

Basic Study

Analysis of tumor-infiltrating gamma delta T cells in rectal cancer

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Author contributions: Rong L designed the study; Li K performed the research; Liu HM contributed new reagents and analytic tools; Rong L and Sun R analyzed the data; Rong L and Li K wrote the paper; all the authors contributed to this manuscript.

Institutional review board statement: The study was revised and approved by the Fifth Affiliated Hospital of Xinjiang Medical University Institutional Review Board.

Institutional animal care and use committee statement: All procedures involving animals were reviewed and approved by the Institutional Animal Care and Use Committee of the First Affiliated Hospital of Soochow University (IACUC protocol number: 2013-10-13).

Conflict-of-interest statement: We declare that there are no conflicts of interest to disclose.

Data sharing statement: No additional data are available.

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Received: November 1, 2015

Peer-review started: November 2, 2015

First decision: November 27, 2015

Revised: December 14, 2015

Accepted: January 11, 2016

Article in press: January 11, 2016

Published online: April 7, 2016

Abstract

AIM: To investigate the regulatory effect of V δ 1 T cells and the antitumor activity of V δ 2 T cells in rectal cancer.

METHODS: Peripheral blood, tumor tissues and para-carcinoma tissues from 20 rectal cancer patients were collected. Naïve CD4 T cells from the peripheral blood of rectal cancer patients were purified by negative selection using a Naive CD4⁺ T Cell Isolation Kit II (Miltenyi Biotec). Tumor tissues and para-carcinoma tissues were minced into small pieces and digested in a triple enzyme mixture containing collagenase type IV, hyaluronidase, and deoxyribonuclease for 2 h at room temperature. After digestion, the cells were washed twice in RPMI1640 and cultured in RPMI1640 containing 10% human serum supplemented with L-glutamine and 2-mercaptoethanol and 1000 U/mL of IL-2 for the generation of T cells. V δ 1 T cells and V δ 2 T cells from tumor tissues and para-carcinoma tissues were expanded by anti-TCR $\gamma\delta$ antibodies. The inhibitory effects of V δ 1 T cells on naïve CD4 T cells were analyzed using the CFSE method. The cytotoxicity of V δ 2 T cells on rectal cancer lines was determined by the LDH method.

RESULTS: The percentage of V δ 1 T cells in rectal tumor

tissues from rectal cancer patients was significantly increased, and positively correlated with the T stage. The percentage of V δ 2 T cells in rectal tumor tissues from rectal cancer patients was significantly decreased, and negatively correlated with the T stage. After culture for 14 d with 1 μ g/mL anti-TCR $\gamma\delta$ antibodies, the percentage of V δ 1 T cells from para-carcinoma tissues was 21.45% \pm 4.64%, and the percentage of V δ 2 T cells was 38.64% \pm 8.05%. After culture for 14 d, the percentage of V δ 1 T cells from rectal cancer tissues was 67.45% \pm 11.75% and the percentage of V δ 2 T cells was 8.94% \pm 2.85%. Tumor-infiltrating V δ 1 T cells had strong inhibitory effects, and tumor-infiltrating V δ 2 T cells showed strong cytolytic activity. The inhibitory effects of V δ 1 T cells from para-carcinoma tissues and from rectal cancer tissue were not significantly different. In addition, the cytolytic activities of V δ 2 T cells from para-carcinoma tissues and from rectal cancer tissues were not significantly different.

CONCLUSION: A percentage imbalance in V δ 1 and V δ 2 T cells in rectal cancer patients may contribute to the development of rectal cancer.

Key words: Rectal cancer; T cells; V δ 1 T cells; V δ 2 T cells; Foxp3; Cytotoxicity

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Core tip: The percentage of tumor-infiltrating V δ 1 T cells in rectal cancer patients increased when T stage increased, whereas the percentage of tumor-infiltrating V δ 2 T cells in rectal cancer patients decreased as T stage increased. V δ 1 T cells from rectal cancer tissues had strong regulatory effects, and in rectal cancer tissues the main infiltrating $\gamma\delta$ T cells were V δ 1 T cells. Although V δ 2 T cells from rectal cancer tissues have strong cytotoxic effects, there was little infiltration of V δ 2 T cells in rectal cancer tissues. Thus, an immunosuppressant microenvironment was formed in rectal cancer tissues, which may limit antitumor immunity and allow tumors in rectal cancer patients to evade immune surveillance.

Rong L, Li K, Li R, Liu HM, Sun R, Liu XY. Analysis of tumor-infiltrating gamma delta T cells in rectal cancer. *World J Gastroenterol* 2016; 22(13): 3573-3580 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i13/3573.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i13.3573>

INTRODUCTION

T cells can be divided into two major subsets according to their expression of rearranged adaptive T cell receptors (TCRs, $\gamma\delta$ T cells and $\alpha\beta$ T cells)^[1]. $\gamma\delta$ T cells, which represent a small subset (1%-10%) of CD3⁺ cells^[2], can be divided into two subsets: V δ 1 T

cells in the epithelial-associated lymphoid tissue and V δ 2 T cells in the peripheral blood^[3,4]. They differ in their cytokine production and receptor expression, V δ 2 T cells being more inflammatory^[5], and V δ 1 T cells having more of a regulatory phenotype^[6]. It has been demonstrated that V δ 1 T cells express Foxp3, and their number is substantially decreased in the peripheral blood of patients with new-onset systemic lupus erythematosus (SLE)^[6]. V δ 2 T cells have predominantly been investigated in the context of tumor immunosurveillance and host defense against viral invasion^[7-10].

Rectal cancer is one of the most common causes of cancer deaths worldwide^[11]. In recent years, combined chemoradiotherapy followed by total mesorectal excision (TME) has become the standard treatment for patients with locally advanced rectal cancer^[12]. Local excision is often considered a curative treatment alternative to TME in early rectal cancer^[13], however, one of the limitations of this approach is that it is impossible to determine the pN-category^[13]. Lymph node involvement in rectal cancer is known to correlate with T stage^[14,15]. There is increasing evidence that immune-profiling may help to predict clinical outcomes in rectal cancer, possibly more reliably than TNM classification or grading. There is evidence that a low number of tumor-infiltrating lymphocytes (TILs) predicts lymph node involvement in melanoma, gastric cancer, breast cancer, and cervical cancer^[13,16-18]. The precise role of $\gamma\delta$ T cells in the development of rectal cancer remains elusive.

In this study, we found that the percentage of V δ 1 T cells in rectal tumor tissues from rectal cancer patients was significantly increased, whereas the percentage of V δ 2 T cells in these tissues was significantly decreased. The percentages of V δ 1 and V δ 2 T cells correlated with the T stage of rectal cancer patients. To obtain V δ 1 and V δ 2 T cells, tumor tissues and para-carcinoma tissues were minced into small pieces, digested with a triple enzyme mixture containing collagenase type IV, hyaluronidase, and deoxyribonuclease and stimulated by anti-TCR $\gamma\delta$ antibodies. After 14 d culture, the percentage of V δ 1 T cells from para-carcinoma tissues was 21.45% \pm 4.64% and the percentage of V δ 2 T cells was 38.64% \pm 8.05%. The percentage of V δ 1 T cells from rectal cancer tissues was 67.45% \pm 11.75% and the percentage of V δ 2 T cells was 8.94% \pm 2.85%. Functional assays demonstrated that tumor-infiltrating V δ 1 T cells in rectal cancer patients have strong inhibitory effects, and tumor-infiltrating V δ 2 T cells displayed strong cytolytic activity. The inhibitory effects of V δ 1 T cells and the cytolytic activity of V δ 2 T cells from para-carcinoma tissues and from rectal cancer tissues were not significantly different. Collectively, these data suggest that an imbalance in the V δ 1 and V δ 2 T cell percentages creates an immunosuppressant microenvironment in rectal cancer tissues, which may allow tumors to limit antitumor immunity and evade

immune surveillance in rectal cancer patients.

MATERIALS AND METHODS

Patients

Twenty patients with rectal cancer were enrolled in this study. The study was approved by the Fifth Affiliated Hospital of Xinjiang Medical University (Xinjiang, China) and written informed consent was obtained from each participating patient. Peripheral blood, tumor tissues, and para-carcinoma tissues were collected from the patients.

Antibodies and reagents

RPMI-1640 medium and fetal bovine serum (FBS) were obtained from Gibco; FITC-conjugated anti-human TCR $\gamma\delta$ (IMMU510) was purchased from Beckman Coulter Immunotech; APC-conjugated anti-human CD3 (HIT3a) and FITC-conjugated anti-human TCR V δ 2 (B6) were purchased from Biolegend; FITC-conjugated anti-human TCR V δ 1 (TS8.2) was obtained from Pierce; a CellTrace™ CFSE Cell Proliferation Kit was purchased from Invitrogen; the CytoTox 96® Non-Radioactive Cytotoxicity Assay was purchased from Promega; interleukin 2 was purchased from Read United Cross Pharmaceutical Co., Ltd.

Cells

To expand V δ 1 T cells and V δ 2 T cells *in vitro*, tumor tissues and para-carcinoma tissues were minced into small pieces and then digested with a triple enzyme mixture containing collagenase type IV, hyaluronidase, and deoxyribonuclease for 2 h at room temperature. After digestion, the cells were washed twice in RPMI-1640 and cultured in RPMI-1640 medium with 10% FBS and 200 IU/mL interleukin 2 in 24-well culture plates coated with 1 μ g/mL anti-pan-TCR $\gamma\delta$ mAb. The HR8348 (human rectal carcinoma) cell line was obtained from the Cell Culture Center, Institute of Basic Medicine, Chinese Academy of Medical Sciences. HR8348 cells were cultured in complete RPMI-1640 medium with 10% FBS.

Flow cytometric analysis

Cells were washed with PBS containing 1% bovine serum albumin (BSA) and incubated with surface-staining antibodies for 30 min at 4 °C. The cells were then washed and resuspended in PBS. Cytometry data were acquired using a BD Accuri C6 flow cytometer (Becton Dickinson). Data analysis was carried out with FlowJo software (Tree Star Inc.).

CFSE proliferation assay

Naïve CD4 T cells were labeled with CFSE and used as the responder cells. The cells were cultured with V δ 1 T cells in the dark at a ratio of 1:2. After 5 d in culture, the cells were collected and washed twice with PBS

containing 1% BSA. The cells were analyzed using a BD Accuri C6 flow cytometer (Becton Dickinson). Data analysis was performed using FlowJo software (Tree Star Inc.).

LDH assay

To determine specific cytotoxicity, we used the CytoTox 96® Non-Radioactive Cytotoxicity Assay (Promega) based on the calorimetric detection of the released enzyme lactate dehydrogenase (LDH). HR8348 and V δ 2 T cells were co-cultured at the ratios of 10:1, 20:1 and 30:1. Assays were performed in triplicate. After 6 h at 37 °C, 50 μ L supernatant was assayed for LDH activity following the manufacturer's protocol. Controls for spontaneous LDH release in effector and target cells, as well as target maximum release, were prepared. The percentage of cytotoxicity was calculated as follows:

$$\% \text{Cytotoxicity} = \frac{([\text{Experimental} - \text{Effector spontaneous}] - [\text{Target spontaneous}])}{[\text{Target maximum} - \text{Target spontaneous}]} \times 100$$

Statistical analysis

The results are expressed as mean \pm SD. Data were analyzed by *t*-test or one-way analysis of variance (ANOVA) (SPSS version 16.0), followed by Tukey-Kramer multiple comparisons. In all analyses, the minimum acceptable level of significance was $P < 0.05$.

RESULTS

Percentage of V δ 1 and V δ 2 T cells in tumor tissue and para-carcinoma tissue from rectal cancer patients

We first compared the percentages of total $\gamma\delta$ T cells and the V δ 1 and V δ 2 T subsets in tumor tissues and para-carcinoma tissues from rectal cancer patients. There was no significant difference in the percentage of total $\gamma\delta$ T cells in the tumor tissues and para-carcinoma tissues of rectal cancer patients ($4.32\% \pm 0.026\%$ vs $4.30\% \pm 0.037\%$, $P > 0.05$) (Figure 1A). The percentage of V δ 1 T cells in tumor tissues was significantly greater than in para-carcinoma tissues ($2.58\% \pm 0.017\%$ vs $1.03\% \pm 0.008\%$, $P < 0.01$) (Figure 1B), and the percentage of V δ 2 T cells was significantly lower in tumor tissue than in para-carcinoma tissue ($1.75\% \pm 0.012\%$ vs $3.27\% \pm 0.032\%$, $P < 0.05$) (Figure 1C).

Correlation of V δ 1 and V δ 2 T cells with TNM stage in rectal cancer patients

The percentage of peripheral V δ 1 T cells in rectal cancer patients increased as T stage increased (Figure 2A), whereas the percentage of peripheral V δ 2 T cells decreased as T stage increased (Figure 2B). However, there was no significant correlation between N category or M category and the percentage of V δ 1 or V δ 2 T cells (data not shown).

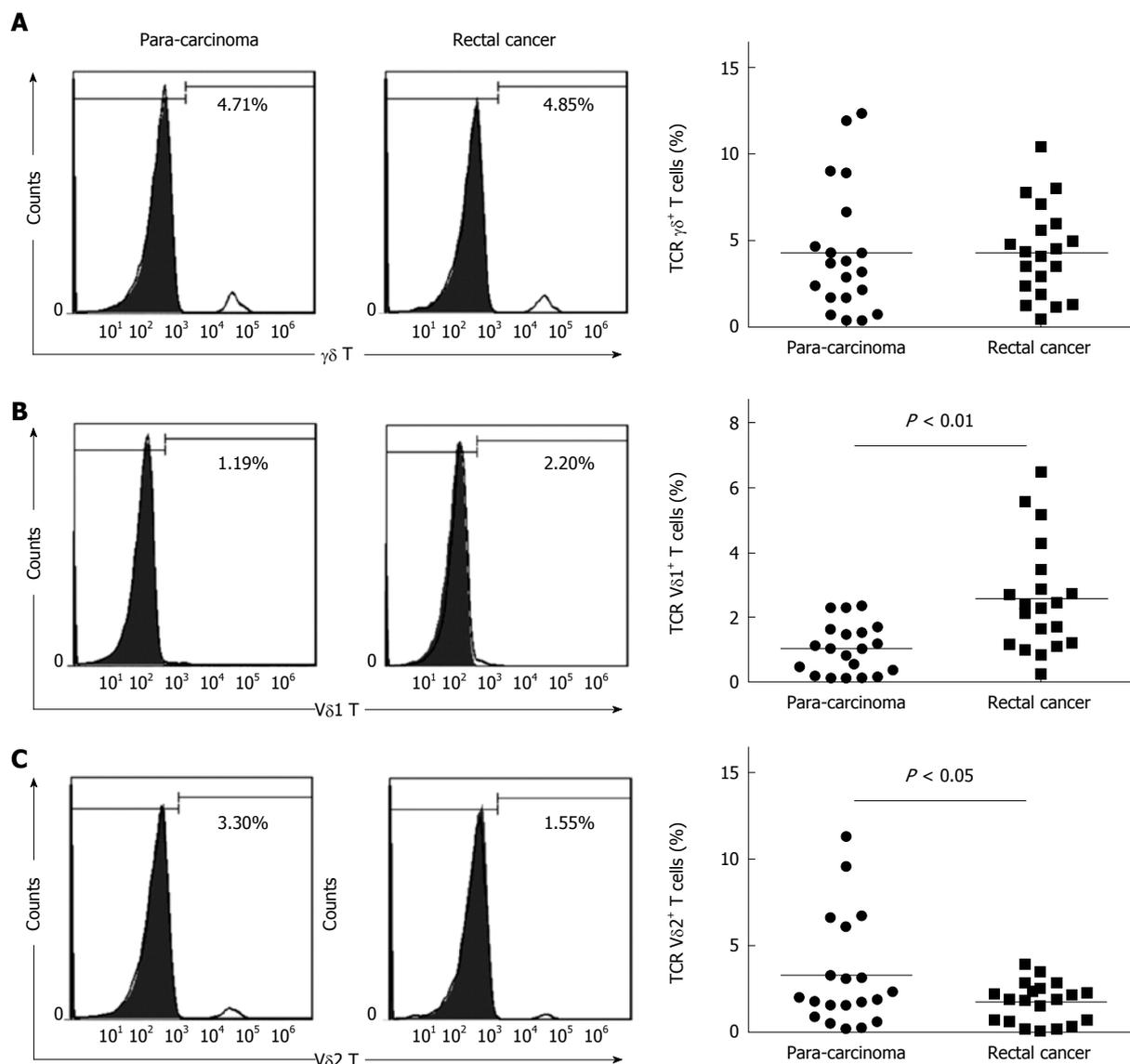


Figure 1 Percentage of infiltrating $\gamma\delta$ T cells in 20 rectal cancer patients. Cells were stained with an anti- $\gamma\delta$ TCR mAb, anti-V δ 1 mAb or anti-V δ 2 mAb and analyzed by flow cytometry. The left panels show representative histogram results from flow cytometry for $\gamma\delta$ T cells (A), V δ 1 T cells (B), and V δ 2 T cells (C). The right panels show bar graphs of the percentage of positively stained cells from the patients.

Percentage of V δ 1 and V δ 2 T cells after 14 d amplification with anti-TCR $\gamma\delta$ antibody

After culture in RPMI-1640 medium containing 10% FBS in 24-well culture plates coated with 1 μ g/mL anti-TCR $\gamma\delta$ antibody for 14 d, the percentage of V δ 1 T cells from para-carcinoma tissues was 21.45% \pm 4.64%, and the percentage of V δ 2 T cells was 38.64% \pm 8.05% (Figure 3A). After culture for 14 d, the percentage of V δ 1 T cells from rectal cancer tissues was 67.45% \pm 11.75%, and the percentage of V δ 2 T cells was 8.94% \pm 2.85% (Figure 3B).

Regulatory effects of tumor-infiltrating V δ 1 T cells and cytolytic activity of tumor-infiltrating V δ 2 T cells

The proliferation rate of naïve CD4T cells from the blood of rectal cancer patients was 80.23% \pm 11.86%; when co-cultured with V δ 1 T cells from para-carcinoma

tissues, the proliferation rate was 53.45% \pm 7.95%, and when co-cultured with V δ 1 T cells from rectal cancer tissues, the proliferation rate was 52.53% \pm 8.52% (Figure 4A). The inhibitory effects of V δ 1 T cells from para-carcinoma tissues and from rectal cancer tissues were not significantly different (Figure 4B). In addition, the cytolytic activities of V δ 2 T cells from para-carcinoma tissues and from rectal cancer tissues were not significantly different (Figure 4C).

DISCUSSION

The major finding of this study is that the percentage of V δ 1 T cells in the rectal tumor tissues of rectal cancer patients significantly increased, and the percentage of V δ 2 T cells in the rectal tumor tissues of rectal cancer patients significantly decreased, when

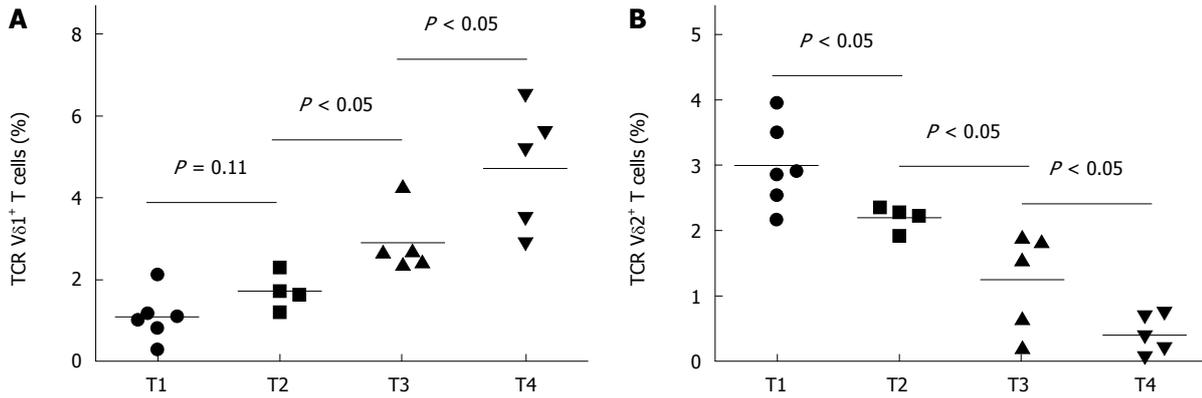


Figure 2 Percentage of tumor-infiltrating Vδ1 and Vδ2 T cells correlated with disease T stage. A: Tumor-infiltrating Vδ1 T cells positively correlated with disease T stage; B: Tumor-infiltrating Vδ2 T cells negatively correlated with disease T stage.

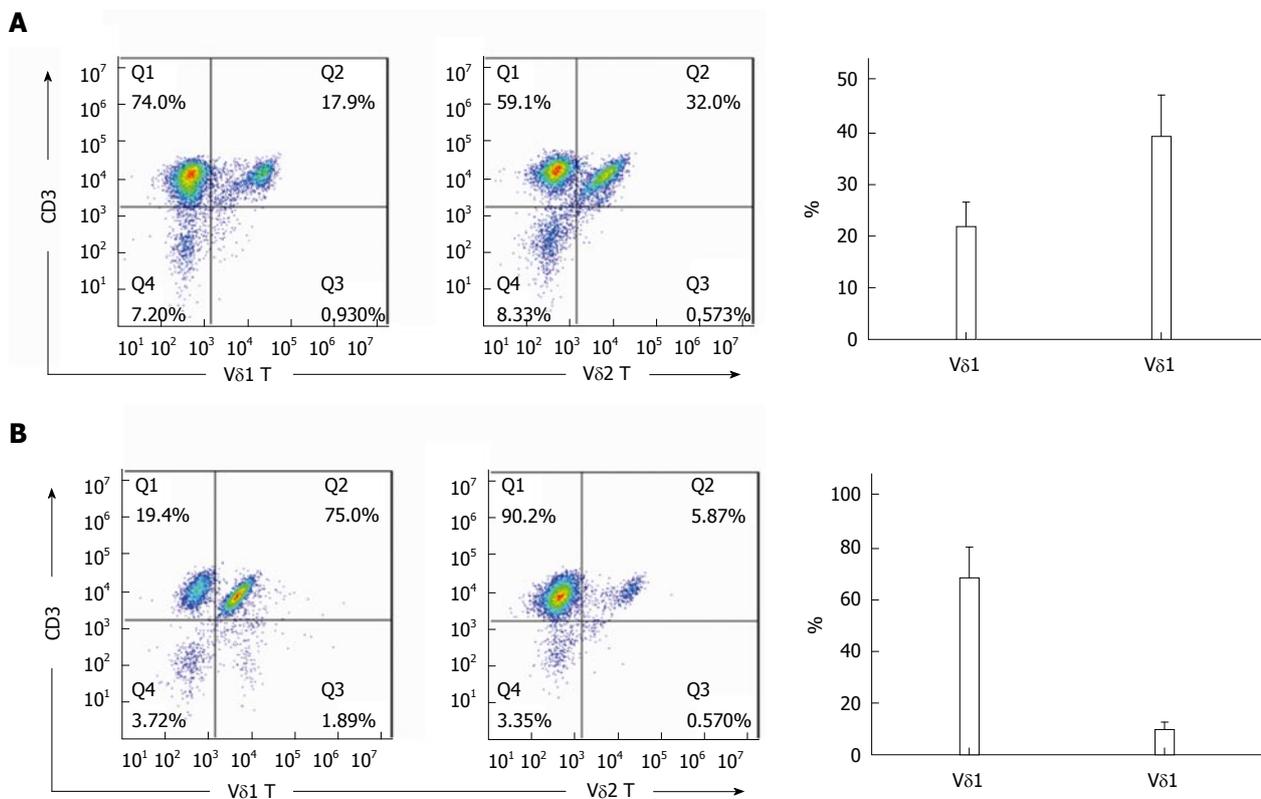


Figure 3 Percentage of Vδ1 and Vδ2 T cells from cancer tissues or para-carcinoma tissues after expansion. Cells were stained with anti-CD3 mAb, anti-Vδ1 mAb or anti-Vδ2 mAb and analyzed by flow cytometry. The left panels show representative dot plots from flow cytometry for Vδ1 and Vδ2 T cells from para-carcinoma tissues (A) and rectal cancer tissues (B). The right panels show bar graphs of the percentage of positively stained cells from 20 rectal cancer patients.

compared with para-carcinoma tissues. $\gamma\delta$ T cells can be divided into two subsets: Vδ1 T cells found in the epithelial-associated lymphoid tissue and Vδ2 T cells in the peripheral blood. Our results showed that after culture for 14 d with 1 μ g/mL anti-TCR $\gamma\delta$ antibody, the major subset from para-carcinoma tissues was Vδ1 T cells, and the major subset from rectal cancer tissues was Vδ2 T cells. Tumor-infiltrating Vδ1 T cells had strong inhibitory effects, and tumor-infiltrating Vδ2 T cells showed strong cytolytic activity. Although there were no significant differences in the cytolytic activities of Vδ2 T cells from para-carcinoma tissues

and from rectal cancer tissues, the predominant subset in rectal cancer tissues was Vδ1 T cells. Thus, tumors may limit antitumor immunity and evade immune surveillance in rectal cancer patients by forming an immunosuppressant microenvironment.

The MHC-independent antigen recognition and strong cytotoxicity to tumor cells make $\gamma\delta$ T cells attractive candidate effector cells for cancer immunotherapy^[19-25]. Administration of Vδ2 T cells at suitable intervals after chemotherapy and zoledronate may substantially increase antitumor activity in a range of malignancies^[26], whereas tumor-infiltrating Vδ1 T

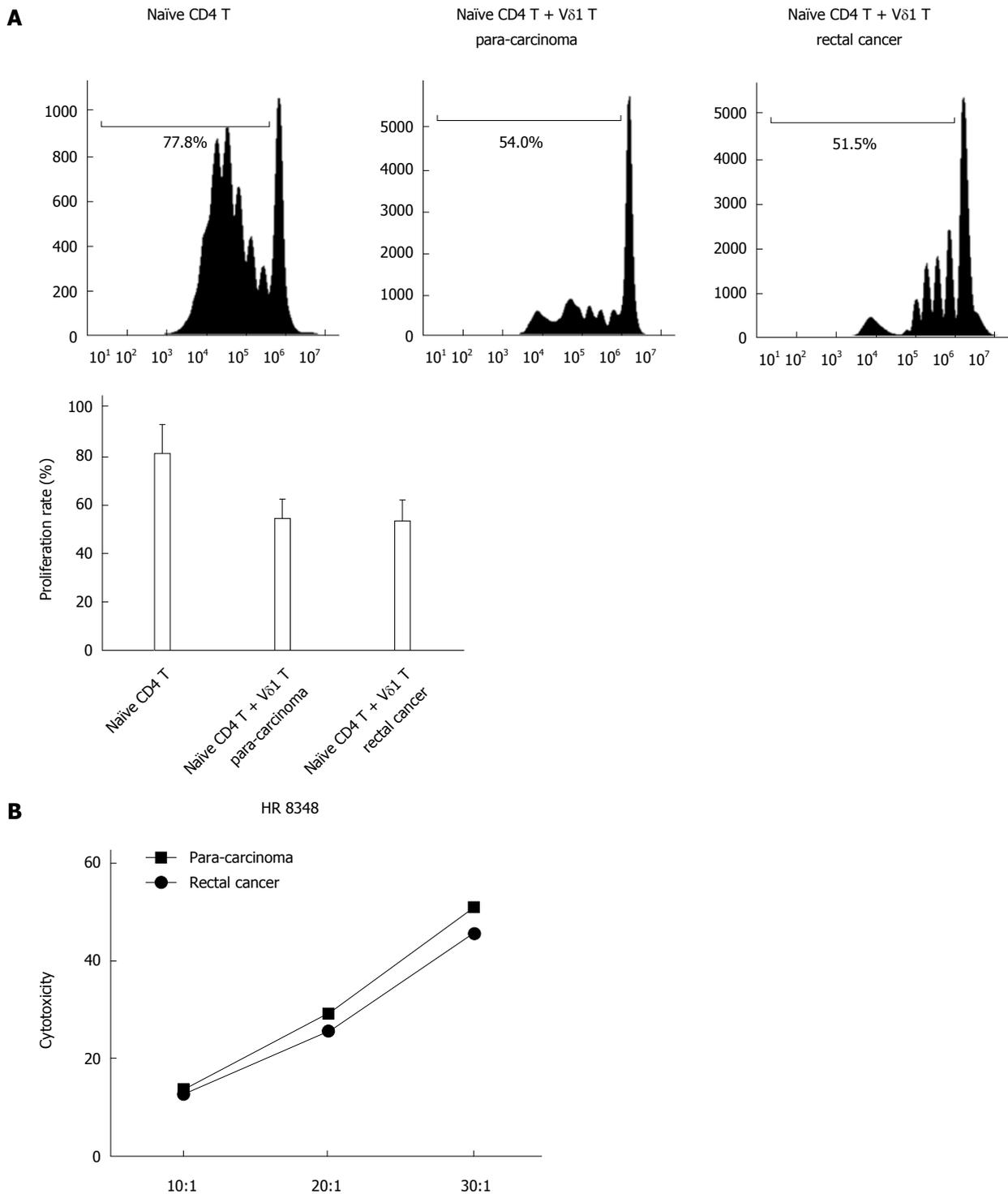


Figure 4 Functional analysis of Vδ1 and Vδ2 T cells from cancer tissues or para-carcinoma tissues. A: Fresh naïve CD4 T cells were labeled with CFSE and co-cultured with Vδ1 T cells from cancer tissues or para-carcinoma tissues; B: Lysis of HR8348 cells by Vδ2 T cells at effector to target cell ratios (E:T) of 10:1, 20:1 and 40:1.

cells mainly have an immunosuppressive function and promote cancer development^[27,28]. The infiltration of $\gamma\delta$ T cells in cancer tissues has been reported in some tumors^[29-32]. However, to date, there has been no research on the percentages of Vδ1 and Vδ2 T cells in rectal cancer tissues. In this study, we used the FACS method to analyze the percentage of Vδ1 T cells and Vδ2 T cells in tumor tissues and para-carcinoma tissues

from 20 rectal cancer patients. The results showed that the percentage of Vδ1 T cells in the rectal tumor tissues of these patients was significantly increased and positively correlated with the T stage, whereas the percentage of Vδ2 T cells in the rectal tumor tissues was significantly decreased and negatively correlated with the T stage.

We also discovered that after culture for 14 d with

1 µg/mL anti-TCR $\gamma\delta$ antibody, the major subset of $\gamma\delta$ T cells from para-carcinoma tissues was V δ 1, and the major subset from rectal cancer tissues was V δ 2. This result is consistent with the predominant subset in rectal cancer tissues being V δ 1 T cells, and the predominant subset in para-carcinoma tissues being V δ 2 T cells, and with the findings of a previous study which showed that the major $\gamma\delta$ T cells infiltrating breast cancer tissues were V δ 1 T cells^[28]. V δ 2 T cells have a more inflammatory phenotype; V δ 1 T cells have a more regulatory phenotype and have been shown to express Foxp3^[6]. V δ 2 T cells have a major cytotoxicity function, and have predominantly been investigated in tumor immunosurveillance and host defense against viral invasion^[7-10]. Our results demonstrate that tumor-infiltrating V δ 1 T cells have strong inhibitory effects, and tumor-infiltrating V δ 2 T cells have strong cytolytic activity, consistent with previous studies on the function of V δ 1 T cells and V δ 2 T cells.

The findings in this study suggest that a percentage imbalance in V δ 1 and V δ 2 T cells creates an immunosuppressant microenvironment in rectal cancer tissues, which may enable tumors to limit antitumor immunity and evade immune surveillance in rectal cancer patients. This is the first report on the percentages of V δ 1 and V δ 2 T cells in rectal cancer tissues. We demonstrate that an imbalance in V δ 1 and V δ 2 T cell percentages in cancer tissues may facilitate the development of rectal cancer. The results of this study provide a new insight into immunotherapy for rectal cancer.

COMMENTS

Background

$\gamma\delta$ T cells can be divided into two subsets: V δ 1 T cells and V δ 2 T cells. V δ 1 T cells, which have been shown to express Foxp3, have a regulatory function. V δ 2 T cells have largely been investigated in the context of tumor immunosurveillance and host defense against viral invasion. An imbalance of V δ 1 and V δ 2 T cell percentages in cancer tissues may facilitate the development of rectal cancer.

Research frontiers

$\gamma\delta$ T cells have been shown to be useful in cancer immunotherapy. They can be divided into two subsets: V δ 1 and V δ 2. Administration of V δ 2 T cells at suitable intervals after chemotherapy and zoledronate may substantially increase antitumor activity in a range of malignancies, whereas tumor-infiltrating V δ 1 T cells mainly have an immune repression function and promote cancer development. However, there has been no research on the percentages of V δ 1 and V δ 2 T cells in rectal cancer tissues.

Innovations and breakthroughs

This is the first study to report the percentages of V δ 1 and V δ 2 T cells in rectal cancer tissues. The authors demonstrate that an imbalance in the percentages of V δ 1 and V δ 2 T cells in cancer tissues may facilitate the development of rectal cancer.

Applications

This study provides new insight into immunotherapy for rectal cancer.

Peer-review

This study is meaningful and the findings that the imbalance of V δ 1 and V δ 2 T cell percentages in rectal cancer tissues are interesting, and provide new insight into immunotherapy for rectal cancer.

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ISSN 1007-9327

