

## Basic Study

# Transient receptor potential vanilloid 4-dependent calcium influx and ATP release in mouse and rat gastric epithelia

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## Abstract

**AIM:** To explore the expression of transient receptor potential vanilloid 4 (TRPV4) and its physiological meaning in mouse and rat gastric epithelia.

**METHODS:** RT-PCR and immunochemistry were used to detect TRPV4 mRNA and protein expression in mouse stomach and a rat normal gastric epithelial cell line (RGE1-01), while  $\text{Ca}^{2+}$ -imaging and electrophysiology were used to evaluate TRPV4 channel activity. ATP release was measured by a luciferin-luciferase assay. Gastric emptying was also compared between WT and TRPV4 knockout mice.

**RESULTS:** TRPV4 mRNA and protein were detected in mouse tissues and RGE1-01 cells. A TRPV4-specific agonist (GSK1016790A) increased intracellular  $\text{Ca}^{2+}$  concentrations and/or evoked TRPV4-like current activities in WT mouse gastric epithelial cells and

RGE1-01 cells, but not TRPV4KO cells. GSK1016790A or mechanical stimuli induced ATP release from RGE1-01 cells while TRPV4 knockout mice displayed delayed gastric emptying *in vivo*.

**CONCLUSION:** TRPV4 is expressed in mouse and rat gastric epithelium and contributes to ATP release and gastric emptying.

**Key words:** Transient receptor potential vanilloid 4; Stomach; Gastric emptying; ATP

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**Core tip:** A mechano-sensitive ion channel, transient receptor potential vanilloid 4 (TRPV4), is expressed in gastric epithelium and contributes to ATP release and gastric emptying. These findings suggest that gastric distension stimulates TRPV4 on gastric epithelium and released ATP stimulates sub-epithelial nerve fibers or acts on visceral smooth muscles. TRPV4 might be a promising novel diagnostic and therapeutic target for functional gastric disorders.

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## INTRODUCTION

The transient receptor potential vanilloid 4 channel (TRPV4) is a non-selective cation channel that is involved in various cellular functions<sup>[1]</sup> and is activated by several physical and chemical stimuli, including mechanical stimuli, endogenous arachidonic acid metabolites (epoxyeicosatrienoic acids)<sup>[2]</sup>, and heat. TRPV4 is also activated by the specific agonist GSK1016790A that elicits whole-cell currents in mouse and rat TRPV4-expressing cells with EC<sub>50</sub> values of 18.5 and 10 nmol/L, respectively<sup>[3]</sup>. TRPV4 is widely expressed throughout the body, including gastrointestinal tract epithelium and the esophagus<sup>[4]</sup>. Although the physiological function of TRPV4 expression in intestinal epithelial cells is unknown, TRPV4 activation in these cells causes increases in intracellular calcium concentrations, chemokine release, and incidence of colitis<sup>[5]</sup>, as well as increased paracellular permeability<sup>[6]</sup>. Furthermore, TRPV4 antagonists are promising therapeutic options for colitis<sup>[7,8]</sup>. However, TRPV4 expression in the gastric epithelium awaits evaluation.

In addition to its function as an intracellular energy donor, ATP is recognized as an important signaling molecule that mediates diverse biological effects *via*

cell surface receptors: the purinergic receptors<sup>[9]</sup>. ATP is released by neurons of the central, peripheral, and enteric nervous system<sup>[10,11]</sup>, and acts as a non-adrenergic non-cholinergic (NANC) neurotransmitter that causes different responses or effects (either excitatory or inhibitory depending on the P2 receptor subtype upon which they act as well as the animal species under study). Several studies showed that purinergic neurotransmission (assuming that gut neurons are the sole source of released ATP) affects gastric motility<sup>[12]</sup>. Recent reports showed that ATP is also released from non-neuronal tissues and has an effect on tissue function. Moreover, we found that ATP release in the esophagus and urothelium was mediated by TRPV4 stimulation<sup>[4,13,14]</sup>. However, there are no data concerning whether TRPV4 is expressed in the stomach and, if so, whether TRPV4 stimulation plays a role in mediating ATP release. Therefore, this study explored the morphological (RT-PCR and immunostaining) and functional (Ca<sup>2+</sup>-imaging, patch clamp and gastric emptying) expression of TRPV4 in mouse and rat stomach with special focus on gastric epithelium.

## MATERIALS AND METHODS

### Animals

Eight week-old male C57BL/6NCr (SLC) and TRPV4-knockout (TRPV4KO) mice<sup>[15]</sup> weighing between 23 and 25 g were housed in a controlled environment (12-h light/12-h dark cycle; room temperature, 22-24 °C; 50%-60% relative humidity) with free access to food and water. All procedures involving the care and use of animals were approved by The Institutional Animal Care and Use Committee of the National Institutes of Natural Sciences.

### Cell lines

RGE1-01 is an immortalized rat gastric mucosal cell line that shows distinct cell differentiation types and preserves some epithelial cell characteristics. RGE1-01 cells were maintained at 34 °C in Dulbecco's modified Eagle medium supplemented with 10% heat-inactivated fetal bovine serum, 100 µg/mL streptomycin and 100 U/mL penicillin with the addition of ITES (see reference<sup>[16]</sup> for details).

### Acute isolated mouse gastric epithelium

WT and TRPV4KO mice were sacrificed by cervical dislocation. The stomachs were washed in cold (4 °C) PBS (-) and then incubated in trypsin solution (Invitrogen) at 4 °C for 1 h. Gastric epithelial cells were harvested and plated on CELL-TAK (BD Biosciences)-coated glass cover slips and used for Ca<sup>2+</sup>-imaging and patch clamp experiments.

### Reverse transcription PCR analysis

RT-PCR was performed as previously described<sup>[4,17]</sup>. Total RNA (1 µg) was isolated using the RNeasy Mini

**Table 1** Primer sequences for RT-PCR

Primer name	Sequence (5'→3')
mTRPV4-F	ACAACACCCGAGAGAACACC
mTRPV4-R	CCCAAACCTTACGCCACTTGT
mGAPDH-F	TGAAGGGTGGAGCCAAAAGG
mGAPDH-R	GGAAGAGTGGGAGTTGCTGTG
rTRPV4-F	CCTGGCAGGGATCGAGGCCT
rTRPV4-R	GGATGGTGGTGGCCCACTGC
rGAPDH-F	GCCAAGGCTGTGGGCAAGGT
rGAPDH-R	GAGCAATGCCAGCCCCAGCA

Kit (Qiagen, Courtaboeuf, France) and measured with a NanoDrop device (Thermo Fisher Scientific Inc., Wilmington, United States). Genomic DNA was eliminated in the process of reverse transcription (QuantiTect Reverse Transcription Kit, QIAGEN). PCR was performed using rTaq DNA polymerase (TaKaRa) in an iCycler (Bio-Rad) with specific primer sets (Table 1).

### Immunocytochemistry

Immunocytochemistry was performed as previously described<sup>[4]</sup> using the antibodies summarized in Table 2. For section preparation, mouse stomachs were fixed at 4 °C for 6 h. Tissues were placed in PBS-sucrose and embedded in OCT compound (Tissue Tek, Elkhart, IN, United States). Non-specific antibody binding was reduced by incubation in BlockAce (Yukijirushi, Sapporo, Japan) for 1 h at room temperature prior to antibody exposure. Preparations were analyzed using a confocal laser scanning microscope (LSM 700, Carl Zeiss). For immunocytochemistry, RGE1-01 cells were fixed at 4 °C for 20 min with the same fixative. Bovine serum albumin (3% BSA; Sigma) was used as a blocking solution.

### Ca<sup>2+</sup>-imaging

Fura-2 fluorescence was measured in primary mouse gastric epithelial cells and RGE1-01 cells with a standard bath solution containing 140 mmol/L NaCl, 5 mmol/L KCl, 2 mmol/L MgCl<sub>2</sub>, 2 mmol/L CaCl<sub>2</sub>, 10 mmol/L HEPES, and 10 mmol/L glucose at pH 7.4 (adjusted with NaOH) at 25 °C. Results are presented as ratios of fluorescence intensities obtained with fura-2 emissions at 340 nm and 380 nm. GSK1016790A<sup>[3]</sup> and ionomycin (both from Sigma) were used as a TRPV4 agonist and a positive control, respectively.  $F_{340}/F_{380}$  was calculated and acquired with an image processing system (IP-Lab, Scanalytics Inc., Rockville, MD or AQUA COSMOS, Hamamatsu Co., Japan) and ImageJ software (<http://rsb.info.nih.gov/ij/>). Changes in ratio ( $\Delta$ ) were calculated by subtracting the mean basal values from peak values. Since the degree of responses (strong, weak, or no response) to GSK varied from cell to cell in WT gastric epithelial cells, we expressed the observed changes in all ionomycin-responsive cells as a ratio between WT and TRPV4 knockout mice ( $\Delta$ ). We evaluated 53 and 41 ionomycin-responsive cells from six WT mice

**Table 2** Primary and secondary antisera for immunochemistry

Tissue antigen/host	Dilution	Source
TRPV4/rabbit	1:500	B. Nilius, or Abcam
Goat anti-rabbit IgG-Alexa488	1:1500	Invitrogen, Inc.

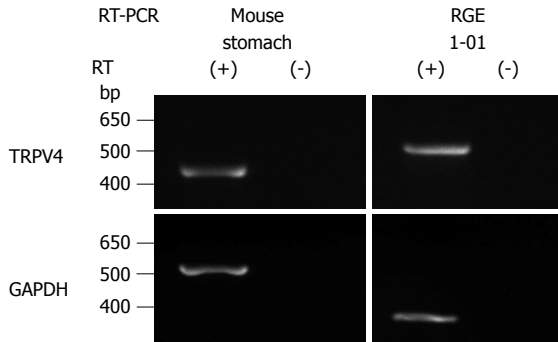
and five TRPV4 knockout mice, respectively. Given the variations in response times, and that some WT cells responded 30 s after GSK application, we decided to incubate cells with GSK for 90 s.

### Electrophysiology

The standard bath solution was the same as that used in the Ca<sup>2+</sup>-imaging experiments. Pipette solutions for whole-cell recordings contained 140 mmol/L KCl, 5 mmol/L EGTA and 10 mmol/L HEPES, pH 7.4. Whole-cell recording data on primary gastric epithelial cells three hours after insolation and RGE1-01 were sampled at 10 kHz and filtered at 5 kHz for analysis (Axon 200B amplifier with pCLAMP software, Molecular Devices, Foster City, CA, United States). Voltage ramp-pulses from -100 mV to +100 mV (500 ms) were applied every 5 s to generate an I-V curve.

### ATP release measurement

ATP concentrations released from RGE1-01 rat gastric epithelial cells cultured in 12-well plates or stretch silicon chambers (STB-10-04 from STREX Inc., Osaka, Japan) were measured by a luciferin-luciferase assay (ATP Bioluminescence assay kit CLS II, Roche Diagnostics) and a luminometer (Lumat LB 9507, Berthold Technologies, Japan), using a previously described method that was slightly modified<sup>[4]</sup>. For chemical stimuli, cells cultured to 70%-80% confluence and incubated in 500  $\mu$ L bath solution for 30 min at room temperature (25 °C) were used to measure basal ATP release. The superfusate was collected and replaced gently with another 500  $\mu$ L of bath solution with or without the TRPV4 agonists GSK1016790A or 5,6-EET. The superfusate was collected after 15 min and the ratio of released ATP (15 min stimulation/30 min basal condition) was calculated. An aliquot (200  $\mu$ L) of superfusate was then mixed with 200  $\mu$ L luciferin-luciferase reagent for luminometric ATP measurements. For mechanical stimuli, stretching was quantitatively applied with a STB-10 stretch machine (STREX Inc.) to RGE1-01 cells cultured on a silicon chamber. Three minutes after chamber placement in the stretch machine, the superfusate was washed away to exclude artificial ATP release and replaced with 500  $\mu$ L of bath solution for basal ATP measurement. After a further 3 min, the superfusate was collected and replaced, whereupon mechanical stimuli was applied for 3 min and the ratio of released ATP (3 min stimulation/3 min basal condition) was calculated. To block TRPV4 channels, cells were pre-treated with the specific TRPV4 antagonist HC 067047 (Sigma, 1  $\mu$ mol/L) for 3 min<sup>[18]</sup>.



**Figure 1** *TRPV4* mRNA expression in mouse stomach and RGE1-01 cells. *TRPV4* and *GAPDH* mRNA levels were examined with (+) and without (-) RT reaction. The expected sizes of the amplified fragments for *TRPV4* and *GAPDH* were 404 and 545 bp for mouse and 524 and 268 bp for rat, respectively. *TRPV4* mRNA was detected in mouse stomach and RGE1-01 cells (RGE1-01: normal rat gastric epithelial cell line). Band positions differed due to the use of different primers.

### Gastric emptying

Eight-week-old WT and TRPV4KO male mice were used with a modified version of previously reported methods<sup>[17]</sup>. Gastric emptying was determined by transit of a test meal containing phenol red. Mice were fasted for 14 h with *ad libitum* water before the experiment. Five mg/kg (200  $\mu$ L) of the test meal was administered into the stomach using a feeding needle. Fifteen minutes later, the mice were euthanized by cervical dislocation and the gastrointestinal tract was removed. The stomach was minced and the remaining phenol red concentration was measured. Gastric emptying was expressed as mean  $\pm$  SEM for each group.

### Data analysis

Values for  $\text{Ca}^{2+}$ -imaging, patch-clamp experiments, ATP measurements, and gastric emptying are presented as mean  $\pm$  SEM from three or more independent experiments. A Student's *t*-test or non-parametric Bonferroni-type multiple comparison was used. Significance was accepted for  $P < 0.05$ .

## RESULTS

### *TRPV4* expression in mouse and rat gastric epithelia

Given that TRPV4 was shown to be expressed in the esophagus and intestinal epithelia<sup>[4-6,19,20]</sup>, we examined *TRPV4* mRNA expression in mouse stomach and a rat gastric epithelial cell line, RGE1-01. *TRPV4* mRNA was detected in mouse stomach and RGE1-01 cells (Figure 1). We next examined TRPV4 protein expression in mouse and the RGE1-01 cells. A strong homogenous immunofluorescent signal was confined to the epithelial cell layer of the WT mouse gastric corpus and antrum but not in cells from TRPV4KO mice (Figure 2A). Meanwhile, Z-stack images obtained by confocal microscopy of RGE1-01 cells displayed apical

TRPV4 expression (Figure 2B).

### *TRPV4*-mediated increase in cytosolic $\text{Ca}^{2+}$ ( $[\text{Ca}^{2+}]_i$ ) in mouse primary gastric epithelial cells

To confirm functional TRPV4 expression in primary gastric epithelial cells and RGE1-01 cells, we examined the response to the reported specific TRPV4 agonist, GSK1016790A (GSK)<sup>[3]</sup>, using a fluorescent  $\text{Ca}^{2+}$ -imaging system (10  $\mu$ mol/L, fura-2/AM). Response traces of  $[\text{Ca}^{2+}]_i$  for WT and TRPV4KO gastric epithelial cells in the presence of GSK (100 nmol/L) showed that almost all cells isolated from WT stomach responded to GSK (Figure 3A) and the  $[\text{Ca}^{2+}]_i$  increases were significantly larger in WT cells compared to TRPV4KO cells (Figure 3B). This finding suggests that the majority of gastric epithelial cells expressed TRPV4 and  $[\text{Ca}^{2+}]_i$  responses to GSK were TRPV4 specific.

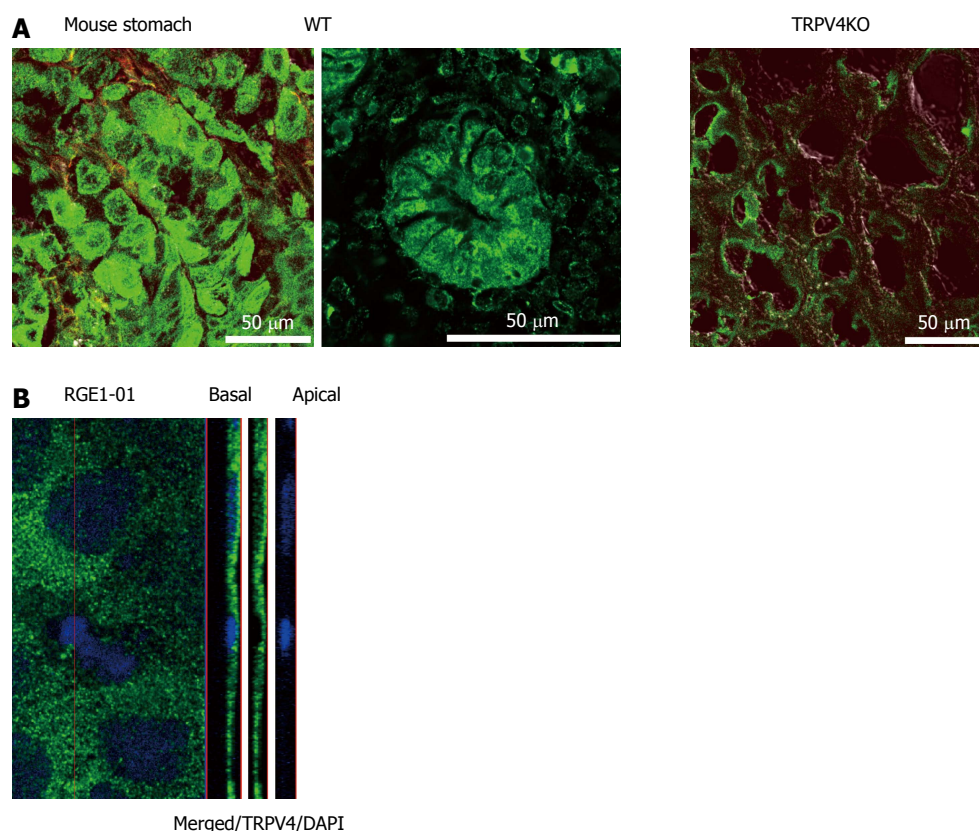
### *TRPV4*-mediated current responses in mouse primary gastric epithelial cells and RGE1-01 cells

We next performed patch-clamp experiments with acute isolated mouse gastric epithelial cells in the presence of GSK (300 nmol/L) and observed inward current responses with an outwardly rectifying IV-relationship in WT but not TRPV4KO cells (Figure 4A)<sup>[3]</sup>. Current responses were observed in all 5 trials with WT gastric epithelial cells but were completely absent with TRPV4KO cells, which indicates that the majority of gastric epithelial cells expressed TRPV4. Similar chemical stimulation with GSK (300 nmol/L) induced TRPV4-like current responses in the rat gastric epithelial cell line, RGE1-01 (Figure 4B). These data strongly indicated functional expression of TRPV4 in mouse and rat gastric epithelial cells.

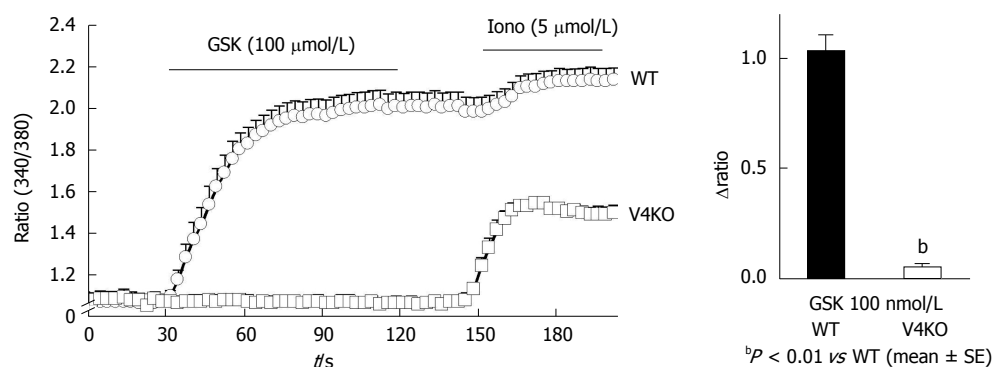
### *TRPV4* activators induced ATP release from RGE1-01 cells

Mechanical stimuli reportedly activate TRPV4 expressed in esophageal keratinocytes that in turn leads to increased ATP release<sup>[4]</sup>. To examine whether TRPV4 stimulation has a similar effect in gastric epithelium, we measured ATP release in chemically- or mechanically-stimulated RGE1-01 cells using a luciferin-luciferase assay. TRPV4 agonists GSK1016790A (GSK, 100 nmol/L) or 5,6-EET (500 nmol/L)<sup>[2]</sup> significantly increased ATP release in RGE1-01 cells (Figure 5A, 2- to 3-fold higher vs control,  $P < 0.05$ ). Additionally, 120% lateral stretch applied for 3 min to RGE1-01 cells cultured on a silicon chamber induced significantly higher amounts of ATP release (Figure 5B, about 2-fold vs without stretch (control),  $P < 0.05$ ), and these responses were inhibited by the specific TRPV4 inhibitor HC 067047 (1  $\mu$ mol/L) (stretched cells showed no detachment over the 3-min stretch period). These results suggested that chemical and mechanical stimuli-induced ATP release in RGE1-01 cells was mediated by TRPV4 channel activation.





**Figure 2** TRPV4 protein expression in mouse stomach and RGE1-01 cells. A: TRPV4 expression was homogeneously observed in WT but not TRPV4KO mouse gastric epithelium. Bars indicate 50  $\mu$ m; B: Z-stack image of RGE1-01 cells demonstrated apical TRPV4 expression.



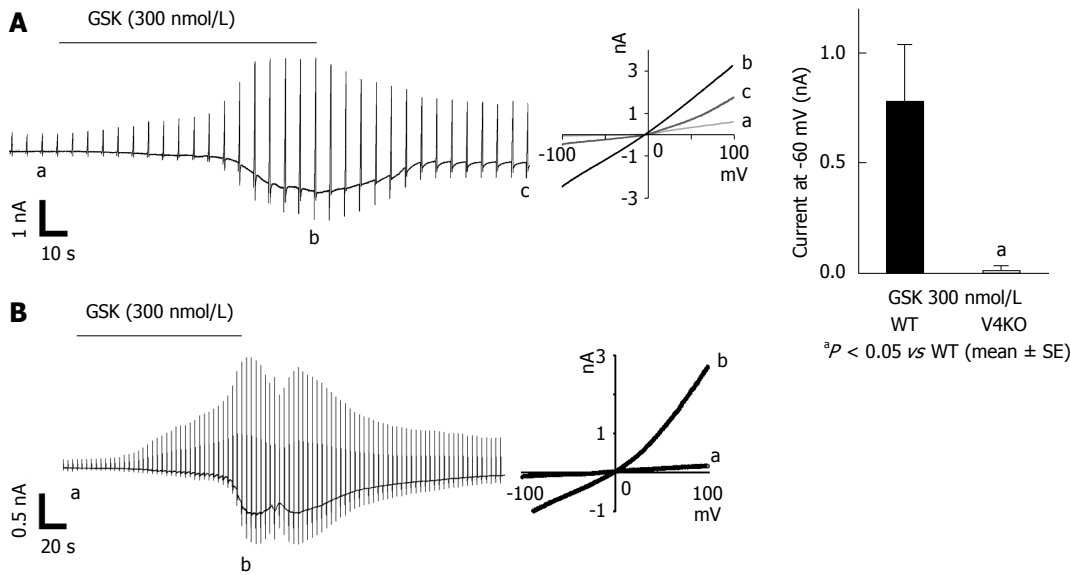
**Figure 3** TRPV4-mediated increases in cytosolic  $\text{Ca}^{2+}$  ( $[\text{Ca}^{2+}]_i$ ) in mouse primary gastric epithelial cells. A:  $[\text{Ca}^{2+}]_i$  changes (340/380 ratio) in response to the TRPV4 specific agonist GSK1016790A (GSK, 100 nmol/L) in WT or TRPV4KO (V4KO) primary gastric epithelial cells (mean  $\pm$  SEM). Ionomycin (iono, 5  $\mu$ mol/L) was used as a positive control. Bars indicate the period of chemical application; B: GSK significantly increased  $[\text{Ca}^{2+}]_i$  in WT cells (means  $\pm$  SD;  $1.03 \pm 0.07$ ,  $n = 20$ ) compared to TRPV4KO cells ( $0.05 \pm 0.01$ ,  $n = 20$ ) ( $^bP < 0.01$  vs WT).

### Delayed gastric emptying in TRPV4KO mice

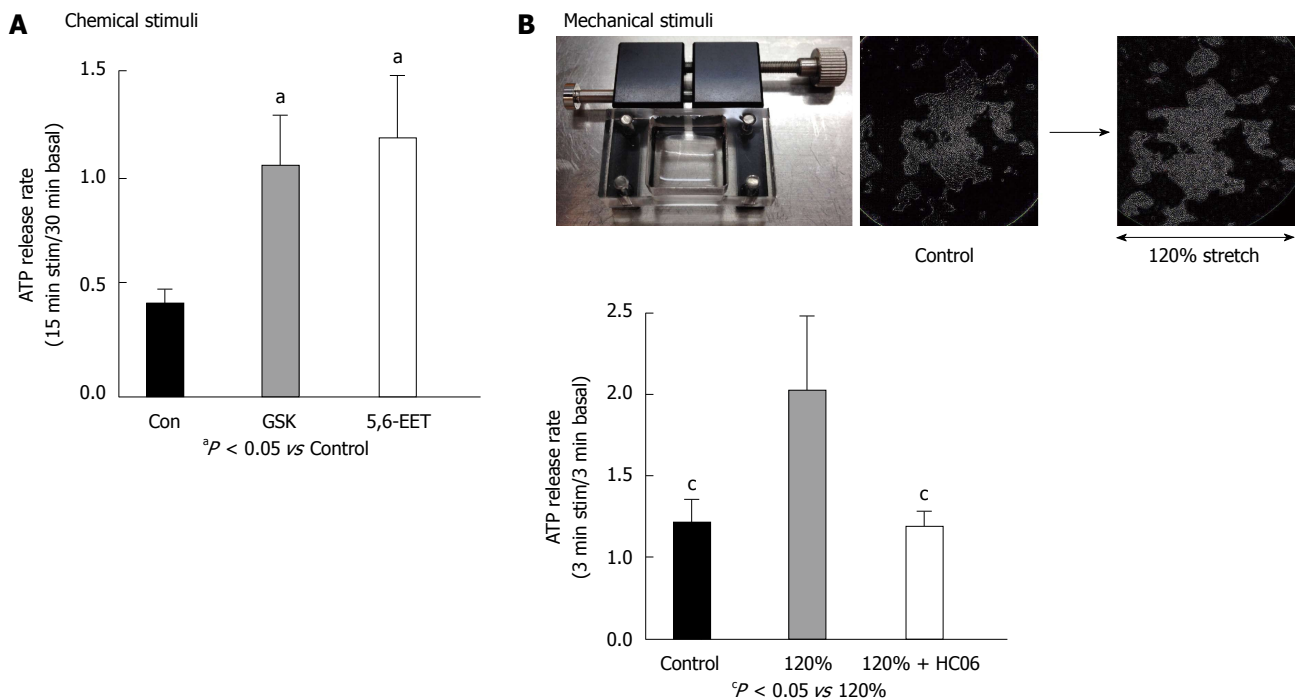
Since TRPV4 has been shown to sense chemical and mechanical stimuli and contribute to ATP release from gastric epithelial cells, we hypothesized that TRPV4KO mice would exhibit altered gastric motility. To evaluate the physiological role of TRPV4 expressed in the gastric epithelium, we performed an *in vivo* experiment to compare gastric emptying rates of WT and TRPV4KO mice. Gastric emptying rates in TRPV4KO mice were about 2/3 of those in WT (Figure 6), suggesting that TRPV4 contributes to gastric motor function.

## DISCUSSION

We identified morphological and functional TRPV4 expression in mouse gastric epithelial cells as well as the rat gastric epithelial cell line RGE1-01 (Figures 1-4). Furthermore, we demonstrated that chemical and mechanical stimuli can induce TRPV4-dependent ATP release from RGE1-01 cells (Figure 5), and that stimulation of gastric epithelial TRPV4 enhances gastric emptying *in vivo* (Figure 6). Using immunohistochemistry,  $\text{Ca}^{2+}$  imaging,



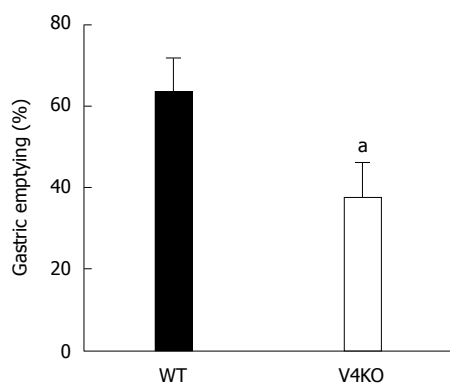
**Figure 4** TRPV4-mediated current responses in mouse primary gastric epithelial cells and RGE1-01 cells. A: GSK (300 nmol/L) evoked inward current responses in WT primary gastric epithelial cells. Currents in response to ramp-pulses at points a, b and c (left in panel B) are shown (middle), with a strongly outwardly rectifying current-voltage relationship. Significantly larger inward currents at -60 mV were obtained from WT cells (means ± SEM;  $0.76 \pm 0.27$  nA,  $n = 5$ ) than in TRPV4KO cells ( $0.01 \pm 0.00$  nA,  $n = 5$ ) (<sup>a</sup>*P* < 0.05 vs WT). B: Similar current responses were also obtained in RGE1-01 cells.



**Figure 5** TRPV4 activator-induced ATP release in the RGE1-01 cells. A: GSK1016790A (GSK, 100 nmol/L) or 5,6-EET (500 nmol/L) induced significantly higher ATP release in RGE1-01 cells (<sup>a</sup>*P* < 0.05 vs Control). B: Mechanical stimuli were quantitatively applied with a stretch apparatus. Microscopy images demonstrated that cells were stretched laterally without detachment. A 120% stretch induced significantly higher amounts of ATP release from RGE1-01 cells [<sup>c</sup>*P* < 0.05 vs 0% stretch (control)] that could be inhibited by pre-treatment with specific TRPV4 antagonist HC 067047 (1  $\mu$ mol/L). TRPV4: Transient receptor potential vanilloid 4.

and electrophysiology, we found that the majority of mouse gastric epithelial cells exhibited abundant TRPV4 expression and responded to TRPV4 agonists, suggesting that TRPV4 is a candidate mechanoreceptor in gastric epithelial cells. In fact, ATP release was several hundred nmol/L in our *in vitro* study, suggesting that the corresponding ATP concentration *in vivo* might be estimated to be several  $\mu$ mol/L, which

would be sufficient to activate the P2 receptor present in the wall of mouse stomach<sup>[21,22]</sup>. These results suggested the hypothesis that luminal distension stimulates TRPV4 on gastric epithelial cells that in turn release ATP. The released ATP either stimulates subepithelial sensory nerve fibers that form the afferent limb of short or long gastrointestinal reflex arcs or acts directly on visceral smooth muscles expressing



**Figure 6 Delayed gastric emptying in TRPV4 knockout mice.** Gastric emptying rates *in vivo* in TRPV4KO mice were significantly delayed relative to WT mice ( $^aP < 0.05$ , vs WT,  $n = 7-9$ ).

purinergic receptors. The first possibility is supported by morphological evidence showing that purinergic receptors, mainly P2X2 and P2X3, were identified on putative gastric mechanosensing structures, including the vagal afferent intraganglionic laminar endings that are located in close proximity to the epithelium<sup>[23,24]</sup>. These vagal afferents form the afferent limb of the central vago-vagal reflex (long reflex arc) and are known to increase gastric motility following stimulation<sup>[25]</sup>. The hypothesis is also supported by findings from a previous study wherein P2X3-knockout mice show a blunted neural response to gastric distension and no differences in distension-evoked ATP release between knockout and control mice<sup>[26]</sup>. The released ATP could also trigger a local reflex arc intrinsic to the stomach wall, which is supported by results from a previous study wherein ATP was shown to induce tetrodotoxin-sensitive contraction responses mediated by neuronal P2X receptors in an *in vitro* whole-stomach preparation<sup>[27]</sup>.

The possibility that ATP released from gastric epithelium could directly stimulate purinergic receptors expressed on gastric visceral smooth muscles is rather unlikely considering the short half-life of ATP and the distance that ATP must cross while diffusing from the gastric epithelium to visceral smooth muscles. Moreover, gastric smooth muscles are known to functionally express P2Y receptors that mediate relaxation in response to ATP<sup>[27]</sup> and would be expected, upon stimulation, to delay gastric emptying, which is opposite to our current findings. This outcome further decreases the likelihood of a direct effect for ATP released from gastric epithelium on smooth muscles. However, the purinergic signaling pathway that mediates gastric distension-induced epithelial TRPV4 stimulation requires further future characterization.

In conclusion, TRPV4 is morphologically and functionally expressed in mouse and rat gastric epithelia and contributes to ATP release and gastric emptying. Our results suggest that TRPV4 could be a promising novel diagnostic and therapeutic target for functional gastrointestinal disorders.

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## COMMENTS

### Background

The transient receptor potential vanilloid 4 channel (TRPV4) is a non-selective cation channel that is activated by mechanical stimuli. ATP has been recognized as an important signaling molecule via cell surface ATP receptors. This study explored TRPV4 expression in mouse and rat stomach and whether the stimulation mediated ATP release.

### Research frontiers

The authors have reported that ATP release in the esophagus and urothelium is mediated by TRPV4 stimulation.

### Innovations and breakthroughs

This is the first study showing TRPV4 expression and ATP release by its stimulation in the mouse stomach and/or rat gastric epithelial cells.

### Applications

These data suggested the hypothesis that luminal distension stimulates TRPV4 on gastric epithelial cells that in turn release ATP. However TRPV4 expression in human gastric epithelium and the purinergic signaling pathway requires further evaluation.

### Terminology

TRPV4 is a non-selective cation channel, that is activated by several physical and chemical stimuli, including mechanical stimuli. The channel activation increases intracellular  $Ca^{2+}$  concentration and elicits whole-cell currents in TRPV4-expressing cells.

### Peer-review

The manuscript describes an original research performed in mouse stomach and in rat epithelium cell line focusing on the expression and function of TRPV4 ion channels. The study concept is based on previous findings of the authors regarding TRPV4-mediated ATP release on the esophagus. The series of experiments in this manuscript demonstrated delayed gastric emptying in TRPV4 knockout mice compared to their WT littermates, as well as TRPV4 expression in mRNA and protein levels in both mouse and rat gastric epithelial cell line. In general, the idea is interesting, the various morphological and functional methods are sophisticated, the figures are demonstrative.

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