

## 2016 Pancreatic Cancer: Global view

# Molecular targets for the treatment of pancreatic cancer: Clinical and experimental studies

Tasuku Matsuoka, Masakazu Yashiro

Tasuku Matsuoka, Masakazu Yashiro, Department of Surgical Oncology, Osaka City University Graduate School of Medicine, Osaka 545-8585, Japan

Masakazu Yashiro, Oncology Institute of Geriatrics and Medical Science, Osaka City University Graduate School of Medicine, Osaka 545-8585, Japan

**Author contributions:** Matsuoka T and Yashiro M designed this review; Matsuoka T wrote the manuscript; and Yashiro M edited the manuscript.

**Supported by** (in part) Grant-in-Aid for Scientific Research, No. 23390329.

**Conflict-of-interest statement:** There are not any financial or other interests with regard to the submitted manuscript that might be construed as a conflict of interest.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Correspondence to:** Masakazu Yashiro, MD, Department of Surgical Oncology, Osaka City University Graduate School of Medicine, 1-4-3 Asahi-machi, Abeno-ku, Osaka 545-8585, Japan. [m9312510@med.osaka-cu.ac.jp](mailto:m9312510@med.osaka-cu.ac.jp)  
Telephone: +81-6-66453838  
Fax: +81-6-66466450

Received: June 1, 2015

Peer-review started: June 3, 2015

First decision: July 20, 2015

Revised: August 13, 2015

Accepted: September 28, 2015

Article in press: September 30, 2015

Published online: January 14, 2016

## Abstract

Pancreatic cancer is the fourth most common cause of cancer deaths worldwide. Although recent therapeutic developments for patients with pancreatic cancer have provided survival benefits, the outcomes for patients with pancreatic cancer remain unsatisfactory. Molecularly targeted cancer therapy has advanced in the past decade with the use of a number of pathways as candidates of therapeutic targets. This review summarizes the molecular features of this refractory disease while focusing on the recent clinical and experimental findings on pancreatic cancer. It also discusses the data supporting current standard clinical outcomes, and offers conclusions that may improve the management of pancreatic cancer in the future.

**Key words:** Pancreatic cancer; Targeted therapy; Tyrosine kinase inhibitors; Microenvironment; Cancer stem cell

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** Pancreatic cancer-related mortality is almost consistently caused by local recurrence and metastasis. The survival of patients after surgical resection remains poor, and the results of adjuvant chemotherapy and radiotherapy are still unsatisfactory. Therefore, new treatments are urgently needed. Recent developments in our knowledge of the underlying biological features of pancreatic cancer may be useful in establishing molecularly targeted therapy as a new strategy, similar to those used to treat other types of malignancies.

Matsuoka T, Yashiro M. Molecular targets for the treatment of pancreatic cancer: Clinical and experimental studies. *World J Gastroenterol* 2016; 22(2): 776-789 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i2/776.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i2.776>

## INTRODUCTION

Pancreatic cancer is the fourth most common cause of cancer deaths, resulting in 330000 deaths per year worldwide<sup>[1]</sup>. Many patients with pancreatic cancer are diagnosed at advanced incurable stages because of the absence of screening. Although advances in a variety of approaches have improved the management of pancreatic cancer, the 5-year survival rate remains lower than 5%<sup>[1]</sup>. Surgical resection is currently the only potentially curative treatment. However, even after resection, the 5-year survival rate is less than 20% due to the high frequency of distal metastasis and local recurrence<sup>[2]</sup>.

The treatment of inoperable pancreatic cancer has traditionally involved the use of gemcitabine with low response rates and a marginal survival benefit. The failure of clinical treatment in patients with pancreatic cancer is often due to the heterogeneous nature of the disease. This type of tumor involves not only cancer cells, but stellate cells and stroma, which were known as microenvironment. Stromal proliferation and reduced angiogenesis have been shown to contribute to therapeutic resistance despite the efficacy in experimental studies utilizing cell lines or animal models. A recent European study found that the combination chemotherapy of FOLFIRINOX and gemcitabine is more effective than the use of gemcitabine alone. Albumin-bound paclitaxel (nab-paclitaxel), which was approved by the FDA in 2013, can also be used in conjunction with gemcitabine to treat pancreatic cancer<sup>[3]</sup>. However, in most patients with advanced stages of the disease, these treatments only prolong survival by a few months, while combination therapy can also lead to significantly increased toxicity<sup>[4]</sup>. The development of effective pancreatic cancer treatments is urgently needed to overcome these obstacles.

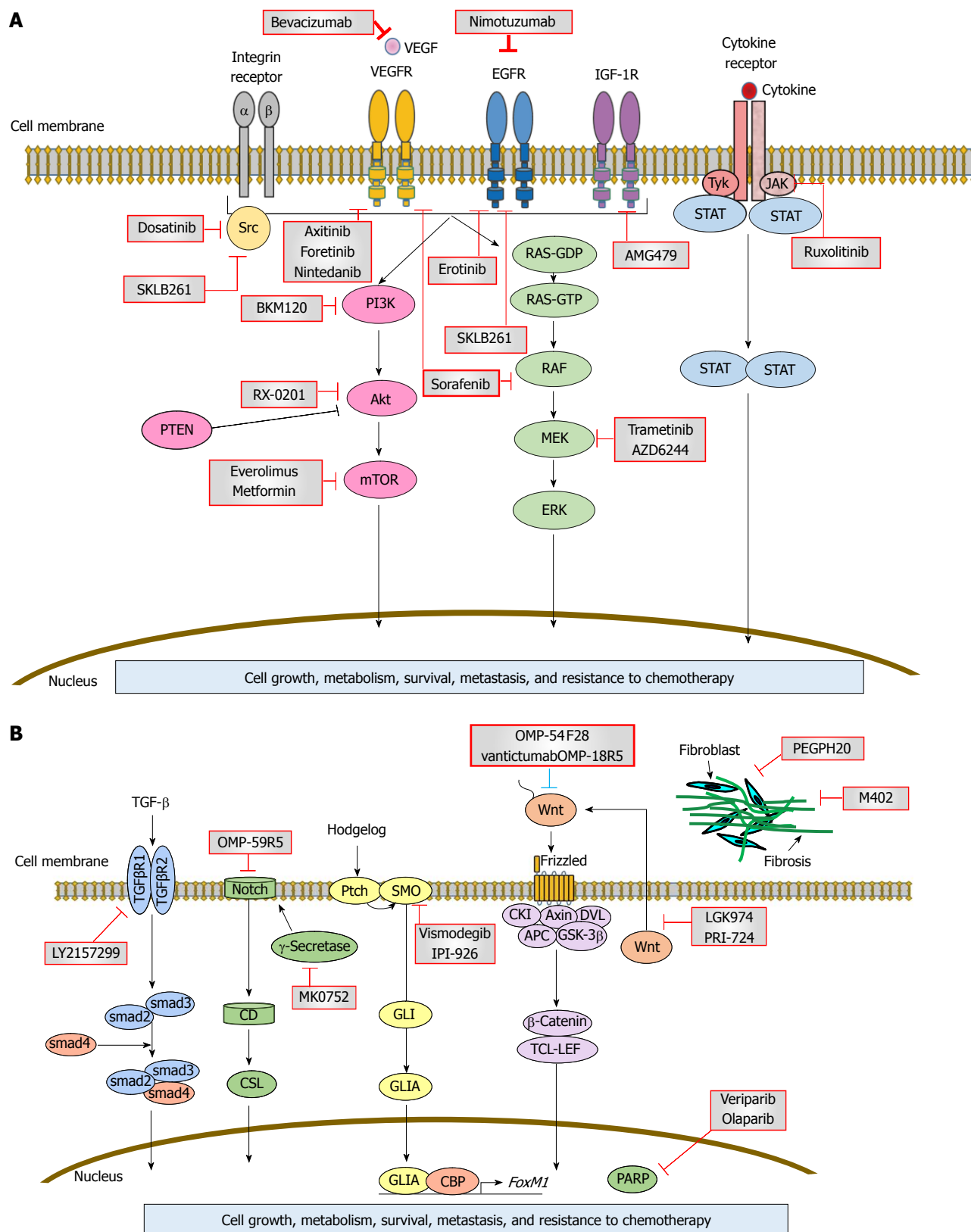
To date, knowledge of the molecular basis of tumor initiation has led to the use of various kinds of targeting agents to produce better prognoses for some types of solid tumors. These agents, including those targeting the angiogenesis pathways, the epidermal growth factor receptor (EGFR), the mitogen-activated ERK kinase (MEK), the fibroblast growth factor receptor (FGFR), the phosphatidylinositol-3 kinase/protein kinase-B/mammalian target of rapamycin (PI3K/mTOR), and the cancer stem cell compartment, may lead to significant advancements in pancreatic cancer treatment. In this study, we will review the current clinical and experimental results regarding molecular targets for the treatment of pancreatic cancer, and discuss potential future treatments.

## GENE ALTERATION AND MOLECULAR PATHOLOGY OF PANCREATIC CANCER

Recent studies have shown that pancreatic cancers

include an average of 63 genetic alterations<sup>[5]</sup>. Therefore, in order to develop effective treatments for pancreatic cancer, the complicated gene alterations and pathological features of this tumor type need to be elucidated. The molecular analysis of pancreatic cancer has often shown the involvement of known cancer genes and traditional cancer signaling pathways. The *KRAS* gene, which encodes a small GTPase that regulates the downstream signaling of growth factor receptors, is a known mutated oncogene found in most pancreatic cancers at advanced stages<sup>[6]</sup>. Missense mutations in the *KRAS* cluster have been found in specific hotspots (most generally codon 12)<sup>[7]</sup>. Recent studies have demonstrated that *KRAS* mutations are one of the earliest genetic events seen in human pancreatic intraepithelial neoplasia (PanIN) progression<sup>[8,9]</sup>. In addition to *KRAS* mutations, alterations in tumor suppressor genes such as *INK4A*, *BRCA2*, and *LKB1* occur frequently in pancreatic cancer. The tumor suppressor gene, *P16/CDKN2A*, which encodes a critical cell cycle regulator, is inactivated in > 90% of pancreatic cancer<sup>[10]</sup>. Mutation of the *p53* gene is closely associated with cellular responses to cytotoxic stress by contributing to both cell cycle arrest and cell apoptosis<sup>[11]</sup>. Mutations in *p53* are also common in pancreatic cancer, as these have been reported in approximately 75% of patients and most frequently characterized by small intragenic mutations coupled with a loss of heterozygosity<sup>[5]</sup>. The missense mutation of *SMAD4*, a tumor suppressor gene that encodes the transforming growth factor beta (TGF $\beta$ ) signaling pathway, is found in approximately 55% of patients with pancreatic cancer<sup>[12]</sup>. Mutations in *SMAD4* are associated with a poorer prognosis and widespread metastases, which suggests potential clinical implications<sup>[13]</sup>. The mismatch repair gene, *MLH1*, and the cationic trypsinogen gene, *PRSS1*, are also often mutated in pancreatic cancer<sup>[14]</sup>. Based on the typical age of onset related to the aforementioned mutations, these genetic lesions are thought to impact malignant progression rather than cancer initiation.

Some pancreatic cancers harbor activating mutations of *BRAF* rather than *KRAS*<sup>[15]</sup>. *BRAF* encodes RAF, a serine/threonine kinase belonging to a family of MEK (Figure 1A). MEK activates ERK, which forms the MAPK signaling pathway. Thus, active mutations of *KRAS* and *BRAF* eventually result in triggering the MAPK signaling, which is critical for the development of pancreatic cancer. Activation of the MAPK pathway is found not only in benign lesions, but also in late-stage pancreatic cancer<sup>[16]</sup>. Overexpressed MAPK through a constitutively active form of RAF results in PanIN/pancreatic ductal adenocarcinoma formation; conversely, the silencing of MAPK signaling inhibits tumor initiation<sup>[17,18]</sup>. PI3K signaling is another important pathway that has been studied in great detail in pancreatic cancer along with the MAPK pathway. The PI3K signaling mediates cell growth



**Figure 1 Signaling cascade (A) and therapeutic inhibitor (B) in pancreatic cancer.** Black circles indicated the critical signaling for the development of pancreas cancer. Red squares indicated the molecularly targeted agents for the treatment of pancreatic cancer. Adapted from Matsuoka T *et al.*

and survival via several downstream substrates such as Akt, p70-S6K, and mTOR (Figure 1A). Similar to the MAPK pathway, the consistent activation of PI3K has been shown to be closely associated with

the carcinogenesis of pancreatic cancer<sup>[19]</sup>. The PI3K downstream effector, *Akt*, is amplified in 10%-20% of pancreatic cancers, providing genetic evidence to support the importance of this pathway in this

type of cancer<sup>[20]</sup>. Several growth factor receptors, including vascular endothelial growth factor (VEGF) and insulin-like growth factor 1 receptor (IGF1R), are aberrantly expressed in pancreatic cancer<sup>[21]</sup>. These pathways mediate the important genes involved in a variety of cellular functions such as growth, apoptosis, differentiation, and metastasis *via* these two pathways (Figure 1A).

## MOLECULARLY TARGETED AGENTS FOR PANCREATIC CANCER TREATMENT

Within the last decade, numerous targeted agents have been examined individually or in combination with cytotoxic agents for the treatment of pancreatic cancer. The growth stimulating signaling described above has been targeted by molecular therapies for many kinds of cancer. Taken together, a number of paracrine signaling pathways, such as Hedgehog, Wnt, Notch, and TGF $\beta$ , might also contribute to cancer stem cell signaling and tumorigenesis (Figure 1B)<sup>[22]</sup>. These characteristics of pancreatic cancer may contribute to the development of molecularly targeted therapies. Figure 1 schematically summarizes the current understanding of inhibitors in pancreatic cancer. Table 1 summarizes clinical trials using molecular targeting agents.

### Signaling pathway

**Angiogenesis pathway:** Angiogenesis is crucial for the growth of malignancies. Anti-angiogenic therapies have shown efficacy in renal cell carcinoma, colorectal cancer, lung cancer, glioblastoma, and ovarian epithelial cancers<sup>[23]</sup>. VEGF is one of the key factors of angiogenesis that promotes tumor growth and metastasis<sup>[24]</sup>. VEGF is overexpressed in over 90% of patients with pancreatic cancer<sup>[25]</sup>, thus providing justification for VEGF-targeted therapy for pancreatic cancer. In contrast, a randomized phase III trial (CALGB 80303) found no improvements in the survival of patients who were given a combination of bevacizumab, which is a monoclonal antibody to VEGF, and gemcitabine, compared to the results of those treated with gemcitabine and a placebo, despite promising outcomes in phase II<sup>[26]</sup>. Taken together, efforts to use targeted agents such as sorafenib and axitinib have been unfavorable<sup>[27-29]</sup>. A phase II trial using TL-118, a novel anti-angiogenic drug, combined with gemcitabine to treat metastatic pancreatic cancer (NCT01509911) is currently ongoing (Table 1). Foretinib was developed as an ATP-binding site competitor to inhibit receptor tyrosine kinases with reported activity against VEGFRs, RON, c-Met, c-KIT, FLT-3, and platelet-derived growth factor receptors (PDGFRs)<sup>[30]</sup>. Mounting evidence has indicated that foretinib targets multi-additional kinases, resulting in the growth inhibition of tumors. Since hepatocyte growth factor (HGF) and c-MET are frequently

overexpressed in pancreatic cancer<sup>[31]</sup>, targeting these pathways has attracted much attention. A recent study concluded that foretinib inhibits tumor growth, angiogenesis, and lymphangiogenesis in xenograft animals by inhibiting not only c-MET but VEGFR-2, VEGFR-3, and TIE-2 signaling as well. These results suggest that simultaneous inhibitory effects to reduce pancreatic tumor growth can be expected from multikinase inhibition<sup>[32]</sup>.

**EGFR pathway:** EGFR, a transmembrane tyrosine kinase receptor of the ErbB family, plays an important role in tumor cell behavior. Aberrant EGFR activity leads to receptor dimerization and subsequently activates downstream signals, including members of the RAS and PI3K/Akt/mTOR pathways<sup>[33]</sup>. Overexpression of this growth factor receptor is seen in over 90% of pancreatic cancers<sup>[34]</sup>. Sustained EGFR activation has been reported in pancreatic cancer cell lines, and EGFR inhibitors have been found to lead to decreased proliferation<sup>[35]</sup>. Thus, EGFR pathway activation seems to provide a rationale for EGFR-targeted inhibition strategies. However, clinical trials using the anti-EGFR and anti-ErbB2 antibodies have yielded negative results<sup>[36,37]</sup>. In contrast, a large randomized phase III in which patients with pancreatic cancer were assigned to receive gemcitabine with or without erlotinib<sup>[38]</sup> reported that the patients who received the combination treatment had a modest but statistically significant improvement in overall median survival (OS) ( $P = 0.038$ ) and progression-free survival ( $P = 0.004$ ). The data from a subset analysis of this trial failed to indicate whether the KRAS mutation status or EGFR was a predictive marker for the therapeutic response to erlotinib<sup>[39]</sup>. Even though the median OS was only prolonged by 2 wk, this trial is remarkable because it is the only one to have shown an improvement in survival outcomes with combination gemcitabine/erlotinib in metastatic pancreatic cancer. On the other hand, the oncogenic benefit of erlotinib should be balanced with its potential complications, some of which have been reported to be fatal<sup>[40]</sup>. Another EGFR monoclonal antibody, nimotuzumab, achieved survival benefits when added to gemcitabine (8.7 mo vs 6.1 mo) with tolerable toxicity in a recent phase II trial involving patients with locally advanced pancreatic cancer (*J Clin Oncol* 2013;31:abstr 4009). Clinical trials to evaluate the effects of nimotuzumab combined with gemcitabine are ongoing in patients with the RAS wild type of locally advanced or metastatic pancreatic cancer (NCT 02395016).

**IGF1R pathway:** IGF1R belongs to the insulin receptor family. IGF1R signaling is highly expressed in pancreatic cancer, and this activation leads to a signaling cascade that triggers pathways such as ERK and PI3K/Akt/mTOR. It also plays a role in cancer survival and proliferation through RAS-dependent and -independent pathways. Inhibition of IGF1R signaling



**Table 1** Current clinical trials for pancreatic cancer

Target molecule	ClinicalTrials.gov identifier	Sponsor	Agent	Treatment setting	Study phase	Comments
EGFR	NCT00561990	Oncoscience AG	Nimotuzumab	First line	II / III	GEM ± nimotuzumab
	NCT02395016	Biotech Pharmaceutical	Nimotuzumab	First line	III	GEM ± nimotuzumab
MEK	NCT01222689	National Cancer Institute	selumetinib	First line	II	Selumetinib + erlotinib
PI3K	NCT01571024	UNC Lineberger Comprehensive Cancer Center	BKM120	First line	I	BKM120 + mFOLFOX6
Akt	NCT01028495	Rexahn Pharmaceuticals	RX-0201	First line	II	RX0201 + GEM
mTOR	NCT00981162	Roswell Park Cancer Institute	Everolimus	Second line	I / II	Everolimus + soresfenib
Angiogenesis	NCT01509911	Tiltan Pharma Ltd	TL-118	First line	II	TL-118 + GEM
Src	FOLFOX-D, NCT01652976	University of Florida	Dasatinib	First line	II	5-Fluorouracil + leucovorin + oxaliplatin + dasatinib
Jak	NCT01423604	Incyte Corporation	Ruxolitinib	Second line	II	Ruxolitinib + capecitabine
	NCT01822756	Incyte Corporation	Ruxolitinib	First line	I	Ruxolitinib + gemcitabine or nab-paclitaxel
Notch	NCT01098344	Cancer Research UK	MK0752	First line	I	MK0752 + GEM
Hedgehog	NCT01130142	Infinity Pharmaceuticals, Inc.	IP1-926	First line	I / II	IP1-926 + GEM
Wnt	NCT01351103	Novartis Pharmaceutical	LGK974	First line	I	LGK974 alone
	NCT01302405	Prism Pharma Co., Ltd.	PRI-724	First line	I	PRI-724 alone
	NCT02050178	OncoMed Pharmaceuticals, Inc	OMP-54F28	First line	I	GEM + nab-paclitaxel + OMP-54F28
	NCT02005315	OncoMed Pharmaceuticals, Inc	Vantictumab (OMP-18R5)	First line	I	GEM + nab-paclitaxel + vantictumab
Stroma	Halo-109-202, NCT01839487	Halozyme Therapeutics	PEGPH20 (hyaluronidase)	First line	II	GEM + nab-paclitaxel ± PEGPH20
	S1313, NCT01959139	Southwest Oncology Group	PEGPH20	First line	I / II	FOLFIRINOX ± PEGPH20
PARP	NCT01585805	National Cancer Institute	Veriparib	First line	II	GEM + cisplatin ± veriparib
	NCT01296763	Sidney Kimmel Comprehensive Cancer Center	Olaparib	First line	I / II	Irinotecan + cisplatin + mitomycin C ± olaparib
Others	NCT01210911	Academisch Medisch Centrum	Metformin	First line	II	Erlotinib + metformin + GEM
	NCT01373164	Eli Lilly and Company	LY2157299 (TGF- $\beta$ inhibitor)	First line	II	LY2157299 + GEM
	NCT01621243	Momenta Pharmaceuticals, Inc	M402 (heparan sulfate)	First line	I / II	GEM + nab-paclitaxel ± M402
	NCT01783171	National Cancer Institute (NCI)	Dinaciclib	First line	I	Dinaciclib + MK-226

GEM: Gemcitabine.

enhances the cytotoxicity of gemcitabine in pancreatic cancer xenografts<sup>[41]</sup>. However, the IGF1R inhibitor, AMG-479, and the monoclonal antibody, cixutumumab, failed to provide any survival benefits in a previous study (NCT01231347) (*J Clin Oncol* 2012;30:abstr 198). Meanwhile, the use of IGF1R in conjunction with ErbB would appear to be a good strategy for overcoming the chemoresistance common in pancreatic cancer. A previous study demonstrated that the simultaneous blockade of IGF1R and EGFR/Her-2 synergistically inhibits the pancreatic tumor growth and completely abolishes the activation of IRS-1, Akt, and MAPK phosphorylation. These results suggest that the combined application of these two inhibitors averts the resistance associated with monotherapy<sup>[42]</sup>.

**RAS pathway:** The RAS/RAF/MEK/ERK (MAPK) pathway is activated by numerous growth signals via their receptors, including EGFR, and is crucial in mediating uncontrolled growth and survival<sup>[43]</sup>. As previously discussed, MAPK plays a crucial role in the development of pancreatic cancer. Although trametinib, a MEK inhibitor, is currently approved for the treatment of melanoma, this drug has failed to show survival

benefit when combined with gemcitabine in advanced pancreatic cancer (*J Clin Oncol* 2013;31:abstr 291). A phase II trial of another MEK inhibitor, AZD6244, in combination with erlotinib as a second line treatment of advanced pancreatic cancer is currently underway (NCT01222689).

**PI3K/Akt/mTOR pathway:** Upon activation by RAS or EGFR, PI3K activates Akt, which subsequently triggers multiple downstream targets such as mTOR, leading to the regulation of many essential cellular processes, including cell growth, metabolism, survival, metastasis, and resistance to chemotherapy<sup>[44]</sup>. The PI3K-Akt pathway is activated in 59% of patients with pancreatic cancer<sup>[45]</sup>. Deregulation of this pathway through absent or reduced expression of PTEN (phosphatase and tensin homolog, a natural antagonist of PI3K) is frequently found in pancreatic cancer<sup>[46]</sup>. A phase II trial is currently testing the combination of an Akt antisense oligonucleotide, RX-0201, with gemcitabine in metastatic pancreatic cancer (NCT01028495). A phase I study of BKM120, a pan-class 1A PI3K inhibitor, with mFOLFOX6 in patients with metastatic pancreatic cancer is also now

recruiting (NCT01571024). Everolimus, an mTOR inhibitor, has shown antitumor activities, including the inhibition of cell proliferation, apoptosis, and angiogenesis, and displayed synergistic effects when combined with other anticancer agents<sup>[47]</sup>. A phase II study was conducted to explore treatment activity of the combination of capecitabine with everolimus in patients with advanced pancreatic cancer. The results revealed a response rate (RR) of 6.5% and an OS of 8.9 mo, suggesting that this combination therapy might enhance the efficacy of capecitabine<sup>[48]</sup>. Sorafenib, a multikinase inhibitor targeting Raf-1, BRAf, VEGFR1, VEGFR2, VEGFR3, and PDGFR $\beta$ , has been confirmed to be efficacious against advanced hepatocellular carcinoma<sup>[49]</sup>. It acts by inhibiting Raf-1 and BRAf as well as the activities of VEGFR1, VEGFR2, VEGFR3, and PDGFR $\beta$ . A phase I / II combination trial of sorafenib and everolimus in advanced pancreatic cancer has been completed, but the results have yet to be released (NCT00981162). Metformin, an antidiabetic drug that has direct metabolic effects through the activation of adenosine monophosphate-activated protein kinases, can inhibit the mTOR pathway by activating the tumor suppressor gene, TSC2. A randomized phase II study exploring the activity and safety of erlotinib and metformin combined with gemcitabine in patients with metastatic pancreatic cancer is also currently underway (NCT01210911).

**Src pathway:** Src, a family of proto-oncogenic non-receptor protein tyrosine kinases, plays a pivotal role in regulating multiple signal transduction pathways *via* its interactions with a number of proteins, including receptor tyrosine kinases and G-protein coupled receptors. C-Src is frequently overexpressed and/or aberrantly activated in a number of malignancies including 70% of pancreatic cancers<sup>[50]</sup>. Dasatinib, a compound related to saracatinib, was examined in a phase II trial in patients with metastatic pancreatic cancer; however, encouraging results were not obtained. A phase II trial to explore the efficacy of dasatinib combined with 5-fluorouracil, leucovorin, and oxaliplatin against metastatic pancreatic cancer is currently recruiting subjects (FOLFOX-D, NCT01652976).

**JAK/STAT pathways:** Activation of the Janus kinase/signal transducer and transcription (JAK/STAT) pathway has been found in many human cancers<sup>[51]</sup>. JAKs are a family of cytoplasmic tyrosine kinases, comprised of four members-JAK1, JAK2, JAK3, and Tyk2. JAK activation occurs upon the binding of a ligand to cell surface receptors, which leads to the creation of sites for interaction with proteins that contain phosphotyrosine-binding Src homology 2 (SH2) domains. STATs, a family of downstream transcription factors for JAKs<sup>[52]</sup>, contain a tyrosine residue phosphorylated by JAKs, leading to nuclear translocation. In the nucleus, STATs serve as transcription factors that initiate the transcription

of downstream target genes. Abnormalities of the JAK/STAT pathway contribute directly to cellular transformation, increased cell proliferation, apoptosis, and angiogenesis. JAK mutations and STAT activation have been reported in pancreatic cancer<sup>[53,54]</sup>. In a randomized phase II study of capecitabine plus either ruxolitinib or placebo, patients with metastatic pancreatic cancer demonstrated an improvement in survival (NCT01423604). An early phase clinical trial of ruxolitinib and gemcitabine with or without nab-paclitaxel is also currently underway (NCT01822756).

### Cancer stem cells

A small population of pancreatic cancer stem cells (CSCs) has been suggested to be resistant to chemotherapy and radiation therapy. CSCs are believed to be responsible for tumor carcinogenesis, progression, and metastasis in cancers including the pancreatic type<sup>[55,56]</sup>. Hedgehog, Notch, and Wnt have been shown to play a pivotal role in the development of pancreatic cancer stem cells<sup>[57]</sup>. Remarkable progress in understanding the involvement of CSCs in pancreatic cancer might highlight these cells as attractive targets for therapy.

**Notch signaling:** Recent evidence has suggested that Notch signaling is implicated in tumor growth and survival as well as involved in the development and function of many organs<sup>[58]</sup>. This pathway is thought to sustain a pool of pancreatic progenitor cells at an early stage of pancreatic development, and regulates pancreatic ductal cell differentiation<sup>[59]</sup>. Notch ligands and receptors have been shown to be highly expressed in pancreatic cancer; they also promote epithelial-mesenchymal transition (EMT) by regulating several transcription factors such as Snail, Slug, and TGF $\beta$ <sup>[60]</sup>. A number of studies have shown that chemotherapy-resistant pancreatic CSCs are related to Notch signaling activation<sup>[61]</sup>. The ALPINE trial is a phase Ib study exploring the anti-Notch2/3 inhibitor, OMP-59R5, in combination with nab-paclitaxel and gemcitabine in patients with untreated metastatic pancreatic cancer (*J Clin Oncol* 2014;32:abstr). This study showed good tolerability and positive responses (partial response = 46%, durable complete response = 77%). One appealing target for blocking this pathway is the  $\gamma$ -secretase enzyme, which causes proteolytic cleavage and controls the release of the Notch intracellular domain as well as the production of its active form. Clinical trials are currently underway to investigate the effects of MK0752, a  $\gamma$ -secretase inhibitor, combined with gemcitabine in patients with advanced pancreatic cancer (NCT01098344).

**Hedgehog signaling:** The Hedgehog (HH) pathway regulates embryogenesis, which is undetectable in normal pancreatic tissue. HH binds to the extracellular receptor and the transcriptional target gene, Patched

(Ptch), the latter of which releases the Smoothed seven-transmembrane protein (SMO). This allows SMO to translocate to the cell surface and results in the activation of GLI transcription factors and the consequent induction of HH target genes, including GLI and Ptch1 (Figure 1B). The pathological behavior of HH signaling is well known, and an increased expression has been observed during pancreatic tumorigenesis<sup>[62]</sup>. It has been found that hedgehog signaling is phosphorylated in earlier pancreatic tumor lesions, and the expression of pathway substrates becomes elevated during the progression to an advanced stage<sup>[63]</sup>. HH signaling has also been shown to be closely associated with KRAS mutations, which drive the early stages of pancreatic neoplasia<sup>[64]</sup>. Interestingly, HH signaling in pancreatic cancer is localized to the stromal compartment, and the overexpression of sonic HH in the pancreas is sufficient to initiate precancerous lesions in transgenic mice, which contributes to maintaining the tumor microenvironment<sup>[65,66]</sup>. Interestingly, the attenuated action of sonic HH has resulted in improved gemcitabine delivery, the depletion of dense stroma, and an enhanced vascularization of the tumors in mouse models<sup>[67]</sup>, suggesting that this pathway could be an appealing target for drug development. GDC-0449, also known as vismodegib, a small-molecule SMO antagonist, inhibits the HH signaling pathway. A pilot study evaluating the effects of GDC-0449 in combination with gemcitabine was performed in patients with metastatic pancreatic adenocarcinoma; however, the joint use of GDC-0449 and gemcitabine was not found to be superior to the sole use of gemcitabine in the treatment of metastatic pancreatic cancer<sup>[68]</sup>. Similarly, a randomized phase II trial of gemcitabine with or without vismodegib in treating patients with recurrent or metastatic pancreatic cancer also yielded disappointing results (NCT01064622). A phase II randomized study evaluating IPI-926, a small molecule SMO antagonist, in combination with gemcitabine in metastatic pancreatic cancer patients was recently completed, and the publication of its results is highly anticipated (NCT01130142).

**Wnt signaling:** Evidence that the Wnt signaling pathway plays a pivotal role in the regulation of stem cells has been accumulating<sup>[69,70]</sup>. Recent studies of Wnt signaling have suggested its roles in tumor biology and the pathogenesis of pancreatic cancer. Based on findings, it is plausible that the dysregulation of Wnt signaling pathway is closely associated with pancreatic cancer chemoresistance<sup>[71]</sup>. Wnt signaling inhibitors such as LGK974 and PRI-724 are currently under investigation in active phase I clinical studies on advanced solid tumors including pancreatic cancers (NCT01302405 and NCT01351103, respectively). An open-label phase 1b dose-escalation study to elucidate the safety and tolerability of OMP-54F28 (NCT02050178) and Vantictumab/OMP-18R5

(NCT02005315) when combined with nab-paclitaxel and gemcitabine is currently in progress.

**TGFβ:** TGF-β is intimately involved in regulating numerous physiological processes, including cellular differentiation, homeostasis, and EMT in pancreatic cancer<sup>[72]</sup>. A phase II study of gemcitabine in combination with LY2157299, a specific type 1 receptor inhibitor of TGFβ, or a placebo is currently being implemented (NCT01373164).

### **Stromal environment**

One of the most important obstacles that impairs the effects of anticancer agents is the extracellular matrix (ECM) and stromal cells<sup>[73]</sup>. It has been reported that tumor-stroma interactions result in a complicated signaling network that leads to tumor progression in many kinds of solid cancers<sup>[74,75]</sup>. Pancreatic cancer shows abundant stroma in the tumor microenvironment<sup>[76]</sup>. Pancreatic cancer is uniquely characterized by a rich tumor stroma that might interfere with the delivery of agents to tumors and induce a complex interplay of intercellular signaling. Stromal depletion strategies such as the degradation of hyaluronic acid could potentially facilitate drug delivery to tumor sites<sup>[67]</sup>. Currently, there is a randomized phase II trial comparing the treatment effects of PEGPH20, a pegylated formulation of recombinant hyaluronidase, in combination with nab-paclitaxel and gemcitabine comparing the treatment effects of PEGPH20 with the treatment effects of nab-paclitaxel alone in metastatic pancreatic cancer (Halo-109-202, NCT01839487). A partially randomized phase I / II evaluation of modified FOLFIRINOX with or without PEGPH20 in patients with newly diagnosed metastatic pancreatic cancer is also now under testing (S1313, NCT01959139). Heparan sulfate proteoglycans (HSPGs) are complex polysaccharides that regulate several aspects of cancer biology, including tumorigenesis, tumor development, and metastasis<sup>[77]</sup>. HSPGs have been shown to be associated with the tumor microenvironment by binding to factors that support tumor proliferation. M402, a mimetic of heparan sulfate, blocks the multiple interactions associated with heparan sulfate<sup>[77]</sup>. A phase II clinical trial evaluating the effects of M402 in conjunction with standard chemotherapy is actively ongoing (NCT01621243).

### **Alternative poly (ADP-ribose) polymerase pathway**

Breaks in the DNA double-strand are generally repaired by homologous recombination, which is mediated by BRCA1 and BRCA2 proteins that sustain genomic stability and cell death. The poly (ADP-ribose) polymerase pathway takes on the main role in DNA repair when BRCA dysfunction occurs<sup>[78]</sup>. The PARP proteins undertake roles in a wide range of cellular functions including DNA transcription, DNA damage response, genomic stability maintenance,

and cell cycle regulation. Inhibitors of PARP enzymes result in synthetic lethality in cancers with DNA repair failure or homologous repair deficiency. They have also been shown to be clinically effective in cancers with poor DNA repair due to germ-line mutations in BRCA1 and BRCA2, which are estimated to be 5%-7% of patients with pancreatic cancer. A randomized phase II trial of cisplatin plus gemcitabine with or without veliparib, which is a selective PARP inhibitor, in locally advanced or metastatic pancreatic cancer patients is ongoing (NCT01585805). Another PARP inhibitor, Olaparib, at 100 mg b.i.d. with 600 mg/m<sup>2</sup> of gemcitabine is tolerated in patients with advanced pancreatic cancer<sup>[79]</sup>. A randomized multi-center phase I / II trial is currently investigating the effects of irinotecan, cisplatin, and mitomycin C combined with olaparib in patients with advanced pancreatic cancer (NCT01296763). Conversely, a randomized, double blind phase III study of olaparib monotherapy in patients with BRCA1/2 mutated metastatic pancreatic cancer who have not progressed from first line platinum-based chemotherapy is also now underway (NCT02184195).

## RECENT EXPERIMENTAL STUDIES ON THE TREATMENT OF PANCREATIC CANCER

Despite numerous clinical trials utilizing known targeted agents, the overall advancement made in pancreatic cancer treatment has been relatively modest in comparison to the advancement made in the treatment of other types of tumors. Therefore, the exploration of novel agents targeted at certain signaling pathways is one of the most important undertakings to improve the outcome of patients with lethal pancreatic cancer. In this section, we will focus on the recent experimental studies that may open the door to the development of novel and hopefully more effective strategies for the treatment of pancreatic cancer. A list of novel therapeutic targets and drugs is presented in Table 2.

### Novel tyrosine kinase inhibitor

There is growing evidence to suggest that novel tyrosine kinase inhibitors can target multiple different pathways and/or signaling processes that have never been seen before in pancreatic cancer. As previously explained, utilizing the multikinase inhibitor, foretinib, may provide a simultaneous inhibitory effect on pancreatic cancer<sup>[32]</sup>. SKLB261 is a multikinase inhibitor obtained recently through lead optimization with reported activity against EGFR, Src, and VEGFR2. The application of SKLB261 has resulted in more potent antitumor activities than that of dasatinib, gemcitabine, or erlotinib in pancreatic cancer xenografts as well as more prolonged survival in mice compared with

gemcitabine-treated groups<sup>[80]</sup>. Nintedanib, a triple angiokinase inhibitor that targets the VEGFR1/2/3, FGFR1/2/3, and PDGFR $\alpha/\beta$  pathways, has been shown to strongly inhibit the growth of pancreatic cancer cell lines in addition to enhancing the inhibitory effects of gemcitabine. Nintedanib also induces apoptosis in pancreatic cancer cells associated with stromal cells, providing a strong rationale for the clinical evaluation of nintedanib combined with conventional cytotoxic agents<sup>[81]</sup>.

Masitinib, a multi-targeted protein tyrosine kinase inhibitor with possible anticancer activity, selectively binds to and inhibits both the wild-type and mutated forms of the stem cell factor receptor (c-Kit; SCFR), PDGFR, and FGFR3. In the recently conducted randomized phase III trial, masitinib combined with gemcitabine prolonged the survival of patients in subgroups defined by an overexpression of acyl-CoA oxidase-1 (ACOX1) in secondary analyses. In the ACOX1 subgroup, the patients treated with masitinib plus gemcitabine showed a median OS of 11.7 mo (95%CI: 8.3-19.9) compared with a median OS of 5.6 mo (95%CI: 3.7-12.9) for the placebo plus gemcitabine treatment-arm<sup>[82]</sup>.

Hypoxia inducible factor-1 (HIF-1) is a principal mediator of cell adaption to hypoxia, and is extensively expressed in 88% of pancreatic tumors<sup>[83]</sup>. PX-478, an HIF-1 inhibitor, has been shown to promote the anti-cancer effects of gemcitabine, which are closely associated with immunogenic cell death in pancreatic cancer<sup>[84]</sup>. Recent studies have demonstrated that GSK3 $\beta$  is closely related to pancreatic cancer cell growth, providing a rationale for targeting GSK3 $\beta$  in the treatment of patients with pancreatic cancer<sup>[85]</sup>. GSK3 $\beta$  inhibition has been shown to induce apoptosis by a mechanism involving JNK-cJUN activation<sup>[86]</sup>. CXCR4 (CXC chemokine receptor type 4), a G-protein coupled receptor of CXCL12 (SDF-1), has been found to promote GSK3 $\beta$  expression and the invasion ability of pancreatic cancer cells by Akt signaling. This finding suggests that CXCR4 inhibition may open a new therapeutic avenue that will impact the capacity to effectively treat pancreatic cancer patients<sup>[87]</sup>. Forkhead box M1 (FOXM1) is a transcription factor in the FOX protein superfamily containing a conserved winged helix DNA-binding domain<sup>[88]</sup>. FOXM1 is an important transcription factor for many genes key to regulating a variety of processes in tumor pathogenesis, such as tumor cell survival, growth, and EMT. Several bodies of evidence have demonstrated that FOXM1 plays a crucial role in pancreatic cancer progression<sup>[89]</sup>. A recent paper asserted that FOXM1 contributes to the development of pancreatic cancer by enhancing the *uPAR* gene transcription and subsequently the EMT of cancer<sup>[90]</sup>. These findings suggest that the deregulation of FOXM1 signaling may be a new attractive strategy in the development of novel therapeutic targets to



**Table 2** Recent experimental studies of targeted therapy for pancreatic cancer

Targeted therapeutics	Ref.	Cell lines (cell type)	Main results
Multikinase inhibitors			
Foretinib	Chen <i>et al</i> <sup>[32]</sup>	Panc-1 (P)	Foretinib inhibited tumor growth, angiogenesis and lymphangiogenesis in xenograft animals, by inhibiting c-MET but VEGFR-2, VEGFR-3, and TIE-2 signaling
SKLB261	Pan <i>et al</i> <sup>[80]</sup>	BxPC-3 (P), Panc-1 (P), AsPC-1 (S), HPAC (P)	Application of SKLB261 resulted in more potent antitumor activities than dasatinib, gemcitabine, or erlotinib in pancreatic cancer xenografts
Nintedanib	Awasthi <i>et al</i> <sup>[81]</sup>	AsPC-1 (S), BxPC-3 (P), Panc-1 (P), MIA-PaCa-2 (P),	A triple angiokinase inhibitor, nintedanib inhibited growth of pancreatic cancer cell lines, with gemcitabine enhancing inhibitory effects
Dual inhibition			
Lapatinib and trametinib	Lindberg <i>et al</i> <sup>[101]</sup>	MAD 08-608, 08-738, 09-366 (P)	Dual anti-EGFR and anti-HER2 therapy significantly enhanced the growth inhibitory effects of the MEK1/2 inhibitor trametinib
ZSTK474 and RO5126766	Van Dort <i>et al</i> <sup>[99]</sup>	Panc-1 (P)	PI3K inhibitor and the Raf/MEK inhibitor RO5126766 resulted in high in vitro inhibition of both PI3K and MEK1 also decreased cellular viability in pancreatic cancer cell line
NVP-AEW541 and lapatinib	Urtasun <i>et al</i> <sup>[42]</sup>	NP-9, -18, -29 (P) CP15T, 15A (p)	Combined treatment with the IGF-1R and EGFR/Her-2 inhibitors synergistically inhibited pancreatic cancer cell growth which is associated with abolishment of Akt, Efk, and IRS-1 activity
Novel pathways			
PX-478 (HIF-a)	Zhao <i>et al</i> <sup>[84]</sup>	CFPAC-1 (S), BxPC-3 (P), Panc-1 (P), MIA-PaCa-2 (P)	Combined treatment with gemcitabine/PX-478 significantly enhanced the anti-tumor effect which is associated with immunogenic cell death
SB216763(GSK-3b)	Marchand <i>et al</i> <sup>[86]</sup>	Panc-1 (P), MIA-PaCa-2 (P), BxPC-3 (P)	Inhibition of GSK-3 $\beta$ induced apoptosis by mechanism involving
Micro RNA			
miR-142-3p	MacKenzie <i>et al</i> <sup>[95]</sup>	MIA-PaCa-2 (P), Capan-1(S), HEK-293 (P), S2-013 (P)	A unique HSP70 inhibiting compounds, miR-142-3p regulate triptolide-induced inhibition of pancreatic cancer growth
miR-146a	Li <i>et al</i> <sup>[96]</sup>	AsPC-1 (s), Panc-1 (P)	miR-146a takes significant roles in pancreatic cancer invasion and metastasis but lower expressed in pancreatic cancer compared with normal pancreatic tissue
miR-494	Li <i>et al</i> <sup>[97]</sup>	Colo357 (s), Panc-1 (P)	miR-494, identified to affect levels of FOXM1 in pancreatic cancer cell lines and act as a negative regulator of this transcriptional activator, blocked nuclear translocation $\beta$ -catenin

VEGF: Vascular endothelial growth factor; EGFR: Epidermal growth factor receptor; HER2: Human epidermal growth factor receptor 2; MEK: Mitogen-activated ERK kinase; PI3K: Phosphatidylinositol-3 kinase; IGF-1R: Insulin-like growth factor 1 receptor; FGFR: Fibroblast growth factor receptor; HSP: Heat shock protein; FOXM1: Forkhead box M1; P: Primary; S: Secondary.

control pancreatic cancer.

### Micro RNA

Recently discovered micro RNA (miRNA) are short non-coding RNAs involved in the negative regulation of miRNAs translation. It is important to note that miRNAs mediate a variety of cellular functions and their dysregulation is a crucial event in tumor initiation<sup>[91]</sup>. The expression of miRNAs in pancreatic cancer cells differs from those in normal pancreatic cells and in patients with chronic pancreatitis. Over 130 miRNAs have been proven to be deregulated in pancreatic cancer<sup>[92]</sup>. Several of these miRNAs play a role in new therapeutic prospects in the treatment of pancreatic cancer. It has been shown that nanomolar concentrations of antisense miR-21 and miR-221 oligonucleotides significantly repress their targets and reduce the growth of pancreatic cancer cell lines<sup>[93,94]</sup>. A unique HSP70 inhibiting compound, miR-142-3p, regulates the triptolide, a diterpene triepoxide extract from the Chinese herb *Tripterygium wilfordii*, -induced

inhibition of pancreatic cancer growth<sup>[95]</sup>. The miR-146a compound plays a significant role in pancreatic cancer invasion and metastasis<sup>[96]</sup>. Interestingly, miR-146a has been shown to restore the proteolytic activity of pancreatic cancer cells but with less expression in pancreatic cancer than in normal pancreatic tissue. Utilizing isoflavones or 3,3'-diinodolylmethane has been found to increase the expression of miR-146a, which may be a promising approach to blocking the invasion of pancreatic cancer. MicroRNA-494 (miR-494) is known to affect levels of FOXM1 in pancreatic cancer cell lines and act as a negative regulator of this transcriptional activator. It has also been shown to block nuclear translocation of  $\beta$ -catenin, which leads to cell proliferation, migration, and the increase of the sensitivity of pancreatic cancer cells to gemcitabine<sup>[97]</sup>.

### CONCLUSION

Pancreatic cancer illustrates genetic heterogeneity, and the complications in molecular signaling crosstalk

cause the failure of existing treatment strategies. Currently, only a few targeted agents, including erlotinib, have yielded a significant survival benefit for patients with pancreatic cancer. To overcome these obstacles, a considerable effort needs to be put into investigating the effective use of these therapies. Another challenge in evaluating novel targeted therapies in pancreatic cancer treatment is to identify underlying pathological features and incorporate them into trials with molecular biomarkers. Analysis of pancreatic cancer pathogenesis with its molecular characteristics may help to identify the biomarker-defined subsets of patients that can be targeted to optimize the therapeutic benefit<sup>[98]</sup>.

The identification of effective targeted combination therapies may be useful for generating enhanced strategies of treating pancreatic cancer. For instance, suppression of the PI3K/Akt/mTOR pathway may result in an escape *via* the MAPK pathway as well as in diminished effects due to the intensive crosstalk between these pathways. Therefore, a combination of agents that inhibit separate pathways may be crucial for achieving the desired efficacy against tumors. A recent study found that the application of a prototype dual-acting agent designed using the PI3K inhibitor, ZSTK474, and the Raf/MEK inhibitor, RO5126766, resulted in high *in vitro* inhibition of both PI3K and MEK1 as well as decreased cellular viability in pancreatic cancer cell lines<sup>[99]</sup>. In a previous study, the combined effect of the cyclin-dependent kinase inhibitor, dinaciclib, and the pan-Akt inhibitor, MK-2206, dramatically inhibited tumor growth and metastasis in eight pancreatic cancer models. Remarkably, several complete responses were induced by this combination treatment. These results suggest that blocking RAF in combination with other effector pathways downstream from KRAS may provide increased efficacy in pancreatic cancer treatment<sup>[100]</sup>. Notably, preclinical studies predicting the effects of combination therapies with EGFR and other pathway inhibitors in pancreatic cancer xenografts have presented promising results. A recent study asserted that dual anti-EGFR and anti-HER2 therapy significantly enhanced the growth inhibitory effects of the MEK1/2 inhibitor, trametinib, in different patient-derived pancreatic cancer xenografts. This highlights the importance of designing therapeutic interventions that target not only the EGFR-HER2 but also the KRAS pathways to achieve maximal therapeutic efficacy<sup>[101]</sup>.

Selecting drug combinations with novel agents that target not only tumor initiation but also the surrounding stroma may be one such approach. There is growing evidence to suggest that proteins expressed by stromal cells (Cox-2, stromal-derived factor, integrins, secreted protein acidic and rich in cysteine, and HH elements) are related to poor outcomes and the resistance to current therapies<sup>[102]</sup>. In the meantime, it is also essential to pay attention to the

toxicity resulting from the use of targeted agents with conventional chemotherapy or agents targeting other pathways. A thorough understanding of the underlying mechanisms involved in toxicity will be crucial to furthering the development of pancreatic cancer therapies.

Thus far, most clinical trials involving targeted therapies for pancreatic cancer have not yielded successful results. The lack of effectiveness of several targeted agents in pancreatic cancer, despite their success in treating other types of malignancies, suggests that pancreatic cancer has particularly challenging biological characteristics that have yet to be well defined. As discussed in this review, several experimental studies have demonstrated promising new pathogenesises and efficacy in the treatment of pancreatic cancer. Therefore, continuing to develop and investigate treatments that fight pancreatic cancer from as many angles as possible will likely increase the chances of achieving positive outcomes for patients.

## REFERENCES

- 1 Siegel R, Naishadham D, Jemal A. Cancer statistics, 2012. *CA Cancer J Clin* 2012; **62**: 10-29 [PMID: 22237781 DOI: 10.3322/caac.20138]
- 2 Oettle H, Post S, Neuhaus P, Gellert K, Langrehr J, Ridwelski K, Schramm H, Fahlke J, Zuelke C, Burkart C, Gutterer K, Kettner E, Schmalenberg H, Weigang-Koehler K, Bechstein WO, Niedergethmann M, Schmidt-Wolf I, Roll L, Doerken B, Riess H. Adjuvant chemotherapy with gemcitabine vs observation in patients undergoing curative-intent resection of pancreatic cancer: a randomized controlled trial. *JAMA* 2007; **297**: 267-277 [PMID: 17227978 DOI: 10.1001/jama.297.3.267]
- 3 Borazanci E, Von Hoff DD. Nab-paclitaxel and gemcitabine for the treatment of patients with metastatic pancreatic cancer. *Expert Rev Gastroenterol Hepatol* 2014; **8**: 739-747 [PMID: 24882381 DOI: 10.1586/17474124.2014.925799]
- 4 Thota R, Pauff JM, Berlin JD. Treatment of metastatic pancreatic adenocarcinoma: a review. *Oncology (Williston Park)* 2014; **28**: 70-74 [PMID: 24683721]
- 5 Jones S, Zhang X, Parsons DW, Lin JC, Leary RJ, Angenendt P, Mankoo P, Carter H, Kamiyama H, Jimeno A, Hong SM, Fu B, Lin MT, Calhoun ES, Kamiyama M, Walter K, Nikolskaya T, Nikolsky Y, Hartigan J, Smith DR, Hidalgo M, Leach SD, Klein AP, Jaffee EM, Goggins M, Maitra A, Jacobuzio-Donahue C, Eshleman JR, Kern SE, Hruban RH, Karchin R, Papadopoulos N, Parmigiani G, Vogelstein B, Velculescu VE, Kinzler KW. Core signaling pathways in human pancreatic cancers revealed by global genomic analyses. *Science* 2008; **321**: 1801-1806 [PMID: 18772397]
- 6 Rozenblum E, Schutte M, Goggins M, Hahn SA, Panzer S, Zahurak M, Goodman SN, Sohn TA, Hruban RH, Yeo CJ, Kern SE. Tumor-suppressive pathways in pancreatic carcinoma. *Cancer Res* 1997; **57**: 1731-1734 [PMID: 9135016]
- 7 Yashiro M, Carethers JM, Laghi L, Saito K, Slezak P, Jaramillo E, Rubio C, Koizumi K, Hirakawa K, Boland CR. Genetic pathways in the evolution of morphologically distinct colorectal neoplasms. *Cancer Res* 2001; **61**: 2676-2683 [PMID: 11289147]
- 8 Moskaluk CA, Hruban RH, Kern SE. p16 and K-ras gene mutations in the intraductal precursors of human pancreatic adenocarcinoma. *Cancer Res* 1997; **57**: 2140-2143 [PMID: 9187111]
- 9 Hruban RH, van Mansfeld AD, Offerhaus GJ, van Weering DH, Allison DC, Goodman SN, Kensler TW, Bose KK, Cameron JL, Bos JL. K-ras oncogene activation in adenocarcinoma of the human pancreas. A study of 82 carcinomas using a combination of mutant-

- enriched polymerase chain reaction analysis and allele-specific oligonucleotide hybridization. *Am J Pathol* 1993; **143**: 545-554 [PMID: 8342602]
- 10 **Schutte M**, Hruban RH, Geradts J, Maynard R, Hilgers W, Rabindran SK, Moskaluk CA, Hahn SA, Schwarte-Waldhoff I, Schmiegel W, Baylin SB, Kern SE, Herman JG. Abrogation of the Rb/p16 tumor-suppressive pathway in virtually all pancreatic carcinomas. *Cancer Res* 1997; **57**: 3126-3130 [PMID: 9242437]
  - 11 **Muller PA**, Vousden KH. p53 mutations in cancer. *Nat Cell Biol* 2013; **15**: 2-8 [PMID: 23263379 DOI: 10.1038/ncb2641]
  - 12 **Iacobuzio-Donahue CA**, Song J, Parmigiani G, Yeo CJ, Hruban RH, Kern SE. Missense mutations of MADH4: characterization of the mutational hot spot and functional consequences in human tumors. *Clin Cancer Res* 2004; **10**: 1597-1604 [PMID: 15014009 DOI: 10.1158/1078-0432.CCR-1121-3]
  - 13 **Blackford A**, Serrano OK, Wolfgang CL, Parmigiani G, Jones S, Zhang X, Parsons DW, Lin JC, Leary RJ, Eshleman JR, Goggins M, Jaffee EM, Iacobuzio-Donahue CA, Maitra A, Cameron JL, Olin K, Schulick R, Winter J, Herman JM, Laheru D, Klein AP, Vogelstein B, Kinzler KW, Velculescu VE, Hruban RH. SMAD4 gene mutations are associated with poor prognosis in pancreatic cancer. *Clin Cancer Res* 2009; **15**: 4674-4679 [PMID: 19584151 DOI: 10.1158/1078-0432.CCR-09-0227]
  - 14 **Hezel AF**, Kimmelman AC, Stanger BZ, Bardeesy N, Depinho RA. Genetics and biology of pancreatic ductal adenocarcinoma. *Genes Dev* 2006; **20**: 1218-1249 [PMID: 16702400 DOI: 10.1101/gad.1415606]
  - 15 **Calhoun ES**, Jones JB, Ashfaq R, Adsay V, Baker SJ, Valentine V, Hempen PM, Hilgers W, Yeo CJ, Hruban RH, Kern SE. BRAF and FBXW7 (CDC4, FBW7, AGO, SEL10) mutations in distinct subsets of pancreatic cancer: potential therapeutic targets. *Am J Pathol* 2003; **163**: 1255-1260 [PMID: 14507635 DOI: 10.1016/S0002-9440(10)63485-2]
  - 16 **Hingorani SR**, Petricoin EF, Maitra A, Rajapakse V, King C, Jacobetz MA, Ross S, Conrads TP, Veenstra TD, Hitt BA, Kawaguchi Y, Johann D, Liotta LA, Crawford HC, Putt ME, Jacks T, Wright CV, Hruban RH, Lowy AM, Tuveson DA. Preinvasive and invasive ductal pancreatic cancer and its early detection in the mouse. *Cancer Cell* 2003; **4**: 437-450 [PMID: 14706336 DOI: 10.1016/S1535-6108(03)00309-X]
  - 17 **Ardito CM**, Grüner BM, Takeuchi KK, Lubeseder-Martellato C, Teichmann N, Mazur PK, Delgiorno KE, Carpenter ES, Halbrook CJ, Hall JC, Pal D, Briel T, Herner A, Trajkovic-Arsic M, Sipos B, Liou GY, Storz P, Murray NR, Threadgill DW, Sibilia M, Washington MK, Wilson CL, Schmid RM, Raines EW, Crawford HC, Siveke JT. EGF receptor is required for KRAS-induced pancreatic tumorigenesis. *Cancer Cell* 2012; **22**: 304-317 [PMID: 22975374 DOI: 10.1016/j.ccr.2012.07.024]
  - 18 **Collins MA**, Bednar F, Zhang Y, Brisset JC, Galbán S, Galbán CJ, Rakshit S, Flannagan KS, Adsay NV, Pasca di Magliano M. Oncogenic Kras is required for both the initiation and maintenance of pancreatic cancer in mice. *J Clin Invest* 2012; **122**: 639-653 [PMID: 22232209 DOI: 10.1172/JCI59227]
  - 19 **Cantley LC**. The phosphoinositide 3-kinase pathway. *Science* 2002; **296**: 1655-1657 [PMID: 12040186]
  - 20 **Altomare DA**, Tanno S, De Rienzo A, Klein-Szanto AJ, Tanno S, Skele KL, Hoffman JP, Testa JR. Frequent activation of AKT2 kinase in human pancreatic carcinomas. *J Cell Biochem* 2002; **87**: 470-476 [PMID: 14735903 DOI: 10.1002/jcb.10287]
  - 21 **Hirakawa T**, Yashiro M, Murata A, Hirata K, Kimura K, Amano R, Yamada N, Nakata B, Hirakawa K. IGF-1 receptor and IGF binding protein-3 might predict prognosis of patients with resectable pancreatic cancer. *BMC Cancer* 2013; **13**: 392 [PMID: 23962053 DOI: 10.1186/1471-2407-13-392]
  - 22 **Neesse A**, Michl P, Frese KK, Feig C, Cook N, Jacobetz MA, Lolkema MP, Buchholz M, Olive KP, Gress TM, Tuveson DA. Stromal biology and therapy in pancreatic cancer. *Gut* 2011; **60**: 861-868 [PMID: 20966025 DOI: 10.1136/gut.2010.226092]
  - 23 **Whipple C**, Korc M. Targeting angiogenesis in pancreatic cancer: rationale and pitfalls. *Langenbecks Arch Surg* 2008; **393**: 901-910 [PMID: 18210149]
  - 24 **Koch S**, Claesson-Welsh L. Signal transduction by vascular endothelial growth factor receptors. *Cold Spring Harb Perspect Med* 2012; **2**: a006502 [PMID: 22762016 DOI: 10.1101/cshperspect.a006502]
  - 25 **Seo Y**, Baba H, Fukuda T, Takashima M, Sugimachi K. High expression of vascular endothelial growth factor is associated with liver metastasis and a poor prognosis for patients with ductal pancreatic adenocarcinoma. *Cancer* 2000; **88**: 2239-2245 [PMID: 10820344]
  - 26 **Kindler HL**, Niedzwiecki D, Hollis D, Sutherland S, Schrag D, Hurwitz H, Innocenti F, Mulcahy MF, O'Reilly E, Wozniak TF, Picus J, Bhargava P, Mayer RJ, Schilsky RL, Goldberg RM. Gemcitabine plus bevacizumab compared with gemcitabine plus placebo in patients with advanced pancreatic cancer: phase III trial of the Cancer and Leukemia Group B (CALGB 80303). *J Clin Oncol* 2010; **28**: 3617-3622 [PMID: 20606091 DOI: 10.1200/JCO.2010.28.1386]
  - 27 **Cardin DB**, Goff L, Li CI, Shyr Y, Winkler C, DeVore R, Schlabach L, Holloway M, McClanahan P, Meyer K, Grigorieva J, Berlin J, Chan E. Phase II trial of sorafenib and erlotinib in advanced pancreatic cancer. *Cancer Med* 2014; **3**: 572-579 [PMID: 24574334 DOI: 10.1002/cam4.208]
  - 28 **Saif MW**. Pancreatic cancer: Sorafenib: no effect on efficacy of chemotherapy in pancreatic cancer. *Nat Rev Gastroenterol Hepatol* 2014; **11**: 8-9 [PMID: 24322903]
  - 29 **Ioka T**, Okusaka T, Ohkawa S, Boku N, Sawaki A, Fujii Y, Kamei Y, Takahashi S, Namazu K, Umeyama Y, Bycott P, Furuse J. Efficacy and safety of axitinib in combination with gemcitabine in advanced pancreatic cancer: subgroup analyses by region, including Japan, from the global randomized Phase III trial. *Jpn J Clin Oncol* 2015; **45**: 439-448 [PMID: 25647781 DOI: 10.1093/jjco/hyv011]
  - 30 **Kataoka Y**, Mukohara T, Tomioka H, Funakoshi Y, Kiyota N, Fujiwara Y, Yashiro M, Hirakawa K, Hirai M, Minami H. Foretinib (GSK1363089), a multi-kinase inhibitor of MET and VEGFRs, inhibits growth of gastric cancer cell lines by blocking inter-receptor tyrosine kinase networks. *Invest New Drugs* 2012; **30**: 1352-1360 [PMID: 21655918 DOI: 10.1007/s10637-011-9699-0]
  - 31 **Kiehne K**, Herzig KH, Fölsch UR. c-met expression in pancreatic cancer and effects of hepatocyte growth factor on pancreatic cancer cell growth. *Pancreas* 1997; **15**: 35-40 [PMID: 9211490 DOI: 10.1097/00006676-199707000-00005]
  - 32 **Chen HM**, Tsai CH, Hung WC. Foretinib inhibits angiogenesis, lymphangiogenesis and tumor growth of pancreatic cancer in vivo by decreasing VEGFR-2/3 and TIE-2 signaling. *Oncotarget* 2015; **6**: 14940-14952 [PMID: 25909285 DOI: 10.18632/oncotarget.3613]
  - 33 **Voldborg BR**, Damstrup L, Spang-Thomsen M, Poulsen HS. Epidermal growth factor receptor (EGFR) and EGFR mutations, function and possible role in clinical trials. *Ann Oncol* 1997; **8**: 1197-1206 [PMID: 9496384 DOI: 10.1023/A:1008209720526]
  - 34 **Tobita K**, Kijima H, Dowaki S, Kashiwagi H, Ohtani Y, Oida Y, Yamazaki H, Nakamura M, Ueyama Y, Tanaka M, Inokuchi S, Makuuchi H. Epidermal growth factor receptor expression in human pancreatic cancer: Significance for liver metastasis. *Int J Mol Med* 2003; **11**: 305-309 [PMID: 12579331 DOI: 10.3892/ijmm.11.3.305]
  - 35 **Yamanaka Y**, Friess H, Kobrin MS, Buchler M, Beger HG, Korc M. Coexpression of epidermal growth factor receptor and ligands in human pancreatic cancer is associated with enhanced tumor aggressiveness. *Anticancer Res* 1993; **13**: 565-569 [PMID: 8317885]
  - 36 **Philip PA**, Benedetti J, Corless CL, Wong R, O'Reilly EM, Flynn PJ, Rowland KM, Atkins JN, Mirtsching BC, Rivkin SE, Khorana AA, Goldman B, Fenoglio-Preiser CM, Abbruzzese JL, Blanke CD. Phase III study comparing gemcitabine plus cetuximab versus gemcitabine in patients with advanced pancreatic adenocarcinoma: Southwest Oncology Group-directed intergroup trial S0205. *J Clin Oncol* 2010; **28**: 3605-3610 [PMID: 20606093 DOI: 10.1200/JCO.2009.25.7550]
  - 37 **Safran H**, Iannitti D, Ramanathan R, Schwartz JD, Steinhoff M, Nauman C, Hesketh P, Rathore R, Wolff R, Tantravahi U, Hughes TM, Maia C, Pasquariello T, Goldstein L, King T, Tsai JY, Kennedy



- T. Herceptin and gemcitabine for metastatic pancreatic cancers that overexpress HER-2/neu. *Cancer Invest* 2004; **22**: 706-712 [PMID: 15581051 DOI: 10.1081/CNV-200032974]
- 38 **Moore MJ**, Goldstein D, Hamm J, Figer A, Hecht JR, Gallinger S, Au HJ, Murawa P, Walde D, Wolff RA, Campos D, Lim R, Ding K, Clark G, Voskoglou-Nomikos T, Ptasynski M, Parulekar W. Erlotinib plus gemcitabine compared with gemcitabine alone in patients with advanced pancreatic cancer: a phase III trial of the National Cancer Institute of Canada Clinical Trials Group. *J Clin Oncol* 2007; **25**: 1960-1966 [PMID: 17452677 DOI: 10.1200/JCO.2006.07.9525]
  - 39 **da Cunha Santos G**, Dhani N, Tu D, Chin K, Ludkovski O, Kamel-Reid S, Squire J, Parulekar W, Moore MJ, Tsao MS. Molecular predictors of outcome in a phase 3 study of gemcitabine and erlotinib therapy in patients with advanced pancreatic cancer: National Cancer Institute of Canada Clinical Trials Group Study PA.3. *Cancer* 2010; **116**: 5599-5607 [PMID: 20824720 DOI: 10.1002/cncr.25393]
  - 40 **Togashi Y**, Hayashi H, Nakagawa K, Nishio K. Clinical utility of erlotinib for the treatment of non-small-cell lung cancer in Japanese patients: current evidence. *Drug Des Devel Ther* 2014; **8**: 1037-1046 [PMID: 25114510 DOI: 10.2147/DDDT.S50358]
  - 41 **Pollak MN**, Schernhammer ES, Hankinson SE. Insulin-like growth factors and neoplasia. *Nat Rev Cancer* 2004; **4**: 505-518 [PMID: 15229476]
  - 42 **Urtasun N**, Vidal-Pla A, Pérez-Torras S, Mazo A. Human pancreatic cancer stem cells are sensitive to dual inhibition of IGF-IR and ErbB receptors. *BMC Cancer* 2015; **15**: 223 [PMID: 25886138 DOI: 10.1186/s12885-015-1249-2]
  - 43 **De Luca A**, Maiello MR, D'Alessio A, Pergameno M, Normanno N. The RAS/RAF/MEK/ERK and the PI3K/AKT signalling pathways: role in cancer pathogenesis and implications for therapeutic approaches. *Expert Opin Ther Targets* 2012; **16** Suppl 2: S17-S27 [PMID: 22443084 DOI: 10.1517/14728222.2011.639361]
  - 44 **Willems L**, Tamburini J, Chapuis N, Lacombe C, Mayeux P, Bouscary D. PI3K and mTOR signaling pathways in cancer: new data on targeted therapies. *Curr Oncol Rep* 2012; **14**: 129-138 [PMID: 22350330 DOI: 10.1007/s11912-012-0227-y]
  - 45 **Schlieman MG**, Fahy BN, Ramsamooj R, Beckett L, Bold RJ. Incidence, mechanism and prognostic value of activated AKT in pancreas cancer. *Br J Cancer* 2003; **89**: 2110-2115 [PMID: 14647146 DOI: 10.1038/sj.bjc.6601396]
  - 46 **Asano T**, Yao Y, Zhu J, Li D, Abbruzzese JL, Reddy SA. The PI 3-kinase/Akt signaling pathway is activated due to aberrant Pten expression and targets transcription factors NF-kappaB and c-Myc in pancreatic cancer cells. *Oncogene* 2004; **23**: 8571-8580 [PMID: 15467756 DOI: 10.1038/sj.onc.1207902]
  - 47 **Matsuzaki T**, Yashiro M, Kaizaki R, Yasuda K, Doi Y, Sawada T, Ohira M, Hirakawa K. Synergistic antiproliferative effect of mTOR inhibitors in combination with 5-fluorouracil in scirrhous gastric cancer. *Cancer Sci* 2009; **100**: 2402-2410 [PMID: 19764996 DOI: 10.1111/j.1349-7006.2009.01315.x]
  - 48 **Kordes S**, Klumpen HJ, Weterman MJ, Schellens JH, Richel DJ, Wilmink JW. Phase II study of capecitabine and the oral mTOR inhibitor everolimus in patients with advanced pancreatic cancer. *Cancer Chemother Pharmacol* 2015; **75**: 1135-1141 [PMID: 25822310 DOI: 10.1007/s00280-015-2730-y]
  - 49 **Llovet JM**, Ricci S, Mazzaferro V, Hilgard P, Gane E, Blanc JF, de Oliveira AC, Santoro A, Raoul JL, Forner A, Schwartz M, Porta C, Zeuzem S, Bolondi L, Greten TF, Galle PR, Seitz JF, Borbath I, Häussinger D, Giannaris T, Shan M, Moscovici M, Voliotis D, Bruix J. Sorafenib in advanced hepatocellular carcinoma. *N Engl J Med* 2008; **359**: 378-390 [PMID: 18650514 DOI: 10.1056/NEJMoa0708857]
  - 50 **Thomas SM**, Brugge JS. Cellular functions regulated by Src family kinases. *Annu Rev Cell Dev Biol* 1997; **13**: 513-609 [PMID: 9442882 DOI: 10.1146/annurev.cellbio.13.1.513]
  - 51 **Jatiani SS**, Baker SJ, Silverman LR, Reddy EP. Jak/STAT pathways in cytokine signaling and myeloproliferative disorders: approaches for targeted therapies. *Genes Cancer* 2010; **1**: 979-993 [PMID: 21442038 DOI: 10.1177/1947601910397187]
  - 52 **Seidel HM**, Lamb P, Rosen J. Pharmaceutical intervention in the JAK/STAT signaling pathway. *Oncogene* 2000; **19**: 2645-2656 [PMID: 10851064 DOI: 10.1038/sj.onc.1203550]
  - 53 **Müller S**, Raulefs S, Bruns P, Afonso-Grünz F, Plötner A, Thermann R, Jäger C, Schlitter AM, Kong B, Regel I, Roth WK, Rotter B, Hoffmeier K, Kahl G, Koch I, Theis FJ, Kleeff J, Winter P, Michalski CW. Next-generation sequencing reveals novel differentially regulated mRNAs, lncRNAs, miRNAs, sRNAs and a piRNA in pancreatic cancer. *Mol Cancer* 2015; **14**: 94 [PMID: 25910082 DOI: 10.1186/s12943-015-0358-5]
  - 54 **Lili LN**, Matyunina LV, Walker LD, Daneker GW, McDonald JF. Evidence for the importance of personalized molecular profiling in pancreatic cancer. *Pancreas* 2014; **43**: 198-211 [PMID: 24518497 DOI: 10.1097/MPA.000000000000020]
  - 55 **Tanase CP**, Neagu AI, Necula LG, Mambet C, Enciu AM, Calenic B, Cruceru ML, Albulescu R. Cancer stem cells: involvement in pancreatic cancer pathogenesis and perspectives on cancer therapeutics. *World J Gastroenterol* 2014; **20**: 10790-10801 [PMID: 25152582 DOI: 10.3748/wjg.v20.i31.10790]
  - 56 **Rao CV**, Mohammed A. New insights into pancreatic cancer stem cells. *World J Stem Cells* 2015; **7**: 547-555 [PMID: 25914762 DOI: 10.4252/wjsc.v7.i3.547]
  - 57 **Castellanos JA**, Merchant NB, Nagathihalli NS. Emerging targets in pancreatic cancer: epithelial-mesenchymal transition and cancer stem cells. *Onco Targets Ther* 2013; **6**: 1261-1267 [PMID: 24049451]
  - 58 **Leach SD**. Epithelial differentiation in pancreatic development and neoplasia: new niches for nestin and Notch. *J Clin Gastroenterol* 2005; **39**: S78-S82 [PMID: 15758664 DOI: 10.1097/01.mcg.0000155547.83901.a3]
  - 59 **Avila JL**, Kissil JL. Notch signaling in pancreatic cancer: oncogene or tumor suppressor? *Trends Mol Med* 2013; **19**: 320-327 [PMID: 23545339 DOI: 10.1016/j.molmed.2013.03.003]
  - 60 **Ristorcelli E**, Lombardo D. Targeting Notch signaling in pancreatic cancer. *Expert Opin Ther Targets* 2010; **14**: 541-552 [PMID: 20392166 DOI: 10.1517/14728221003769895]
  - 61 **Wang Z**, Ahmad A, Li Y, Azmi AS, Miele L, Sarkar FH. Targeting notch to eradicate pancreatic cancer stem cells for cancer therapy. *Anticancer Res* 2011; **31**: 1105-1113 [PMID: 21508353]
  - 62 **Chen JK**, Taipale J, Young KE, Maiti T, Beachy PA. Small molecule modulation of Smoothened activity. *Proc Natl Acad Sci USA* 2002; **99**: 14071-14076 [PMID: 12391318 DOI: 10.1073/pnas.182542899]
  - 63 **Thayer SP**, di Magliano MP, Heiser PW, Nielsen CM, Roberts DJ, Lauwers GY, Qi YP, Gysin S, Fernández-del Castillo C, Yajnik V, Antoniù B, McMahon M, Warshaw AL, Hebrok M. Hedgehog is an early and late mediator of pancreatic cancer tumorigenesis. *Nature* 2003; **425**: 851-856 [PMID: 14520413 DOI: 10.1038/nature02009]
  - 64 **Pasca di Magliano M**, Sekine S, Ermilov A, Ferris J, Dlugosz AA, Hebrok M. Hedgehog/Ras interactions regulate early stages of pancreatic cancer. *Genes Dev* 2006; **20**: 3161-3173 [PMID: 17114586 DOI: 10.1101/gad.1470806]
  - 65 **Bailey JM**, Swanson BJ, Hamada T, Eggers JP, Singh PK, Caffery T, Ouellette MM, Hollingsworth MA. Sonic hedgehog promotes desmoplasia in pancreatic cancer. *Clin Cancer Res* 2008; **14**: 5995-6004 [PMID: 18829478 DOI: 10.1158/1078-0432.CCR-08-0291]
  - 66 **Tanaka S**. Cancer stem cells as therapeutic targets of hepato-biliary-pancreatic cancers. *J Hepatobiliary Pancreat Sci* 2015; **22**: 531-537 [PMID: 25874410 DOI: 10.1002/jhpb.248]
  - 67 **Olive KP**, Jacobetz MA, Davidson CJ, Gopinathan A, McIntyre D, Honess D, Madhu B, Goldgraben MA, Caldwell ME, Allard D, Frese KK, Denicola G, Feig C, Combs C, Winter SP, Ireland-Zecchini H, Reichelt S, Howat WJ, Chang A, Dhara M, Wang L, Rückert F, Grützmann R, Pilarsky C, Izeradjene K, Hingorani SR, Huang P, Davies SE, Plunkett W, Egorin M, Hruban RH, Whitebread N, McGovern K, Adams J, Iacobuzio-Donahue C, Griffiths J, Tuveson DA. Inhibition of Hedgehog signaling enhances delivery of chemotherapy in a mouse model of pancreatic cancer. *Science* 2009; **324**: 1457-1461 [PMID: 19460966]
  - 68 **Kim EJ**, Sahai V, Abel EV, Griffith KA, Greenson JK, Takebe N, Khan GN, Blau JL, Craig R, Balis UG, Zalupski MM, Simeone DM. Pilot clinical trial of hedgehog pathway inhibitor GDC-0449



- (vismodegib) in combination with gemcitabine in patients with metastatic pancreatic adenocarcinoma. *Clin Cancer Res* 2014; **20**: 5937-5945 [PMID: 25278454 DOI: 10.1158/1078-0432.CCR-14-1269]
- 69 **Baarsma HA**, Königshoff M, Gosens R. The WNT signaling pathway from ligand secretion to gene transcription: molecular mechanisms and pharmacological targets. *Pharmacol Ther* 2013; **138**: 66-83 [PMID: 23328704 DOI: 10.1016/j.pharmthera.2013.01.002]
  - 70 **Espada J**, Calvo MB, Diaz-Prado S, Medina V. Wnt signalling and cancer stem cells. *Clin Transl Oncol* 2009; **11**: 411-427 [PMID: 19574199 DOI: 10.1007/s12094-009-0380-4]
  - 71 **Cui J**, Jiang W, Wang S, Wang L, Xie K. Role of Wnt/ $\beta$ -catenin signaling in drug resistance of pancreatic cancer. *Curr Pharm Des* 2012; **18**: 2464-2471 [PMID: 22372504 DOI: 10.2174/13816128112092464]
  - 72 **Fuxe J**, Karlsson MC. TGF- $\beta$ -induced epithelial-mesenchymal transition: a link between cancer and inflammation. *Semin Cancer Biol* 2012; **22**: 455-461 [PMID: 22627188 DOI: 10.1016/j.semcancer.2012.05.004]
  - 73 **Oberstein PE**, Olive KP. Pancreatic cancer: why is it so hard to treat? *Therap Adv Gastroenterol* 2013; **6**: 321-337 [PMID: 23814611 DOI: 10.1177/1756283X13478680]
  - 74 **Yashiro M**, Chung YS, Sowa M. Role of orthotopic fibroblasts in the development of scirrhous gastric carcinoma. *Jpn J Cancer Res* 1994; **85**: 883-886 [PMID: 7961114 DOI: 10.1111/j.1349-7006.1994.tb02963.x]
  - 75 **Yashiro M**, Ikeda K, Tendo M, Ishikawa T, Hirakawa K. Effect of organ-specific fibroblasts on proliferation and differentiation of breast cancer cells. *Breast Cancer Res Treat* 2005; **90**: 307-313 [PMID: 15830145 DOI: 10.1007/s10549-004-5364-z]
  - 76 **Rucki AA**, Zheng L. Pancreatic cancer stroma: understanding biology leads to new therapeutic strategies. *World J Gastroenterol* 2014; **20**: 2237-2246 [PMID: 24605023 DOI: 10.3748/wjg.v20.i9.2237]
  - 77 **Zhou H**, Roy S, Cochran E, Zouaoui R, Chu CL, Duffner J, Zhao G, Smith S, Galcheva-Gargova Z, Karlgren J, Dussault N, Kwan RY, Moy E, Barnes M, Long A, Honan C, Qi YW, Shriver Z, Ganguly T, Schultes B, Venkataraman G, Kishimoto TK. M402, a novel heparan sulfate mimetic, targets multiple pathways implicated in tumor progression and metastasis. *PLoS One* 2011; **6**: e21106 [PMID: 21698156 DOI: 10.1371/journal.pone.0021106]
  - 78 **Helleday T**, Bryant HE, Schultz N. Poly(ADP-ribose) polymerase (PARP-1) in homologous recombination and as a target for cancer therapy. *Cell Cycle* 2005; **4**: 1176-1178 [PMID: 16123586]
  - 79 **Bendell J**, O'Reilly EM, Middleton MR, Chau I, Hochster H, Fielding A, Burke W, Burris H. Phase I study of olaparib plus gemcitabine in patients with advanced solid tumours and comparison with gemcitabine alone in patients with locally advanced/metastatic pancreatic cancer. *Ann Oncol* 2015; **26**: 804-811 [PMID: 25573533 DOI: 10.1093/annonc/mdl581]
  - 80 **Pan Y**, Zheng M, Zhong L, Yang J, Zhou S, Qin Y, Xiang R, Chen Y, Yang SY. A preclinical evaluation of SKLB261, a multikinase inhibitor of EGFR/Src/VEGFR2, as a therapeutic agent against pancreatic cancer. *Mol Cancer Ther* 2015; **14**: 407-418 [PMID: 25519702 DOI: 10.1158/1535-7163.MCT-14-0485]
  - 81 **Awasthi N**, Hinz S, Brekken RA, Schwarz MA, Schwarz RE. Nintedanib, a triple angiokinase inhibitor, enhances cytotoxic therapy response in pancreatic cancer. *Cancer Lett* 2015; **358**: 59-66 [PMID: 25527450 DOI: 10.1016/j.canlet.2014.12.027]
  - 82 **Deplanque G**, Demarchi M, Hebbat M, Flynn P, Melichar B, Atkins J, Nowara E, Moyé L, Piquemal D, Ritter D, Dubreuil P, Mansfield CD, Acin Y, Moussy A, Hermine O, Hammel P. A randomized, placebo-controlled phase III trial of masitinib plus gemcitabine in the treatment of advanced pancreatic cancer. *Ann Oncol* 2015; **26**: 1194-1200 [PMID: 25858497 DOI: 10.1093/annonc/mdv133]
  - 83 **Shibaji T**, Nagao M, Ikeda N, Kanehiro H, Hisanaga M, Ko S, Fukumoto A, Nakajima Y. Prognostic significance of HIF-1  $\alpha$  overexpression in human pancreatic cancer. *Anticancer Res* 2003; **23**: 4721-4727 [PMID: 14981919]
  - 84 **Zhao T**, Ren H, Jia L, Chen J, Xin W, Yan F, Li J, Wang X, Gao S, Qian D, Huang C, Hao J. Inhibition of HIF-1 $\alpha$  by PX-478 enhances the anti-tumor effect of gemcitabine by inducing immunogenic cell death in pancreatic ductal adenocarcinoma. *Oncotarget* 2015; **6**: 2250-2262 [PMID: 25544770 DOI: 10.18632/oncotarget.2948]
  - 85 **Miyashita K**, Nakada M, Shakoori A, Ishigaki Y, Shimasaki T, Motoo Y, Kawakami K, Minamoto T. An emerging strategy for cancer treatment targeting aberrant glycogen synthase kinase 3  $\beta$ . *Anticancer Agents Med Chem* 2009; **9**: 1114-1122 [PMID: 19925395 DOI: 10.2174/187152009789734982]
  - 86 **Marchand B**, Tremblay I, Cagnol S, Boucher MJ. Inhibition of glycogen synthase kinase-3 activity triggers an apoptotic response in pancreatic cancer cells through JNK-dependent mechanisms. *Carcinogenesis* 2012; **33**: 529-537 [PMID: 22201186 DOI: 10.1093/carcin/bgr309]
  - 87 **Ma S**, Li Q, Pan F. CXCR4 promotes GSK3 $\beta$  expression in pancreatic cancer cells via the Akt pathway. *Int J Clin Oncol* 2015; **20**: 525-530 [PMID: 25145299]
  - 88 **Clark KL**, Halay ED, Lai E, Burley SK. Co-crystal structure of the HNF-3/fork head DNA-recognition motif resembles histone H5. *Nature* 1993; **364**: 412-420 [PMID: 8332212 DOI: 10.1038/364412a0]
  - 89 **Wang Z**, Banerjee S, Kong D, Li Y, Sarkar FH. Down-regulation of Forkhead Box M1 transcription factor leads to the inhibition of invasion and angiogenesis of pancreatic cancer cells. *Cancer Res* 2007; **67**: 8293-8300 [PMID: 17804744 DOI: 10.1158/0008-5472.CAN-07-1265]
  - 90 **Huang C**, Xie D, Cui J, Li Q, Gao Y, Xie K. FOXM1c promotes pancreatic cancer epithelial-to-mesenchymal transition and metastasis via upregulation of expression of the urokinase plasminogen activator system. *Clin Cancer Res* 2014; **20**: 1477-1488 [PMID: 24452790 DOI: 10.1158/1078-0432.CCR-13-2311]
  - 91 **Bartel DP**. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004; **116**: 281-297 [PMID: 14744438 DOI: 10.1016/S0092-8674(04)00045-5]
  - 92 **Rachagani S**, Kumar S, Batra SK. MicroRNA in pancreatic cancer: pathological, diagnostic and therapeutic implications. *Cancer Lett* 2010; **292**: 8-16 [PMID: 20004512 DOI: 10.1016/j.canlet.2009.11.010]
  - 93 **Park WG**, Yan BM, Schellenberg D, Kim J, Chang DT, Koong A, Patalano C, Van Dam J. EUS-guided gold fiducial insertion for image-guided radiation therapy of pancreatic cancer: 50 successful cases without fluoroscopy. *Gastrointest Endosc* 2010; **71**: 513-518 [PMID: 20189509 DOI: 10.1016/j.gie.2009.10.030]
  - 94 **Hwang JH**, Voortman J, Giovannetti E, Steinberg SM, Leon LG, Kim YT, Funel N, Park JK, Kim MA, Kang GH, Kim SW, Del Chiaro M, Peters GJ, Giaccone G. Identification of microRNA-21 as a biomarker for chemoresistance and clinical outcome following adjuvant therapy in resectable pancreatic cancer. *PLoS One* 2010; **5**: e10630 [PMID: 20498843 DOI: 10.1371/journal.pone.0010630]
  - 95 **MacKenzie TN**, Mujumdar N, Banerjee S, Sangwan V, Sarver A, Vickers S, Subramanian S, Saluja AK. Triptolide induces the expression of miR-142-3p: a negative regulator of heat shock protein 70 and pancreatic cancer cell proliferation. *Mol Cancer Ther* 2013; **12**: 1266-1275 [PMID: 23635652 DOI: 10.1158/1535-7163.MCT-12-1231]
  - 96 **Li Y**, Vandenboom TG, Wang Z, Kong D, Ali S, Philip PA, Sarkar FH. miR-146a suppresses invasion of pancreatic cancer cells. *Cancer Res* 2010; **70**: 1486-1495 [PMID: 20124483 DOI: 10.1158/0008-5472.CAN-09-2792]
  - 97 **Li L**, Li Z, Kong X, Xie D, Jia Z, Jiang W, Cui J, Du Y, Wei D, Huang S, Xie K. Down-regulation of microRNA-494 via loss of SMAD4 increases FOXM1 and  $\beta$ -catenin signaling in pancreatic ductal adenocarcinoma cells. *Gastroenterology* 2014; **147**: 485-497. e18 [PMID: 24859161]
  - 98 **Kern SE**, Shi C, Hruban RH. The complexity of pancreatic ductal cancers and multidimensional strategies for therapeutic targeting. *J Pathol* 2011; **223**: 295-306 [PMID: 21125682 DOI: 10.1002/path.2813]
  - 99 **Van Dort ME**, Galbán S, Wang H, Sebolt-Leopold J, Whitehead

- C, Hong H, Rehemtulla A, Ross BD. Dual inhibition of allosteric mitogen-activated protein kinase (MEK) and phosphatidylinositol 3-kinase (PI3K) oncogenic targets with a bifunctional inhibitor. *Bioorg Med Chem* 2015; **23**: 1386-1394 [PMID: 25766633 DOI: 10.1016/j.bmc.2015.02.053]
- 100 **Hu C**, Dadon T, Chenna V, Yabuuchi S, Bannerji R, Booher R, Strack P, Azad N, Nelkin BD, Maitra A. Combined Inhibition of Cyclin-Dependent Kinases (Dinaciclib) and AKT (MK-2206) Blocks Pancreatic Tumor Growth and Metastases in Patient-Derived Xenograft Models. *Mol Cancer Ther* 2015; **14**: 1532-1539 [PMID: 25931518 DOI: 10.1158/1535-7163.MCT-15-0028]
- 101 **Lindberg JM**, Newhook TE, Adair SJ, Walters DM, Kim AJ, Stelow EB, Parsons JT, Bauer TW. Co-treatment with panitumumab and trastuzumab augments response to the MEK inhibitor trametinib in a patient-derived xenograft model of pancreatic cancer. *Neoplasia* 2014; **16**: 562-571 [PMID: 25117978 DOI: 10.1016/j.neo.2014.06.004]
- 102 **Tang SC**, Chen YC. Novel therapeutic targets for pancreatic cancer. *World J Gastroenterol* 2014; **20**: 10825-10844 [PMID: 25152585 DOI: 10.3748/wjg.v20.i31.10825]

**P- Reviewer:** Talukdar R **S- Editor:** Ma YJ  
**L- Editor:** Filipodia **E- Editor:** Wang CH





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>



ISSN 1007-9327



9 771007 932045