

World Journal of *Methodology*

World J Methodol 2017 June 26; 7(2): 33-72





FIELD OF VISION

- 33 Synergetic role of integrating the departments of cancer registry and clinical research at an academic comprehensive cancer center

Bedra M, Vyskocil T, Emel J, Edwards C, Boutros C

REVIEW

- 37 Antioxidants in experimental ischemia-reperfusion injury of the testis: Where are we heading towards?

Vaos G, Zavras N

- 46 Role of metabolic stress for enhancing muscle adaptations: Practical applications

de Freitas MC, Gerosa-Neto J, Zanchi NE, Lira FS, Rossi FE

- 55 Targeted temperature management in neurological intensive care unit

Muengtaweepongsa S, Srivilaithon W

ORIGINAL ARTICLE

Basic Study

- 68 Nutech functional score: A novel scoring system to assess spinal cord injury patients

Shroff G, Barthakur JK

ABOUT COVER

Editorial Board Member of *World Journal of Methodology*, Wei Liu, PhD, Assistant Professor, Department of Radiation Physics, the University of Texas MD Anderson Cancer Center, Houston, TX 77030, United States

AIM AND SCOPE

World Journal of Methodology (*World J Methodol*, *WJM*, online ISSN 2222-0682, DOI: 10.5662) is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

The primary task of *WJM* is to rapidly publish high-quality original articles, reviews, and commentaries that deal with the methodology to develop, validate, modify and promote diagnostic and therapeutic modalities and techniques in preclinical and clinical applications. *WJM* covers topics concerning the subspecialties including but not exclusively anesthesiology, cardiac medicine, clinical genetics, clinical neurology, critical care, dentistry, dermatology, emergency medicine, endocrinology, family medicine, gastroenterology and hepatology, geriatrics and gerontology, hematology, immunology, infectious diseases, internal medicine, obstetrics and gynecology, oncology, ophthalmology, orthopedics, otolaryngology, radiology, serology, pathology, pediatrics, peripheral vascular disease, psychiatry, radiology, rehabilitation, respiratory medicine, rheumatology, surgery, toxicology, transplantation, and urology and nephrology.

INDEXING/ABSTRACTING

World Journal of Methodology is now indexed in PubMed, PubMed Central.

FLYLEAF

I-V Editorial Board

EDITORS FOR THIS ISSUE

Responsible Assistant Editor: *Xiang Li*
Responsible Electronic Editor: *Huan-Liang Wu*
Proofing Editor-in-Chief: *Lian-Sheng Ma*

Responsible Science Editor: *Fang-Fang Ji*
Proofing Editorial Office Director: *Ze-Mao Gong*

NAME OF JOURNAL
World Journal of Methodology

ISSN
ISSN 2222-0682 (online)

LAUNCH DATE
September 26, 2011

FREQUENCY
Quarterly

EDITOR-IN-CHIEF
Yicheng Ni, MD, PhD, Professor, Department of Radiology, University Hospitals, KU, Leuven, Herestraat 49, B-3000, Leuven, Belgium

EDITORIAL BOARD MEMBERS
All editorial board members resources online at <http://www.wjgnet.com/2222-0682/editorialboard.htm>

EDITORIAL OFFICE
Xiu-Xia Song, Director
World Journal of Methodology
Baishideng Publishing Group Inc
7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
Telephone: +1-925-2238242
Fax: +1-925-2238243
E-mail: editorialoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>

PUBLISHER
Baishideng Publishing Group Inc
7901 Stoneridge Drive,
Suite 501, Pleasanton, CA 94588, USA
Telephone: +1-925-2238242
Fax: +1-925-2238243
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>

PUBLICATION DATE
June 26, 2017

COPYRIGHT
© 2016 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT
All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORS
<http://www.wjgnet.com/bpg/gerinfo/204>

ONLINE SUBMISSION
<http://www.f6publishing.com>

Antioxidants in experimental ischemia-reperfusion injury of the testis: Where are we heading towards?

George Vaos, Nick Zavras

George Vaos, Nick Zavras, Department of Paediatric Surgery, "ATTIKON" University General Hospital, National and Kapodistrian University of Athens, School of Medicine, 12462 Haidari, Athens, Greece

Author contributions: All authors equally contributed to this paper with conception and design of the study, literature review and analysis, drafting and critical revision and editing, and final approval of the final version.

Conflict-of-interest statement: No potential conflicts of interest. No financial support.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: George Vaos, MD, PhD, FEBPS, Professor, Department of Paediatric Surgery, "ATTIKON" University General Hospital, National and Kapodistrian University of Athens, School of Medicine, 1 Rimini Street, 12462 Haidari, Athens, Greece. gvaos@med.uoa.gr
Telephone: +30-210-5831299
Fax: +30-210-5326411

Received: January 28, 2017

Peer-review started: February 8, 2017

First decision: March 28, 2017

Revised: April 21, 2017

Accepted: May 12, 2017

Article in press: May 15, 2017

Published online: June 26, 2017

Abstract

Testicular torsion (TT) is a medical emergency that

primary affects newborns and young adolescents. It causes testicular injury due to the torsion of the spermatic cord and its components, initially in the venous blood flow and finally in the arterial blood flow. Prompt diagnosis and early surgical management are necessary in managing this urgent situation. The process of the pathophysiological events in ischemia-reperfusion is multifactorial and deals with the perception of the oxidative stress responsible for the consequences of ischemia/reperfusion (I/R) stress following TT. Duration and severity of torsion also play a significant role in the oxidative stress. A detrimental result of the defense system of the testes takes place resulting finally in testicular atrophy and impaired function. Antioxidant factors have been experimentally studied in an effort to front this state. They have been classified as endogenous or exogenous antioxidants. Endogenous antioxidants comprise a structure of enzymic enzymatic and non-enzymic enzymatic particles presented within cytoplasm and numerous other subunits in the cells. Exogenous antioxidants include a variety of natural and pharmaceutical agents that may prevent or ameliorate the harmful effects of I/R injury. In this study we review those factors and their ability to enhance the oxidative status of the testis. A feature insight into where we are heading is attempted.

Key words: Testis; Torsion; Experimental; Ischemia-reperfusion injury; Antioxidants

© **The Author(s) 2017.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Testicular torsion is an emergency condition, most commonly seen in newborns and adolescents, which can be considered as an ischemia-reperfusion injury. We provide an overview of the molecular pathogenesis of the disease, and the current evidence of antioxidants use in the experimental torsion-detorsion situation. Possible adaptation of the experimental factors in the clinical practice is discussed.

Vaos G, Zavras N. Antioxidants in experimental ischemia-reperfusion injury of the testis: Where are we heading towards? *World J Methodol* 2017; 7(2): 37-45 Available from: URL: <http://www.wjgnet.com/2222-0682/full/v7/i2/37.htm> DOI: <http://dx.doi.org/10.5662/wjmv7.i2.37>

INTRODUCTION

Testicular torsion (TT) is one of the most serious surgical emergencies, deriving from the twisting of the spermatic cord and its contents, and causing decreased blood flow to the affected testis and finally testicular atrophy^[1,2]. The testis is exclusively prone to ischemic insults due to anatomical reasons (terminal arteries without anastomoses) and the inflexible properties of the tunica albuginea which restricts satisfactory expansion of the testis^[3]. Although, TT can be detected at any age, it is usually seen during perinatal period and puberty^[4-6]. Two main types of TT exist: Extravaginal and intravaginal^[3]. Extravaginal TT is usually seen during perinatal period, and is ought to the absence of normal fixation between testicular coverings and tunica vaginalis resulting in abnormal motility of the testis within scrotum. Intravaginal TT is most commonly seen in adolescent boys and results from a long mesorchium which allows a greater mobility of the testis within the tunica albuginea^[4].

TT has an annual incidence of about 3.8 per 100000 males less than 18 years^[7], and in cases of bilateral torsion, there is evidence that may be inherited^[8]. If left untreated within 4 to 6 h, loss of spermatogenic cells will occur^[9] leading to harmful results such as infertility and subfertility^[10]. The degree of twisting of the spermatic cord may also play an important role. In animal studies, 720° torsion caused significant reduction blood flow when compared with a twisted spermatic cord of 360° or less^[11].

There are two kinds of injuries responsible for testicular necrosis after TT: The first is related to ischemia (I) injury during torsion, and the second to reperfusion (R) injury during detorsion^[12]. I/R injury can cause cell damage from generation of reactive oxygen species (ROS), proinflammatory cytokines and adhesion molecules, lipid peroxidation, apoptosis, anoxia and alteration in microvascular blood flow, which finally lead to testicular atrophy^[13]. Although the testicular environment is characterized by low oxygen tensions, testes are susceptible to oxidative stress due to the plethora of highly unsaturated fatty acids and the presence of ROS^[14].

Antioxidants represent the first line defense of the organism in order to prevent the harmful consequences of I/R injury occurring in the environment of the testicular cell^[15]. Antioxidants may be classified as endogenous and exogenous^[15]. Endogenous antioxidants include a variety of enzymatic molecules that are

presented within the cytoplasm. Common existing endogenous antioxidant enzymes include superoxide dismutase (SOD), catalase, and peroxidases^[15,16]. Exogenous antioxidants include natural derived components such herb productions^[17-25], vitamins^[26-31], selenium^[32], hormones^[33-36], hormones receptors^[37,38], vascular agents^[39-41], phosphodiesterase inhibitors^[42,43], anesthetic and non-steroid anti-inflammatory drugs^[44-47], mucolytic agents^[48], and hyperbaric oxygen^[49]. All have been used in an effort to prevent the consequences of the oxidative stress in I/R injury.

The aim of this review is to present the pathophysiological changes that take place during I/R injury and to summarize the current literature regarding the role of antioxidants in the prevention of experimental I/R injury. Possible translation from the experimental laboratory studies to clinical practice is discussed.

SEARCH

Literature search

We conducted a search focusing on TT and experimental I/R injury in PubMed publishing over the last five years, between 2012 and 2016. The following search terms were used: "testicular torsion", "experimental ischemia-reperfusion injury", "protective agents". A total number of 22 full papers were extracted.

Pathophysiologic alterations during I/R injury

The pathophysiological alterations during I/R injury are multifactorial and difficult to understand. A cascade of events take place during the course of ischemia and further perturbations of biomolecules in cells are seeing during the blood re-establishment after reperfusion. The basic mechanisms of I/R are described below.

Ischemia injury: The role of Ca²⁺: During ischemia a decrease of cell pH is observed due to accumulation of lactic acid, protons and NAD⁺. To balance these alterations, the cell forces out H⁺ via the Na⁺/H⁺ exchanger system^[50]. Thereafter, Na⁺ ions are swapped for Ca²⁺ by the plasmalemmal Na⁺/Ca²⁺ exchanger, which results in increase of Ca²⁺, exacerbated furthermore during reperfusion. These huge alterations in Ca²⁺ stimulate an array of systems, which finally contribute to cell death^[50-52]. For instance, Ca²⁺ entry into the mitochondria via a mitochondrial protein further increases the lethal concentration of Ca²⁺^[53-55]. In addition, the Ca²⁺ cytosolic elevation during I/R can trigger the Ca²⁺/calmodulin-dependent protein kinases, which further added to cell death and tissue dysfunction^[53]. Additionally, the activation of calpains, a family of cysteine proteases by Ca²⁺ elevation, further degrades a group of intracellular proteins, including cytoskeletal, endoplasmic reticulum, and mitochondrial proteins^[56]. Furthermore, Ca²⁺ forces the creation of calcium pyrophosphate structures and uric acid, a pair that binds to a protein complex called inflammasomes which in turn increase the production

of cytokines IL-1 β , and TNF, which lead to a cytokine cyclone that irritate further the I/R injury^[53].

Reperfusion injury: Studies have shown that during reperfusion, the returned oxygenated blood restores the ATP production but also results in production of ROS, which in turn may modify every biomolecule found in cells, producing further cell dysfunction (oxygen paradox)^[57,58]. Redox molecules derived from nitric oxide (NO), the so called reactive nitrogen species (RNS) interact with ROS and lead to the production of reactive nitric oxide species (RNOS), such as peroxynitrite, responsible for harmful damage of macromolecules, initiation of death of endothelial and parenchymal cells, stimulation and release of pro-inflammatory mediators by various cell groups, and induction of adhesion molecules supporting leukocyte/lymphocyte-endothelial cells interactions, and reduction of protective NO^[57-59].

Oxidative stress: The classic theory of oxidative stress was that it arises from an imbalance between pro-oxidants vs antioxidants intracellular compounds^[39]. Currently, it is believed that oxidative stress is involved in three mechanisms in I/R injury: (1) indirect, through non-radical oxidants such as hydrogen peroxide (H₂O₂); (2) modulator, *via* molecular bond, oxidative or nitrosative modification of principle regulatory proteins; and (3) direct damage by oxidant radicals of DNA, proteins, lipids and carbohydrates^[53,60].

Superoxide anion radical (O₂⁻) is the first product of ROS during I/R injury, and subsequently all the other reactive species are derived from interactions or dismutation with other reactive species^[39]. This is supported by experimental studies showing that I/R were considerably attenuated by treatment with SOD or SOD analogues^[53,61,62]. O₂⁻ oxidizes various biomolecules and inactivates enzymes such as NADH, creatine kinase, and calcineurin^[58]. Sources of O₂⁻ are xanthine oxidoreductase, NADPH oxidase, cytochrome P450, and uncoupled nitric oxide species (NOS)^[53].

Nitric oxide stress: Nitric oxide (NO⁻) is elicited during oxidation of arginine to citrulline, through nitrite or nitrate through the action of xanthine oxidoreductase, or by mitochondrial cytochrome c^[63,64]. NO⁻ plays a protective role in the vascular system by producing dilation of blood vessels, modulating platelets aggregation and adhesions, and inhibiting leukocyte-endothelial adhesive interactions and angiogenesis^[53]. Interactions of NO with O₂ or O₂⁻ forming N₂O₃ or peroxynitrite, are associated with overproduction of NO and O₂⁻ resulting in pathophysiological nitrosative and oxidative stress^[53].

In summary, the oxidative/nitric oxide stress may have negative impact on the cell function in I/R stress through three ways: (1) destruction of cellular macromolecules such as membrane lipids, proteins, and DNA; (2) production of possibly toxic peroxynitrite and other RNOS; and (3) side effects on distinct cellular

systems and functions^[53].

Current antioxidant treatment of I/R injury in experimental TT

Comparable to other tissue-cells which live under aerobic conditions, spermatozoa produce ROS which is a physiological process activity^[65]. Moreover, spermatozoa contain an array of ROS scavengers such as SOD, catalase, and substances such as ascorbic acid, taurine, hypotaurine, albumin, and carnitine to balance any ROS high concentration. However, any increased concentration of toxic metabolic products over the ROS scavenging ability, may cause loss of sperm motility and viability^[66-68].

A substantial number of experimental studies by using different agents have studied experimental TT focusing on the effect of I/R injury on ipsilateral and contralateral testis, on treatment and prevention of this injury^[53]. However, conflicting results are raised due to different animal species, such as rats or pigs, model of I/R injury, age, and technique that has been performed to evaluate the I/R damage^[69]. Furthermore, several experimental studies proposed that the contralateral testis is not affected by unilateral torsion^[70-72]. Nevertheless, there is evidence that both testes are affected, and contralateral testis is not disturbed by initial removal of the torsed testis and pretreatment with antioxidants^[73-75].

There are two therapeutic opportunities to counteract oxidative stress. In the first, the superoxide radical and hydrogen peroxide are eliminated by using specific enzymes such as SOD, catalase, and glutathione peroxidase (GPX) either by administration of these enzymes or by increasing them *in vivo* actions. In the second, radical production is prevented by antioxidant scavenging systems^[66].

Some authors showed that apigenin may prevent lipid peroxidation and protect the antioxidant system^[76,77]. We also found a decrease in immunoreactivity of TNF and IL-10, suggesting a synergistic action of apigenin with endogenous IL-10. This antioxidant effect may be due to the H⁺ donation of the OH⁻ aromatic group^[6]. Among others, we demonstrated^[42] that intraperitoneal injection of erythropoietin and sildenafil protects against I/R injury.

Amlodipine is a calcium channel blocker with antioxidants properties, effectively decreasing experimental vascular ischemia-induced damage in the liver and other tissues^[78]. Dogan *et al.*^[79] examined the effect of amlodipine in a rat model of TT injury. They found a significant decrease of TNF and transforming growth factor-beta in the treatment group, decreases in free radicals and increases in antioxidants such as SOD and GSH.

Goji berry (GB) is a traditional Chinese plant product, from the Solanaceae family with antioxidant effects. In experimental studies, GB has been shown to reduce blood sugar and lipid levels, and exhibits male fertility-enhancing effects, immunomodulating,

antitumor, and anti-fatigue properties. GB is composed from six monosaccharides and influences its effects *via* ion exchange chromatography. In a rat experimental study of TT, administration of GB reduced I/R injury by the antioxidant effects of GB^[9].

Mannitol is usually administered before partial nephrectomy to reduce renal damage due to intravascular volume expansion and its free-radical scavenging^[80]. Kurt *et al*^[81] in a rat model of TT, demonstrated that the treatment with mannitol group had less seminiferous tubules disruptions when compared to the TT group without mannitol treatment.

Hesperidin, is another antioxidant compound belonging to flavones with significant antioxidant effects in many tissues^[82,83]. Hesperidin was given intraperitoneally by Celik *et al*^[12] in an experimental group of rats underwent TT and the sample was compared to control group. They found a reduced effect on histological examinations of the hesperidin group when compared to control, while MDA levels were increased, and SOD, catalase and GSH levels were decreased as compared to the control group, concluding that hesperidin has positive results in cases of TT.

Polyphenolic catechins are components of green tea and comprise (-)- epicatechin, (-)- epigallocatechin, (-)- epicatechin gallate, and (-)- epigallocatechin gallate (EGCG)^[84]. Sugiyama *et al*^[85] studied an experimental rat model by producing 4 hours' ischemia and giving orally a single dose of (-)- EGCG 1 h before reperfusion. Histologic examination 4 wk after reperfusion found that EGCG protected against testicular damage from I/R injury and inhibited a further decrease in the activity of SOD.

Dexketoprofen, is a racemic mixture from the arylpropionic acid family of NSAIDS. Yildirim *et al*^[86] studied the intraperitoneal effect of dexketoprofen in a rat model of I/R injury. Malondialdehyde (MDA) levels were investigated in tissue and serum of torsioned testicles in the dexketoprofen group and control group. They found a statistically lower serum MDA levels in the dexketoprofen group compared to control group, and decreased, but not statistically significant, pathological changes in the spermatogenic cells of the control group.

Tyrphostin AG 556 is a tyrosine kinase inhibitor and belongs to the tyrphostin group which has been assessed in animal models of spinal cord and coronary I/R injury^[87,88]. Karaguzel *et al*^[89] investigated the effect of Tyrphostin AG 556 by giving it intraperitoneally and measured the following biochemical parameters: MDA, ischemia modified albumin, signal peptide-CUB (complement C1r/C1s, Uegf, and Bmp1), epidermal growth factor like domain-containing protein1, oxidative stress index, total oxidant status, and total antioxidant status. They concluded that tyrphostin AG 556 has a protective effect on I/R injury

The protective effect of udenafil citrate, piracetam and dexmedetomidine in different doses was evaluated by Tuglu *et al*^[90] and found that all these agents have antioxidant effects on I/R injury.

Grape seed proanthocyanidin extract has been reported to display better antioxidant activity than other antioxidants such as vitamin C, vitamin E, and gallic acid^[91]. Bayatli *et al*^[92] examined the protective effect of grape seed proanthocyanidin after TT performed for 2 h and administered it daily for a week prior to torsion/detorsion. They reported that grape seed proanthocyanidine prohibited the rise of MDA, apoptosis and endothelial nitric oxide synthase expression and enhanced testicular morphology.

Carnosine, is a dipeptide found in high amounts in mammalian tissues^[93]. Abbasoglu *et al*^[94] demonstrated that carnosine treatment has a protective effect on pro-oxidant and antioxidant status in rat testes with I/R injury.

Ozone has been studied as a potential therapeutic agent for the treatment of various physio-pathologic conditions expressing high levels of ROS^[95,96]. Ekici *et al*^[97] assessed the potential effects of ozone in testicular function and morphology in a rat experimental study, in a mixture of ozone/oxygen and compared the results with those of melatonin. They found similar results in the amelioration of I/R injury between melatonin and ozone, but in different pathways.

Ethyl pyruvate, a ROS scavenger, has been found to ameliorate in different conditions such as sepsis, acute pancreatitis, burn, radiation injury and hemorrhagic shock^[98,99]. Turkmen *et al*^[100] reported that ethyl pyruvate has a positive effect on torsion-detorsion associated I/R injury in an experimental rat model.

Carvedilol is a third generation vasodilator agent which has been used in the treatment of hypertension, congestive heart failure and ischemic heart disease^[101,102]. Parlaktas *et al*^[103] investigated the antioxidant effects of carvedilol against I/R injury, and found a decrease in MDA and protein carbonyl and an increase in the level of antioxidant enzymes SOD, and GPX, but not histopathological changes against the control group. They concluded, that carvedilol may have a potential therapeutic value and improve fertility in the clinical practice in patients with TT.

Jiang *et al*^[104] investigated the effect of intraperitoneally injected hydrogen rich saline solution on the protection against testicular damage induced by I/R injury in rats. They found a significant decrease of MDA and a significant improvement of SOD activity in the group of rats which received hydrogen rich saline solution. Therefore, hydrogen rich saline solution may have a protective and therapeutic action against testicular damage.

Inhaled hydrogen gas has been shown to produce a therapeutic activity in a middle cerebral artery occlusion in a rat model and reduce infarct volumes of brain, liver, and myocardium^[105,106]. Lee *et al*^[107] studied the possible therapeutic properties of inhaled 2% hydrogen in pubertal rat model underwent testicular I/R injury. The results of histopathological and biochemical studies suggested that inhalation of hydrogen gas has anti-apoptotic and anti-oxidant properties in cases of TT.

Alpha-lipoic acid is an eight-carbon endogenous cofactor which works against oxygen radicals^[108]. It is established that α -lipoic acid catches hydroxyl and nitric oxide radicals, peroxynitrite anions and hydrogen peroxide. Moreover, α -lipoic acid may act indirectly by enhancing the level of other natural antioxidants such as glutathione, ascorbic acid and tocopherol^[31,109-111]. Ozbal *et al.*^[108] investigated the role of α -lipoic acid in testicular I/R injury in rats and concluded that it is a potential beneficial agent in preserving testicular function.

Genistein is an isoflavone extracted by soy^[112] which displays anti-oxidant and anti-inflammatory properties^[113]. Furthermore, genistein promotes steroidogenesis by restriction progesterone synthesis and decreases secretion of cortisol and corticosterone in mature female pigs^[114]. In addition, it has a protective role against gamma irradiation-induced testicular dysfunction^[115]. Recently, Al-Maghrebi *et al.*^[116] reported that genistein protects the extracellular matrix of the testis which is responsible for the structural integrity of the testicular components, and prevents spermatogenesis's suppression, mitigating oxidative stress and apoptosis in experimental testicular I/R injury.

Nuclear factor kappa B plays a crucial role in immune response, cellular proliferation, inflammatory, and apoptosis^[117]. Pyrrolidine dithiocarbamate (PDTC) is a stable low-molecular thiol compound which acts by neutralizing ROS^[118]. Kemahli *et al.*^[118] studied the antioxidant effect of PDTC in a TT model and found that administration of PDTC exaggerates the antioxidant system by lowering MDA levels, increasing SOD activity and improving Johnson scores of biopsy specimen.

Urocortin, is a 40-amino acid peptide found in different organs, such as digestive tract, cardiovascular and reproductive system^[119]. For instance, urocortin has been shown that protect cardiovascular system against I/R injury^[120]. Sumii *et al.*^[121] investigated the role of urocortin in testicular apoptosis in an experimental I/R rat model and found a cytoprotective role in germ cells through the activation of anti-apoptotic proteins.

Melatonin is an endogenous compound secreted by the pineal gland and influences reproduction *via* its activity on the hypothalamus^[122]. Kurcer *et al.*^[123] reported that melatonin protects testicular tissue against oxidation and alleviates histopathologic changes after experimental testicular I/R injury. Metformin belongs to the biguanide family and has the capacity to reduce ROS^[124]. Asghari *et al.*^[125] investigated a combined use of melatonin and metformin in a rat model and found that may protect the testes from I/R injury by restoring SOD activity, and MDA and myeloperoxidase levels.

Very recently Erol *et al.*^[126] investigated the effect of a antioxidant factors combination, constituting either by L-carnitine, fructose, citric acid, ascorbic acid, cyanocobalamin, selenium, coenzyme Q10, zinc and folic acid or fructose, cellulose microcrystalline, pygeum shell, L-arginine, L-carnitine, zinc, vitamin E, folic acid, vitamin B6, sodium selenite, and hydroxypropyl methyl cellulose. They found that combined antioxidants were

more effective than one protective antioxidant by reducing apoptosis and preventing I/R injury.

Antioxidants and I/R injury in clinical practice

The large body of experimental studies demonstrated undoubtedly that oxidative stress is a dominant factor in the creation of testis impairment after I/R injury. Furthermore, all these antioxidative compounds have been sought to be clearly capable to protect testicular function from oxidative stress. However the relationship between experimental results and clinical practice has not come together until now. A feature mandatory pursuit is to advance understanding of the basic mechanism of oxidative stress in the male reproductive tract and to develop optimizing antioxidant factors in order to treat the pathological consequences from imbalance in the oxidation state of testicular tissue. These mandatory demands are beyond laboratory ways that outline the present approach to counterbalance the deleterious effects of TT.

CONCLUSION

Currently, a large number of studies investigate the role of I/R injury in experimental animal models and many antioxidants and free radical scavengers have been studied to indicate their possible application in human beings. However, the molecular mechanism by which these agents may control the harmful effect of TT has to be clarified. Moreover, experimentally checked drugs or compounds still anticipate clinical utilization. Additional experimental and future clinical studies have to be performed to further assess the effects on antioxidant therapy.

REFERENCES

- 1 Moslemi MK, Kamalimotlagh S. Evaluation of acute scrotum in our consecutive operated cases: a one-center study. *Int J Gen Med* 2014; **7**: 75-78 [PMID: 24470769 DOI: 10.2147/IJGM.S52413]
- 2 Sharp VJ, Kieran K, Arlen AM. Testicular torsion: diagnosis, evaluation, and management. *Am Fam Physician* 2013; **88**: 835-840 [PMID: 24364548]
- 3 Karaguzel E, Kadihasanoglu M, Kutlu O. Mechanisms of testicular torsion and potential protective agents. *Nat Rev Urol* 2014; **11**: 391-399 [PMID: 24934447 DOI: 10.1038/nrurol.2014.135]
- 4 Bowlin PR, Gatti JM, Murphy JP. Pediatric Testicular Torsion. *Surg Clin North Am* 2017; **97**: 161-172 [PMID: 27894425 DOI: 10.1016/j.suc.2016.08.012]
- 5 Drlik M, Kočvara R. Torsion of spermatic cord in children: a review. *J Pediatr Urol* 2013; **9**: 259-266 [PMID: 22763105 DOI: 10.1016/j.jpuro.2012.05.016]
- 6 Skondras I, Lambropoulou M, Tsaroucha A, Gardikis S, Tripsianis G, Simopoulos C, Vaos G. The role of Apigenin in testicular damage in experimental ischemia-reperfusion injury in rats. *Hippokratia* 2015; **19**: 225-230 [PMID: 27418781]
- 7 Zhao LC, Lautz TB, Meeks JJ, Maizels M. Pediatric testicular torsion epidemiology using a national database: incidence, risk of orchiectomy and possible measures toward improving the quality of care. *J Urol* 2011; **186**: 2009-2013 [PMID: 21944120 DOI: 10.1016/j.juro.2011.07.024]
- 8 Shteynshlyuger A, Yu J. Familial testicular torsion: a meta analysis suggests inheritance. *J Pediatr Urol* 2013; **9**: 683-690 [PMID: 23811111]

- 23017841 DOI: 10.1016/j.jpuro.2012.08.002]
- 9 **Dursun R**, Zengin Y, Gündüz E, İçer M, Durgun HM, Dağgüllü M, Kaplan İ, Alabalık U, Güloğlu C. The protective effect of goji berry extract in ischemic reperfusion in testis torsion. *Int J Clin Exp Med* 2015; **8**: 2727-2733 [PMID: 25932226]
- 10 **Parlaktas BS**, Atilgan D, Ozyurt H, Gençten Y, Akbas A, Erdemir F, Uluocak N. The biochemical effects of ischemia-reperfusion injury in the ipsilateral and contralateral testes of rats and the protective role of melatonin. *Asian J Androl* 2014; **16**: 314-318 [PMID: 24407181 DOI: 10.4103/1008-682X.122202]
- 11 **Lievano G**, Nguyen L, Radhakrishnan J, Fornell L, John E. New animal model to evaluate testicular blood flow during testicular torsion. *J Pediatr Surg* 1999; **34**: 1004-1006 [PMID: 10392923 DOI: 10.1016/S0022-3468(99)90778-9]
- 12 **Celik E**, Oguzturk H, Sahin N, Turtay MG, Oguz F, Ciftci O. Protective effects of hesperidin in experimental testicular ischemia/reperfusion injury in rats. *Arch Med Sci* 2016; **12**: 928-934 [PMID: 27695481 DOI: 10.5114/aoms.2015.47697]
- 13 **Cuzzocrea S**, Riley DP, Caputi AP, Salvemini D. Antioxidant therapy: a new pharmacological approach in shock, inflammation, and ischemia/reperfusion injury. *Pharmacol Rev* 2001; **53**: 135-159 [PMID: 11171943]
- 14 **Aitken RJ**, Roman SD. Antioxidant systems and oxidative stress in the testes. *Adv Exp Med Biol* 2008; **636**: 154-171 [PMID: 19856167 DOI: 10.1007/978-0-387-09597-4_9]
- 15 **Rahal A**, Kumar A, Singh V, Yadav B, Tiwari R, Chakraborty S, Dhama K. Oxidative stress, prooxidants, and antioxidants: the interplay. *Biomed Res Int* 2014; **2014**: 761264 [PMID: 24587990 DOI: 10.1155/2014/761264]
- 16 **Turner TT**, Lysiak JJ. Oxidative stress: a common factor in testicular dysfunction. *J Androl* 2008; **29**: 488-498 [PMID: 18567643 DOI: 10.2164/jandrol.108.005132]
- 17 **Lin Y**, Shi R, Wang X, Shen HM. Luteolin, a flavonoid with potential for cancer prevention and therapy. *Curr Cancer Drug Targets* 2008; **8**: 634-646 [PMID: 18991571 DOI: 10.2174/156800908786241050]
- 18 **Diplock AT**, Charleux JL, Crozier-Willi G, Kok FJ, Rice-Evans C, Roberfroid M, Stahl W, Viña-Ribes J. Functional food science and defence against reactive oxidative species. *Br J Nutr* 1998; **80** Suppl 1: S77-112 [PMID: 9849355 DOI: 10.1079/BJN19980106]
- 19 **Jaganathan SK**, Mandal M. Antiproliferative effects of honey and of its polyphenols: a review. *J Biomed Biotechnol* 2009; **2009**: 830616 [PMID: 19636435 DOI: 10.1155/2009/830616]
- 20 **Korkmaz A**, Kolankaya D. Protective effect of rutin on the ischemia/reperfusion induced damage in rat kidney. *J Surg Res* 2010; **164**: 309-315 [PMID: 19592016 DOI: 10.1016/j.jss.2009.03.022]
- 21 **Guimarães SB**, Santos JM, Aragão AA, Kimura OS, Silveira ER, Vasconcelos PR. Ternatin pretreatment attenuates testicular injury induced by torsion/detorsion in Wistar rats. *Acta Cir Bras* 2011; **26**: 325-328 [PMID: 21808848 DOI: 10.1590/S0102-86502011000400014]
- 22 **Aktoz T**, Kanter M, Aktas C. Protective effects of quercetin on testicular torsion/detorsion-induced ischaemia-reperfusion injury in rats. *Andrologia* 2010; **42**: 376-383 [PMID: 21105888 DOI: 10.1111/j.1439-0272.2010.01044.x]
- 23 **Hekimoglu A**, Kurcer Z, Aral F, Baba F, Sahna E, Atessahin A. Lycopene, an antioxidant carotenoid, attenuates testicular injury caused by ischemia/reperfusion in rats. *Tohoku J Exp Med* 2009; **218**: 141-147 [PMID: 19478470 DOI: 10.1620/tjem.218.141]
- 24 **Kim YH**, Kim GH, Shin JH, Kim KS, Lim JS. Effect of Korean red ginseng on testicular tissue injury after torsion and detorsion. *Korean J Urol* 2010; **51**: 794-799 [PMID: 21165202 DOI: 10.4111/kju.2010.51.11.794]
- 25 **Maffei Facino R**, Carini M, Aldini G, Berti F, Rossoni G. Panax ginseng administration in the rat prevents myocardial ischemia-reperfusion damage induced by hyperbaric oxygen: evidence for an antioxidant intervention. *Planta Med* 1999; **65**: 614-619 [PMID: 10575376 DOI: 10.1055/s-1999-14034]
- 26 **Azizollahi S**, Babaei H, Derakhshanfar A, Oloumi MM. Effects of co-administration of dopamine and vitamin C on ischaemia-reperfusion injury after experimental testicular torsion-detorsion in rats. *Andrologia* 2011; **43**: 100-105 [PMID: 21382063 DOI: 10.1111/j.1439-0272.2009.01028.x]
- 27 **Romeo C**, Antonuccio P, Esposito M, Marini H, Impellizzeri P, Turiaco N, Altavilla D, Bitto A, Zuccarello B, Squadrito F. Raxofelast, a hydrophilic vitamin E-like antioxidant, reduces testicular ischemia-reperfusion injury. *Urol Res* 2004; **32**: 367-371 [PMID: 15316698 DOI: 10.1007/s00240-004-0436-4]
- 28 **Antonuccio P**, Minutoli L, Romeo C, Nicotina PA, Bitto A, Arena S, Altavilla D, Zuccarello B, Polito F, Squadrito F. Lipid peroxidation activates mitogen-activated protein kinases in testicular ischemia-reperfusion injury. *J Urol* 2006; **176**: 1666-1672 [PMID: 16952711 DOI: 10.1016/j.juro.2006.06.086]
- 29 **Slyshenkov VS**, Dymkowska D, Wojtczak L. Pantothenic acid and pantothenol increase biosynthesis of glutathione by boosting cell energetics. *FEBS Lett* 2004; **569**: 169-172 [PMID: 15225628 DOI: 10.1016/j.febslet.2004.05.044]
- 30 **Etensel B**, Ozkısacık S, Ozkara E, Serbest YA, Oztan O, Yazıcı M, Gürsoy H. The protective effect of dexpanthenol on testicular atrophy at 60th day following experimental testicular torsion. *Pediatr Surg Int* 2007; **23**: 271-275 [PMID: 17205291 DOI: 10.1007/s00383-006-1871-9]
- 31 **Petersen Shay K**, Moreau RF, Smith EJ, Hagen TM. Is alpha-lipoic acid a scavenger of reactive oxygen species in vivo? Evidence for its initiation of stress signaling pathways that promote endogenous antioxidant capacity. *IUBMB Life* 2008; **60**: 362-367 [PMID: 18409172 DOI: 10.1002/iub.40]
- 32 **Ursini F**, Bindoli A. The role of selenium peroxidases in the protection against oxidative damage of membranes. *Chem Phys Lipids* 1987; **44**: 255-276 [PMID: 3311419 DOI: 10.1016/0009-3084(87)90053-3]
- 33 **Katavetin P**, Tungsanga K, Eiam-Ong S, Nangaku M. Antioxidative effects of erythropoietin. *Kidney Int Suppl* 2007; **(107)**: S10-S15 [PMID: 17943138 DOI: 10.1038/sj.ki.5002482]
- 34 **Yazihan N**, Ataoglu H, Koku N, Erdemli E, Sargin AK. Protective role of erythropoietin during testicular torsion of the rats. *World J Urol* 2007; **25**: 531-536 [PMID: 17690891 DOI: 10.1007/s00345-007-0200-9]
- 35 **Chikuma M**, Masuda S, Kobayashi T, Nagao M, Sasaki R. Tissue-specific regulation of erythropoietin production in the murine kidney, brain, and uterus. *Am J Physiol Endocrinol Metab* 2000; **279**: E1242-E1248 [PMID: 11093910]
- 36 **Koksall M**, Oguz E, Baba F, Eren MA, Ciftci H, Demir ME, Kurcer Z, Take G, Aral F, Ocak AR, Aksoy N, Ulas T. Effects of melatonin on testis histology, oxidative stress and spermatogenesis after experimental testis ischemia-reperfusion in rats. *Eur Rev Med Pharmacol Sci* 2012; **16**: 582-588 [PMID: 22774397]
- 37 **Yapanoglu T**, Aksoy Y, Gursan N, Ozbey I, Ziyapak T, Calik M. Antiapoptotic effects of dehydroepiandrosterone on testicular torsion/detorsion in rats. *Andrologia* 2008; **40**: 38-43 [PMID: 18211300 DOI: 10.1111/j.1439-0272.2008.00806.x]
- 38 **Minutoli L**, Bitto A, Squadrito F, Irrera N, Rinaldi M, Nicotina PA, Arena S, Magno C, Marini H, Spaccapelo L, Ottani A, Giuliani D, Romeo C, Guarini S, Antonuccio P, Altavilla D. Melanocortin 4 receptor activation protects against testicular ischemia-reperfusion injury by triggering the cholinergic antiinflammatory pathway. *Endocrinology* 2011; **152**: 3852-3861 [PMID: 21828180 DOI: 10.1210/en.2011-1016]
- 39 **Mak IT**, Freedman AM, Dickens BF, Weglicki WB. Protective effects of sulfhydryl-containing angiotensin converting enzyme inhibitors against free radical injury in endothelial cells. *Biochem Pharmacol* 1990; **40**: 2169-2175 [PMID: 2173602 DOI: 10.1016/0006-2952(90)90250-O]
- 40 **Yancopoulos GD**, Davis S, Gale NW, Rudge JS, Wiegand SJ, Holash J. Vascular-specific growth factors and blood vessel formation. *Nature* 2000; **407**: 242-248 [PMID: 11001067 DOI: 10.1038/35025215]
- 41 **Shirazi M**, Noorafshan A, Karbalay-Doust S, Ardeshiri M, Afrasiabi MA, Monabati A. Comparison of the protective effects of papaverine, lidocaine and verapamil on the sperm quality of the

- testis after induced torsion-detorsion in rats. *Scand J Urol Nephrol* 2010; **44**: 133-137 [PMID: 20166843 DOI: 10.3109/00365591003636588]
- 42 **Zavras N**, Kostakis ID, Sakellariou S, Damaskos C, Roupakas E, Tsagkari E, Spartalis E, Velaoras K, Dontas IA, Karatzas T. Comparison of erythropoietin and sildenafil protective role against ischemia/reperfusion injury of the testis in adult rats. *Int Urol Nephrol* 2014; **46**: 731-736 [PMID: 24097275 DOI: 10.1007/s11255-013-0569-x]
- 43 **Istanbulluoglu MO**, Zor M, Celik A, Cicek T, Basal S, Ozgok A, Ustun H, Ozgok Y. Effects of vardenafil on testicular torsion/detorsion damage: an experimental study in pigs. *Urol Int* 2011; **86**: 228-232 [PMID: 21124003 DOI: 10.1159/000321492]
- 44 **Kidambi S**, Yarmush J, Fong W, Kamath S, Schianodicola J, Nahmias Y. Propofol induces ERK-dependant expression of c-Fos and Egr-1 in neuronal cells. *Neuroreport* 2009; **20**: 657-662 [PMID: 19349923 DOI: 10.1097/WNR.0b013e328329a449]
- 45 **Yagmurdu H**, Ayyildiz A, Karaguzel E, Ogus E, Surer H, Caydere M, Nuhoglu B, Germiyanoglu C. The preventive effects of thiopental and propofol on testicular ischemia-reperfusion injury. *Acta Anaesthesiol Scand* 2006; **50**: 1238-1243 [PMID: 17067323 DOI: 10.1111/j.1399-6576.2006.01145.x]
- 46 **Hanci V**, Erol B, Bektaş S, Mungan G, Yurtlu S, Tokgöz H, Can M, Ozkoçak Turan I. Effect of dexmedetomidine on testicular torsion/detorsion damage in rats. *Urol Int* 2010; **84**: 105-111 [PMID: 20173379 DOI: 10.1159/000273476]
- 47 **Dokmeci D**, Kanter M, Inan M, Aydogdu N, Basaran UN, Yalcin O, Turan FN. Protective effects of ibuprofen on testicular torsion/detorsion-induced ischemia/reperfusion injury in rats. *Arch Toxicol* 2007; **81**: 655-663 [PMID: 17345063 DOI: 10.1007/s00204-007-0189-2]
- 48 **Aktaş BK**, Bulut S, Bulut S, Baykam MM, Ozden C, Senes M, Yücel D, Memiş A. The effects of N-acetylcysteine on testicular damage in experimental testicular ischemia/reperfusion injury. *Pediatr Surg Int* 2010; **26**: 293-298 [PMID: 19911182 DOI: 10.1007/s00383-009-2538-0]
- 49 **Zhang Y**, Lv Y, Liu YJ, Yang C, Hu HJ, Meng XE, Li MX, Pan SY. Hyperbaric oxygen therapy in rats attenuates ischemia-reperfusion testicular injury through blockade of oxidative stress, suppression of inflammation, and reduction of nitric oxide formation. *Urology* 2013; **82**: 489.e9-489.e15 [PMID: 23769121 DOI: 10.1016/j.urology.2013.04.016]
- 50 **Sanada S**, Komuro I, Kitakaze M. Pathophysiology of myocardial reperfusion injury: preconditioning, postconditioning, and translational aspects of protective measures. *Am J Physiol Heart Circ Physiol* 2011; **301**: H1723-H1741 [PMID: 21856909 DOI: 10.1152/ajpheart.00553.2011]
- 51 **Baines CP**. The mitochondrial permeability transition pore and ischemia-reperfusion injury. *Basic Res Cardiol* 2009; **104**: 181-188 [PMID: 19242640 DOI: 10.1007/s00395-009-0004-8]
- 52 **Baines CP**. The molecular composition of the mitochondrial permeability transition pore. *J Mol Cell Cardiol* 2009; **46**: 850-857 [PMID: 19233198 DOI: 10.1016/j.jmcc.2009.02.007]
- 53 **Kalogeris T**, Baines CP, Krenz M, Korthuis RJ. Cell biology of ischemia/reperfusion injury. *Int Rev Cell Mol Biol* 2012; **298**: 229-317 [PMID: 22878108 DOI: 10.1016/B978-0-12-394309-5.0006-7]
- 54 **Contreras L**, Drago I, Zampese E, Pozzan T. Mitochondria: the calcium connection. *Biochim Biophys Acta* 2010; **1797**: 607-618 [PMID: 20470749 DOI: 10.1016/j.bbabi.2010.05.005]
- 55 **Szydlowska K**, Tymianski M. Calcium, ischemia and excitotoxicity. *Cell Calcium* 2010; **47**: 122-129 [PMID: 20167368 DOI: 10.1016/j.ceca.2010.01.003]
- 56 **Croall DE**, Ersfeld K. The calpains: modular designs and functional diversity. *Genome Biol* 2007; **8**: 218 [PMID: 17608959 DOI: 10.1186/gb-2007-8-6-218]
- 57 **Kvietys PR**, Granger DN. Role of reactive oxygen and nitrogen species in the vascular responses to inflammation. *Free Radic Biol Med* 2012; **52**: 556-592 [PMID: 22154653 DOI: 10.1016/j.freeradbiomed.2011.11.002]
- 58 **Raedschelders K**, Ansley DM, Chen DD. The cellular and molecular origin of reactive oxygen species generation during myocardial ischemia and reperfusion. *Pharmacol Ther* 2012; **133**: 230-255 [PMID: 22138603 DOI: 10.1016/j.pharmthera.2011.11.004]
- 59 **Granger DN**. Ischemia-reperfusion: mechanisms of microvascular dysfunction and the influence of risk factors for cardiovascular disease. *Microcirculation* 1999; **6**: 167-178 [PMID: 10501090 DOI: 10.1111/j.1549-8719.1999.tb00099.x]
- 60 **Granger DN**, Korthuis RJ. Physiologic mechanisms of postischemic tissue injury. *Annu Rev Physiol* 1995; **57**: 311-332 [PMID: 7778871 DOI: 10.1146/annurev.ph.57.030195.001523]
- 61 **Lima B**, Forrester MT, Hess DT, Stamler JS. S-nitrosylation in cardiovascular signaling. *Circ Res* 2010; **106**: 633-646 [PMID: 20203313 DOI: 10.1161/CIRCRESAHA.109.207381]
- 62 **Granger DN**. Role of xanthine oxidase and granulocytes in ischemia-reperfusion injury. *Am J Physiol* 1988; **255**: H1269-H1275 [PMID: 3059826]
- 63 **Horie Y**, Ishii H. Liver dysfunction elicited by gut ischemia-reperfusion. *Pathophysiology* 2001; **8**: 11-20 [PMID: 11476968 DOI: 10.1016/S0928-4680(01)00063-3]
- 64 **Golwala NH**, Hodenette C, Murthy SN, Nossaman BD, Kadowitz PJ. Vascular responses to nitrite are mediated by xanthine oxidoreductase and mitochondrial aldehyde dehydrogenase in the rat. *Can J Physiol Pharmacol* 2009; **87**: 1095-1101 [PMID: 20029546 DOI: 10.1139/Y09-101]
- 65 **Bansal AK**, Bilaspuri GS. Impacts of oxidative stress and antioxidants on semen functions. *Vet Med Int* 2010; **2010**: pii: 686137 [PMID: 20871827 DOI: 10.4061/2011/686137]
- 66 **Dockmeci D**. Oxidative stress and testicular torsion. In: Agarwal A, et al (eds). *Studies on Men's Health and Fertility. Oxidative stress in Applied Basic Research and Clinical Practice*. Springer Science and Business Media, LLC, 2012: 355-398 [DOI: 10.1007/978-1-61779-776-7_17]
- 67 **Said TM**, Agarwal A, Sharma RK, Mascha E, Sikka SC, Thomas AJ. Human sperm superoxide anion generation and correlation with semen quality in patients with male infertility. *Fertil Steril* 2004; **82**: 871-877 [PMID: 15482762 DOI: 10.1016/j.fertnstert.2004.02.132]
- 68 **Saleh RA**, Agarwal A. Oxidative stress and male infertility: from research bench to clinical practice. *J Androl* 2002; **23**: 737-752 [PMID: 12399514 DOI: 10.1007/978-3-642-02062-9_7]
- 69 **Romero FR**, Gomes RP, Lorenzini F, Erdmann TR, Tambara Filho R. Ipsilateral testicular necrosis and atrophy after 1,080-degree torsion of the spermatic cord in rats. *Acta Cir Bras* 2009; **24**: 118-123 [PMID: 19377780 DOI: 10.1590/S0102-86502009000200008]
- 70 **Akgür FM**, Kiliç K, Aktuğ T. Is ipsilateral testis mandatory for the occurrence of contralateral intratesticular biochemical changes indicative of hypoxia after unilateral spermatic cord torsion? *Eur Urol* 1995; **28**: 143-146 [PMID: 8529740]
- 71 **Becker EJ**, Turner TT. Endocrine and exocrine effects of testicular torsion in the prepubertal and adult rat. *J Androl* 1995; **16**: 342-351 [PMID: 8537252]
- 72 **Turner TT**. Acute experimental testicular torsion. No effect on the contralateral testis. *J Androl* 1985; **6**: 65-72 [PMID: 3972720 DOI: 10.1002/j.1939-4640.1985.tb00817.x]
- 73 **Cosentino MJ**, Rabinowitz R, Valvo JR, Cockett AT. The effect of prepubertal spermatic cord torsion on subsequent fertility in rats. *J Androl* 1984; **5**: 93-98 [PMID: 6715258 DOI: 10.1002/j.1939-4640.1984.tb00781.x]
- 74 **Tanyel FC**, Büyükpamukçu N, Hiçsönmez A. Contralateral testicular blood flow during unilateral testicular torsion. *Br J Urol* 1989; **63**: 522-524 [PMID: 2731010 DOI: 10.1111/j.1464-410X.1989.tb05949.x]
- 75 **Thomas WE**, Cooper MJ, Crane GA, Lee G, Williamson RC. Testicular exocrine malfunction after torsion. *Lancet* 1984; **2**: 1357-1360 [PMID: 6150364 DOI: 10.1016/S0140-6736(84)92056-7]
- 76 **Singh JP**, Selvendiran K, Banu SM, Padmavathi R, Sakthisekaran D. Protective role of Apigenin on the status of lipid peroxidation and antioxidant defense against hepatocarcinogenesis in Wistar albino rats. *Phytomedicine* 2004; **11**: 309-314 [PMID: 15185843]

- DOI: 10.1078/0944711041495254]
- 77 **Nagaraja HS**, Anupama BK. Apigenin reduces cyclosporine-A induced changes in lipid hydroperoxides and total antioxidants in Sprague-Dawley rats. *J Chin Clin Med* 2009; **4**: 26-31
- 78 **Halici Z**, Karaca M, Keles ON, Borekci B, Odabasoglu F, Suleyman H, Cadirci E, Bayir Y, Unal B. Protective effects of amlodipine on ischemia-reperfusion injury of rat ovary: biochemical and histopathologic evaluation. *Fertil Steril* 2008; **90**: 2408-2415 [PMID: 18178199 DOI: 10.1016/j.fertnstert.2007.10.007]
- 79 **Dogan C**, Halici Z, Topcu A, Cadirci E, Karakus E, Bayir Y, Selli J. Effects of amlodipine on ischaemia/reperfusion injury in the rat testis. *Andrologia* 2016; **48**: 441-452 [PMID: 26259852 DOI: 10.1111/and.12464]
- 80 **Zager RA**, Mahan J, Merola AJ. Effects of mannitol on the postischemic kidney. Biochemical, functional, and morphologic assessments. *Lab Invest* 1985; **53**: 433-442 [PMID: 3930877]
- 81 **Kurt O**, Yazici CM, Erboglu M, Turan C, Bozdemir Y, Akbas A, Turker P, Aktas C, Aydin M, Yesildag E. Mannitol has a protective effect on testicular torsion: An experimental rat model. *J Pediatr Urol* 2016; **12**: 167.e1-167.e8 [PMID: 26879410 DOI: 10.1016/j.jpuro.2016.01.004]
- 82 **Shagirtha K**, Pari L. Hesperetin, a citrus flavonone, protects potentially cadmium induced oxidative testicular dysfunction in rats. *Ecotoxicol Environ Saf* 2011; **74**: 2105-2111 [PMID: 21719105 DOI: 10.1016/j.ecoenv.2011.06.002]
- 83 **Gaur V**, Kumar A. Hesperidin pre-treatment attenuates NO-mediated cerebral ischemic reperfusion injury and memory dysfunction. *Pharmacol Rep* 2010; **62**: 635-648 [PMID: 20885004 DOI: 10.1016/S1734-1140(10)70321-2]
- 84 **Nagle DG**, Ferreira D, Zhou YD. Epigallocatechin-3-gallate (EGCG): chemical and biomedical perspectives. *Phytochemistry* 2006; **67**: 1849-1855 [PMID: 16876833 DOI: 10.1016/j.phytochem.2006.06.020]
- 85 **Sugiyama A**, Chiba M, Nakagami T, Kawano S, Sanada Y, Tajiri T, Toki A. Beneficial effects of (-)-epigallocatechin gallate on ischemia-reperfusion testicular injury in rats. *J Pediatr Surg* 2012; **47**: 1427-1432 [PMID: 22813808 DOI: 10.1016/j.jpedsurg.2012.01.069]
- 86 **Yildirim Y**, Karakaya D, Kelsaka E, Aksoy A, Gülbahar MY, Bedir A. The effect of dexamethasone on ischemia reperfusion injury. *Bratisl Lek Listy* 2014; **115**: 256-259 [PMID: 25174054 DOI: 10.4149/bll_2014_053]
- 87 **Usul H**, Kahir E, Cobanoglu U, Alver A, Peksoylu B, Topbas M, Baykal S. The effects of tyrophostine AG 556 on experimental spinal cord ischemia reperfusion injury. *Surg Neurol* 2004; **61**: 45-54; discussion 54 [PMID: 14706378 DOI: 10.1016/S0090-3019(03)00539-1]
- 88 **Altavilla D**, Squadrito F, Campo GM, Saitta A, Squadrito G, Quartarone C, Deodato B, Arlotta M, Ferlito M, Minutoli L, Tringali M, Urna G, Sardella A, Caputi AP. The reduction of myocardial damage and leukocyte polymorphonuclear accumulation following coronary artery occlusion by the tyrosine kinase inhibitor tyrphostin AG 556. *Life Sci* 2000; **67**: 2615-2629 [PMID: 11104363 DOI: 10.1016/S0024-3205(00)00845-6]
- 89 **Karaguzel E**, Sivrikaya A, Mentese A, Yulug E, Turkmen S, Kutlu O, Guler Y, Us D, Turedi S, Alver A, Kazaz IO. Investigation of tyrphostin AG 556 for testicular torsion-induced ischemia reperfusion injury in rat. *J Pediatr Urol* 2014; **10**: 223-229 [PMID: 24070787 DOI: 10.1016/j.jpuro.2013.08.007]
- 90 **Tuglu D**, Yuvanc E, Ozan T, Bal F, Yilmaz E, Atasoy P, Kisa U, Batislam E. Protective effects of udenafil citrate, piracetam and dexmedetomidine treatment on testicular torsion/detorsion-induced ischaemia/reperfusion injury in rats. *Andrologia* 2016; **48**: 676-682 [PMID: 26589469 DOI: 10.1111/and.12499]
- 91 **Ariga T**. The antioxidative function, preventive action on disease and utilization of proanthocyanidins. *Biofactors* 2004; **21**: 197-201 [PMID: 15630197 DOI: 10.1002/biof.552210140]
- 92 **Bayatli F**, Akkuş D, Kilic E, Saraymen R, Sönmez MF. The protective effects of grape seed extract on MDA, AOPP, apoptosis and eNOS expression in testicular torsion: an experimental study. *World J Urol* 2013; **31**: 615-622 [PMID: 23475212 DOI: 10.1007/s00345-013-1049-8]
- 93 **Aldini G**, Facino RM, Beretta G, Carini M. Carnosine and related dipeptides as quenchers of reactive carbonyl species: from structural studies to therapeutic perspectives. *Biofactors* 2005; **24**: 77-87 [PMID: 16403966 DOI: 10.1002/biof.5520240109]
- 94 **Abbasoglu L**, Kalaz EB, Soluk-Tekkesin M, Olgaç V, Doğru-Abbasoglu S, Uysal M. Beneficial effects of taurine and carnosine in experimental ischemia/reperfusion injury in testis. *Pediatr Surg Int* 2012; **28**: 1125-1131 [PMID: 22961384 DOI: 10.1007/s00383-012-3168-5]
- 95 **Peralta C**, Xaus C, Bartrons R, Leon OS, Gelpi E, Roselló-Catafau J. Effect of ozone treatment on reactive oxygen species and adenosine production during hepatic ischemia-reperfusion. *Free Radic Res* 2000; **33**: 595-605 [PMID: 11200091 DOI: 10.1080/1071576000301121]
- 96 **Yu G**, Liu X, Chen Z, Chen H, Wang L, Wang Z, Qiu T, Weng X. Ozone therapy could attenuate tubulointerstitial injury in adenine-induced CKD rats by mediating Nrf2 and NF-κB. *Iran J Basic Med Sci* 2016; **19**: 1136-1143 [PMID: 27872711]
- 97 **Ekici S**, Doğan Ekici AI, Öztürk G, Benli Aksungar F, Sinanoğlu O, Turan G, Lüleci N. Comparison of melatonin and ozone in the prevention of reperfusion injury following unilateral testicular torsion in rats. *Urology* 2012; **80**: 899-906 [PMID: 22950989 DOI: 10.1016/j.urology.2012.06.049]
- 98 **Park SY**, Yi EY, Jung M, Lee YM, Kim YJ. Ethyl pyruvate, an anti-inflammatory agent, inhibits tumor angiogenesis through inhibition of the NF-κB signaling pathway. *Cancer Lett* 2011; **303**: 150-154 [PMID: 21333439 DOI: 10.1016/j.canlet.2010.12.024]
- 99 **Turkmen S**, Cekic Gonenc O, Karaca Y, Mentese A, Demir S, Beyhun E, Sahin A, Gunduz A, Yulug E, Turedi S. The effect of ethyl pyruvate and N-acetylcysteine on ischemia-reperfusion injury in an experimental model of ischemic stroke. *Am J Emerg Med* 2016; **34**: 1804-1807 [PMID: 27324856 DOI: 10.1016/j.ajem.2016.06.003]
- 100 **Turkmen S**, Mentese A, Karaguzel E, Karaca Y, Kucuk A, Uzun A, Yulug E, Turedi S. A comparison of the effects of N-acetylcysteine and ethyl pyruvate on experimental testicular ischemia-reperfusion injury. *Fertil Steril* 2012; **98**: 626-631 [PMID: 22717346 DOI: 10.1016/j.fertnstert.2012.05.034]
- 101 **Singh D**, Chander V, Chopra K. Carvedilol attenuates ischemia-reperfusion-induced oxidative renal injury in rats. *Fundam Clin Pharmacol* 2004; **18**: 627-634 [PMID: 15548233 DOI: 10.1111/j.1472-8206.2004.00279.x]
- 102 **Hayashi T**, De Velasco MA, Saitou Y, Nose K, Nishioka T, Ishii T, Uemura H. Carvedilol protects tubular epithelial cells from ischemia-reperfusion injury by inhibiting oxidative stress. *Int J Urol* 2010; **17**: 989-995 [PMID: 20946473 DOI: 10.1111/j.1442-2042.2010.02644.x]
- 103 **Parlaktas BS**, Atılgan D, Gençten Y, Akbas A, Markoc F, Erdemir F, Ozyurt H, Uluocak N. The effects of carvedilol on ischemia-reperfusion injury in the rat testis. *Int Braz J Urol* 2014; **40**: 109-117 [PMID: 24642157 DOI: 10.1590/S1677-5538.IBJU.2014.01.16]
- 104 **Jiang D**, Wu D, Zhang Y, Xu B, Sun X, Li Z. Protective effects of hydrogen rich saline solution on experimental testicular ischemia-reperfusion injury in rats. *J Urol* 2012; **187**: 2249-2253 [PMID: 22503049 DOI: 10.1016/j.juro.2012.01.029]
- 105 **Fukuda K**, Asoh S, Ishikawa M, Yamamoto Y, Ohsawa I, Ohta S. Inhalation of hydrogen gas suppresses hepatic injury caused by ischemia/reperfusion through reducing oxidative stress. *Biochem Biophys Res Commun* 2007; **361**: 670-674 [PMID: 17673169 DOI: 10.1016/j.bbrc.2007.07.088]
- 106 **Hayashida K**, Sano M, Ohsawa I, Shinmura K, Tamaki K, Kimura K, Endo J, Katayama T, Kawamura A, Kohsaka S, Makino S, Ohta S, Ogawa S, Fukuda K. Inhalation of hydrogen gas reduces infarct size in the rat model of myocardial ischemia-reperfusion injury. *Biochem Biophys Res Commun* 2008; **373**: 30-35 [PMID: 18541148 DOI: 10.1016/j.bbrc.2008.05.165]
- 107 **Lee JW**, Kim JI, Lee YA, Lee DH, Song CS, Cho YJ, Han JS. Inhaled hydrogen gas therapy for prevention of testicular ischemia/reperfusion injury in rats. *J Pediatr Surg* 2012; **47**: 736-742 [PMID: 22498389 DOI: 10.1016/j.jpedsurg.2011.09.035]
- 108 **Ozbal S**, Ergur BU, Erbil G, Tekmen I, Bagrıyanık A, Cavdar Z.

- The effects of α -lipoic acid against testicular ischemia-reperfusion injury in Rats. *ScientificWorldJournal* 2012; **2012**: 489248 [PMID: 23193380 DOI: 10.1100/2012/489248]
- 109 **Shay KP**, Moreau RF, Smith EJ, Smith AR, Hagen TM. Alpha-lipoic acid as a dietary supplement: molecular mechanisms and therapeutic potential. *Biochim Biophys Acta* 2009; **1790**: 1149-1160 [PMID: 19664690 DOI: 10.1016/j.bbagen.2009.07.026]
 - 110 **Biewenga GP**, Haenen GR, Bast A. The pharmacology of the antioxidant lipoic acid. *Gen Pharmacol* 1997; **29**: 315-331 [PMID: 9378235 DOI: 10.1016/S0306-3623(96)00474-0]
 - 111 **Gorąca A**, Huk-Kolega H, Piechota A, Kleniewska P, Ciejk E, Skibska B. Lipoic acid - biological activity and therapeutic potential. *Pharmacol Rep* 2011; **63**: 849-858 [PMID: 22001972 DOI: 10.1016/S1734-1140(11)70600-4]
 - 112 **Nagaraju GP**, Zafar SF, El-Rayes BF. Pleiotropic effects of genistein in metabolic, inflammatory, and malignant diseases. *Nutr Rev* 2013; **71**: 562-572 [PMID: 23865800 DOI: 10.1111/nure.12044]
 - 113 **Kim SH**, Kim SH, Kim YB, Jeon YT, Lee SC, Song YS. Genistein inhibits cell growth by modulating various mitogen-activated protein kinases and AKT in cervical cancer cells. *Ann N Y Acad Sci* 2009; **1171**: 495-500 [PMID: 19723095 DOI: 10.1111/j.1749-6632.2009.04899.x]
 - 114 **Kaminska B**, Ciereszko R, Kiezun M, Dusza L. In vitro effects of genistein and daidzein on the activity of adrenocortical steroidogenic enzymes in mature female pigs. *J Physiol Pharmacol* 2013; **64**: 103-108 [PMID: 23568977]
 - 115 **Kim JS**, Heo K, Yi JM, Gong EJ, Yang K, Moon C, Kim SH. Genistein mitigates radiation-induced testicular injury. *Phytother Res* 2012; **26**: 1119-1125 [PMID: 22162311 DOI: 10.1002/ptr.3689]
 - 116 **Al-Maghrebi M**, Renno WM. Genistein alleviates testicular ischemia and reperfusion injury-induced spermatogenic damage and oxidative stress by suppressing abnormal testicular matrix metalloproteinase system via the Notch 2/Jagged 1/Hes-1 and caspase-8 pathways. *J Physiol Pharmacol* 2016; **67**: 129-137 [PMID: 27010902]
 - 117 **Ahn KS**, Sethi G, Aggarwal BB. Nuclear factor-kappa B: from clone to clinic. *Curr Mol Med* 2007; **7**: 619-637 [PMID: 18045141 DOI: 10.2174/156652407782564363]
 - 118 **Kemahli E**, Yildiz M, Firat T, Özyalvaçlı ME, Üyetürk U, Yilmaz B, Güçük A. An experimental study on effects of pyrrolidine dithiocarbamate on ischemia-reperfusion injury in testis. *Can Urol Assoc J* 2016; **10**: E104-E109 [PMID: 27330576 DOI: 10.5489/cuaj.3160]
 - 119 **Scarabelli T**, Knight R. Urocortins: take them to heart. *Curr Med Chem Cardiovasc Hematol Agents* 2004; **2**: 335-342 [PMID: 15320777 DOI: 10.2174/1568016043356174]
 - 120 **Takahashi K**. Distribution of urocortins and corticotropin-releasing factor receptors in the cardiovascular system. *Int J Endocrinol* 2012; **2012**: 395284 [PMID: 22675352 DOI: 10.1155/2012/395284]
 - 121 **Sumii K**, Miyake H, Enatsu N, Chiba K, Fujisawa M. Characterization of urocortin as an anti-apoptotic protein in experimental ischemia-reperfusion model of the rat testis. *Biochem Biophys Res Commun* 2016; **479**: 387-392 [PMID: 27659706 DOI: 10.1016/j.bbrc.2016.09.091]
 - 122 **Take G**, Erdogan D, Helvacioğlu F, Göktas G, Ozbey G, Uluoglu C, Yücel B, Güney Y, Hicsonmez A, Ozkan S. Effect of melatonin and time of administration on irradiation-induced damage to rat testes. *Braz J Med Biol Res* 2009; **42**: 621-628 [PMID: 19578641 DOI: 10.1590/S0100-879X2009000700006]
 - 123 **Kurcer Z**, Oguz E, Ozbilge H, Baba F, Aksoy N, Celik N. Effect of melatonin on testicular ischemia/reperfusion injury in rats: is this effect related to the proinflammatory cytokines? *Fertil Steril* 2008; **89**: 1468-1473 [PMID: 17681337 DOI: 10.1016/j.fertnstert.2007.04.065]
 - 124 **Esteghamati A**, Eskandari D, Mirmiranpour H, Noshad S, Mousavizadeh M, Hedayati M, Nakhjavani M. Effects of metformin on markers of oxidative stress and antioxidant reserve in patients with newly diagnosed type 2 diabetes: a randomized clinical trial. *Clin Nutr* 2013; **32**: 179-185 [PMID: 22963881 DOI: 10.1016/j.clnu.2012.08.006]
 - 125 **Asghari A**, Akbari G, Meghdadi A, Mortazavi P. Effects of melatonin and metformin co-administration on testicular ischemia/reperfusion injury in rats. *J Pediatr Urol* 2016; **12**: 410.e1-410.e7 [PMID: 27595505 DOI: 10.1016/j.jpuro.2016.06.017]
 - 126 **Erol B**, Sari U, Amasyali AS, Ozkanli S, Sogut S, Hanci V, Efiloglu O, Danacioglu YO, Engin P, Yencilek F, Atis G, Yildirim A, Alkoc OA, Caskurlu T. Comparison of combined antioxidants and thymoquinone in the prevention of testis ischemia - reperfusion injury. *Andrology* 2017; **5**: 119-124 [PMID: 27748062 DOI: 10.1111/andr.12268]

P- Reviewer: Chan WH, Weng CF **S- Editor:** Ji FF **L- Editor:** A
E- Editor: Wu HL





Published by **Baishideng Publishing Group Inc**
7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
Telephone: +1-925-223-8242
Fax: +1-925-223-8243
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>

