

Gene regulatory networks in atrial fibrillation

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10%-12% in 80+ years. Thus, AF represents nowadays a highly prevalent medical problem generating a large economic burden. At the electrophysiological level, distinct mechanisms have been elucidated. Yet, despite its prevalence, the genetic and molecular culprits of this pandemic cardiac electrophysiological abnormality have remained largely obscure. Molecular genetics of AF familiar cases have demonstrated that single nucleotide mutations in distinct genes encoding for ion channels underlie the onset of AF, albeit such alterations only explain a minor subset of patients with AF. In recent years, analyses by means of genome-wide association studies have unraveled a more complex picture of the etiology of AF, pointing out to distinct cardiac-enriched transcription factors, as well as to other regulatory genes. Furthermore a new layer of regulatory mechanisms have emerged, *i.e.*, post-transcriptional regulation mediated by non-coding RNA, which have been demonstrated to exert pivotal roles in cardiac electrophysiology. In this manuscript, we aim to provide a comprehensive review of the genetic regulatory networks that if impaired exert electrophysiological abnormalities that contribute to the onset, and subsequently, on self-perpetuation of AF.

Key words: Atrial fibrillation; Genetics; MicroRNAs; Genome-wide association studies; PITX2

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Core tip: Atrial fibrillation (AF) is the most prevalent arrhythmogenic defect in the human population. Genetic factors such as mutations in distinct ion channel encoding genes have been described, yet representing less than 10% of all AF cases. Genome wide association studies have widened the genetic culprits contributing to AF. We provide herein a state-of-the art review on the genetic components underlying AF. Experimental evidences demonstrated that PITX2 plays a pivotal role regulating cellular, molecular and electrophysiological characteristics of the developing and adult heart that, if impaired, predispose to AF, leading to complex regulatory networks with transcriptional and post-transcriptional (microRNA)

Abstract

Atrial fibrillation (AF) is the most frequent arrhythmogenic syndrome in humans. With an estimate incidence of 1%-2% in the general population, AF raises up to almost

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BACKGROUND

Atrial fibrillation (AF) is the most frequent arrhythmogenic syndrome in humans. With an estimate incidence of 1%-2% in the general population, AF raises up to almost 10%-12% in 80+ years^[1-5]. Cardiovascular risk factors such diabetes, obesity, hypertension and hyperthyroidism promote AF^[6-9]. In addition, the occurrence of AF can be also triggered by preceding cardiovascular diseases such as hypertrophic cardiomyopathy and valvular heart diseases^[10,11] as well as it can be boosted by cardiac surgery, obstructive sleep apnea and inflammatory processes^[12-15]. Importantly, besides the global risk factors for AF, it is well-established that the debut of an AF episode triggers subsequent and more severe AF episodes, leading to electrical and structural remodeling of the diseased heart, a condition quoted as "AF begets AF"^[16,17]. Thus AF *per se* is a progressive disease. Electrical remodeling involves progressive changes in the cardiac electrical properties, possibly triggered by oxidative stress imbalance, leading to EADs, DADs and/or changes in the ADP configuration^[18,19], culminating thus in rotor formation^[20] (Figure 1). Structural remodeling involves atrial dilation, fibrosis and/or inflammation^[21,22] which then indirectly promotes the generation of rotors and thus electrical re-entry circuitries^[20] (Figure 1). Moreover, suffering from AF predisposes to additional health problems, ranging from bleeding and stroke^[23-25], cognitive decline and dementia^[26,27], ventricular dysfunction^[28,29] and even sudden death^[30] (Figure 1). Thus, these data support the notion that AF is an epidemic disease with large socio-economic burden worldwide.

While large epidemiological data support the previously mentioned predictive risk factors for AF, there are also unquestionable evidences that in a subset of AF patients no concurrent previous risk factors are diagnosed and thus AF occurs in an isolated manner, a condition dubbed lone AF^[31-34]. These observations, in addition to seminal observations supporting that AF can also be familial^[35], strengthen the notion of a genetic component on the onset of AF. In this context, two waves of genetic information have been gained over the last decades. Seminal studies on familiar AF cases followed by genetic linkage analyses, and subsequently by genetic screening of candidate genes identified a large number of point mutations in distinct genes encoding for proteins involved in cardiac electrophysiology^[36-38]. These data, along with meticulous electrophysiological mapping analyses and *in silico* data modeling, provided an important cornerstone to understand AF pathophysiology^[39-43].

However, genetic identification of candidate AF genes can only explained around 10%-15% of all AF patients. Thus, new approaches to untangle the genetic bases of AF were envisioned. Genome-wide association analyses (GWAS) lightened the discovery of new genes associated to AF. Seminal worked by Gudbjartsson *et al*^[44] firstly identify common risk variants highly associated to the onset of lone AF in distinct large European and Chinese patient cohorts. Subsequent GWAS studies and meta-GWAS have further identified new candidate genes for AF pathophysiology^[45-49], yet the functional link between these variants and the AF pathophysiology is a rather unexplored. A tortuous route to dissect their functional roles is envisioned, given that most of these risk variants are located rather apart from gene coding sequences. Thus, a first proposal based on these findings is that risk variants might affect gene regulatory networks within the vicinity and thus underscore AF onset. In addition, a novel layer of complexity in gene regulatory networks has emerged with the discovery of non-coding RNAs that can mediate post-transcriptional regulation. In this review, we aim to elaborate on the current state-of-the-art of the gene regulatory networks involved in AF pathophysiology.

CONTRIBUTION OF THE IMPAIRED CARDIAC ACTION POTENTIAL TO THE AF PATHOPHYSIOLOGY

The cardiac action potential is driven by inward and outward flow of distinct ion currents along the cardiomyocyte membrane. In atrial and ventricular cardiomyocytes, the configuration of the action potential is initiated by the upstroke depolarization of the sodium current, followed by a repolarization mediated by distinct potassium currents until the resting membrane potential is restored. A conduction-contraction coupling is modulated within each action potential by a complex regulatory network of calcium handling^[50]. Interestingly, nodal conductive cells display a distinct upstroke configuration which is modulated by cation selective currents, yet repolarization is rather similar to working cardiomyocytes. A large number of mutations in the *SCN5A* gene, coding for the pore-forming subunits of the I_{Na} current, have been associated to AF, alone^[51-56] or in combination to other concurrent cardiac pathologies^[57-62]. Similarly, mutations in *SCN1B*, *SCN2B*, *SCN3B* and *SCN4B* ancillary subunits, respectively, have also been linked to AF^[63-67]. At present, the functional interpretation of these findings suggest that impaired I_{Na} current can promote AF onset, yet electrophysiological evidences are only available for a subset of these mutations. Importantly, mutations in *SCN5A* are also linked to other cardiac arrhythmias such as long QT and Brugada^[68-70] demonstrating a pivotal role for *SCN5A* on cardiac electrophysiology and illustrating that distinct phenotype might be acquired depending on subtle differences on the mutation location as well as on plausible modulatory accompanying proteins. In addition, mutations in the *HCN4* cation channel, responsible for the cardiac

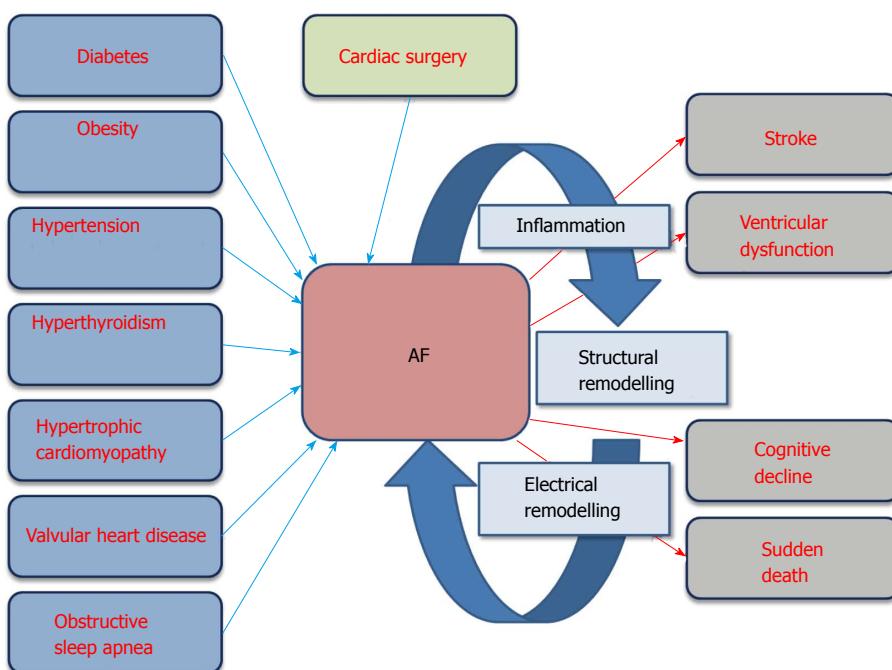


Figure 1 Schematic representation of the clinically relevant risk factors influencing the onset of atrial fibrillation. Atrial fibrillation (AF) leads thereafter to distinct cellular and molecular remodeling events, including structural and electrical remodeling, in part as caused by inflammatory process. In addition, progression of AF is highly associated with the onset of additional cardiac pathophysiologies.

action potential upstroke of conductive cells, have also been reported in AF^[71].

Mutations in genes encoding for proteins controlling the repolarization phase of the cardiac action potential have also been associated to AF. In particular, *KCNQ1*^[36,72-79], *KCNA5*^[80] and *KCNE2*^[38,81] mutations have been identified. In this setting, point mutations seem to shorten the repolarization phase providing an electrophysiological substrate for AF onset. Similarly, alteration of the resting membrane potential have been related to trigger AF onset and a large number of mutations in the gene coding for *KCNJ2* have been reported^[37,38,82,83]. Similarly as for the sodium channels, mutation in potassium channels have been associated to AF and concomitantly with other arrhythmogenic syndromes such as long^[72] and short^[84] QT, respectively.

Multiple lines of evidences have demonstrated the essential role of impaired calcium homeostasis as a triggering factor of AF^[2,85-90]. In this context, mutations in *RYR2* are associated to AF^[91] and in conjunction with other cardiac pathophysiological conditions such as catecholaminergic polymorphic ventricular tachycardia^[92,93]. Curiously, *RYR2* mutations can also affect sodium channel expression^[94]. Importantly, murine model of *RYR2* mutation leads to AF, reinforcing the genetic evidences of impaired calcium homeostasis as a trigger of AF^[95].

In addition, mutations in gap junction proteins such as *GJA1* (connexin43)^[96,97] and *GJA5* (connexin40)^[98-104], which are critical for the coordinated transmission of the electrical impulse among cardiomyocytes, have also been identified in association to AF. Seminal work described that mutation in connexin proteins were occurring in somatic cells (*i.e.*, cardiomyocytes)^[98], whereas more recently germline mutations have also been identified^[98-104] and the concept of somatic cell mutation challenged^[105]. In this setting, electrophysiological analyses of connexin

mutations in the AF context demonstrated that cardiomyocyte electrical transmission is impaired^[106-111] and thus can result in cardiac re-entry circuitries and then on AF.

Furthermore, mutations in several other genes, besides those directly contributing to the electrophysiological properties of the heart, have also been identified in the AF setting. In particular, mutations in the transcription factors *TBX5*^[112], *NKX2.5*^[113-117] and *NKX2.6*^[118] as well as in the nuclear pore component *NUP155*^[119] and the atrial natriuretic factor (*NNP4*^[120]). A summary of gene mutations associated to AF is provided on Figure 2. It is important to highlight in this context that mutation in *NUP155* was found in patients with AF and early sudden death^[118], while *TBX5* and *NKX2.5* are developmental transcription factors that play determinant roles during cardiogenesis and mutations in these genes have also been reported in distinct congenital heart diseases as well as GWAS candidate genes in other cardiac electrophysiological defects^[121-128].

NOVEL GENES LINKED TO AF

As previously said, classical genetic approaches provided an entry site to discover discrete genes involved in AF pathophysiology, but felt short to explain most of the diagnosed AF cases. Genome-wide association studies (GWAS) introduced a revolutionary genetic approach to understand AF, and cardiac arrhythmias at large^[129,130]. Seminal work by Gudbjartsson et al^[44] identified risk variant at 4q25 highly associated to lone AF. Soon thereafter these observations were corroborated in distinct studies worldwide^[131-136], yet with some controversial findings^[137,138]. Surprisingly, 4q25 risk variants are located in a gene desert, being the closest annotated gene a homeobox transcription factor, *PITX2*, around 150 kb downstream. Soon thereafter, experimental observations provided

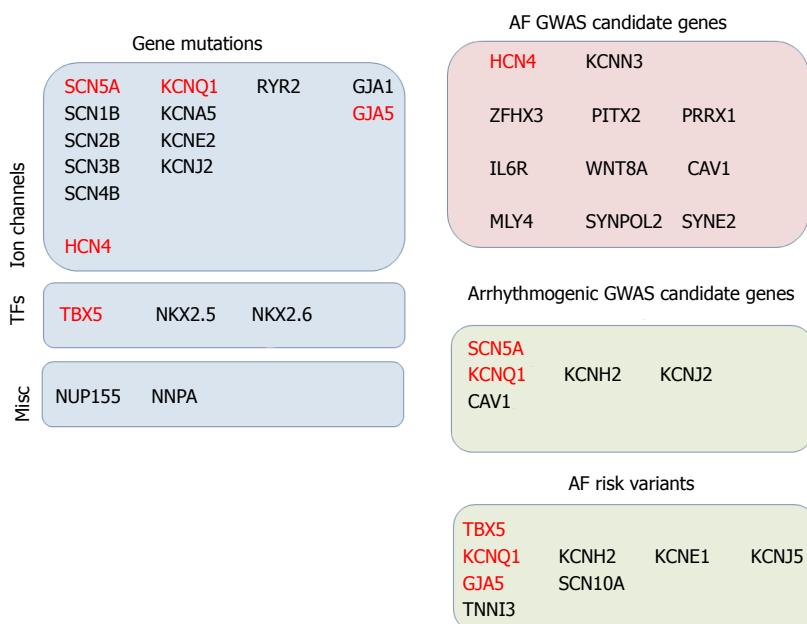


Figure 2 Schematic representation genes associated to atrial fibrillation. On the one hand, gene mutations associated to atrial fibrillation (AF) are illustrated. On the other hand, genes associated to AF by means of GWAS are also illustrated. In the latter, three subset of genes are provided: (1) those directly linked to AF GWAS analyses; (2) those related to AF, but identified in arrhythmogenic syndromes GWAS analyses; and (3) risk variants association studies in small AF cohorts by a candidate approach strategy. Genes highlighted in bold represent those genes identified in AF by multiple approaches. GWAS: Genome-wide association analyses.

evidences that impaired *Pitx2* function in animal models triggered increase atrial arrhythmias susceptibility^[139,140]. However, how risk variants can influence *PITX2* expression and/or function have only been partially revealed^[141], since regulatory elements containing the risk variants can molecularly interact with *PITX2* but also *ENPEP* promoter elements in a tissue- and developmental-specific manner^[141]. Importantly, controversial findings in humans have been reported. Chinchilla et al^[136] firstly described decreased *PITX2* expression in AF patients, yet these claims were challenged since no correlation between risk variants and *PITX2C* expression was observed^[142]. However, more recently a correlation with *PITX2A* expression has been reported, limiting their functional significance since data were obtained from right atrial biopsies^[143]. Thus, further investigations are required to clarify these controversial findings.

Besides *PITX2*, additional GWAS have enlightened the putative role of other novel genes in AF pathophysiology, such as *KCNN3*^[47], *ZFHX3*^[45,48], *IL6R*^[49] and *MLY4*^[144]. In most cases, these findings have been corroborated in independent studies with distinct ethnicity cohorts as for *ZFHX3*^[145,146], and *IL6R*^[147], but to date no additional reports are available for *KCNN3* and *MLY4*.

While arguments for the plausible involvement in AF have been provided in all cases, experimental evidences remain largely missing. *KCNN3* loss-of-function mutant mice display no overt cardiac electrophysiological defects, yet overexpression leads to sudden death^[148]. Similarly *ZFHX3* function role in the heart is scarce, yet recent findings suggests that atrial arrhythmias^[149] and pacing^[150] leads to significant impaired *ZFHX3* expression. Importantly, risk variants associated to *KCNN3*, *ZFHX3* and *IL6R* lie within intronic regions, respectively, and thus have no predictive functional consequences. Therefore, as in the cases of 4q25 risk variants (*PITX2*), a functional role on plausible regulatory elements is hypothesized,

but functional evidences are yet missing.

In addition to these GWAS studies, at least two meta-GWAS studies have digged for additional risk variants associated to AF^[46,151]. In this context, six new loci have been uncovered using such approach; *CAV1*, *HCN4*, *SYNE2*, *SYNPOL2*, *PRRX1* and *WTN8A*. Replication analyses of the risk variants have been confirmed for *SYNE2*^[152] and *PRRX1*^[149], are controversial for *CAV1*^[53,153,154] or have not been reported to date for *HCN4* and *SYNPOL2*. Importantly, a 12-SNPs risk score have been developed that can individually identify the risk of AF and stroke^[155].

While this approach broadens the plausible genetic bases of AF, it is also true that their contribution to explain AF pathophysiology would be directed linked to the statistical significance associated with those findings. It is important to say in this context that risk variants associated to *PITX2* display highest significance values, *KCNN3*, *ZFHX3* and *IL6R* are moderately significant and those revealed by meta-GWAS display the lowest, yet statistical significant, values. In a simplistic way, this could be transferred as *PITX2* being contributing to larger spectrum of AF pathophysiology and thus being in the upper level of a hierarchical signaling pathway while *CAV1*, *HCN4*, *SYNE2*, *SYNPOL2*, *PRRX1* and *WTN8A* will be rather discretely contributing to AF pathophysiology in a rather discrete, small subset of AF patients, and therefore being at the bottom of such hierarchy. We have recently provided experimental evidences supporting this notion, as detailed below.

GWAS studies have also been reported in other electrophysiological parameters such as PR interval^[129,156-158], heart rate^[157], QRS interval^[157] as well as physiopathological conditions such as Brugada syndrome^[159] and long QT^[160-162]. Several of these cardiac arrhythmogenic conditions are sometimes directly associated to AF, such as for example impaired PR interval or Brugada syndrome. Importantly,

such GWAS studies have identified multiple genes, some of which are shared with AF, such as *CAV1* (PR interval^[129,156-158]) while other have been previously involved in AF pathophysiology, such as *KCNQ1*, *KCNH2*, *SCN5A*, *KCNJ2* (long QT GWAS analyses^[160-162]). In addition to GWAS studies, risk variants have been associated to AF using more discrete/small AF patient cohorts. Among them it is important to highlight risk variants at the following loci; *KCNQ1*^[163-165], *KCNH2*^[163], *KCNE1*^[163], *KCNJ5*^[166], *GJA5*^[167] and *TBX5*^[168]. In addition to these GWAS studies, several independent AF cohort studies have reported risk variants associated to AF in other loci, such as *TNNI3*^[169] and *SCN10A*^[170,171], yet the functional relevance of these findings remains unsolved. These data suggest that complex and intricate gene networks are likely to be operative between these regulatory genes resulting in AF (Figure 3) as well as other electrophysiological associated disorders.

A NOVEL AND EMERGING LAYER OF GENE REGULATORY CONTROL

Gene regulatory networks have been classically associated as hierarchical interactions between master genes, such as transcription factors, and structural genes, such as actin and myosin genes forming the sarcomere or genes encoding ion channels configuring therefore the cardiac action potential. In this setting, transcriptional regulation has been considered the master regulatory point, deciding whether to go or not to go, whereas translation and post-translational modifications are considered as minor regulatory nodes. Over the last decade, we have witnessed the discovery that large part of the so called “rubbish DNA” in fact codes for a large variety of non-coding RNAs species with highly relevant functional properties. In broad sense, non-coding RNAs can be sub-classified into long non-coding (lncRNAs) and small non-coding RNAs, exerting a distinct and variable functional roles^[172]. Among small non-coding RNAs, microRNAs have emerged as the larger group, playing a pivotal role on post-transcriptional regulation of mRNA transcript expression^[173]. MicroRNAs are 22-24 nt non-coding RNAs which can block translation or trigger transcript degradation by binding to complementary seed sequence in the 3' untranslated region (3'UTR) of nascent mRNA transcripts^[173]. At present the hallmark of microRNA transcriptomics has been uncovered in the developing and adult heart in both normal and pathological conditions^[174,175]. In the context of AF the microRNA fingerprint has been profiled in different settings^[176-178], providing highly valuable biomarkers^[179-183] as well as opening new avenues to dissect the functional role of these differentially expressed microRNAs in AF pathology. In addition distinct sets of microRNAs have also been reported to selectively modulate expression of distinct gene encoding for ion channels, opening thus new pathways to dissect their genetic contribution to AF.

AN INTEGRATIVE GENE REGULATORY NETWORK LINKED TO AF

As stated above, evidence on the functional role of multiple genes have been provided by the identification and experimental testing of AF patient gene mutations. In this setting, a causative link can, in most cases, be established. However, GWAS analyses provide circumstantial evidences whereas functional linkage between the risk variants and coding genes, whether protective or deleterious, is not directly established. To date, an example of successful linkage have been reported^[184] in the cardiovascular context, however in most cases, this is not yet established. Therefore, dissecting the functional role of GWAS candidate genes represents the first step for the analyses of their causative link.

Large set of evidences suggest that multiple genetic pathways can contribute to the onset of AF. Over the last years we have carefully dissected the role of the homeobox transcription factor PITX2 in this context and we have made efforts to dissect the regulatory interaction of *PITX2* with those genes previously reported to play a role in AF pathophysiology. Several studies have demonstrated that *PITX2* can influence expression of distinct ion channels in the developing and adult hearts. Wang et al^[138] firstly demonstrate that *PITX2* haploinsufficiency is sufficient to increase AF susceptibility. These authors demonstrated that *PITX2* haploinsufficiency led to developmental impaired expression of key transcription factors (*SHOX2*, *TBX3*) directing sinoatrial morphogenesis and thus impacting on the normal onset of the cardiac pacemaker structures. However, in this context, no basal AF occurs and no ECG defects were observed. In this line of thinking soon thereafter Kirchoff et al^[139] demonstrate similar findings and furthermore reported that expression of multiple ion channels were impaired as well as cell-cell interacting proteins (Figure 3). However, none of these genes were reported to play a pivotal role on cardiac action potential and thus, molecular and electrophysiological links were still on its infancy. Using a distinct experimental approach, i.e., an atrial-specific *PITX2* conditional transgenic mouse line, Chinchilla et al^[135] revealed that ECG defects were observed at rest (no triggering needed), electrophysiological measurements were impaired, i.e., increased resting membrane potential and prolonged action potential duration, and furthermore using a candidate approach analyses, distinct ion channels were also impaired, particularly *SCN5A* (I_{Na}), *KCNJ2* (I_{K1}) and *KCNJ12* (I_{K1}). Thus, these data demonstrate for the first time that functional impairment of *PITX2* leads to molecular defects which provoke electrophysiological alterations and thus increase rate of atrial arrhythmias. In line with these findings, Tao et al^[185] using a *PITX2* deletion conditional approach in the adult heart also demonstrated that basal ECG defects were observed, similar to those already reported by Chinchilla et al^[135], and moreover they

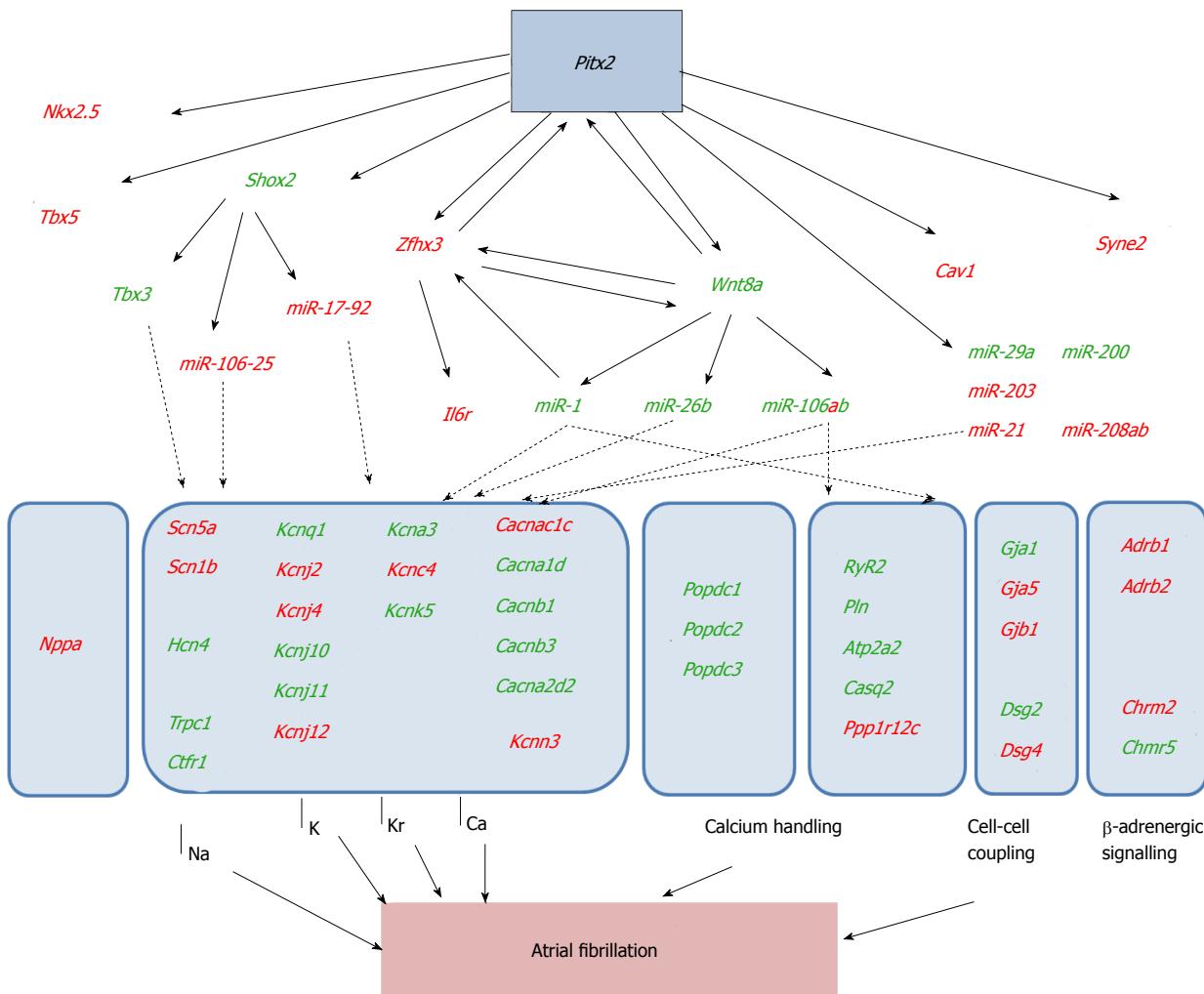


Figure 3 Schematic representation of the *Pitx2* downstream pathways involved in pro-arrhythmic events leading to atrial fibrillation. Multiple lines of evidences demonstrate that *PITX2* exerts a pivotal role regulating expression of distinct ion channels, cell-cell coupling, blood pressure controllers and beta-adrenergic stimulation. Most of these pathways are modulated by microRNAs which are under control of *PITX2* action. Red denotes down-regulation and green upregulation.

demonstrated that key determinants of the cardiac action potential were impaired, such as *KCNQ1* (I_{Ks}), *KCNH2* (I_{Kr}), *CACNA1C* (I_{Ca}), among others. Overall these data demonstrated that *Pitx2* insufficiency can both promote and trigger molecular and electrophysiological alterations, namely affecting the depolarization, repolarization and resting membrane potential characteristics of the cardiac action potential. They also illustrate that alteration of a single transcription factor can influence the functional role of multiple ion channels previously reported to be associated to AF. Therefore it can be inferred that *PITX2* is upstream of this hierarchical signaling pathway and its impairment would explain a large fraction of AF patients.

In addition to *PITX2*, other GWAS studies have provided evidence on the plausible role of distinct gene in AF pathophysiology, yet as previously said, their functional implication has been poorly documented. We investigated whether these genes would be under control of *PITX2*. Using a series of gain and loss-of-function approaches, Lozano-Velasco et al^[149] demonstrated that *PITX2* can modulate expression of *ZFHx3*, *KCNN3* and *IL6R*. The

role of *ZFHx3* in atrial arrhythmias is basically unexplored. However, given its transcriptional capacity it might be plausible that it can modulate multiple genes previously related to AF. The role of *KCNN3* in atrial arrhythmias is under debate, its plausible contribution to the configuration of the cardiac action potential, if any, seems to be residual. On the other hand, the involvement of *IL6R* could be related to inflammatory processes that, if impaired, are associated to AF as previously mentioned. We have noticed that *IL6R* expression in the atrial chambers is impaired in *PITX2* deficient mice yet plasma levels of soluble *IL6* and *TFN α* ligands are not altered. While these data support a tissue-specific involvement of *IL6R*, its links to AF remain to be elucidated.

We also investigated if *PITX2* can influence the role of those genes related to AF by meta-GWAS analyses. Among them, previous evidences support that *PITX2* can modulate expression of *Hcn4*^[138]. More recently, Lozano-Velasco et al^[151] demonstrated that also *CAV1*, *SYNE2* and *WNT8A* are modulated by *PITX2*. It is important nonetheless to highlight that risk variants associated

to *CAV1* and *SYNE2* have also been reported in other cardiac electrophysiological disorders yet their functional role remains poorly characterized. Importantly, WNT signaling has been largely documented to play pivotal role during cardiac development and homeostasis^[186,187], however WNT8 has not been reported to play fundamental roles therein^[188].

Given the potential signaling properties of WNT8A and those transcriptional capacity exerted by ZFHX3 we explored in any of those pathways might be involved on the regulation of distinct ion channels previously reported to be modulated by PITX2 and related to AF. Gain- and loss-of-function approaches demonstrated that *WNT8A* but not *ZFHX3* can modulate expression of calcium handling proteins^[151]. Importantly, impaired expression of *WNT8A* is observed in *PITX2* insufficiency models with basal ECG alterations but not in those with normal ECG recordings^[151], supporting the notion that WNT signaling plays a fundamental role conferring susceptibility vs triggering capacity to the atrial chambers during AF onset.

In addition to gene regulatory mechanisms driven by *PITX2*, several lines of evidence support that *PITX2*-regulated microRNAs also contribute to AF pathophysiology. Chinchilla et al^[135] already demonstrated that impaired miR-1 expression in *PITX2* deficient mice underlie abnormal resting membrane potential configuration (by modulating *KCNJ2* and *KCNJ12* post-transcriptional regulation). Huang et al^[153] has confirmed that *PITX2* inhibits miR-1 expression, which negatively regulates *ZFHX3*. Wang et al^[189] demonstrated that two distinct genomic microRNA clusters, namely *miR-17-92* and *miR-106b-25*, as well as *miR-335* and *miR-423*, are under control of *PITX2*. Genetic deletion of *miR-17-92* and *miR-106b-25* microRNA clusters, respectively display normal baseline electrophysiological parameters but were susceptible to pacing-induced AF. Cardiac-specific deletion (Nkx2.5Cre) of *miR-17-92* cluster resulted in prolonged PR interval. Compound double mutant mice using conditional Nkx2.5Cre mediated deletion resulted in sinus node dysfunction, a condition reminiscent of sick sinus syndrome in humans. Molecular analyses revealed that *SHOX2* and *TBX3* expression were impaired. Furthermore, RYR2-mediated calcium-leak was reported in *miR-106b-25* deficient mice^[190] since several members of this cluster, namely *miR-106b*, *miR-93* and *miR-25* directly target the 3'UTR of Ryr2. More recently, Lozano-Velasco et al^[149] demonstrate that a large number of microRNAs, previously reported to be associated to AF in patients, are also modulated by *PITX2*. In particular, *miR-1*, *miR-26b*, *miR-29a*, *miR-30e*, *miR-106b*, *miR-133* and *miR-200* were up-regulated in absence of *PITX2* whereas *miR-21*, *miR-106a*, *miR-203*, *miR-208a* and *miR-208b* were down-regulated. Several of these microRNAs, such as *miR-1*, *miR-133*, *miR-21*, *miR-106b* and *miR-26* have been previously reported to regulate calcium (*CACNA1C*^[191]; *RYR2*^[190]), sodium (*SCN5A*^[192]), potassium (*KCNJ2*^[193], *KCNE1*^[194], *KCNB2*^[194]), cation (*HCN4*^[195]) channel subunits, respectively. Overall, these

data demonstrate a highly complex gene regulatory networks leading to AF as summarized in Figure 3.

CONCLUSION

In this review we have highlighted the complexity of risk factors influencing the onset of AF, both clinically and genetically. Several lines of evidences demonstrated that *PITX2* exerts a pivotal role on the genetic determinants of AF^[136,139,140,149,185]. While it has been robustly demonstrated that *PITX2* insufficiency predisposes to atrial arrhythmia genesis in experimental mouse models^[136,139,140,149,185], discrepancies remain as whether *PITX2* is impaired in AF risk variants human patient carriers^[136,142,143]. Importantly some discrepancies also exist as whether predisposes or triggers AF^[136,139]. Our recent findings suggest the notion that *PITX2* insufficiency in the embryo predisposes whereas *PITX2* insufficiency in the adult atrial chamber triggers AF. In this context, we have provided evidences that such switching mechanisms seems to be regulated by WNT signaling and the downstream activation/repression of key microRNAs^[149,165], yet future experiments are needed to provide additional evidences on this front. In the last decade, our understanding of the downstream pathways controlled by *PITX2* has advanced at a quick pace. Several studies have demonstrated that *PITX2* controls signaling pathways regulation sinus node formation^[189,196], including therein the pivotal role of a microRNA cluster^[189,190]. These data shed light thus on the concurrence of AF with other cardiac physiopathological conditions such as sick sinus syndrome^[190]. Similarly we and others have demonstrated the key role exerted by *PITX2* regulating microRNAs impacting on calcium handling and thus on fibrillatory risk factors^[136,153,189]. It remains nonetheless to be established if *PITX2* also leads to impaired regulation of inflammatory processes and/or redox signaling which might impact on structural remodeling of the atrial chambers. To date, no evidences of atrial fibrosis have been reported in any of the *PITX2* deficient mouse models^[136,139,140,149,185], while some incipient evidences have been reported on the inflammatory link^[149] but not on redox signaling. Thus, future lines of research should be envisioned to clarify this point.

In addition, whereas our understanding of the *PITX2* downstream signaling pathways in the context of AF have progressively increased, scarce insights are currently available on the impact of AF clinically related risk factors on *PITX2*. A link between 4q25 risk variant carriers and increase left atrial volume has been recently reported^[197] in AF patients, however it remains to be established this is modulated by *PITX2*, as suggested by previous evidence in *PITX2* deficient mice in which atrial volume is already increased at fetal developmental stages^[136]. A recent study reported that aging and hypertension, two well-established risk factors for AF, severely decreased *PITX2* expression in a rat experimental model when both risk factors were combined^[198]. We recently reported that *Pitx2* is severely impaired in dilated cardiomyopathy patients as well as in an experimental heart failure pig model, shedding light

into a possible connection between ventricular dysfunction and AF^[199,200]. Yet, it remains to be established if other clinically relevant AF risk factors, such as diabetes, obesity, hyperthyroidism, valvular heart disease and/or obstructive sleep apnea are also impairing *PITX2* function and thus predisposing to AF. Overall these data demonstrate the pivotal role of *PITX2* regulating multiple aspect that if impaired are pro-arrhythmic and they also open new pathways to explore therapeutical approaches that could eventually lead to minimize the burden of AF in the human population.

REFERENCES

- 1 **Hakim FA**, Shen WK. Atrial fibrillation in the elderly: a review. *Future Cardiol* 2014; **10**: 745-758 [PMID: 25495816 DOI: 10.2217/fca.14.32]
- 2 **Nattel S**, Dobrev D. The multidimensional role of calcium in atrial fibrillation pathophysiology: mechanistic insights and therapeutic opportunities. *Eur Heart J* 2012; **33**: 1870-1877 [PMID: 22507975 DOI: 10.1093/euroheartj/ehs079]
- 3 **Nalliah CJ**, Sanders P, Kottkamp H, Kalman JM. The role of obesity in atrial fibrillation. *Eur Heart J* 2015; **pii:** ehv486 [PMID: 26371114]
- 4 **Elahi MM**, Flatman S, Matata BM. Tracing the origins of postoperative atrial fibrillation: the concept of oxidative stress-mediated myocardial injury phenomenon. *Eur J Cardiovasc Prev Rehabil* 2008; **15**: 735-741 [PMID: 19020458 DOI: 10.1097/HJR.0b013e328317f38a]
- 5 **Zhang Q**, Liu T, Ng CY, Li G. Diabetes mellitus and atrial remodeling: mechanisms and potential upstream therapies. *Cardiovasc Ther* 2014; **32**: 233-241 [PMID: 25065462 DOI: 10.1111/1755-5922.12089]
- 6 **Abed HS**, Wittert GA. Obesity and atrial fibrillation. *Obes Rev* 2013; **14**: 929-938 [PMID: 23879190 DOI: 10.1111/obr.12056]
- 7 **Vargas-Uricoechea H**, Sierra-Torres CH. Thyroid hormones and the heart. *Horm Mol Biol Clin Investig* 2014; **18**: 15-26 [PMID: 25389997 DOI: 10.1515/hmbci-2013-0059]
- 8 **Goudis CA**, Korantzopoulos P, Ntalas IV, Kallergis EM, Ketikoglou DG. Obesity and atrial fibrillation: A comprehensive review of the pathophysiological mechanisms and links. *J Cardiol* 2015; **66**: 361-369 [PMID: 25959929 DOI: 10.1016/j.jcc.2015.04.002]
- 9 **Goudis CA**, Korantzopoulos P, Ntalas IV, Kallergis EM, Liu T, Ketikoglou DG. Diabetes mellitus and atrial fibrillation: Pathophysiological mechanisms and potential upstream therapies. *Int J Cardiol* 2015; **184**: 617-622 [PMID: 25770841 DOI: 10.1016/j.ijcard.2015.03.052]
- 10 **De Caterina R**, Camm AJ. What is 'valvular' atrial fibrillation? A reappraisal. *Eur Heart J* 2014; **35**: 3328-3335 [PMID: 25265975 DOI: 10.1093/euroheartj/ehu352]
- 11 **Kumar KR**, Mandleywala SN, Link MS. Atrial and ventricular arrhythmias in hypertrophic cardiomyopathy. *Card Electrophysiol Clin* 2015; **7**: 173-186 [PMID: 26002384 DOI: 10.1016/j.ccep.2015.03.00]
- 12 **Riber LP**, Larsen TB, Christensen TD. Postoperative atrial fibrillation prophylaxis after lung surgery: systematic review and meta-analysis. *Ann Thorac Surg* 2014; **98**: 1989-1997 [PMID: 25283696 DOI: 10.1016/j.athoracsur.2014.06.069]
- 13 **Qaddoura A**, Kabali C, Drew D, van Oosten EM, Michael KA, Redfearn DP, Simpson CS, Baranchuk A. Obstructive sleep apnea as a predictor of atrial fibrillation after coronary artery bypass grafting: a systematic review and meta-analysis. *Can J Cardiol* 2014; **30**: 1516-1522 [PMID: 25475456 DOI: 10.1016/j.cjca.2014.10.014]
- 14 **Anumonwo JM**, Kalifa J. Risk factors and genetics of atrial fibrillation. *Cardiol Clin* 2014; **32**: 485-494 [PMID: 25443231 DOI: 10.1016/j.ccl.2014.07.007]
- 15 **Yadava M**, Hughey AB, Crawford TC. Postoperative atrial fibrillation: incidence, mechanisms, and clinical correlates. *Cardiol Clin* 2014; **32**: 627-636 [PMID: 25443241 DOI: 10.1016/j.ccl.2014.07.002]
- 16 **Akoum N**, Marrouche N. Assessment and impact of cardiac fibrosis on atrial fibrillation. *Curr Cardiol Rep* 2014; **16**: 518 [PMID: 24950676 DOI: 10.1007/s11886-014-0518]
- 17 **Berenfeld O**, Jalife J. Mechanisms of atrial fibrillation: rotors, ionic determinants, and excitation frequency. *Cardiol Clin* 2014; **32**: 495-506 [PMID: 25443232 DOI: 10.1016/j.ccl.2014.07.001]
- 18 **Heijman J**, Voigt N, Wehrens XH, Dobrev D. Calcium dysregulation in atrial fibrillation: the role of CaMKII. *Front Pharmacol* 2014; **5**: 30 [PMID: 24624086 DOI: 10.3389/fphar.2014.00030]
- 19 **Wolke C**, Bukowska A, Goette A, Lendeckel U. Redox control of cardiac remodeling in atrial fibrillation. *Biochim Biophys Acta* 2015; **1850**: 1555-1565 [PMID: 25513966 DOI: 10.1016/j.bbagen.2014.12.012]
- 20 **Jalife J**, Kaur K. Atrial remodeling, fibrosis, and atrial fibrillation. *Trends Cardiovasc Med* 2015; **25**: 475-484 [PMID: 25661032 DOI: 10.1016/j.tcm.2014.12.015]
- 21 **Corradi D**. Atrial fibrillation from the pathologist's perspective. *Cardiovasc Pathol* 2014; **23**: 71-84 [PMID: 24462196 DOI: 10.1016/j.carpath.2013.12.001]
- 22 **Dzeshka MS**, Lip GY, Snezhitskiy V, Shantsila E. Cardiac Fibrosis in Patients With Atrial Fibrillation: Mechanisms and Clinical Implications. *J Am Coll Cardiol* 2015; **66**: 943-959 [PMID: 26293766 DOI: 10.1016/j.jacc.2015.06.1313]
- 23 **Senoo K**, Lane D, Lip GY. Stroke and bleeding risk in atrial fibrillation. *Korean Circ J* 2014; **44**: 281-290 [PMID: 25278980 DOI: 10.4070/kcj.2014.44.5.281]
- 24 **Zimetbaum P**, Waks JW, Ellis ER, Glotzer TV, Passman RS. Role of atrial fibrillation burden in assessing thromboembolic risk. *Circ Arrhythm Electrophysiol* 2014; **7**: 1223-1229 [PMID: 25516580 DOI: 10.1161/CIRCEP.114.001356]
- 25 **Hirsh BJ**, Copeland-Halperin RS, Halperin JL. Fibrotic atrial cardiomyopathy, atrial fibrillation, and thromboembolism: mechanistic links and clinical inferences. *J Am Coll Cardiol* 2015; **65**: 2239-2251 [PMID: 25998669 DOI: 10.1016/j.jacc.2015.03.557]
- 26 **Hui DS**, Morley JE, Mikolajczak PC, Lee R. Atrial fibrillation: A major risk factor for cognitive decline. *Am Heart J* 2015; **169**: 448-456 [PMID: 25819850 DOI: 10.1016/j.ahj.2014.12.015]
- 27 **Jacobs V**, Cutler MJ, Day JD, Bunch TJ. Atrial fibrillation and dementia. *Trends Cardiovasc Med* 2015; **25**: 44-51 [PMID: 25442732]
- 28 **Luong C**, Barnes ME, Tsang TS. Atrial fibrillation and heart failure: cause or effect? *Curr Heart Fail Rep* 2014; **11**: 463-470 [PMID: 25258196 DOI: 10.1007/s11897-014-0229-1]
- 29 **Wijesurendra RS**, Casadei B. Atrial fibrillation: effects beyond the atrium? *Cardiovasc Res* 2015; **105**: 238-247 [PMID: 25587048 DOI: 10.1093/cvr/cvv001]
- 30 **Chen LY**, Benditt DG, Alonso A. Atrial fibrillation and its association with sudden cardiac death. *Circ J* 2014; **78**: 2588-2593 [PMID: 25262841]
- 31 **Potpata TS**, Lip GY. A brief history of 'lone' atrial fibrillation: from 'a peculiar pulse irregularity' to a modern public health concern. *Curr Pharm Des* 2015; **21**: 679-696 [PMID: 25269559]
- 32 **Andreasen L**, Nielsen JB, Olesen MS. Genetic aspects of lone atrial fibrillation: what do we know? *Curr Pharm Des* 2015; **21**: 667-678 [PMID: 25175087]
- 33 **Tello-Montoliu A**, Hernández-Romero D, Sanchez-Martínez M, Valdes M, Marín F. Lone atrial fibrillation - a diagnosis of exclusion. *Curr Pharm Des* 2015; **21**: 544-550 [PMID: 25175090]
- 34 **Andreasen L**, Nielsen JB, Christophersen IE, Holst AG, Sajadieh A, Tveit A, Haunso S, Svendsen JH, Schmitt N, Olesen MS. Genetic modifier of the QTc interval associated with early-onset atrial fibrillation. *Can J Cardiol* 2013; **29**: 1234-1240 [PMID: 24074973 DOI: 10.1016/j.cjca.2013.06.009]
- 35 **Brugada R**, Tapscott T, Czernuszewicz GZ, Marian AJ, Iglesias A, Mont L, Brugada J, Girona J, Domingo A, Bachinski LL, Roberts R. Identification of a genetic locus for familial atrial fibrillation. *N Engl J Med* 1997; **336**: 905-911 [PMID: 9070470]

- 36 **Chen YH**, Xu SJ, Bendahhou S, Wang XL, Wang Y, Xu WY, Jin HW, Sun H, Su XY, Zhuang QN, Yang YQ, Li YB, Liu Y, Xu HJ, Li XF, Ma N, Mou CP, Chen Z, Barhanin J, Huang W. KCNQ1 gain-of-function mutation in familial atrial fibrillation. *Science* 2003; **299**: 251-254 [PMID: 12522251]
- 37 **Xia M**, Jin Q, Bendahhou S, He Y, Larroque MM, Chen Y, Zhou Q, Yang Y, Liu Y, Liu B, Zhu Q, Zhou Y, Lin J, Liang B, Li L, Dong X, Pan Z, Wang R, Wan H, Qiu W, Xu W, Eurlings P, Barhanin J, Chen Y. A Kir2.1 gain-of-function mutation underlies familial atrial fibrillation. *Biochem Biophys Res Commun* 2005; **332**: 1012-1019 [PMID: 15922306]
- 38 **Yang Y**, Xia M, Jin Q, Bendahhou S, Shi J, Chen Y, Liang B, Lin J, Liu Y, Liu B, Zhou Q, Zhang D, Wang R, Ma N, Su X, Niu K, Pei Y, Xu W, Chen Z, Wan H, Cui J, Barhanin J, Chen Y. Identification of a KCNE2 gain-of-function mutation in patients with familial atrial fibrillation. *Am J Hum Genet* 2004; **75**: 899-905 [PMID: 15368194]
- 39 **Berenfeld O**, Zaitsev AV, Mironov SF, Pertsov AM, Jalife J. Frequency-dependent breakdown of wave propagation into fibrillatory conduction across the pectinate muscle network in the isolated sheep right atrium. *Circ Res* 2002; **90**: 1173-1180 [PMID: 12065320]
- 40 **Chen J**, Mandapati R, Berenfeld O, Skanes AC, Gray RA, Jalife J. Dynamics of wavelets and their role in atrial fibrillation in the isolated sheep heart. *Cardiovasc Res* 2000; **48**: 220-232 [PMID: 11054469]
- 41 **Mandapati R**, Skanes A, Chen J, Berenfeld O, Jalife J. Stable microreentrant sources as a mechanism of atrial fibrillation in the isolated sheep heart. *Circulation* 2000; **101**: 194-199 [PMID: 10637208]
- 42 **Skanes AC**, Mandapati R, Berenfeld O, Davidenko JM, Jalife J. Spatiotemporal periodicity during atrial fibrillation in the isolated sheep heart. *Circulation* 1998; **98**: 1236-1248 [PMID: 9743516]
- 43 **Tanaka K**, Zlochiver S, Vikstrom KL, Yamazaki M, Moreno J, Klos M, Zaitsev AV, Vaidyanathan R, Auerbach DS, Landas S, Guiraudon G, Jalife J, Berenfeld O, Kalifa J. Spatial distribution of fibrosis governs fibrillation wave dynamics in the posterior left atrium during heart failure. *Circ Res* 2007; **101**: 839-847 [PMID: 17704207]
- 44 **Gudbjartsson DF**, Arnar DO, Helgadottir A, Gretarsdottir S, Holm H, Sigurdsson A, Jonasdottir A, Baker A, Thorleifsson G, Kristjansson K, Palsson A, Blöndal T, Sulem P, Backman VM, Hardarson GA, Palsdottir E, Helgason A, Sigurjonsdottir R, Sverrisson JT, Kostulas K, Ng MC, Baum L, So WY, Wong KS, Chan JC, Furie KL, Greenberg SM, Sale M, Kelly P, MacRae CA, Smith EE, Rosand J, Hillert J, Ma RC, Ellinor PT, Thorgeirsson G, Gulcher JR, Kong A, Thorsteinsdottir U, Stefansson K. Variants conferring risk of atrial fibrillation on chromosome 4q25. *Nature* 2007; **448**: 353-357 [PMID: 17603472]
- 45 **Benjamin EJ**, Rice KM, Arking DE, Pfeifer A, van Noord C, Smith AV, Schnabel RB, Bis JC, Boerwinkle E, Sinner MF, Dehghan A, Lubitz SA, D'Agostino RB, Lumley T, Ehret GB, Heeringa J, Aspelund T, Newton-Cheh C, Larson MG, Marcante KD, Soliman EZ, Rivadeneira F, Wang TJ, Eiriksdottir G, Levy D, Psaty BM, Li M, Chamberlain AM, Hofman A, Vasan RS, Harris TB, Rotter JI, Kao WH, Agarwal SK, Stricker BH, Wang K, Launer LJ, Smith NL, Chakravarti A, Uitterlinden AG, Wolf PA, Sotoodehnia N, Köttgen A, van Duijn CM, Meitinger T, Mueller M, Perz S, Steinbeck G, Wichmann HE, Lunetta KL, Heckbert SR, Gudnason V, Alonso A, Kääb S, Ellinor PT, Witteman JC. Variants in ZFHX3 are associated with atrial fibrillation in individuals of European ancestry. *Nat Genet* 2009; **41**: 879-881 [PMID: 19597492 DOI: 10.1038/ng.416]
- 46 **Ellinor PT**, Lunetta KL, Albert CM, Glazer NL, Ritchie MD, Smith AV, Arking DE, Müller-Nurasyid M, Krijthe BP, Lubitz SA, Bis JC, Chung MK, Dörr M, Ozaki K, Roberts JD, Smith JG, Pfeifer A, Sinner MF, Lohman K, Ding J, Smith NL, Smith JD, Rienstra M, Rice KM, Van Wagoner DR, Magnani JW, Wakili R, Clauss S, Rotter JI, Steinbeck G, Launer LJ, Davies RW, Borkovich M, Harris TB, Lin H, Völker U, Völzke H, Milan DJ, Hofman A, Boerwinkle E, Chen LY, Soliman EZ, Voight BF, Li G, Chakravarti A, Kubo M, Tedrow UB, Rose LM, Ridker PM, Conen D, Tsunoda T, Furukawa T, Sotoodehnia N, Xu S, Kamatani N, Levy D, Nakamura Y, Parvez B, Mahida S, Furie KL, Rosand J, Muhammad R, Psaty BM, Meitinger T, Perz S, Wichmann HE, Witteman JC, Kao WH, Kathiresan S, Roden DM, Uitterlinden AG, Rivadeneira F, McKnight B, Sjögren M, Newman AB, Liu Y, Gollob MH, Melander O, Tanaka T, Stricker BH, Felix SB, Alonso A, Darbar D, Barnard J, Chasman DI, Heckbert SR, Benjamin EJ, Gudnason V, Kääb S. Meta-analysis identifies six new susceptibility loci for atrial fibrillation. *Nat Genet* 2012; **44**: 670-675 [PMID: 22544366 DOI: 10.1038/ng.2261]
- 47 **Ellinor PT**, Lunetta KL, Glazer NL, Pfeifer A, Alonso A, Chung MK, Sinner MF, de Bakker PI, Mueller M, Lubitz SA, Fox E, Darbar D, Smith NL, Smith JD, Schnabel RB, Soliman EZ, Rice KM, Van Wagoner DR, Beckmann BM, van Noord C, Wang K, Ehret GB, Rotter JI, Hazen SL, Steinbeck G, Smith AV, Launer LJ, Harris TB, Makino S, Nelis M, Milan DJ, Perz S, Esko T, Köttgen A, Moebus S, Newton-Cheh C, Li M, Möhlenkamp S, Wang TJ, Kao WH, Vasan RS, Nöthen MM, MacRae CA, Stricker BH, Hofman A, Uitterlinden AG, Levy D, Boerwinkle E, Metspalu A, Topol EJ, Chakravarti A, Gudnason V, Psaty BM, Roden DM, Meitinger T, Wichmann HE, Witteman JC, Barnard J, Arking DE, Benjamin EJ, Heckbert SR, Kääb S. Common variants in KCNN3 are associated with lone atrial fibrillation. *Nat Genet* 2010; **42**: 240-244 [PMID: 20173747 DOI: 10.1038/ng.537]
- 48 **Gudbjartsson DF**, Holm H, Gretarsdottir S, Thorleifsson G, Walters GB, Thorgeirsson G, Gulcher J, Mathiesen EB, Njølstad I, Nyrnes A, Wilsgaard T, Hald EM, Hveem K, Stoltenberg C, Kucera G, Stubblefield T, Carter S, Roden D, Ng MC, Baum L, So WY, Wong KS, Chan JC, Gieger C, Wichmann HE, Gschwendtner A, Dichgans M, Kuhlenbäumer G, Berger K, Ringelstein EB, Bevan S, Markus HS, Kostulas K, Hillert J, Sveinbjörnsdóttir S, Valdimarsson EM, Löchen ML, Ma RC, Darbar D, Kong A, Arnar DO, Thorsteinsdottir U, Stefansson K. A sequence variant in ZFHX3 on 16q22 associates with atrial fibrillation and ischemic stroke. *Nat Genet* 2009; **41**: 876-878 [PMID: 19597491 DOI: 10.1038/ng.417]
- 49 **Schnabel RB**, Kerr KF, Lubitz SA, Alkylbekova EL, Marcus GM, Sinner MF, Magnani JW, Wolf PA, Deo R, Lloyd-Jones DM, Lunetta KL, Mehra R, Levy D, Fox ER, Arking DE, Mosley TH, Müller-Nurasyid M, Young TR, Wichmann HE, Seshadri S, Farlow DN, Rotter JI, Soliman EZ, Glazer NL, Wilson JG, Breteler MM, Sotoodehnia N, Newton-Cheh C, Kääb S, Ellinor PT, Alonso A, Benjamin EJ, Heckbert SR. Large-scale candidate gene analysis in whites and African Americans identifies IL6R polymorphism in relation to atrial fibrillation: the National Heart, Lung, and Blood Institute's Candidate Gene Association Resource (CARE) project. *Circ Cardiovasc Genet* 2011; **4**: 557-564 [PMID: 21846873 DOI: 10.1161/CIRGENETICS.110.959197]
- 50 **Amin AS**, Tan HL, Wilde AA. Cardiac ion channels in health and disease. *Heart Rhythm* 2010; **7**: 117-126 [PMID: 19875343 DOI: 10.1016/j.hrthm.2009.08.005]
- 51 **Amin AS**, Bhuiyan ZA. SCN5A mutations in atrial fibrillation. *Heart Rhythm* 2010; **7**: 1870-1871 [PMID: 20850563 DOI: 10.1016/j.hrthm.2010.09.012]
- 52 **Blana A**, Kaese S, Fortmüller L, Laakmann S, Damke D, van Bragt K, Eckstein J, Piccini I, Kirchhefer U, Nattel S, Breithardt G, Carmeliet P, Carmeliet E, Schotten U, Verheule S, Kirchhof P, Fabritz L. Knock-in gain-of-function sodium channel mutation prolongs atrial action potentials and alters atrial vulnerability. *Heart Rhythm* 2010; **7**: 1862-1869 [PMID: 20728579 DOI: 10.1016/j.hrthm.2010.08.016]
- 53 **Chen S**, Wang C, Wang X, Xu C, Wu M, Wang P, Tu X, Wang QK. Significant Association Between CAV1 Variant rs3807989 on 7p31 and Atrial Fibrillation in a Chinese Han Population. *J Am Heart Assoc* 2015; **4**: pii: e001980 [PMID: 25953654 DOI: 10.1161/JAHA.115.001980]
- 54 **Laitinen-Forsblom PJ**, Mäkinen P, Mäkinen H, Yli-Mäyry S, Virtanen V, Kontula K, Aalto-Setälä K. SCN5A mutation

- associated with cardiac conduction defect and atrial arrhythmias. *J Cardiovasc Electrophysiol* 2006; **17**: 480-485 [PMID: 16684018]
- 55 **Li Q**, Huang H, Liu G, Lam K, Rutberg J, Green MS, Birnie DH, Lemery R, Chahine M, Gollobo MH. Gain-of-function mutation of Nav1.5 in atrial fibrillation enhances cellular excitability and lowers the threshold for action potential firing. *Biochem Biophys Res Commun* 2009; **380**: 132-137 [PMID: 19167345 DOI: 10.1016/j.bbrc.2009.01.052]
- 56 **Makiyama T**, Akao M, Shizuta S, Doi T, Nishiyama K, Oka Y, Ohno S, Nishio Y, Tsuji K, Itoh H, Kimura T, Kita T, Horie M. A novel SCN5A gain-of-function mutation M1875T associated with familial atrial fibrillation. *J Am Coll Cardiol* 2008; **52**: 1326-1334 [PMID: 18929244 DOI: 10.1016/j.jacc.2008.07.013]
- 57 **Benito B**, Brugada R, Perich RM, Lizotte E, Cinca J, Mont L, Berrueto A, Tolosana JM, Freixa X, Brugada P, Brugada J. A mutation in the sodium channel is responsible for the association of long QT syndrome and familial atrial fibrillation. *Heart Rhythm* 2008; **5**: 1434-1440 [PMID: 18929331 DOI: 10.1016/j.hrthm.2008.07.013]
- 58 **Calloe K**, Schmitt N, Grubb S, Pfeiffer R, David JP, Kanter R, Cordeiro JM, Antzelevitch C. Multiple arrhythmic syndromes in a newborn, owing to a novel mutation in SCN5A. *Can J Physiol Pharmacol* 2011; **89**: 723-736 [PMID: 21895525 DOI: 10.1139/y11-070]
- 59 **Dolz-Gaitón P**, Núñez M, Núñez L, Barana A, Amorós I, Matamoros M, Pérez-Hernández M, González de la Fuente M, Alvarez-López M, Macías-Ruiz R, Tercedor-Sánchez L, Jiménez-Jáimez J, Delpón E, Caballero R, Tamargo J. Functional characterization of a novel frameshift mutation in the C-terminus of the Nav1.5 channel underlying a Brugada syndrome with variable expression in a Spanish family. *PLoS One* 2013; **8**: e81493 [PMID: 24363796 DOI: 10.1371/journal.pone.0081493]
- 60 **Olson TM**, Michels VV, Ballew JD, Reyna SP, Karst ML, Herron KJ, Horton SC, Rodeheffer RJ, Anderson JL. Sodium channel mutations and susceptibility to heart failure and atrial fibrillation. *JAMA* 2005; **293**: 447-454 [PMID: 15671429]
- 61 **Ziyadeh-Isleem A**, Clatot J, Duchatelet S, Gandjbakhch E, Denjoy I, Hidden-Lucet F, Hatem S, Deschênes I, Coulombe A, Neyroud N, Guicheney P. A truncating SCN5A mutation combined with genetic variability causes sick sinus syndrome and early atrial fibrillation. *Heart Rhythm* 2014; **11**: 1015-1023 [PMID: 24582607 DOI: 10.1016/j.hrthm.2014.02.021]
- 62 **Rossenbacher T**, Carroll SJ, Liu H, Kuipéri C, de Ravel TJ, Devriendt K, Carmeliet P, Kass RS, Heidbüchel H. Novel pore mutation in SCN5A manifests as a spectrum of phenotypes ranging from atrial flutter, conduction disease, and Brugada syndrome to sudden cardiac death. *Heart Rhythm* 2004; **1**: 610-615 [PMID: 15851228]
- 63 **Watanabe H**, Darbar D, Kaiser DW, Jiramongkolchai K, Chopra S, Donahue BS, Kannankeril PJ, Roden DM. Mutations in sodium channel $\beta 1$ - and $\beta 2$ -subunits associated with atrial fibrillation. *Circ Arrhythm Electrophysiol* 2009; **2**: 268-275 [PMID: 19808477 DOI: 10.1161/CIRCEP.108.779181]
- 64 **Wang P**, Yang Q, Wu X, Yang Y, Shi L, Wang C, Wu G, Xia Y, Yang B, Zhang R, Xu C, Cheng X, Li S, Zhao Y, Fu F, Liao Y, Fang F, Chen Q, Tu X, Wang QK. Functional dominant-negative mutation of sodium channel subunit gene SCN3B associated with atrial fibrillation in a Chinese GenEID population. *Biochem Biophys Res Commun* 2010; **398**: 98-104 [PMID: 20558140 DOI: 10.1016/j.bbrc.2010.06.042]
- 65 **Olesen MS**, Jespersen T, Nielsen JB, Liang B, Møller DV, Hedley P, Christiansen M, Varró A, Olesen SP, Haunso S, Schmitt N, Svendsen JH. Mutations in sodium channel β -subunit SCN3B are associated with early-onset lone atrial fibrillation. *Cardiovasc Res* 2011; **89**: 786-793 [PMID: 21051419 DOI: 10.1093/cvr/cvq348]
- 66 **Li RG**, Wang Q, Xu YJ, Zhang M, Qu XK, Liu X, Fang WY, Yang YQ. Mutations of the SCN4B-encoded sodium channel $\beta 4$ subunit in familial atrial fibrillation. *Int J Mol Med* 2013; **32**: 144-150 [PMID: 23604097 DOI: 10.3892/ijmm.2013.1355]
- 67 **Olesen MS**, Holst AG, Svendsen JH, Haunso S, Tfelt-Hansen J, SCN1Bb R214Q found in 3 patients: 1 with Brugada syndrome and 2 with lone atrial fibrillation. *Heart Rhythm* 2012; **9**: 770-773 [PMID: 22155598 DOI: 10.1016/j.hrthm.2011.12.005]
- 68 **Remme CA**, Wilde AA, Bezzina CR. Cardiac sodium channel overlap syndromes: different faces of SCN5A mutations. *Trends Cardiovasc Med* 2008; **18**: 78-87 [PMID: 18436145 DOI: 10.1016/j.tcm.2008.01.002]
- 69 **Rook MB**, Evers MM, Vos MA, Bierhuizen MF. Biology of cardiac sodium channel Nav1.5 expression. *Cardiovasc Res* 2012; **93**: 12-23 [PMID: 21937582 DOI: 10.1093/cvr/cvr252]
- 70 **Veerman CC**, Wilde AA, Lodder EM. The cardiac sodium channel gene SCN5A and its gene product NaV1.5: Role in physiology and pathophysiology. *Gene* 2015; **573**: 177-187 [PMID: 26361848 DOI: 10.1016/j.gene.2015.08.062]
- 71 **Macri V**, Mahida SN, Zhang ML, Sinner MF, Dolmatova EV, Tucker NR, McLellan M, Shea MA, Milan DJ, Lunetta KL, Benjamin EJ, Ellinor PT. A novel trafficking-defective HCN4 mutation is associated with early-onset atrial fibrillation. *Heart Rhythm* 2014; **11**: 1055-1062 [PMID: 24607718 DOI: 10.1016/j.hrthm.2014.03.002]
- 72 **Hong K**, Piper DR, Diaz-Valdecantos A, Brugada J, Oliva A, Burashnikov E, Santos-de-Soto J, Grueso-Montero J, Diaz-Enfante E, Brugada P, Sachse F, Sanguineti MC, Brugada R. De novo KCNQ1 mutation responsible for atrial fibrillation and short QT syndrome in utero. *Cardiovasc Res* 2005; **68**: 433-440 [PMID: 16109388]
- 73 **Lundby A**, Ravn LS, Svendsen JH, Olesen SP, Schmitt N. KCNQ1 mutation Q147R is associated with atrial fibrillation and prolonged QT interval. *Heart Rhythm* 2007; **4**: 1532-1541 [PMID: 17997361]
- 74 **Kharche S**, Adeniran I, Stott J, Law P, Boyett MR, Hancox JC, Zhang H. Pro-arrhythmogenic effects of the S140G KCNQ1 mutation in human atrial fibrillation - insights from modelling. *J Physiol* 2012; **590**: 4501-4514 [PMID: 22508963 DOI: 10.1113/jphysiol.2012.229146]
- 75 **Das S**, Makino S, Melman YF, Shea MA, Goyal SB, Rosenzweig A, Macrae CA, Ellinor PT. Mutation in the S3 segment of KCNQ1 results in familial lone atrial fibrillation. *Heart Rhythm* 2009; **6**: 1146-1153 [PMID: 19632626 DOI: 10.1016/j.hrthm.2009.04.015]
- 76 **El Harchi A**, Zhang H, Hancox JC. The S140G KCNQ1 atrial fibrillation mutation affects 'I(KS)' profile during both atrial and ventricular action potentials. *J Physiol Pharmacol* 2010; **61**: 759-764 [PMID: 21224508]
- 77 **Bartos DC**, Duchatelet S, Burgess DE, Klug D, Denjoy I, Peat R, Lupoglazoff JM, Fressart V, Berthet M, Ackerman MJ, January CT, Guicheney P, Delisle BP. R231C mutation in KCNQ1 causes long QT syndrome type 1 and familial atrial fibrillation. *Heart Rhythm* 2011; **8**: 48-55 [PMID: 20850564 DOI: 10.1016/j.hrthm.2010.09.010]
- 78 **Hasegawa K**, Ohno S, Ashihara T, Itoh H, Ding WG, Toyoda F, Makiyama T, Aoki H, Nakamura Y, Delisle BP, Matsuura H, Horie M. A novel KCNQ1 missense mutation identified in a patient with juvenile-onset atrial fibrillation causes constitutively open IKs channels. *Heart Rhythm* 2014; **11**: 67-75 [PMID: 24096004 DOI: 10.1016/j.hrthm.2013.09.073]
- 79 **Ki CS**, Jung CL, Kim HJ, Baek KH, Park SJ, On YK, Kim KS, Noh SJ, Youm JB, Kim JS, Cho H. A KCNQ1 mutation causes age-dependant bradycardia and persistent atrial fibrillation. *Pflugers Arch* 2014; **466**: 529-540 [PMID: 23989646 DOI: 10.1007/s00424-013-1337-6]
- 80 **Ravn LS**, Aizawa Y, Pollevick GD, Hofman-Bang J, Cordeiro JM, Dixen U, Jensen G, Wu Y, Burashnikov E, Haunso S, Guerchicoff A, Hu D, Svendsen JH, Christiansen M, Antzelevitch C. Gain of function in IKs secondary to a mutation in KCNE5 associated with atrial fibrillation. *Heart Rhythm* 2008; **5**: 427-435 [PMID: 18313602 DOI: 10.1016/j.hrthm.2007.12.019]
- 81 **Nielsen JB**, Bentzen BH, Olesen MS, David JP, Olesen SP, Haunso S, Svendsen JH, Schmitt N. Gain-of-function mutations in potassium channel subunit KCNE2 associated with early-onset lone atrial fibrillation. *Biomark Med* 2014; **8**: 557-570 [PMID: 24796621 DOI: 10.2217/bmm.13.137]

- 82 **Deo M**, Ruan Y, Pandit SV, Shah K, Berenfeld O, Blaufox A, Cerrone M, Noujaim SF, Denegri M, Jalife J, Priori SG. KCNJ2 mutation in short QT syndrome 3 results in atrial fibrillation and ventricular proarrhythmia. *Proc Natl Acad Sci USA* 2013; **110**: 4291-4296 [PMID: 23440193 DOI: 10.1073/pnas.1218154110]
- 83 **Kharche S**, Garratt CJ, Boyett MR, Inada S, Holden AV, Hancox JC, Zhang H. Atrial proarrhythmia due to increased inward rectifier current (I(K1)) arising from KCNJ2 mutation--a simulation study. *Prog Biophys Mol Biol* 2008; **98**: 186-197 [PMID: 19041665 DOI: 10.1016/j.pbiomolbio.2008.10.010]
- 84 **Villafañe J**, Fischbach P, Gebauer R. Short QT syndrome manifesting with neonatal atrial fibrillation and bradycardia. *Cardiology* 2014; **128**: 236-240 [PMID: 24818999 DOI: 10.1159/000360758]
- 85 **Chelu MG**, Sarma S, Sood S, Wang S, van Oort RJ, Skapura DG, Li N, Santonastasi M, Müller FU, Schmitz W, Schotten U, Anderson ME, Valderrábano M, Dobrev D, Wehrens XH. Calmodulin kinase II-mediated sarcoplasmic reticulum Ca²⁺ leak promotes atrial fibrillation in mice. *J Clin Invest* 2009; **119**: 1940-1951 [PMID: 19603549]
- 86 **Shan J**, Xie W, Betzenhauser M, Reiken S, Chen BX, Wronski A, Marks AR. Calcium leak through ryanodine receptors leads to atrial fibrillation in 3 mouse models of catecholaminergic polymorphic ventricular tachycardia. *Circ Res* 2012; **111**: 708-717 [PMID: 22828895 DOI: 10.1161/CIRCRESAHA.112.273342]
- 87 **Li N**, Wang T, Wang W, Cutler MJ, Wang Q, Voigt N, Rosenbaum DS, Dobrev D, Wehrens XH. Inhibition of CaMKII phosphorylation of RyR2 prevents induction of atrial fibrillation in FKBP12.6 knockout mice. *Circ Res* 2012; **110**: 465-470 [PMID: 22158709 DOI: 10.1161/CIRCRESAHA.111.253229]
- 88 **Zhang Y**, Matthews GD, Lei M, Huang CL. Abnormal Ca(2+) homeostasis, atrial arrhythmogenesis, and sinus node dysfunction in murine hearts modeling RyR2 modification. *Front Physiol* 2013; **4**: 150 [PMID: 23805105 DOI: 10.3389/fphys.2013.00150]
- 89 **Beavers DL**, Wang W, Ather S, Voigt N, Garbino A, Dixit SS, Landstrom AP, Li N, Wang Q, Olivotto I, Dobrev D, Ackerman MJ, Wehrens XH. Mutation E169K in junctophilin-2 causes atrial fibrillation due to impaired RyR2 stabilization. *J Am Coll Cardiol* 2013; **62**: 2010-2019 [PMID: 23973696 DOI: 10.1016/j.jacc.2013.06.052]
- 90 **Chiang DY**, Li N, Wang Q, Alsina KM, Quick AP, Reynolds JO, Wang G, Skapura D, Voigt N, Dobrev D, Wehrens XH. Impaired local regulation of ryanodine receptor type 2 by protein phosphatase 1 promotes atrial fibrillation. *Cardiovasc Res* 2014; **103**: 178-187 [PMID: 24812280 DOI: 10.1093/cvr/cvu123]
- 91 **Di Pino A**, Caruso E, Costanzo L, Guccione P. A novel RyR2 mutation in a 2-year-old baby presenting with atrial fibrillation, atrial flutter, and atrial ectopic tachycardia. *Heart Rhythm* 2014; **11**: 1480-1483 [PMID: 24793461 DOI: 10.1016/j.hrthm.2014.04.037]
- 92 **Zhabayev P**, Hiess F, Wang R, Liu Y, Wayne Chen SR, Oudit GY. S4153R is a gain-of-function mutation in the cardiac Ca(2+) release channel ryanodine receptor associated with catecholaminergic polymorphic ventricular tachycardia and paroxysmal atrial fibrillation. *Can J Cardiol* 2013; **29**: 993-996 [PMID: 23498838 DOI: 10.1016/j.cjca.2012.12.019]
- 93 **Kazemian P**, Gollob MH, Pantano A, Oudit GY. A novel mutation in the RYR2 gene leading to catecholaminergic polymorphic ventricular tachycardia and paroxysmal atrial fibrillation: dose-dependent arrhythmia-event suppression by β-blocker therapy. *Can J Cardiol* 2011; **27**: 870.e7-870.e10 [PMID: 21652165 DOI: 10.1016/j.cjca.2011.02.003]
- 94 **King JH**, Wickramarachchi C, Kua K, Du Y, Jeevaratnam K, Matthews HR, Grace AA, Huang CL, Fraser JA. Loss of Nav1.5 expression and function in murine atria containing the RyR2-P2328S gain-of-function mutation. *Cardiovasc Res* 2013; **99**: 751-759 [PMID: 23723061 DOI: 10.1093/cvr/cvt141]
- 95 **Zhang Y**, Fraser JA, Jeevaratnam K, Hao X, Hothi SS, Grace AA, Lei M, Huang CL. Acute atrial arrhythmogenicity and altered Ca(2+) homeostasis in murine RyR2-P2328S hearts. *Cardiovasc Res* 2011; **89**: 794-804 [PMID: 20621925 DOI: 10.1093/cvr/cvq229]
- 96 **Thibodeau IL**, Xu J, Li Q, Liu G, Lam K, Veinot JP, Birnie DH, Jones DL, Krahn AD, Lemery R, Nicholson BJ, Gollob MH. Paradigm of genetic mosaicism and lone atrial fibrillation: physiological characterization of a connexin 43-deletion mutant identified from atrial tissue. *Circulation* 2010; **122**: 236-244 [PMID: 20606116 DOI: 10.1161/CIRCULATIONAHA.110.961227]
- 97 **Tuomi JM**, Tyml K, Jones DL. Atrial tachycardia/fibrillation in the connexin 43 G60S mutant (Oculodentodigital dysplasia) mouse. *Am J Physiol Heart Circ Physiol* 2011; **300**: H1402-H1411 [PMID: 21239638 DOI: 10.1152/ajpheart.01094.2010]
- 98 **Delmar M**, Makita N. Cardiac connexins, mutations and arrhythmias. *Curr Opin Cardiol* 2012; **27**: 236-241 [PMID: 22382502 DOI: 10.1097/HCO.0b013e328352220e]
- 99 **Gollob MH**, Jones DL, Krahn AD, Danis L, Gong XQ, Shao Q, Liu X, Veinot JP, Tang AS, Stewart AF, Tesson F, Klein GJ, Yee R, Skanes AC, Guiraudon GM, Ebihara L, Bai D. Somatic mutations in the connexin 40 gene (GJA5) in atrial fibrillation. *N Engl J Med* 2006; **354**: 2677-2688 [PMID: 16790700]
- 100 **Yang YQ**, Liu X, Zhang XL, Wang XH, Tan HW, Shi HF, Jiang WF, Fang WY. Novel connexin40 missense mutations in patients with familial atrial fibrillation. *Europace* 2010; **12**: 1421-1427 [PMID: 20650941 DOI: 10.1093/europace/euq274]
- 101 **Yang YQ**, Zhang XL, Wang XH, Tan HW, Shi HF, Jiang WF, Fang WY, Liu X. Connexin40 nonsense mutation in familial atrial fibrillation. *Int J Mol Med* 2010; **26**: 605-610 [PMID: 20818502]
- 102 **Christophersen IE**, Holmegård HN, Jabbari J, Sajadieh A, Haunø S, Tveit A, Svendsen JH, Olesen MS. Rare variants in GJA5 are associated with early-onset lone atrial fibrillation. *Can J Cardiol* 2013; **29**: 111-116 [PMID: 23040431 DOI: 10.1016/j.cjca.2012.08.002]
- 103 **Shi HF**, Yang JF, Wang Q, Li RG, Xu YJ, Qu XK, Fang WY, Liu X, Yang YQ. Prevalence and spectrum of GJA5 mutations associated with lone atrial fibrillation. *Mol Med Rep* 2013; **7**: 767-774 [PMID: 23292621 DOI: 10.3892/mmr.2012.1252]
- 104 **Sun Y**, Yang YQ, Gong XQ, Wang XH, Li RG, Tan HW, Liu X, Fang WY, Bai D. Novel germline GJA5/connexin40 mutations associated with lone atrial fibrillation impair gap junctional intercellular communication. *Hum Mutat* 2013; **34**: 603-609 [PMID: 23348765 DOI: 10.1002/humu.22278]
- 105 **Roberts JD**, Longoria J, Poon A, Gollob MH, Dewland TA, Kwok PY, Olglin JE, Deo RC, Marcus GM. Targeted deep sequencing reveals no definitive evidence for somatic mosaicism in atrial fibrillation. *Circ Cardiovasc Genet* 2015; **8**: 50-57 [PMID: 25406240 DOI: 10.1161/CIRCGENETICS.114.000650]
- 106 **Gemel J**, Simon AR, Patel D, Xu Q, Matiukas A, Veenstra RD, Beyer EC. Degradation of a connexin40 mutant linked to atrial fibrillation is accelerated. *J Mol Cell Cardiol* 2014; **74**: 330-339 [PMID: 24973497 DOI: 10.1016/j.yjmcc.2014.06.010]
- 107 **Sun Y**, Hills MD, Ye WG, Tong X, Bai D. Atrial fibrillation-linked germline GJA5/connexin40 mutants showed an increased hemichannel function. *PLoS One* 2014; **9**: e95125 [PMID: 24733048 DOI: 10.1371/journal.pone.0095125]
- 108 **Sun Y**, Tong X, Chen H, Huang T, Shao Q, Huang W, Laird DW, Bai D. An atrial-fibrillation-linked connexin40 mutant is retained in the endoplasmic reticulum and impairs the function of atrial gap-junction channels. *Dis Model Mech* 2014; **7**: 561-569 [PMID: 24626989 DOI: 10.1242/dmm.013813]
- 109 **Patel D**, Gemel J, Xu Q, Simon AR, Lin X, Matiukas A, Beyer EC, Veenstra RD. Atrial fibrillation-associated connexin40 mutants make hemichannels and synergistically form gap junction channels with novel properties. *FEBS Lett* 2014; **588**: 1458-1464 [PMID: 24457199 DOI: 10.1016/j.febslet.2014.01.010]
- 110 **Bai D**. Atrial fibrillation-linked GJA5/connexin40 mutants impaired gap junctions via different mechanisms. *FEBS Lett* 2014; **588**: 1238-1243 [PMID: 24656738 DOI: 10.1016/j.febslet.2014.02.064]
- 111 **Lübkemeier I**, Andrié R, Lickfett L, Bosen F, Stöckigt F, Dobrowolski R, Draftehn AM, Fregeac J, Schultze JL, Bukauskas FF, Schrickel JW, Willecke K. The Connexin40A96S mutation from a patient with atrial fibrillation causes decreased atrial conduction velocities and sustained episodes of induced atrial

- fibrillation in mice. *J Mol Cell Cardiol* 2013; **65**: 19-32 [PMID: 24060583 DOI: 10.1016/j.yjmcc.2013.09.008]
- 112 **Postma AV**, van de Meerakker JB, Mathijssen IB, Barnett P, Christoffels VM, Ilgun A, Lam J, Wilde AA, Lekanne Deprez RH, Moorman AF. A gain-of-function TBX5 mutation is associated with atypical Holt-Oram syndrome and paroxysmal atrial fibrillation. *Circ Res* 2008; **102**: 1433-1442 [PMID: 18451335 DOI: 10.1161/CIRCRESAHA.107.168294]
- 113 **Gutierrez-Roelens I**, De Roy L, Ovaert C, Sluysmans T, Devriendt K, Brunner HG, Viikula M. A novel CSX/NKX2-5 mutation causes autosomal-dominant AV block: are atrial fibrillation and syncopes part of the phenotype? *Eur J Hum Genet* 2006; **14**: 1313-1316 [PMID: 16896344]
- 114 **Huang RT**, Xue S, Xu YJ, Zhou M, Yang YQ. A novel NKX2.5 loss-of-function mutation responsible for familial atrial fibrillation. *Int J Mol Med* 2013; **31**: 1119-1126 [PMID: 23525379 DOI: 10.3892/ijmm.2013.1316]
- 115 **Xie WH**, Chang C, Xu YJ, Li RG, Qu XK, Fang WY, Liu X, Yang YQ. Prevalence and spectrum of Nkx2.5 mutations associated with idiopathic atrial fibrillation. *Clinics (Sao Paulo)* 2013; **68**: 777-784 [PMID: 23778487 DOI: 10.6061/clinics/2013(06)09]
- 116 **Yu H**, Xu JH, Song HM, Zhao L, Xu WJ, Wang J, Li RG, Xu L, Jiang WF, Qiu XB, Jiang JQ, Qu XK, Liu X, Fang WY, Jiang JF, Yang YQ. Mutational spectrum of the NKX2-5 gene in patients with lone atrial fibrillation. *Int J Med Sci* 2014; **11**: 554-563 [PMID: 24782644 DOI: 10.7150/ijms.8407]
- 117 **Yuan F**, Qiu XB, Li RG, Qu XK, Wang J, Xu YJ, Liu X, Fang WY, Yang YQ, Liao DN. A novel NKX2-5 loss-of-function mutation predisposes to familial dilated cardiomyopathy and arrhythmias. *Int J Mol Med* 2015; **35**: 478-486 [PMID: 25503402 DOI: 10.3892/ijmm.2014.2029]
- 118 **Wang J**, Zhang DF, Sun YM, Li RG, Qiu XB, Qu XK, Liu X, Fang WY, Yang YQ. NKX2-6 mutation predisposes to familial atrial fibrillation. *Int J Mol Med* 2014; **34**: 1581-1590 [PMID: 25319568 DOI: 10.3892/ijmm.2014.1971]
- 119 **Zhang X**, Chen S, Yoo S, Chakrabarti S, Zhang T, Ke T, Oberti C, Yong SL, Fang F, Li L, de la Fuente R, Wang L, Chen Q, Wang QK. Mutation in nuclear pore component NUP155 leads to atrial fibrillation and early sudden cardiac death. *Cell* 2008; **135**: 1017-1027 [PMID: 19070573 DOI: 10.1016/j.cell.2008.10.022]
- 120 **Hua R**, MacLeod SL, Polina I, Moghtadaei M, Jansen HJ, Bogachev O, O'Bleness SB, Sapp JL, Legare JF, Rose RA. Effects of Wild-Type and Mutant Forms of Atrial Natriuretic Peptide on Atrial Electrophysiology and Arrhythmogenesis. *Circ Arrhythm Electrophysiol* 2015; **8**: 1240-1254 [PMID: 26227000]
- 121 **Li QY**, Newbury-Ecob RA, Terrett JA, Wilson DI, Curtis AR, Yi CH, Gebuhr T, Bullen PJ, Robson SC, Strachan T, Bonnet D, Lyonnet S, Young ID, Raeburn JA, Buckler AJ, Law DJ, Brook JD. Holt-Oram syndrome is caused by mutations in TBX5, a member of the Brachyury (T) gene family. *Nat Genet* 1997; **15**: 21-29 [PMID: 8988164]
- 122 **Costa MW**, Guo G, Wolstein O, Vale M, Castro ML, Wang L, Otway R, Riek P, Cochrane N, Furtado M, Semsarian C, Weintraub RG, Yeoh T, Hayward C, Keogh A, Macdonald P, Feneley M, Graham RM, Seidman JG, Seidman CE, Rosenthal N, Fatkin D, Harvey RP. Functional characterization of a novel mutation in NKX2-5 associated with congenital heart disease and adult-onset cardiomyopathy. *Circ Cardiovasc Genet* 2013; **6**: 238-247 [PMID: 23661673 DOI: 10.1161/CIRCGENETICS.113.000057]
- 123 **Qu XK**, Qiu XB, Yuan F, Wang J, Zhao CM, Liu XY, Zhang XL, Li RG, Xu YJ, Hou XM, Fang WY, Liu X, Yang YQ. A novel NKX2.5 loss-of-function mutation associated with congenital bicuspid aortic valve. *Am J Cardiol* 2014; **114**: 1891-1895 [PMID: 25438918 DOI: 10.1016/j.amjcard.2014.09.028]
- 124 **Wang J**, Mao JH, Ding KK, Xu WJ, Liu XY, Qiu XB, Li RG, Qu XK, Xu YJ, Huang RT, Xue S, Yang YQ. A novel NKX2.6 mutation associated with congenital ventricular septal defect. *Pediatr Cardiol* 2015; **36**: 646-656 [PMID: 25380965 DOI: 10.1007/s00246-014-1060-x]
- 125 **Lin Y**, Guo X, Zhao B, Liu J, Da M, Wen Y, Hu Y, Ni B, Zhang K, Yang S, Xu J, Dai J, Wang X, Xia Y, Ma H, Jin G, Yu S, Liu J, Keavney BD, Goodship JA, Cordell HJ, Wang X, Shen H, Sha J, Zhou Z, Chen Y, Mo X, Luo L, Hu Z. Association analysis identifies new risk loci for congenital heart disease in Chinese populations. *Nat Commun* 2015; **6**: 8082 [PMID: 26283027 DOI: 10.1038/ncomms9082]
- 126 **Chowdhury R**, Ashraf H, Melanson M, Tanada Y, Nguyen M, Silberbach M, Wakimoto H, Benson DW, Anderson RH, Kasahara H. Mouse Model of Human Congenital Heart Disease: Progressive Atrioventricular Block Induced by a Heterozygous Nkx2-5 Homeodomain Missense Mutation. *Circ Arrhythm Electrophysiol* 2015; **8**: 1255-1264 [PMID: 26226998]
- 127 **Zhou W**, Zhao L, Jiang JQ, Jiang WF, Yang YQ, Qiu XB. A novel TBX5 loss-of-function mutation associated with sporadic dilated cardiomyopathy. *Int J Mol Med* 2015; **36**: 282-288 [PMID: 25963046 DOI: 10.3892/ijmm.2015.2206]
- 128 **Zhao L**, Ni SH, Liu XY, Wei D, Yuan F, Xu L, Xin-Li RG, Qu XK, Xu YJ, Fang WY, Yang YQ, Qiu XB. Prevalence and spectrum of Nkx2.6 mutations in patients with congenital heart disease. *Eur J Med Genet* 2014; **57**: 579-586 [PMID: 25195019 DOI: 10.1016/j.ejmg.2014.08.005]
- 129 **Smith JG**, Magnani JW, Palmer C, Meng YA, Soliman EZ, Musani SK, Kerr KF, Schnabel RB, Lubitz SA, Sotoodehnia N, Redline S, Pfeifer A, Müller M, Evans DS, Nalls MA, Liu Y, Newman AB, Zonderman AB, Evans MK, Deo R, Ellinor PT, Paltolo DN, Newton-Cheh C, Benjamin EJ, Mehra R, Alonso A, Heckbert SR, Fox ER. Genome-wide association studies of the PR interval in African Americans. *PLoS Genet* 2011; **7**: e1001304 [PMID: 21347284 DOI: 10.1371/journal.pgen.1001304]
- 130 **den Hoed M**, Eijgelsheim M, Esko T, Brundel BJ, Peal DS, Evans DM, Nolte IM, Segrè AV, Holm H, Handsaker RE, Westra HJ, Johnson T, Isaacs A, Yang J, Lundby A, Zhao JH, Kim YJ, Go MJ, Almgren P, Bochud M, Boucher G, Cornelis MC, Gudbjartsson D, Hadley D, van der Harst P, Hayward C, den Heijer M, Igli W, Jackson AU, Katalik Z, Luan J, Kemp JP, Kristiansson K, Ladenvall C, Lorentzon M, Montasser ME, Njajou OT, O'Reilly PF, Padmanabhan S, St Pourcain B, Rankinen T, Salo P, Tanaka T, Timpton NJ, Vitart V, Waite L, Wheeler W, Zhang W, Draisma HH, Feitosa MF, Kerr KF, Lind PA, Mihailov E, Onland-Moret NC, Song C, Weedon MN, Xie W, Yengo L, Absher D, Albert CM, Alonso A, Arking DE, de Bakker PI, Balkau B, Barlassina C, Benaglio P, Bis JC, Bouatia-Naji N, Brage S, Chanock SJ, Chines PS, Chung M, Darbar D, Dina C, Dörö M, Elliott P, Felix SB, Fischer K, Fuchsberger C, de Geus EJ, Goyette P, Gudnason V, Harris TB, Hartikainen AL, Havulinna AS, Heckbert SR, Hicks AA, Hofman A, Holewijn S, Hoogstra-Berends F, Hottenga JJ, Jensen MK, Johansson A, Junnila J, Kääb S, Kanon B, Ketkar S, Khaw KT, Knowles JW, Kooper AS, Kors JA, Kumari M, Milani L, Laiho P, Lakatta EG, Langenberg C, Leusink M, Liu Y, Luben RN, Lunetta KL, Lynch SN, Markus MR, Marques-Vidal P, Mateo Leach I, McArdle WL, McCarroll SA, Medland SE, Miller KA, Montgomery GW, Morrison AC, Müller-Nurasyid M, Navarro P, Nelis M, O'Connell JR, O'Donnell CJ, Ong KK, Newman AB, Peters A, Polasek O, Pouta A, Pramstaller PP, Psaty BM, Rao DC, Ring SM, Rossin EJ, Rudan D, Sanna S, Scott RA, Sehmi JS, Sharp S, Shin JT, Singleton AB, Smith AV, Soranzo N, Spector TD, Stewart C, Stringham HM, Tarasov KV, Uitterlinden AG, Vandendput L, Hwang SJ, Whitfield JB, Wijmenga C, Wild SH, Willemse G, Wilson JF, Witteman JC, Wong A, Wong Q, Jamshidi Y, Zitting P, Boer JM, Boomsma DI, Borecki IB, van Duijn CM, Ekelund U, Forouhi NG, Froguel P, Hingorani A, Ingelsson E, Kivimaki M, Kronmal RA, Kuh D, Lind L, Martin NG, Oostra BA, Pedersen NL, Quertermous T, Rotter JI, van der Schouw YT, Verschuren WM, Walker M, Albanes D, Arnar DO, Assimes TL, Bandinelli S, Boehnke M, de Boer RA, Bouchard C, Caulfield WL, Chambers JC, Curhan G, Cusi D, Eriksson J, Ferrucci L, van Gilst WH, Glorioso N, de Graaf J, Groop L, Gyllensten U, Hsueh WC, Hu FB, Huikuri HV, Hunter DJ, Iribarren C, Isomaa B, Jarvelin MR, Jula A, Kähönen M, Kiemeney LA, van der Klauw MM, Kooper JS, Kraft P, Iacoviello L, Lehtimäki T, Lokki ML,

- Mitchell BD, Navis G, Nieminen MS, Ohlsson C, Poulter NR, Qi L, Raitakari OT, Rimm EB, Rioux JD, Rizzi F, Rudan I, Salomaa V, Sever PS, Shields DC, Shuldiner AR, Sinisalo J, Stanton AV, Stolk RP, Strachan DP, Tardif JC, Thorsteinsdottir U, Tuomilehto J, van Veldhuisen DJ, Virtamo J, Viikari J, Vollenweider P, Waeber G, Widen E, Cho YS, Olsen JV, Visscher PM, Willer C, Franke L, Erdmann J, Thompson JR, Pfeifer A, Sotoodehnia N, Newton-Cheh C, Ellinor PT, Stricker BH, Metspalu A, Perola M, Beckmann JS, Smith GD, Stefansson K, Wareham NJ, Munroe PB, Sibon OC, Milan DJ, Snieder H, Samani NJ, Loos RJ. Identification of heart rate-associated loci and their effects on cardiac conduction and rhythm disorders. *Nat Genet* 2013; **45**: 621-631 [PMID: 23583979 DOI: 10.1038/ng.2610]
- 131 **Parvez B**, Shoemaker MB, Muhammad R, Richardson R, Jiang L, Blair MA, Roden DM, Darbar D. Common genetic polymorphism at 4q25 locus predicts atrial fibrillation recurrence after successful cardioversion. *Heart Rhythm* 2013; **10**: 849-855 [PMID: 23428961 DOI: 10.1016/j.hrthm.2013.02.018]
- 132 **Mohanty S**, Santangeli P, Bai R, Di Biase L, Mohanty P, Pump A, Natale A. Variant rs2200733 on chromosome 4q25 confers increased risk of atrial fibrillation: evidence from a meta-analysis. *J Cardiovasc Electrophysiol* 2013; **24**: 155-161 [PMID: 23130627 DOI: 10.1111/jce.12017]
- 133 **Olesen MS**, Holst AG, Jabbari J, Nielsen JB, Christophersen IE, Sajadieh A, Haunsoe S, Svendsen JH. Genetic loci on chromosomes 4q25, 7p31, and 12p12 are associated with onset of lone atrial fibrillation before the age of 40 years. *Can J Cardiol* 2012; **28**: 191-195 [PMID: 22336519 DOI: 10.1016/j.cjca.2011.11.016]
- 134 **Henningsen KM**, Olesen MS, Haunsoe S, Svendsen JH. Association of rs2200733 at 4q25 with early onset of lone atrial fibrillation in young patients. *Scand Cardiovasc J* 2011; **45**: 324-326 [PMID: 21793630 DOI: 10.3109/14017431.2011.594081]
- 135 **Kiliszek M**, Franaszczyk M, Kozluk E, Lodziński P, Piatkowska A, Broda G, Płoski R, Opolski G. Association between variants on chromosome 4q25, 16q22 and 1q21 and atrial fibrillation in the Polish population. *PLoS One* 2011; **6**: e21790 [PMID: 21760908 DOI: 10.1371/journal.pone.0021790]
- 136 **Chinchilla A**, Daimi H, Lozano-Velasco E, Dominguez JN, Caballero R, Delpón E, Tamargo J, Cinca J, Hove-Madsen L, Aranega AE, Franco D. PITX2 insufficiency leads to atrial electrical and structural remodeling linked to arrhythmogenesis. *Circ Cardiovasc Genet* 2011; **4**: 269-279 [PMID: 21511879 DOI: 10.1161/CIRCGENETICS.110.958116]
- 137 **Sinner MF**, Lubitz SA, Pfeifer A, Makino S, Beckmann BM, Lunetta KL, Steinbeck G, Perz S, Rahman R, Sonni A, Greenberg SM, Furie KL, Wichmann HE, Meitinger T, Peters A, Benjamin EJ, Rosand J, Ellinor PT, Kääb S. Lack of replication in polymorphisms reported to be associated with atrial fibrillation. *Heart Rhythm* 2011; **8**: 403-409 [PMID: 21056700 DOI: 10.1016/j.hrthm.2010.11.003]
- 138 **Lee KT**, Yeh HY, Tung CP, Chu CS, Cheng KH, Tsai WC, Lu YH, Chang JG, Sheu SH, Lai WT. Association of RS2200733 but not RS10033464 on 4q25 with atrial fibrillation based on the recessive model in a Taiwanese population. *Cardiology* 2010; **116**: 151-156 [PMID: 20606429 DOI: 10.1159/000318172]
- 139 **Wang J**, Klysik E, Sood S, Johnson RL, Wehrens XH, Martin JF. Pitx2 prevents susceptibility to atrial arrhythmias by inhibiting left-sided pacemaker specification. *Proc Natl Acad Sci USA* 2010; **107**: 9753-9758 [PMID: 20457925 DOI: 10.1073/pnas.0912585107]
- 140 **Kirchhof P**, Kahr PC, Kaese S, Piccini I, Vokshi I, Scheld HH, Rotering H, Fortmueller L, Laakmann S, Verheule S, Schotten U, Fabritz L, Brown NA. PITX2c is expressed in the adult left atrium, and reducing Pitx2c expression promotes atrial fibrillation inducibility and complex changes in gene expression. *Circ Cardiovasc Genet* 2011; **4**: 123-133 [PMID: 21282332 DOI: 10.1161/CIRCGENETICS.110.958058]
- 141 **Aguirre LA**, Alonso ME, Badía-Careaga C, Rollán I, Arias C, Fernández-Miñán A, López-Jiménez E, Aránega A, Gómez-Skarmeta JL, Franco D, Manzanares M. Long-range regulatory interactions at the 4q25 atrial fibrillation risk locus involve PITX2c and ENPEP. *BMC Biol* 2015; **13**: 26 [PMID: 2588893 DOI: 10.1186/s12915-015-0138-0]
- 142 **Gore-Panter SR**, Hsu J, Hanna P, Gillinov AM, Pettersson G, Newton DW, Moravec CS, Van Wagoner DR, Chung MK, Barnard J, Smith JD. Atrial Fibrillation associated chromosome 4q25 variants are not associated with PITX2c expression in human adult left atrial appendages. *PLoS One* 2014; **9**: e86245 [PMID: 24465984 DOI: 10.1371/journal.pone.0086245]
- 143 **Martin RI**, Babaei MS, Choy MK, Owens WA, Chico TJ, Keenan D, Yonan N, Koref MS, Keavney BD. Genetic variants associated with risk of atrial fibrillation regulate expression of PITX2, CAV1, MYOZ1, C9orf3 and FANCC. *J Mol Cell Cardiol* 2015; **85**: 207-214 [PMID: 26073630 DOI: 10.1016/j.jmcc.2015.06.005]
- 144 **Gudbjartsson DF**, Helgason H, Gudjonsson SA, Zink F, Oddson A, Gylfason A, Besenbacher S, Magnusson G, Halldorsson BV, Hjartarson E, Sigurdsson GT, Stacey SN, Frigge ML, Holm H, Saemundsdottir J, Helgadottir HT, Johannsdottir H, Sigfusson G, Thorgeirsson G, Sverrisson JT, Gretarsdottir S, Walters GB, Rafnar T, Thjodleifsson B, Bjornsson ES, Olafsson S, Thorarinsson H, Steingrimsdottir T, Gudmundsdottir TS, Theodors A, Jonasson JG, Sigurdsson A, Bjornsdottir G, Jonsson JJ, Thorarensen O, Ludvigsson P, Gudbjartsson H, Eyjolfsson GI, Sigurdardottir O, Olafsson I, Arnar DO, Magnusson OT, Kong A, Masson G, Thorsteinsdottir U, Helgason A, Sulem P, Stefansson K. Large-scale whole-genome sequencing of the Icelandic population. *Nat Genet* 2015; **47**: 435-444 [PMID: 25807286 DOI: 10.1038/ng.3247]
- 145 **Li C**, Wang F, Yang Y, Fu F, Xu C, Shi L, Li S, Xia Y, Wu G, Cheng X, Liu H, Wang C, Wang P, Hao J, Ke Y, Zhao Y, Liu M, Zhang R, Gao L, Yu B, Zeng Q, Liao Y, Yang B, Tu X, Wang QK. Significant association of SNP rs2106261 in the ZFHX3 gene with atrial fibrillation in a Chinese Han GeneID population. *Hum Genet* 2011; **129**: 239-246 [PMID: 21107608 DOI: 10.1007/s00439-010-0912-6]
- 146 **Zhai C**, Cong H, Liu Y, Zhang Y, Liu X, Zhang H, Ren Z. Rs1793343 polymorphism in zinc finger homeobox 3 (ZFHX3) gene and atrial fibrillation: an updated meta-analysis of 10 case-control comparisons. *BMC Cardiovasc Disord* 2015; **15**: 58 [PMID: 26112950 DOI: 10.1186/s12872-015-0044-y]
- 147 **Lin H**, Sinner MF, Brody JA, Arking DE, Lunetta KL, Rienstra M, Lubitz SA, Magnani JW, Sotoodehnia N, McKnight B, McManus DD, Boerwinkle E, Psaty BM, Rotter JI, Bis JC, Gibbs RA, Muzny D, Kovar CL, Morrison AC, Gupta M, Folsom AR, Kääb S, Heckbert SR, Alonso A, Ellinor PT, Benjamin EJ. Targeted sequencing in candidate genes for atrial fibrillation: the Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE) Targeted Sequencing Study. *Heart Rhythm* 2014; **11**: 452-457 [PMID: 24239840 DOI: 10.1016/j.hrthm.2013.11.012]
- 148 **Mahida S**, Mills RW, Tucker NR, Simonson B, Macri V, Lemoine MD, Das S, Milan DJ, Ellinor PT. Overexpression of KCNN3 results in sudden cardiac death. *Cardiovasc Res* 2014; **101**: 326-334 [PMID: 24296650 DOI: 10.1093/cvr/cvt269]
- 149 **Lozano-Velasco E**, Hernández-Torres F, Daimi H, Serra SA, Herráez A, Hove-Madsen L, Aránega A, Franco D. Pitx2 impairs calcium handling in a dose-dependent manner by modulating Wnt signalling. *Cardiovasc Res* 2016; **109**: 55-66 [PMID: 26243430]
- 150 **Jiang Q**, Ni B, Shi J, Han Z, Qi R, Xu W, Wang D, Wang DW, Chen M. Down-regulation of ATBF1 activates STAT3 signalling via PIAS3 in pacing-induced HL-1 atrial myocytes. *Biochem Biophys Res Commun* 2014; **449**: 278-283 [PMID: 24857987 DOI: 10.1016/j.bbrc.2014.05.041]
- 151 **Kääb S**, Darbar D, van Noord C, Dupuis J, Pfeifer A, Newton-Cheh C, Schnabel R, Makino S, Sinner MF, Kannankeril PJ, Beckmann BM, Choudry S, Donahue BS, Heeringa J, Perz S, Lunetta KL, Larson MG, Levy D, MacRae CA, Ruskin JN, Wacker A, Schömöig A, Wichmann HE, Steinbeck G, Meitinger T, Uitterlinden AG, Witteman JC, Roden DM, Benjamin EJ, Ellinor PT. Large scale replication and meta-analysis of variants on chromosome 4q25 associated with atrial fibrillation. *Eur Heart J* 2009; **30**: 813-819 [PMID: 19141561 DOI: 10.1093/euroheartj/

- ehn578]
- 152 **Tsai CT**, Hsieh CS, Chang SN, Chuang EY, Juang JM, Lin LY, Lai LP, Hwang JJ, Chiang FT, Lin JL. Next-generation sequencing of nine atrial fibrillation candidate genes identified novel de novo mutations in patients with extreme trait of atrial fibrillation. *J Med Genet* 2015; **52**: 28-36 [PMID: 25391453 DOI: 10.1136/jmedgenet-2014-102618]
- 153 **Huang Y**, Wang C, Yao Y, Zuo X, Chen S, Xu C, Zhang H, Lu Q, Chang L, Wang F, Wang P, Zhang R, Hu Z, Song Q, Yang X, Li C, Li S, Zhao Y, Yang Q, Yin D, Wang X, Si W, Li X, Xiong X, Wang D, Huang Y, Luo C, Li J, Wang J, Chen J, Wang L, Wang L, Han M, Ye J, Chen F, Liu J, Liu Y, Wu G, Yang B, Cheng X, Liao Y, Wu Y, Ke T, Chen Q, Tu X, Elston R, Rao S, Yang Y, Xia Y, Wang QK. Molecular Basis of Gene-Gene Interaction: Cyclic Cross-Regulation of Gene Expression and Post-GWAS Gene-Gene Interaction Involved in Atrial Fibrillation. *PLoS Genet* 2015; **11**: e1005393 [PMID: 26267381 DOI: 10.1371/journal.pgen.1005393]
- 154 **Li G**, Zhang R, Gao L, Zhang S, Dong Y, Yin X, Chang D, Yang Y, Xia Y. Lack of association between rs3807989 in cav1 and atrial fibrillation. *Int J Clin Exp Pathol* 2014; **7**: 4339-4344 [PMID: 25120818]
- 155 **Tada H**, Shiffman D, Smith JG, Sjögren M, Lubitz SA, Ellinor PT, Louie JZ, Catanese JJ, Engström G, Devlin JJ, Kathiresan S, Melander O. Twelve-single nucleotide polymorphism genetic risk score identifies individuals at increased risk for future atrial fibrillation and stroke. *Stroke* 2014; **45**: 2856-2862 [PMID: 25123217 DOI: 10.1161/STROKEAHA.114.006072]
- 156 **Pfeifer A**, van Noord C, Marciante KD, Arking DE, Larson MG, Smith AV, Tarasov KV, Müller M, Sotoodehnia N, Sinner MF, Verwoert GC, Li M, Kao WH, Köttgen A, Coresh J, Bis JC, Psaty BM, Rice K, Rotter JI, Rivadeneira F, Hofman A, Kors JA, Stricker BH, Uitterlinden AG, van Duijn CM, Beckmann BM, Sauter W, Gieger C, Lubitz SA, Newton-Cheh C, Wang TJ, Magnani JW, Schnabel RB, Chung MK, Barnard J, Smith JD, Van Wagoner DR, Vasan RS, Aspelund T, Eiriksdottir G, Harris TB, Launer LJ, Najjar SS, Lakatta E, Schlessinger D, Uda M, Abecasis GR, Müller-Myhsok B, Ehret GB, Boerwinkle E, Chakravarti A, Soliman EZ, Lunetta KL, Perz S, Wichmann HE, Meitinger T, Levy D, Gudnason V, Ellinor PT, Sanna S, Kääb S, Witteman JC, Alonso A, Benjamin EJ, Heckbert SR. Genome-wide association study of PR interval. *Nat Genet* 2010; **42**: 153-159 [PMID: 20062060 DOI: 10.1038/ng.517]
- 157 **Holm H**, Gudbjartsson DF, Arnar DO, Thorleifsson G, Thorgeirsson G, Stefansdottir H, Gudjonsson SA, Jonasdottir A, Mathiesen EB, Njølstad I, Nyrnes A, Wilsgaard T, Hald EM, Hveem K, Stoltenberg C, Løchen ML, Kong A, Thorsteinsdottir U, Stefansson K. Several common variants modulate heart rate, PR interval and QRS duration. *Nat Genet* 2010; **42**: 117-122 [PMID: 20062063 DOI: 10.1038/ng.511]
- 158 **Butler AM**, Yin X, Evans DS, Nalls MA, Smith EN, Tanaka T, Li G, Buxbaum SG, Whitsel EA, Alonso A, Arking DE, Benjamin EJ, Berenson GS, Bis JC, Chen W, Deo R, Ellinor PT, Heckbert SR, Heiss G, Hsueh WC, Keating BJ, Kerr KF, Li Y, Limacher MC, Liu Y, Lubitz SA, Marciante KD, Mehra R, Meng YA, Newman AB, Newton-Cheh C, North KE, Palmer CD, Psaty BM, Quirbura PM, Redline S, Reiner AP, Rotter JI, Schnabel RB, Schork NJ, Singleton AB, Smith JG, Soliman EZ, Srinivasan SR, Zhang ZM, Zonderman AB, Ferrucci L, Murray SS, Evans MK, Sotoodehnia N, Magnani JW, Avery CL. Novel loci associated with PR interval in a genome-wide association study of 10 African American cohorts. *Circ Cardiovasc Genet* 2012; **5**: 639-646 [PMID: 23139255 DOI: 10.1161/CIRCGENETICS.112.963991]
- 159 **Bezzina CR**, Barc J, Mizusawa Y, Remme CA, Gourraud JB, Simonet F, Verkerk AO, Schwartz PJ, Crotti L, Dagradis F, Guicheney P, Fressart V, Leenhardt A, Antzelevitch C, Bartkowiak S, Borggrefe M, Schimpf R, Schulze-Bahr E, Zumhagen S, Behr ER, Bastiaenen R, Tfelt-Hansen J, Olesen MS, Kääb S, Beckmann BM, Weeke P, Watanabe H, Endo N, Minamino T, Horie M, Ohno S, Hasegawa K, Makita N, Nogami A, Shimizu W, Aiba T, Froguel P, Balkau B, Lantieri O, Torchio M, Wiese C, Weber D, Wolswinkel R, Coronel R, Boukens BJ, Bézieau S, Charpentier E, Chatel S, Despres A, Gros F, Kyndt F, Lecointe S, Lindenbaum P, Portero V, Violleau J, Gessler M, Tan HL, Roden DM, Christoffels VM, Le Marec H, Wilde AA, Probst V, Schott JJ, Dina C, Redon R. Common variants at SCN5A-SCN10A and HEY2 are associated with Brugada syndrome, a rare disease with high risk of sudden cardiac death. *Nat Genet* 2013; **45**: 1044-1049 [PMID: 23872634 DOI: 10.1038/ng.2712]
- 160 **Newton-Cheh C**, Eijgelsheim M, Rice KM, de Bakker PI, Yin X, Estrada K, Bis JC, Marciante K, Rivadeneira F, Noseworthy PA, Sotoodehnia N, Smith NL, Rotter JI, Kors JA, Witteman JC, Hofman A, Heckbert SR, O'Donnell CJ, Uitterlinden AG, Psaty BM, Lumley T, Larson MG, Stricker BH. Common variants at ten loci influence QT interval duration in the QTGEN Study. *Nat Genet* 2009; **41**: 399-406 [PMID: 19305408 DOI: 10.1038/ng.364]
- 161 **Pfeifer A**, Sanna S, Arking DE, Müller M, Gateva V, Fuchsberger C, Ehret GB, Orrù M, Pattaro C, Köttgen A, Perz S, Usala G, Barbalic M, Li M, Pütz B, Scuteri A, Prineas RJ, Sinner MF, Gieger C, Najjar SS, Kao WH, Mühlleisen TW, Dei M, Happel C, Möhlenkamp S, Crisponi L, Erbel R, Jöckel KH, Naitza S, Steinbeck G, Marroni F, Hicks AA, Lakatta E, Müller-Myhsok B, Pramstaller PP, Wichmann HE, Schlessinger D, Boerwinkle E, Meitinger T, Uda M, Coresh J, Kääb S, Abecasis GR, Chakravarti A. Common variants at ten loci modulate the QT interval duration in the QTSCD Study. *Nat Genet* 2009; **41**: 407-414 [PMID: 19305409 DOI: 10.1038/ng.362]
- 162 **Arking DE**, Pulit SL, Crotti L, van der Harst P, Munroe PB, Koopmann TT, Sotoodehnia N, Rossin EJ, Morley M, Wang X, Johnson AD, Lundby A, Gudbjartsson DF, Noseworthy PA, Eijgelsheim M, Bradford Y, Tarasov KV, Dörr M, Müller-Nurasyid M, Lahtinen AM, Nolte IM, Smith AV, Bis JC, Isaacs A, Newhouse SJ, Evans DS, Post WS, Waggett D, Lyytikäinen LP, Hicks AA, Eisele L, Ellinghaus D, Hayward C, Navarro P, Ulivi S, Tanaka T, Tester DJ, Chatel S, Gustafsson S, Kumari M, Morris RW, Naluaï ÄT, Padmanabhan S, Klutig A, Strohmer B, Panayiotou AG, Torres M, Knoflach M, Hubacek JA, Slowikowski K, Raychaudhuri S, Kumar RD, Harris TB, Launer LJ, Shuldiner AR, Alonso A, Bader JS, Ehret G, Huang H, Kao WH, Strait JB, Macfarlane PW, Brown M, Caulfield MJ, Samani NJ, Kronenberg F, Willeit J, Smith JG, Greiser KH, Meyer Zu Schwabedissen H, Werdan K, Carella M, Zelante L, Heckbert SR, Psaty BM, Rotter JI, Kolcic I, Polašek O, Wright AF, Griffin M, Daly MJ, Arnar DO, Hölm H, Thorsteinsdottir U, Denny JC, Roden DM, Zuvich RL, Emilsson V, Plump AS, Larson MG, O'Donnell CJ, Yin X, Bobbo M, D'Adamo AP, Iorio A, Sinagra G, Carracedo A, Cummings SR, Nalls MA, Jula A, Kontula KK, Marjamaa A, Oikarinen L, Perola M, Porthan K, Erbel R, Hoffmann P, Jöckel KH, Kälsch H, Nöthen MM, den Hoed M, Loos RJ, Thelle DS, Gieger C, Meitinger T, Perz S, Peters A, Prucha H, Sinner MF, Waldenberger M, de Boer RA, Franke L, van der Vleuten PA, Beckmann BM, Martens E, Bardai A, Hofman N, Wilde AA, Behr ER, Dalageorgou C, Giudicessi JR, Medeiros-Domingo A, Barc J, Kyndt F, Probst V, Ghidoni A, Insolia R, Hamilton RM, Scherer SW, Brandimarto J, Margulies K, Moravec CE, del Greco M F, Fuchsberger C, O'Connell JR, Lee WK, Watt GC, Campbell H, Wild SH, El Mokhtari NE, Frey N, Asselbergs FW, Mateo Leach I, Navis G, van den Berg MP, van Veldhuisen DJ, Kellis M, Krijthe BP, Franco OH, Hofman A, Kors JA, Uitterlinden AG, Witteman JC, Kedenko L, Lamina C, Oostra BA, Abecasis GR, Lakatta EG, Mulas A, Orrù M, Schlessinger D, Uda M, Markus MR, Völker U, Snieder H, Spector TD, Årnlöv J, Lind L, Sundström J, Syvänen AC, Kivimaki M, Kähönen M, Mononen N, Raitakari OT, Viikari JS, Adamkova V, Kiechl S, Brion M, Nicolaides AN, Paulweber B, Haerting J, Dominiczak AF, Nyberg F, Whincup PH, Hingorani AD, Schott JJ, Bezzina CR, Ingelsson E, Ferrucci L, Gasparini P, Wilson JF, Rudan I, Franke A, Mühlleisen TW, Pramstaller PP, Lehtimäki TJ, Paterson AD, Parsa A, Liu Y, van Duijn CM, Siscovick DS, Gudnason V, Jamshidi Y, Salomaa V, Felix SB, Sanna S, Ritchie MD, Stricker BH, Stefansson K, Boyer LA, Cappola TP, Olsen JV, Lage K, Schwartz PJ, Kääb S, Chakravarti A, Ackerman MJ, Pfeifer A, de Bakker PI, Newton-Cheh C. Genetic association

- study of QT interval highlights role for calcium signaling pathways in myocardial repolarization. *Nat Genet* 2014; **46**: 826-836 [PMID: 24952745 DOI: 10.1038/ng.3014]
- 163 **Li L**, Shen C, Yao Z, Liang J, Huang C. Genetic variants of potassium voltage-gated channel genes (KCNQ1, KCNH2, and KCNE1) affected the risk of atrial fibrillation in elderly patients. *Genet Test Mol Biomarkers* 2015; **19**: 359-365 [PMID: 26066992 DOI: 10.1089/gtmb.2014.0307]
- 164 **Chu HM**, Feng MJ, Li YG, Zhang YX, Ma JF, He B, Yu YB, Liu J, Chen XM. Polymorphisms but not mutations of the KCNQ1 gene are associated with lone atrial fibrillation in the Chinese Han population. *ScientificWorldJournal* 2013; **2013**: 373454 [PMID: 23710137 DOI: 10.1155/2013/373454]
- 165 **Zeng Z**, Tan C, Teng S, Chen J, Su S, Zhou X, Wang F, Zhang S, Gu D, Makielinski JC, Pu J. The single nucleotide polymorphisms of I(Ks) potassium channel genes and their association with atrial fibrillation in a Chinese population. *Cardiology* 2007; **108**: 97-103 [PMID: 17016049]
- 166 **Jabbari J**, Olesen MS, Holst AG, Nielsen JB, Haunso S, Svendsen JH. Common polymorphisms in KCNJ5 [corrected] are associated with early-onset lone atrial fibrillation in Caucasians. *Cardiology* 2011; **118**: 116-120 [PMID: 21555883 DOI: 10.1159/000323840]
- 167 **Hauer RN**, Groenewegen WA, Firouzi M, Ramanna H, Jongasma HJ. Cx40 polymorphism in human atrial fibrillation. *Adv Cardiol* 2006; **42**: 284-291 [PMID: 16646598]
- 168 **Zang X**, Zhang S, Xia Y, Li S, Fu F, Li X, Wang F, Zhang R, Tian X, Gao L, Zhang J, Yang Y, Tu X, Wang Q. SNP rs3825214 in TBX5 is associated with lone atrial fibrillation in Chinese Han population. *PLoS One* 2013; **8**: e64966 [PMID: 23717681 DOI: 10.1371/journal.pone.0064966]
- 169 **Wang C**, Wu M, Qian J, Li B, Tu X, Xu C, Li S, Chen S, Zhao Y, Huang Y, Shi L, Cheng X, Liao Y, Chen Q, Xia Y, Yao W, Wu G, Cheng M, Wang QK. Identification of rare variants in TNNI3 with atrial fibrillation in a Chinese GenoID population. *Mol Genet Genomics* 2015; **291**: 79-92 [PMID: 26169204]
- 170 **Jabbari J**, Olesen MS, Yuan L, Nielsen JB, Liang B, Maeri V, Christoffersen IE, Nielsen N, Sajadiyah A, Ellinor PT, Grunnet M, Haunso S, Holst AG, Svendsen JH, Jespersen T. Common and rare variants in SCN10A modulate the risk of atrial fibrillation. *Circ Cardiovasc Genet* 2015; **8**: 64-73 [PMID: 25691686 DOI: 10.1161/HGCAAA000000000000022]
- 171 **Delaney JT**, Muhammad R, Shi Y, Schilderout JS, Blair M, Short L, Roden DM, Darbar D. Common SCN10A variants modulate PR interval and heart rate response during atrial fibrillation. *Europace* 2014; **16**: 485-490 [PMID: 24072447 DOI: 10.1093/europace/eut278]
- 172 **Franco D**, Aranega A. Post-transcriptional regulatory mechanisms. Clinic, Genetics and Molecular Pathways of Congenital Heart Diseases. In: Sperling S, Driscoll D, Kelly R, editors. Springer, 2015; In press
- 173 **Espinosa-Lewis RA**, Wang DZ. MicroRNAs in heart development. *Curr Top Dev Biol* 2012; **100**: 279-317 [PMID: 22449848 DOI: 10.1016/B978-0-12-387786-4.00009-9]
- 174 **Zhou J**, Dong X, Zhou Q, Wang H, Qian Y, Tian W, Ma D, Li X. microRNA expression profiling of heart tissue during fetal development. *Int J Mol Med* 2014; **33**: 1250-1260 [PMID: 24604530 DOI: 10.3892/ijmm.2014.1691]
- 175 **Hu DL**, Liu YQ, Chen FK, Sheng YH, Yang R, Kong XQ, Cao KJ, Zhang JS, Qian LM. Differential expression of microRNAs in cardiac myocytes compared to undifferentiated P19 cells. *Int J Mol Med* 2011; **28**: 59-64 [PMID: 21455566 DOI: 10.3892/ijmm.2011.664]
- 176 **Synnergren J**, Améen C, Lindahl A, Olsson B, Sartipy P. Expression of microRNAs and their target mRNAs in human stem cell-derived cardiomyocyte clusters and in heart tissue. *Physiol Genomics* 2011; **43**: 581-594 [PMID: 20841501 DOI: 10.1152/physiolgenomics.00074.2010]
- 177 **Nishi H**, Sakaguchi T, Miyagawa S, Yoshikawa Y, Fukushima S, Saito S, Ueno T, Kurata T, Sawa Y. Impact of microRNA expression in human atrial tissue in patients with atrial fibrillation undergoing cardiac surgery. *PLoS One* 2013; **8**: e73397 [PMID: 24069193 DOI: 10.1371/journal.pone.0073397]
- 178 **Wang J**, Wang Y, Han J, Li Y, Xie C, Xie L, Shi J, Zhang J, Yang B, Chen D, Meng X. Integrated analysis of microRNA and mRNA expression profiles in the left atrium of patients with nonvalvular paroxysmal atrial fibrillation: Role of miR-146b-5p in atrial fibrosis. *Heart Rhythm* 2015; **12**: 1018-1026 [PMID: 25617731 DOI: 10.1016/j.hrthm.2015.01.026]
- 179 **Li M**, Zhang J. Circulating MicroRNAs: Potential and Emerging Biomarkers for Diagnosis of Cardiovascular and Cerebrovascular Diseases. *Biomed Res Int* 2015; **2015**: 730535 [PMID: 26180810 DOI: 10.1155/2015/730535]
- 180 **Orenes-Piñero E**, Montoro-García S, Patel JV, Valdés M, Marín F, Lip GY. Role of microRNAs in cardiac remodelling: new insights and future perspectives. *Int J Cardiol* 2013; **167**: 1651-1659 [PMID: 23063140 DOI: 10.1016/j.ijcard.2012.09.120]
- 181 **Poudel P**, Xu Y, Cui Z, Sharma D, Tian B, Paudel S. Atrial fibrillation: recent advances in understanding the role of microRNAs in atrial remodeling with an electrophysiological overview. *Cardiology* 2015; **131**: 58-67 [PMID: 25871909 DOI: 10.1159/000375403]
- 182 **Gomes da Silva AM**, Silbiger VN. miRNAs as biomarkers of atrial fibrillation. *Biomarkers* 2014; **19**: 631-636 [PMID: 25171770 DOI: 10.3109/1354750X.2014.954001]
- 183 **Liu Z**, Zhou C, Liu Y, Wang S, Ye P, Miao X, Xia J. The expression levels of plasma microRNAs in atrial fibrillation patients. *PLoS One* 2012; **7**: e44906 [PMID: 23028671 DOI: 10.1371/journal.pone.0044906]
- 184 **van den Boogaard M**, Smemo S, Burnicka-Turek O, Arnolds DE, van de Werken HJ, Klous P, McKean D, Muehlschlegel JD, Moosmann J, Toka O, Yang XH, Koopmann TT, Adriaens ME, Bezzina CR, de Laat W, Seidman C, Seidman JG, Christoffels VM, Nobrega MA, Barnett P, Moskowitz IP. A common genetic variant within SCN10A modulates cardiac SCN5A expression. *J Clin Invest* 2014; **124**: 1844-1852 [PMID: 24642470 DOI: 10.1172/JCI73140]
- 185 **Tao Y**, Zhang M, Li L, Bai Y, Zhou Y, Moon AM, Kaminski HJ, Martin JF. Pitx2, an atrial fibrillation predisposition gene, directly regulates ion transport and intercalated disc genes. *Circ Cardiovasc Genet* 2014; **7**: 23-32 [PMID: 24395921 DOI: 10.1161/CIRCGENETICS.113.000259]
- 186 **Nagy II**, Railo A, Rapila R, Hast T, Sormunen R, Tavi P, Räsänen J, Vainio SJ. Wnt-11 signalling controls ventricular myocardium development by patterning N-cadherin and beta-catenin expression. *Cardiovasc Res* 2010; **85**: 100-109 [PMID: 19622544 DOI: 10.1093/cvr/cvp254]
- 187 **Pandur P**, Lásche M, Eisenberg LM, Kühl M. Wnt-11 activation of a non-canonical Wnt signalling pathway is required for cardiogenesis. *Nature* 2002; **418**: 636-641 [PMID: 12167861]
- 188 **Martin A**, Maher S, Summerhurst K, Davidson D, Murphy P. Differential deployment of paralogous Wnt genes in the mouse and chick embryo during development. *Evol Dev* 2012; **14**: 178-195 [PMID: 23017026 DOI: 10.1111/j.1525-142X.2012.00534.x]
- 189 **Wang J**, Bai Y, Li N, Ye W, Zhang M, Greene SB, Tao Y, Chen Y, Wehrens XH, Martin JF. Pitx2-microRNA pathway that delimits sinoatrial node development and inhibits predisposition to atrial fibrillation. *Proc Natl Acad Sci USA* 2014; **111**: 9181-9186 [PMID: 24927531 DOI: 10.1073/pnas.1405411111]
- 190 **Chiang DY**, Kongchan N, Beavers DL, Alsina KM, Voigt N, Neilson JR, Jakob H, Martin JF, Dobrev D, Wehrens XH, Li N. Loss of microRNA-106b-25 cluster promotes atrial fibrillation by enhancing ryanodine receptor type-2 expression and calcium release. *Circ Arrhythm Electrophysiol* 2014; **7**: 1214-1222 [PMID: 25389315 DOI: 10.1161/CIRCEP.114.001973]
- 191 **Barana A**, Matamoros M, Dolz-Gaitón P, Pérez-Hernández M, Amorós I, Núñez M, Sacristán S, Pedraz Á, Pinto Á, Fernández-Avilés F, Tamargo J, Delpón E, Caballero R. Chronic atrial fibrillation increases microRNA-21 in human atrial myocytes decreasing L-type calcium current. *Circ Arrhythm Electrophysiol* 2014; **7**: 861-868 [PMID: 25107449 DOI: 10.1161/CIRCEP.114.001709]
- 192 **Daimi H**, Lozano-Velasco E, Haj Khelil A, Chibani JB, Barana A, Amorós I, González de la Fuente M, Caballero R, Aranega

- A, Franco D. Regulation of SCN5A by microRNAs: miR-219 modulates SCN5A transcript expression and the effects of flecainide intoxication in mice. *Heart Rhytm* 2015; **12**: 1333-1342 [PMID: 25701775 DOI: 10.1016/j.hrthm.2015.02.018]
- 193 **Luo X**, Pan Z, Shan H, Xiao J, Sun X, Wang N, Lin H, Xiao L, Maguy A, Qi XY, Li Y, Gao X, Dong D, Zhang Y, Bai Y, Ai J, Sun L, Lu H, Luo XY, Wang Z, Lu Y, Yang B, Nattel S. MicroRNA-26 governs profibrillatory inward-rectifier potassium current changes in atrial fibrillation. *J Clin Invest* 2013; **123**: 1939-1951 [PMID: 23543060 DOI: 10.1172/JCI62185]
- 194 **Jia X**, Zheng S, Xie X, Zhang Y, Wang W, Wang Z, Zhang Y, Wang J, Gao M, Hou Y. MicroRNA-1 accelerates the shortening of atrial effective refractory period by regulating KCNE1 and KCNB2 expression: an atrial tachypacing rabbit model. *PLoS One* 2013; **8**: e85639 [PMID: 24386485 DOI: 10.1371/journal.pone.0085639]
- 195 **Li YD**, Hong YF, Yusufuaji Y, Tang BP, Zhou XH, Xu GJ, Li JX, Sun L, Zhang JH, Xin Q, Xiong J, Ji YT, Zhang Y. Altered expression of hyperpolarization-activated cyclic nucleotide-gated channels and microRNA-1 and -133 in patients with age-associated atrial fibrillation. *Mol Med Rep* 2015; **12**: 3243-3248 [PMID: 26005035 DOI: 10.3892/mmr.2015.3831]
- 196 **Mommersteeg MT**, Brown NA, Prall OW, de Gier-de Vries C, Harvey RP, Moorman AF, Christoffels VM. Pitx2c and Nkx2-5 are required for the formation and identity of the pulmonary myocardium. *Circ Res* 2007; **101**: 902-909 [PMID: 17823370]
- 197 **Mints Y**, Yarmohammadi H, Khurram IM, Hoyt H, Hansford R, Zimmerman SL, Steinberg SJ, Judge DP, Tomaselli GF, Calkins H, Zipunnikov V, Nazarian S. Association of common variations on chromosome 4q25 and left atrial volume in patients with atrial fibrillation. *Clin Med Insights Cardiol* 2015; **9**: 39-45 [PMID: 26005361 DOI: 10.4137/CMC.S21712]
- 198 **Scridon A**, Fouilloux-Meugnier E, Loizon E, Rome S, Julien C, Barrès C, Chevalier P. Long-standing arterial hypertension is associated with Pitx2 down-regulation in a rat model of spontaneous atrial tachyarrhythmias. *Europace* 2015; **17**: 160-165 [PMID: 24908044 DOI: 10.1093/europace/euu139]
- 199 **Torrado M**, Franco D, Hernández-Torres F, Crespo-Leiro MG, Iglesias-Gil C, Castro-Beiras A, Mikhailov AT. Pitx2c is reactivated in the failing myocardium and stimulates myf5 expression in cultured cardiomyocytes. *PLoS One* 2014; **9**: e90561 [PMID: 24595098 DOI: 10.1371/journal.pone.0090561]
- 200 **Torrado M**, Franco D, Lozano-Velasco E, Hernández-Torres F, Calviño R, Aldama G, Centeno A, Castro-Beiras A, Mikhailov A. A MicroRNA-Transcription Factor Blueprint for Early Atrial Arrhythmogenic Remodeling. *Biomed Res Int* 2015; **2015**: 263151 [PMID: 26221584 DOI: 10.1155/2015/263151]

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