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**Quo vadis motor neuron disease?**

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**Abstract**

Motor neuron disease (MND), also known as amyotrophic lateral sclerosis (ALS), is a relentlessly progressive neurodegenerative condition that is invariably fatal, usually within 3 to 5 years of diagnosis. The aetio-pathogenesis of MND remains unresolved and no effective treatments exist. The only FDA approved disease modifying therapy is riluzole, a glutamate antagonist, which prolongs survival by up to 3 mo. Current management is largely symptomatic/supportive. There is therefore a desperate and unmet clinical need for discovery of disease mechanisms to guide novel therapeutic strategy. In this review, we start by introducing the organizational anatomy of the motor system, before providing a clinical overview of its dysfunction specifically in MND. We then summarize what insights have been gained from pathological, genetic and animal models and conclude by speculating on optimal strategies to drive the step change in discovery, which is so desperately needed in this arena.

**Key words:** Motor neuron disease; Amyotrophic lateral sclerosis; Neurodegeneration; Disease models

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**Core tip:** Motor neuron disease (MND) is a fatal neurodegenerative disorder with no known cure. Here we discuss the organization of the motor system and the clinical presentation of MND. We detail the diagnostic criteria for ALS including electrophysiological studies and potential future diagnostic markers of disease. We discuss the staging of disease progression in MND. We then provide an overview of disease management and end with insights into molecular pathogenesis of the disease and the use of disease models.

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**Organizational anatomy of the motor system**

The complexity and capacity of the vertebrate nervous system is largely directed at the generation and control of movement by a carefully choreographed activation of muscles responsible for locomotion, swallowing and breathing. The motor system can be categorized most simply into upper and lower divisions. Within the brain, Betz cells in cortical layer V (the motor cortex) are classically large pyramidal upper motor neurons (MNs). Their smaller cortical counterparts densely populate the motor and premotor cortices. Upper MNs control lower MNs in the spinal cord either directly (monosynaptic input) or indirectly (through spinal interneurons). Descending motor neurons in the spinal cord travel in laterally partitioned corticospinal tracts, most of which cross the midline at the level of the lower brainstem medullary pyramids to synapse contralaterally within the spinal cord. There are also anterior corticospinal tracts, which do not cross at the medullary pyramids but remain ipsilateral. Notably, a minority of spinal cord regions are innervated by these anterior corticospinal projections, which branch and innervate on both sides of the spinal cord, crossing at the appropriate spinal segment. Direct synaptic connection between upper and lower MNs is likely a recent development in evolution, given that it is confined to the primary motor cortex and exclusive to higher primates.

Lower MNs are anatomically positioned in the ventral horns of the spinal cord and motor nuclei within the brainstem; these in turn synapse at neuromuscular junctions and muscle spindles forming a final common pathway for voluntary movement. Spinal MNs are large, polarized cells with long axons, and are the conduit through which the motor cortex in the brain activates contraction of skeletal muscles. These multipolar cells can project axons over a meter long and each innervate up to 1000 muscle fibres. Remarkably, their extensive dentritic arborisation can accommodate up to 10000 synaptic terminals, receiving input from descending upper MNs and spinal interneurons. Despite certain generic properties, distinct molecular phenotypes of MN exist. Even seemingly simple motor actions require collaboration and coordination of multiple MN subtypes, which are anatomically organized into motor columns and further grouped into motor pools in a muscle-specific manner. The generation of MN subtype diversity is an absolute pre-requisite to survival. In total, the human body has more than 100000 spinal MNs, which innervate 600 peripheral muscle targets organized into bilateral pairs. MNs can be classified according to the type of motor unit they generate into alpha, beta, and gamma. Alpha MNs are most abundant of all and are responsible for innervating extrafusal skeletal muscle, generating contractile force to move the skeleton. Indeed alpha MNs themselves can be further sub-divided by the contractile properties of the muscle fibres they innervate into fast-twitch fatigable (FF), fast-twitch fatigue-resistant (FR), and slow-twitch fatigue-resistant[1]. Beta MNs innervate both intra- and extrafusal fibres, although these are the least well-understood MN class. Gamma MNs innervate intrafusal muscle fibres of the muscle spindle, modulating their sensitivity to stretch[2,3]. Gamma MN cell somae are smaller than alpha MNs and they generally have slower axonal conduction velocities[4-6]. The dendritic arrangements of gamma MNs are significantly less complex when compared to an alpha MN arborisation[4]. Furthermore, all gamma MNs lack monosynaptic input from proprioceptive sensory neurons, whereas most alpha-MNs receive direct input[7,8]. This degree of structural and functional diversity commands distinct developmental lineage restriction programs for each different class of MN.

MN subclasses are spatially allocated into groups that reflect both their developmental origins and also their adult function. This coupling of developmental origin to adult function is depicted in Figure 1. MNs are developmentally partitioned into discrete motor columns, which extend along the rostro-caudal (R-C) neural tube. Within a column, the group of MNs responsible for innervating a single skeletal muscle is termed a motor pool, each of which is also arranged by an anatomical logic related to the muscle target(s) of its projections. The medial motor column (MMC) contains MNs that innervate dorsal epaxial muscles, which mainly subserve postural functions. Hypaxial motor column (HMC) MNs projects to the ventral hypaxial muscles, which is mainly involved in respiration. The lateral motor columns (LMC) are responsible for innervating limb muscles. The preganglionic motor column (PGC) is present at thoracic levels and MNs originating from here innervate sympathetic ganglia. The MMCs run throughout the R-C extent of the spinal cord, while the LMCs, HMCs and PGCs occur only at brachio-lumbar (LMCs) and thoracic (HMCs and PGCs) foci (Figure 1). The descriptive term ‘MN’ is thus an oversimplification and the numerous motor neuronal subtype differences described above (including R-C position, column, pool, axonal trajection) begin to demonstrate some of the necessary complexity inherent in MN diversification.

**Motor neuron disease (MND) – a clinical perspective**

MND causes progressive motor neuron (MN) degeneration in the anterior horn of the spinal cord, brain stem and motor cortex[9-12], invariably leading to fatal paralysis usually through respiratory failure[13,14]. The lifetime risk of MND is 1:400 in those of European ancestry[15]. Most cases (90%) are sporadic and affect men more than women. It can present at any age, but with a peak incidence in the sixth to seventh decades of life. Familial MND is caused by mutations in a variety of genes, about 60% of which are now identified[16-18]. Clinically, the patient history and examination typically suggest evidence of upper (UMN) and lower motor neuron (LMN) dysfunction in the absence of sensory or autonomic symptoms or signs. A striking clinical feature of this condition is the near universal sparing of the oculomotor nerves and the MNs in the sacral spinal cord that are responsible for pelvic sphincter control, called Onufrowicz nucleus.

Although initial presentation is quite variable, limb muscle weakness often begins focally (over 60% of cases, approximately equally distributed over upper and lower limb) and spreads in an orderly/stereotyped fashion, although overall patterns of motor weakness do vary quite widely between patients. While not pathognomonic, the so called ‘split-hand phenomenon’ is certainly a well-recognized feature of MND, clinically presenting as lateral hand muscle atrophy (*i.e.,* thenar eminence and first dorsal interosseous) with comparative normality of the medial hand muscles. Approximately 30% of patients present with bulbar symptoms, which include dysarthria, dysphagia and sialorrhoea (Table 1). Sialorrhoea is caused by inability to inability to swallow secretions due to a combination of tongue spasticity, weakness of the facial, mouth and pharyngeal muscles, and loss of oropharyngeal co-ordination and function[19]. Pseudobulbar palsy is also a recognized feature of MND, which can manifest clinically with spasticity of the tongue or of speech, a brisk jaw jerk, a positive gag reflex and mood incongruent emotionality. Muscle cramps and hypersalivation are common symptoms, and head drop, bilateral tongue wasting and widespread fasciculations important physical signs. Fasciculations can be a prominent and early sign in the disease[20]. Although only a minority of patients with MND initially present with acute respiratory failure, the majority do progress to this; indeed it is often the cause of their ultimate demise. Combined upper and lower MN dysfunction can be difficult to detect in early disease, sometimes explaining diagnostic uncertainty between different MND subtypes (Table 2) and some conceivable differential diagnoses (Table 2). Although a period of observation can be valuable for diagnostic clarification in this context (as concurrent upper and lower MN involvement will typically become more evident as the disease progresses), one must take into careful consideration the importance of making a timely diagnosis. Most MND patients who present with predominantly UMN pathology will develop lower MN signs within 3 or 4 years. The clinical diagnosis of MND is usually fairly self-evident, however it is critical not to miss any possible differential diagnoses listed in Table 1, as suggested by the history, examination and paraclinical tests.

The Revised EL Escorial diagnostic criteria and the Awaji electrodiagnostic criteria are well established for the clinical diagnosis of ALS and evaluate evidence for progressive degeneration of upper and/or lower motor neurons in the absence of other disease processes that could explain the clinical findings[21-23]. There are three diagnostic categories: clinically definite, probable or possible ALS (Table 1). Importantly, the Awaji criteria established equivalent importance of both clinical and electrophysiological findings when detecting chronic neurogenic changes[24]. A study prior to the introduction of the Awaji criteria found that 29% of ALS patients died without a diagnosis of definite ALS[25]. The Awaji diagnostic criteria have been shown to increase the sensitivity of ALS diagnosis[24,26]. As the diagnosis is made on the basis of upper and lower motor neuron involvement in bulbar and spinal regions, the addition of electrophysiology for more sensitive detection of lower motor neuron involvement facilitates the diagnosis. Evidence for neurogenic changes on the electromyography (EMG) should be sought[23]. Chronic neurogenic change may be demonstrated by motor unit potentials (MUPs) of increased amplitude and duration usually with increased number of phases; decreased motor unit recruitment or using a narrow pass filter to detect unstable or complex MUPs. Fibrillation potentials with positive sharp waves may be observed and fasciculation potentials with complex morphology, in the presence of chronic neurogenic change on needle EMG, may also be seen. The Revised El Escorial and Awaji criteria have proved very useful for diagnosis, especially for determining patient inclusion for clinical trials, however for use in clinical practice it is proposed that these criteria should be updated, to reflect the phenotypic heterogeneity of ALS, the stages of disease and the presence of familial disease[27].

Similarly the use of investigations to support upper motor neuron involvement would add further diagnostic certainty. Transcranial magnetic stimulation (TMS) is a technique used to measure corticomotoneuronal function with the parameters of motor threshold, motor evoked potential amplitude, central motor conduction time, cortical silent period, intracortical inhibition and facilitation[28]. Early cortical hyperexcitability, which may reflect glutamate excitotoxicity, precedes lower motor neuron involvement in ALS, and through the course of the disease this hyperexcitability decreases[28-32]. Threshold tracking TMS has the potential for use as a diagnostic marker and distinguishes ALS from non-ALS disorders with a sensitivity of 73.21% and specificity of 80.88% at an early disease stage[33]. Three hypotheses for motor neuron death have been proposed: a “dying-forward” phenomenon, where diseases initiates in corticomotoneurons, leading to excitotoxic death of lower motor neurons, a “dying-back phenomenon”, where disease begins at the lower motor neuron level and progresses back to the upper motor neurons, or an independent-degeneration phenomenon. The finding cortical hyperexcitability starts below lower motor neuron involvement supports the “dying-forward” hypothesis. Furthermore neuroimaging techniques, such as diffusion tensor MRI, are showing promise for determining motor cortical and corticospinal tract involvement in disease, and could be used as biomarkers of disease and predictors of prognosis[34].

Various staging systems have been devised to measure disease progression in ALS[35-40]. Individuals can progress through the disease at very variable rates[41,42], and as each clinical stage is reached at a consistent proportion through the disease process, staging can be used to make more useful comparisons between patients[35,36]. Furthermore, incremental stages correspond to decreasing function and health utility, and can be used in cost-benefit analyses of new treatments[38]. An important application of staging is as an endpoint in clinical trial design. The goal is to develop therapies which would prolong time in the earlier stages of disease, when function and quality of life are better, as compared to the later stages.

Cognitive impairment is recognized in up to half of patients with MND, usually detectable on neuropsychological testing rather than from routine clinical evaluation. However, frank dementia of the frontotemporal lobar degeneration (FTLD) type is increasingly diagnosed against the background of pathological and genetic discoveries that have mechanistically linked these two conditions together over the last decade[43]. Conversely, some patients presenting with FTLD will have clinical and para-clinical evidence of MND the mode of presentation here is likely determined by same pathomechanistic process starting/predominating at different neuraxial sites. Approximately 15% of ALS patients have a clinical diagnosis of FTLD and 15% of FTLD patients have a diagnosis of ALS[43,44].

Both European Federation of Neurological Societies and American Academy of Neurology guidelines for the management of MND patients have guided management to some degree in the United Kingdom[45-48]. Following a review decision in November 2014, NICE is currently developing a guideline for the management of MND. This will ultimately replace the current NICE guideline on non-invasive ventilation in MND. The MND Association website offers a comprehensive list of available regional and national/international guidelines in specific MND-related areas, with direct links to documents. Indeed the support of the MNDA in all respects is frequently fed back as being highly valued by patients and carers. Most patients will experience hypoventilation/orthopnea as the disease progresses, justifying proactive interval monitoring of respiratory performance (including nocturnal oximetry, dynamic forced vital capacity, and maximal inspiratory pressure). Noninvasive positive pressure ventilation (NIPPV) should be accessible when needed. Importantly, the management of MND should be in a multidisciplinary clinical setting, including experts in neurology, respiratory medicine, nutrition, psychology/psychiatry, speech therapy, physical and occupational therapy, social work, and case management. Other supportive measures include reactive and proactive interval examination of swallowing function as MND increases risk of aspiration. It is noteworthy that parotid/submandibular botulinum toxin injections can be helpful for sialorrhoea[19,49]. Consideration of a percutaneous gastrostomy (PEG) tube can help to maintain body weight and hydration in MND. Pseudobulbar affect is often treated off-licence with selective serotonin reuptake inhibitors or tricyclic antidepressants. In October 2010, FDA approved a dextromethorphan-quinidine combination for symptomatic relief of pseudobulbar affect.

**Lessons from pathological, genetic, animal and cellular models**

Various experimental strategies including *in-vivo* studies, cell based *in-vitro* approaches and human post-mortem neuropathological specimens from MND patients have been employed in order to improve understanding of this disease. Human stem cell strategies are becoming an increasingly important component of the armoury of investigative tools used to study disease mechanisms and identify potential therapeutic targets[50,51].

Historically, the most intensively studied cause of familial MND have been mutations in the copper/zinc superoxide dismutase (SOD1) gene, which account for approximately 15% of cases of familial ALS and less than 5% of sporadic ALS cases. The mutant SOD1 protein characteristically maintains its dismutase function, but appears to cause MN degeneration through alternative mechanisms, including a possible toxic gain of function[52]. Well over 100 individual point mutations located throughout the primary structure of SOD1 are sufficient to cause disease, suggesting protein-folding abnormalities as a possible initiating event. Transgenic mice globally expressing mutant forms of human SOD1 exhibit selective MN degeneration, which broadly mirrors the pathology of human sporadic and familial MND. Unfortunately, despite countless pre-clinical and clinical trials based on SOD1 models, not one of these has led to a significant therapeutic advance in MND. A landmark study in 2006 then discovered that the pathological hallmark of >95% MND cases (sporadic and familial) is cytoplasmic misaccumulation of ubiquitinated and hyperphosphorylated *transactive response DNA-binding* protein (TDP-43)[53], a highly conserved, ubiquitously expressed and multifunctional nuclear protein with both DNA and RNA binding capacities[54-56]. A striking observation made in this work was that TDP-43 appeared mislocalised from the nucleus to the cytoplasm in MND and FTLD, although the pathophysiological significance of this remains incompletely understood. Interestingly, TDP-43 immunoreactive inclusions are found in both neurons and glia in MND and FTLD, hence their proposed taxonomic reclassification as TDP-43 “proteinopathies”. SOD 1 mutations do not produce this common hallmark of MND and may not therefore be pathomechanistically representative of the majority of MND. Different subtypes of FTLD are based upon the protein found in pathological inclusions: in 45% of cases this is TDP-43, in another 45% of cases this is tau, and in 10% of cases this is *fused in sarcoma* (FUS)[43,57].

Other recent discoveries identified MND-causing gene mutations in TDP-43 and FUS[58,59]; findings that both complement and extend previous pathological studies. Furthermore, two recent contemporaneous studies have identified another MND-causing intronic mutation that introduces long hexanucleotide repeats into *C9orf72* pre-mRNA[60,61], which is the most frequent genetic cause of ALS and a common cause of FTLD. TDP-43 and FUS are both RNA-binding proteins (RBPs). Collectively, these discoveries implicate a deregulation of RNA metabolism as playing a crucial role in MND pathogenesis. In addition to these genes, several further mutations have been discovered including in the following genes: *PGRN, UBQLN2, SQSTM1, PFN1, ANG, VCP, MATR3, TUB4A*. Taken together, gene mutations and pathological studies implicate both deregulated protein homeostasis and RNA metabolism as underlying key pathways in molecular pathogenesis[43,58,59,62-66].

A widely held view regarding the pathogenesis of neurodegenerative disease posits that selective injury to a disease-specific subclass of neurons is mechanistically cell autonomous. This ‘neuron-centric’ view has been increasingly challenged by pivotal mice-chimera studies using lineage-specific expression of mutant SOD-1 and subsequent related investigation, which confirmed a major non cell-autonomous role for astrocytes and microglia in SOD1-related MND pathogenesis[67-69]. Non cell-autonomous injury has also recently been implicated in sporadic MND, raising the possibility of common pathogenic mechanisms[70,71].

The discovery of induced pluripotent stem cells (iPSC) enables patient-specific fibroblasts to be virally transduced with up to 4 transcription factors and ‘reprogrammed’ into embryonic-like stem cells[72]. Using insights from developmental neurobiology, these cells can subsequently be treated with a programme of extrinsic cues to direct their differentiation into a range of regionally defined neurons and glia for further study[73-76]. Importantly, a variety of studies have confirmed the capacity of these terminally differentiated cells to recapitulate key pathological hallmarks of a range of different neurodegenerative diseases[71,77-79]. In particular, several important studies have already demonstrated that iPSC-derived neurons and glia from patients with monogenic and sporadic MND show pathological phenotypes when compared to their control counterparts. Furthermore, this reductionist and human *in vitro* model system allows assays that directly elucidate non cell autonomous mechanisms of disease[80]. Several studies have also confirmed the utility in this model system as a pre-clinical test-bed for drug discovery[81-83], including the practical feasibility of high throughput automated approaches[84].

**Future strategies**

We conclude that the integration of human experimental approaches is required to drive the desperately needed discovery of disease mechanisms and therapeutic strategy in MND. Unfortunately animal models have failed to deliver a significant therapeutic advance in MND, despite numerous efforts and important discoveries. Human iPSCs models can better approximate clinical MND not only by virtue of species, but also because they express mutations at accurate pathophysiological levels and thus bypass the need for artificial overexpression, knock down or knock out experiments. A multitude of studies have now validated the human iPSC technology for disease modeling of both developmental and adult-onset conditions and drug discovery. However, this remains an *in vitro* system and thus lacks the dynamic cellular and signaling environments of an *in vivo* model. The integration of transgenic animal models that recapitulate MND pathogenesis together with patient-specific iPSCs represents an unprecedented opportunity to capture the complexity of pathogenic events underlying this devastating condition. By combining these approaches at the pre-clinical phase, we firmly believe that the translational yield of clinical trials will increase in MND.

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**Table 1 Possible differential diagnoses and diagnostic clues to discriminate from Motor neuron disease[23]**

|  |  |
| --- | --- |
| **Alternative diagnosis** | **Diagnostic clue** |
| Cervical (myelo) neuropathy | Cervicalgia, osteopaenia/porosis, abnormal cervical MRI  |
| Benign fasciculations | Absence of weakness, limited distribution, young age |
| Nutritional (B12 or Cu deficiency) | Usually have sensory impairment |
| Motor predominant CIDP | Relapsing-remitting course, evidence of demyelination on NCS, IVIG-responsive |
| Multifocal motor neuropathy with conduction block | Weakness with little wasting, distal and slowly progressive, absent bulbar involvement, conduction block on NCS  |
| Autoimmune and paraneoplastic | *e.g.,* stiff person’s syndrome: GAD, amphiphysin, gephyrin antibodies, EMG differences |
| HIV, HTLV1 | HIV: history, sensory neuropathy, opportunistic infections |
| Parsonage-Turner syndrome (or brachial neuritis) | Preceded by pain, preceding vaccination/viral illness, process arrests and followed by recovery, usually upper limb |
| Inclusion body myositis | Distribution – forearm and quadriceps, raised CK, muscle biopsy  |
| Hirayama’s disease | Upper limb, young males from Asia, unilateral, may arrest after a few years |
| Radiation-induced motor neuropathies | History and distribution of radiotherapy |
| Kennedy’s disease  | Family history (X-linked), Gynecomastia  |
| Spinal muscular atrophy  | Only affects LMNs |
| Primary progressive multiple sclerosis | MRI and/or cerebrospinal fluid (oligoclonal bands) |
| Adrenoleucodystrophy | Family history (X-linked), adult onset, slowly progressive, usually have sensory ataxia and sphincteric involvement |
| Hexosaminidase A deficiency  | Family history, dystonia, ataxia, psychosis |
| Poliomyelitis or post-polio syndrome | Clinical history and NCS/EMG  |
| Hereditary spastic paraparesis | Family history and genetic testing |

Cu: Copper; CIDP: Chronic inflammatory demyelinating polyneuropathy; NCS: Nerve conduction studies; IVIG: Intravenous immunoglobulin; GAD: Glutamic acid decarboxylase; EMG: Electromyography; HIV: Human immunodeficiency virus; HTLV: Human T-cell lymphotropic virus; LMN: Lower motor neuron.

**Table 2 Motor neuron disease subtypes, discriminating features and possible differential diagnoses**

|  |  |  |
| --- | --- | --- |
| **MND subtype** |  **Clinical features** | **Possible differential diagnoses** |
| Amyotrophic lateral sclerosis (ALS) | Affect both UMNs and LMNsOnset 50 s or 60 sMedian survival 3 to 5 years | Cervical myelomeuropathy HIV |
| Primary lateral sclerosis (PLS) | Only affect UMNs 3 yr from onsetOnset 50 s Profound spastricity Progressive quadriparesisLate cranial nerve involvementRarely bulbar onsetSlow progressionMedian survival 5 to 10 yr | Cervical myelopathy Nutritional (B12 or Cu deficiency)Primary progressive multiple sclerosisHereditary spastic paraparesis Stiff person syndromeTropical spastic paraparesis (HTLV1)AdremomyeloneuropathyHexosaminidase A deficiencyCorticobasal degeneration |
| Progressive muscular atrophy (PMA) | Only affect UMNs 3 yr from onsetFocal asymmetric distal weakneaa, followed by proximal involvementLate bullar/respiratory involvementEarlier onset than ALS Raised CK (< 10 × normal)Median survival 3 to 5 yr | Benign fasciculationsPost-polio syndromeAdult onset spinal muscular atrophy Inclusion body myositis |

UMN: Upper motor neuron; LMN: Lower motor neuron; HIV: Human immunodeficiency virus; Cu: Copper; CK: Creatinine kinase.

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**Figure 1 The motor columns of the spinal cord.** The LMCs innervate the muscles of the upper and lower limbs, the MMC innervates axial musculature and the HMC and PGC are in the thoracic spinal cord and innervate the intercostal musculature and sympathetic ganglia respectively. LMC: Lateral motor column; MMC: Medial motor column; HMC: Hypaxial motor column; PGC: Preganglionic motor column.