

World Journal of *Nephrology*

World J Nephrol 2017 May 6; 6(3): 86-167



REVIEW

- 86 Contrast-induced acute kidney injury: A review of practical points
Ozkok S, Ozkok A
- 100 Role of different imaging modalities of vascular calcification in predicting outcomes in chronic kidney disease
Disthabanchong S, Boongird S
- 111 Targeting cannabinoid signaling for peritoneal dialysis-induced oxidative stress and fibrosis
Yang CY, Chau YP, Chen A, Lee OKS, Tarng DC, Yang AH

MINIREVIEWS

- 119 Cold dialysis and its impact on renal patients' health: An evidence-based mini review
Sakkas GK, Krase AA, Giannaki CD, Karatzaferi C
- 123 Role of imaging in the evaluation of renal dysfunction in heart failure patients
Grande D, Terlizzese P, Iacoviello M

ORIGINAL ARTICLE

Case Control Study

- 132 Any link of gout disease control among hypertensive patients and onset of end-stage renal disease? Results from a population-based study
Perreault S, Nuevo J, Baumgartner S, Morlock R

Retrospective Study

- 143 Advanced wasting in peritoneal dialysis patients
Xu Z, Murata GH, Glew RH, Sun Y, Vigil D, Servilla KS, Tzamaloukas AH
- 150 Clinicopathological spectrum of snake bite-induced acute kidney injury from India
Vikrant S, Jaryal A, Parashar A
- 162 Acute kidney injury from different poisonous substances
Naqvi R

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World Journal of Nephrology (*World J Nephrol*, *WJN*, online ISSN 2220-6124, DOI: 10.5527) is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

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World Journal of Nephrology is now indexed in PubMed, PubMed Central.

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NAME OF JOURNAL
World Journal of Nephrology

ISSN
 ISSN 2220-6124 (online)

LAUNCH DATE
 February 6, 2012

FREQUENCY
 Bimonthly

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PUBLICATION DATE
 May 6, 2017

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Targeting cannabinoid signaling for peritoneal dialysis-induced oxidative stress and fibrosis

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Author contributions: All authors contributed to this paper.

Supported by The Ministry Of Science and Technology, Taiwan, Nos. NSC 96-2628-B-075-003-MY3, MOST 104-2314-B-075-031, and MOST 105-2628-B-075-008-MY3; a grant from Taipei Veterans General Hospital, Taipei, Taiwan, No. V106D25-003-MY3-1; Taipei Veterans General Hospital, National Yang-Ming University Excellent Physician Scientists Cultivation Program, No. 103-V-B-024.

Conflict-of-interest statement: None.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external

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Manuscript source: Unsolicited manuscript

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Received: November 4, 2016

Peer-review started: November 9, 2016

First decision: November 30, 2016

Revised: January 28, 2017

Accepted: February 18, 2017

Article in press: February 20, 2017

Published online: May 6, 2017

Abstract

Long-term exposure to bioincompatible peritoneal dialysis (PD) solutions frequently results in peritoneal fibrosis and ultrafiltration failure, which limits the life-long use of and leads to the cessation of PD therapy. Therefore, it is important to elucidate the pathogenesis of peritoneal fibrosis in order to design therapeutic strategies to prevent its occurrence. Peritoneal fibrosis is associated with a chronic inflammatory status as well as an elevated oxidative stress (OS) status. Beyond uremia *per se*, OS also results from chronic exposure to high glucose load, glucose degradation products, advanced glycation end products, and hypertonic stress. Therapy targeting the cannabinoid (CB) signaling pathway has been reported in several chronic inflammatory diseases with elevated OS. We recently reported that the intra-peritoneal administration of CB receptor ligands, including CB₁ receptor antagonists

and CB₂ receptor agonists, ameliorated dialysis-related peritoneal fibrosis. As targeting the CB signaling pathway has been reported to be beneficial in attenuating the processes of several chronic inflammatory diseases, we reviewed the interaction among the cannabinoid system, inflammation, and OS, through which clinicians ultimately aim to prolong the peritoneal survival of PD patients.

Key words: Reactive oxygen species; Peritoneal fibrosis; Peritoneal dialysis; Cannabinoid signaling; Oxidative stress

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Core tip: Long-term exposure to bioincompatible peritoneal dialysis (PD) solutions frequently results in peritoneal fibrosis and ultrafiltration failure, which limits the life-long use of PD therapy. Beyond uremia *per se*, oxidative stress (OS) also results from chronic exposure to high glucose load, glucose degradation products, advanced glycation end products, and hypertonic stress in PD patients. Therapy targeting the cannabinoid signaling pathway has been reported in several chronic inflammatory diseases with elevated OS. In this article, we review the interaction among the cannabinoid system, inflammation, and OS, through which the health-care professionals ultimately aim to prolong the peritoneal survival of PD patients.

Yang CY, Chau YP, Chen A, Lee OKS, Tarn DC, Yang AH. Targeting cannabinoid signaling for peritoneal dialysis-induced oxidative stress and fibrosis. *World J Nephrol* 2017; 6(3): 111-118 Available from: URL: <http://www.wjgnet.com/2220-6124/full/v6/i3/111.htm> DOI: <http://dx.doi.org/10.5527/wjn.v6.i3.111>

REVIEW ARTICLE

As chronic kidney disease progresses to end-stage renal disease, uremia ensues requiring the use of long-term dialysis therapy. Both uremia and dialysis give rise to elevated oxidative stress (OS)^[1-4], which is detrimental to the patients' health. A recent survey indicated that approximately 11% of dialysis patients undergo peritoneal dialysis (PD) therapy worldwide, estimating to be more than 272000 patients with an 8% annual growth rate globally^[5]. Beyond uremia *per se*, PD patients are at an increased risk of inflammation and OS, both systemically and locally, because of the chronic exposure to high glucose load^[6-15], advanced glycosylated end products (AGEs)^[16-18], glucose degradation products (GDPs)^[18-21], and hypertonic stress^[9,22-24]. After long-term exposure to various GDPs and AGEs, mesothelial cells of PD patients undergo a de-differentiation process, followed by peritoneal fibrosis and ultrafiltration failure^[25-29]. In a large peritoneal biopsy study, peritoneal tissue samples from 212 subjects including healthy controls, hemodialysis and PD patients were examined. They found that peritoneal fibrosis was absent in normal individuals but was present

in 28% of samples from hemodialysis patients and up to 56% of biopsies from PD patients^[30]. Although whether a uremia-induced chronic inflammation status causes OS or OS leads to a proinflammatory process in uremic patients remains uncertain^[1,4,31], therapeutic strategies for peritoneal damage targeting OS have been reported, such as antioxidants^[12,18,32,33], scavenging agents for reactive oxygen species (ROS)^[34-36], the trace element selenium^[37], and gaseous mediators^[38,39].

Cannabinoid (CB) signaling has also been reported to be effective in treating a variety of disease entities with elevated OS, including diabetic macrovascular and microvascular complications^[40-50], cardiomyopathies^[51-57], liver injury and fibrosis^[58-64], cholangiopathies^[65], colitis^[66,67], drug nephrotoxicity^[68,69], and autoimmune diseases^[70] (Table 1). Since the peritoneum of long-term PD patients is under a chronic pro-inflammatory status, it is possible that the CB receptor (CBR) signaling system may be inappropriately modulated. In this article, we reviewed the sources and influence of OS in PD patients, and the therapeutic rationale and mechanisms of targeting the CB signaling system to reduce OS.

HIGH GLUCOSE AND GDP INCREASE OS

Uremia *per se* and dialysis therapy both lead to a pro-oxidant status, which can lead to increased OS in patients receiving hemodialysis or PD therapy^[1-4]. In particular, PD patients are exposed to hypertonic glucose solution on a long-term basis, which is not only toxic to mesothelial cells^[6,71] but also promotes immune cell apoptosis^[72]. Moreover, the high-temperature sterilization process produces GDPs such as methylglyoxal (MGO), acetaldehyde, formaldehyde, and 3-deoxyglucosone in PD dialysate^[73-76]. GDPs possess strong oxidative properties and toxicity, and can induce AGEs^[77]. In addition, it has been demonstrated that MGO, a key GDP, in PD dialysates inhibits the insulin signaling pathway, resulting in increased endogenous ROS production and subsequent cell injury^[78]. Furthermore, 2-33 μmol/L of MGO has been reported to be present in commercial glucose-based PD fluids^[79,80].

After long-term exposure to various GDPs and AGEs, mesothelial cells undergo a de-differentiation process and peritoneal fibrosis ensues^[25-29]. Furthermore, these sites of chronic inflammatory have been associated with progressive peritoneal angiogenesis^[29,81-83], and finally a reduction in the efficacy of PD. However, therapeutic strategies for these pathogenic processes have not been fully developed^[81], and so some PD patients still develop peritoneal fibrosis or even encapsulating peritoneal sclerosis, a disastrous and highly fatal condition.

Low GDP PD dialysates can prevent peritoneal injury by PD-induced OS. However, the relatively high cost limits their full implementation. Moreover, even though the concentration of GDPs in the new generation of PD dialysates is low, it still exists^[84-86]. Meanwhile, as long as the PD dialysate is glucose-based, glucose load *per se* results in ROS production^[6-15]. Fortunately,

Table 1 Summary of cannabinoid signaling on modulation of diseases with increased oxidative stress

Disease	Species	Disease model	Ref.
Diabetes mellitus			
Microvascular complications			
Nephropathy	Mouse	<i>In vivo</i>	[49,50]
Retinopathy	Mouse, rat	<i>In vivo</i>	[43,44]
Neuropathy	Mouse, rat	<i>In vivo</i>	[45-47]
Macrovascular complications	Mouse, rat	<i>In vivo</i>	[40,41]
Cardiomyopathy			
Diabetic cardiomyopathy	Mouse	<i>In vivo</i>	[57]
	Human cardiomyocyte	<i>In vitro</i>	[57]
Myocardial ischemia/reperfusion	Mouse	<i>In vivo</i>	[51,53,54]
Ageing-related cardiomyopathy	Mouse	<i>In vivo</i>	[55]
Doxorubicin-induced cardiomyopathy	Mouse	<i>In vivo</i>	[52,56]
	Human cardiomyocyte	<i>In vitro</i>	[56]
	Rat cardiomyocyte	<i>In vitro</i>	[52]
	Mouse	<i>In vivo</i>	[68,69]
Cisplatin nephropathy			
Liver injury/fibrosis			
Ischemia/reperfusion hepatocyte injury	Mouse	<i>In vivo</i>	[59]
CCl ₄ -induced hepatocyte injury	Mouse	<i>In vivo</i>	[59]
Alcoholic liver fibrosis	Mouse	<i>In vivo</i>	[62]
Fibrotic activation of hepatic stellate cells	Human, rat, mouse	<i>In vitro</i>	[58,60,63,64]
Biliary diseases			
Cholangiopathies	Mouse	<i>In vivo</i>	[65]
	Mouse cholangiocyte	<i>In vitro</i>	[65]
Enteric diseases			
Inflammatory bowel disease	Mouse	<i>In vivo</i>	[66]
Colitis	Mouse	<i>In vivo</i>	[67]
Autoimmune systemic sclerosis	Mouse	<i>In vivo</i>	[70]

therapies reducing peritoneal OS are under investigation, and include antioxidants^[12,18,32,33], ROS scavengers^[34-36], selenium^[37], and gaseous mediators^[38,39] (Figure 1).

HYPERTONIC DIALYSATE-INDUCED OSMOTIC STRESS AND OXIDATIVE INJURY

In addition to low GDP PD dialysates, non-glucose-based PD dialysates such as icodextrin are free of GDPs and have been shown to be beneficial in fluid control and small solute clearance^[87]. It has also been reported that peritoneal OS is reduced when using icodextrin compared with conventional PD dialysates^[88].

However, other studies have reported conflicting results in that the osmotic stress, a type of stress resulted from hypertonic PD dialysate exposure, leads to oxidative DNA damage of peritoneal mesothelial cells through lipid peroxidation. Such peritoneal oxidative injury may then lead to mesothelial cell death either through apoptosis or necrosis^[9,22-24]. Therefore, persistent efforts are warranted to develop an optimal solution.

THE CANNABINOID SIGNALING PATHWAY AND ITS MOLECULAR MECHANISMS ON INFLAMMATION AND FIBROSIS

Our recent study suggested that using CBR ligands as an

additive in PD dialysate may be a promising solution to treat dialysis-induced peritoneal inflammation^[89]. There are two subtypes of CBRs, type 1 CB receptor (CB₁R) and type 2 CB receptor (CB₂R). The former mainly exists in the brain and regulates inhibitory neurotransmitters on neurons through the psychoactive drug cannabis or endocannabinoids such as anandamide. Nevertheless, it has recently been found that CB₁R also exists in tissues other than that of the central nervous system, and that its function varies in different organs^[73]. CB₁R antagonists and CB₂R agonists have been shown to decrease inflammation and OS^[48], and previous studies have also shown that CBR plays an important role in liver fibrogenesis^[90-94]. Moreover, hepatic fibrosis can be rescued by knockout of the CB₁R gene or by administration of the CB₁R antagonist^[93,95,96]. In contrast, CB₂R is located on immune cells and modulates cytokine release^[97,98]. Recent studies have shown that the activation of CB₂R ameliorates liver fibrogenesis through inhibiting myofibroblast cell proliferation^[92,99]. Furthermore, CBR ligands such as cannabidiol have been proven to be well-tolerated without adverse effects when administered to humans on a long-term basis^[48].

Only a few studies have been published on pharmacological modulation targeting peritoneal inflammation and fibrogenesis using CBR ligands^[100]. Our recent study indicated that the pharmacological effects of CBR ligands against dialysate-induced peritoneal fibrosis may involve a diverse signaling system including the TGF- β 1-PI3K pathway^[89], and that this offers a promising therapeutic strategy for the prevention of peritoneal fibrosis in patients receiving long-term PD. Therefore, we suggest that CBR signaling might play an important role in the patho-

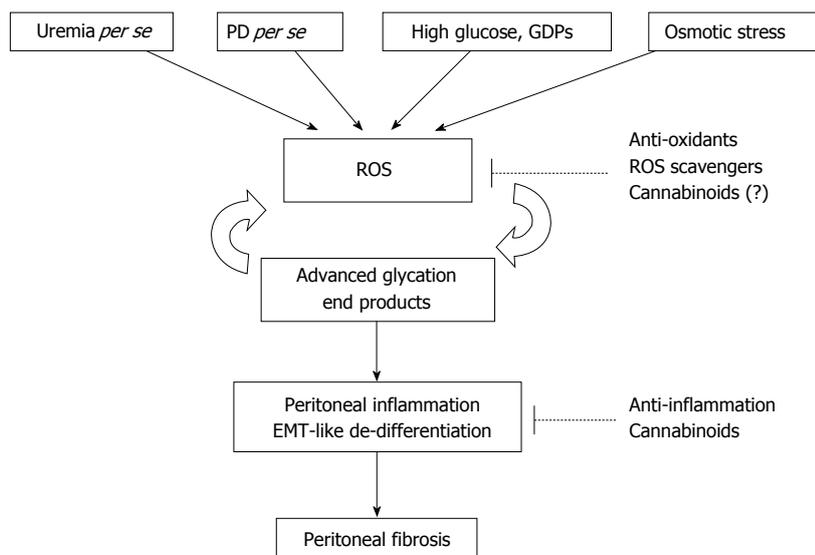


Figure 1 Peritoneal dialysis-induced oxidative stress and peritoneal fibrosis. PD: Peritoneal dialysis; GDPs: Glucose degradation products; EMT: Epithelial-mesenchymal transition; ROS: Reactive oxygen species.

genesis of dialysis-induced peritoneal inflammation and ROS production, which required further studies.

CELLULAR SOURCES OF REACTIVE OXYGEN SPECIES IN PD-INDUCED FIBROSIS

In vitro data showed that, upon GDP and AGE exposure during PD, cellular OS of human peritoneal mesothelial cells were induced through activation of protein kinase C, nicotinamide adenine dinucleotide phosphate oxidase, and mitochondrial metabolism. In turn, the generated ROS upregulate fibronectin expression by mesothelial cells^[7]. An *in vivo* study demonstrated 8-hydroxy-2'-deoxyguanosine (8-OHdG)-positive cells, indicating cells with increased OS, were observed throughout the fibrotic peritoneal tissue. Further immunofluorescent analysis revealed that 8-OHdG-positive cells also co-stained with mesothelin (mesothelial cells), CD68 (macrophages), CD31 (vascular endothelial cells), and α -smooth muscle actin (fibroblasts)^[34], suggesting that OS was also increased in cells other than mesothelial cells. However, whether these fibroblasts with increased cellular OS were derived from an epithelial-mesenchymal transition (EMT)-like process of mesothelial cells is unknown.

As aforementioned, it has been reported that CB₂R is located on immune cells and modulates cytokine release^[97,98]. The CB₂R expression of human lymphocytes was downregulated by TGF- β stimulation^[101], which was not seen in human mesothelial cells^[89]. These findings suggest that TGF- β 1 might have different physiological function in different cell types. Meanwhile, CB₂R activation might exert its anti-fibrotic effects not directly to cells undergoing fibrotic change but indirectly through modulating the immune cells. Such interaction among different types of cells underlines the pathophysiological role of CBR signaling pathway in uremic and/or dialysis

injuries, which is partly supported by a recent study showing that systemic administration of interleukin-10, an anti-inflammatory cytokines secreted by M2 macrophages, significantly reduced fibrous peritoneal thickening^[102]. Therefore, current evidence indicates that beyond mesothelial cells, macrophages and vascular endothelial cells also contribute to ROS production during PD-induced peritoneal fibrosis.

THERAPEUTIC POTENTIAL OF CANNABINOID SIGNALING ON PD-INDUCED OS AND INFLAMMATION

It has been reported that modulating the CB signaling system is beneficial in treating various diseases resulting from increased OS including diabetic macrovascular and microvascular complications^[41-43,48,57], cardiomyopathies^[51,55-57], liver injury and fibrosis^[58,60-64,103], cholangiopathies^[65], colitis^[66,67], drug nephrotoxicity^[68,69], and autoimmune diseases^[70]. At present, evidences of the therapeutic benefit of CBR ligands on peritoneal OS are lacking and deserve investigations.

Furthermore, recent studies have also shown significant anti-fibrogenic effects of CBR ligands in the liver^[91-93,95,96,99]. However, the effects of CBR ligands on peritoneal tissue have rarely been studied, with only one recent study reporting that the CB₂R agonist reduced the number of peritoneal macrophages in a murine peritonitis model induced by thioglycollate, an AGE derivative^[104]. Furthermore, we recently demonstrated that both the selective CB₁R antagonist (AM281) and the selective CB₂R agonist (AM1241) were able to ameliorate MGO-induced peritoneal fibrosis *in vivo*, indicating that pharmacological modulation of CBR may be a feasible approach to optimize the biocompatibility of peritoneal dialysis fluid. However, ACEA, a CB₁R agonist, has been shown to have an opposite effect to AM281 with regards type I collagen

expression in cultured mesothelial cells, indicating specific anti-fibrogenic activity of the CB₁R antagonist^[89].

During peritoneal fibrosis, mesothelial cells undergo a process of myofibroblastic conversion. This is a complex process which has been reported to be associated with increased levels of TGF- β 1, leptin, metalloproteinase-2, vascular endothelial growth factor, Snail, and the receptor for advanced glycosylated end products^[79,105,106]. TGF- β 1 has long been known to play crucial roles in the fibrogenic process of the peritoneum^[107,108]. A previous study demonstrated that a high glucose load stimulates the production of TGF- β 1 in peritoneal mesothelial cells^[109]. Moreover, AGEs have been shown to increase the expression of TGF- β 1, contribute to the development of sub-mesothelial fibrosis^[110], and significantly contribute to increases in peritoneal OS. Meanwhile, our recent study showed that such EMT-like processes can be attenuated by the selective CB₁R antagonist, AM281^[89]. It is quite possible that OS is involved in the CBR-related pharmacological effects against peritoneal fibrosis. However, the exact pathogenic mechanisms between the CBR signaling pathway and uremic and/or dialysis injuries remain largely unknown.

CONCLUSION

Compared with hemodialysis patients, the chronic use of PD dialysate exposes PD patients to additional OS. The influence of such OS on the patients' health can be both systemic and local, leading to cardiovascular diseases and peritoneal fibrosis, respectively. It has been shown that OS plays a critical role in the pathogenesis of chronic inflammatory diseases, and therefore targeting the CB signaling system may offer a potential therapeutic strategy to reduce dialysis-induced peritoneal fibrosis and eventually to prolong the peritoneal survival of PD patients.

REFERENCES

- 1 **Marques de Mattos A**, Marino LV, Ovidio PP, Jordão AA, Almeida CC, Chiarello PG. Protein oxidative stress and dyslipidemia in dialysis patients. *Ther Apher Dial* 2012; **16**: 68-74 [PMID: 22248198 DOI: 10.1111/j.1744-9987.2011.01009.x]
- 2 **Diepeveen SH**, Verhoeven GH, van der Palen J, Dikkeschei BL, van Tits BL, Kolsters G, Offerman JJ, Bilo HJ, Stalenhoef AF. The effect of the initiation of renal replacement therapy on lipid profile and oxidative stress during the first 6 months of treatment. *Clin Chim Acta* 2005; **361**: 112-118 [PMID: 16122722 DOI: 10.1016/j.cccn.2005.05.007]
- 3 **Witko-Sarsat V**, Friedlander M, Capeillère-Blandin C, Nguyen-Khoa T, Nguyen AT, Zingraff J, Jungers P, Descamps-Latscha B. Advanced oxidation protein products as a novel marker of oxidative stress in uremia. *Kidney Int* 1996; **49**: 1304-1313 [PMID: 8731095 DOI: 10.1038/ki.1996.186]
- 4 **Himmelfarb J**, Stenvinkel P, Ikizler TA, Hakim RM. The elephant in uremia: oxidant stress as a unifying concept of cardiovascular disease in uremia. *Kidney Int* 2002; **62**: 1524-1538 [PMID: 12371953 DOI: 10.1046/j.1523-1755.2002.00600.x]
- 5 **Li PK**, Chow KM, Van de Luitgaarden MW, Johnson DW, Jager KJ, Mehrotra R, Naicker S, Pecoits-Filho R, Yu XQ, Lameire N. Changes in the worldwide epidemiology of peritoneal dialysis. *Nat Rev Nephrol* 2017; **13**: 90-103 [PMID: 28029154 DOI: 10.1038/nrneph.2016.181]
- 6 **Gotloib L**, Shostak A, Wajsbrodt V, Kushnier R. High glucose induces a hypertrophic, senescent mesothelial cell phenotype after long in vivo exposure. *Nephron* 1999; **82**: 164-173 [PMID: 10364709 DOI: 10.1159/000045393]
- 7 **Lee HB**, Yu MR, Song JS, Ha H. Reactive oxygen species amplify protein kinase C signaling in high glucose-induced fibronectin expression by human peritoneal mesothelial cells. *Kidney Int* 2004; **65**: 1170-1179 [PMID: 15086456 DOI: 10.1111/j.1523-1755.2004.00491.x]
- 8 **Hung KY**, Liu SY, Yang TC, Liao TL, Kao SH. High-dialysate-glucose-induced oxidative stress and mitochondrial-mediated apoptosis in human peritoneal mesothelial cells. *Oxid Med Cell Longev* 2014; **2014**: 642793 [PMID: 24891925]
- 9 **Gotloib L**. Mechanisms of cell death during peritoneal dialysis. A role for osmotic and oxidative stress. *Contrib Nephrol* 2009; **163**: 35-44 [PMID: 19494593 DOI: 10.1159/000223778]
- 10 **Doñate T**, Herreros A, Martinez E, Martinez J, Andrés E, Cabezas A, Ortiz A, de Prado A, Pou JM, Pamplona R, Portero Otin M, Bellmunt MJ. Protein oxidative stress in dialysis patients. *Adv Perit Dial* 2002; **18**: 15-17 [PMID: 12402579]
- 11 **Zhu X**, Wen F, Yang D, Liu J, Yuan S, Li J, Liu H, Xu X, Sun L, Liu F. [Effect of high glucose peritoneal dialysis solution on PGC-1 α expression and mitochondrial related oxidative injury in human peritoneal mesothelial cells]. *ZhongNan DaXue XueBao YiXue Ban* 2013; **38**: 1085-1091 [PMID: 24316930]
- 12 **Gotloib L**, Wajsbrodt V, Cuperman Y, Shostak A. Acute oxidative stress induces peritoneal hyperpermeability, mesothelial loss, and fibrosis. *J Lab Clin Med* 2004; **143**: 31-40 [PMID: 14749683 DOI: 10.1016/j.lab.2003.09.005]
- 13 **Duan S**, Yu J, Liu Q, Wang Y, Pan P, Xiao L, Ling G, Liu F. Epithelial-to-mesenchymal transdifferentiation of peritoneal mesothelial cells mediated by oxidative stress in peritoneal fibrosis rats. *ZhongNan DaXue XueBao YiXue Ban* 2011; **36**: 34-43 [PMID: 21311137]
- 14 **Nishikawa T**, Edelstein D, Du XL, Yamagishi S, Matsumura T, Kaneda Y, Yorek MA, Beebe D, Oates PJ, Hammes HP, Giardino I, Brownlee M. Normalizing mitochondrial superoxide production blocks three pathways of hyperglycaemic damage. *Nature* 2000; **404**: 787-790 [PMID: 10783895 DOI: 10.1038/35008121]
- 15 **Ishibashi Y**, Sugimoto T, Ichikawa Y, Akatsuka A, Miyata T, Nangaku M, Tagawa H, Kurokawa K. Glucose dialysate induces mitochondrial DNA damage in peritoneal mesothelial cells. *Perit Dial Int* 2002; **22**: 11-21 [PMID: 11929138]
- 16 **Yim MB**, Yim HS, Lee C, Kang SO, Chock PB. Protein glycation: creation of catalytic sites for free radical generation. *Ann N Y Acad Sci* 2001; **928**: 48-53 [PMID: 11795527 DOI: 10.1111/j.1749-6632.2001.tb05634.x]
- 17 **Wautier MP**, Chappey O, Corda S, Stern DM, Schmidt AM, Wautier JL. Activation of NADPH oxidase by AGE links oxidant stress to altered gene expression via RAGE. *Am J Physiol Endocrinol Metab* 2001; **280**: E685-E694 [PMID: 11287350]
- 18 **Noh H**, Kim JS, Han KH, Lee GT, Song JS, Chung SH, Jeon JS, Ha H, Lee HB. Oxidative stress during peritoneal dialysis: implications in functional and structural changes in the membrane. *Kidney Int* 2006; **69**: 2022-2028 [PMID: 16641917 DOI: 10.1038/sj.ki.5001506]
- 19 **Müller-Krebs S**, Kihm LP, Zeier B, Gross ML, Wieslander A, Haug U, Zeier M, Schwenger V. Glucose degradation products result in cardiovascular toxicity in a rat model of renal failure. *Perit Dial Int* 2010; **30**: 35-40 [PMID: 20056977 DOI: 10.3747/pdi.2009.00031]
- 20 **Diaz-Buxo JA**, Gotloib L. Peritoneal dialysis solutions--at a crossroad. *Minerva Urol Nefrol* 2006; **58**: 145-160 [PMID: 16767068]
- 21 **Zeier M**, Schwenger V, Deppisch R, Haug U, Weigel K, Bahner U, Wanner C, Schneider H, Henle T, Ritz E. Glucose degradation products in PD fluids: do they disappear from the peritoneal cavity and enter the systemic circulation? *Kidney Int* 2003; **63**: 298-305 [PMID: 12472796 DOI: 10.1046/j.1523-1755.2003.00705.x]
- 22 **Gotloib L**, Wajsbrodt V, Shostak A. Mesothelial dysplastic changes and lipid peroxidation induced by 7.5% icodextrin. *Nephron* 2002;

- 92: 142-155 [PMID: 12187097 DOI: 10.1159/000064482]
- 23 **Gastaldello K**, Husson C, Dondeyne JP, Vanherweghem JL, Tielemans C. Cytotoxicity of mononuclear cells as induced by peritoneal dialysis fluids: insight into mechanisms that regulate osmotic stress-related apoptosis. *Perit Dial Int* 2008; **28**: 655-666 [PMID: 18981398]
 - 24 **Gotloib L**, Wajsbrot V, Shostak A. Icodextrin-induced lipid peroxidation disrupts the mesothelial cell cycle engine. *Free Radic Biol Med* 2003; **34**: 419-428 [PMID: 12566067 DOI: 10.1016/S0891-5849(02)01296-0]
 - 25 **De Vriese AS**, Tilton RG, Mortier S, Lameire NH. Myofibroblast transdifferentiation of mesothelial cells is mediated by RAGE and contributes to peritoneal fibrosis in uraemia. *Nephrol Dial Transplant* 2006; **21**: 2549-2555 [PMID: 16757496 DOI: 10.1093/ndt/gfl271]
 - 26 **Jiménez-Heffernan JA**, Aguilera A, Aroeira LS, Lara-Pezzi E, Bajo MA, del Peso G, Ramirez M, Gamallo C, Sánchez-Tomero JA, Alvarez V, López-Cabrera M, Selgas R. Immunohistochemical characterization of fibroblast subpopulations in normal peritoneal tissue and in peritoneal dialysis-induced fibrosis. *Virchows Arch* 2004; **444**: 247-256 [PMID: 14749928 DOI: 10.1007/s00428-003-0963-3]
 - 27 **Radisky DC**. Epithelial-mesenchymal transition. *J Cell Sci* 2005; **118**: 4325-4326 [PMID: 16179603 DOI: 10.1242/jcs.02552]
 - 28 **Yáñez-Mó M**, Lara-Pezzi E, Selgas R, Ramírez-Huesca M, Dominguez-Jiménez C, Jiménez-Heffernan JA, Aguilera A, Sánchez-Tomero JA, Bajo MA, Alvarez V, Castro MA, del Peso G, Cirujeda A, Gamallo C, Sánchez-Madrid F, López-Cabrera M. Peritoneal dialysis and epithelial-to-mesenchymal transition of mesothelial cells. *N Engl J Med* 2003; **348**: 403-413 [PMID: 12556543]
 - 29 **Selgas R**, Bajo A, Jiménez-Heffernan JA, Sánchez-Tomero JA, Del Peso G, Aguilera A, López-Cabrera M. Epithelial-to-mesenchymal transition of the mesothelial cell--its role in the response of the peritoneum to dialysis. *Nephrol Dial Transplant* 2006; **21** Suppl 2: ii2-ii7 [PMID: 16825254 DOI: 10.1093/ndt/gfl183]
 - 30 **Williams JD**, Craig KJ, Topley N, Von Ruhland C, Fallon M, Newman GR, Mackenzie RK, Williams GT. Morphologic changes in the peritoneal membrane of patients with renal disease. *J Am Soc Nephrol* 2002; **13**: 470-479 [PMID: 11805177]
 - 31 **Carrero JJ**, Axelsson J, Avesani CM, Heimbürger O, Lindholm B, Stenvinkel P. Being an inflamed peritoneal dialysis patient - a Dante's journey. *Contrib Nephrol* 2006; **150**: 144-151 [PMID: 16721004 DOI: 10.1159/000093514]
 - 32 **Kihm LP**, Müller-Krebs S, Klein J, Ehrlich G, Mertes L, Gross ML, Adaikalakoteswari A, Thornalley PJ, Hammes HP, Nawroth PP, Zeier M, Schwenger V. Benfotiamine protects against peritoneal and kidney damage in peritoneal dialysis. *J Am Soc Nephrol* 2011; **22**: 914-926 [PMID: 21511829 DOI: 10.1681/ASN.2010070750]
 - 33 **Nakayama M**, Izumi G, Nemoto Y, Shibata K, Hasegawa T, Numata M, Wang K, Kawaguchi Y, Hosoya T. Suppression of N(epsilon)-(carboxymethyl)lysine generation by the antioxidant N-acetylcysteine. *Perit Dial Int* 1999; **19**: 207-210 [PMID: 10433156]
 - 34 **Wakabayashi K**, Hamada C, Kanda R, Nakano T, Io H, Horikoshi S, Tomino Y. Oral Astaxanthin Supplementation Prevents Peritoneal Fibrosis in Rats. *Perit Dial Int* 2014; **35**: 506-516 [PMID: 25292409 DOI: 10.3747/pdi.2013.00317]
 - 35 **Zarjie M**, Tangelder GJ, ter Wee PM, Hekking LH, van Lambalgen AA, Keuning ED, Schadee-Eestermans IL, Schalkwijk CG, Beelen RH, van den Born J. Beneficial effects of aminoguanidine on peritoneal microcirculation and tissue remodelling in a rat model of PD. *Nephrol Dial Transplant* 2005; **20**: 2783-2792 [PMID: 16204296 DOI: 10.1093/ndt/gfi138]
 - 36 **Inagi R**, Miyata T, Ueda Y, Yoshino A, Nangaku M, van Ypersele de Strihou C, Kurokawa K. Efficient in vitro lowering of carbonyl stress by the glyoxalase system in conventional glucose peritoneal dialysis fluid. *Kidney Int* 2002; **62**: 679-687 [PMID: 12110033 DOI: 10.1046/j.1523-1755.2002.00488.x]
 - 37 **Liu J**, Zeng L, Zhao Y, Zhu B, Ren W, Wu C. Selenium suppresses lipopolysaccharide-induced fibrosis in peritoneal mesothelial cells through inhibition of epithelial-to-mesenchymal transition. *Biol Trace Elem Res* 2014; **161**: 202-209 [PMID: 25108639 DOI: 10.1007/s12011-014-0091-8]
 - 38 **Lu Y**, Shen H, Shi X, Feng S, Wang Z, Shi Y. Hydrogen sulfide ameliorates high-glucose toxicity in rat peritoneal mesothelial cells by attenuating oxidative stress. *Nephron Exp Nephrol* 2014; **126**: 157-165 [PMID: 24863338 DOI: 10.1159/000358436]
 - 39 **Terawaki H**, Hayashi Y, Zhu WJ, Matsuyama Y, Terada T, Kabayama S, Watanabe T, Era S, Sato B, Nakayama M. Transperitoneal administration of dissolved hydrogen for peritoneal dialysis patients: a novel approach to suppress oxidative stress in the peritoneal cavity. *Med Gas Res* 2013; **3**: 14 [PMID: 23816239 DOI: 10.1186/2045-9912-3-14]
 - 40 **Steffens S**, Veillard NR, Arnaud C, Pelli G, Burger F, Staub C, Karsak M, Zimmer A, Frossard JL, Mach F. Low dose oral cannabinoid therapy reduces progression of atherosclerosis in mice. *Nature* 2005; **434**: 782-786 [PMID: 15815632 DOI: 10.1038/nature03389]
 - 41 **Wheal AJ**, Cipriano M, Fowler CJ, Randall MD, O'Sullivan SE. Cannabidiol improves vasorelaxation in Zucker diabetic fatty rats through cyclooxygenase activation. *J Pharmacol Exp Ther* 2014; **351**: 457-466 [PMID: 25212218 DOI: 10.1124/jpet.114.217125]
 - 42 **Wang D**, Couture R, Hong Y. Activated microglia in the spinal cord underlies diabetic neuropathic pain. *Eur J Pharmacol* 2014; **728**: 59-66 [PMID: 24508519 DOI: 10.1016/j.ejphar.2014.01.057]
 - 43 **El-Remessy AB**, Rajesh M, Mukhopadhyay P, Horváth B, Patel V, Al-Gayyar MM, Pillai BA, Pacher P. Cannabinoid 1 receptor activation contributes to vascular inflammation and cell death in a mouse model of diabetic retinopathy and a human retinal cell line. *Diabetologia* 2011; **54**: 1567-1578 [PMID: 21373835 DOI: 10.1007/s00125-011-2061-4]
 - 44 **El-Remessy AB**, Al-Shabraway M, Khalifa Y, Tsai NT, Caldwell RB, Liou GI. Neuroprotective and blood-retinal barrier-preserving effects of cannabidiol in experimental diabetes. *Am J Pathol* 2006; **168**: 235-244 [PMID: 16400026 DOI: 10.2353/ajpath.2006.050500]
 - 45 **Ulugol A**, Karadag HC, Ipci Y, Tamer M, Dokmeci I. The effect of WIN 55,212-2, a cannabinoid agonist, on tactile allodynia in diabetic rats. *Neurosci Lett* 2004; **371**: 167-170 [PMID: 15519750 DOI: 10.1016/j.neulet.2004.08.061]
 - 46 **Toth CC**, Jedrzejewski NM, Ellis CL, Frey WH. Cannabinoid-mediated modulation of neuropathic pain and microglial accumulation in a model of murine type I diabetic peripheral neuropathic pain. *Mol Pain* 2010; **6**: 16 [PMID: 20236533 DOI: 10.1186/1744-8069-6-16]
 - 47 **Ellington HC**, Cotter MA, Cameron NE, Ross RA. The effect of cannabinoids on capsaicin-evoked calcitonin gene-related peptide (CGRP) release from the isolated paw skin of diabetic and non-diabetic rats. *Neuropharmacology* 2002; **42**: 966-975 [PMID: 12069907 DOI: 10.1016/S0028-3908(02)00040-0]
 - 48 **Horváth B**, Mukhopadhyay P, Haskó G, Pacher P. The endocannabinoid system and plant-derived cannabinoids in diabetes and diabetic complications. *Am J Pathol* 2012; **180**: 432-442 [PMID: 22155112 DOI: 10.1016/j.ajpath.2011.11.003]
 - 49 **Barutta F**, Corbelli A, Mastrocola R, Gambino R, Di Marzo V, Pinach S, Rastaldi MP, Perin PC, Gruden G. Cannabinoid receptor 1 blockade ameliorates albuminuria in experimental diabetic nephropathy. *Diabetes* 2010; **59**: 1046-1054 [PMID: 20068137 DOI: 10.2337/db09-1336]
 - 50 **Barutta F**, Piscitelli F, Pinach S, Bruno G, Gambino R, Rastaldi MP, Salvadio G, Di Marzo V, Cavallo Perin P, Gruden G. Protective role of cannabinoid receptor type 2 in a mouse model of diabetic nephropathy. *Diabetes* 2011; **60**: 2386-2396 [PMID: 21810593 DOI: 10.2337/db10-1809]
 - 51 **Defer N**, Wan J, Souktani R, Escoubet B, Perier M, Caramelle P, Manin S, Deveaux V, Bourin MC, Zimmer A, Lotersztajn S, Pecker F, Pavoine C. The cannabinoid receptor type 2 promotes cardiac myocyte and fibroblast survival and protects against ischemia/reperfusion-induced cardiomyopathy. *FASEB J* 2009; **23**: 2120-2130 [PMID: 19246487 DOI: 10.1096/fj.09-129478]
 - 52 **Mukhopadhyay P**, Bátkai S, Rajesh M, Czifra N, Harvey-White J, Haskó G, Zsengeller Z, Gerard NP, Liaudet L, Kunos G, Pacher P. Pharmacological inhibition of CB1 cannabinoid receptor protects against doxorubicin-induced cardiotoxicity. *J Am Coll*

- Cardiol* 2007; **50**: 528-536 [PMID: 17678736 DOI: 10.1016/j.jacc.2007.03.057]
- 53 **Lim SY**, Davidson SM, Yellon DM, Smith CC. The cannabinoid CB1 receptor antagonist, rimonabant, protects against acute myocardial infarction. *Basic Res Cardiol* 2009; **104**: 781-792 [PMID: 19462153 DOI: 10.1007/s00395-009-0034-2]
- 54 **Montecucco F**, Lenglet S, Braunersreuther V, Burger F, Pelli G, Bertolotto M, Mach F, Steffens S. CB(2) cannabinoid receptor activation is cardioprotective in a mouse model of ischemia/reperfusion. *J Mol Cell Cardiol* 2009; **46**: 612-620 [PMID: 19162037 DOI: 10.1016/j.yjmcc.2008.12.014]
- 55 **Bátkai S**, Rajesh M, Mukhopadhyay P, Haskó G, Liaudet L, Cravatt BF, Csizsár A, Ungvári Z, Pacher P. Decreased age-related cardiac dysfunction, myocardial nitrate stress, inflammatory gene expression, and apoptosis in mice lacking fatty acid amide hydrolase. *Am J Physiol Heart Circ Physiol* 2007; **293**: H909-H918 [PMID: 17434980 DOI: 10.1152/ajpheart.00373.2007]
- 56 **Mukhopadhyay P**, Rajesh M, Bátkaí S, Patel V, Kashiwaya Y, Liaudet L, Evgenov OV, Mackie K, Haskó G, Pacher P. CB1 cannabinoid receptors promote oxidative stress and cell death in murine models of doxorubicin-induced cardiomyopathy and in human cardiomyocytes. *Cardiovasc Res* 2010; **85**: 773-784 [PMID: 19942623 DOI: 10.1093/cvr/cvp369]
- 57 **Rajesh M**, Mukhopadhyay P, Bátkaí S, Patel V, Saito K, Matsumoto S, Kashiwaya Y, Horváth B, Mukhopadhyay B, Becker L, Haskó G, Liaudet L, Wink DA, Vives A, Mechoulam R, Pacher P. Cannabidiol attenuates cardiac dysfunction, oxidative stress, fibrosis, and inflammatory and cell death signaling pathways in diabetic cardiomyopathy. *J Am Coll Cardiol* 2010; **56**: 2115-2125 [PMID: 21144973 DOI: 10.1016/j.jacc.2010.07.033]
- 58 **Siegmund SV**, Qian T, de Minicis S, Harvey-White J, Kunos G, Vinod KY, Hungund B, Schwaibe RF. The endocannabinoid 2-arachidonoyl glycerol induces death of hepatic stellate cells via mitochondrial reactive oxygen species. *FASEB J* 2007; **21**: 2798-2806 [PMID: 17440119 DOI: 10.1096/fj.06-7717com]
- 59 **Cao Z**, Mulvihill MM, Mukhopadhyay P, Xu H, Erdélyi K, Hao E, Holovac E, Haskó G, Cravatt BF, Nomura DK, Pacher P. Monoacylglycerol lipase controls endocannabinoid and eicosanoid signaling and hepatic injury in mice. *Gastroenterology* 2013; **144**: 808-817.e15 [PMID: 23295443 DOI: 10.1053/j.gastro.2012.12.028]
- 60 **Wang M**, Abais JM, Meng N, Zhang Y, Ritter JK, Li PL, Tang WX. Upregulation of cannabinoid receptor-1 and fibrotic activation of mouse hepatic stellate cells during *Schistosoma* J. infection: role of NADPH oxidase. *Free Radic Biol Med* 2014; **71**: 109-120 [PMID: 24657416 DOI: 10.1016/j.freeradbiomed.2014.03.015]
- 61 **Wei Y**, Kang XL, Wang X. The peripheral cannabinoid receptor 1 antagonist VD60 efficiently inhibits carbon tetrachloride-intoxicated hepatic fibrosis progression. *Exp Biol Med* (Maywood) 2014; **239**: 183-192 [PMID: 24459189 DOI: 10.1177/1535370213514922]
- 62 **Patsenker E**, Stoll M, Millonig G, Agaimy A, Wissniewski T, Schneider V, Mueller S, Brenneisen R, Seitz HK, Ocker M, Stickel F. Cannabinoid receptor type 1 modulates alcohol-induced liver fibrosis. *Mol Med* 2011; **17**: 1285-1294 [PMID: 21863215 DOI: 10.2119/molmed.2011.00149]
- 63 **Siegmund SV**, Wojtalla A, Schlosser M, Zimmer A, Singer MV. Fatty acid amide hydrolase but not monoacyl glycerol lipase controls cell death induced by the endocannabinoid 2-arachidonoyl glycerol in hepatic cell populations. *Biochem Biophys Res Commun* 2013; **437**: 48-54 [PMID: 23806692 DOI: 10.1016/j.bbrc.2013.06.033]
- 64 **Lee PJ**, Woo SJ, Jee JG, Sung SH, Kim HP. Bisdemethoxycurcumin Induces apoptosis in activated hepatic stellate cells via cannabinoid receptor 2. *Molecules* 2015; **20**: 1277-1292 [PMID: 25594342 DOI: 10.3390/molecules20011277]
- 65 **DeMorrow S**, Francis H, Gaudio E, Ueno Y, Venter J, Onori P, Franchitto A, Vaculin B, Vaculin S, Alpini G. Anandamide inhibits cholangiocyte hyperplastic proliferation via activation of thioredoxin 1/redox factor 1 and AP-1 activation. *Am J Physiol Gastrointest Liver Physiol* 2008; **294**: G506-G519 [PMID: 18096608 DOI: 10.1152/ajpgi.00304.2007]
- 66 **Borrelli F**, Fasolino I, Romano B, Capasso R, Maiello F, Coppola D, Orlando P, Battista G, Pagano E, Di Marzo V, Izzo AA. Beneficial effect of the non-psychotropic plant cannabinoid cannabigerol on experimental inflammatory bowel disease. *Biochem Pharmacol* 2013; **85**: 1306-1316 [PMID: 23415610 DOI: 10.1016/j.bcp.2013.01.017]
- 67 **Borrelli F**, Aviello G, Romano B, Orlando P, Capasso R, Maiello F, Guadagno F, Petrosino S, Capasso F, Di Marzo V, Izzo AA. Cannabidiol, a safe and non-psychotropic ingredient of the marijuana plant *Cannabis sativa*, is protective in a murine model of colitis. *J Mol Med (Berl)* 2009; **87**: 1111-1121 [PMID: 19690824 DOI: 10.1007/s00109-009-0512-x]
- 68 **Mukhopadhyay P**, Rajesh M, Pan H, Patel V, Mukhopadhyay B, Bátkaí S, Gao B, Haskó G, Pacher P. Cannabinoid-2 receptor limits inflammation, oxidative/nitrosative stress, and cell death in nephropathy. *Free Radic Biol Med* 2010; **48**: 457-467 [PMID: 19969072 DOI: 10.1016/j.freeradbiomed.2009.11.022]
- 69 **Pan H**, Mukhopadhyay P, Rajesh M, Patel V, Mukhopadhyay B, Gao B, Haskó G, Pacher P. Cannabidiol attenuates cisplatin-induced nephrotoxicity by decreasing oxidative/nitrosative stress, inflammation, and cell death. *J Pharmacol Exp Ther* 2009; **328**: 708-714 [PMID: 19074681 DOI: 10.1124/jpet.108.147181]
- 70 **Servettaz A**, Kaviani N, Nicco C, Deveaux V, Chéreau C, Wang A, Zimmer A, Lotersztajn S, Weill B, Batteux F. Targeting the cannabinoid pathway limits the development of fibrosis and autoimmunity in a mouse model of systemic sclerosis. *Am J Pathol* 2010; **177**: 187-196 [PMID: 20508030 DOI: 10.2353/ajpath.2010.090763]
- 71 **Yang AH**, Chen JY, Lin YP, Huang TP, Wu CW. Peritoneal dialysis solution induces apoptosis of mesothelial cells. *Kidney Int* 1997; **51**: 1280-1288 [PMID: 9083298 DOI: 10.1038/ki.1997.175]
- 72 **Jin HM**, Di YD, Xu QJ. Effects of commercial glucose-based peritoneal dialysates on peripheral blood phagocytes apoptosis. *Perit Dial Int* 1999; **19** Suppl 2: S388-S393 [PMID: 10406552]
- 73 **Kjellstrand P**, Martinson E, Wieslander A, Holmquist B. Development of toxic degradation products during heat sterilization of glucose-containing fluids for peritoneal dialysis: influence of time and temperature. *Perit Dial Int* 1995; **15**: 26-32 [PMID: 7734557]
- 74 **Nilsson-Thorell CB**, Muscalu N, Andrén AH, Kjellstrand PT, Wieslander AP. Heat sterilization of fluids for peritoneal dialysis gives rise to aldehydes. *Perit Dial Int* 1993; **13**: 208-213 [PMID: 8369351]
- 75 **Jörres A**. Glucose degradation products in peritoneal dialysis: from bench to bedside. *Kidney Blood Press Res* 2003; **26**: 113-117 [PMID: 12771536 DOI: 10.1159/000070993]
- 76 **Linden T**, Forsbäck G, Deppisch R, Henle T, Wieslander A. 3-Deoxyglucosone, a promoter of advanced glycation end products in fluids for peritoneal dialysis. *Perit Dial Int* 1998; **18**: 290-293 [PMID: 9663893]
- 77 **Nakayama M**, Sakai A, Numata M, Hosoya T. Hyper-vascular change and formation of advanced glycation endproducts in the peritoneum caused by methylglyoxal and the effect of an antioxidant, sodium sulfite. *Am J Nephrol* 2003; **23**: 390-394 [PMID: 14551463 DOI: 10.1159/000074065]
- 78 **Riboulet-Chavey A**, Pierron A, Durand I, Murdaca J, Giudicelli J, Van Obberghen E. Methylglyoxal impairs the insulin signaling pathways independently of the formation of intracellular reactive oxygen species. *Diabetes* 2006; **55**: 1289-1299 [PMID: 16644685 DOI: 10.2337/db05-0857]
- 79 **Hirahara I**, Ishibashi Y, Kaname S, Kusano E, Fujita T. Methylglyoxal induces peritoneal thickening by mesenchymal-like mesothelial cells in rats. *Nephrol Dial Transplant* 2009; **24**: 437-447 [PMID: 18790810 DOI: 10.1093/ndt/gfn495]
- 80 **Schalkwijk CG**, ter Wee PM, Teerlink T. Reduced 1,2-dicarbonyl compounds in bicarbonate/lactate-buffered peritoneal dialysis (PD) fluids and PD fluids based on glucose polymers or amino acids. *Perit Dial Int* 2000; **20**: 796-798 [PMID: 11216581]
- 81 **Aroeira LS**, Aguilera A, Sánchez-Tomero JA, Bajo MA, del Peso G, Jiménez-Heffernan JA, Selgas R, López-Cabrera M. Epithelial to mesenchymal transition and peritoneal membrane failure in

- peritoneal dialysis patients: pathologic significance and potential therapeutic interventions. *J Am Soc Nephrol* 2007; **18**: 2004-2013 [PMID: 17568021 DOI: 10.1681/ASN.2006111292]
- 82 **Stavenuiter AW**, Schilte MN, Ter Wee PM, Beelen RH. Angiogenesis in peritoneal dialysis. *Kidney Blood Press Res* 2011; **34**: 245-252 [PMID: 21691127 DOI: 10.1159/000326953]
- 83 **Schilte MN**, Celie JW, Wee PM, Beelen RH, van den Born J. Factors contributing to peritoneal tissue remodeling in peritoneal dialysis. *Perit Dial Int* 2009; **29**: 605-617 [PMID: 19910560]
- 84 **Jörres A**, Bender TO, Witowski J. Glucose degradation products and the peritoneal mesothelium. *Perit Dial Int* 2000; **20** Suppl 5: S19-S22 [PMID: 11229607]
- 85 **Mortier S**, Faict D, Schalkwijk CG, Lameire NH, De Vriese AS. Long-term exposure to new peritoneal dialysis solutions: Effects on the peritoneal membrane. *Kidney Int* 2004; **66**: 1257-1265 [PMID: 15327425 DOI: 10.1111/j.1523-1755.2004.00879.x]
- 86 **Mittelmaier S**, Niwa T, Pischetsrieder M. Chemical and physiological relevance of glucose degradation products in peritoneal dialysis. *J Ren Nutr* 2012; **22**: 181-185 [PMID: 22200439 DOI: 10.1053/j.jrn.2011.10.014]
- 87 **Qi H**, Xu C, Yan H, Ma J. Comparison of icodextrin and glucose solutions for long dwell exchange in peritoneal dialysis: a meta-analysis of randomized controlled trials. *Perit Dial Int* 2011; **31**: 179-188 [PMID: 21119069 DOI: 10.3747/pdi.2009.00264]
- 88 **Canepa A**, Verrina E, Perfumo F. Use of new peritoneal dialysis solutions in children. *Kidney Int Suppl* 2008; **108**: S137-S144 [PMID: 18379537 DOI: 10.1038/sj.ki.5002615]
- 89 **Yang CY**, Chau YP, Lee HT, Kuo HY, Lee OK, Yang AH. Cannabinoid receptors as therapeutic targets for dialysis-induced peritoneal fibrosis. *Am J Nephrol* 2013; **37**: 50-58 [PMID: 23296044 DOI: 10.1159/000345726]
- 90 **Bátkai S**, Járjai Z, Wagner JA, Goparaju SK, Varga K, Liu J, Wang L, Mirshahi F, Khanolkar AD, Makriyannis A, Urbaschek R, Garcia N, Sanyal AJ, Kunos G. Endocannabinoids acting at vascular CB1 receptors mediate the vasodilated state in advanced liver cirrhosis. *Nat Med* 2001; **7**: 827-832 [PMID: 11433348 DOI: 10.1038/89953]
- 91 **Julien B**, Grenard P, Teixeira-Clerc F, Van Nhieu JT, Li L, Karsak M, Zimmer A, Mallat A, Lotersztajn S. Antifibrogenic role of the cannabinoid receptor CB2 in the liver. *Gastroenterology* 2005; **128**: 742-755 [PMID: 15765409 DOI: 10.1053/j.gastro.2004.12.050]
- 92 **Lotersztajn S**, Teixeira-Clerc F, Julien B, Deveaux V, Ichigotani Y, Manin S, Tran-Van-Nhieu J, Karsak M, Zimmer A, Mallat A. CB2 receptors as new therapeutic targets for liver diseases. *Br J Pharmacol* 2008; **153**: 286-289 [PMID: 17952109 DOI: 10.1038/sj.bjp.0707511]
- 93 **Teixeira-Clerc F**, Julien B, Grenard P, Tran Van Nhieu J, Deveaux V, Li L, Serriere-Lanneau V, Ledent C, Mallat A, Lotersztajn S. CB1 cannabinoid receptor antagonism: a new strategy for the treatment of liver fibrosis. *Nat Med* 2006; **12**: 671-676 [PMID: 16715087 DOI: 10.1038/nm1421]
- 94 **Caraceni P**, Pertosa AM, Giannone F, Domenicali M, Grattagliano I, Principe A, Mastroleone C, Perrelli MG, Cutrin J, Trevisani F, Croci T, Bernardi M. Antagonism of the cannabinoid CB-1 receptor protects rat liver against ischaemia-reperfusion injury complicated by endotoxaemia. *Gut* 2009; **58**: 1135-1143 [PMID: 19282305 DOI: 10.1136/gut.2007.147652]
- 95 **Giannone FA**, Baldassarre M, Domenicali M, Zaccherini G, Trevisani F, Bernardi M, Caraceni P. Reversal of liver fibrosis by the antagonism of endocannabinoid CB1 receptor in a rat model of CCl(4)-induced advanced cirrhosis. *Lab Invest* 2012; **92**: 384-395 [PMID: 22184091 DOI: 10.1038/labinvest.2011.191]
- 96 **Yang YY**, Lin HC, Huang YT, Lee TY, Hou MC, Wang YW, Lee FY, Lee SD. Effect of chronic CB1 cannabinoid receptor antagonism on livers of rats with biliary cirrhosis. *Clin Sci (Lond)* 2007; **112**: 533-542 [PMID: 17176248 DOI: 10.1042/CS20060260]
- 97 **Demuth DG**, Molleman A. Cannabinoid signalling. *Life Sci* 2006; **78**: 549-563 [PMID: 16109430 DOI: 10.1016/j.lfs.2005.05.055]
- 98 **Pertwee RG**. Cannabinoid pharmacology: the first 66 years. *Br J Pharmacol* 2006; **147** Suppl 1: S163-S171 [PMID: 16402100 DOI: 10.1038/sj.bjp.0706406]
- 99 **Montecucco F**, Burger F, Mach F, Steffens S. CB2 cannabinoid receptor agonist JWH-015 modulates human monocyte migration through defined intracellular signaling pathways. *Am J Physiol Heart Circ Physiol* 2008; **294**: H1145-H1155 [PMID: 18178718 DOI: 10.1152/ajpheart.01328.2007]
- 100 **González-Mateo GT**, Aroeira LS, López-Cabrera M, Ruiz-Ortega M, Ortiz A, Selgas R. Pharmacological modulation of peritoneal injury induced by dialysis fluids: is it an option? *Nephrol Dial Transplant* 2012; **27**: 478-481 [PMID: 21965583 DOI: 10.1093/ndt/gfr543]
- 101 **Gardner B**, Zu LX, Sharma S, Liu Q, Makriyannis A, Tashkin DP, Dubinett SM. Autocrine and paracrine regulation of lymphocyte CB2 receptor expression by TGF-beta. *Biochem Biophys Res Commun* 2002; **290**: 91-96 [PMID: 11779138 DOI: 10.1006/bbrc.2001.6179]
- 102 **Onishi A**, Akimoto T, Urabe M, Hirahara I, Muto S, Ozawa K, Nagata D, Kusano E. Attenuation of methylglyoxal-induced peritoneal fibrosis: immunomodulation by interleukin-10. *Lab Invest* 2015; **95**: 1353-1362 [PMID: 26367488 DOI: 10.1038/labinvest.2015.110]
- 103 **Siegmund SV**, Uchinami H, Osawa Y, Brenner DA, Schwabe RF. Anandamide induces necrosis in primary hepatic stellate cells. *Hepatology* 2005; **41**: 1085-1095 [PMID: 15841466 DOI: 10.1002/hep.20667]
- 104 **Willecke F**, Zeschky K, Ortiz Rodriguez A, Colberg C, Auwärter V, Kneisel S, Hutter M, Lozhkin A, Hoppe N, Wolf D, von zur Mühlen C, Moser M, Hilgendorf I, Bode C, Zirlik A. Cannabinoid receptor 2 signaling does not modulate atherogenesis in mice. *PLoS One* 2011; **6**: e19405 [PMID: 21541300 DOI: 10.1371/journal.pone.0019405]
- 105 **Hirahara I**, Kusano E, Yanagiba S, Miyata Y, Ando Y, Muto S, Asano Y. Peritoneal injury by methylglyoxal in peritoneal dialysis. *Perit Dial Int* 2006; **26**: 380-392 [PMID: 16722033]
- 106 **Yang AH**, Huang SW, Chen JY, Lin JK, Chen CY. Leptin augments myofibroblastic conversion and fibrogenic activity of human peritoneal mesothelial cells: a functional implication for peritoneal fibrosis. *Nephrol Dial Transplant* 2007; **22**: 756-762 [PMID: 17142261 DOI: 10.1093/ndt/gfl715]
- 107 **van Westrhenen R**, Aten J, Hajji N, de Boer OJ, Kunne C, de Waart DR, Krediet RT. Cyclosporin A induces peritoneal fibrosis and angiogenesis during chronic peritoneal exposure to a glucose-based, lactate-buffered dialysis solution in the rat. *Blood Purif* 2007; **25**: 466-472 [PMID: 18087149 DOI: 10.1159/000112475]
- 108 **Yang AH**, Chen JY, Lin JK. Myofibroblastic conversion of mesothelial cells. *Kidney Int* 2003; **63**: 1530-1539 [PMID: 12631370 DOI: 10.1046/j.1523-1755.2003.00861.x]
- 109 **Kang DH**, Hong YS, Lim HJ, Choi JH, Han DS, Yoon KI. High glucose solution and spent dialysate stimulate the synthesis of transforming growth factor-beta1 of human peritoneal mesothelial cells: effect of cytokine costimulation. *Perit Dial Int* 1999; **19**: 221-230 [PMID: 10433158]
- 110 **De Vriese AS**, Flyvbjerg A, Mortier S, Tilton RG, Lameire NH. Inhibition of the interaction of AGE-RAGE prevents hyperglycemia-induced fibrosis of the peritoneal membrane. *J Am Soc Nephrol* 2003; **14**: 2109-2118 [PMID: 12874465]

P- Reviewer: Demonacos C, Riutta AA, Swierczynski JT

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