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***Observational Study***

**Circulating miR-125a but not miR-125b is decreased in active disease status and negatively correlates with disease severity as well as inflammatory cytokines in patients with Crohn's disease**

Sun CM *et al*. Circulating miR-125a/b in CD

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**Abstract**

***AIM***

To determine the association of circulating miR-125a/b expression with the risk and disease severity of Crohn's disease (CD), and with inflammatory cytokines.

***METHODS***

Plasma samples were collected from patients with active CD (A-CD), or CD in remission (R-CD) and from health controls (HCs). The levels of the inflammatory cytokines interleukin 17 (IL-17), tumour necrosis factor-α (TNF-α) and interferon-γ (IFN-γ) were measured by enzyme-linked immunosorbent assay (ELISA). The expression of miR-125a/b was assessed by quantitative polymerase chain reaction (qPCR).

***RESULTS***

Twenty-nine A-CD patients, 37 R-CD patients and 37 HCs were included in the study. Plasma miR-125a expression was decreased in A-CD patients compared with that in R-CD patients (*P* < 0.001) and HCs (*P* < 0.001). MiR-125a expression values enabled the differentiation of A-CD from R-CD patients (AUC = 0.854) and from HCs (AUC = 0.780), whereas miR-125b expression did not. miR-125a was negatively correlated with C-reaction protein (CRP) (*P* = 0.017), erythrocyte sedimentation rate (ESR) (*P* = 0.026), Crohn’s disease activity index (CDAI) (*P* = 0.003), IL-17 (*P* = 0.015) and TNF-α (*P* = 0.004) in A-CD patients. Furthermore, miR-125a was negatively association with CRP (*P* = 0.038) and CDAI (*P* = 0.021) in R-CD patients. Regarding miR-125b, no association with CRP, CDAI, IL-17, TNF-α or IFN-γ was found in A-CD or in R-CD patients (all *P* > 0.05). MiR-125a levels gradually increased in A-CD patients who achieved clinical remission (*P* = 0.009) after a 3-month treatment, whereas they remained unchanged among patients who failed to achieve remission (*P* > 0.05). No changes in miR-125b expression were detected in remission or non-remission patients after treatment (both *P* > 0.05).

***CONCLUSION***

Circulating miR-125a but not miR-125b is decreased in patients with active disease status and negatively correlates with disease severity and inflammatory cytokines in patients with CD.

**Key words:** Crohn's disease; miR-125; Disease risk; Disease severity; Inflammatory cytokines

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**Core tip：**This study aimed to investigate the association of circulating miR-125a/b expression with the risk and severity of Crohn's disease (CD) and with inflammatory cytokines. Our results showed that miR-125a but not miR-125b is negatively correlated with the risk of active CD and disease severity and with inflammatory cytokines.

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**INTRODUCTION**

Crohn’s disease (CD), an idiopathic chronic inflammatory disease, is an inflammatory bowel diseases (IBD) and primarily affects the gastrointestinal tract. This disease causes the development of ulcers and complications, including abscesses and fistulas, affecting all layers of the intestinal wall[1,2]. It is reported that CD has an annual incidence of approximately 24 per 100000 in Europe and 19 per 100000 in North America. However, the incidence rate of CD is still increasing in developing countries, particularly in China due to its considerable increase in gross domestic product (GDP)[3,4] and improvement in quality of life[5].

miRNAs are small endogenous RNAs that can degrade targeted mRNA or inhibit protein synthesis, and increasing evidence shows that miRNAs play a key role in regulating the intestinal immune system[6-9]. For example, miR-29b inhibits transforming growth factor-β (TGF-β)-induced intestinal fibrosis, and Let-7 regulates the activity of nuclear factor-κB (NF-κB) and mediates IL-6 down-regulation in CD patients[10,11]. Furthermore, miR-192, miR-142-3p, and miR-21 are notably upregulated in paediatric IBD patients[12], whereas miR-495-5p and miR-19b are down-regulated in the inflamed bowel of CD patients[13,14].

microRNA-125 family (miR-125), an miRNA family highly conserved throughout evolution, consists of miR-125a and miR-125b[15]. It has been shown that miR-125a inhibits innate macrophage responses by suppressing macrophage differentiation, and the expression level of miR-125a is down-regulated in systemic lupus erythematosus (SLE)[16,17]. miR-125b was correlated with rheumatoid arthritis (RA) disease activity and may serve as a potential biomarker for treatment response in early RA[18]. To date, few studies have investigated the impact of dysregulated miR-125a/b expression in CD patients. Therefore, the aim of our study was to determine the association of circulating miR-125a/b expression with the risk and disease severity of CD and with inflammatory cytokines.

**MATERIALS AND METHODS**

***Participants***

Twenty-nine clinically active CD (A-CD) patients and 37 CD patients in clinical remission (R-CD) were enrolled in this study from May 2014 to June 2016 at the Department of Gastroenterology, The Central Hospital of Wuhan, Tongji Medical College, Huazhong University of Science and Technology. The diagnosis of CD was determined according to the Lennard-Jones criteria[19], and active clinical disease was defined as Crohn’s disease activity index (CDAI) above 150 points. Patients with the following conditions were excluded: arthritis with complications or other autoimmune diseases; history of malignant tumour or complications; severe renal and/or kidney dysfunction; history of serious surgery or severe infection. Concurrently, 37 health controls (HCs) age and gender matched to the CD patients were enrolled.

The study was approved by the Ethics Committee of The Central Hospital of Wuhan, Tongji Medical College, Huazhong University of Science and Technology, and all participants signed the informed consent forms.

***Sample collection and total RNA extraction***

Blood samples were collected from all participants into EDTA tubes when they enrolled in this study and after a 3-month treatment in the case of A-CD patients. After collection, the blood samples were centrifuged at 1000 g for 15 mins at 4 ℃; then, the supernatant (plasma) was removed and stored at -80 ℃ for further analysis. If red blood cell lysis was discovered, a repeat blood sample was collected from patients or HCs. Total RNA was extracted from the samples with TRIzol reagent (Invitrogen, CA, United States) according to the manufacturer’s instructions.

***miR-125a/b determination by qPCR***

RNA was translated by reverse transcription using the PrimerScript Real-time reagent kit (TAKARA BIO Inc. Shiga, Japan). Subsequently, miR-125 a/b expression levels were quantitated by SYBR Premix Ex TaqTM II (TAKARA BIO Inc. Shiga, Japan). The expression level of miR-125a/b was calculated using the 2-△△t method and U6 was used as the internal reference.

***IL-17, TNF-α and IFN-γ* measurement by enzyme-linked immunosorbent assay**

The measurement of interleukin 17 (IL-17), tumour necrosis factor-α (TNF-α) and interferon-γ (IFN-γ) expression levels in plasma samples from A-CD and R-CD patients (but not HCs) was performed using commercial ELISA kits, according to the manufacturer’s instructions (all purchased from eBioscience, CA, United States).

***Disease evaluation and treatment of A-CD patients***

C-reaction protein (CRP), erythrocyte sedimentation rate (ESR), and Crohn’s disease activity index (CDAI) were used to assess the severity of CD and to examine the association of miR-125a/b expression with disease severity. In A-CD patients, effective treatments, including immunosuppressive agents and biologics, among others, were used, according to disease status and clinical experience. After a 3-month treatment, the CDAI and blood of patients were assessed again, and changes in miR-125a/b expression were analysed based on clinical remission achievement.

***Statistics***

Statistical analysis was performed using the SPSS 21.0 software program (IBM, United States) and MS Office 2016 (Microsoft, United States). Data were mainly expressed as mean±standard deviation, median (1/4-3/4 quarters) or counts. Three-group comparisons were performed by one-way analysis of variance (ANOVA), Kruskal-Wallis test by ranks or Chi-squared test. Two-group comparisons were performed using the Wilcoxon signed-rank test. Comparisons between each visit in the same group were analysed using the Wilcoxon signed-rank test. Receiver operating characteristic (ROC) curve analysis was performed to assess the value of miR-125a/b expression in differentiating A-CD from R-CD patients and patients from HCs. Spearman’s test was used to analyse the correlation of miR-125a/b expression with disease severity and inflammatory cytokines. *P* < 0.05 was considered significant.

**RESULTS**

***Characteristics***

As shown in Table 1, the 29 patients of the A-CD group had a mean age of 31.38 ± 9.51 years, whereas the 37 patients of the R-CD and the 37 patients of the HCs had mean ages of 31.97 ± 8.86 and 31.41 ± 5.69 years, respectively. No difference in age or gender was found (both *P* > 0.05), although significant differences in CRP and ESR (both *P* < 0.001) were observed between groups. Significant differences in inflammatory cytokines IL-17, TNF-α, and IFN and in CDAI were also found between A-CD and R-CD patients (all *P* < 0.001).

***miR-125a/b expression in A-CD patients, R-CD patients and HCs***

Plasma miR-125a expression was decreased in A-CD patients [0.719 (0.534-1.072)] compared with that of R-CD patients [1.564 (1.159-2.851), *P* < 0.001] and HCs [1.781 (0.874-2.873), *P* < 0.001]. However, no significant difference between R-CD patients and HCs (*P* = 0.297) was found (Figure 1A). No difference in miR-125b expression was observed between A-CD [2.707 (2.168-3.531)] and R-CD patients [2.600 (2.154-3.064)] and between patients and HCs [2.393 (1.705-2.852)], as shown in Figure 1B (all *P* > 0.05).

Interestingly, diagnostic values were discovered for miR-125a in differentiating A-CD from R-CD patients (AUC = 0.854, 95%CI: 0.758-0.949) and patients from HCs (AUC = 0.780, 95%CI: 0.665-0.895) (Figure 2A and B). However, miR-125b failed as a predictive value in differentiating A-CD from R-CD patients (AUC = 0.571, 95%CI: 0.423-0.718) and patients from HCs (AUC = 0.629, 95%CI: 0.492-0.766) (Figure 2A and B). Moreover, neither miR-125a nor miR-125b were able to differentiate R-CD patients from HCs (AUC = 0.537, 95%CI: 0.406-0.668; AUC = 0.590, 95%CI: 0.461-0.720, respectively) (Figure 2C).

***Correlations of miR-125a/b expression with disease severity in CD patients***

Negative correlations of miR-125a with CRP (*r* = -0.439, *P* = 0.017), ESR (*r* = -0.412, *P* = 0.026) and CDAI (*r* = -0.526, *P* = 0.003) in A-CD patients were observed, as shown in Figure 3A-C. However, no association was found between miR-125b and CRP, ESR or CDAI in A-CD patients (Figure 3D-F) (all *P* > 0.05).

Regarding R-CD patients, we found that the expression of miR-125a was negatively correlated with CRP (*r* = -0.342, *P* = 0.038) and CDAI (*r* = -0.379, *P* = 0.021) but not ESR (*r* = -0.140, *P* = 0.410) (Figure 4A-C). No significant association of miR-125b was observed in R-CD patients, except for ESR (r = 0.326, *P* = 0.049); CRP(*r* = -0.068, *P* = 0.691); CDAI (r = 0.126, *P* = 0.457); (Figure 4D-F).

***Correlations of miR-125a/b expression with inflammatory cytokines in CD patients***

Negative correlations were detected for miR-125a with the inflammatory factors IL-17 (*r* = -0.446, *P* = 0.015) and TNF-α (*r* = -0.518, *P* = 0.004), but not with IFN-γ (*r* =- 0.203, *P* = 0.290), in A-CD patients, as shown in Figure 5A-C. However, no association of miR-125b with the inflammatory factors IL-17 (*r* = -0.027, *P* = 0.890), TNF-α (*r* = 0.091, *P* = 0.640) and IFN-γ (*r* = -0.198, *P* = 0.304) was found in A-CD patients (Figure 5D-F). Regarding R-CD patients, we observed no miR-125a or miR-125b correlation with the inflammatory factors IL-17, TNF-α or IFN-γ, as shown in Figure 6A-F (all *P* > 0.05).

***miR-125a/b expression after treatment in A-CD patients***

After a 3-month treatment, 19 of 29 A-CD patients achieved clinical remission (CDAI < 150), whereas 10 patients failed to achieve clinical remission. In clinical remission patients, the miR-125a levels dramatically increased compared with the baseline (*P* = 0.009); conversely, in non-remission patients, miR-125a levels remained unchanged compared with the baseline (*P* > 0.05). No changes in miR-125b expression were detected, either in remission or non-remission patients (both *P* > 0.05), after treatment **(**Figure 7).

**DISCUSSION**

CD, a chronic debilitating syndrome, affects all layers of the gastrointestinal tract and causes severe diarrhea, abdominal pain, weight loss, metabolic disorder, and malabsorption and is a major health concern, leading to huge financial losses of up to $2.2 billion dollars per year in the United States alone[20-23]. Although the pathogenesis of CD is still unclear, accumulating evidence shows that environmental factors, genetics, autoimmunity and dietary habits may contribute to its development and progression[24, 25]. As an important genetic factor, miRNA dysregulation is involved in the aetiological mechanism of several autoimmune diseases, including CD[26]; however, the role of miR-125a/b expression in CD risk and management is largely unstudied[27].

In this prospective study, we observed three notable findings. First, miR-125a expression was decreased in A-CD patients compared with R-CD patients and HCs. The results also showed the ability to distinguish A-CD from R-CD patients and from HCs using miR-125a expression levels, which was not possible using miR-125b expression levels. Second, miR-125a was negatively correlated with disease severity in A-CD patients, and negative correlations of miR-125a with inflammatory cytokines were also found in A-CD patients.Third, miR-125a levels were dramatically increased in A-CD patients who achieved clinical remission after a 3-mo treatment, whereas miR-125a levels remained unchanged in non-remission patients. No changes in miR-125b expression were detected in remission or non-remission A-CD patients after treatment.

The miR-125a gene is located on chromosome 19 and in a cluster with miR-99b and miR-7e[28]. In health population, the largest contributor of circulating miR-125a may be germinal center (GC) and hematopoietic stem cells (HSC). Shaham et al. suggests that miR-125a is enriched in HSCs (up to 23-fold more than in total bone marrow), particularly in long-term HSCs (up to 6-fold). Moreover, miR-125a is not restricted to the stem cell population, and its cluster members, miR-99b and let-7e, are preferentially expressed by centroblasts in the GC[16]. Studies show that miR-125a is down-regulated in peripheral CD3+ T cells and negatively correlated with RANTES (also known as CCL5 chemokine) expression by targeting the KLF13 gene in SLE patients[17]. Interestingly, miR-125a expression is also decreased in oral lichen planus, which is a T-cell-mediated autoimmune disease of the oral mucosa[29]. Furthermore, miR-125a is identified as a key regulator of CD4+ T-cell differentiation that prevents autoimmune pathogenesis by controlling the balance between tolerance and autoimmunity[30]. Recently, it was reported that miR-125a participates in immune thrombocytopenic purpura (ITP), by modulating Tregs and Th17[31], which play a key role in CD development and progression and are correlated with CD disease activity[32-34]. These studies indicate that miR-125a is an anti-inflammatory gene that plays a key role in regulating autoimmune diseases, which is consistent with our results showing that miR-125a expression is decreased in A-CD patients and may be used to differentiate A-CD from R-CD patients and from HCs. This might be due to the anti-inflammatory effect of miR-125a, because the levels of inflammatory cytokines were markedly increased in A-CD patients compared with those of R-CD patients and HCs, whereas the extent of inflammation in R-CD patients was similar to that of HCs. The results showing a negative correlation between miR-125a and the inflammatory cytokines IL-17 and TNF-α further confirmed this point of view.

In addition, we found a negative association of miR-125a with disease severity in A-CD patients. Partly in line with our results, Murata et al. reported that miR-125a was negatively correlated with some indices of disease activity including CRP in rheumatoid arthritis (RA)[35], and reduced levels of miR-125a were associated with severe trauma through inflammatory cytokine IL-10 regulation, as shown in polytrauma patients[36], most likely, because of the powerful anti-inflammatory effect of miR-125a. Thus, miR-125a was able to predict the disease severity of CD. Furthermore, we found that the miR-125a levels were dramatically increased in A-CD patients who achieved clinical remission after a 3-mo treatment, whereas they remained unchanged in patients who failed to achieve remission. The results proved once again that miR-125a is closely related to inflammation and might be a therapeutic target for CD in the future.

miR125-b has been shown to target the 3’UTR region of the TNF-α gene to negatively regulate the inflammatory response[37]. However, another study has shown that miR-125b could promote macrophage mediated inflammation by increasing the expression of co-stimulatory factor[38]. These studies suggest that miR-125b has both anti- and pro-inflammatory effects. We found no miR-125b correlation with inflammatory factors in A-CD patients, and miR-125b was not able to differentiate A-CD from R-CD patients and from HCs, most likely because miR-125b has both anti- and pro-inflammatory effects, which has been reported in previous studies. Thus, the role of miR-125b in regulating inflammation in CD remains unclear[16,39,40]. Then, we analysed the target genes of miR-125a and miR-125b by Validated Target Module-MicroRNA-gene analysis using the software miRWalk 2.0[41], which showed that miR-125a had 234 reported target genes, whereas miR-125b had 391 reported target genes, and that they shared 110 similar target genes. Conversely, most miR-125a and miR-125b target genes were different, which may explain the differences between miR-125a and miR-125b in CD.

To the best of our knowledge, this was the first study investigating the correlation between circulating miR-125a/b expression and the risk and disease severity of CD and with inflammatory cytokines. Notwithstanding, there were still some limitations in our study: (1) we did not detect the expression of miR-125a/b in the intestinal tract, where in miRNA dys-regulation might be more frequent than in blood. However, it is difficult to obtain normal intestinal tract tissue from HCs and CD patients who remain remission. (2) The sample size of our study was relatively small, and a larger sample is needed for further studies. (3) IL-17, TNF-α, and IFN-γ expression was not detected in HCs in this study, which would help to further elucidate the role of miR-125a/b in inflammation, which we will investigate in future studies. (4) The levels of Tregs and Th17 cells, which are essential for CD development and progression, were not determined in the present study. We will also assess these levels in future studies.

In summary, circulating miR-125a but not miR-125b is decreased in active disease status and negatively correlates with disease severity and inflammatory cytokines in patients with Crohn's disease. Therefore, this study shed some light on the measurement of circulating miR-125a in the diagnosis and treatment of CD patients.

**COMMENTS**

***Background***

Crohn’s disease (CD), an idiopathic chronic inflammatory disease, is an inflammatory bowel diseases (IBD) and primarily affects the gastrointestinal tract. Few studies have investigated the impact of dysregulated miR-125a/b expression on Crohn's disease patients.

***Research frontiers***

Accumulating evidence shows that environmental factors, genetics, autoimmunity and dietary habits may contribute to the development and progression of CD. As an important genetic factor, dysregulated miRNA has been involved in the aetiological mechanism of several autoimmune diseases, including CD; however, the role of miR-125a/b expression in CD risk and management is largely unstudied.

***Innovations and breakthroughs***

This was the first study that investigated the correlation between circulating miR-125a/b expression and the risk and disease severity of CD and inflammatory cytokines. The authors showed that miR-125a but not miR-125b is negatively associated with the risk for A-CD and disease severity and with inflammatory cytokines.

***Applications***

miR-125a is negatively associated with the risk for A-CD patients and disease severity and with inflammatory cytokines. In the near future, miR-125a may be an important marker for the diagnosis and treatment of CD patients.

***Terminology***

Crohn’s disease activity index (CDAI) was used to differentiate A-CD from R-CD patients. Patients with a CDAI above 150 points were defined as A-CD patients, and those with a CDAI below 150 points were defined as R-CD patients.

***Peer-review***

This work focused on two mRNA molecules and their potential relevance to IBD. It’s well written and interesting manuscript.

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**Table 1 Demographic and clinical characteristics of Active Crohn's disease patients, Crohn's disease in remission patients and health controls**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Parameters** | **A-CD patients (*n* = 29)** | **R-CD patients (*n* = 37)** | **HCs (*n* = 37)** | ***P* value** |
| Age (yr) | 31.38 ± 9.51 | 31.97 ± 8.86 | 31.41 ± 5.69 | 0.941a |
| Gender (female/male) | 17/12 | 20/17 | 18/19 | 0.719a |
| CRP (mg/L) | 45.96 (32.27-67.26) | 18.94 (10.78-26.71) | 3.99 (3.33-6.06) | < 0.001a |
| ESR (mm/H) | 43.93 (37.46-57.82) | 16.73 (11.36-20.40) | 13.02 (5.82-15.27) | < 0.001a |
| CDAI Score | 206.0 (170.5-253.0) | 95.0 (75.5-112.0) | - | < 0.001c |
| IL-17 (pg/mL) | 46.90 (38.46-60.06) | 18.07 (10.21-20.17) | - | < 0.001c |
| TNF-α (pg/mL) | 72.77 (53.46-83.39) | 22.17 (19.49-30.43) | - | < 0.001c |
| INF-γ (pg/mL) | 33.23 (24.59-39.54) | 11.62 (9.16-15.07) | - | < 0.001c |

Data were presented as mean±standard deviation, median (1/4-3/4 quarters) or counts. *P* < 0.05 was considered significant: aComparison among three groups, determined by One-way analysis of variance (ANOVA), Kruskal-Wallis test by ranks or Chi-squared test; cComparison between A-CD and R-CD groups, determined by Wilcoxon signed-rank test. A-CD: Active Crohn's disease; R-CD: Crohn's disease in remission; HCs: Health controls; CRP: C reactive protein; ESR: Erythrocyte sedimentation rate; CDAI: Crohn’s disease activity index.

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**Figure 1 miR-125a/b expression in active Crohn's disease patients and Crohn's disease in remission patients and in health controls.** miR-125a was decreased in A-CD patients compared with R-CD patients and HCs (A), while no differences in miR-125b were detected between groups. Comparison between two groups was performed by the Wilcoxon signed-rank test. *P* < 0.05 was considered significant. A-CD: Active Crohn's disease; R-CD: Crohn's disease in remission; HCs: Health controls.

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**Figure 2 Receiver operating characteristic curve analysis of miR-125a/b for active Crohn's disease patients and Crohn's disease in remission patients prediction.** Receiver operating characteristic curve was operated to assess miR-125a/b expression in differentiating A-CD patients from R-CD patients and from HCs. miR-125a was able to differentiate A-CD patients from R-CD (A) patients and from HCs (B), whereas miR-125b was not (A, B). Neither miR-125a nor miR-125b could differentiate R-CD from HCs (C). A-CD: Active Crohn's disease; R-CD: Crohn's disease in remission; HCs: Health controls.

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**Figure 3 Correlations of miR-125a/b expression with disease severityin active Crohn's disease patients.** A-C: Correlations of miR-125a expression with disease severityin A-CD patients; D-F: Correlations of miR-125b expression with disease severityin A-CD patients. Spearman’s test was used to analyse the correlation of miR-125a/b expression with disease severity. *P* < 0.05 was considered significant. A-CD: Active Crohn's disease; R-CD: Crohn's disease in remission; HCs: Health controls.

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**Figure 4 Correlations of miR-125a/b expression with disease severity in Crohn's disease patients in remission.** A-C: Correlations of miR-125a expression with disease severityin R-CD patients; D-F: Correlations of miR-125b expression with disease severityin R-CD patients. Spearman’s test was used to analyse the correlation of miR-125a/b expression with disease severity. *P* < 0.05 was considered significant. A-CD: Active Crohn's disease; R-CD: Crohn's disease in remission; HCs: Health controls.

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**Figure 5 Correlations of miR-125a/b expression with inflammatory cytokinesin active Crohn's disease patients.** A-C: Correlations of miR-125a expression with inflammatory cytokinesin A-CD patients; D-F: Correlations of miR-125b expression with inflammatory cytokinesin A-CD patients. Spearman’s test was used to analyse the correlation of miR-125a/b expression with inflammatory cytokines. *P* < 0.05 was considered significant. A-CD: Active Crohn's disease; R-CD: Crohn's disease in remission; HCs: Health controls.

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**Figure 6 Correlations of miR-125a/b expression with inflammatory cytokines in Crohn's disease patients in remission.** A-C: Correlations of miR-125a expression with inflammatory cytokinesin R-CD patients; D-F: Correlations of miR-125b expression with inflammatory cytokinesin R-CD patients. Spearman’s test was used to analyse the correlation of miR-125a/b expression with inflammatory cytokines. *P* < 0.05 was considered significant. A-CD: Active Crohn's disease; R-CD: Crohn's disease in remission; HCs: Health controls.

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**Figure 7 miR-125a/b expression after treatment in active Crohn's disease patients.** After a 3-mo treatment, miR-125a expression was increased in A-CD patients who achieved clinical remission but remained stable in patients who failed to achieve remission. Conversely, no changes in miR-125b expression were observed after a 3-mo treatment in either remission or non remission patients. Comparison between visits in the same group was performed by the Wilcoxon signed-rank test. *P* < 0.05 was considered significant. A-CD: Active Crohn's disease.