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**Monitoring photodynamic therapy of head and neck malignancies with optical spectroscopies**

Sunar U. Optical monitoring of clinical PDT of H and N

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**Abstract**

In recent years there has been significant developments in photosensitizers, light sources and light delivery systems that have allowed decreasing the treatment time and skin phototoxicity resulting in more frequent use of photodynamic therapy (PDT) in the clinical settings. Compared to standard treatment approaches such as chemo-radiation and surgery, PDT has much reduced morbidity for head and neck malignancies and is becoming an alternative treatment option. It can be used as an adjunct therapy to other treatment modalities without any additive cumulative side effects. Surface illumination can be an option for pre-malignant and early-stage malignancies while interstitial treatment is for debulking of thick tumors in the head and neck region. PDT can achieve equivalent or greater efficacy in treating head and neck malignancies, suggesting that it may be considered as a first line therapy in the future. Despite progressive development, clinical PDT needs improvement in several topics for wider acceptance including standardization of protocols that involve the same administrated light and photosensitizer doses and establishing quantitative tools for PDT dosimetry planning and response monitoring. Quantitative measures such as optical parameters, photosensitizer concentration, tissue oxygenation and blood flow are essential for accurate PDT dosimetry as well as PDT response monitoring and assessing therapy outcome. Unlike conventional imaging modalities like MRI, novel optical imaging techniques can quantify PDT-related parameters without any contrast agent administration and enable real-time assessment during PDT for providing on-line feedback to clinicians. Ongoing developments in optical imaging offer the promise of optimization of PDT protocols with improved outcomes.

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**Key words:** Head and neck cancer; Photodynamic therapy; Monitoring and predicting response; Blood flow; Oxygenation; Oxygen metabolism; Diffuse optical imaging

**Core tip:**

Most treatment approaches including chemo-radiation and surgery can induce prolonged morbidity and functional loss resulting in severe impairment of patients’ quality of life. Photodynamic therapy (PDT) is an emerging alternative treatment option without any significant accumulative side effects due to targeted light illumination and preferential accumulation of photosensitizers. However, PDT has not found widespread applications at the clinic mainly due to variable responses that originated from unstandardized treatment protocols such as different light and photosensitizer doses. Novel optical imaging techniques can quantify PDT-dosimetry related parameters such as local light and photosensitizer dose in tissue and PDT response related parameters such as tissue oxygenation and blood flow noninvasively without any contrast agent administration, thereby providing real-time feedback about the treatment efficacy for optimizing and standardizing PDT.

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**INTRODUCTION**

Head and neck malignancies refer to malignancies arising from the oral cavity, pharynx, nasal cavity and sinuses[[1-3](#_ENREF_1)]. Head and neck squamous cell carcinoma (HNSCC), constituting approximately 90% of malignancies in the head and neck region, remains the fifth most common form of cancer worldwide with an incidence of approximately 800000 new cases per year[[4](#_ENREF_4)]. Most of these tumors may be attributed to risk factors such as tobacco and alcohol consumption. HNSCC is a heterogeneous disease with different stages ranging from benign squamous hyperplasia, dysplasia, carcinoma *in situ* to invasive carcinoma[[3](#_ENREF_3)]. Early stage diagnosis and treatment of HNSCC increases the likelihood of successful treatment and improves patients’ quality of life, lowers risk of mortality and health costs[[5](#_ENREF_5),[6](#_ENREF_6)].

Substantial efforts concentrate on early detection with fair success, but still many patients present with clinically evident tumors that require effective treatment[[7](#_ENREF_7)]. Several treatment options are available including surgery, chemotherapy, radiation therapy or combinations thereof[[8](#_ENREF_8)]. In spite of improvements in these treatment modalities, they have their own limitations. For example, surgery may require resection of vital tissue such as part of the tongue resulting in functional loss. On the other hand, organ-preserving surgery can result in high recurrence rates. Nonsurgical management with chemo and radiation therapies to improve local-regional disease resulting in only modest or suboptimal improvements in survival but with significantly high cost side effects including speech and swallow function[[9](#_ENREF_9)]. These conventional therapies may induce permanent vasculature dysfunction and necrosis, severe toxicities and irreversible injuries to non-tumor tissue such as the oral mucosa and the salivary glands, often resulting in morbidity and severe impairment of patients’ quality of life. Further, normal tissue toxicity such as mucositis, bleeding and imflammation may lead to changes in applied dose quantity, and treatment re-schedule, which may affect treatment efficacy and outcome. For these reasons, an alternative treatment modality that would be effective, safe, repeatable, minimally invasive and non-surgical is desired for the management of head and neck malignancies.

Photodynamic therapy (PDT) uses light to activate a photosensitizer in the presence of oxygen for local tissue destruction, has potential in these respects and is particularly attractive due to its significant level of normal tissue preservation and its repeatability without cumulative side effects[[10](#_ENREF_10)]. It has potential impact particularly for cases with multiple lesions and wide-spread early stage head and neck diseases (*e.g.*, leukoplakia, invasive carcinoma) in the oral cavity[[11](#_ENREF_11)]. However, PDT has not found widespread applications at the clinic mainly due to variable responses that originated from unstandardized treatment protocols such as different light and photosensitizer doses. Optical imaging can quantify local light and photosensitizer dose in tissue and monitor PDT; and therefore can provide feedback about the treatment efficacy. Thus, we expect optical imaging modalities will help in optimizing and standardizing PDT. Below we will detail PDT treatment and optical imaging for monitoring and ultimately predicting the PDT response.

**CLINICAL PHOTODYNAMIC THERAPY**

PDT is an emerging treatment option for many malignancies including head and neck. It is minimally invasive with much less side effects compared to conventional therapies. Since it does not have any significant accumulative side effects, it can be repeated many times and be applied before or after chemotherapy, radiation therapy. It can also be used as an adjuvant therapy to these therapies and surgery to eliminate residual microscopic tumor cells. PDT light can be delivered at the surface for wide and superficial malignancies and pre-malignancies such as mucosal dysplasia and carcinoma in situ in the oral cavity. Interstitial light delivery is applied in treating thick and deep tumors for the aim of debulking tumors as an adjuvant to surgery.

***Basics of PDT***

PDT efficacy depends on three main elements: a sufficient amount of light, photosensitizing drug (also called photosensitizer) and available oxygen in tissue. The photosensitizer (PS) is activated during light illumination and the active PS reacts with molecular oxygen to produce singlet oxygen that induces direct cell killing, vascular destruction and immune response[[12](#_ENREF_12),[13](#_ENREF_13)]. Most PSs are administered systematically but some can be applied topically for head and neck lesions in the oral cavity and nonmelanoma skin tumors. After a specific time, depending on the PS itself, PS accumulates specifically more in the diseased site compared to normal and surrounding periphery sites. Tumor to normal tissue contrast is generally 2-3 fold with a passive targeting mechanism, but even 10-fold contrast has been reported[[14](#_ENREF_14)]. At the optimal time point of accumulation, a specific wavelength of light depending on the optical absorption properties of the PS is shined at a predetermined power to activate the PS to create a photodynamic reaction. Due to specific accumulation of the PS and localized light illumination, PDT is a local therapy rather than a systemic therapy like chemotherapy. The treatment volume depends on both photosensitizer and light penetration depth. For example, for the cases of Photofrin®, which is the first FDA-approved photosensitizer that was developed here at Roswell Park Cancer Institute, light illumination is at approximately 630 nm with a penetration depth of 5 mm or less. Thus, Photofrin® has been in use worldwide to treat early stage carcinomas in many organs including the head and neck.

***Superficial and interstitial PDT approaches for head and neck diseases***

Previous studies have shown that PDT is safe and effective in the treatment of early carcinomas of the head and neck[[2](#_ENREF_2),[10](#_ENREF_10),11,[13](#_ENREF_13),[14-35](#_ENREF_15)]. PDT is an excellent choice for the early-stage malignancies since local treatment and limited light penetration eliminates the side effects that can occur in the sensitive areas of the oral cavity such as soft palate. Lasers are the choice for the light sources and laser light is delivered *via* surface illumination by using a micro-lens as shown in Figure 1A. For deeper and thicker tumors, however, superficial illumination is not suitable. In this case, light is delivered by feeding laser fibers through needles placed directly into the tumor (Figure 1B). This approach is very similar to brachytherapy or interstitial radiotherapy[[36](#_ENREF_37),[37](#_ENREF_38)].

**CHALLENGES**

One of the main challenges of PDT is treating deeper and thicker tissues. However, this is not an issue for superficial malignancies. Pain management is a frequently reported challenge[[38](#_ENREF_39)]. Another common side effect of PDT is the long-term skin photosensitivity, especially for the cases of systemic administration of PSs such as Photofrin® (porfimer sodium). ALA-PDT is another widely used treatment option for early stage malignancies with much reduced skin photosensitization, but with the drawback of severe pain during treatment, often necessitating anesthesia. Therefore, the development of PSs that do not induce long-term photosensitivity, produce durable results and are patient friendly is of significant clinical benefit. In this respect the second generation PSs, such as Photoclor (HPPH) used in our clinical trials, have shown clinical promise with their improved efficacy, higher penetration depths and significantly less skin photosensitivity.

Variable outcomes are the main roadblock to wider use of PDT. The lack of standardized protocols with the same light and photosensitizer type and doses, as well as imprecise dosimetry drives the variable PDT responses[[36](#_ENREF_37),[37](#_ENREF_38)]. There is strong evidence that variations in clinical response are a direct result of dosimetry that does not take into account individual differences[[39](#_ENREF_40)]. In order to bring PDT to a full realization of its potential benefits, quantitative tools are likely to play an essential role. They can provide standardization of site-specific individualized protocols by assessing light and photosensitizer doses.

Another challenge for clinical PDT of the head and neck is the difficulty in predicting the responders and non-responders[[36](#_ENREF_37)]. Quantitative optical imaging tools can play a crucial role in filling this niche. These tools are currently in primitive stages and not widely used in clinical settings for monitoring PDT mainly because optical measurements may require extra clinical time and extra fiber replacements during PDT. The techniques are limited to pre- and post-PDT measurements but with the advent of new technologies they can be adapted for monitoring during PDT, which would have three-fold benefits: (1) Reduced required clinical time, (2) no interruptions of treatment light for the optical measurements, and (3) more accurate quantification of kinetics of PDT-related parameters such as photobleaching and blood flow kinetics, which have been shown to be predictors of PDT response[[36](#_ENREF_37),[40-50](#_ENREF_41)].

**CLINICAL OPTICAL IMAGING FOR PDT MONITORING**

Tissue oxygen level is crucial for effective PDT since the PS initiates chemical reactions that result in cellular and vascular damage in targeted tissue in the presence of oxygen. Tissue oxygenation is highly affected by vascular parameters such as blood flow and blood oxygenation. During the PDT process, PS is consumed continuously. Thus, the efficacy of PDT is dependent on the vascular parameters and PS level and consumption (photobleaching)[[50](#_ENREF_51),[51](#_ENREF_52)]. Vascular parameters and PS level change during PDT and these changes may be useful early markers for therapy response[[36](#_ENREF_37),[44](#_ENREF_45),[52-54](#_ENREF_53)].

Optical imaging is a wide topic that includes many different imaging approaches. Here we will focus on a subdivision called Diffuse Optical Spectroscopies (DOS) for probing millimeter to centimeter deep tissue[[55-61](#_ENREF_56)]. In this context, DOS includes diffuse reflectance spectroscopy (DRS)[[62-67](#_ENREF_63)], diffuse fluorescence spectroscopy (DFS)[[40](#_ENREF_41),[67-70](#_ENREF_68)] and diffuse correlation spectroscopy (DCS)[[71](#_ENREF_72),[72](#_ENREF_73)]. We have recently developed a multi-modal optical imaging technique that combines DRS, DFS and DCS in a single instrument and showed the feasibility of quantification of optical parameters (absorption and scattering), drug concentration and vascular parameters such as blood flow and oxygenation in a clinical setting[[44](#_ENREF_45),[73](#_ENREF_74)].

***Multi-modal optical instrument***

The technical details of our multi-modal optical system can be found elsewhere[[44](#_ENREF_45),[73](#_ENREF_74)], but here we briefly mention the basic working principles. The instrument performs measurements sequentially in the order of blood flow (DCS), optical parameters, blood oxygenation and volume (DRS), and fluorescence (DFS). Figure 2A and Bshows the picture and schematic diagram of the instrument, respectively. The DCS instrument has a 785 nm, long coherence length laser (CrystaLaser), four single photon-counting detectors (SPCD, Perkin-Elmer), and a custom-built autocorrelator board (Correlator.com). Photodetector outputs were fed into a correlator board and intensity autocorrelation functions and photon arrival times were recorded by a computer. After blood flow measurements, the second laptop initiates fluorescence (DFS) and reflectance (DRS) data acquisition by utilizing TTL switching via a data acquisition card (DAQ, National Instruments). In absorption (DRS) mode, broadband diffuse reflectance measurements were taken by exciting the tissue with tungsten halogen lamp (Ocean Optics) and collecting the light with the Master channel of a two-channel spectrometer (Ocean Optics). In fluorescence (DFS) mode, a 410 nm laser diode (Power Technology) excites the photosensitizer in Soret band and the Slave channel of the spectrometer collects the fluorescence spectra.

A hand-held “surface” probe that holds the light source and detector fibers can be used for measuring superficial malignancies by directly placing the tip of the probe on the tissue surface (Figure 2C). Although the instrument stays the same, the hand-held surface probe is ill-suited for interstitial light delivery and noninvasive measurements and the probe-tissue interface must be changed accordingly. For an “interstitial” probe, source and detector fibers are placed inside a catheter (Figure 2D).

***Optical parameters and local light dose distribution by DRS***

Currently the standard PDT light dosimetry at the clinics is based on the prescribed incident dose, which does not take into account reflected and scattered light in the lesion. Head and neck malignancies can exhibit a multi-focal, wide-field nature of invasion and they may occur at diverse sites (*e.g.*, tongue, lip, palate, *etc.*). Therefore, they can have different optical parameters resulting in considerable inter- and intra-patient variations in the deposited local dose[[11](#_ENREF_11)]. It has been shown that the measured effective local dose can be more than 5-fold greater than the incident administrated dose, illustrating the need for *in situ* dose monitoring on an individual basis[[39](#_ENREF_40)]. Dosimetry systems using isotropic light detectors to measure both incident and scattered light are becoming more available in clinical systems[[36](#_ENREF_37),[37](#_ENREF_38)]. Multi-channel systems that can measure light dose at multiple points of interest in real time can provide on-line feedback to clinicians during treatment planning.

Tissue absorption and tissue scattering parameters modify light attenuation and thus affect the true light dose delivered to the whole three-dimensional tissue volume. Thus, direct light dose measurements may not be sufficient to quantify volumetric light distribution. Since malignancies can be highly heterogeneous, three dimensional optical parameter mapping can show heterogeneity of local light dose to the whole lesion volume***.*** Several techniques are available for mapping of optical parameters (optical absorption and scattering) *in vivo*. Most of them are based on photon diffusion equation with multi source-detector separations. Photon fluence (rate) is measured as a function of source-detector distance and measured data is fit to the diffusion model to extract optical parameters.

***Local PS dose distribution by DFS and DRS***

It has been demonstrated that PSs demonstrate significant inter- and intra-patient heterogeneity in distribution, leading to variations in the accumulated PDT dose and treatment failures[[36](#_ENREF_37),[74](#_ENREF_75),[75](#_ENREF_76)]. It has been also suggested that the variation of the treatment outcome can be reduced by adjusting the light dose based on the pretreatment PS distribution so that PDT dose is uniform in the whole disease[[36](#_ENREF_37),[75-78](#_ENREF_76)]. Although DRS can be used to quantify PS concentration by using the absorption peak of PSs, DFS is the preferred choice for this aim, since the fluorescence contrast is usually higher than the absorption contrast *in vivo*. However, fluorescence signal is affected by the tissue optical properties, and thus is not directly related to PS concentration. Ratiometric methods (with respect to optical attenuation and autofluorescence) may correct this signal distortion significantly[[79](#_ENREF_80),[80](#_ENREF_81)]. Moreover, short source-detector separation (or single source-detector) based optical probes and empirical calibration techniques that calibrate the system with respect to reference optical phantoms may allow quantification of drug concentration. For quantifying PS concentration using DFS data, background subtracted fluorescence signal is usually normalized with the reflectance data obtained by DRS[[65](#_ENREF_66),[66](#_ENREF_67),[70](#_ENREF_71),[81](#_ENREF_82)]. Fluorescence signal is assumed to be a linear combination of contributing components (*i.e.*, PS fluorescence, tissue autofluorescence, *etc*.). The normalized tissue fluorescence is fit to the modeled tissue fluorescence to extract PS concentration[[44](#_ENREF_45),[73](#_ENREF_74)].

***Tissue response monitoring by DRS and DCS***

Tissue oxygen is crucial for effective PDT[[36](#_ENREF_37),[82-84](#_ENREF_83)]. Tissue oxygen, in turn, is affected by vascular parameters such as blood oxygenation, blood volume and blood flow[[50](#_ENREF_51),[52](#_ENREF_53)]. Most photosensitizers have significant vascular disrupting effects, and can create substantial vascular changes. All these parameters are inter-dependent to each other and can change continuously during PDT[[4](#_ENREF_4),[36](#_ENREF_37)]. Blood flow changes during PDT correlated strongly with tumor growth delay, and blood oxygenation and volume changes were correlated with PDT outcome[[50](#_ENREF_51),[52](#_ENREF_53),[85](#_ENREF_86)]. Moreover, photosensitizer photobleaching has been shown to be a surrogate marker of PDT response[[40](#_ENREF_41),[86-90](#_ENREF_87)]. Therefore, continuous monitoring of these parameters could be useful for providing real-time treatment feedback, and may serve as quantitative *in vivo* markers for assessing treatment response[[4](#_ENREF_4),[36](#_ENREF_37),[63](#_ENREF_92)].

For quantifying vascular parameters such as blood oxygenation and blood volume, an analytic diffuse reflectance modeling can be utilized to fit the diffusion model to experimental diffuse reflectance data obtained by DRS. We assume tissue absorption is composed of linear contribution from oxy-hemoglobin and deoxy-hemoglobin in blood, and PS absorption. Blood volume is related to total hemoglobin concentration and is defined as the sum of oxy-hemoglobin and dexoy-hemoglobin concentrations, and blood oxygen saturation is defined as the ratio of oxy-hemoglobin concentration to total hemoglobin concentration. Tissue scattering is usually modeled as Mie type behavior that is related to scatterer size and concentration[[91](#_ENREF_93)]. A multi-wavelength fitting algorithm is usually used to directly extract the hemoglobin concentrations or blood oxygen saturation and blood volume[[63](#_ENREF_64),[92](#_ENREF_94),[93](#_ENREF_95)]. Blood oxygen saturation is related to tissue oxygen and hypoxia[[52](#_ENREF_53),[94](#_ENREF_96)] and blood volume is related to microvessel density[[95](#_ENREF_97)].

Tissueblood flow is measured using a previously described and validated DCS instrument, which measures rapid light intensity temporal fluctuations in tissue and then uses the autocorrelation functions associated with these fluctuations to extract information about the speed of moving tissue scatterers, in this case red blood cells[[44](#_ENREF_45),[49](#_ENREF_50),[96-101](#_ENREF_98)]. Decay rate of the autocorrelation function is related to blood flow[[99-101](#_ENREF_101)]. DCS is advantageous compared to conventional imaging modalities in that it measures directly blood cell movements and does not need any contrast agent administration and pharmacokinetic models to quantify blood flow.

***A surrogate molecular marker for PDT efficacy***

It is often desired to correlate noninvasive parameters with other techniques such as molecular biomarkers of a treatment response. We have shown previously in preclinical models and clinical biopsy samples that the cross-linking of the signal transducer and activator of transcription 3 (STAT3) correlates with the accumulated PDT dose and can be a quantitative biomarker of cellular killing[[102](#_ENREF_104),[103](#_ENREF_105)]. The crosslinking is identified by immunoblot analysis for STAT3 protein in the extracts from tumor tissue sections calculated as homodimeric complex I relative to total STAT3 signal[[102](#_ENREF_104),[103](#_ENREF_105)]. We compared our measured indices with the STAT3 crosslinking as showcased below.

***A clinical case report***

In our previous work we demonstrated the assessment of PDT response-related multi-parameters of blood flow, oxygenation, blood volume, PS concentration in the same clinical setting of Photoclor (HPPH)-mediated PDT in head and neck lesions in the oral cavity[[44](#_ENREF_45)]. We reported an interesting case where two patients had lesions treated with the same administered photosensitizer dose (HPPH, 4.0 mg/m2) and a similar delivered light dose (approximately 125 J/cm2), but the accumulated local doses were more than 100-fold different as determined by the STAT3 crosslinking (Table 1). The first patient (P-1) had a large carcinoma *in situ* (CIS) of the hard palate on the roof of the mouth and PDT induced photoreaction with 35% STAT3 crosslinking, and the second patient (P-2) had high-grade dysplasia in a papilloma of the buccal mucosa with only 0.3% STAT3 crosslinking (Figure 3). We quantified local PDT-related parameters with diffuse optical methods to investigate whether this substantial difference could be detected noninvasively since these parameters can affect accumulated local dose.

As Table 1 summarizes, PDT-induced changes in the quantified optical parameters were significantly different between these lesions. Changes in photosensitizer concentration (ΔcHPPH), blood flow index (ΔBFI)and blood volume fraction (ΔBVf) were significantly higher in Patient-1 (P1) than in Patient-2 (P2), but the changes in blood oxygen saturation (StO2) were similar for both patients, though the trend was different: P1 had an increase and P2 showed a decrease trend.

We further investigated whether this difference could be observed before therapy by quantifying pre-PDT contrasts (mean ± SE) by noninvasive methods. All parameters except blood volume fraction were significantly different between the lesions (Table 2). The lesion of Patient-1 (P1) had more favorable properties related to accumulated local PDT dose, since its photosensitizer content as well as blood flow, blood volume and blood oxygen saturation were higher than Patient-2 (P2).

Our results indicated that parameters quantified with diffuse optical spectroscopies at pre-PDT as well as PDT-induced changes may be indicative of local PDT reaction and may be *in vivo* predictors of PDT outcome. Since each parameter showed different contrast and therapy-induced changes, one parameter alone may not be a strong indicator of PDT response and multi-parameters assessed by optical methods may provide accurate measure of PDT response[[44](#_ENREF_45)].

**CONCLUSION**

In summary, PDT is regarded as an emerging treatment option for the head and neck malignancies. PDT can be applied repetitively if the previous treatment fails. With the advent of newly developed photosensitizers, specificity and penetration depth can be improved. The simplicity of the PDT treatment and reduced cost of technology such as light sources and light delivery devices can help wide usage at the clinical settings. Moreover, standardization of clinical protocols by using the same light and drug types and doses is a desired need. Novel optical methods can provide PDT-dose related parameters such as optical parameters and photosensitizer concentration in the whole lesion, as well as can quantify blood flow, oxygenation and photosensitizer photobleaching for assessing the PDT response and providing feedback to clinicians for optimization and standardization of PDT in clinics.

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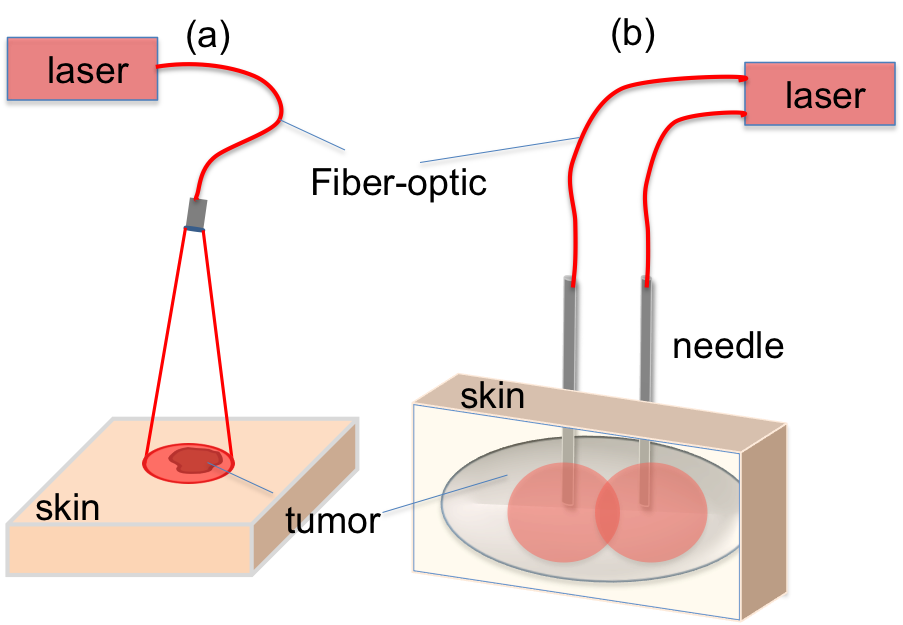
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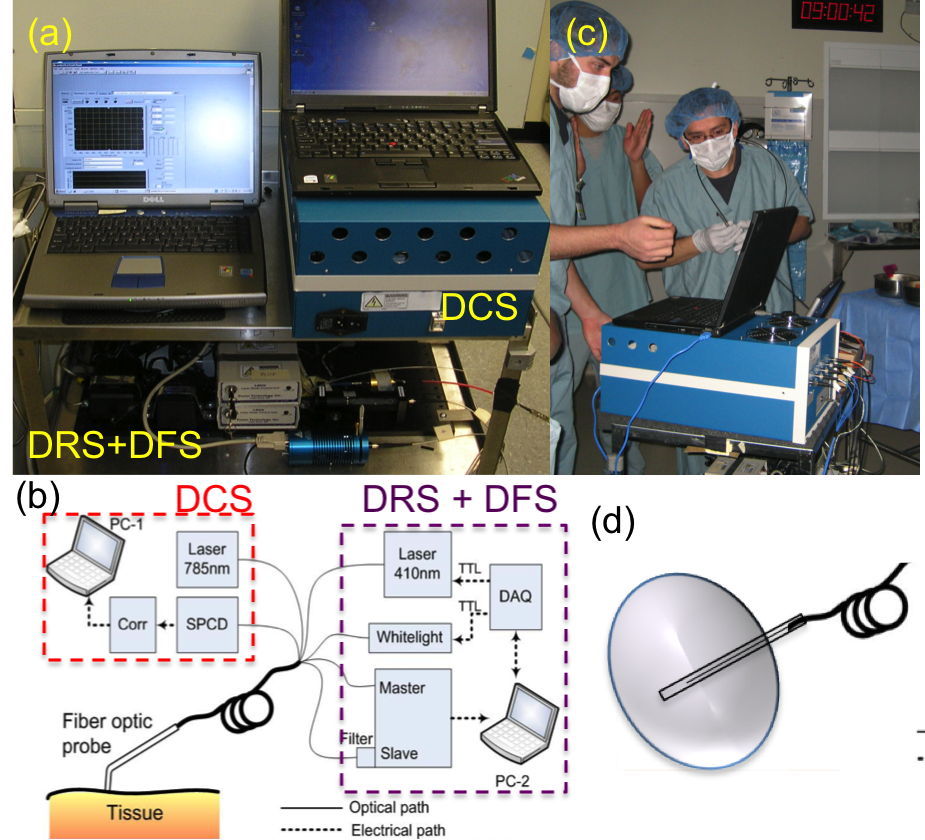
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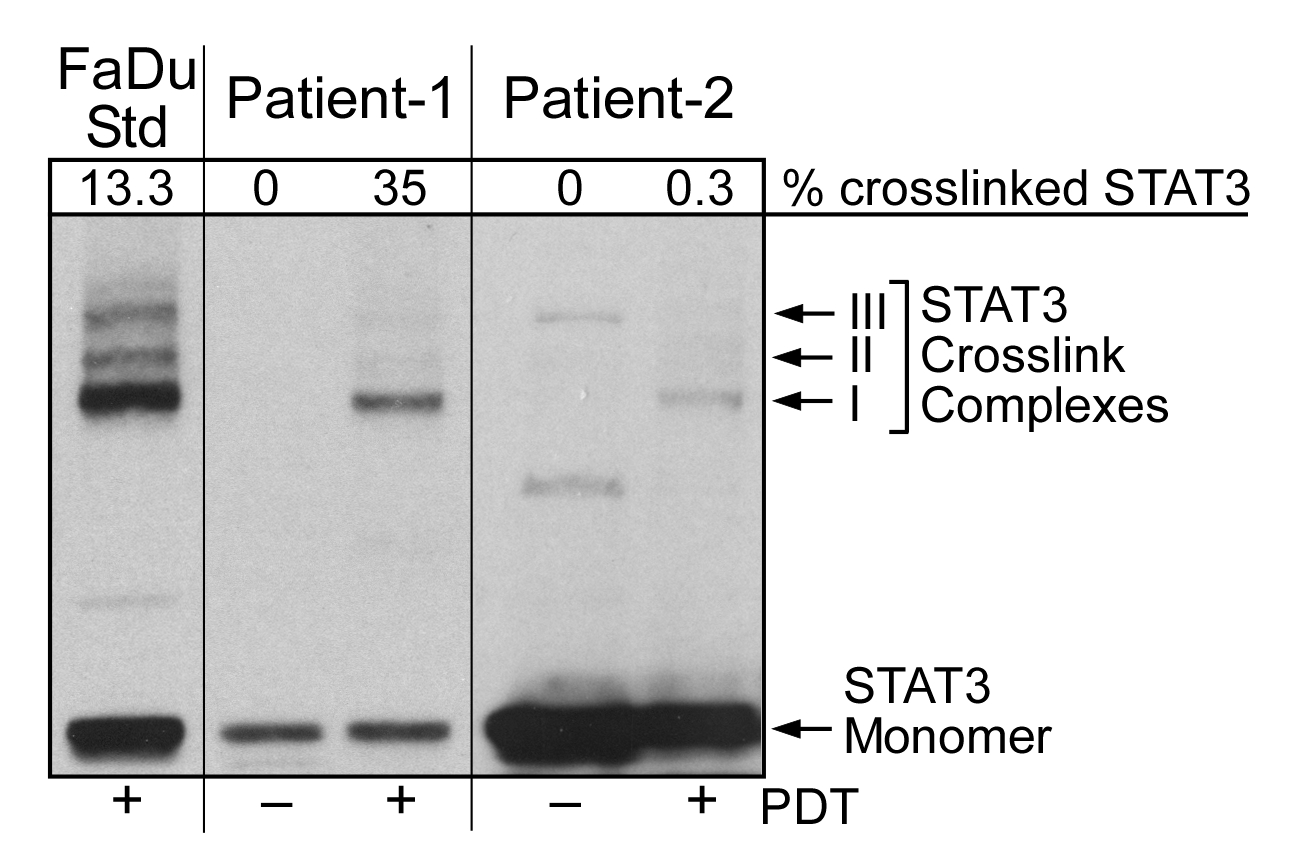
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**Figure 1 Representation for light delivery during surface and interstitial photodynamic therapy.** A: Surface illumination photodynamic therapy (PDT) for treating superficial malignancies. Laser light is directed to tissue surface *via* micro-lens fiber. Tumor is located superficially; B: Interstitial PDT treatment for deeper and thicker malignancies. Individual fibers are placed inside 19-gauge needles and inserted into tissue. Number of fibers is selected according to treated volume.



**Figure 2 Clinical multi-modal optical instrument for photodynamic therapy dosimetry and response monitoring.** A: Picture of multi-modal clinical optical instrument; B: Diagram of multi-modal clinical optical instrument; C: During the measurements at the operating room; D: Interstitial optical probe for measurements in deep and thick tumors. Adapted from the reference[[45](#_ENREF_45)] with the permission.



**Figure 3 Signal transducer and activator of transcription 3 crosslinking as a molecular marker for local photodynamic therapy dose.** Signal transducer and activator of transcription 3 crosslinking for Patient-1 and Patient-2 with a human hypopharyngeal carcinoma cell line (FaDu) shown as a control. Adapted from the reference[[74](#_ENREF_74)] with the permission.

**Table 1** **Photodynamic therapy-induced changes in photodynamic therapy-related parameters for two patients**

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
|  | **Lesion type** | **STAT3**  **(%)** | **ΔBFI**  **(%)** | **ΔBVf**  **(%)** | **ΔStO2**  **(%)** | **ΔcHPPH**  **(%)** |
| **P1** | CIS | 35 | 83.4 | 23.0 | +15.2 | 51.8 |
| **P2** | Dysplasia | 0.3 | 59.2 | 7.5 | −17.0 | 38.6 |

BFI was significant for both patients while changes in cHPPH were only significant for patient-1. Adapted from the reference[[74](#_ENREF_74)] with the permission. STAT3: Signal transducer and activator of transcription 3; cHPPH: Changes in photosensitizer concentration; BFI: Blood flow index; BVf: Blood volume fraction; StO2: Saturation; P1:Patient-1; P2: Patient-2; CIS: carcinoma *in situ*.

**Table 2** **Pretreatment contrasts in photodynamic therapy-related parameters between two patients**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
|  | **Lesion type** | **BFI**  **(a.u.)** | **BVf**  **(%)** | **StO2**  **(%)** | **cHPPH**  **(μmol/L)** |
| **P1** | CIS | 6.7 ± 2.8 | 2.5 ± 0.7 | 74 ± 2 | 0.34 ± 0.02 |
| **P2** | Dysplasia | 1.8 ± 0.5 | 1.3 ± 0.2 | 64 ± 3 | 0.10 ± 0.03 |

All parameters except blood volume fraction showed a significant difference between two patients. Adapted from the reference[[74](#_ENREF_74)] with the permission. cHPPH: Changes in photosensitizer concentration; BFI: Blood flow index; BVf: Blood volume fraction; StO2: Saturation; P1:Patient-1; P2: Patient-2; CIS: carcinoma *in situ*.