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***Elizabethkingia miricola*: A rare non-fermenter causing urinary tract infection**

GuptaP *et al. Elizabethkingia miricola*: A rare uropathogen

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**Abstract**

*Elizabethkingia miricola* is a gram-negative non-fermentative bacterium which is rarely encountered. It is usually misidentified or considered as a contaminant in routine microbiology laboratories due to the limitations in conventional biochemical techniques. However, with the advent of the matrix-assisted laser desorption ionization time-of-flight mass spectrometry (MALDI-TOF-MS), the identification of non-fermenters has become easy and this has led to enhanced understanding of the clinical significance of these uncommonly isolated microorganisms. The genus *Elizabethkingia* has only two species *E. meningoseptica* and *E. miricola*. Both of these organisms are known to be multi-drug resistant and therefore, their accurate identification and antimicrobial susceptibility testing are necessary prior to the initiation of appropriate therapy. In the world literature till date, only 3 cases of sepsis caused by *E. miricola* have been reported. We present the first case of *E. miricola* association with urinary tract infection.

**Key words:** *Elizabethkingia miricola*; Urinary tract infections; Matrix-assisted laser desorption ionization time-of-flight; Non-fermenters; Antibiotics

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**Core tip:** Non-fermenters except *Pseudomonas* and *Acinetobacter* are less commonly associated with urinary tract infection (UTI). But recently an upsurge in a number of reported cases has been noted due to the use of MALDI-TOF which is an easy and reliable identification technique. Till date in literature, there is no reported case of *E. miricola* causing UTI, although its significance in blood and sputum samples of sepsis patients has been demonstrated earlier. This is the first case report showing a clinical association of *E. miricola* with symptomatic UTI and also demonstrating the multidrug resistance nature of this organism.

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**INTRODUCTION**

Urinary tract infections (UTI) are amongst the most common bacterial infections occurring in human beings during their lifetime[1]. The usual organisms responsible for UTI belong to the family *Enterobacteriaceae* and gram-positive bacteria like *Staphylococcus* and *Enterococcus*[2]*.* UTI caused by non-fermenters (NF) is being increasingly reported especially in the nosocomial settings, with *Pseudomonas and Acinetobacter spp.* being the most common agents. However, UTI due to other NFs like *Alcaligenes, Flavobacterium, Oligella, Flavimonas, Agrobacter, Weeksiella* are also on the rise[3]. Routine laboratory identification of NF is difficult and labour-intensive, which often misclassifies or misidentify these agents and thereby may mask the exact clinical significance of these isolates. Nowadays, the identification of these NF has become easy by the advent of matrix-assisted laser desorption ionization time-of-flight mass spectrometry (MALDI-TOF-MS). We recently encountered a case of UTI caused by rare multidrug resistant non-fermenter *E. miricola*, which was identified by MALDI-TOF.

**CASE REPORT**

A 25-year-old female presented with complaints of increased bowel frequency, oliguria, fever and abdominal pain since one month. Detailed history revealed that the patient had difficulty in micturition for past two weeks. The routine laboratory investigations revealed a haemoglobin of 7.8 gm%, total leucocytes count 3200 cells/ mm3, platelets count of 70000 cells/mm3. Renal function tests revealed normal sodium concentration (139 mEq/L), hyperkalemia (8.2 mEq/L), hyperureamia (74 mg/dL) and elevated creatinine levels (7.5 mg/dL). Coagulation profile was normal. Ultrasonography (USG) revealed bilateral hydroureteronephrosis with normal renal parenchyma and features of vesicoureteric reflux. The midstream urine sample was subjected to microbiological testing. The wet mount microscopic examination showed 1-2 RBCs, numerous pus cells and bacteria per high-power field[4]. The semi-quantitative culture done on the cysteine lysine electrolyte deficient agar showed significant bacterial growth (colony count > 105 CFU/mL). The colonies were non-lactose fermenting, translucent, greenish blue, smooth having entire edges and became mucoid on prolonged incubation. Subculture on MacConkey agar showed pale, translucent, glistening colonies with entire edges (Figure 1). Gram staining showed 0.5 µm × 2 µm gram-negative bacilli, with no spores and no capsule. The isolate was also subjected to conventional identification using a battery of biochemical tests. The isolate was catalase positive, oxidase positive, produced indole, non-nitrate reducing, mannitol fermenting, esculin and gelatinase hydrolysis positive. Urease was produced and this test helped to distinguish it from *E. meningoseptica*. The isolate was confirmed as *Elizabethkingia miricola* (identification score of 2.29) by using MALDI-TOF-MS (BrukerDaltonics, Bremen, Germany). The antimicrobial susceptibility was carried out using Kirby-Bauer disc diffusion method and the antibiotics tested were chosen from the available literature as there are no CLSI guidelines available till now[5,6].The isolate was sensitive to gentamicin, ceftriaxone, aztreonam, piperacillin-tazobactam and imipenem, and resistant to ampicillin, ciprofloxacin, levofloxacin, vancomycin and colistin. The patient was started on piperacillin-tazobactam and responded well to the treatment. The patient improved clinically and the follow-up urine culture after two weeks of therapy was sterile.

**DISCUSSION**

*Elizabethkingia miricola* was first isolated from Mir space station, Russia and hence named as *E. miricola*[7]. Previously, it was classified into genus *Chryseobacterium* but later in 2005, the genus was changed to *Elizabethkingea* on the basis of the comparative analytical studies involving DNA hybridization and sequencing of the 16S rRNA region[8].*Elizabethkingia miricola* is a gram-negative (0.5 µm × 1-2.5 µm), non-motile, non-spore-forming bacterium. It grows well on blood and MacConkey agar producing non-fermenting sticky colonies. Biochemical reactions show indole positive, citrate positive, produce acid from D-glucose, D-fructose, D-lactose, trehalose, D-mannitol, D-maltose, but not from D-xylose, L-arabinose, D-cellobiose, sucrose and raffinose. It can be differentiated from *Chryseobacterium* because of the absence of yellow pigment in culture. Urease production is the test used to differentiate *E. miricola* from *E. meningoseptica*[8]. Till date*, Elizabethkingia miricola* has been isolated from blood and sputum and has been found to be responsible for sepsis. The first case of *E. miricola* was reported in 2008 in an adult with mantle cell carcinoma, who underwent stem cell transplant[5]. In this case, *E. miricola* was isolated from sputum and blood and the identification was confirmed using 16S rRNA sequencing. Later on, *E. miricola* was isolated from the blood sample of a young female with alcoholic pancreatitis[6].More recently, *E. miricola* has been isolated from a patient with severe sepsis and pulmonary abscess[9]. In both the above cases, the isolate was identified by MALDI-TOF. In the present case, *Elizabethkingia miricola* was isolated from the urine sample of a young female with clinical features of UTI and bilateral hydroureteronephrosis. The clinical presentation pointed towards differential diagnosis like pyelonephritis, renal abscess, renal infarction, venous obstruction or ATN. However, the USG findings of bilateral hydroureteronephrosis and sterile blood culture pointed towards localised urinary tract infection.

*Elizabethkingia miricola* has been found to be multidrug resistant similar to as *E. meningoseptica* which are known to harbor β-lactamases showing resistant to β-lactams and carbapenems[10]. The *E. miricola* isolates have been found to be resistant to many antibiotics. Previous studies have shown resistance to ampicillin, ceftazidime, imipenem, gentamicin, cotrimoxazole, colistin and with variable susceptibility to ciprofloxacin, vancomycin and rifampicin[5,6,11]. It is interesting to note that, *E. miricola* isolates in previous studies were sensitive to levofloxacin, but in our case, the isolate was resistant to both ciprofloxacin and levofloxacin. With limited clinical reports, varied susceptibility profiles, lack of antimicrobial susceptibility breakpoint and no defined consensus for the empiric treatment regimen makes it difficult to treat such rare organisms.

We present the first case report of human UTI caused by rare multidrug resistant *E. miricola*. The present case emphasizes the clinical importance of rare non-fermenter like *E. miricola* in human infections especially in the case of UTI*.* The knowledge of newer species and their antimicrobial susceptibility profile will help in formulating appropriate antibiotic treatment regimens to tackle such rarely encountered bacteria.

**COMMENTS**

***Case characteristics***

A 25-year-old female complaining of difficulty in micturition, oliguria fever with abdominal pain.

***Clinical diagnosis***

Urinary tract infections (UTI) with bilateral hydroureteronephrosis

***Differential diagnosis***

Chronic pyelonephritis.

***Laboratory diagnosis***

The routine laboratory investigations revealed anemia, leucopenia, hyperkalemia, hyperuricaemia and elevated creatinine levels. Urine culture had significant bacterial growth (colony count >105 CFU/mL) of *E. miricola*.

***Imaging diagnosis***

Bilateral hydroureteronephrosis.

***Pathological diagnosis***

Bilateral hydroureteronephrosis with urinary tract infection.

***Treatment***

Piperacillin-tazobactam.

***Related reports***

*E. miricola* has been reported to cause sepsis and pulmonary infection.

***Experiences and lessons***

Rare non-fermenters can cause UTI and prompt identification is required to guide proper antimicrobial therapy. CLSI/EUCAST guidelines need to be developed.

***Peer-review***

Interesting case of unusual bacterial cause of UTI with a severe clinical scenario.

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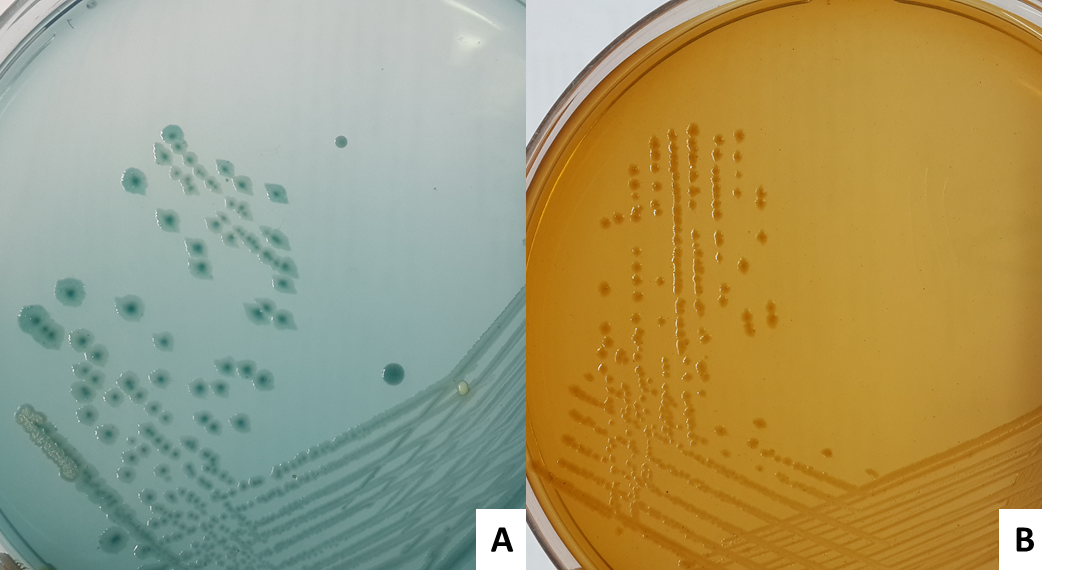
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**Figure 1 Culture plates showing the growth of *Elizabethkingia miricola* on (A) CLED agar and (B) MacConkey agar**.