**Name of Journal: *World Journal of Transplantation***

**Manuscript NO: 32642**

**Manuscript Type: Minireviews**

**Polyoma virus nephropathy in kidney transplantation**

Scadden JRW *et al.* BKVN in kidney transplantation

**Jacob RW Scadden, Adnan Sharif, Kassi Skordilis, Richard Borrows**

**Jacob RW Scadden,** University of Birmingham, Edgbaston, Birmingham B15 2TH, United Kingdom

**Adnan Sharif,** **Richard Borrows,** Department of Kidney Transplantation, Queen Elizabeth Hospital Birmingham, Birmingham B15 2TH, United Kingdom

**Kassi Skordilis,** Department of Renal Histopathology, Queen Elizabeth Hospital Birmingham, Birmingham B15 2TH, United Kingdom

**Author contributions:** Scadden JRW, Sharif A and Borrows R devised, wrote and edited manuscript; Skordilis K edited manuscript and provided histology.

**Conflict-of-interest statement:** Dr. Borrows has nothing to disclose.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Correspondence to: Richard Borrows, Consultant Nephrologist,** Department of Kidney Transplantation, Queen Elizabeth Hospital Birmingham, Edgbaston, Birmingham B15 2TH, United Kingdom. richard.borrows@uhb.nhs.uk

**Telephone:** +44-121-3716099

**Fax:** +44**-**121-3716098

**Received:** January 18, 2017

**Peer-review started:** January 19, 2017

**First decision:** February 15, 2017

**Revised:** November 17, 2017

**Accepted:** December 1, 2017

**Article in press:**

**Published online:**

**Abstract**

BK virus (BKV) is a polyomavirus that is able to cause renal dysfunction in transplanted grafts *via* BK virus-associated nephritis (BKVAN). This condition was mis-diagnosed in the past due to clinical and histopthological similarities with acute rejection. Due to the prevalence of the virus in the population, it is an important pathogen in this context, and so it is important to understand how this virus functions and its’ relationship with the pathogenesis of BKVN. Screening for BKV often reveals viruria and/or viremia, which then manifests as BKVN, which can be asymptomatic or result in clinical features namely renal dysfunction. The pathogenesis of BKV infection is still unclear and needs to be further investigated; nevertheless there are a variety of hypotheses that indicate that there are a host of factors that play important roles. Treatments for BKVAN include a reduction in immunosuppression, the use of antiviral therapy or the combination of both treatment options.

**Key words:** Polyoma; Kidney; Transplant; Infection; Virus

**© The Author(s) 2017.** Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** Prior to its recognition as a separate entity, kidney transplant infection with the polyoma virus, BK virus (BKV), and the ensuing viral nephropathy (BKVN) portended a poor prognosis. But with the advent of heightened clinical suspicion and improved diagnostics the prognosis has improved considerably. Blood and urine polymerase chain reaction testing allows invasive investigation (*i.e*., transplant biopsy) to be selective and appropriate. Peripheral blood assays of anti-BKV cell mediated immunity offers potential for refining risk stratification. While conventional antiviral agents have failed to show utility to date, reduction of immunosuppression currently represents the most effective and proven treatment for BKVN.

Scadden JRW, Sharif A, Skordilis K, Borrows R. Polyoma virus nephropathy in kidney transplantation. *World J Transplant* 2017; In press

**BACKGROUND**

BK virus (BKV) was first isolated from the urine of a patient with transplant ureteric stenosis[1] in 1971. BKV is a member of the *Polyomaviridae* family, falling into the *Betapolyomavirus* subcategory with JC virus and Simian virus 40 (SV40)[2]. BKV and SV40 have approximately 70% similarity within their respective genomes[3,4]. This similarity between the genomes of these viruses enables SV40 to be a marker for immunohistochemical staining, which is vital in diagnosis of BKV-associated nephropathy (BKVAN)[4]. BKV primary infection occurs in early childhood and is asymptomatic in the majority of cases[5,6]. Transmission of BKV is thought to involve respiratory and oral routes[7], and results in a seroprevalence of 82% in adulthood[8]. BKV is a latent infection, which can lie dormant in tissues, the kidney being the most notable. Heritage *et al*[9] showed that BKV was present in 50% of kidneys based on DNA sequencing of BKV in renal samples. The virus is able to reside in renal tubular epithelial cells and in the uroepithelium. Therefore, another “artificial” route of BKV transmission is through kidney transplantation, particularly on the background of the immunosuppression required in this context to prevent organ rejection. Moreover, although primary infection in non-transplanted patients is generally asymptomatic, viral infection in the context of transplantation and immunosuppression may result in viral replication within epithelial tissues (in this case renal tubules), and the development of inflammation, BKVAN, which resembles other forms of tubule-interstitial nephritis and transplant rejection. If left untreated, these processes progress to result in allograft dysfunction and failure[10]. Indeed, North American and European series suggest that although not the leading overall cause of graft failure, it represents an important and potentially treatable (even preventable) cause in many[11,12]. For many years, BKVAN was mis-diagnosed as rejection, and therefore inappropriately treated, resulting in graft failure in many patients. Nevertheless, greater understanding of BKV has resulted in clinical advances in the field. In this literature review the virology of BKV, the mechanism of BKVAN pathogenesis, and advances in therapeutic strategies will be addressed.

**VIROLOGY OF BKV**

BKV is a member of the *Polyomavirus* genus of the *Polyomaviridae* family. These viruses are 40-45 nm in diameter[1,13] comprising of an icosahedral capsid surrounding double-stranded DNA, which is able to replicate in the host cell nucleus[14]. The BKV genome contains 5153 base pairs that can be translated bi-directionally[6,15,16]. However, recent analysis of BKV from a kidney transplant patient observed a genome size of 5141 base pairs[17]. The BKV genome is divided into three regions: Early, late and regulatory (non-coding control region or NCCR) regions. The early stage encompasses regulatory proteins such as small tumour antigens (tAg) and large tumour antigens (TAg) as well as late structural capsid proteins - viral protein (VP) 1, VP2, VP3 and the agnoprotein[18]. However, these late proteins are produced after the genome of the virus has be replicated[19]. VP1 is the most common protein found on the outer layer of the capsid and contains a small groove used for host cell receptor binding[20]. The NCCR encodes transcriptional control elements, such as the origin of replication and promoters for genes encoded within the early and late regions[11,21]. BKV enters cells *via* VP1 binding to sialic acid residues of glycoprotein receptors[22,23]. After receptor binding BKV is internalised *via* a caveolae-mediated endocytosis pathway, the virus then travelling to the cell nucleus to launch either a latent or acute infection[13,24,25]. According to Jin *et al*[26], four serotypes (I-IV) of BKV are recognised based on the differences between amino acids 61-83 in the region coding for VP1, with the similarity of this region between different serotypes at 61% to 70%[27]. Serotype I is the most common within the worldwide population (80%), followed by Type IV (15%)[28]. However, Sharma *et al*[29], using a phylogenetic whole-genome approach, suggested a classification system of BKV which contain serotypes V and VI.

**PATHOGENESIS OF BKVAN**

There are many proposed factors relating to the pathogenesis of BKVAN as shown in Figure 1: Source of the viral infection; host cellular immunity to BKV; the influence of immunosuppression; HLA matching; recipient and donor blood group matching.

***Source: Donor vs recipient***

BKVAN within transplanted kidneys arises from either primary infection from the transplanted kidney itself, or following reactivation from latency in the patient’s native urinary tract. Epidemiological work has striven to understand the predominant mechanism in this context. Andrews *et al*[30] were the first to show that recipients of transplants from seropositive donors (either deceased or living), was associated with increased rate of BKV infection within the transplant kidney, thereby suggesting the importance of donor-derived infection in the pathogenesis of BKVAN. This was supported by data from Bohl *et al*[31] who observed BKV infection in 25 of 54 (46%) recipients of kidneys from seropositive donors compared with 4 of 27 (15%) recipients of kidneys from seronegative donors (*P* = 0.007). These authors also noted that the rate at which BK viruria occurred was faster in patients receiving kidneys from seropositive versus seronegative donors (median onset 45 d *vs* 370 d respectively; *P* < 0.001). The duration of BK viruria was also longer in the context of seropositive donors (median duration 157 d *vs* 7 d; *P* = 0.009). The authors also recorded that none of the 27 recipients from seronegative donors developed viremia or sustained viremia, whereas these numbers were 7 and 4 respectively in recipients from seropositive donors. In addition, this study demonstrated donor BKV antibody titre inversely correlated with the time to onset of post-transplant viruria (*P* = 0.001), and was positively correlated with duration of viruria (*P* = 0.014) and peak urine viral titres (*P* = 0.005). These studies did not evaluate the importance of recipient BKV serostatus, but this was done in a study of paediatric recipients[32], which suggested the importance of recipient BK seronegativity. In this study, all patients developed BKV viruria, with recipient seronegativity strongly associated with the development of nephropathy (*P* = 0.01). Finally, Saundh *et al*[33] studied 112 renal transplant patients before and after transplantation, and conducted a phylogenetic analysis of VP1 sequences and serotypes. Twelve patients developed BKV viremia, and 8 had a sufficiently high viral load to allow amplification of VP1. Based on this analysis the authors concluded that donor-derived infection was responsible for the majority of cases of BKV infection. A single patient had two differing VP1 subgroups present (Ib-1 after 6 mo followed by Ia after 12 mo post transplantation), perhaps suggesting a potential switch between donor and recipient strains, and means that cases of BKV infection due to reactivation from the recipient may be a real phenomenon. However, the burden of evidence from this, and the other aforementioned studies, is that donor-derived infection represents the major risk confronting the kidney transplant recipient.

***Cellular and humoral immunity***

Humoral and cellular immunity is thought to be implicated with the pathogenesis of BKVAN, and both CD8+ and CD4+ T cells are involved in the recognition and clearance of viruses such as BKV. The lack of BKV specific IgG may be important in the development of BKVAN[12]. As mentioned above, there is a greater risk for patients that are BKV seronegative at the time of their transplant, as they will have no BKV specific antibody; patients with previous exposure and who have developed immunity to BKV, may not develop the infection[34]. Yet the presence of a BKV-specific antibody response is clearly not protective, and it is likely that cellular immunity plays the central role in viral control. Certainly, patients with BKV specific antibodies remain at risk of developing BKVAN[35]. Comoli *et al*[36] showed that patients that had BKVAN had fewer BKV-specific lymphocytes that secreted IFN-γ, with the mean frequency of BKV specific, IFN-γ being 151/106 cells. This was approximately 10 times less than other viruses related to transplantation, such as EBV. The researchers concluded that, based on their data that there is reduced BKV immunity, which in turn would increase the rate of active BKV infection.

***Immunosuppression burden***

Immunosuppression is required to ameliorate rejection of the transplanted kidney by the host immune system. However, with developments in immunosuppressive drugs such as mycophenolate mofetil (MMF) and tacrolimus (Tac), the reduction in rejection rate has been inversely paralleled by an increased incidence of BKV infection. Calcineurin inhibitor (CNI) based therapies have been shown to increase the risk of BKVAN and subsequent nephrotoxicity following renal transplant[37]. However, a recent study by Jacobi *et al*[38] observed no significant change in the number of patients with BKV infection (*n* = 352) when using CNI of either tacrolimus or cyclosporine A (CyA). This led to the conclusion that CNI immunosuppressants were not associated in BK viremia.

Mengel *et al*[39] suggested particularly increased risk of BKV nephropathy with a combination of Tac and MMF. Similarly, Brennan *et al*[40] showed that viruria was most common with a drug combination of Tac-MMF (46%) and lowest with cyclosporine-MMF (13%). These authors also confirmed the association between viruria and subsequent BKV viremia. In addition, when surveillance for BK viremia was undertaken for the purposes of this study, a reduction in immunosuppression in response to detectable viremia resulted in reductions in viral load in 95% of patients, and without increased risk of rejection. A more recent randomised study also supports the role of immunosuppression in this context. In a study of 682 patients, the combination of Tac and MMF was associated with greater rates of viremia at 6 and 12 mo post-transplantation than the combination of cyclosporine and MMF (16.3% *vs* 10.6%, *P* = 0.048 and 12.1% and 4.8%, *P* = 0.004 respectively). Cumulative steroid dose up to month 3 was also a risk factor for viremia in this study, highlighting the role of overall immunosuppression burden in this disease[41]. Clinical observations of campth therapy were made by Kayler *et al*[42], they administered campth to two patients with BKV viruria and one with nephropathy. In all cases, there was increased viral replication and one of the patients with viruria developed nephropathy. The authors concluded that campth treatment does not permanently remove immune cells that are able to respond against BKV and that the therapy does not prevent stop the clearance of BKV from the blood.

Recent attention has focussed on the role of lytic antibody induction, and particularly the role of alemtuzumab (anti-CD52 monoclonal antibody; “Campath”) which is undergoing more recent widespread usage. A large study (“3C study”) demonstrated double the incidence of BKV infection with alemtuzumab compared with basiliximab induction. However, absolute incidence was low (8% *vs* 4%), and this difference was driven by BK viremia (7% *vs* 3%) rather than BKV nephropathy, the rates of which were very low in both alemtuzumab and basiliximab arms of the study (1% and 2% respectively)[43]. Conversely, it has been shown that alemtuzumab did not remain significant risk factor after the adjusted hazard ratio for each variable had been calculated[44].

A US OPTN database review showed that there was an increased risk of BKV infection with an induction therapy using thymoglobulin (*P* < 0.0001)[44]. In a study by Ott *et al*[45] renal transplants patients, under either basiliximab (*n* = 22) or thymoglobulin (*n* = 27) treatment regimens, were assessed for complication in a mean follow up period of three years. Of the 27 patients treated with thymoglobulin, two developed BKVAN whereas no patients had a BKV infection when treated with basiliximab. However, CMV infections were observed in both patient cohorts, with four and three patients infected for basiliximab and thymoglobulin therapies, respectively. This indicated that treatments using thymoglobulin carry a greater risk of BKV infection to renal transplant patients post transplant.

***Effect of HLA matching***

The adaptive immune response to viral infection is dependent on T-cell recognition of viral antigen presented in the context of self-MHC. In other transplant settings, it has been shown that immune responses to viral antigen presented in the context of donor-derived MHC (in this case cytomegalovirus) do not develop[46]. The donor-derived nature of BKV may therefore impair the magnitude or timing of effective immune clearance. In keeping with this concept, a study by Lee *et al*[47] used a mouse model to show ineffective clearance of BKV in the context of MHC mismatching. Clinical data also supports this concept of increased HLA mismatch as a risk factor for BKVAN[48-50]. However, in contrast, a clinical study from Drachenberg *et al*[51] found lesser degrees of HLA matching was actually associated with maintained graft function in patients with established BKVAN. This led to the proposal that even though reduction in HLA matching would decrease the recipient’s ability to mount an effective immune response to BKV, there would be less tissue damage, thus reduced risk of graft loss. This in turn raises questions in regard to the mechanism of viral clearance in this context, and whether this is dependent on the T-cell response or other viral elimination mechanisms such as NK and NKT cell activation. Clinical data from other cohorts may also serve to clarify the current understanding.

Aside from HLA matching, the question arises as to whether BKV infection and nephropathy is associated with specific (donor or recipient) HLA alleles. Awadalla *et al*[50] found no such association(s), but another study from Bohl *et al*[31] found a possible link between HLA C7 and the severity of BKV infection. Although there was no association between BK viremia and either donor or recipient HLA-A, -B or –DR type, all 11 transplant recipients with persistent BK viremia received kidneys from HLA C7 negative donors, and 10 of these 11 recipients also lacked HLA C7. The possible mechanism underlying this observation is unclear. However, if confirmed, this raises implications for the relevance of HLA-C typing in transplantation, which is not currently recommended or undertaken, but which may identify individuals at greater risk of refractory infection.

***Donor and recipient blood group***

A fundamental feature of transplantation is the matching of blood groups of donor and recipient in order to avoid the risk of hyper-acute rejection. Nevertheless, with intensified preconditioning and antibody removal, blood group-incompatible transplantation is now commonplace[52]. However, Sharif *et al*[53] suggest a high rate of BKVAN in such patients. In a study of 62 blood group incompatible transplantations between 1998 and 2010 the risk of BKVAN was 17.7% (compared to 3% risk among blood group compatible patients). This data has been replicated in a different cohort by Bentall *et al*[54]. While we may infer this is due to desensitisation and/or heightened immunosuppression for incompatible patients, the authors actually observed a lower risk among a cotemporaneous group of 221 HLA antibody incompatible transplants (5.9%, *P* = 0.008) who also underwent intensified preconditioning and received stronger induction therapy (ATG) compared to either blood group incompatible or compatible patients (Basiliximab). Therefore, pre-conditioning or heightened immunosuppression cannot be the sole explanation for this observation. Interestingly, the authors identified the lack of a typical accommodation-like phenotype (defined as C4d deposition in the absence of any micro-circulation inflammation) among blood-group incompatible transplant recipients with BKVAN compared to those without BKVAN (40.0% *vs* 75.8% respectively, *P* = 0.04). However, it is unclear from this data whether lack of accommodation-like phenotype development increases the risk for BKVAN or whether blood-group incompatible patients with BKVAN lose their accommodation-like phenotype but further studies are warranted to research this further.

**OTHER RISK FACTORS**

There are also a number of risk factors between the donor and recipient that can increase the risk of a BKV infection, including gender, race, age, diabetes mellitus or where the organ was sourced from a deceased donor[39,44,55 56].

**CLINICAL FEATURES AND DIAGNOSIS OF BKVAN**

The median time to clinically apparent BKVAN is within the first year after transplantation[57,58]. The recipient is characteristically asymptomatic, with the infection presenting as progressively worsening renal function, usually in the absence of significant or new-onset proteinuria[59]. This presentation generally results in a ‘for cause’ biopsy, which shows the characteristic features of BKVAN. A number of histological grading systems have attempted to classify BKVAN[60-63], and whilst differences exist between these alternative systems, recurring themes are:

The separation into stages of BKVAN depending on the presence of viral infection in the absence of inflammation or significant chronic damage (Grade A), with inflammation dominating over chronic damage (Grade B), and with chronic damage (fibrosis and tubular atrophy) as a notable component, with or without inflammation (Grade C).

That prognosis is correlated with these stages of BKVAN, and especially with the presence of significant chronic damage (Grade C nephropathy).

In simultaneous biopsy cores, discordant findings (*i.e*., the lack of evidence of BKVAN in one of the cores) was found in around a third of cases. Of note, in the core without evidence of virus, interstitial inflammation and/or acute tubular injury were frequent findings (approximately 80%), raising implications for the clinical interpretation of kidney biopsy specimens where only one core is retrieved, where the sampling is inadequate, and where there is collateral evidence that BKVAN may be a diagnosis[64].

Whilst concurrent viremia is almost universal with the finding of BKVAN on microscopy, the magnitude of circulating viral load seems to have little or no relationship with the extent of nephropathy[64]. Representative examples of the histological appearances of BKVAN are shown in Figures 2-4.

For many years, BKVAN was confused with acute rejection, as both have the appearance of an “interstitial nephritis”. With more widespread recognition of BKVAN, the availability of blood and urine testing for viral load, and the utility of SV40 staining on biopsy samples, the pathological diagnosis of BKVAN has become more straightforward. However, acute rejection and BKVN are not mutually exclusive, and particularly in the period following BKVAN treatment (see below), the two may coincide, and it may be unclear which represents the dominant process. Despite efforts, there remains no consensus in regard to the most accurate way to separate these entities, although the presence of macro- or micro-vascular inflammation points to rejection as a component at least.

Aside from the classical presentation described above, BKVAN may present in a “subclinical” manner, as is seen with other forms of transplant-related renal injury including “subclinical rejection”. Recent clinical studies have used protocol renal biopsies to test for the presence of BKV in this setting. Buehrig *et al*[65] concluded that allograft biopsy allowed earlier detection of BKVAN, and the potential for this to enable earlier treatment, although this proposed approach has not yet been evaluated. Whether such a strategy translates into improved overall clinical outcome (and justifies the risk, inconvenience and cost of the biopsy) remains to be seen.

**SCREENING FOR BKV INFECTION**

Established BKV screening methods include testing urine for decoy cells, viral particles by electron microscopy, and viral DNA by PCR. However, plasma PCR for detection of viremia remains a more common approach to screening[66]. It has also been suggested that circulating viral loads above certain thresholds (approximately > 4 log copies/mL) can be considered presumptive of nephropathy even in the absence of histological evidence (see above). Whilst inter-laboratory standardisation of such PCR assays is awaited, such discrete values remain subject to interpretation by individual centres.

A more recent study carried out by Singh *et al*[67] investigated whether the qualitative detection of three-dimensional aggregates of polyomavirus (Haufen crystals) within a patient’s urine could be used as a diagnostic test for patients BKVAN. Of 21 patients known to have BKVAN 77 of the 143 samples taken contained Haufen. During follow up, the presence or absence of Haufen matched the course of renal disease. All control samples (194) were negative. The predictive values of Haufen for BKVAN were 97% for positive and 100% for negative. This leads to the conclusion that Haufen testing in urine is a more accurate approach than detection of viral DNA in urine or plasma, although the reproducibility and generalisability of these findings requires further clarification.

Finally, the role of urine profiling for VP1 (BK capsid protein) mRNA is under investigation, with exploratory[68] and validation studies[69] suggesting potential utility in predicting nephropathy, albeit in small patient numbers Further generalisation may yet provide additional important information in this regard.

**CURRENT AND FUTURE TREATMENTS FOR BKV INFECTION**

The first step in the treatment of BKV infection is reduction in immunosuppression. Certainly, this approach is not disputed for cases of nephropathy, although evidence guiding the order with which the component immunosuppressants are withdrawn is lacking. Less clear is whether immunosuppression should be altered in the face of viremia and in the absence of overt nephropathy. This question was addressed in a large and important study of 200 patients by Brennan *et al*[40], where reduction in immunosuppression in response to detectable viremia on protocolised plasma samples resolved 95% cases of viremia with no signal towards graft rejection, dysfunction or loss. Other smaller studies also support this approach of “pre-emptive” therapy[70]. The efficacy of this strategy is also supported by Saad *et al*[71]. In this study, MMF and/or Tac doses were reduced for the patients, who in this study were not restricted to those with viremia (24 patients: 66% BKVN, 34% Viremia). Overall, a decline in BKV viral load was seen. However, three patients developed acute cellular rejection, albeit with successful treatment with intravenous bolus steroids. One patient experienced BKVN relapse during pregnancy and lost the graft. Seventeen patients maintained or improved their graft function following this reduction in immunosuppression. In summary, the evidence from these studies support decreasing immunosuppression as the first line of treatment for patients that present with BKVAN, and possibly with detectable viremia, although clearly the risk of rejection needs careful consideration. Controlled studies are required to solidify these findings, although in the meantime the recommendation from KDIGO (Kidney Disease: Improving Global Outcomes) expert panel is to reduce immunosuppression when plasma viral loads exceed a certain threshold (10000 copies per ml, whilst accounting for inter-laboratory variation)[72].

BKV interacts with the AKT/mammalian target of rapamycin (mTOR) pathway[73]. Everolimus and sirolimus are examples of mTOR inhibitors (mTOR-i)[73-76]. Everolimus was observed by Polanco *et al*[75] to increase renal function in BKVAN positive patients that had their treatments converted from tacrolimus to everolimus, with a suspension of mycophenolate. This study involved 15 patients, all presenting with BKVAN of which 9 underwent the immunosuppressant conversation. The serum creatinine of these patients decreased from 2 (± 0.21) mg/dL at the time of conversion to 1.6 (± 0.39) mg/dL at the final follow up. BK viremia became negative in 5 of the 9 patients and the remain 4 had a > 95% decrease in BKV. This decrease in BKVAN is also seen in conversion to sirolimus. In a recent single centre retrospective study by Tohme *et al*[77], patients were either placed on a tacrolimus or sirolimus based immunosuppression therapy. If the patients were < 62 years old they were converted from tacrolimus to sirolimus. Clinically significant BK viremia fell when converting from tacrolimus (*P* = 0.04) to sirolimus (*P* = 0.02), 17.9% to 4.3%, respectively. However, the hazard ratio for the male gender was also associated with the incidence of BK viremia (*P* = 0.03). Discontinuation of the sirolimus treatment occurred in 34% of patients due to various side effects. Thus, the use of mTOR-i as a treatment option of not only provides immunosuppression, reducing the risk of acute rejection, but also due to its behaviour as a metabolic pathway inhibitor for BKV it can also aid in the reduction in viral load, hence a lower risk of developing BKVAN.

The next question is whether detection of virus in urine (rather than waiting for it to appear in plasma) might represent a more efficient screening and intervention biomarker. In this regard, the clinical data is less optimistic. Specifically a series of retrospective studies have suggested increased rates of (or episodes of) acute rejection in the presence of viruria, even in the absence of viremia[78-80]. Whilst a proportion of these episodes were likely a response to immunosuppression weaning, there were clearly others which were unrelated, and which may potentially be a manifestation of low grade viral reactivation and inflammation inciting a secondary alloimmune response. However, irrespective of the mechanism, these observations (although limited by study design and interpretation) suggest that immunosuppression weaning in the context of viruria should not be recommended until and unless further information comes to light.

Even with successful treatment with immunosuppression reduction, the timeline of viral clearance is variable, although the reported median time to complete plasma clearance is 9 mo[81]. Serial renal histopathology following treatment is interesting, although reports are limited due to the nature of such studies. Of relevance though, the report from Menter *et al*[81] suggests that a self-limited “interstitial nephritis” is common during the phase of viral clearance and that this may represent an appropriate antiviral response rather than alloimmunity.

Specific antiviral therapy is generally used as a secondary line of treatment for BKVAN, and although an attractive approach, the role(s) of multiple agents remain unproven and unclear. Although better recognised as antibacterial agents, the quinolone antibiotics do display in vitro activity against polyoma viruses. Arroyo *et al*[82] retrospectively investigated the effects of ciprofloxacin on patients with BK viruria and viremia, after clinical failure with prior reduction in immunosuppression. The study showed that there were no adverse effects of ciprofloxacin and that out of the nine patients that received the treatment, three showed complete clearance of the virus and another three had the viral load in the plasma reduced by ≥ 50%. Unfortunately, a subsequent randomised controlled trial of 3 mo levofloxacin (from post-operative day 5) in 154 kidney transplant recipients showed no effect on the development of BKV viruria compared with the control group (29% *vs* 33%)[83]. In addition, an increased incidence of antibiotic resistance to bacterial isolates, and also a signal towards increased tendonitis was seen in the levofloxacin treated arm. Observational data also comes from Jung *et al*[84], this time studying the effect of leflunomide on biopsy-proven BKVN in paediatric patients. Tac dosage was reduced and leflunomide and intravenous immunoglobulin treatment was instituted. Viral load then decreased and remained below 100 copies/mL over an 18 mo period with no loss of renal function, from a value of 474140 copies/mL of BKV viral load in patient serum. Intravenous immunoglobulin in the absence of adjunctive antiviral agents has also been reported as a treatment for BKV infection[85], and observational data supports a potential role for another antiviral agent, cidofovir[86]. Yet in the absence of more robust data, few conclusions can be drawn; it is also relevant to highlight the conclusion of a 2010 systematic review, which found no evidence of an effect for either leflunomide or cidofovir in treating this infection[87]. Adoptive cell therapy in the context of transplant-associated infections is perhaps best known in the context of EBV and post-transplant lymphoproliferative disease. Whilst no data exists for this strategy in the setting of BKV infection, it is possible this approach might hold promise. In the context of infection with the related polyoma virus, JC virus, a report describes a positive clinical response to this form of therapy in a patient following hematopoietic cell transplantation[88]. Intuitively, the same approach may be worthwhile for BKV infection.

**CONCLUSION**

This review focusses on the pathogenesis, risk factors, presentation and treatment of BKV infection in the setting of kidney transplantation, which remains clearly the most common scenario in which this polyoma virus infection is encountered. Whilst important understanding has accumulated over recent years, and has certainly led to improved recognition of this infection and clinical management of patients, there is much more to be discovered and studied. We believe the most important tasks at hand are now to: (1) more accurately risk-stratify patients prior to (and also following) transplantation, with aim of individualising immunosuppression and reducing the risk of (or duration/consequences of) BKV infection. This may include developing understanding of, and then monitoring strategies for, cell-mediated immune responses to this virus, which can then be interpreted in combination with peripheral blood and renal biopsy measures of viral load and (admittedly currently unavailable) standardised assays of alloreactivity to garner a more “holistic” understanding of the overall and antigen-specific immunosuppressive burden; and (2) to enhance the sizeable observational experience of treatment strategies with controlled studies of immunosuppression weaning and/or adjunctive antiviral agents. It is not unconceivable that with such refined approaches BKV infection (whilst not eradicated) may present a far less sinister complication for kidney transplant patients in the future.

**REFERENCES**

1 **Gardner SD**, Field AM, Coleman DV, Hulme B. New human papovavirus (B.K.) isolated from urine after renal transplantation. *Lancet* 1971; **1**: 1253-1257 [PMID: 4104714 DOI: 10.1016/S0140-6736(71)91776-4]

2 **Shah KV**. Polyomaviruses. In: Fields Virology. Knipe DM, *et al*. 2nd ed. 1990: 1609-1623

3 **Butel JS**, Lednicky JA. Cell and molecular biology of simian virus 40: implications for human infections and disease. *J Natl Cancer Inst* 1999; **91**: 119-134 [PMID: 9923853 DOI: 10.1093/jnci/91.2.119]

4 **Li RM**, Mannon RB, Kleiner D, Tsokos M, Bynum M, Kirk AD, Kopp JB. BK virus and SV40 co-infection in polyomavirus nephropathy. *Transplantation* 2002; **74**: 1497-1504 [PMID: 12490781 DOI: 10.1097/01.TP.0000042485.92082.5B]

5 **Shah KV**. Human polyomavirus BKV and renal disease. *Nephrol Dial Transplant* 2000; **15**: 754-755 [PMID: 10831621 DOI: 10.1093/ndt/15.6.754]

6 **Bohl DL**, Brennan DC. BK virus nephropathy and kidney transplantation. *Clin J Am Soc Nephrol* 2007; **2** Suppl 1: S36-S46 [PMID: 17699509 DOI: 10.2215/CJN.00920207]

7 **Imperiale MJ**, Major EO. Polyomaviruses. In: Fields Virology. Knipe DM, *et al*. 5th ed. 2007: 2175-2195

8 **Egli A**, Infanti L, Dumoulin A, Buser A, Samaridis J, Stebler C, Gosert R, Hirsch HH. Prevalence of polyomavirus BK and JC infection and replication in 400 healthy blood donors. *J Infect Dis* 2009; **199**: 837-846 [PMID: 19434930 DOI: 10.1086/597126]

9 **Heritage J**, Chesters PM, McCance DJ. The persistence of papovavirus BK DNA sequences in normal human renal tissue. *J Med Virol* 1981; **8**: 143-150 [PMID: 6271922 DOI: 10.1002/jmv.1890080208]

10 **Sawinski D**, Goral S. BK virus infection: an update on diagnosis and treatment. *Nephrol Dial Transplant* 2015; **30**: 209-217 [PMID: 24574543 DOI: 10.1093/ndt/gfu023]

11 **El-Zoghby ZM**, Stegall MD, Lager DJ, Kremers WK, Amer H, Gloor JM, Cosio FG. Identifying specific causes of kidney allograft loss. *Am J Transplant* 2009; **9**: 527-535 [PMID: 19191769 DOI: 10.1111/j.1600-6143.2008.02519.x]

12 **Chand S**, Atkinson D, Collins C, Briggs D, Ball S, Sharif A, Skordilis K, Vydianath B, Neil D, Borrows R. The Spectrum of Renal Allograft Failure. *PLoS One* 2016; **11**: e0162278 [PMID: 27649571 DOI: 10.1371/journal.pone.0162278]

13 **Hirsch HH**, Steiger J. Polyomavirus BK. *Lancet Infect Dis* 2003; **3**: 611-623 [PMID: 14522260 DOI: 10.1016/S1473-3099(03)00770-9]

14 **Hariharan S**. BK virus nephritis after renal transplantation. *Kidney Int* 2006; **69**: 655-662 [PMID: 16395271 DOI: 10.1038/sj.ki.5000040]

15 **Garces JC**. BK Virus-Associated Nephropathy in Kidney Transplant Recipients. *Ochsner J* 2010; **10**: 245-249 [PMID: 21603392]

16 **Henriksen S**, Mittelholzer C, Gosert R, Hirsch HH, Rinaldo CH. Human BK Polyomavirus Plasmid pBKV (34-2) (Dunlop) Contains Mutations Not Found in the Originally Published Sequences. *Genome Announc* 2015; **3**: pii: e00046-15 [PMID: 25814590 DOI: 10.1128/genomeA.00046-15]

17 **Ranjan R**, Rani A, Brennan DC, Finn PW, Perkins DL. Complete Genome Sequence of BK Polyomavirus Subtype Ib-1 Detected in a Kidney Transplant Patient with BK Viremia Using Shotgun Sequencing. *Genome Announc* 2017; **5**: pii: e01474-16 [PMID: 28183780 DOI: 10.1128/genomeA.01474-16]

18 **Leuenberger D**, Andresen PA, Gosert R, Binggeli S, Ström EH, Bodaghi S, Rinaldo CH, Hirsch HH. Human polyomavirus type 1 (BK virus) agnoprotein is abundantly expressed but immunologically ignored. *Clin Vaccine Immunol* 2007; **14**: 959-968 [PMID: 17538118 DOI: 10.1128/CVI.00123-07]

19 **Barbanti-Brodano G**, Sabbioni S, Martini F, Negrini M, Corallini A, Tognon M. BK virus, JC virus and Simian Virus 40 infection in humans, and association with human tumors. *Adv Exp Med Biol* 2006; **577**: 319-341 [PMID: 16626046 DOI: 10.1007/0-387-32957-9\_23]

20 **Anzivino E**, Bellizzi A, Mitterhofer AP, Tinti F, Barile M, Colosimo MT, Fioriti D, Mischitelli M, Chiarini F, Ferretti G, Taliani G, Pietropaolo V. Early monitoring of the human polyomavirus BK replication and sequencing analysis in a cohort of adult kidney transplant patients treated with basiliximab. *Virol J* 2011; **8**: 407 [PMID: 21849069 DOI: 10.1186/1743-422X-8-407]

21 **Mischitelli M**, Bellizzi A, Anzivino E, Fioriti D, Boldorini R, Miglio U, Chiarini F, Di Monaco F, Pietropaolo V. Complications post renal transplantation: literature focus on BK virus nephropathy and diagnostic tools actually available. *Virol J* 2008; **5**: 38 [PMID: 18315864 DOI: 10.1186/1743-422X-5-38]

22 **Dugan AS**, Eash S, Atwood WJ. An N-linked glycoprotein with alpha(2,3)-linked sialic acid is a receptor for BK virus. *J Virol* 2005; **79**: 14442-14445 [PMID: 16254379 DOI: 10.1128/JVI.79.22.14442-14445.2005]

23 **Low JA**, Magnuson B, Tsai B, Imperiale MJ. Identification of gangliosides GD1b and GT1b as receptors for BK virus. *J Virol* 2006; **80**: 1361-1366 [PMID: 16415013 DOI: 10.1128/JVI.80.3.1361-1366.2006]

24 **Moriyama T**, Marquez JP, Wakatsuki T, Sorokin A. Caveolar endocytosis is critical for BK virus infection of human renal proximal tubular epithelial cells. *J Virol* 2007; **81**: 8552-8562 [PMID: 17553887 DOI: 10.1128/JVI.00924-07]

25 **Eash S**, Querbes W, Atwood WJ. Infection of vero cells by BK virus is dependent on caveolae. *J Virol* 2004; **78**: 11583-11590 [PMID: 15479799 DOI: 10.1128/JVI.78.21.11583-11590.2004]

26 **Jin L**, Gibson PE, Knowles WA, Clewley JP. BK virus antigenic variants: sequence analysis within the capsid VP1 epitope. *J Med Virol* 1993; **39**: 50-56 [PMID: 7678637 DOI: 10.1002/jmv.1890390110]

27 **Knowles WA**, Gibson PE, Gardner SD. Serological typing scheme for BK-like isolates of human polyomavirus. *J Med Virol* 1989; **28**: 118-123 [PMID: 2544676 DOI: 10.1002/jmv.1890280212]

28 **Takasaka T**, Goya N, Tokumoto T, Tanabe K, Toma H, Ogawa Y, Hokama S, Momose A, Funyu T, Fujioka T, Omori S, Akiyama H, Chen Q, Zheng HY, Ohta N, Kitamura T, Yogo Y. Subtypes of BK virus prevalent in Japan and variation in their transcriptional control region. *J Gen Virol* 2004; **85**: 2821-2827 [PMID: 15448343 DOI: 10.1099/vir.0.80363-0]

29 **Sharma PM**, Gupta G, Vats A, Shapiro R, Randhawa P. Phylogenetic analysis of polyomavirus BK sequences. *J Virol* 2006; **80**: 8869-8879 [PMID: 16940499 DOI: 10.1128/JVI.00510-06]

30 **Andrews CA**, Shah KV, Daniel RW, Hirsch MS, Rubin RH. A serological investigation of BK virus and JC virus infections in recipients of renal allografts. *J Infect Dis* 1988; **158**: 176-181 [PMID: 2839580 DOI: 10.1093/infdis/158.1.176]

31 **Bohl DL**, Storch GA, Ryschkewitsch C, Gaudreault-Keener M, Schnitzler MA, Major EO, Brennan DC. Donor origin of BK virus in renal transplantation and role of HLA C7 in susceptibility to sustained BK viremia. *Am J Transplant* 2005; **5**: 2213-2221 [PMID: 16095500 DOI: 10.1111/j.1600-6143.2005.01000.x]

32 **Smith JM**, McDonald RA, Finn LS, Healey PJ, Davis CL, Limaye AP. Polyomavirus nephropathy in pediatric kidney transplant recipients. *Am J Transplant* 2004; **4**: 2109-2117 [PMID: 15575916 DOI: 10.1111/j.1600-6143.2004.00629.x]

33 **Saundh BK**, Baker R, Harris M, Welberry Smith MP, Cherukuri A, Hale A. Early BK polyomavirus (BKV) reactivation in donor kidney is a risk factor for development of BKV-associated nephropathy. *J Infect Dis* 2013; **207**: 137-141 [PMID: 23100561 DOI: 10.1093/infdis/ji642]

34 **Beimler J**, Sommerer C, Zeier M. The influence of immunosuppression on the development of BK virus nephropathy-- does it matter? *Nephrol Dial Transplant* 2007; **22** Suppl 8: viii66-viii71 [PMID: 17890267 DOI: 10.1093/ndt/gfm646]

35 **Comoli P**, Cioni M, Basso S, Gagliardone C, Potenza L, Verrina E, Luppi M, Zecca M, Ghiggeri GM, Ginevri F. Immunity to Polyomavirus BK Infection: Immune Monitoring to Regulate the Balance between Risk of BKV Nephropathy and Induction of Alloimmunity. *Clin Dev Immunol* 2013; **2013**: 256923 [PMID: 24000288 DOI: 10.1155/2013/256923]

36 **Comoli P**, Azzi A, Maccario R, Basso S, Botti G, Basile G, Fontana I, Labirio M, Cometa A, Poli F, Perfumo F, Locatelli F, Ginevri F. Polyomavirus BK-specific immunity after kidney transplantation. *Transplantation* 2004; **78**: 1229-1232 [PMID: 15502726 DOI: 10.1097/01.TP.0000137932.44791.D3]

37 **Kuppachi S**, Kaur D, Holanda DG, Thomas CP. BK polyoma virus infection and renal disease in non-renal solid organ transplantation. *Clin Kidney J* 2016; **9**: 310-318 [PMID: 26985385 DOI: 10.1093/ckj/sfv143]

38 **Jacobi J**, Prignitz A, Büttner M, Korn K, Weidemann A, Hilgers KF, Heller K, Velden J, Knöll A, Wullich B, May C, Eckardt KU, Amann KU. BK viremia and polyomavirus nephropathy in 352 kidney transplants; risk factors and potential role of mTOR inhibition. *BMC Nephrol* 2013; **14**: 207 [PMID: 24088187 DOI: 10.1186/1471-2369-14-207]

39 **Mengel M**, Marwedel M, Radermacher J, Eden G, Schwarz A, Haller H, Kreipe H. Incidence of polyomavirus-nephropathy in renal allografts: influence of modern immunosuppressive drugs. *Nephrol Dial Transplant* 2003; **18**: 1190-1196 [PMID: 12748354 DOI: 10.1093/ndt/gfg072]

40 **Brennan DC**, Agha I, Bohl DL, Schnitzler MA, Hardinger KL, Lockwood M, Torrence S, Schuessler R, Roby T, Gaudreault-Keener M, Storch GA. Incidence of BK with tacrolimus versus cyclosporine and impact of preemptive immunosuppression reduction. *Am J Transplant* 2005; **5**: 582-594 [PMID: 15707414 DOI: 10.1111/j.1600-6143.2005.00742.x]

41 **Hirsch HH**, Vincenti F, Friman S, Tuncer M, Citterio F, Wiecek A, Scheuermann EH, Klinger M, Russ G, Pescovitz MD, Prestele H. Polyomavirus BK replication in de novo kidney transplant patients receiving tacrolimus or cyclosporine: a prospective, randomized, multicenter study. *Am J Transplant* 2013; **13**: 136-145 [PMID: 23137180 DOI: 10.1111/j.1600-6143.2012.04320.x]

42 **Kayler LK**, Mohanka R, Morgan C, Basu A, Shapiro R, Randhawa PS. Clinical course of kidney transplant patients with acute rejection and BK virus replication following Campath therapy. *Clin Transplant* 2008; **22**: 348-353 [PMID: 18279421 DOI: 10.1111/j.1399-0012.2008.00791.x]

43 **3C Study Collaborative Group**, Haynes R, Harden P, Judge P, Blackwell L, Emberson J, Landray MJ, Baigent C, Friend PJ. Alemtuzumab-based induction treatment versus basiliximab-based induction treatment in kidney transplantation (the 3C Study): a randomised trial. *Lancet* 2014; **384**: 1684-1690 [PMID: 25078310 DOI: 10.1016/S0140-6736(14)61095-3]

44 **Dharnidharka VR**, Cherikh WS, Abbott KC. An OPTN analysis of national registry data on treatment of BK virus allograft nephropathy in the United States. *Transplantation* 2009; **87**: 1019-1026 [PMID: 19352121 DOI: 10.1097/TP.0b013e31819cc383]

45 **Ott U**, Busch M, Steiner T, Schubert J, Wolf G. Renal retransplantation: a retrospective monocentric study. *Transplant Proc* 2008; **40**: 1345-1348 [PMID: 18589102 DOI: 10.1016/j.transproceed.2008.01.068]

46 **Egli A**, Binggeli S, Bodaghi S, Dumoulin A, Funk GA, Khanna N, Leuenberger D, Gosert R, Hirsch HH. Cytomegalovirus and polyomavirus BK posttransplant. *Nephrol Dial Transplant* 2007; **22** Suppl 8: viii72-viii82 [PMID: 17890268 DOI: 10.1093/ndt/gfm648]

47 **Lee EDH**, Wang J, Dong Y, Lukacher AE, Newell KA, Pearson TC, Larsen CP. Inability to clear polyoma virus in MHC disparate murine kidney transplant model results in accelerated graft rejection [Abstract]. *Am J Kidney Dis* 2004; **4**: 599

48 **Hirsch HH**, Knowles W, Dickenmann M, Passweg J, Klimkait T, Mihatsch MJ, Steiger J. Prospective study of polyomavirus type BK replication and nephropathy in renal-transplant recipients. *N Engl J Med* 2002; **347**: 488-496 [PMID: 12181403 DOI: 10.1056/NEJMoa020439]

49 **Masutani K**, Ninomiya T, Randhawa P. HLA-A2, HLA-B44 and HLA-DR15 are associated with lower risk of BK viremia. *Nephrol Dial Transplant* 2013; **28**: 3119-3126 [PMID: 24084328 DOI: 10.1093/ndt/gft298]

50 **Awadalla Y**, Randhawa P, Ruppert K, Zeevi A, Duquesnoy RJ. HLA mismatching increases the risk of BK virus nephropathy in renal transplant recipients. *Am J Transplant* 2004; **4**: 1691-1696 [PMID: 15367226 DOI: 10.1111/j.1600-6143.2004.00563.x]

51 **Drachenberg CB**, Papadimitriou JC, Mann D, Hirsch HH, Wali R, Ramos E. Negative impact of human leukocyte antigen matching in the outcome of polyomavirus nephropathy. *Transplantation* 2005; **80**: 276-278 [PMID: 16041275 DOI: 10.1097/01.TP.0000165096.01034.15]

52 **Muramatsu M**, Gonzalez HD, Cacciola R, Aikawa A, Yaqoob MM, Puliatti C. ABO incompatible renal transplants: Good or bad? *World J Transplant* 2014; **4**: 18-29 [PMID: 24669364 DOI: 10.5500/wjt.v4.i1.18]

53 **Sharif A**, Alachkar N, Bagnasco S, Geetha D, Gupta G, Womer K, Arend L, Racusen L, Montgomery R, Kraus E. Incidence and outcomes of BK virus allograft nephropathy among ABO- and HLA-incompatible kidney transplant recipients. *Clin J Am Soc Nephrol* 2012; **7**: 1320-1327 [PMID: 22626962 DOI: 10.2215/CJN.00770112]

54 **Bentall A**, Neil D, Sharif A, Ball S. ABO-incompatible kidney transplantation is a novel risk factor for BK nephropathy. *Transplantation* 2015; **99**: e8-e9 [PMID: 25651124 DOI: 10.1097/TP.0000000000000483]

55 **Merlino C**, Bergallo M, Gribaudo G, Gregori G, Paolo Segoloni G, Giacchino F, Ponzi AN, Cavallo R. Polyomavirus BK DNA quantification assay to evaluate viral load in renal transplant recipients. *J Clin Virol* 2003; **28**: 265-274 [PMID: 14522065 DOI: 10.1016/S1386-6532(03)00012-X]

56 **Vasudev B**, Hariharan S, Hussain SA, Zhu YR, Bresnahan BA, Cohen EP. BK virus nephritis: risk factors, timing, and outcome in renal transplant recipients. *Kidney Int* 2005; **68**: 1834-1839 [PMID: 16164661 DOI: 10.1111/j.1523-1755.2005.00602.x]

57 **Nickeleit V**, Klimkait T, Binet IF, Dalquen P, Del Zenero V, Thiel G, Mihatsch MJ, Hirsch HH. Testing for polyomavirus type BK DNA in plasma to identify renal-allograft recipients with viral nephropathy. *N Engl J Med* 2000; **342**: 1309-1315 [PMID: 10793163 DOI: 10.1056/NEJM200005043421802]

58 **Drachenberg CB**, Beskow CO, Cangro CB, Bourquin PM, Simsir A, Fink J, Weir MR, Klassen DK, Bartlett ST, Papadimitriou JC. Human polyoma virus in renal allograft biopsies: morphological findings and correlation with urine cytology. *Hum Pathol* 1999; **30**: 970-977 [PMID: 10452511 DOI: 10.1016/S0046-8177(99)90252-6]

59 **Krishna A**, Prasad N. BK virus nephropathy in renal transplantation. *Ind J Transplant* 2011; **5**: 182-190 [DOI: 10.1016/S2212-0017(11)60038-5]

60 **Drachenberg RC**, Drachenberg CB, Papadimitriou JC, Ramos E, Fink JC, Wali R, Weir MR, Cangro CB, Klassen DK, Khaled A, Cunningham R, Bartlett ST. Morphological spectrum of polyoma virus disease in renal allografts: diagnostic accuracy of urine cytology. *Am J Transplant* 2001; **1**: 373-381 [PMID: 12099383 DOI: 10.1034/j.1600-6143.2001.10414.x]

61 **Sar A**, Worawichawong S, Benediktsson H, Zhang J, Yilmaz S, Trpkov K. Interobserver agreement for Polyomavirus nephropathy grading in renal allografts using the working proposal from the 10th Banff Conference on Allograft Pathology. *Hum Pathol* 2011; **42**: 2018-2024 [PMID: 21733554 DOI: 10.1016/j.humpath.2011.03.008]

62 **Masutani K**, Shapiro R, Basu A, Tan H, Wijkstrom M, Randhawa P. The Banff 2009 Working Proposal for polyomavirus nephropathy: a critical evaluation of its utility as a determinant of clinical outcome. *Am J Transplant* 2012; **12**: 907-918 [PMID: 22390378 DOI: 10.1111/j.1600-6143.2012.03993.x]

63 **Hirsch HH**, Randhawa P; AST Infectious Diseases Community of Practice. BK polyomavirus in solid organ transplantation. *Am J Transplant* 2013; **13** Suppl 4: 179-188 [PMID: 23465010 DOI: 10.1111/ajt.12110]

64 **Drachenberg CB**, Papadimitriou JC, Hirsch HH, Wali R, Crowder C, Nogueira J, Cangro CB, Mendley S, Mian A, Ramos E. Histological patterns of polyomavirus nephropathy: correlation with graft outcome and viral load. *Am J Transplant* 2004; **4**: 2082-2092 [PMID: 15575913 DOI: 10.1046/j.1600-6143.2004.00603.x]

65 **Buehrig CK**, Lager DJ, Stegall MD, Kreps MA, Kremers WK, Gloor JM, Schwab TR, Velosa JA, Fidler ME, Larson TS, Griffin MD. Influence of surveillance renal allograft biopsy on diagnosis and prognosis of polyomavirus-associated nephropathy. *Kidney Int* 2003; **64**: 665-673 [PMID: 12846764 DOI: 10.1046/j.1523-1755.2003.00103.x]

66 **Boan P**, Hewison C, Swaminathan R, Irish A, Warr K, Sinniah R, Pryce TM, Flexman J. Optimal use of plasma and urine BK viral loads for screening and predicting BK nephropathy. *BMC Infect Dis* 2016; **16**: 342 [PMID: 27448566 DOI: 10.1186/s12879-016-1652-6]

67 **Singh HK**, Andreoni KA, Madden V, True K, Detwiler R, Weck K, Nickeleit V. Presence of urinary Haufen accurately predicts polyomavirus nephropathy. *J Am Soc Nephrol* 2009; **20**: 416-427 [PMID: 19158358 DOI: 10.1681/ASN.2008010117]

68 **Ding R**, Medeiros M, Dadhania D, Muthukumar T, Kracker D, Kong JM, Epstein SR, Sharma VK, Seshan SV, Li B, Suthanthiran M. Noninvasive diagnosis of BK virus nephritis by measurement of messenger RNA for BK virus VP1 in urine. *Transplantation* 2002; **74**: 987-994 [PMID: 12394843 DOI: 10.1097/01.TP.0000032151.07990.1E]

69 **Dadhania D**, Snopkowski C, Ding R, Muthukumar T, Lee J, Bang H, Sharma VK, Seshan S, August P, Kapur S, Suthanthiran M. Validation of noninvasive diagnosis of BK virus nephropathy and identification of prognostic biomarkers. *Transplantation* 2010; **90**: 189-197 [PMID: 20526237 DOI: 10.1097/TP.0b013e3181e2a932]

70 **Hussain S**, Orentas R, Walczak J. Prevention of BKV nephritis by monitoring BK viremia in renal transplant recipients: a prospective study. *Graft* 2004; **7**: 28-30

71 **Saad ER**, Bresnahan BA, Cohen EP, Lu N, Orentas RJ, Vasudev B, Hariharan S. Successful treatment of BK viremia using reduction in immunosuppression without antiviral therapy. *Transplantation* 2008; **85**: 850-854 [PMID: 18360267 DOI: 10.1097/TP.0b013e318166cba8]

72 **Eckardt KU**, Kasiske BL. Kidney disease: improving global outcomes. *Nat Rev Nephrol* 2009; **5**: 650-657 [PMID: 19786993 DOI: 10.1038/nrneph.2009.153]

73 **Liacini A**, Seamone ME, Muruve DA, Tibbles LA. Anti-BK virus mechanisms of sirolimus and leflunomide alone and in combination: toward a new therapy for BK virus infection. *Transplantation* 2010; **90**: 1450-1457 [PMID: 21079551 DOI: 10.1097/TP.0b013e3182007be2]

74 **Morath C**, Arns W, Schwenger V, Mehrabi A, Fonouni H, Schmidt J, Zeier M. Sirolimus in renal transplantation. *Nephrol Dial Transplant* 2007; **22** Suppl 8: viii61-viii65 [PMID: 17890266]

75 **Polanco N**, González Monte E, Folgueira MD, Morales E, Gutiérrez Martínez E, Bengoa I, Hernández A, Morales JM, Praga M, Andrés A. Everolimus-based immunosuppression therapy for BK virus nephropathy. *Transplant Proc* 2015; **47**: 57-61 [PMID: 25645770 DOI: 10.1016/j.transproceed.2014.11.008]

76 **Jouve T**, Rostaing L, Malvezzi P. Place of mTOR inhibitors in management of BKV infection after kidney transplantation. *J Nephropathol* 2016; **5**: 1-7 [PMID: 27047803 DOI: 10.15171/jnp.2016.01]

77 **Tohme FA**, Kalil RS, Thomas CP. Conversion to a sirolimus-based regimen is associated with lower incidence of BK viremia in low-risk kidney transplant recipients. *Transpl Infect Dis* 2015; **17**: 66-72 [PMID: 25582442 DOI: 10.1111/tid.12347]

78 **Masutani K**, Shapiro R, Basu A, Tan H, Ninomiya T, Randhawa P. Putative episodes of T-cell-mediated rejection in patients with sustained BK viruria but no viremia. *Transplantation* 2012; **94**: 43-49 [PMID: 22691957 DOI: 10.1097/TP.0b013e318253e7a4]

79 **Babel N**, Fendt J, Karaivanov S, Bold G, Arnold S, Sefrin A, Lieske E, Hoffzimmer M, Dziubianau M, Bethke N, Meisel C, Grütz G, Reinke P. Sustained BK viruria as an early marker for the development of BKV-associated nephropathy: analysis of 4128 urine and serum samples. *Transplantation* 2009; **88**: 89-95 [PMID: 19584686 DOI: 10.1097/TP.0b013e3181aa8f62]

80 **Hardinger KL**, Koch MJ, Bohl DJ, Storch GA, Brennan DC. BK-virus and the impact of pre-emptive immunosuppression reduction: 5-year results. *Am J Transplant* 2010; **10**: 407-415 [PMID: 20055811 DOI: 10.1111/j.1600-6143.2009.02952.x]

81 **Menter T**, Mayr M, Schaub S, Mihatsch MJ, Hirsch HH, Hopfer H. Pathology of resolving polyomavirus-associated nephropathy. *Am J Transplant* 2013; **13**: 1474-1483 [PMID: 23721552 DOI: 10.1111/ajt.12218]

82 **Arroyo D**, Chandran S, Vagefi PA, Wojciechowski D. Adjuvant Ciprofloxacin for Persistent BK Polyomavirus Infection in Kidney Transplant Recipients. *J Transplant* 2014; **2014**: 107459 [PMID: 25349720 DOI: 10.1155/2014/107459]

83 **Knoll GA**, Humar A, Fergusson D, Johnston O, House AA, Kim SJ, Ramsay T, Chassé M, Pang X, Zaltzman J, Cockfield S, Cantarovich M, Karpinski M, Lebel L, Gill JS. Levofloxacin for BK virus prophylaxis following kidney transplantation: a randomized clinical trial. *JAMA* 2014; **312**: 2106-2114 [PMID: 25399012 DOI: 10.1001/jama.2014.14721]

84 **Jung YH**, Moon KC, Ha JW, Kim SJ, Ha IS, Cheong HI, Kang HG. Leflunomide therapy for BK virus allograft nephropathy after pediatric kidney transplantation. *Pediatr Transplant* 2013; **17**: E50-E54 [PMID: 23210794 DOI: 10.1111/petr.12029]

85 **Shah T**, Vu D, Naraghi R, Campbell A, Min D. Efficacy of Intravenous Immunoglobulin in the Treatment of Persistent BK Viremia and BK Virus Nephropathy in Renal Transplant Recipients. *Clin Transpl* 2014: 109-116 [PMID: 26281134]

86 **Kuypers DR**, Vandooren AK, Lerut E, Evenepoel P, Claes K, Snoeck R, Naesens L, Vanrenterghem Y. Adjuvant low-dose cidofovir therapy for BK polyomavirus interstitial nephritis in renal transplant recipients. *Am J Transplant* 2005; **5**: 1997-2004 [PMID: 15996251 DOI: 10.1111/j.1600-6143.2005.00980.x]

87 **Johnston O**, Jaswal D, Gill JS, Doucette S, Fergusson DA, Knoll GA. Treatment of polyomavirus infection in kidney transplant recipients: a systematic review. *Transplantation* 2010; **89**: 1057-1070 [PMID: 20090569 DOI: 10.1097/TP.0b013e3181d0e15e]

88 **Balduzzi A**, Lucchini G, Hirsch HH, Basso S, Cioni M, Rovelli A, Zincone A, Grimaldi M, Corti P, Bonanomi S, Biondi A, Locatelli F, Biagi E, Comoli P. Polyomavirus JC-targeted T-cell therapy for progressive multiple leukoencephalopathy in a hematopoietic cell transplantation recipient. *Bone Marrow Transplant* 2011; **46**: 987-992 [PMID: 20921942 DOI: 10.1038/bmt.2010.221]

**P-Reviewer:** Esposito P, Friedman EA, Moens U **S-Editor:** Ji FF **L-Editor: E-Editor:**

**Specialty type:** Transplantation

**Country of origin:** United Kingdom

**Peer-review report classification**

Grade A (Excellent): 0

Grade B (Very good): B

Grade C (Good): C, C

Grade D (Fair): 0

Grade E (Poor): 0



**Figure 1 Proposed mechanisms for the pathogenesis of BK virus-associated nephritis after BK virus infection has occurred resulting in BK viruria or BK viremia.** These mechanisms include immunological factors, such as alterations to immunosuppressive therapy and cellular and humoral immunity, the source of BKV, either from the recipient or the donor, HLA matching, donor and recipient blood group. The two main treatment options for BKVN are a reduction in immunosuppression and the use of antiviral therapies. These treatments can also be used for BK viruria and viremia in order to prevent progression to BKVAN. BKV: BK virus; BKVAN: BK virus-associated nephritis.

A B C

  

**Figure 2 Histological features of BK virus nephropathy by light microscopy.** A: Tubule-interstitial infiltrate and tubulitis classical for BKVN, but also compatible with any other form of interstitial nephritis such as acute cellular rejection; B: Higher power view of same biopsy sample, with characteristic viral inclusions seen within epithelial cells (circled); C: Positive SV40 immunoperoxidase staining on same specimen, confirming diagnosis of BKVN. BKVN: BK virus nephropathy.



**Figure 3 Kidney with preserved tubular architecture, without significant chronic damage or interstitial inflammation, but with BK virus nephropathy confirmed by virtue of positive SV40 staining (as shown in insert taken from immunoperoxidase sample from same biopsy specimen).**

  

**Figure 4 Histological features of BK virus nephropathy by electron microscopy.** A: Electron microscopy evidence of viral inclusions (arrow) within epithelial cells, equivalent to those seen and circled in the light microscopy sample shown in Figure 2B; B: Higher power magnification of epithelial viral inclusions; C: Highest magnification demonstrating characteristic appearance and size (labelled) of BK virions.