**Name of journal:** *World Journal of Gastroenterology*

**ESPS Manuscript NO: 20131**

**Manuscript Type:** **MINIREVIEWS**

**Biomarkers of skin toxicity induced by anti-epidermal growth factor receptor antibody treatment in colorectal cancer**

Kubo A *et al.* Skin toxicity of anti-EGFR antibody

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**Author contributions:** Kubo A and Hashimoto H contributed equally to this work, generated the figure and wrote the manuscript; Takahashi N and Yamada Y contributed to the writing of the manuscript and designed the aim of the mini-review.

**Conflict-of-interest** **statement:** The authors have declared no conflicts of interest.

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**Received:** May 28, 2015

**Peer-review started:** June 1, 2015

**First decision:** September 29, 2015

**Revised:** October 20, 2015

**Accepted:** November 30, 2015

**Article in press:**

**Published online:**

**Abstract**

Skin toxicity is a common symptom of anti-epidermal growth factor receptor (EGFR) antibody treatment and is also a predictive marker of its efficacy in colorectal cancer patients. However, severe skin disorders induced by such antibodies negatively impact on the quality of life of patients and decreases drug compliance during treatment. If we can predict the high-risk group susceptible to severe skin toxicity before treatment, we can undertake the early management of any arising skin disorders and formulate a more accurate prognosis for anti-EGFR antibody treatment. Previous studies have identified molecular markers of skin toxicity induced by anti-EGFR antibody, such as *EGFR* polymorphisms, the expression of inflammatory chemokines and serum levels of EGFR ligands. A clinical trial was undertaken involving the escalation of cetuximab doses, guided by the grade of skin toxicity observed, such as no or low-grade, in metastatic colorectal cancer (the EVEREST study). The dose escalation of cetuximab was confirmed by a safety profile and had the tendency to achieve a higher response rate in *KRAS* wild-type patients. A large, prospective randomized trial is now ongoing (EVEREST 2) and the results of this trial may contribute to personalized medicine in *KRAS* wild-type colorectal cancer patients.

**Key words:** Colorectal cancer; Skin toxicity; Epidermal growth factor receptor; *EGFR* polymorphism; Ligand

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**Core tip:** Skin toxicity is a well-known biomarker used in the prognosis of anti- epidermal growth factor receptor (EGFR) antibody treatment of colorectal cancer patients. Previous retrospective studies indicated a change of the polymorphism of *EGFR* intron-1, chemokines and ligands were predictive markers of skin toxicity induced by anti-EGFR antibody. Such biomarkers used in predicting skin toxicity will enable the earlier management of skin toxicity as well as improve patients’ quality of life; however, further validations of prospective studies are needed.For patients with no/mild skin toxicity, a clinical trial of a dose escalation strategy is under evaluation and ongoing in the form of the EVEREST 2 study.

Kubo A, Hashimoto H, Takahashi N, Yamada Y. Biomarkers of skin toxicity induced by anti-epidermal growth factor receptor antibody treatment in colorectal cancer. *World J Gastroenterol* 2015; In press

**INTRODUCTION**

Colorectal cancer is one of the most common causes of death from cancer, in both men and women, around the world[1]. Owing to the development of diagnostic skills and chemotherapeutic drugs, prognoses concerning colorectal cancer patients have improved in the last decade. Although patients with early-stage colorectal cancer can undergo curative resection by endoscopy or surgery to achieve long survival after treatment, the 5-year survival rate of advanced colorectal cancer patients continues to be low because of a high rate of recurrence after surgical treatment. For the treatment of patients with metastatic or recurrent colorectal cancer, a variety of agents, including anti-vascular endothelial growth factor (VEGF) antibody, anti-epithelial growth factor receptor (EGFR) antibody, regorafenib and TAS-102 have recently been approved in Japan[2-7]. Unfortunately, most patients eventually acquire resistance to these drugs, leading to poor survival times.

Cetuximab (Erbitax®, Merck Serono) and panitummab (Vectibix®, Amgen) are anti-EGFR antibodies, which were initially approved for *KRAS* exon 2 wild-type patients with metastatic or recurrent colorectal cancer. Recently, genomic analyses of the EGFR downstream signal pathway, such as minor *KRAS* (exon 3 and 4)*, NRAS* (exon 3, 4 and 5), *BRAF* V600E and *PIK3CA* (exon 9, 20) were performed and it was found that these genomic alterations were associated with a poor prognosis in *KRAS* exon2 wild-type patients treated with anti-EGFR antibodies[8-10]. Retrospective analyses of several prospective trials indicated that the *RAS* mutation, which consists of *KRAS* (exon 2, 3, 4) and *NRAS* (exon 2, 3, 4) mutations, is a newly predictive biomarker. The *BRAF* V600E mutation is also considered a prognostic factor in anti-EGFR antibody treatment of patients with metastatic colorectal cancer[11-13].

Besides the genomic mutations of the EGFR downstream pathway, several studies have indicated that the grade of skin toxicity is a biomarker for predicting the efficacy of anti-EGFR antibody treatment for several cancers [14-16]. Skin toxicity is a typical side effect of anti-EGFR antibodies and causes various types of cutaneous changes, such as acneiform eruptions, dry skin and paronychia, during treatment. Although severe skin toxicity is associated with a better response to anti-EGFR antibodies, it negatively affects the quality of life (QOL) of patients and decreases drug compliance. Prophylaxis for skin toxicity, such as moisturizers, sunscreen, topical steroids, and oral doxycycline, is known to decrease the frequency of cutaneous disorders due to anti-EGFR antibodies and to improve the QOL of patients[17]. Molecular biomarkers for predicting the subgroup that will have severe skin toxicity due to anti-EGFR antibodies before treatment have been investigated, but there are no established markers for use in clinical practice.

In this review, we describe previous findings concerning the mechanism of skin toxicity in EGFR inhibition, biomarkers of skin toxicity for anti-EGFR antibodies, and treatment approaches guided by the severity of skin toxicity of anti-EGFR antibodies in colorectal cancer.

**MECHANISM OF SKIN TOXICITY INDUCED BY EGFR INHIBITION**

EGFR inhibition induces various symptoms of skin disorders and an acneiform rash is commonly observed on the scalp and face, particularly the cheeks, nose, nasolabial folds, chin, perioral regions, and the forehead, within the first 2–4 wk of treatment[18,19]. The EGFR is normally expressed in proliferating keratinocytes in the basal and supra-basal layers of the epidermis, outer layers of the hair follicle, sebaceous and eccrine sweat glands. It is believed that the EGFR plays a significant role in several processes of skin homeostasis, such as the regulation of cell survival, keratinocyte proliferation, differentiation and migration, wound healing and carcinogenesis[20]. Inhibition of the EGFR leads to the impairment of epidermal thickness and barrier function, and to the abnormal proliferation and differentiation of cells that express the EGFR in the skin and follicular epithelium[21,22]. Intracellular signaling pathway and mechanism of EGFR inhibition are shown in Figure 1.

According to previous reports, several ligands of EGFR as well as chemokines play significant roles in the skin inflammatory reaction caused by EGFR inhibition. Keratinocytes in cutaneous tissues are a rich source of EGFR ligands, such as transforming growth factor-alpha (TGF-alpha), epiregulin (EREG), amphiregulin (AREG) and heparin-binding EGF (HB-EGF)[23]. These ligands are known to stimulate the EGFR and maintain the cutaneous cell cycle of wound healing and EGFR-driven inflammatory reactions in keratinocytes[24]. EREG is known as an autocrine growth factor in normal human keratinocytes, and organizes the epidermal structure by regulating keratinocyte proliferation and differentiation[25]. The expression of AREG is developmentally regulated in the epithelium and mesenchyme of human skin during morphogenesis[26]. In addition, the EGFR is also activated by hepatocyte growth factor (HGF) stimulation in human epidermal keratinocytes. Transactivation of the EGFR by HGF-induced stimulation is associated with wound healing and mortality in epithelial cells[27,28]. EGFR inhibition causes an imbalance of cutaneous homeostasis, which is controlled by several ligands, and a skin inflammatory reaction induced by several chemokines.

An EGFR inhibitor decreases the activity of EGFR downstream signals and induces the expression of several chemokines, which enhance the skin’s inflammatory and immune response[29,30]. Previous reports indicated that the inhibition of EGFR induces tumor necrosis factor-alpha (TNF-alpha) and interleukin-1 (IL-1) during the development of skin rash in mice[31]. These chemokines induce IL-8 secretion by fibroblasts and keratinocytes, which activate neutrophil migration in cutaneous tissues[32-34]. Bangsgaard *et al*[35] revealed that the neutralization of IL-8 by HuMab-10F8 prevented the skin toxicity induced by an EGFR inhibitor. In addition, EGFR blockage is known to increase CC-chemokine ligand 2 (CCL2), CCL5, and C-X-C motif chemokine 10 (CXCL10), and reduce CXCL8 expression in keratinocytes[36]. Recently, Paul *et al*[37] reported that increased CCL2, CCL5 and decreased IL-8 or CXCL8 expression was observed in keratinocytes treated by EGFR inhibitor. In patients treated with EGFR inhibitor, a low level of serum CXCL8, corresponding to stronger EGFR inhibition, was associated with a higher grade of skin toxicity.

**PREDICTIVE MARKERS OF SKIN TOXICITY INDUCED BY ANTI-EGFR ANTIBODY IN ADVANCED COLORECTAL CANCER PATIENTS**

Most patients treated with anti-EGFR antibodies have an increased risk of skin toxicity. If we could predict the development of skin toxicities before the initiation of anti-EGFR antibody treatment, we could manage skin toxicity early and improve the patient’s QOL. Previous reports which evaluated the predictive factors of skin toxicity of anti-EGFR antibody treatment in patients with colorectal cancer are summarized in Table 1.

A North Central Cancer Treatment Group (NCCTG) N0147 trial, which was a randomized phase III trial of oxaliplatin plus 5-fluorouracil/leucovorin (FOLFOX6), with or without cetuximab, after curative resection of stage III colon cancer, investigated the risk factors for severe rash (≥ grade 3) in 933 patients[38,39]. More men (odds rate 2.12, *P =* 0.017) and younger patients (< 70-year-old; odds rate 0.21, *P =* 0.032) developed a severe rash compared with women and older patients.

Pharmacogenomic analyses of *EGFR* polymorphisms and several genomic mutations have been undertaken to determine their predictive value in the development of skin toxicity after anti-EGFR antibody treatment. A previous study indicated that a polymorphism of the *EGFR* intron-1 (CA single sequence repeat; short [S]/long [L] variant) was associated with the severity of skin toxicity (grade 0–1 *vs* 2–3) in colorectal cancer patients treated with irinotecan plus cetuximab as second-line chemotherapy[40]. *EGFR* intron-1 S/S carriers showed significantly more frequent grade 2–3 skin toxicity (*P =* 0.001) and a treatment response (*P =* 0.008) than *EGFR* intron-1 L/L carriers. The *EGFR* intron-1 S/S genotype was also associated with better survival in patients treated with cetuximab. Other polymorphisms of *EGF* and *EGFR,* such as *EGF* 61A>G, *EGFR* 216G>Tand *EGFR* 497G>A were not associated with the severity of skin toxicity after cetuximab treatment in this study. On the other hand, prognostic analyses of a randomized phase II trial of first-line chemotherapy with cetuximab in the AIO CRC Study Group indicated that the LL variant of the CA repeat in the *EGFR* intron-1 had a significant tendency to predict the severity of skin toxicity compared with the SS variant (*P* = 0.07)[41]. A heterogeneity in results was apparent in terms of polymorphisms of the CA repeat in the *EGFR* intron-1 because these studies were relatively small-sized and the optimized cut-off values for the CA repeat were not determined in colorectal cancer. Further validation by other translational analyses in a large prospective study is required to solve the heterogeneity of biomarkers.

As well as the above, a small scale analysis was conducted that measured the mRNA expression of cyclo-oxygenase 2 (Cox-2), cyclin D1 (CCND1), IL-8 and VEGF,and evaluated their predictive role in skin toxicity by cetuximab in patients of a phase II open-label multicenter study (IMCL-0144). Of these genes, the low expression of *Cox-2* was associated with a high grade of skin toxicity[42]. There was no association between the expression level of other genes and the severity of skin toxicity.

EGFR ligands have been considered significant modulators of cutaneous homeostasis and the inflammatory reaction in cutaneous tissues according to previous reports. As alterative markers of skin toxicity, we focused on the serum levels of ligands, which were associated with the EGFR signaling pathway. We evaluated the relationship between serum levels of EGFR ligands and the severity of skin toxicity, and the prognostic roles of these serum ligands in metastatic colorectal cancer patients who received anti-EGFR antibodies[43,44]. Our study indicated that low pre-treatment levels of serum AREG, EREG and HGF were associated with severe skin toxicity induced by anti-EGFR antibodies and a better prognosis in *KRAS* wild-type patients with metastatic colorectal cancer.

Molecular markers predicting skin toxicity, such as the CA repeat variant of the *EGFR* intron-1 and serum levels of particular ligands, are associated with the efficacy of the anti-EGFR antibody. Unfortunately, these potent findings were evaluated in small-scale studies and have not been validated as yet by other research studies; therefore the role of these biomarkers for clinical use is, at the moment, inconclusive.

**DOSE ESCALATING STRATEGY OF CETUXIMAB GUIDED BY THE SEVERITY OF SKIN TOXICITY IN METASTATIC COLORECTAL CANCER PATIENTS WITHOUT HIGH-GRADE SKIN TOXICITY**

Several prospective studies of colorectal cancer have described the severity and frequency of acneiform skin toxicity as a significant signature of the activity of anti-EGFR antibody, and are summarized in Tables 2 and 3[4,41,45-48]. These reports on the treatment of colorectal cancer with anti-EGFR antibody indicated that severe skin toxicity was associated with a higher response to antibody and a better prognosis, compared with no or mild skin toxicity. For patients with genetic alterations in the EGFR downstream pathway such as a *RAS* mutation, dose escalation of cetuximab for those without a high-grade skin reaction may be inefficient because of the permanent activation of the EGFR downstream pathway by such genetic alterations. On the other hand, in patients without genetic alterations of the EGFR downstream pathway, a dose escalation of anti-EGFR antibody may be effective and improve patient survival, especially for patients with tumor tissues not saturated with anti-EGFR antibodies. Skin toxicity may be a significant signature that affects the saturation of the anti-EGFR antibody in tumor tissues.

Two prospective studies have evaluated the dose escalation of anti-EGFR antibodies in colorectal cancer[49,50]. Fora *et al*[49] reported on a phase II trial of high-dose cetuximab (500 mg/m2) plus irinotecan in *KRAS* wild-type patients after progression of standard-dose cetuximab plus irinotecan. This study was small-scale, but nine out of twenty patients who received high-dose cetuximab plus irinotecan achieved disease control lasting more than 12 wk; the toxicity profile was also tolerable, except for grade 3/4 hypomagnesemia (25%). The efficacy of dose escalation of cetuximab after progression of a standard dose of cetuximab was limited and further selection of patients, such as those with low-grade skin toxicity or a response to a previous standard dose of cetuximab, may have been necessary in this study.

The EVEREST study was a prospective randomized study to evaluate the efficacy of dose escalation of cetuximab, compared with the 250 mg/m2 per week standard regimen, in patients who had developed no or a mild skin reaction in the first three weeks[50]. This study revealed that a dose escalation of cetuximab up to 500 mg/m2 achieved a safety profile comparable to a standard dose of cetuximab (250 mg/m2). Among *KRAS* mutant-type patients, there was no efficacy such as an increased objective response rate (ORR) and overall survival (OS) by the dose escalation of cetuximab. Among *KRAS* wild-type patients, the results of an ORR, for this subgroup, of a dose escalation of cetuximab were better than that for the subgroup with a standard dose of cetuximab (ORR: 43% *vs* 30%). However, the OS was similar between the two subgroups. The reasons for these results were considered, such as the small sample size of each group and the effects of negative genomic biomarkers such as *NRAS*, *BRAF* and other mutations. Data was not presented in this study on the difference in the change of severity of skin toxicity after dose escalation of cetuximab in *KRAS* wild-type and mutant-type patients.

Another EVEREST study has not drawn a conclusion on the efficacy of a dose escalation strategy for anti-EGFR antibody in a large-scale phase II trial (EVEREST 2), which was designed to compare the efficacy of dose escalation of a cetuximab plus 5-fluorouracil/leucovorin/irinotecan (FOLFIRI) regimen, as first-line chemotherapy, with a standard dose of FOLFIRI plus cetuximab in *KRAS* wild-type patients with metastatic colorectal cancer; this study is ongoing (NCT01251536) and the schema of this trial is shown in Figure 2. If such a dose escalation of anti-EGFR antibody achieves favorable results, a therapeutic decision on anti-EGFR antibody treatment for *RAS* wild-type patients with a low-grade skin toxicity may change dramatically and contribute to personalized treatment for this target population in future.

**CONCLUSION**

Although several studies have investigated the predictive markers of skin toxicity induced by anti-EGFR antibody in colorectal cancer patients, potent markers such as polymorphisms of *EREG* intron1, the expression of Cox-2 and serum levels of several ligands, such as HGF, EREG and AREG, have already been identified. Further validation of these biomarkers by other, larger studies, and translational research of novel markers to predict the skin toxicity of anti-EGFR antibody, are presently required.

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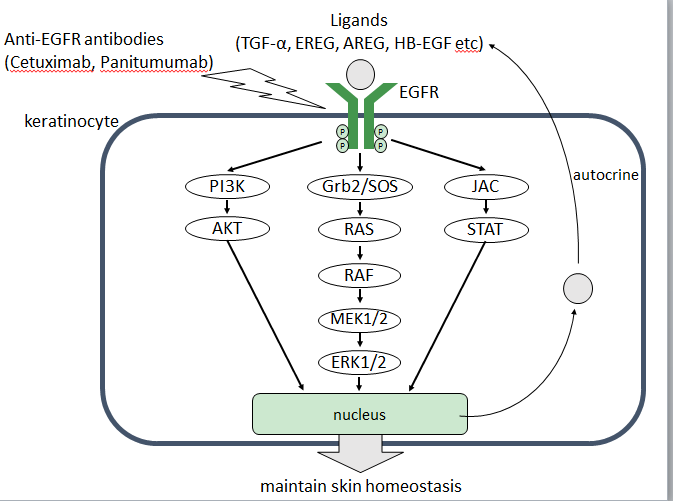
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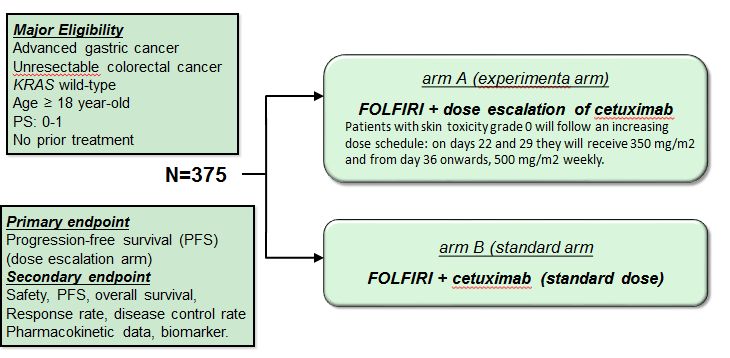
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**P-Reviewer:** Ju JF **S-Editor:** Yu J **L-Editor:** **E-Editor:**



**Figure 1 Homeostasis and epidermal growth factor receptor downstream signaling pathway in keratinocyte.** EGFR: epidermal growth factor receptor; TGF-α: Transforming growth factor-alpha; AREG: Amphiregulin; HB-EGF: Heparin-binding epithelial growth factor.



**Figure 2 Schema of EVEREST II trial (a non-randomized phase II study).**

**Table 1 Previous studies which evaluated the predictive biomarker of severity of skin toxicity induced by anti-** **epidermal growth factor receptor antibody treatment in colorectal cancer**

|  |  |  |  |
| --- | --- | --- | --- |
| **Ref.** | **Methods** | **Predictive markers** | **Risk factors of severe skin toxicity** |
| Jatoi *et al*[39] (2009) | Patient's backgrounds | Age  Gender | Younger  Men |
| Graziano *et al*[40] (2008) | *EGFR* polymorphism | CA repeat in EGFR intron-1 | EGFR intron-1 S/S variant |
| Vallböhmer *et al*[42] (2005) | mRNA expression | Cox-2 | Low expression |
| Takahashi *et al*[43,44] (2014, 2015) | Serum levels of ligands | AREG, EREG, HGF | Low levels of ligands at pre-treatment |

EGFR: Epidermal growth factor receptor; Cox-2: Cyclo-oxygenase 2, AREG: Amphilegulin; EREG: Epiregulin; HGF: Hepatocyte growth factor.

**Table 2 Skin toxicity as a biomarker of predicting the efficacy of anti-epidermal growth factor receptor antibody treatment in prospective phase II-III trial of unresectable colorectal cancer - Objective response rate**

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
|  | **Number** | **Treatment** | **Clinical trial** |  | **Objective response rate** | |
| **Ref.** |  |  |  |  | **Grade of skin toxicity** | ***P* value** |
| [Jonker *et al[*45]](http://www.ncbi.nlm.nih.gov/pubmed?term=Metzger%20R%5BAuthor%5D&cauthor=true&cauthor_uid=9440758) (2007) | 283 | Cetuximab | Phase III |  | ― | ― |
| [Van Cutsem *et al*[46] (2007)](http://www.ncbi.nlm.nih.gov/pubmed?term=Napieralski%20R%5BAuthor%5D&cauthor=true&cauthor_uid=15837757) | 200 | Panitumumab + BSC | Phase III |  | 14% *vs* 86% (grade 1 *vs* grade 2-3) | ― |
| Cunningham *et al*[4] (2004) | 218 | IRI + cetuximab | Phase III |  | 6.3% *vs* 25.8% (grade 0 *vs* any grade) | 0.005 |
| 111 | Cetuximab |  | 0% *vs* 13.0% (grade 0 *vs* any grade) |
| Sobrero *et al*[47] (2008) | 648 | IRI+cetuximab (2nd-line) | Phase III |  | ― | ― |
| Bokemeyer *et al*[48] (2009) | 169 | FOLFOX+cetuximab (1st-line) | Phase II |  | 13.0 *vs* 43.2% *vs* 53.2% *vs* 66.7% (grade 0 *vs* 1 *vs* 2 *vs* 3) | ― |
| Stintzing *et al*[41] (2013) | 149 | CAPIRI/CapOX+cetuximab (1st-line) | Phase II |  | 41% *vs* 62% (grade 0-1 *vs* grade 2-3) | 0.021 |

HR: Hazard ratio; CI: Confidence interval; IRI: Irinotecan; ND: Not described; FOLFOX: 5-fluorouracil/leucovorin/oxaliplati; CAPIRI: Capecitabine/ irinotecan; CapeOX: Capecitabine/oxaliplatin; BSC: Best supportive care.

**Table 3 Skin toxicity as a biomarker of predicting the efficacy of anti-epidermal growth factor receptor antibody treatment in prospective phase II-III trial of unresectable colorectal cancer - Overall survival**

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  |  |  |  |  | **Overall survival** | | | |
|  | **Number** | **Treatment** | **Clinical trial** |  | **Grade of skin toxicity (mo)** | | ***P* value** | **HR** |
| [Jonker *et al[*45]](http://www.ncbi.nlm.nih.gov/pubmed?term=Metzger%20R%5BAuthor%5D&cauthor=true&cauthor_uid=9440758) (2007) | 283 | Cetuximab | Phase III |  | 2.6 *vs* 4.8 *vs* 8.4 (grade 0 *vs* grade 1 *vs* ≥ grade 2) | | < 0.001 | ― |
| [Van Cutsem *et al*[46] (2007)](http://www.ncbi.nlm.nih.gov/pubmed?term=Napieralski%20R%5BAuthor%5D&cauthor=true&cauthor_uid=15837757) | 200 | Panitumumab+BSC | Phase III |  | ND | | ― | 0.59 (95%CI: 0.42-0.85) |
| Cunningham *et al*[4] (2004)  Sobrero *et al*[47] (2008) | 218 | IRI+cetuximab | Phase III  Phase III |  | 3.0 *vs* 9.1 (grade 0 *vs* any grade) | | ― | ― |
| 111 | Cetuximab |  | 2.5 *vs* 8.1 (grade 0 *vs* any grade) | | ― | ― |
|  | 648 | IRI+cetuximab (2nd-line) |  |  | 5.8 *vs* 11.7 *vs* 15.6 (grade 0 *vs* grade 1-2 *vs* ≥ grade 3) | | ― | ― |
| Bokemeyer *et al*[48] (2009) | 169 | FOLFOX+cetuximab (first-line) | Phase II |  | ― | ― | ― | ― |
| Stintzing *et al*[41] (2013) | 149 | CAPIRI/CapOX+cetuximab (first-line) | Phase II |  | 18.0 *vs* 30.3 (grade 0-1 *vs* grade 2-3) | | 0.161 | 0.75 (95%CI: 0.50-1.12) |

HR: Hazard ratio; CI: Confidence interval; IRI: Irinotecan; ND: Not described; FOLFOX: 5-fluorouracil/leucovorin/oxaliplati; BSC: Best supportive care.