleotides) to that of a PCR clone referred as elastase IV (ELA4)^[12]. Comparison of the amino acid sequences encoded by these two cDNAs indicated that the central region of caldecrin differed from that of ELA4 due to a frame shift caused by this minor nucleotide change (Figure 1). The amino acid sequences of the purified caldecrin fragments, including the central region, were consistent with the deduced amino acid sequence of caldecrin but not with that of ELA4. Over-expression of the ELA4 PCR clone in Sf9 cells caused a complete loss of secretion, low expression levels, and much lower protease activity^[13]. Furthermore, the rat genomic DNA

Table 1 Amino acid sequence similarity with rat caldecrin

Species	Pancreatic protease	Identity (%)	Similarity (%)
Rat	Caldecrin	100	100
	Chymotrypsin B	41	55
	Elastase I	51	67
	Elastase II A	59	72
	Elastase III B	57	71
Human	Caldecrin	78	88
	Chymotrypsin B	41	56
	Elastase II A	61	74
	Elastase II B	56	70
	Elastase III A	57	70
	Elastase III B	55	69
Cow	Chymotrypsin A	39	57

Sequence identity: Percent of same amino residues in a sequence alignment between 2 sequences; Sequence similarity: Percent amino acid sequence identity and percent positive substitutions between 2 sequences.

sequence matched that of the caldecrin cDNA, but not that of the ELA4 clone^[13]. Therefore, the ELA4 PCR clone may be a cloning artifact or represent a mutant caldecrin gene. In 1995, the crystalline structure of bovine chymotrypsinogen C was reported^[14-16] and its amino acid sequence was very close to that of rat caldecrin, thereby suggesting a similarity between caldecrin and chymotrypsin C (CTRC). It is now known that CTCR, caldecrin, and ELA4 are the same protein, which is encoded by the *CTRC* gene and known officially as CTCR (caldecrin), according to the *HUGO* Gene Nomenclature Committee. Table 1 compares the amino acid sequence of rat caldecrin with that of other members of the rat and human pancreatic chymotrypsin and elastase families. Caldecrin shows a greater similarity with elastase than with chymotrypsin. In addition, expressed recombinant human caldecrin also showed serum calcium-decreasing activity, even following phenylmethylsulfonyl fluoride treatment to abolish its protease activity^[17].

In 1996, another research group purified a calcium metabolism-regulating factor from the porcine pancreas by determining its stimulatory effects on proliferation of the osteosarcoma MG-63 cell line and its inhibition of 1, 25 vitamin D₃-stimulated calcium release in organ cultures^[18]. The terminal sequence of the 28-kDa protein that was isolated corresponded to that of human elastase III B. Recombinant elastase III B decreased interleukin-1-induced hypercalcemia and this effect was dependent on its protease activity. Although both have been isolated from the pancreas, caldecrin and elastase III B were found to be different molecules that exerted their hypocalcemic effects *via* different mechanisms of action.

PROTEIN STRUCTURE AND PROTEASE ACTIVITY OF CALDECRIN

The human *CTRC* gene maps to chromosome 1p36.21. The homologous mouse and rat genes are located on chromosomes 4E1 and 5q36, respectively. The *CTRC* genes consist of 8 exons in these species. Northern blot

rCal:	ATG	TTG	GGA	ATT	ACG	GTC	CTC	GCT	GCC	ATC	CTG	GCC	TGC	GCC	TCT	TGC	TGC	GGG	AAC	CCC	60
Ela4:	M	L	G	I	T	V	L	A	A	I	L	A	C	A	S	C	C	G	N	P	4
	60
	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	4
rCal:	GCC	TTC	CCA	CCT	AAC	CTG	TCA	ACC	AGA	GTG	GTA	GGA	GGA	GAG	GAT	GCT	GTC	CCC	AAC	AGC	120
Ela4:	A	F	P	P	N	L	S	T	R	V	V	G	G	E	D	A	V	P	N	S	24
	120
	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	24
rCal:	TGG	CCT	TGG	CAG	GTC	TCT	CTC	CAG	TAC	CTC	AAG	GAC	GAC	ACA	TGG	AGG	CAC	ACC	TGT	GGG	180
Ela4:	W	P	W	Q	V	S	L	Q	Y	L	K	D	D	T	W	R	H	T	C	G	44
	...	G	180
	*	A	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	44
rCal:	GGA	AGT	CTC	ATC	ACC	ACC	AGC	CAC	GTC	CTC	ACT	GCC	GCC	CAC	TGC	ATC	AAC	AAA	GAC	TTC	240
Ela4:	G	S	L	I	T	T	S	H	V	L	T	A	A	H	C	I	N	K	D	F	64
	240
	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	64
rCal:	ACT	TAC	CGT	GTG	GGC	CTG	GGG	AAG	TAT	AAT	CTG	ACA	GTG	GAG	GAT	GAG	GAA	GGC	TCC	GTG	300
Ela4:	T	Y	R	V	G	L	G	K	Y	N	L	T	V	E	D	E	E	G	S	V	84
	300
	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	A	*	A	P	C	84
rCal:	TAC	GCT	GAG	GTG	GAC	ACC	ATC	TAC	GTC	CAT	GAG	AAG	TGG	AAC	CGA	CTC	TTC	CTG	TGG	AAC	360
Ela4:	Y	A	E	V	D	T	I	Y	V	H	E	K	W	N	R	L	F	L	W	N	104
	360
	A	C	T	G	A	C	C	T	C	A	G	A	G	A	C	T	C	T	G	A	360
	T	L	R	W	T	P	S	T	S	M	R	S	G	T	D	S	S	C	G	T	104
rCal:	GAC	ATC	GCT	ATC	ATT	AAG	TTG	GCT	GAG	CCT	GTG	GAA	CTG	AGC	AAC	ACC	ATC	CAG	GTG	GCC	420
Ela4:	D	I	A	I	I	K	L	A	E	P	V	E	L	S	N	T	I	Q	V	A	124
	420
	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	124
rCal:	TGC	ATC	CCA	GAG	GAA	GGT	TCC	CTG	CTG	CCT	CAG	GAC	TAT	CCC	TGC	TAT	GTC	ACG	GGC	TGG	480
Ela4:	C	I	P	E	E	G	S	L	L	P	Q	D	Y	P	C	Y	V	T	G	W	144
	480
	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	144
rCal:	GGT	CGC	CTC	TGG	ACC	AAT	GGT	CCC	ATC	GCT	GAA	GTG	CTC	CAG	CAG	GGC	CTG	CAG	CCC	ATC	540
Ela4:	G	R	L	W	T	N	G	P	I	A	E	V	L	Q	Q	G	L	Q	P	I	164
	540
	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	164
rCal:	GTG	AGC	CAT	GCC	ACG	TGC	TCC	AGG	TTG	GAC	TGG	TGG	TTC	ATC	AAG	GTC	CGG	AAG	ACG	ATG	600
Ela4:	V	S	H	A	T	C	S	R	L	D	W	W	F	I	K	V	R	K	T	M	184
	600
	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	184
rCal:	GTG	TGC	GCT	GGG	GGT	GAT	GGC	GTC	ATC	TCT	GCC	TGT	AAC	GGA	GAT	TCT	GGC	GGC	CCA	CTG	660
Ela4:	V	C	A	G	G	D	G	V	I	S	A	C	N	G	D	S	G	G	P	L	204
	660
	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	204
rCal:	AAC	TGC	CAA	GCA	GAA	GAC	GGC	TCA	TGG	CAG	GTG	CAC	GGC	ATC	GTG	AGC	TTC	GGT	TCC	AGT	720
Ela4:	N	C	Q	A	E	D	G	S	W	Q	V	H	G	I	V	S	F	G	S	S	224
	720
	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	244
rCal:	AGC	GGC	TGC	AAC	GTA	CAC	AAG	AAA	CCG	GTA	GTC	TTC	ACC	CGA	GTG	TCT	GCC	TAC	AAT	GAC	780
Ela4:	S	G	C	N	V	H	K	K	P	V	V	F	T	R	V	S	A	Y	N	D	244
	780
	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	244
rCal:	TGG	ATC	AAC	GAG	AAA	ATA	CAA	CTG													
Ela4:	W	I	N	E	K	I	Q	L													
													
	*	*	*	*	*	*	*	*													

Figure 1 Nucleotide and deduced amino acid sequences of rat caldecrin (rCal) and elastase IV (Ela4). The nucleotide (upper row) and amino acid (lower row) sequences of the indicated molecules are shown. The dots and asterisks indicate nucleotides and amino acid residues, respectively, that are conserved between rCal and Ela4. Circle: Charge-relay system; Vertical arrowhead: Proteolytic cleavage site.

analysis has indicated that caldecrin is mainly expressed in the pancreas (Figure 2A).

CTRC (caldecrin) is a single protein consisting of 268 amino acids, with a signal peptide (16 amino acids), pro-peptide (13 amino acids), and the mature protein (239 amino acids; Figure 2B). The three-dimensional structure demonstrated that five disulfide bridges were formed at Cys1-Cys125 (according to the chymotrypsin numbering),

Cys43-Cys59, Cys139-Cys206, Cys170-Cys186, and Cys196-Cys227 (Figure 2B). CTCR (caldecrin) was shown to have a two-barrel structure, each composed of 6-7 β -sheets and a C-terminal α -helix long tail^[14-16] (Figure 2C). Following tryptic cleavage at Arg13-Val14, the caldecrin pro-peptide remains associated with the mature enzyme *via* the Cys1-Cys125 disulfide bridge; this generates a structure resembling those of chymotrypsin A and B, as

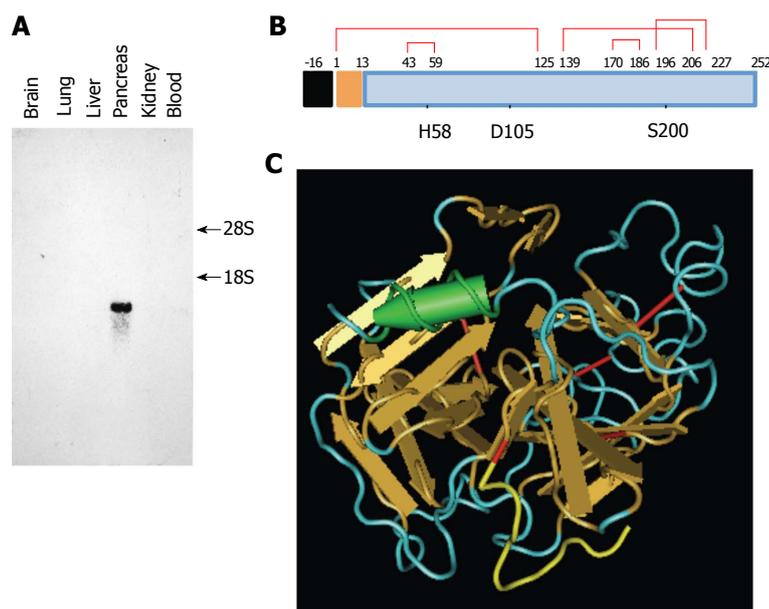


Figure 2 Caldecrin expression and protein structure. A: Caldecrin expression was analyzed by Northern blot. 18S, 28S: 18S, 28S ribosomal RNA; B: Domain structures of caldecrin. Black box: signal peptide; orange box: pro-peptide; blue box: mature protein; red line: disulfide bridges with cysteine number; the H (histidine), D (aspartic acid), S (serine) catalytic triad; C: Ribbon diagram of the crystal structure of human caldecrin (adapted from PDB ID: 4H4F, prepared from [16]). Red line: Disulfide bridge; Yellow line: Pro-peptide; Arrow: β -sheet structure; Cylinder: α -helix structure.

well as elastase II A, but not those of elastase I, III A, and III B, where the pro-peptide is removed from the mature enzyme after tryptic activation^[11,14-16].

CTRC (caldecrin) is a serine protease with the characteristic charge-relayed catalytic triad (His58, Asp105, and Ser200), located in the active site cleft between the barrel structures^[14-16]. After tryptic activation, caldecrin changes its structure to a substrate-accessible catalytic cleft form. Active caldecrin hydrolyzes the leucyl bond (*e.g.*, in the N-Succinyl-Ala-Ala-Pro-Leu-p-nitroanilide substrate) more efficiently than chymotrypsin A and B; Caldecrin also cleaves the phenylalanyl bond (*e.g.*, in the N-Succinyl-Ala-Ala-Pro-Phe-p-nitroanilide substrate) and the tyrosyl bond (*e.g.*, in the N-Succinyl-Leu-Leu-Val-Tyr-p-nitroanilide substrate)^[10,19-21]. The protease activity of caldecrin is inhibited by serine protease inhibitors (phenylmethylsulfonyl fluoride or diisopropyl fluorophosphate), chymotrypsin inhibitor (chymostatin), and the Bowman-Birk trypsin and chymotrypsin inhibitor. The amino acid sequence and protease activity of caldecrin indicate that it is a hybrid of chymotrypsin and elastase.

CALDECRIN AND BONE METABOLISM

Caldecrin produces dose-dependent decreases in serum calcium concentrations^[10]. The administration of purified porcine and rat caldecrin *via* the tail vein of mice decreased their serum calcium concentration dose-dependently and the maximum effect was attained 2-4 h post-injection with 20-100 μ g (about 0.7-3.6 nmol)/kg body weight. The hypocalcemic potency of caldecrin was almost equivalent to that of porcine calcitonin (1 nmol/kg body weight, Tomomura *et al.*^[10] data not shown). The caldecrin proform (pro calderon), purified from the porcine pancreas in the presence of diisopropyl fluorophosphate, appeared to show time- and concentration-dependent chymotryptic activity following cleavage by trypsin. Administration of activated caldecrin reduced the serum calcium level

in mice, even after treatment with the serine protease inhibitor, phenylmethylsulfonyl fluoride, which abolished the chymotryptic activity. However, administration of procaldecrin did not decrease serum calcium levels^[22]. Recombinant rat^[11] and human^[17] caldecrin also decreased serum calcium levels. In addition, rat protease activity-deficient caldecrin mutants (with His58Ala or Ser200Ala substitutions) decreased the levels of serum calcium. Therefore, the effect of caldecrin on serum calcium levels *in vivo* requires its activation by trypsin cleavage. An intramolecular responsive region required for this calcium decreasing activity may therefore be exposed by trypsin activation.

The caldecrin-induced serum calcium decrease occurred concomitantly with a decrease in the serum concentration of hydroxyproline, which is a marker of bone resorption. This observation suggested that this serum calcium decrease may be due to the suppression of bone resorption^[10]. The effects of caldecrin on osteoclast function have also been investigated; recombinant wild-type and protease activity-deficient mutant caldecrin produced concentration-dependent suppression of bone resorption in isolated rabbit mature osteoclasts^[23].

Osteoclasts execute bone resorption, which is modulated by macrophage colony-stimulating factor and receptor activator of nuclear factor-kappa B (NF- κ B) ligand (RANKL), produced by osteoblasts and osteocytes. An imbalance between bone formation and resorption leads to bone diseases, including osteoporosis. Osteoclast differentiation and maturation involves the following three steps: (1) Osteoclast precursor cells are generated from bone marrow cells in response to macrophage colony-stimulating factor; (2) osteoclasts begin to differentiate from the precursor cells following stimulation by RANKL; and (3) at the later stage of differentiation, osteoclasts fuse to become multinucleated giant cells, leading to the cytoskeletal actin ring formation required for bone resorption. These processes are tightly regulated to

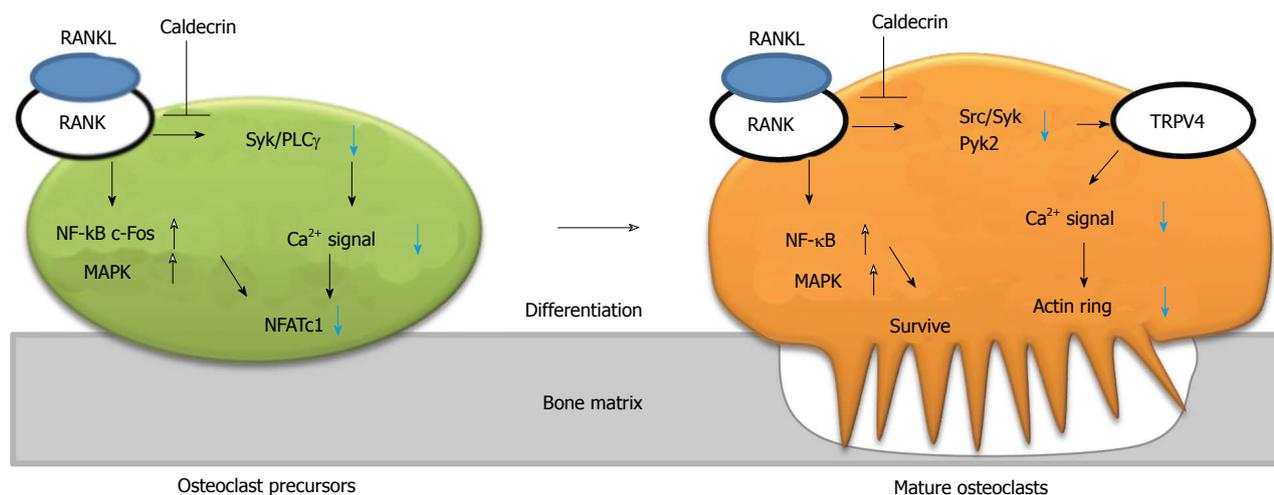


Figure 3 Caldecrin suppresses RANKL-induced osteoclast differentiation and bone resorption. RANKL binds to its receptor (RANK) on the osteoclast precursor, leading to simultaneous activation of two pathways: the NF- κ B/ MAPK/c-Fos axis and the Syk/PLC γ -calcium oscillation- NFATc1 axis. In mature osteoclasts, RANKL also activates the NF- κ B/MAPK and Src/Syk/Pyk2-TRPV4 channel–calcium entry–actin ring formation axes. Caldecrin inhibits the latter pathways (but not the NF- κ B/MAPK pathway) in the precursor and mature osteoclasts. TRPV4: Transient receptor potential vanilloid channel 4; MAPK: Mitogen-activated protein kinase; RANKL: RANK ligand; PLC γ : Phospholipase C γ ; Pyk2: Proline-rich tyrosine kinase 2; NFATc1: Nuclear factor of activated T-cells cytoplasmic 1; NF- κ B: Nuclear factor-kappa B; Syk: Spleen tyrosine kinase.

maintain bone homeostasis, and many molecules are involved in osteoclast differentiation^[24-26]. The key molecule involved in osteoclastogenesis is RANKL, which is a member of the tumor necrosis factor superfamily that is expressed by osteoblasts and osteocytes in membrane-bound and secreted forms^[27-33]. RANKL induces osteoclast differentiation by activating two signaling pathways: the mitogen-activated protein kinase (MAPK), NF- κ B, and c-Fos activation axis and the phospholipase C γ (PLC γ)-mediated calcium oscillation-calcineurin-nuclear factor of activated T-cells, cytoplasmic 1 (NFATc1) axis. Caldecrin did not inhibit macrophage colony-stimulating factor-induced osteoclast progenitor formation from bone marrow cells but did inhibit RANKL-induced osteoclast differentiation, even in the absence of protease activity^[34]. Caldecrin inhibited the RANKL-stimulated spleen tyrosine kinase- and PLC γ -induced calcium oscillation, leading to an inhibition of calcineurin and NFATc1 activity (Figure 3). Caldecrin also inhibited the RANKL-mediated actin ring formation in mature osteoclasts, which is associated with RANKL-evoked calcium entry *via* the transient receptor potential vanilloid channel 4^[35]. Caldecrin significantly inhibited RANKL-stimulated phosphorylation of c-Src in association with spleen tyrosine kinase, which is upstream of transient receptor potential vanilloid channel 4 and actin ring formation. On the other hand, caldecrin did not inhibit RANKL-mediated stimulation of MAPK, NF- κ B, and c-Fos activation in osteoclast precursors or mature osteoclasts^[34,35]. Therefore, caldecrin antagonized the RANKL-stimulated calcium signaling pathway involved in both osteoclast differentiation and activation.

Caldecrin is a therapeutic target in osteoporosis. The ovariectomized mouse provides a model of postmenopausal osteoporosis and exhibits an increased serum calcium level due to elevated bone resorption. This is

evidenced by an increase in the bone surface to bone volume ratio, increased trabecular separation, decreased bone volume density, and decreased trabecular thickness and number. Expression of the caldecrin plasmid vector, which harbors the wild-type rat caldecrin cDNA, in the femoral muscle of this mouse model reversed this increase in serum calcium levels and restored bone resorption parameters to normal levels^[36].

An important, but unaddressed, question relates to how the caldecrin released from the pancreas targets the bone. Recently, osteocalcin, which is osteoblast-derived, stored in the bone matrix, and then released by osteoclastic bone resorption, was shown to increase insulin secretion from pancreatic islets^[37,38]. This activity appears to provide a physiological link between bone and pancreas, in relation to the regulation of energy metabolism. It is possible that some of the caldecrin derived from the pancreas enters the circulation and then inhibits osteoclasts, in order to regulate calcium homeostasis. The physiological activation and functions of caldecrin are not defined; however, considering its obvious effects on serum calcium levels and osteogenesis, caldecrin might be an intrinsic calcium regulating factor. The expression and distribution of caldecrin, peptide fragments of caldecrin, and its binding proteins should be explored in order to determine their physiological roles in bone metabolism.

OTHER BIOLOGICAL ASPECTS

The *CTRC* gene modulates risk for pancreatitis. Rosendahl *et al.*^[39] reported that *CTRC* gene mutations were significantly associated with hereditary chronic pancreatitis. Masson *et al.*^[40] also identified a *CTRC* mutation in patients with idiopathic chronic pancreatitis. *CTRC* hydrolyzes the pro-peptide and calcium-binding

loop of the trypsinogens, enhancing their activation and degradation, respectively^[41-43]. Loss-of-function *CTRC* variants increase the risk for chronic pancreatitis. *CTRC* is also a susceptibility gene for tropical calcific pancreatitis, which is a juvenile form of chronic nonalcoholic pancreatitis that occurs in Asians and Africans and is associated with nearly 90% pancreatic calcium deposition^[44].

It is of clinical interest that five decades ago, Takaoka *et al.*^[8,9,45] administered pancreatic extract to patients diagnosed with myasthenia gravis and muscular dystrophy. The symptoms of the patients treated with the extract improved progressively, suggesting that the hypocalcemic effect of the extract could have contributed to protecting them against the development of muscular dystrophy. The effect of caldecrin was also investigated in the *dy/dy* muscular dystrophic mouse model^[46]. These mice genetically lack M-laminin and exhibit defective muscle basement membranes. Peritoneal administration of caldecrin protein or intramuscular expression of a caldecrin vector inhibited muscular destruction in the *dy/dy* mice. This indicated that caldecrin was responsible for the effects of the pancreatic extract on muscular dystrophy.

In 2011, Lacruz *et al.*^[47] found that *CTRC* (caldecrin) was expressed by ameloblasts and was up-regulated during enamel maturation, suggesting that caldecrin might be involved in tooth development.

CTRC (caldecrin) has been reported to be associated with pancreatic cancer, where its expression is drastically reduced. Individuals with chronic pancreatitis who show low or no activity of caldecrin show an increased risk for pancreatic cancer^[48]. Furthermore, Wang *et al.*^[49] demonstrated that overexpression of *CTRC* (caldecrin) downregulated the migration of human pancreatic adenocarcinoma Aspc-1 cells, whereas the knockdown of *CTRC* (caldecrin) increased cell migration. It would be interesting to explore the potential use of caldecrin in pancreatic cancer diagnosis and treatment. In addition, breast cancer is highly associated with osteolytic metastatic disease. RANKL is important in mammary gland development and also in the progression of metastatic breast cancer cells^[50,51]. RANKL may partly contribute to the activation of metastatic breast cancer *via* the calcineurin/NFAT pathway^[52], which is modulated by caldecrin. It would therefore be interesting to investigate whether caldecrin suppresses RANKL-dependent tumor metastases.

CONCLUSION

The serum calcium-decreasing factor, caldecrin, was discovered in the pancreas. Caldecrin inhibits osteoclast differentiation and bone resorption in mature osteoclasts *via* inhibition of RANKL-induced intracellular calcium signaling. This effect occurs independently of its inherent protease activity. Therefore, caldecrin might be a potential therapeutic target for the treatment of osteolytic diseases such as osteoporosis and osteoarthritis.

REFERENCES

- 1 **Williams GA**, Bowser EN, Henderson WJ. Mode of hypocalcemic action of glucagon in the rat. *Endocrinology* 1969; **85**: 538-541 [PMID: 5793033 DOI: 10.1210/endo-85-3-537]
- 2 **Stern PH**, Bell NH. Effects of glucagon on serum calcium in the rat and on bone resorption in tissue culture. *Endocrinology* 1970; **87**: 111-117 [PMID: 4315590 DOI: 10.1210/endo-87-1-111]
- 3 **Zaidi M**, Datta HK, Bevis PJ, Wimalawansa SJ, MacIntyre I. Amylin-amide: a new bone-conserving peptide from the pancreas. *Exp Physiol* 1990; **75**: 529-536 [PMID: 2223054 DOI: 10.1113/expphysiol.1990.sp003429]
- 4 **Alam AS**, Moonga BS, Bevis PJ, Huang CL, Zaidi M. Amylin inhibits bone resorption by a direct effect on the motility of rat osteoclasts. *Exp Physiol* 1993; **78**: 183-196 [PMID: 8385961 DOI: 10.1113/expphysiol.1993.sp003679]
- 5 **D'Souza SM**, MacIntyre I, Giris SI, Mundy GR. Human synthetic calcitonin gene-related peptide inhibits bone resorption in vitro. *Endocrinology* 1986; **119**: 58-61 [PMID: 3487444 DOI: 10.1210/endo-119-1-58]
- 6 **Zaidi M**, Chambers TJ, Gaines Das RE, Morris HR, MacIntyre I. A direct action of human calcitonin gene-related peptide on isolated osteoclasts. *J Endocrinol* 1987; **115**: 511-518 [PMID: 3502132 DOI: 10.1677/joe.0.1150511]
- 7 **D'Souza A**, Floch MH. Calcium metabolism in pancreatic disease. *Am J Clin Nutr* 1973; **26**: 352-361 [PMID: 4347666]
- 8 **Takaoka Y**, Hiwaki C, Ozawa H, Ichinose M, Otsubo Y, Shikaya T. A pancreatic protein anabolic extract. Proposal of a protein anabolic extract from pancreas. I. Preliminary report. *Acta Med Nagasaki* 1966; **10**: 51-57 [PMID: 5961056]
- 9 **Takaoka Y**, Takamori M, Ichinose M, Shikaya T, Igawa N. Hypocalcemic action of a pancreatic factor and its clinical significance on the myasthenic patients. *Acta Med Nagasaki* 1969; **14**: 28-35 [PMID: 5393331]
- 10 **Tomomura A**, Fukushige T, Noda T, Noikura T, Saheki T. Serum calcium-decreasing factor (caldecrin) from porcine pancreas has proteolytic activity which has no clear connection with the calcium decrease. *FEBS Lett* 1992; **301**: 277-281 [PMID: 1577166 DOI: 10.1016/0014-5793(92)80256-G]
- 11 **Tomomura A**, Tomomura M, Fukushige T, Akiyama M, Kubota N, Kumaki K, Nishii Y, Noikura T, Saheki T. Molecular cloning and expression of serum calcium-decreasing factor (caldecrin). *J Biol Chem* 1995; **270**: 30315-30321 [PMID: 8530454 DOI: 10.1074/jbc.270.51.30315]
- 12 **Kang J**, Wiegand U, Müller-Hill B. Identification of cDNAs encoding two novel rat pancreatic serine proteases. *Gene* 1992; **110**: 181-187 [PMID: 1537555 DOI: 10.1016/0378-1119(92)90646-7]
- 13 **Yoshino-Yasuda I**, Kobayashi K, Akiyama M, Itoh H, Tomomura A, Saheki T. Caldecrin is a novel-type serine protease expressed in pancreas, but its homologue, elastase IV, is an artifact during cloning derived from caldecrin gene. *J Biochem* 1998; **123**: 546-554 [PMID: 9538241 DOI: 10.1093/oxfordjournals.jbchem.a021971]
- 14 **Gomis-Rüth FX**, Gómez M, Bode W, Huber R, Avilés FX. The three-dimensional structure of the native ternary complex of bovine pancreatic procarboxypeptidase A with proproteinase E and chymotrypsinogen C. *EMBO J* 1995; **14**: 4387-4394 [PMID: 7556081]
- 15 **Gomis-Rüth FX**, Gómez-Ortiz M, Vendrell J, Ventura S, Bode W, Huber R, Avilés FX. Crystal structure of an oligomer of proteolytic zymogens: detailed conformational analysis of the bovine ternary complex and implications for their activation. *J Mol Biol* 1997; **269**: 861-880 [PMID: 9223647 DOI: 10.1006/jmbi.1997.1040]
- 16 **Batra J**, Szabó A, Caulfield TR, Soares AS, Sahin-Tóth M, Radisky ES. Long-range electrostatic complementarity governs substrate recognition by human chymotrypsin C, a key regulator of digestive enzyme activation. *J Biol Chem* 2013; **288**: 9848-9859 [PMID: 23430245 DOI: 10.1074/jbc.M113.457382]
- 17 **Tomomura A**, Akiyama M, Itoh H, Yoshino I, Tomomura M, Nishii Y, Noikura T, Saheki T. Molecular cloning and expression of human caldecrin. *FEBS Lett* 1996; **386**: 26-28 [PMID: 8635596 DOI: 10.1016/S0167-4834(96)00026-8]

- 16/0014-5793(96)00377-8]
- 18 **Izbicka E**, Yoneda T, Takaoka Y, Horn D, Williams P, Mundy GR. Identification of a novel bone/calcium metabolism-regulating factor in porcine pancreas. *J Biol Chem* 1996; **271**: 23230-23234 [PMID: 8798519 DOI: 10.1074/jbc.271.38.23230]
 - 19 **Folk JE**, Schirmer EW. Chymotrypsin C. I. Isolation of the zymogen and the active enzyme: preliminary structure and specificities. *J Biol Chem* 1965; **240**: 181-192 [PMID: 14253410]
 - 20 **Folk JE**, Cole PW. Chymotrypsin C. II. Enzymatic specificity toward several polypeptides. *J Biol Chem* 1965; **240**: 193-197 [PMID: 14253411]
 - 21 **Keil-Dlouha V**, Puigserver A, Marie A, Keil B. On subunit II of bovine procarboxypeptidase A. Enzymatic specificity after tryptic activation. *Biochim Biophys Acta* 1972; **276**: 531-535 [PMID: 4672120 DOI: 10.1016/0005-2744(72)91013-3]
 - 22 **Tomomura A**, Fukushima T, Tomomura M, Noikura T, Nishii Y, Saheki T. Caldecrin proform requires trypsin activation for the acquisition of serum calcium-decreasing activity. *FEBS Lett* 1993; **335**: 213-216 [PMID: 8253199 DOI: 10.1016/0014-5793(93)80732-A]
 - 23 **Tomomura A**, Yamada H, Fujimoto K, Inaba A, Katoh S. Determination of amino acid sequence responsible for suppression of bone resorption by serum calcium-decreasing factor (caldecrin). *FEBS Lett* 2001; **508**: 454-458 [PMID: 11728471 DOI: 10.1016/S0014-5793(01)03107-6]
 - 24 **Suda T**, Takahashi N, Martin TJ. Modulation of osteoclast differentiation. *Endocr Rev* 1992; **13**: 66-80 [PMID: 1555533]
 - 25 **Teitelbaum SL**. Bone resorption by osteoclasts. *Science* 2000; **289**: 1504-1508 [PMID: 10968780 DOI: 10.1126/science.289.5484.1504]
 - 26 **Asagiri M**, Takayanagi H. The molecular understanding of osteoclast differentiation. *Bone* 2007; **40**: 251-264 [PMID: 17098490 DOI: 10.1016/j.bone.2006.09.023]
 - 27 **Anderson DM**, Maraskovsky E, Billingsley WL, Dougall WC, Tometsko ME, Roux ER, Teepe MC, DuBose RF, Cosman D, Galibert L. A homologue of the TNF receptor and its ligand enhance T-cell growth and dendritic-cell function. *Nature* 1997; **390**: 175-179 [PMID: 9367155 DOI: 10.1038/36593]
 - 28 **Wong BR**, Rho J, Arron J, Robinson E, Orlinick J, Chao M, Kalachikov S, Cayani E, Bartlett FS, Frankel WN, Lee SY, Choi Y. TRANCE is a novel ligand of the tumor necrosis factor receptor family that activates c-Jun N-terminal kinase in T cells. *J Biol Chem* 1997; **272**: 25190-25194 [PMID: 9312132 DOI: 10.1074/jbc.272.40.25190]
 - 29 **Lacey DL**, Timms E, Tan HL, Kelley MJ, Dunstan CR, Burgess T, Elliott R, Colombero A, Elliott G, Scully S, Hsu H, Sullivan J, Hawkins N, Davy E, Capparelli C, Eli A, Qian YX, Kaufman S, Sarosi I, Shalhoub V, Senaldi G, Guo J, Delaney J, Boyle WJ. Osteoprotegerin ligand is a cytokine that regulates osteoclast differentiation and activation. *Cell* 1998; **93**: 165-176 [PMID: 9568710 DOI: 10.1016/S0092-8674(00)81569-X]
 - 30 **Yasuda H**, Shima N, Nakagawa N, Yamaguchi K, Kinoshita M, Mochizuki S, Tomoyasu A, Yano K, Goto M, Murakami A, Tsuda E, Morinaga T, Higashio K, Udagawa N, Takahashi N, Suda T. Osteoclast differentiation factor is a ligand for osteoprotegerin/osteoclastogenesis-inhibitory factor and is identical to TRANCE/RANKL. *Proc Natl Acad Sci USA* 1998; **95**: 3597-3602 [PMID: 9520411 DOI: 10.1073/pnas.95.7.3597]
 - 31 **Fuller K**, Wong B, Fox S, Choi Y, Chambers TJ. TRANCE is necessary and sufficient for osteoblast-mediated activation of bone resorption in osteoclasts. *J Exp Med* 1998; **188**: 997-1001 [PMID: 9730902 DOI: 10.1084/jem.188.5.997]
 - 32 **Nakashima T**, Hayashi M, Fukunaga T, Kurata K, Oh-Hora M, Feng JQ, Bonewald LF, Kodama T, Wutz A, Wagner EF, Penninger JM, Takayanagi H. Evidence for osteocyte regulation of bone homeostasis through RANKL expression. *Nat Med* 2011; **17**: 1231-1234 [PMID: 21909105 DOI: 10.1038/nm.2452]
 - 33 **Xiong J**, Onal M, Jilka RL, Weinstein RS, Manolagas SC, O'Brien CA. Matrix-embedded cells control osteoclast formation. *Nat Med* 2011; **17**: 1235-1241 [PMID: 21909103 DOI: 10.1038/nm.2448]
 - 34 **Hasegawa H**, Kido S, Tomomura M, Fujimoto K, Ohi M, Kiyomura M, Kanegae H, Inaba A, Sakagami H, Tomomura A. Serum calcium-decreasing factor, caldecrin, inhibits osteoclast differentiation by suppression of NFATc1 activity. *J Biol Chem* 2010; **285**: 25448-25457 [PMID: 20547767 DOI: 10.1074/jbc.M109.068742]
 - 35 **Tomomura M**, Hasegawa H, Suda N, Sakagami H, Tomomura A. Serum calcium-decreasing factor, caldecrin, inhibits receptor activator of NF- κ B ligand (RANKL)-mediated Ca²⁺ signaling and actin ring formation in mature osteoclasts via suppression of Src signaling pathway. *J Biol Chem* 2012; **287**: 17963-17974 [PMID: 22461633 DOI: 10.1074/jbc.M112.358796]
 - 36 **Oi M**, Kido S, Hasegawa H, Fujimoto K, Tomomura M, Kanegae H, Suda N, Tomomura A. Inhibitory effects on bone resorption in postmenopausal osteoporosis model mice by delivery of serum calcium decreasing factor (Caldecrin) gene. *J Meikai Dent Med* 2011; **40**: 146-154
 - 37 **Lee NK**, Sowa H, Hinoi E, Ferron M, Ahn JD, Confavreux C, Dacquin R, Mee PJ, McKee MD, Jung DY, Zhang Z, Kim JK, Mauvais-Jarvis F, Ducy P, Karsenty G. Endocrine regulation of energy metabolism by the skeleton. *Cell* 2007; **130**: 456-469 [PMID: 17693256 DOI: 10.1016/j.cell.2007.05.047]
 - 38 **Confavreux CB**, Levine RL, Karsenty G. A paradigm of integrative physiology, the crosstalk between bone and energy metabolisms. *Mol Cell Endocrinol* 2009; **310**: 21-29 [PMID: 19376193 DOI: 10.1016/j.mce.2009.04.004]
 - 39 **Rosendahl J**, Witt H, Szmola R, Bhatia E, Oszvári B, Landt O, Schulz HU, Gress TM, Pfützner R, Löhr M, Kovacs P, Blüher M, Stumvoll M, Choudhuri G, Hegyi P, te Morsche RH, Drenth JP, Truninger K, Macek M, Puhl G, Witt U, Schmidt H, Büning C, Ockenga J, Kage A, Groneberg DA, Nickel R, Berg T, Wiedenmann B, Bödeker H, Keim V, Mössner J, Teich N, Sahin-Tóth M. Chymotrypsin C (CTRC) variants that diminish activity or secretion are associated with chronic pancreatitis. *Nat Genet* 2008; **40**: 78-82 [PMID: 18059268 DOI: 10.1038/ng.2007.44]
 - 40 **Masson E**, Chen JM, Scotet V, Le Maréchal C, Férec C. Association of rare chymotrypsinogen C (CTRC) gene variations in patients with idiopathic chronic pancreatitis. *Hum Genet* 2008; **123**: 83-91 [PMID: 18172691 DOI: 10.1007/s00439-007-0459-3]
 - 41 **Nemoda Z**, Sahin-Tóth M. Chymotrypsin C (caldecrin) stimulates autoactivation of human cationic trypsinogen. *J Biol Chem* 2006; **281**: 11879-11886 [PMID: 16505482 DOI: 10.1074/jbc.M600124200]
 - 42 **Szmola R**, Sahin-Tóth M. Chymotrypsin C (caldecrin) promotes degradation of human cationic trypsin: identity with Rinderknecht's enzyme Y. *Proc Natl Acad Sci USA* 2007; **104**: 11227-11232 [PMID: 17592142 DOI: 10.1073/pnas.0703714104]
 - 43 **Zhou J**, Sahin-Tóth M. Chymotrypsin C mutations in chronic pancreatitis. *J Gastroenterol Hepatol* 2011; **26**: 1238-1246 [PMID: 21631589 DOI: 10.1111/j.1440-1746.2011.06791.x]
 - 44 **Paliwal S**, Bhaskar S, Mani KR, Reddy DN, Rao GV, Singh SP, Thomas V, Chandak GR. Comprehensive screening of chymotrypsin C (CTRC) gene in tropical calcific pancreatitis identifies novel variants. *Gut* 2013; **62**: 1602-1606 [PMID: 22580415 DOI: 10.1136/gutjnl-2012-302448]
 - 45 **Takaoka Y**, Takamori M, Ichinose M, Tsujihata M, Mori M, Eguchi K. A protein anabolic factor from porcine pancreas and its effects on mice and patients with progressive muscular dystrophy. *IRCS J Med Sci* 1977; **5**: 103-105
 - 46 **Tomomura M**, Fujii T, Sakagami H, Tomomura A. Serum calcium-decreasing factor, caldecrin, ameliorates muscular dystrophy in dy/dy mice. *In Vivo* 2011; **25**: 157-163 [PMID: 21471529]
 - 47 **Lacruz RS**, Smith CE, Smith SM, Hu P, Bringas P, Sahin-Tóth M, Moradian-Oldak J, Paine ML. Chymotrypsin C (caldecrin) is associated with enamel development. *J Dent Res* 2011; **90**: 1228-1233 [PMID: 21828354 DOI: 10.1177/0022034511418231]
 - 48 **Lowenfels AB**, Maisonneuve P, DiMaggio EP, Elitsur Y, Gates LK, Perrault J, Whitcomb DC. Hereditary pancreatitis and the risk of pancreatic cancer. International Hereditary Pancreatitis Study Group. *J Natl Cancer Inst* 1997; **89**: 442-446 [PMID: 9091646 DOI: 10.1093/jnci/89.6.442]
 - 49 **Wang H**, Sha W, Liu Z, Chi CW. Effect of chymotrypsin C and related proteins on pancreatic cancer cell migration. *Acta Biochim*

- Biophys Sin* (Shanghai) 2011; **43**: 362-371 [PMID: 21460362 DOI: 10.1093/abbs/gmr022]
- 50 **Schramek D**, Leibbrandt A, Sigl V, Kenner L, Pospisilik JA, Lee HJ, Hanada R, Joshi PA, Aliprantis A, Glimcher L, Pasparakis M, Khokha R, Ormandy CJ, Widschwendter M, Schett G, Penninger JM. Osteoclast differentiation factor RANKL controls development of progestin-driven mammary cancer. *Nature* 2010; **468**: 98-102 [PMID: 20881962 DOI: 10.1038/nature09387]
- 51 **Gonzalez-Suarez E**, Jacob AP, Jones J, Miller R, Roudier-Meyer MP, Erwert R, Pinkas J, Branstetter D, Dougall WC. RANK ligand mediates progestin-induced mammary epithelial proliferation and carcinogenesis. *Nature* 2010; **468**: 103-107 [PMID: 20881963 DOI: 10.1038/nature09495]
- 52 **Quang CT**, Leboucher S, Passaro D, Fuhrmann L, Nourieh M, Vincent-Salomon A, Ghysdael J. The calcineurin/NFAT pathway is activated in diagnostic breast cancer cases and is essential to survival and metastasis of mammary cancer cells. *Cell Death Dis* 2015; **6**: e1658 [PMID: 25719243 DOI: 10.1038/cddis.2015.14]

P- Reviewer: Hegardt FG, Vlachostergios PJ **S- Editor:** Qiu S
L- Editor: A **E- Editor:** Lu YJ





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>

