

World Journal of *Gastroenterology*

World J Gastroenterol 2017 October 7; 23(37): 6747-6922



**EDITORIAL**

- 6747 Microbial dysbiosis in spouses of ulcerative colitis patients: Any clues to disease pathogenesis?
Sorrentino D

REVIEW

- 6750 Road to stemness in hepatocellular carcinoma
Flores-Téllez TNJ, Villa-Treviño S, Piña-Vázquez C
- 6777 Intrahepatic vascular changes in non-alcoholic fatty liver disease: Potential role of insulin-resistance and endothelial dysfunction
Pasarin M, Abrales JG, Liguori E, Kok B, La Mura V
- 6788 Epidemiological and clinical perspectives on irritable bowel syndrome in India, Bangladesh and Malaysia: A review
Rahman MM, Mahadeva S, Ghoshal UC

ORIGINAL ARTICLE**Basic Study**

- 6802 Estrogen receptor expression in chronic hepatitis C and hepatocellular carcinoma pathogenesis
Iyer JK, Kalra M, Kaul A, Payton ME, Kaul R
- 6817 Glycosylation-related gene expression in HT29-MTX-E12 cells upon infection by *Helicobacter pylori*
Cairns MT, Gupta A, Naughton JA, Kane M, Clyne M, Joshi L
- 6833 STAT3 deficiency prevents hepatocarcinogenesis and promotes biliary proliferation in thioacetamide-induced liver injury
Abe M, Yoshida T, Akiba J, Ikezono Y, Wada F, Masuda A, Sakaue T, Tanaka T, Iwamoto H, Nakamura T, Sata M, Koga H, Yoshimura A, Torimura T
- 6845 Performance verification and comparison of TianLong automatic hypersensitive hepatitis B virus DNA quantification system with Roche CAP/CTM system
Li M, Chen L, Liu LM, Li YL, Li BA, Li B, Mao YL, Xia LF, Wang T, Liu YN, Li Z, Guo TS

Case Control Study

- 6854 Association of insertion-deletions polymorphisms with colorectal cancer risk and clinical features
Marques D, Ferreira-Costa LR, Ferreira-Costa LL, Correa RS, Borges AMP, Ito FR, Ramos CCO, Bortolin RH, Luchessi AD, Ribeiro-dos-Santos A, Santos S, Silbiger VN

Retrospective Cohort Study

- 6868 Hospital readmissions in decompensated cirrhotics: Factors pointing toward a prevention strategy

Seraj SM, Campbell EJ, Argyropoulos SK, Wegermann K, Chung RT, Richter JM

- 6877 Measurement of biological age may help to assess the risk of colorectal adenoma in screening colonoscopy

Kim SJ, Kim BJ, Kang H

Retrospective Study

- 6884 Prognostic factors of response to endoscopic treatment in painful chronic pancreatitis

Tantau A, Mandrutiu A, Leucuta DC, Ciobanu L, Tantau M

- 6894 *In vivo* histological diagnosis for gastric cancer using endocytoscopy

Tsurudome I, Miyahara R, Funasaka K, Furukawa K, Matsushita M, Yamamura T, Ishikawa T, Ohno E, Nakamura M, Kawashima H, Watanabe O, Nakaguro M, Satou A, Hirooka Y, Goto H

CASE REPORT

- 6902 Achalasia after bariatric Roux-en-Y gastric bypass surgery reversal

Abu Ghanimeh M, Qasrawi A, Abughanimeh O, Albadarin S, Clarkston W

- 6907 Persistent severe hypomagnesemia caused by proton pump inhibitor resolved after laparoscopic fundoplication

Semb S, Helgstrand F, Hjørne F, Bytzer P

- 6911 Rupture of small cystic pancreatic neuroendocrine tumor with many microtumors

Sagami R, Nishikiori H, Ikuyama S, Murakami K

LETTERS TO THE EDITOR

- 6920 Resistance of *Helicobacter pylori* to furazolidone and levofloxacin: A viewpoint

Zamani M, Rahbar A, Shokri-Shirvani J

ABOUT COVER

Editorial board member of *World Journal of Gastroenterology*, Toru Mizuguchi, MD, PhD, Associate Professor, Surgeon, Department of Surgery, Surgical Oncology and Science, Sapporo Medical University Hospital, Sapporo 060-8543, Hokkaido, Japan

AIMS AND SCOPE

World Journal of Gastroenterology (*World J Gastroenterol*, *WJG*, print ISSN 1007-9327, online ISSN 2219-2840, DOI: 10.3748) is a peer-reviewed open access journal. *WJG* was established on October 1, 1995. It is published weekly on the 7th, 14th, 21st, and 28th each month. The *WJG* Editorial Board consists of 1375 experts in gastroenterology and hepatology from 68 countries.

The primary task of *WJG* is to rapidly publish high-quality original articles, reviews, and commentaries in the fields of gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, hepatobiliary surgery, gastrointestinal oncology, gastrointestinal radiation oncology, gastrointestinal imaging, gastrointestinal interventional therapy, gastrointestinal infectious diseases, gastrointestinal pharmacology, gastrointestinal pathophysiology, gastrointestinal pathology, evidence-based medicine in gastroenterology, pancreatology, gastrointestinal laboratory medicine, gastrointestinal molecular biology, gastrointestinal immunology, gastrointestinal microbiology, gastrointestinal genetics, gastrointestinal translational medicine, gastrointestinal diagnostics, and gastrointestinal therapeutics. *WJG* is dedicated to become an influential and prestigious journal in gastroenterology and hepatology, to promote the development of above disciplines, and to improve the diagnostic and therapeutic skill and expertise of clinicians.

INDEXING/ABSTRACTING

World Journal of Gastroenterology (*WJG*) is now indexed in Current Contents[®]/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch[®]), Journal Citation Reports[®], Index Medicus, MEDLINE, PubMed, PubMed Central and Directory of Open Access Journals. The 2017 edition of Journal Citation Reports[®] cites the 2016 impact factor for *WJG* as 3.365 (5-year impact factor: 3.176), ranking *WJG* as 29th among 79 journals in gastroenterology and hepatology (quartile in category Q2).

FLYLEAF

I-IX Editorial Board

EDITORS FOR THIS ISSUE

Responsible Assistant Editor: Xiang Li
Responsible Electronic Editor: Yan Huang
Proofing Editor-in-Chief: Lian-Sheng Ma

Responsible Science Editor: Li-Juan Wei
Proofing Editorial Office Director: Jin-Lei Wang

NAME OF JOURNAL
World Journal of Gastroenterology

ISSN
ISSN 1007-9327 (print)
ISSN 2219-2840 (online)

LAUNCH DATE
October 1, 1995

FREQUENCY
Weekly

EDITORS-IN-CHIEF
Damian Garcia-Olmo, MD, PhD, Doctor, Professor, Surgeon, Department of Surgery, Universidad Autonoma de Madrid; Department of General Surgery, Fundacion Jimenez Diaz University Hospital, Madrid 28040, Spain

Stephen C Strom, PhD, Professor, Department of Laboratory Medicine, Division of Pathology, Karolinska Institutet, Stockholm 141-86, Sweden

Andrzej S Tarnawski, MD, PhD, DSc (Med), Professor of Medicine, Chief Gastroenterology, VA Long Beach Health Care System, University of California, Irvine, CA, 5901 E. Seventh Str., Long Beach,

CA 90822, United States

EDITORIAL BOARD MEMBERS
All editorial board members resources online at <http://www.wjgnet.com/1007-9327/editorialboard.htm>

EDITORIAL OFFICE
Jin-Lei Wang, Director
Yuan Qi, Vice Director
Ze-Mao Gong, Vice Director
World Journal of Gastroenterology
Baishideng Publishing Group Inc
7901 Stoneridge Drive, Suite 501,
Pleasanton, CA 94588, USA
Telephone: +1-925-2238242
Fax: +1-925-2238243
E-mail: editorialoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>

PUBLISHER
Baishideng Publishing Group Inc
7901 Stoneridge Drive, Suite 501,
Pleasanton, CA 94588, USA
Telephone: +1-925-2238242
Fax: +1-925-2238243
E-mail: bpoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>

<http://www.wjgnet.com>

PUBLICATION DATE
October 7, 2017

COPYRIGHT
© 2017 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT
All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORS
Full instructions are available online at <http://www.wjgnet.com/bpg/gerinfo/204>

ONLINE SUBMISSION
<http://www.f6publishing.com>

Case Control Study

Association of insertion-deletions polymorphisms with colorectal cancer risk and clinical features

Diego Marques, Layse Raynara Ferreira-Costa, Lorena Larissa Ferreira-Costa, Romualdo da Silva Correa, Aline Maciel Pinheiro Borges, Fernanda Ribeiro Ito, Carlos Cesar de Oliveira Ramos, Raul Hernandes Bortolin, André Ducati Luchessi, Ândrea Ribeiro-dos-Santos, Sidney Santos, Vivian Nogueira Silbiger

Diego Marques, Layse Raynara Ferreira-Costa, Lorena Larissa Ferreira-Costa, Raul Hernandes Bortolin, André Ducati Luchessi, Vivian Nogueira Silbiger, Laboratório de Bioanálise e Biotecnologia Molecular, Universidade Federal do Rio Grande do Norte, Natal 59012-570, Rio Grande do Norte, Brazil

André Ducati Luchessi, Vivian Nogueira Silbiger, Departamento de Análises Clínicas e Toxicológicas, Universidade Federal do Rio Grande do Norte, Natal 59012-570, Rio Grande do Norte, Brazil

Diego Marques, Raul Hernandes Bortolin, André Ducati Luchessi, Vivian Nogueira Silbiger, Programa de Pós-graduação em Ciências Farmacêuticas, Universidade Federal do Rio Grande do Norte, Natal 59012-570, Rio Grande do Norte, Brazil

Romualdo da Silva Correa, Aline Maciel Pinheiro Borges, Fernanda Ribeiro Ito, Departamento de Cirurgia Oncológica, Liga Norte Riograndense Contra o Câncer, Natal 59040-000, Rio Grande do Norte, Brazil

Carlos Cesar de Oliveira Ramos, Laboratório de Patologia e Citopatologia, Liga Norte Riograndense Contra o Câncer, Natal 59040-000, Rio Grande do Norte, Brazil

Diego Marques, Ândrea Ribeiro-dos-Santos, Sidney Santos, Laboratório de Genética Humana e Médica, Universidade Federal do Pará, Belém 66055-080, Pará, Brazil

Ândrea Ribeiro-dos-Santos, Sidney Santos, Núcleo de Pesquisas em Oncologia, Universidade Federal do Pará, Belém 66073-005, Pará, Brazil

Author contributions: Ribeiro-dos-Santos A, Santos S and Silbiger VN designed the research; Correa RS, Borges AMP and Ito FR selected patients and collected clinical data; Ramos CCO collected histopathological data; Marques D, Ferreira-Costa LR and Ferreira-Costa LL collected biological material and performed the assays; Marques D, Bortolin RH and Luchessi AD analyzed the data; Marques D wrote the paper; Bortolin RH, Silbiger VN and Ribeiro-dos-Santos A critically revised the

manuscript; Silbiger VN approved final version of the article to be published.

Supported by the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), No. 483031/2013-5; Rede de Pesquisa em Genômica Populacional Humana, No. Biocomputacional/CAPES-051/2013; Fundação de Amparo à Pesquisa do Estado do Pará, No. 155/2014; and Fundação de Amparo à Pesquisa do Estado do Rio Grande do Norte, No. 005/2011.

Institutional review board statement: This study was reviewed and approved by the Liga Norte Riograndense Contra o Câncer Institutional Review Board.

Informed consent statement: All study participants, or their legal guardian, provided informed written consent prior to study enrollment.

Conflict-of-interest statement: No benefits in any form have been received or will be received from a commercial party related directly or indirectly to the subject of this article.

Data sharing statement: Technical appendix, statistical code, and dataset are available from the corresponding author at viviansilbiger@hotmail.com; viviansilbiger@ufnet.br.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Unsolicited manuscript

Correspondence to: Vivian Nogueira Silbiger, PhD, Professor, Departamento de Análises Clínicas e Toxicológicas, Universidade Federal do Rio Grande do Norte, Av. General

Gustavo Cordeiro de Faria S/N, Petrópolis, Natal 59012-570, Rio Grande do Norte, Brazil. viviansilbiger@ufrnet.br
 Telephone: +55-84-33429807
 Fax: +55-84-33429833

Received: April 17, 2017
 Peer-review started: April 27, 2017
 First decision: June 5, 2017
 Revised: June 24, 2017
 Accepted: August 15, 2017
 Article in press: August 15, 2017
 Published online: October 7, 2017

Abstract

AIM

To investigate the association between 16 insertion-deletions (INDEL) polymorphisms, colorectal cancer (CRC) risk and clinical features in an admixed population.

METHODS

One hundred and forty patients with CRC and 140 cancer-free subjects were examined. Genomic DNA was extracted from peripheral blood samples. Polymorphisms and genomic ancestry distribution were assayed by Multiplex-PCR reaction, separated by capillary electrophoresis on the ABI 3130 Genetic Analyzer instrument and analyzed on GeneMapper ID v3.2. Clinicopathological data were obtained by consulting the patients' clinical charts, intra-operative documentation, and pathology scoring.

RESULTS

Logistic regression analysis showed that polymorphism variations in *IL4* gene was associated with increased CRC risk, while *TYMS* and *UCP2* genes were associated with decreased risk. Reference to anatomical localization of tumor Del allele of *NFKB1* and *CASP8* were associated with more colon related incidents than rectosigmoid. In relation to the INDEL association with tumor node metastasis (TNM) stage risk, the Ins alleles of *ACE*, *HLA*G and *TP53* (6 bp INDEL) were associated with higher TNM stage. Furthermore, regarding INDEL association with relapse risk, the Ins alleles of *ACE*, *HLA*G, and *UGT1A1* were associated with early relapse risk, as well as the Del allele of *TYMS*. Regarding INDEL association with death risk before 10 years, the Ins allele of *SGSM3* and *UGT1A1* were associated with death risk.

CONCLUSION

The INDEL variations in *ACE*, *UCP2*, *TYMS*, *IL4*, *NFKB1*, *CASP8*, *TP53*, *HLA*G, *UGT1A1*, and *SGSM3* were associated with CRC risk and clinical features in an admixed population. These data suggest that this cancer panel might be useful as a complementary tool for better clinical management, and more studies need to be conducted to confirm these findings.

Key words: Colorectal cancer; Ins-del polymorphism;

Admixed population; Potential biomarker; Diagnostic; Risk stratification; Prognostic; Clinical features

© **The Author(s) 2017.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: The insertion-deletions (INDEL) variations in *IL4* gene was associated with increased colorectal cancer (CRC) risk, while *TYMS* and *UCP2* genes were associated with decreased risk. The Del-alleles of *NFKB1* and *CASP8* were associated with more colon related incidents than rectosigmoid. The Ins-alleles of *ACE*, *HLA*G and *TP53* were associated with higher TNM stage. The Ins-allele of *ACE*, *HLA*G, and *UGT1A1* were associated with early relapse risk, as well as the Del-allele of *TYMS*. The Ins-alleles of *SGSM3* and *UGT1A1* were associated with death risk. These data suggest that these INDEL might be useful as a complementary tool for better CRC clinical management.

Marques D, Ferreira-Costa LR, Ferreira-Costa LL, Correa RS, Borges AMP, Ito FR, Ramos CCO, Bortolin RH, Luchessi AD, Ribeiro-dos-Santos A, Santos S, Silbiger VN. Association of insertion-deletions polymorphisms with colorectal cancer risk and clinical features. *World J Gastroenterol* 2017; 23(37): 6854-6867 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v23/i37/6854.htm> DOI: <http://dx.doi.org/10.3748/wjg.v23.i37.6854>

INTRODUCTION

Colorectal cancer (CRC) is the third most common cancer type in men and the second in women, considering 1477402 new cases in both sexes in 2015^[1]. In that same year, Brazil was the tenth country with the highest CRC incidence, with 37167 new cases in both sexes^[1], and making matters worse, its incidence and mortality continue to increase in the country.

Both genetic and environmental factors cause CRC^[2], especially when combined^[3,4]. Interestingly, these factors are ample and they vary pursuant to the cancer geographical regions^[5]. However, inherited susceptibility is a major component of CRC predisposition, with an estimated 12%-35% risk attributed to genetic factors^[6-8].

In relation to genetic factors, there are several mutations that might occur in human DNA, such as substitution, insertion, and deletion^[9]. The second most abundant form of genetic variation in humans, after single nucleotide polymorphisms (SNPs), are the insertion-deletions (INDEL)^[10]. INDELs are important because they are common genetic variations within genomes and among different ethnic groups^[11,12], that may alter human traits and cause diseases^[10,13], including CRC^[14], by modifying the coding region^[10,13] or mRNA stability^[15]. The polymorphisms investigated in this study exhibit common features, given they are

all functional polymorphisms that alter the expression of genes participating in metabolic pathways associated with carcinogenesis. Also, these genes are associated with different types of cancer with high incidence in the Brazilian population, such as stomach and CRC.

Furthermore, allele frequency varies among different populations^[16], and genomic ancestry distribution may influence cancer development^[17,18] by affecting polymorphisms distribution^[19,20]. Few studies have been evaluated INDEL association in CRC in admixed population, mainly in Brazil. Thus, the aim of this study was to determine the association between CRC risk and prognostic follow-up with 16 INDELs in genes involved in apoptosis signaling (*CASP8*), GTPase-activating (*SGSM3*), steroids metabolism (*CYP2E1*, *CYP19A1*, and *UGT1A1*), immune system (*HLA*, *IL1A*, *IL4*, and *NFKB1*), MDM2-P53 pathway (*MDM2* and *TP53*), DNA replication and repair (*TYMS* and *XRCC1*) and angiogenesis (*UCP2* and *ACE*) in an admixed population from Rio Grande do Norte state (in the Northeast Region of Brazil).

MATERIALS AND METHODS

A statement of ethics

The protocol used in this study was approved by the Research Ethics Committee of Liga Norte Riograndense Contra o Câncer (Rio Grande do Norte, Brazil) by number 211/211/2011. Moreover, all participants signed a consent form prior to providing a blood sample.

Casuistic distinctions

The patients in the case group ($n = 140$) were diagnosed with CRC as primary cancer and treated in the Proctologist Clinic and Colorectal Surgery Department of Liga Norte Riograndense Contra o Câncer (Rio Grande do Norte, Brazil). The control subjects were cancer-free blood donors ($n = 140$) from the hemotherapy service (Hemovida, Rio Grande do Norte, Brazil) and were recruited in 2014.

Both Peripheral blood samples and questionnaire answers were collected from all subjects. The clinicopathological data were obtained by consulting the patients' clinical charts, intra-operative documentation, and pathology scoring. Furthermore, the CRC patients were followed up to 20 years by medical records.

Definitions

Alcohol consumption was classified as having the habit of alcohol consumption (Yes) or not having the habit of alcohol consumption (No). The subjects who have the habit of consuming alcohol were subcategorized according to consumption frequency (Eventually: ≤ 3 d per month; Frequently: > 3 d per month). The tobacco consumption was classified as have already smoked (Already) or not (Never). The subjects who have already smoked were subcategorized in Former (Stopped smoking for at least 1 year) and Current.

Tumor location was classified as rectosigmoid (sigmoid colon and rectum) and colon (ascending, transverse and descending colon) based on colonoscopy or on radiographic exam. The relapse records were obtained from histological or radiographic exams with subsequent clinical/radiological progression.

Overall survival was defined as the time from the date of surgery to the date of death or the date of the last follow-up of patients who were still alive. The relapse time was defined as the time from the date of surgery to the date of the first local. Patients with no local or distant relapse evidence at the date of death or the date of the last follow-up were censored.

DNA extraction and quantification

Genomic DNA was extracted by using a DNA extraction commercial kit, QIAamp DNA Blood Mini Kit (Qiagen, Hilden, Germany), and quantified with Qubit® 2.0 Fluorometer (Invitrogen, Carlsbad, CA, United States).

Polymorphism selection

Recently, INDELs have been the focus of multiple investigations^[21-25]. This type of polymorphism presents interesting features as genetic markers: (1) INDELs are spread throughout the human genome; (2) INDELs derive from a single event (they do not present homoplasmy); (3) since the allele frequencies of many INDELs are significantly different in separated populations; (4) small INDELs can be analyzed using short amplicons, which improves the amplification of degraded DNA and facilitates multiplexing; and (5) INDELs can be easily genotyped with a simple dye-labeling electrophoretic approach. Furthermore, all these genes evaluated in the present study show potential activity in pathway and may contribute to the carcinogenesis process (Table 1). Their genetic variations could contribute to: (1) risk of developing CRC; (2) impact on treatment response; or (3) in prognosis.

Genotyping of polymorphism

Multiplex PCR was used to simultaneously amplify the 16 investigated markers, as shown in Supplementary Table 1. The amplification was performed on ABI Verity thermocycler (Life Technologies, Foster City, CA, United States). A single multiplex reaction used Master Mix QIAGEN Multiplex PCR kit (Qiagen, Hilden, Germany) according to the manufacturer's instructions. The samples were incubated at 95 °C for 15 min, followed by 35 cycles at 94 °C for 45 s, 60 °C for 90 s, and 72 °C for 1 min, with a final extension at 70 °C for 30 min.

For fragment analysis, we used capillary electrophoresis on the ABI 3130 Genetic Analyzer instrument (Life Technologies). 1.0 µL of PCR product was added to 8.5 µL of HI-DI deionized formamide (Life Technologies) and 0.5 µL of GeneScan 500 LIZ pattern size standard (Life Technologies). After data collection,

Table 1 Potential biological effects of insertion-deletions polymorphism selected in this study

Gene	dbSNP	Localization ¹	INDEL length	Region	Potential biological effect ^[84]				Potential impact on carcinogenesis
					mRNA splicing	mRNA stability	Gene expression	Protein function	
ACE	rs4646994	17:63488539	289	Intron	X	X		X	Angiogenesis, proliferation, progression and metastases ^[85]
CASP8	rs3834129	2:201232809	6	Promoter			X		Apoptosis ^[86-88]
SGSM3	rs56228771	22:40410092	4	3'-UTR		X	X		Proliferation and apoptosis ^[83,89-91]
CYP2E1	-	-	96	5'-Flanking			X		Metabolism of endo- and exogenous ^[92-97]
CYP19A1	rs11575899	15:51227749	3	Intron	X	X		X	Metabolism of endo- and exogenous ^[92-97]
HLA	rs371194629	6:29830804	14	3'-UTR		X	X		Immune surveillance ^[98-102]
IL1A	rs3783553	2:112774138	4	3'-UTR		X	X		Induce chronic inflammation and proliferation ^[103,104]
IL4	rs79071878	5:132680584	70	Intron	X	X		X	Immune surveillance and proliferation ^[40,39,43,105,106]
MDM2	rs3730485	12:68807065	40	Promoter			X		Proliferation and apoptosis ^[107,108]
NFKB1	rs28362491	4:102500998	4	Promoter			X		Differentiation, proliferation and apoptosis ^[109,110]
TP53	rs17878362	17:7676372	16	Intron	X	X		X	Proliferation, apoptosis, repair, differentiation ^[111-114]
TP53	rs17880560	17:7668169	6	3'-Flanking		X	X		Proliferation, apoptosis, repair, differentiation ^[111-114]
TYMS	rs151264360	18:673444	6	3'-UTR		X	X		Differentiation, replication and repair ^[50,105]
UCP2	-	-	45	3'-UTR		X	X		Tumor aggressiveness and metastasis ^[56]
UGT1A1	rs8175347	2:233760235	2	3'-UTR		X	X		Metabolism of endo- and exogenous ^[92-97]
XRCC1	rs3213239	19:43576907	4	5'- Flanking			X		Repair ^[115-117]

¹According to the single nucleotide polymorphism database (dbSNP); UTR: Untranslated region; INDEL: Insertion-deletions.

samples were analyzed on the GeneMapper ID v.3.7 software (Life Technologies).

Analysis of genetic ancestry

Genomic ancestry analysis was performed based on the method described by Santos *et al.*^[125] using 62 autosomal ancestry informative markers (AIMs). Two multiplex PCR reactions of 20 and 22 markers were performed and amplicons were analyzed by electrophoresis using the ABI Prism 3130 sequencer and GeneMapper ID v.3.2 software. The individual proportions of European, African, and Amerindian genetic ancestries were estimated using STRUCTURE v.2.3.3 software, assuming three parental populations (European, African, and Amerindian).

Statistical analysis

The categorical variables case and control participants were tested by the Chi-squared test. For ancestry index and age at diagnosis variables we used the Mann-Whitney test. Logistic regression analyses between the genotype model and CRC risk were performed by the SNPAssoc package v.1.9-2, along with clinical features variables. The association between genotype and free-relapse survival time was evaluated by Kaplan-Meier plots, performed by the survival package v.2.41-3. Log-rank and Wilcoxon tests were used to examine the genetic effect on survival outcomes. The statistical power was estimated by 10000 simulations. All statistical analyses and plotting were performed with R

package v.3.1.2^[26]. Differences between groups were considered significant at $P < 0.05$.

RESULTS

Demographic characteristics

We analyzed 140 subjects with CRC and 140 cancer-free individuals. The demographic characteristics of participants were summarized in Table 2, which shows demographic features of the groups. Regarding genomic ancestry, significance was observed with the distribution of African ancestry ($P = 0.049$), Table 3. However, there was no difference between groups when an analysis of multinomial logistic regression was performed.

Distribution of genotypes associated with susceptibility to CRC

All INDEL polymorphisms are in Hardy-Weinberg equilibrium ($P > 0.05$). The genotypic and allelic frequencies of the subjects are presented in the Table 4. Genotypic frequency ($P = 0.01$) of *IL4* gene polymorphism was significantly different between case-controls, and higher frequency of Del allele were observed in cases than in controls.

The significant logistic regression analyses between case-controls are summarized in Table 5. Del allele polymorphism in *IL4* gene ($P = 0.0110$) was associated with an increased risk of CRC development, while Ins allele in *UCP2* ($P = 0.0210$) was decreased

CRC risk. Furthermore, the Del allele in the *TYMS* ($P = 0.0120$) gene was associated with decreased CRC risk.

Distribution of genotypes associated with prognostic follow-up in CRC

The baseline characteristics of CRC patients are summarized in Table 6. The follow-up time median was 5.28 years among 78 patients who had complete genotype and follow-up information. The 5-year free-relapse rate was 70% and the 10-year free-relapse rate was 66.4%. The 5-year survival rate was 91.4% and the 10-year survival rate was 87.9%.

We also evaluated the genetic impact in the clinical features. The Del allele in *NFKB1* and *CASP8* were associated with more incidents to colon than rectosigmoid (Table 7). In relation to the INDEL association with TNM stage risk, the Ins alleles of *ACE*, *HLA*G and *TP53* (6 bp INDEL) were associated with a higher TNM stage (Table 8). Regarding the INDEL association with relapse risk, the Ins alleles of *ACE*, *HLA*G, and *UGT1A1* were associated with relapse risk, as well as the Del allele of *TYMS* (Table 9). Moreover, these findings corroborate those observed in the free-relapse survival curve (Figure 1). Regarding INDEL association with death risk, the Ins alleles of *SGSM3* and *UGT1A1* were associated with death risk (Table 10).

DISCUSSION

Despite the effective strategies for prevention, early detection, and treatment^[27-32], there are ethnic differences in the CRC incidence and survival^[33,34], specifically in individuals with African American ancestry, who have higher CRC incidence and lower 5-years survival rates than other ethnic groups^[33-38].

In this work, we evaluated the association between 16 INDEL [*ACE*, *CASP8*, *SGSM3*, *CYP19A1*, *CYP2E1*, *HLA*G, *IL1A*, *MDM2*, *NFKB1*, *TP53* (16 and 6 bp), *TYMS*, *UCP2*, *XRCC1*, *IL4* and *UGT1A1*] and the risk of developing CRC in a Brazilian population, as well as their clinical features. We found significant association between three investigated INDEL polymorphisms and CRC risk, two associated with anatomical localization, three associated with TNM stage, four associated with early relapse risk, and two associated with death risk before 10 years.

Variations in the IL-4 activity or in the IL-4 receptor due to mutations have been associated with cell proliferation and might affect signal transduction pathways in cancer^[39]. We evaluated INDEL of 70 bp in intron 3 of the *IL4* gene (rs79071878), a variation which may influence the production of this cytokine. The higher IL-4 production may result in diminished cell-mediated immune response, and escape from immune surveillance in the tumor cells. The cell-mediated immune response may be inhibited by downregulating the expression of Th1 cytokines, decreasing the CD8+ T-cell response in the tumor microenvironment^[39-41].

Furthermore, this INDEL has been associated with gastric cancer^[39] and other immune diseases^[42,43]. However, this is the first study indicating an association between this *IL4* polymorphism and the risk of developing CRC. Our results indicate that the Del allele in *IL4* was associated with the risk of developing CRC.

The *TYMS* gene plays an essential role in the biosynthesis of the DNA-component thymidylate (dTTP) and is required for DNA replication and repair^[44]. The insertion of 6 bp in the 3'-UTR of *TYMS* primary transcript (rs151264360) may significantly influence gene expression as shown by using a luciferase assay^[15]. Mandola *et al.*^[15] observed that a Del allele might decrease the *TYMS* mRNA stability, and the *TYMS* protein expression. Moreover, Rahman *et al.*^[45] showed in vitro that *TYMS* overexpression might induce the transformation of mammalian cells into a malignant phenotype. Studies indicate that this INDEL is associated with many cancers^[46-49], especially colorectal^[48]. These results suggested that this INDEL variation might decrease CRC risk, as showed in the present work. However, this finding diverges from data from Mexico^[50], in which association was not observed. On the other hand, our results showed that this Del allele was associated with an increase relapse risk.

Uncoupling proteins (UCPs) are a family of mitochondrial proteins, which were originally reported to play essential roles in reducing the reactive oxygen species^[51,52]. *UCP2* plays a role in carcinogenesis in various tissues, including colon cancer, and regulates the responsiveness of carcinomas to chemotherapy^[53-56]. Adaptive mechanisms of cancer cells include resistance to tumor growth inhibition and evasion of apoptosis, and cellular events that are appreciably affected by oxidative stress^[57,58]. The *UCP2* expression level is significantly higher in colon cancer tissue than in its adjacent tissue and *UCP2* may play a role in intestinal epithelial cells from benign to malignant transformation^[59]. However, the role of *UCP2* in development of colon cancer is unclear. INDEL polymorphism may regulate *UCP2* mRNA stability via post-transcriptional modification of *UCP2* protein expression^[60,61]. Indeed, in the present study was observed that INDEL polymorphism might be associated with colorectal cancer. However, this is the first study indicating an association between this *UCP2* polymorphism and the risk of developing CRC.

The renin-angiotensin system (RAS), which regulates systemic blood pressure, also exerts local effects on cell proliferation, apoptosis, inflammation and angiogenesis in different tissues^[62]. In addition, there is evidence linking the RAS with tumorigenesis and tumor angiogenesis^[63]. The polymorphisms in the various components of the RAS that may possess clinical relevance^[62], and the most common polymorphism in the gene encoding angiotensin converting enzyme (*ACE*) is INDEL of a 287-bp fragment in intron 16 and is responsible for the inter-individual variation in the

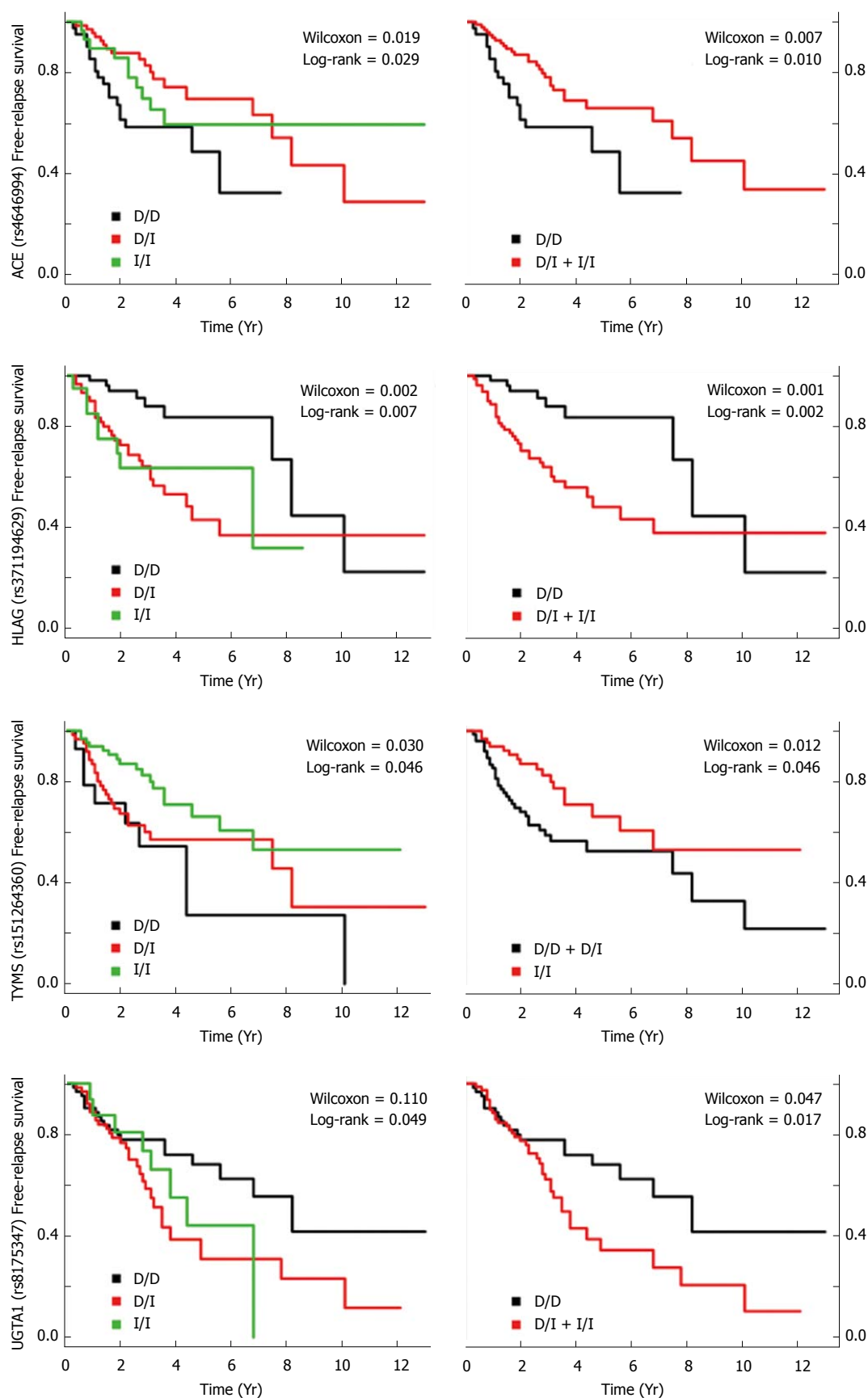


Figure 1 Free-relapse survival of patients with colorectal cancer related with significant insertion-deletions. Logistic regression adjusted for confounders. The analyses and graphic were performed by survival packages, in R statistical software.

Table 2 Participant demographic and clinical characteristics and their stratification by case and control groups

Characteristic	Total, <i>n</i> = 280	Cases, <i>n</i> = 140	Controls, <i>n</i> = 140	<i>P</i> value
Age (yr)	48 (21-93)	59 (23-93)	37 (21-81)	< 0.001
< 45	136 (48.7)	23 (16.5)	113 (80.7)	
≥ 45	144 (51.3)	116 (83.5)	27 (19.3)	
Gender				< 0.001
Male	172 (61.6)	62 (44.6)	110 (78.6)	
Female	108 (38.4)	78 (55.4)	30 (21.4)	
Alcohol consumption				< 0.001
No	180 (65.1)	118 (85.5)	62 (44.9)	
Yes	96 (34.9)	20 (14.5)	76 (55.1)	
Eventually	56 (20.4)	11 (8.0)	45 (32.6)	
Frequently	40 (14.5)	9 (6.5)	31 (22.5)	
Tobacco consumption				< 0.001
Never	182 (65.4)	68 (48.9)	114 (82.0)	
Already	96 (24.6)	71 (48.1)	25 (18.0)	
Former	66 (23.8)	55 (39.6)	11 (7.9)	
Current	30 (10.8)	16 (11.5)	14 (10.1)	

Categorized data are presented by absolute numbers of individuals (percentage) and analyzed by Chi-square test. Continuous data are presented by mean (min-max) and analyzed by Mann-Whitney test.

Table 3 Genetic ancestry distribution between case and control groups

Genetic ancestry (%)	Total, <i>n</i> = 280	Cases, <i>n</i> = 140	Controls, <i>n</i> = 140	OR (95%CI)	<i>P</i> value
European	65.3 ± 15.5	64.2 ± 15.6	66.4 ± 15.3		0.243
95-80	50 (17.9)	21 (15.1)	29 (20.7)	1.0 (Reference)	
80-70	70 (25.1)	33 (23.7)	37 (26.4)	1.23 (0.59-2.56)	0.577
70-60	67 (24.0)	38 (27.3)	29 (20.7)	1.81 (0.86-3.80)	0.117
60-50	47 (16.8)	22 (15.8)	25 (17.9)	1.21 (0.54-2.71)	0.634
50-40	26 (9.3)	14 (10.1)	12 (8.6)	1.61 (0.62-4.18)	0.327
40-30	11 (3.9)	6 (4.3)	5 (3.6)	1.66 (0.45-6.16)	0.451
30-20	7 (2.5)	5 (3.6)	2 (1.4)	3.45 (0.61-19.54)	0.161
20-10	1 (0.4)	-	1 (0.7)	-	
Amerindian	16.2 ± 10.1	16.0 ± 10.3	16.3 ± 9.9		0.645
02-10	90 (32.3)	45 (32.4)	45 (32.1)	1.0 (Reference)	
10-20	113 (40.5)	60 (43.2)	53 (37.9)	1.13 (0.65-1.97)	0.661
20-30	47 (16.8)	18 (12.9)	29 (20.7)	0.62 (0.30-1.27)	0.193
30-40	21 (7.5)	11 (7.9)	10 (7.1)	1.10 (0.42-2.85)	0.844
40-50	7 (2.5)	4 (2.9)	2 (2.1)	2.00 (0.35-11.47)	0.437
50-60	1 (0.4)	1 (0.7)	-	-	
African	18.6 ± 12.0	19.8 ± 12.3	17.3 ± 11.7		0.049
02-10	81 (29.0)	36 (25.9)	45 (32.1)	1.0 (Reference)	
10-20	89 (31.9)	41 (29.5)	48 (34.3)	1.07 (0.58-1.95)	0.832
20-30	66 (23.7)	38 (27.3)	28 (20.0)	1.70 (0.88-3.27)	0.114
30-40	28 (10.0)	15 (10.8)	13 (9.3)	1.44 (0.61-3.42)	0.405
40-50	8 (2.9)	5 (3.6)	3 (2.1)	2.08 (0.47-9.31)	0.337
50-60	5 (1.8)	3 (2.2)	2 (1.4)	1.87 (0.30-11.83)	0.504
60-70	2 (0.7)	1 (0.7)	1 (0.7)	1.25 (0.07-20.68)	0.876

Categorized data are presented by absolute numbers of individuals (percentage) and analyzed by Chi-square test. Continuous data are presented by mean ± standard variation and analyzed by Mann-Whitney test.

ACE levels in blood and tissues^[64]. The insertion allele in this gene was associated with ACE levels, the rate of disease progression, shorter TTF, and lower circulating levels of ACE^[62,65]. This INDEL has been associated with cancer risk susceptibility^[66-68], including CRC^[65,68], and with response to bevacizumab^[62]. Our results indicate that Ins allele was not associated with CRC risk development, as showed by Yang *et al*'s meta-analysis^[69] and Liu *et al*'^[70] case-control study (241 cases and 299 control, China). On the other hand, our results also showed that this INDEL was also associated

with TNM stage risk and relapse risk.

The *HLA-G* is an important immunomodulatory molecule related to several mechanisms of tolerance^[71]. Since the discovery of the HLA-G protein expression in cancer^[72], several pieces of evidence have supported a considerable role for HLA-G in tumor cell escape from immuno-surveillance and antitumor immune responses^[73]. The 14 bp INDEL (rs371194629) has been suggested to have functional significance. The Ins allele has been shown to be associated with alternative splicing, resulting in deletion of 92 bp in

Table 4 Genotype and allele frequency in percentage of patients with colorectal cancer and controls

Gene	dbSNP	Case/control (n = 140/140)						
		Genotype frequency				Allele frequency		HWE
		II	ID	DD	P value	I	D	P value
ACE	rs4646994	21.0/14.3	49.3/54.3	29.7/31.4	0.335	45.7/41.4	54.3/58.6	0.161
CASP8	rs3834129	34.5/30.0	46.0/46.4	19.4/23.6	0.605	57.6/53.2	42.4/46.8	0.424
SGSM3	rs56228771	7.2/3.6	30.2/36.4	62.6/60.0	0.274	22.3/21.8	77.7/78.2	0.415
CYP19A1	rs11575899	35.8/37.9	49.6/50.0	14.6/12.1	0.82	60.3/62.9	39.7/37.1	0.402
CYP2E1	-	0.7/0.7	17.3/12.9	82.0/86.4	0.588	9.4/7.1	90.6/92.6	0.716
HLA-G	rs371194629	14.7/13.6	43.4/50.0	41.9/36.4	0.538	36.4/38.6	63.6/61.4	0.514
IL1A	rs3783553	46.0/52.1	38.8/37.9	15.1/10.0	0.368	65.5/71.1	34.5/28.9	0.348
IL4	rs79071878	48.9/60.0	40.3/37.1	10.8/2.9	0.017	69.1/78.6	30.9/21.4	0.223
MDM2	rs3730485	50.4/50.7	43.2/37.1	6.5/12.1	0.219	71.9/69.3	28.1/30.7	0.132
NFKB1	rs28362491	38.1/40.7	46.0/42.1	15.8/17.1	0.806	61.2/61.8	38.8/38.2	0.203
TP53	rs17878362	3.6/1.4	27.3/32.1	69.1/66.4	0.383	17.3/17.5	82.7/82.5	0.181
TP53	rs17880560	7.2/7.1	39.6/32.9	53.2/60.0	0.489	27.0/23.6	73.0/76.4	0.297
TYMS	rs151264360	46.0/42.9	43.9/42.1	10.1/15.0	0.459	68.0/63.9	32.0/36.1	0.308
UCP2	-	6.6/9.3	35.3/44.3	58.1/46.4	0.149	24.3/31.4	75.7/68.6	0.745
UGT1A1	rs8175347	11.5/10.8	44.6/46.0	43.9/43.2	0.785	33.8/33.8	66.2/66.2	0.735
XRCC1	rs3213239	41.7/42.9	47.5/45.0	10.8/12.1	0.894	65.5/65.4	34.5/34.6	0.941

Genotype frequencies are presented as the percentage of patients with colorectal cancer/percentage of controls, and analysis by Chi-square test. dbSNP: Register of genetic variation on NCBI database; bp: Base pairs of DNA sequence; HWE: Hardy-Weinberg Equilibrium.

Table 5 The logistic regression analyses between case-control and insertion-deletions polymorphism

Gene	Model	OR (95%CI)	P-value
IL4	Ins/Ins vs Del/Ins + Del/Del	2.26 (1.20-4.31)	0.0110
TYMS	Ins/Ins + Del/Ins vs Del/Del	0.26 (0.08-0.75)	0.0120
UCP2	Del/Del vs Del/Ins + Ins/Ins	0.48 (0.25-0.90)	0.0210

Adjusted by age at diagnosis, gender, alcohol consumption, tobacco consumption and ancestry distribution. The Supplementary Table 2 shows the genotype frequency and all logistic regression.

Table 7 The significant insertion-deletions associations with anatomic localization

Gene	Model	OR (95%CI)	P value
CASP8	Ins/Ins vs Del/Ins + Del/Del	0.28 (0.08-0.97)	0.0303
NFKB1	Ins/Ins vs Del/Ins+Del/Del	0.31 (0.10-0.93)	0.0276

Logistic regression adjusted for confounders. The Supplementary Table 3 shows the genotype frequency and all logistic regression.

Table 8 The significant insertion-deletions associations with tumor node metastasis stage risks

Gene	Model	OR (95%CI)	P value
ACE	Del/Del vs Del/Ins + Ins/Ins	2.82 (1.26-6.31)	0.0092
HLA-G	Del/Del + Del/Ins vs Ins/Ins	2.74 (1.01-7.42)	0.0416
TP53 06 bp	Del/Del vs Del/Ins + Ins/Ins	2.50 (1.23-5.06)	0.0099

Logistic regression adjusted for confounders. The Supplementary Table 3 shows the genotype frequency and all logistic regression.

exon 5 from mature mRNA, which then leads to low levels of soluble HLA-G (sHLA-G)^[74]. Furthermore, our results indicated that the Ins allele was associated with a higher TNM stage and relapse up to 5 years.

Table 6 Clinical characteristics of patients with colorectal cancer at diagnosis and follow-up

Characteristics	Cases (n = 140)
Tumor localization	
Colon	25 (17.9)
Rectosigmoid	115 (82.1)
Tumor grade	
G1, G2	130 (92.9)
G3, G4	10 (7.1)
Depth of invasion	
T1, T2	37 (26.6)
T3, T4	89 (64.0)
Tx	13 (9.4)
Lymph node involvement	
N0	77 (55.4)
N1, N2	47 (33.8)
Nx	15 (10.8)
Distant metastasis	
M0	109 (78.4)
M1	13 (9.4)
Mx	17 (12.2)
AJCC stage	
Stage I	31 (22.3)
Stage II	43 (30.9)
Stage III	43 (30.9)
Stage IV	15 (10.8)
Unknown	7 (5.1)
Relapse, Yes	48 (34.5)
Death, Yes	18 (12.9)

Categorized data are presented by absolute numbers (percentage) and continuous data are presented as median (min-max). Tumors were classified according to the guidelines of the American Joint Committee on Cancer (AJCC) staging system.

These findings suggest that low levels of sHLA-G might influence in poor prognostics.

UGT1 is a family of membrane-bound enzymes involved in the inactivation and elimination of lipophilic molecules through glucorination. Moreover, variants

Table 9 The significant insertion-deletions associations with relapse risks

Gene	Model	Time of follow-up	OR (95%CI)	P value
ACE	Del/Del <i>vs</i> Del/Ins + Ins/Ins	2 yr	0.32 (0.13-0.77)	0.0113
ACE	Del/Del <i>vs</i> Del/Ins + Ins/Ins	3 yr	0.37 (0.15-0.91)	0.0298
HLA	Del/Del <i>vs</i> Del/Ins + Ins/Ins	2 yr	2.75 (1.07-7.08)	0.0281
HLA	Del/Del <i>vs</i> Del/Ins + Ins/Ins	4 yr	2.83 (1.07-7.52)	0.0332
HLA	Del/Del <i>vs</i> Del/Ins + Ins/Ins	5 yr	3.47 (1.20-9.99)	0.0194
TYMS	Ins/Ins <i>vs</i> Del/Ins + Del/Del	2 yr	3.35 (1.36-8.28)	0.0058
TYMS	Ins/Ins <i>vs</i> Del/Ins + Del/Del	3 yr	3.42 (1.41-8.28)	0.0046
UGT1A1	Del/Del <i>vs</i> Del/Ins + Ins/Ins	4 yr	3.23 (1.27-8.22)	0.0116
UGT1A1	Del/Del <i>vs</i> Del/Ins + Ins/Ins	5 yr	3.50 (1.24-9.84)	0.0145

Logistic regression adjusted for confounders. The Supplementary Table 4 shows the genotype frequency of all insertion-deletions polymorphism.

Table 10 The significant insertion-deletions associations with death risks

Gene	Model	Time of follow-up	OR (95%CI)	P value
SGSM3	Del/Del <i>vs</i> Del/Ins + Ins/Ins	6 yr	3.61 (1.01-12.92)	0.0487
SGSM3	Del/Del <i>vs</i> Del/Ins + Ins/Ins	7 yr	4.60 (1.16-18.23)	0.0260
UGT1A1	Del/Del <i>vs</i> Del/Ins + Ins/Ins	6 yr	5.30 (1.43-19.73)	0.0084
UGT1A1	Del/Del <i>vs</i> Del/Ins + Ins/Ins	7 yr	4.64 (1.19-18.10)	0.0202
UGT1A1	Del/Del <i>vs</i> Del/Ins + Ins/Ins	8 yr	6.50 (1.47-28.80)	0.0091

Logistic regression adjusted for confounders. The Supplementary Table 5 shows the genotype frequency of all insertion-deletions polymorphism.

in this gene have been shown to be useful tools to identifying patients more likely to experience severe toxicity related to irinotecan-containing regimens^[75]. In particular, INDEL variants in *UGT1A1* (rs8175347) were associated with significantly decreased glucuronidation activity, which results in reduced SN-38 clearance^[76] and an increased risk of these toxicities in patients homozygous for the Ins allele^[75,77-80]. Our results showed that the Ins allele in *UGT1A1* was associated with early relapse risk, as well as with death risk prior to 8 years. This genetic variation may identify patients who might benefit from increased irinotecan dosing, as observed by Chen *et al.*^[75].

The *SGSM3* belongs to a novel protein family consisting of three members and appears to be associated with small G-protein coupled receptor signal transduction pathways, and could control cellular functions by a Ras-mediated signaling pathway^[81]. Studies have linked Rab dysfunction to various human diseases including cancer^[82,83], and our results have shown that the Ins allele might also be associated with death risk prior to 8 years.

The aims of this study were to determine the association between CRC risk and the clinical features with 16 INDEL in genes involved with carcinogenesis pathways in an admixed population from Brazil. Although we have achieved our goal, there are limitations regarding sample number. We suggest, therefore, that an extensive study should be conducted in the Brazilian population to confirm the findings, as well as in other admixed populations.

In summary, the present work indicates that polymorphisms in *ACE* (rs4646994), *TYMS*

(rs151264360), *UCP2* (45 bp), *IL4* (rs79071878), *NFKB1* (rs28362491), *CASP8* (rs3834129), *TP53* (rs17880560), *HLA* (rs371194629), *UGT1A1* (rs3213239), and *SGSM3* (rs56228771) genes were associated with CRC risk and clinical features in an admixed population. These data suggest that this cancer panel might be useful as a complementary tool for better clinical management, and more studies need to be conducted to confirm these findings.

ACKNOWLEDGMENTS

We thank all participants who allowed us to carry out this study. We are also grateful to the students and technicians from LGHM/UFGA/PA, LBBM-LABMULT/UFRN/RN, Laboratory of Pathology and Cytopathology/Liga Norte Riograndense Contra o Câncer/RN and Hemovida/RN. We are also grateful to Philip Barsanti for the invaluable and constructive English review of the manuscript. Furthermore, we are grateful to André Ribeiro-dos-Santos and Ana Paula Schaan for help with statistical revision.

COMMENTS

Background

Colorectal cancer (CRC) is the third most common cancer type in men and the second in women. Despite the effective strategies for prevention, early detection, and treatment, there are ethnic differences in CRC incidence and survival. These variances occur specifically in African Americans, who have higher CRC incidence and lower survival rates than other ethnic groups. Thus, the present study evaluated the association between 16 insertion-deletions (INDEL) polymorphisms with colorectal cancer risk in an admixture population, as well with clinical features.

Research frontiers

The second most abundant form of genetic variation in humans, after single nucleotide polymorphisms (SNPs), are the INDEL. The INDEL understanding is important because they are common genetic variations within genomes, and they may alter human traits and cause diseases, including colorectal cancer, by modifying the coding region or mRNA stability. One of the challenges for genetic polymorphism association studies is the lack of knowledge regarding the frequency of the polymorphism in the targeted population, mainly in admixed populations (e.g. Brazil).

Innovations and breakthroughs

This is the first case-control study to evaluate the association between these 16 INDEL polymorphisms with colorectal risk, clinical features and prognostic follow-up in an admixture population, adopting the methodology that can be easily used to perform multiplexing assays.

Applications

This pilot study is design and findings could be used to determine sample size for a larger randomized controlled study aiming to test the impact of these INDEL polymorphism panel in colorectal risk, clinical features and prognostic follow-up.

Terminology

Ancestry informative marker - In population genetics, an ancestry informative marker (AIM) is a polymorphism that exhibits substantially different frequencies between populations from different geographical regions. A set of many AIMS can be used to estimate the proportion of ancestry of an individual derived from each geographical region.

Peer-review

This is an interesting study aiming to determine the association between CRC risk, and the clinical features with 16 INDEL in genes involved with carcinogenesis pathways in an admixed population from Brazil. The overall structure of the manuscript is complete and conforms to the academic rules.

REFERENCES

- 1 Ferlay J, Soerjomataram I, Ervik M, Dikshit R, Eser S, Mathers C, Rebelo M, Parkin DM, Forman D, Bray F. GLOBOCAN 2012 v1.0, Cancer Incidence and Mortality Worldwide: IARC CancerBase No. 11 [Internet]. Lyon, France: International Agency for Research on Cancer; 2013. Accessed on 2016-2-12 Available from: URL: <http://globocan.iarc.fr/>
- 2 Hong Y, Wu G, Li W, Liu D, He K. A comprehensive meta-analysis of genetic associations between five key SNPs and colorectal cancer risk. *Oncotarget* 2016; **7**: 73945-73959 [PMID: 27661122 DOI: 10.18632/oncotarget.12154]
- 3 Zaridze DG. Molecular epidemiology of cancer. *Biochemistry* (Moscow) 2008; **73**: 532-542 [PMID: 18605978 DOI: 10.1134/S0006297908050064]
- 4 Liu J, He C, Xu Q, Xing C, Yuan Y. NOD2 polymorphisms associated with cancer risk: a meta-analysis. *PLoS One* 2014; **9**: e89340 [PMID: 24586700 DOI: 10.1371/journal.pone.0089340]
- 5 Gomez SL, Shariff-Marco S, DeRouen M, Keegan TH, Yen IH, Mujahid M, Satariano WA, Glaser SL. The impact of neighborhood social and built environment factors across the cancer continuum: Current research, methodological considerations, and future directions. *Cancer* 2015; **121**: 2314-2330 [PMID: 25847484 DOI: 10.1002/cncr.29345]
- 6 Peters U, Bien S, Zubair N. Genetic architecture of colorectal cancer. *Gut* 2015; **64**: 1623-1636 [PMID: 26187503 DOI: 10.1136/gutjnl-2013-306705]
- 7 Lichtenstein P, Holm NV, Verkasalo PK, Iliadou A, Kaprio J, Koskenvuo M, Pukkala E, Skytthe A, Hemminki K. Environmental and heritable factors in the causation of cancer--analyses of cohorts of twins from Sweden, Denmark, and Finland. *N Engl J Med* 2000; **343**: 78-85 [PMID: 10891514 DOI: 10.1056/NEJM200007133430201]
- 8 Czene K, Lichtenstein P, Hemminki K. Environmental and heritable causes of cancer among 9.6 million individuals in the Swedish Family-Cancer Database. *Int J Cancer* 2002; **99**: 260-266 [PMID: 11979442 DOI: 10.1002/ijc.10332]
- 9 Iengar P. An analysis of substitution, deletion and insertion mutations in cancer genes. *Nucleic Acids Res* 2012; **40**: 6401-6413 [PMID: 22492711 DOI: 10.1093/nar/gks290]
- 10 Mullaney JM, Mills RE, Pittard WS, Devine SE. Small insertions and deletions (INDELs) in human genomes. *Hum Mol Genet* 2010; **19**: R131-R136 [PMID: 20858594 DOI: 10.1093/hmg/ddq400]
- 11 Mills RE, Pittard WS, Mullaney JM, Farooq U, Creasy TH, Mahurkar AA, Kemeza DM, Strassler DS, Ponting CP, Webber C, Devine SE. Natural genetic variation caused by small insertions and deletions in the human genome. *Genome Res* 2011; **21**: 830-839 [PMID: 21460062 DOI: 10.1101/gr.115907.110]
- 12 Boschiero C, Gheyas AA, Ralph HK, Eory L, Paton B, Kuo R, Fulton J, Preisinger R, Kaiser P, Burt DW. Detection and characterization of small insertion and deletion genetic variants in modern layer chicken genomes. *BMC Genomics* 2015; **16**: 562 [PMID: 26227840 DOI: 10.1186/s12864-015-1711-1]
- 13 Veerappa AM, Vishweswaraiah S, Lingaiah K, Murthy NM, Suresh RV, Belur K, Ramachandra NB, Tejaswini, Patel NB, Gowda PK. Insertion-deletions burden in copy number polymorphisms of the Tibetan population. *Indian J Hum Genet* 2014; **20**: 166-174 [PMID: 25400346 DOI: 10.4103/0971-6866.142888]
- 14 Gao X, Zhang S, Zhu Z. Genetic variation of ErbB4 confers risk of colorectal cancer in a Chinese Han population. *Cancer Biomark* 2014; **14**: 435-439 [PMID: 25335735 DOI: 10.3233/CBM-140420]
- 15 Mandola MV, Stoecklacher J, Zhang W, Groshen S, Yu MC, Iqbal S, Lenz HJ, Ladner RD. A 6 bp polymorphism in the thymidylate synthase gene causes message instability and is associated with decreased intratumoral TS mRNA levels. *Pharmacogenetics* 2004; **14**: 319-327 [PMID: 15115918]
- 16 Hu Z, Li X, Qu X, He Y, Ring BZ, Song E, Su L. Intron 3 16 bp duplication polymorphism of TP53 contributes to cancer susceptibility: a meta-analysis. *Carcinogenesis* 2010; **31**: 643-647 [PMID: 20089604 DOI: 10.1093/carcin/bgq018]
- 17 Hernandez-Suarez G, Sanabria MC, Serrano M, Herran OF, Perez J, Plata JL, Zabaleta J, Tenesa A. Genetic ancestry is associated with colorectal adenomas and adenocarcinomas in Latino populations. *Eur J Hum Genet* 2014; **22**: 1208-1216 [PMID: 24518838 DOI: 10.1038/ejhg.2013.310]
- 18 Carvalho DC, Wanderley AV, Amador MA, Fernandes MR, Cavalcante GC, Pantoja KB, Mello FA, de Assumpção PP, Khayat AS, Ribeiro-Dos-Santos A, Santos S, Dos Santos NP. Amerindian genetic ancestry and INDEL polymorphisms associated with susceptibility of childhood B-cell Leukemia in an admixed population from the Brazilian Amazon. *Leuk Res* 2015; **39**: 1239-1245 [PMID: 26321572 DOI: 10.1016/j.leukres.2015.08.008]
- 19 Cassiano GC, Santos EJ, Maia MH, Furini Ada C, Storti-Melo LM, Tomaz FM, Trindade PC, Capobianco MP, Amador MA, Viana GM, Póvoa MM, Santos SE, Machado RL. Impact of population admixture on the distribution of immune response co-stimulatory genes polymorphisms in a Brazilian population. *Hum Immunol* 2015; **76**: 836-842 [PMID: 26429313 DOI: 10.1016/j.humimm.2015.09.045]
- 20 Campanella NC, Berardinelli GN, Scapulatempo-Neto C, Viana D, Palmero EI, Pereira R, Reis RM. Optimization of a pentaplex panel for MSI analysis without control DNA in a Brazilian population: correlation with ancestry markers. *Eur J Hum Genet* 2014; **22**: 875-880 [PMID: 24193342 DOI: 10.1038/ejhg.2013.256]
- 21 Bastos-Rodrigues L, Pimenta JR, Pena SD. The genetic structure of human populations studied through short insertion-deletion polymorphisms. *Ann Hum Genet* 2006; **70**: 658-665 [PMID: 16907710 DOI: 10.1111/j.1469-1809.2006.00287.x]
- 22 Mills RE, Luttig CT, Larkins CE, Beauchamp A, Tsui C, Pittard WS, Devine SE. An initial map of insertion and deletion (INDEL) variation in the human genome. *Genome Res* 2006; **16**: 1182-1190 [PMID: 16902084 DOI: 10.1101/gr.4565806]

- 23 **Ribeiro-Rodrigues EM**, dos Santos NP, dos Santos AK, Pereira R, Amorim A, Gusmão L, Zago MA, dos Santos SE. Assessing interethnic admixture using an X-linked insertion-deletion multiplex. *Am J Hum Biol* 2009; **21**: 707-709 [PMID: 19533621 DOI: 10.1002/ajhb.20950]
- 24 **Weber JL**, David D, Heil J, Fan Y, Zhao C, Marth G. Human diallelic insertion/deletion polymorphisms. *Am J Hum Genet* 2002; **71**: 854-862 [PMID: 12205564 DOI: 10.1086/342727]
- 25 **Santos NP**, Ribeiro-Rodrigues EM, Ribeiro-Dos-Santos AK, Pereira R, Gusmão L, Amorim A, Guerreiro JF, Zago MA, Matte C, Hutz MH, Santos SE. Assessing individual interethnic admixture and population substructure using a 48-insertion-deletion (INSEL) ancestry-informative marker (AIM) panel. *Hum Mutat* 2010; **31**: 184-190 [PMID: 19953531 DOI: 10.1002/humu.21159]
- 26 **R Development Core Team**. R: A language and environment for statistical computing. R Found Stat Comput, Vienna, Austria. Available from: URL: <http://www.r-project.org/>
- 27 **Schoen RE**, Pinsky PF, Weissfeld JL, Yokochi LA, Church T, Laiyemo AO, Bresalier R, Andriole GL, Buys SS, Crawford ED, Fouad MN, Isaacs C, Johnson CC, Reding DJ, O'Brien B, Carrick DM, Wright P, Riley TL, Purdue MP, Izmirlian G, Kramer BS, Miller AB, Gohagan JK, Prorok PC, Berg CD; PLCO Project Team. Colorectal-cancer incidence and mortality with screening flexible sigmoidoscopy. *N Engl J Med* 2012; **366**: 2345-2357 [PMID: 22612596 DOI: 10.1056/NEJMoa1114635]
- 28 **Mandel JS**, Bond JH, Church TR, Snover DC, Bradley GM, Schuman LM, Ederer F. Reducing mortality from colorectal cancer by screening for fecal occult blood. Minnesota Colon Cancer Control Study. *N Engl J Med* 1993; **328**: 1365-1371 [PMID: 8474513 DOI: 10.1056/NEJM199305133281901]
- 29 **Nishihara R**, Wu K, Lochhead P, Morikawa T, Liao X, Qian ZR, Inamura K, Kim SA, Kuchiba A, Yamauchi M, Imamura Y, Willett WC, Rosner BA, Fuchs CS, Giovannucci E, Ogino S, Chan AT. Long-term colorectal-cancer incidence and mortality after lower endoscopy. *N Engl J Med* 2013; **369**: 1095-1105 [PMID: 24047059 DOI: 10.1056/NEJMoa1301969]
- 30 **US Preventive Services Task Force**. Screening for colorectal cancer: U.S. Preventive Services Task Force recommendation statement. *Ann Intern Med* 2008; **149**: 627-637 [PMID: 18838716 DOI: 10.7326/0003-4819-149-9-200811040-00243]
- 31 **Scambler G**, Hopkins A. Generating a model of epileptic stigma: the role of qualitative analysis. *Soc Sci Med* 1990; **30**: 1187-1194 [PMID: 2360054 DOI: 10.7326/0003-4819-158-5-201303050-00003]
- 32 **Atkin WS**, Edwards R, Kralj-Hans I, Wooldrage K, Hart AR, Northover JM, Parkin DM, Wardle J, Duffy SW, Cuzick J; UK Flexible Sigmoidoscopy Trial Investigators. Once-only flexible sigmoidoscopy screening in prevention of colorectal cancer: a multicentre randomised controlled trial. *Lancet* 2010; **375**: 1624-1633 [PMID: 20430429 DOI: 10.1016/S0140-6736(10)60551-X]
- 33 **Soneji S**, Iyer SS, Armstrong K, Asch DA. Racial disparities in stage-specific colorectal cancer mortality: 1960-2005. *Am J Public Health* 2010; **100**: 1912-1916 [PMID: 20724684 DOI: 10.2105/AJPH.2009.184192]
- 34 **Robbins AS**, Siegel RL, Jemal A. Racial disparities in stage-specific colorectal cancer mortality rates from 1985 to 2008. *J Clin Oncol* 2012; **30**: 401-405 [PMID: 22184373 DOI: 10.1200/JCO.2011.37.5527]
- 35 **Wu BU**, Longstreth GF, Ngor EW. Screening colonoscopy versus sigmoidoscopy: implications of a negative examination for cancer prevention and racial disparities in average-risk patients. *Gastrointest Endosc* 2014; **80**: 852-861.e1-2 [PMID: 24814774 DOI: 10.1016/j.gie.2014.03.015]
- 36 **Corley DA**, Jensen CD, Marks AR, Zhao WK, de Boer J, Levin TR, Doubeni C, Fireman BH, Quesenberry CP. Variation of adenoma prevalence by age, sex, race, and colon location in a large population: implications for screening and quality programs. *Clin Gastroenterol Hepatol* 2013; **11**: 172-180 [PMID: 22985608 DOI: 10.1016/j.cgh.2012.09.010]
- 37 **Lieberman DA**, Holub JL, Moravec MD, Eisen GM, Peters D, Morris CD. Prevalence of colon polyps detected by colonoscopy screening in asymptomatic black and white patients. *JAMA* 2008; **300**: 1417-1422 [PMID: 18812532 DOI: 10.1001/jama.300.12.1417]
- 38 **Kupfer SS**, Anderson JR, Hooker S, Skol A, Kittles RA, Keku TO, Sandler RS, Ellis NA. Genetic heterogeneity in colorectal cancer associations between African and European Americans. *Gastroenterology* 2010; **139**: 1677-1685, 1685.e1-1685.e8 [PMID: 20659471 DOI: 10.1053/j.gastro.2010.07.038]
- 39 **Bhayal AC**, Krishnaveni D, Rao KP, Kumar AR, Jyothy A, Nallari P, Venkateshwari A. Significant Association of Interleukin4 Intron 3 VNTR Polymorphism with Susceptibility to Gastric Cancer in a South Indian Population from Telangana. *PLoS One* 2015; **10**: e0138442 [PMID: 26383107 DOI: 10.1371/journal.pone.0138442]
- 40 **Nakashima H**, Miyake K, Inoue Y, Shimizu S, Akahoshi M, Tanaka Y, Otsuka T, Harada M. Association between IL-4 genotype and IL-4 production in the Japanese population. *Genes Immun* 2002; **3**: 107-109 [PMID: 11960309 DOI: 10.1038/sj.gene.6363830]
- 41 **Gaur P**, Singh AK, Shukla NK, Das SN. Inter-relation of Th1, Th2, Th17 and Treg cytokines in oral cancer patients and their clinical significance. *Hum Immunol* 2014; **75**: 330-337 [PMID: 24486578 DOI: 10.1016/j.humimm.2014.01.011]
- 42 **Anovazzi G**, Kim YJ, Viana AC, Curtis KM, Orrico SR, Cirelli JA, Scarel-Caminaga RM. Polymorphisms and haplotypes in the interleukin-4 gene are associated with chronic periodontitis in a Brazilian population. *J Periodontol* 2010; **81**: 392-402 [PMID: 20192866 DOI: 10.1902/jop.2009.090392]
- 43 **Cabantous S**, Poudiougou B, Oumar AA, Traore A, Barry A, Vitte J, Bongrand P, Marquet S, Doumbo O, Dessein AJ. Genetic evidence for the aggravation of Plasmodium falciparum malaria by interleukin 4. *J Infect Dis* 2009; **200**: 1530-1539 [PMID: 19835477 DOI: 10.1086/644600]
- 44 **Burdelski C**, Strauss C, Tsourlakis MC, Kluth M, Hube-Magg C, Melling N, Lebok P, Minner S, Koop C, Graefen M, Heinzer H, Wittmer C, Krech T, Sauter G, Wilczak W, Simon R, Schlomm T, Steurer S. Overexpression of thymidylate synthase (TYMS) is associated with aggressive tumor features and early PSA recurrence in prostate cancer. *Oncotarget* 2015; **6**: 8377-8387 [PMID: 25762627 DOI: 10.18632/oncotarget.3107]
- 45 **Rahman L**, Voeller D, Rahman M, Lipkowitz S, Allegra C, Barrett JC, Kaye FJ, Zajac-Kaye M. Thymidylate synthase as an oncogene: a novel role for an essential DNA synthesis enzyme. *Cancer Cell* 2004; **5**: 341-351 [PMID: 15093541 DOI: 10.1016/S1535-6108(04)00080-7]
- 46 **Guan X**, Liu H, Ju J, Li Y, Li P, Wang LE, Brewster AM, Buchholz TA, Arun BK, Wei Q, Liu Z. Genetic variant rs16430 6bp > 0bp at the microRNA-binding site in TYMS and risk of sporadic breast cancer risk in non-Hispanic white women aged ≤ 55 years. *Mol Carcinog* 2015; **54**: 281-290 [PMID: 24166930 DOI: 10.1002/mc.22097]
- 47 **Shen R**, Liu H, Wen J, Liu Z, Wang LE, Wang Q, Tan D, Ajani JA, Wei Q. Genetic polymorphisms in the microRNA binding-sites of the thymidylate synthase gene predict risk and survival in gastric cancer. *Mol Carcinog* 2015; **54**: 880-888 [PMID: 24756984 DOI: 10.1002/mc.22160]
- 48 **Zhou JY**, Shi R, Yu HL, Zeng Y, Zheng WL, Ma WL. The association between two polymorphisms in the TS gene and risk of cancer: a systematic review and pooled analysis. *Int J Cancer* 2012; **131**: 2103-2116 [PMID: 22307944 DOI: 10.1002/ijc.27465]
- 49 **Shaw GM**, Yang W, Perloff S, Shaw NM, Carmichael SL, Zhu H, Lammer EJ. Thymidylate synthase polymorphisms and risks of human orofacial clefts. *Birth Defects Res A Clin Mol Teratol* 2013; **97**: 95-100 [PMID: 23404871 DOI: 10.1002/bdra.23114]
- 50 **Gallegos-Arreola MP**, Peralta-Leal V, Morgan-Villela G, Puebla-Pérez AM. [Frequency of TS1494del6 polymorphism in colorectal patients from west of Mexico]. *Rev Invest Clin* 2008; **60**: 21-30 [PMID: 18589584]
- 51 **Wang M**, Li G, Yang Z, Wang L, Zhang L, Wang T, Zhang Y, Zhang S, Han Y, Jia L. Uncoupling protein 2 downregulation by

- hypoxia through repression of peroxisome proliferator-activated receptor γ promotes chemoresistance of non-small cell lung cancer. *Oncotarget* 2017; **8**: 8083-8094 [PMID: 28042952 DOI: 10.18632/oncotarget.14097]
- 52 **Krauss S**, Zhang CY, Lowell BB. The mitochondrial uncoupling-protein homologues. *Nat Rev Mol Cell Biol* 2005; **6**: 248-261 [PMID: 15738989 DOI: 10.1038/nrm1572]
 - 53 **Baffy G**. Uncoupling protein-2 and cancer. *Mitochondrion* 2010; **10**: 243-252 [PMID: 20005987 DOI: 10.1016/j.mito.2009.12.143]
 - 54 **Esteves P**, Pecqueur C, Ransy C, Esnous C, Lenoir V, Bouillaud F, Bulteau AL, Lombès A, Prip-Buus C, Ricquier D, Alves-Guerra MC. Mitochondrial retrograde signaling mediated by UCP2 inhibits cancer cell proliferation and tumorigenesis. *Cancer Res* 2014; **74**: 3971-3982 [PMID: 24853548 DOI: 10.1158/0008-5472.CAN-13-3383]
 - 55 **Dalla Pozza E**, Fiorini C, Dando I, Menegazzi M, Sgarbossa A, Costanzo C, Palmieri M, Donadelli M. Role of mitochondrial uncoupling protein 2 in cancer cell resistance to gemcitabine. *Biochim Biophys Acta* 2012; **1823**: 1856-1863 [PMID: 22705884 DOI: 10.1016/j.bbamcr.2012.06.007]
 - 56 **Kuai XY**, Ji ZY, Zhang HJ. Mitochondrial uncoupling protein 2 expression in colon cancer and its clinical significance. *World J Gastroenterol* 2010; **16**: 5773-5778 [PMID: 21128330 DOI: 10.3748/wjg.v16.i45.5773]
 - 57 **Benhar M**, Engelberg D, Levitzki A. ROS, stress-activated kinases and stress signaling in cancer. *EMBO Rep* 2002; **3**: 420-425 [PMID: 11991946 DOI: 10.1093/embo-reports/kvf094]
 - 58 **Lenaz G**. The mitochondrial production of reactive oxygen species: mechanisms and implications in human pathology. *IUBMB Life* 2001; **52**: 159-164 [PMID: 11798028 DOI: 10.1080/15216540152845957]
 - 59 **Horimoto M**, Resnick MB, Konkin TA, Routhier J, Wands JR, Baffy G. Expression of uncoupling protein-2 in human colon cancer. *Clin Cancer Res* 2004; **10**: 6203-6207 [PMID: 15448008 DOI: 10.1158/1078-0432.CCR-04-0419]
 - 60 **Lentes KU**, Tu N, Chen H, Winnikes U, Reinert I, Marmann G, Pirke KM. Genomic organization and mutational analysis of the human UCP2 gene, a prime candidate gene for human obesity. *J Recept Signal Transduct Res* 1999; **19**: 229-244 [PMID: 10071761 DOI: 10.3109/10799899909036648]
 - 61 **Tu N**, Chen H, Winnikes U, Reinert I, Marmann G, Pirke KM, Lentes KU. Structural organization and mutational analysis of the human uncoupling protein-2 (hUCP2) gene. *Life Sci* 1999; **64**: PL41-PL50 [PMID: 10027754 DOI: 10.1016/S0024-3205(98)00555-4]
 - 62 **Moreno-Muñoz D**, de la Haba-Rodríguez JR, Conde F, López-Sánchez LM, Valverde A, Hernández V, Martínez A, Villar C, Gómez-España A, Porras I, Rodríguez-Ariza A, Aranda E. Genetic variants in the renin-angiotensin system predict response to bevacizumab in cancer patients. *Eur J Clin Invest* 2015; **45**: 1325-1332 [PMID: 26509357 DOI: 10.1111/eci.12557]
 - 63 **George AJ**, Thomas WG, Hannan RD. The renin-angiotensin system and cancer: old dog, new tricks. *Nat Rev Cancer* 2010; **10**: 745-759 [PMID: 20966920 DOI: 10.1038/nrc2945]
 - 64 **Rodríguez-Pérez JC**, Rodríguez-Esparragón F, Hernández-Perera O, Anabitarte A, Losada A, Medina A, Hernández E, Fiuza D, Avalos O, Yunis C, Ferrario CM. Association of angiotensinogen M235T and A(-)6G gene polymorphisms with coronary heart disease with independence of essential hypertension: the PROCAGENE study. Prospective Cardiac Gene. *J Am Coll Cardiol* 2001; **37**: 1536-1542 [PMID: 11345362 DOI: 10.1016/S0735-1097(01)01186-X]
 - 65 **van der Knaap R**, Siemes C, Coebergh JW, van Duijn CM, Hofman A, Stricker BH. Renin-angiotensin system inhibitors, angiotensin I-converting enzyme gene insertion/deletion polymorphism, and cancer: the Rotterdam Study. *Cancer* 2008; **112**: 748-757 [PMID: 18181094 DOI: 10.1002/ncr.23215]
 - 66 **Srivastava K**, Srivastava A, Mittal B. Angiotensin I-converting enzyme insertion/deletion polymorphism and increased risk of gall bladder cancer in women. *DNA Cell Biol* 2010; **29**: 417-422 [PMID: 20438364 DOI: 10.1089/dna.2010.1033]
 - 67 **de Martino M**, Klatte T, Schatzl G, Waldert M, Remzi M, Haitel A, Kramer G, Marberger M. Insertion/deletion polymorphism of angiotensin I-converting enzyme gene is linked with chromophobe renal cell carcinoma. *Urology* 2011; **77**: 1005.e9-1005.e13 [PMID: 21477733 DOI: 10.1016/j.urology.2010.11.033]
 - 68 **Xie Y**, You C, Chen J. An updated meta-analysis on association between angiotensin I-converting enzyme gene insertion/deletion polymorphism and cancer risk. *Tumour Biol* 2014; **35**: 6567-6579 [PMID: 24691970 DOI: 10.1007/s13277-014-1842-z]
 - 69 **Yang H**, Cai C, Ye L, Rao Y, Wang Q, Hu D, Huang X. The relationship between angiotensin-converting enzyme gene insertion/deletion polymorphism and digestive cancer risk: Insights from a meta-analysis. *J Renin Angiotensin Aldosterone Syst* 2015; **16**: 1306-1313 [PMID: 25990649 DOI: 10.1177/1470320315585908]
 - 70 **Liu SY**, Sima X, Wang CH, Gao M. The association between ACE polymorphism and risk of colorectal cancer in a Chinese population. *Clin Biochem* 2011; **44**: 1223-1226 [PMID: 21843521 DOI: 10.1016/j.clinbiochem.2011.07.016]
 - 71 **Zambra FM**, Biolchi V, de Cerqueira CC, Brum IS, Castelli EC, Chies JA. Immunogenetics of prostate cancer and benign hyperplasia--the potential use of an HLA-G variant as a tag SNP for prostate cancer risk. *HLA* 2016; **87**: 79-88 [PMID: 26889902 DOI: 10.1111/tan.12741]
 - 72 **Paul P**, Rouas-Freiss N, Khalil-Daher I, Moreau P, Riteau B, Le Gal FA, Avril MF, Dausset J, Guillet JG, Carosella ED. HLA-G expression in melanoma: a way for tumor cells to escape from immunosurveillance. *Proc Natl Acad Sci USA* 1998; **95**: 4510-4515 [PMID: 9539768]
 - 73 **Amiot L**, Ferrone S, Grosse-Wilde H, Seliger B. Biology of HLA-G in cancer: a candidate molecule for therapeutic intervention? *Cell Mol Life Sci* 2011; **68**: 417-431 [PMID: 21063893 DOI: 10.1007/s00018-010-0583-4]
 - 74 **Hviid TV**, Høylenius S, Rørbye C, Nielsen LG. HLA-G allelic variants are associated with differences in the HLA-G mRNA isoform profile and HLA-G mRNA levels. *Immunogenetics* 2003; **55**: 63-79 [PMID: 12712263 DOI: 10.1007/s00251-003-0547-z]
 - 75 **Chen S**, Laverdiere I, Tourancheau A, Jonker D, Couture F, Cecchin E, Villeneuve L, Harvey M, Court MH, Innocenti F, Toffoli G, Lévesque E, Guillemette C. A novel UGT1 marker associated with better tolerance against irinotecan-induced severe neutropenia in metastatic colorectal cancer patients. *Pharmacogenomics J* 2015; **15**: 513-520 [PMID: 25778466 DOI: 10.1038/tpj.2015.12]
 - 76 **Beutler E**, Gelbart T, Demina A. Racial variability in the UDP-glucuronosyltransferase 1 (UGT1A1) promoter: a balanced polymorphism for regulation of bilirubin metabolism? *Proc Natl Acad Sci USA* 1998; **95**: 8170-8174 [PMID: 9653159]
 - 77 **Barbarino JM**, Haidar CE, Klein TE, Altman RB. PharmGKB summary: very important pharmacogene information for UGT1A1. *Pharmacogenet Genomics* 2014; **24**: 177-183 [PMID: 24492252 DOI: 10.1097/FPC.0000000000000024]
 - 78 **Hu ZY**, Yu Q, Pei Q, Guo C. Dose-dependent association between UGT1A1*28 genotype and irinotecan-induced neutropenia: low doses also increase risk. *Clin Cancer Res* 2010; **16**: 3832-3842 [PMID: 20562211 DOI: 10.1158/1078-0432.CCR-10-1122]
 - 79 **Hu ZY**, Yu Q, Zhao YS. Dose-dependent association between UGT1A1*28 polymorphism and irinotecan-induced diarrhoea: a meta-analysis. *Eur J Cancer* 2010; **46**: 1856-1865 [PMID: 20335017 DOI: 10.1016/j.ejca.2010.02.049]
 - 80 **Dias MM**, McKinnon RA, Sorich MJ. Impact of the UGT1A1*28 allele on response to irinotecan: a systematic review and meta-analysis. *Pharmacogenomics* 2012; **13**: 889-899 [PMID: 22676194 DOI: 10.2217/pgs.12.68]
 - 81 **Yang H**, Sasaki T, Minoshima S, Shimizu N. Identification of three novel proteins (SGSM1, 2, 3) which modulate small G protein (RAP and RAB)-mediated signaling pathway. *Genomics* 2007; **90**: 249-260 [PMID: 17509819 DOI: 10.1016/j.ygeno.2007.03.013]
 - 82 **Lee IK**, Kim KS, Kim H, Lee JY, Ryu CH, Chun HJ, Lee KU, Lim

- Y, Kim YH, Huh PW, Lee KH, Han SI, Jun TY, Rha HK. MAP, a protein interacting with a tumor suppressor, merlin, through the run domain. *Biochem Biophys Res Commun* 2004; **325**: 774-783 [PMID: 15541357 DOI: 10.1016/j.bbrc.2004.10.095]
- 83 Wang C, Zhao H, Zhao X, Wan J, Wang D, Bi W, Jiang X, Gao Y. Association between an insertion/deletion polymorphism within 3' UTR of SGSM3 and risk of hepatocellular carcinoma. *Tumour Biol* 2014; **35**: 295-301 [PMID: 23918301 DOI: 10.1007/s13277-013-1039-x]
 - 84 Savas S, Liu G. Studying genetic variations in cancer prognosis (and risk): a primer for clinicians. *Oncologist* 2009; **14**: 657-666 [PMID: 19581524 DOI: 10.1634/theoncologist.2009-0042]
 - 85 Röcken C, Lendeckel U, Dierkes J, Westphal S, Carl-McGrath S, Peters B, Krüger S, Malfertheiner P, Roessner A, Ebert MP. The number of lymph node metastases in gastric cancer correlates with the angiotensin I-converting enzyme gene insertion/deletion polymorphism. *Clin Cancer Res* 2005; **11**: 2526-2530 [PMID: 15814629 DOI: 10.1158/1078-0432.CCR-04-1922]
 - 86 Tang YI, Liu Y, Zhao W, Yu T, Yu H. Caspase-8 polymorphisms and risk of oral squamous cell carcinoma. *Exp Ther Med* 2015; **10**: 2267-2276 [PMID: 26668627 DOI: 10.3892/etm.2015.2832]
 - 87 Hengartner MO. The biochemistry of apoptosis. *Nature* 2000; **407**: 770-776 [PMID: 11048727 DOI: 10.1038/35037710]
 - 88 Siegel RM. Caspases at the crossroads of immune-cell life and death. *Nat Rev Immunol* 2006; **6**: 308-317 [PMID: 16557262 DOI: 10.1038/nri1809]
 - 89 Halder G, Johnson RL. Hippo signaling: growth control and beyond. *Development* 2011; **138**: 9-22 [PMID: 21138973 DOI: 10.1242/dev.045500]
 - 90 Harvey K, Tapon N. The Salvador-Warts-Hippo pathway - an emerging tumour-suppressor network. *Nat Rev Cancer* 2007; **7**: 182-191 [PMID: 17318211 DOI: 10.1038/nrc2070]
 - 91 Pan D. The hippo signaling pathway in development and cancer. *Dev Cell* 2010; **19**: 491-505 [PMID: 20951342 DOI: 10.1016/j.devcel.2010.09.011]
 - 92 Na HK, Lee JY. Molecular Basis of Alcohol-Related Gastric and Colon Cancer. *Int J Mol Sci* 2017; **18**: [PMID: 28538665 DOI: 10.3390/ijms18061116]
 - 93 Fernandes GM, Russo A, Proença MA, Gazola NF, Rodrigues GH, Biselli-Chicote PM, Silva AE, Netinho JG, Pavarino EC, Goloni-Bertollo EM. CYP1A1, CYP2E1 and EPHX1 polymorphisms in sporadic colorectal neoplasms. *World J Gastroenterol* 2016; **22**: 9974-9983 [PMID: 28018104 DOI: 10.3748/wjg.v22.i45.9974]
 - 94 Nebert DW, Dalton TP. The role of cytochrome P450 enzymes in endogenous signalling pathways and environmental carcinogenesis. *Nat Rev Cancer* 2006; **6**: 947-960 [PMID: 17128211 DOI: 10.1038/nrc2015]
 - 95 Pfeifer GP, Denissenko MF, Olivier M, Tretyakova N, Hecht SS, Hainaut P. Tobacco smoke carcinogens, DNA damage and p53 mutations in smoking-associated cancers. *Oncogene* 2002; **21**: 7435-7451 [PMID: 12379884 DOI: 10.1038/sj.onc.1205803]
 - 96 Cury NM, Russo A, Galbiatti AL, Ruiz MT, Raposo LS, Maniglia JV, Pavarino EC, Goloni-Bertollo EM. Polymorphisms of the CYP1A1 and CYP2E1 genes in head and neck squamous cell carcinoma risk. *Mol Biol Rep* 2012; **39**: 1055-1063 [PMID: 21590276 DOI: 10.1007/s11033-011-0831-1]
 - 97 Lu Y, Zhu X, Zhang C, Jiang K, Huang C, Qin X. Role of CYP2E1 polymorphisms in breast cancer: a systematic review and meta-analysis. *Cancer Cell Int* 2017; **17**: 11 [PMID: 28074086 DOI: 10.1186/s12935-016-0371-9]
 - 98 Garziera M, Toffoli G. Inhibition of host immune response in colorectal cancer: human leukocyte antigen-G and beyond. *World J Gastroenterol* 2014; **20**: 3778-3794 [PMID: 24744572 DOI: 10.3748/wjg.v20.i14.3778]
 - 99 Garziera M, Bidoli E, Cecchin E, Mini E, Nobili S, Lonardi S, Buonadonna A, Errante D, Pella N, D'Andrea M, De Marchi F, De Paoli A, Zanuso C, De Mattia E, Tassi R, Toffoli G. HLA-G 3'UTR Polymorphisms Impact the Prognosis of Stage II-III CRC Patients in Fluoropyrimidine-Based Treatment. *PLoS One* 2015; **10**: e0144000 [PMID: 26633805 DOI: 10.1371/journal.pone.0144000]
 - 100 Guo ZY, Lv YG, Wang L, Shi SJ, Yang F, Zheng GX, Wen WH, Yang AG. Predictive value of HLA-G and HLA-E in the prognosis of colorectal cancer patients. *Cell Immunol* 2015; **293**: 10-16 [PMID: 25461612 DOI: 10.1016/j.cellimm.2014.10.003]
 - 101 Ye SR, Yang H, Li K, Dong DD, Lin XM, Yie SM. Human leukocyte antigen G expression: as a significant prognostic indicator for patients with colorectal cancer. *Mod Pathol* 2007; **20**: 375-383 [PMID: 17277760 DOI: 10.1038/modpathol.3800751]
 - 102 Zeestraten EC, Reimers MS, Saadatmand S, Goossens-Beumer IJ, Dekker JW, Liefers GJ, van den Elsen PJ, van de Velde CJ, Kuppen PJ. Combined analysis of HLA class I, HLA-E and HLA-G predicts prognosis in colon cancer patients. *Br J Cancer* 2014; **110**: 459-468 [PMID: 24196788 DOI: 10.1038/bjc.2013.696]
 - 103 Gao Y, He Y, Ding J, Wu K, Hu B, Liu Y, Wu Y, Guo B, Shen Y, Landi D, Landi S, Zhou Y, Liu H. An insertion/deletion polymorphism at miRNA-122-binding site in the interleukin-1alpha 3' untranslated region confers risk for hepatocellular carcinoma. *Carcinogenesis* 2009; **30**: 2064-2069 [PMID: 19917630 DOI: 10.1093/carcin/bgp283]
 - 104 Trabert B, Pinto L, Hartge P, Kemp T, Black A, Sherman ME, Brinton LA, Pfeiffer RM, Shiels MS, Chaturvedi AK, Hildesheim A, Wentzensen N. Pre-diagnostic serum levels of inflammation markers and risk of ovarian cancer in the prostate, lung, colorectal and ovarian cancer (PLCO) screening trial. *Gynecol Oncol* 2014; **135**: 297-304 [PMID: 25158036 DOI: 10.1016/j.ygyno.2014.08.025]
 - 105 Volonté A, Di Tomaso T, Spinelli M, Todaro M, Sanvito F, Albarello L, Bissolati M, Ghirardelli L, Orsenigo E, Ferrone S, Doglioni C, Stassi G, Dellabona P, Staudacher C, Parmiani G, Maccalli C. Cancer-initiating cells from colorectal cancer patients escape from T cell-mediated immunosurveillance in vitro through membrane-bound IL-4. *J Immunol* 2014; **192**: 523-532 [PMID: 24277698 DOI: 10.4049/jimmunol.1301342]
 - 106 Koller FL, Hwang DG, Dozier EA, Fingleton B. Epithelial interleukin-4 receptor expression promotes colon tumor growth. *Carcinogenesis* 2010; **31**: 1010-1017 [PMID: 20176658 DOI: 10.1093/carcin/bgq044]
 - 107 Brooks CL, Gu W. p53 ubiquitination: Mdm2 and beyond. *Mol Cell* 2006; **21**: 307-315 [PMID: 16455486 DOI: 10.1016/j.molcel.2006.01.020]
 - 108 Bi H, Tian T, Zhu L, Zhou H, Hu H, Liu Y, Li X, Hu F, Zhao Y, Wang G. Copy number variation of E3 ubiquitin ligase genes in peripheral blood leukocyte and colorectal cancer. *Sci Rep* 2016; **6**: 29869 [PMID: 27417709 DOI: 10.1038/srep29869]
 - 109 Cerhan JR, Liu-Mares W, Fredericksen ZS, Novak AJ, Cunningham JM, Kay NE, Dogan A, Liebow M, Wang AH, Call TG, Habermann TM, Ansell SM, Slager SL. Genetic variation in tumor necrosis factor and the nuclear factor-kappaB canonical pathway and risk of non-Hodgkin's lymphoma. *Cancer Epidemiol Biomarkers Prev* 2008; **17**: 3161-3169 [PMID: 18990758 DOI: 10.1158/1055-9965.EPI-08-0536]
 - 110 Chen F, Castranova V, Shi X, Demers LM. New insights into the role of nuclear factor-kappaB, a ubiquitous transcription factor in the initiation of diseases. *Clin Chem* 1999; **45**: 7-17 [PMID: 9895331]
 - 111 McLure KG, Takagi M, Kastan MB. NAD⁺ modulates p53 DNA binding specificity and function. *Mol Cell Biol* 2004; **24**: 9958-9967 [PMID: 15509798 DOI: 10.1128/MCB.24.22.9958-9967.2004]
 - 112 Riley T, Sontag E, Chen P, Levine A. Transcriptional control of human p53-regulated genes. *Nat Rev Mol Cell Biol* 2008; **9**: 402-412 [PMID: 18431400 DOI: 10.1038/nrm2395]
 - 113 Sun Y, Zheng W, Guo Z, Ju Q, Zhu L, Gao J, Zhou L, Liu F, Xu Y, Zhan Q, Zhou Z, Sun W, Zhao X. A novel TP53 pathway influences the HGS-mediated exosome formation in colorectal cancer. *Sci Rep* 2016; **6**: 28083 [PMID: 27312428 DOI: 10.1038/srep28083]
 - 114 Brosh R, Rotter V. When mutants gain new powers: news from

- the mutant p53 field. *Nat Rev Cancer* 2009; **9**: 701-713 [PMID: 19693097 DOI: 10.1038/nrc2693]
- 115 **Siewchaisakul P**, Suwanrungruang K, Poomphakwaen K, Wiangnon S, Promthet S. Lack of Association between an XRCC1 Gene Polymorphism and Colorectal Cancer Survival in Thailand. *Asian Pac J Cancer Prev* 2016; **17**: 2055-2060 [PMID: 27221895]
- 116 **de Boer JG**. Polymorphisms in DNA repair and environmental interactions. *Mutat Res* 2002; **509**: 201-210 [PMID: 12427539]
- 117 **Huang Y**, Li X, He J, Chen L, Huang H, Liang M, Zhu Q, Huang Y, Wang L, Pan C, Xia T. Genetic polymorphisms in XRCC1 genes and colorectal cancer susceptibility. *World J Surg Oncol* 2015; **13**: 244 [PMID: 26271249 DOI: 10.1186/s12957-015-0650-2]

P- Reviewer: Dai ZJ, Engin AB, Wang YH **S- Editor:** Gong ZM

L- Editor: A **E- Editor:** Huang Y





Published by **Baishideng Publishing Group Inc**
7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
Telephone: +1-925-223-8242
Fax: +1-925-223-8243
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>



ISSN 1007-9327



9 771007 932045