

World Journal of *Cardiology*

World J Cardiol 2018 September 26; 10(9): 74-122



**REVIEW**

- 74 Myocardial reperfusion injury and oxidative stress: Therapeutic opportunities
González-Montero J, Brito R, Gajardo ALJ, Rodrigo R

MINIREVIEWS

- 87 Established and novel pathophysiological mechanisms of pericardial injury and constrictive pericarditis
Ramasamy V, Mayosi BM, Sturrock ED, Ntsekhe M

ORIGINAL ARTICLE**Basic Study**

- 97 NBCe1 $\text{Na}^+/\text{HCO}_3^-$ cotransporter ablation causes reduced apoptosis following cardiac ischemia-reperfusion injury *in vivo*
Vairamani K, Prasad V, Wang Y, Huang W, Chen Y, Medvedovic M, Lorenz JN, Shull GE

Clinical Trials Study

- 110 Accuracy of myocardial viability imaging by cardiac MRI and PET depending on left ventricular function
Hunold P, Jakob H, Erbel R, Barkhausen J, Heilmair C

LETTERS TO THE EDITOR

- 119 Snugger method - The Oldenburg modification of percutaneous implantation technique
Mashhour A, Zhigalov K, Szczechowicz M, Mkalaluh S, Easo J, Eichstaedt H, Borodin D, Ennker J, Weymann A

ABOUT COVER

Editorial Board Member of *World Journal of Cardiology*, Ioanna Andreadou, PharmD, PhD, Associate Professor, Department of Pharmaceutical Chemistry, University of Athens School of Pharmacy, Athens, Greece

AIM AND SCOPE

World Journal of Cardiology (*World J Cardiol*, *WJC*, online ISSN 1949-8462, DOI: 10.4330) is a peer-reviewed open access journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

WJC covers topics concerning arrhythmia, heart failure, vascular disease, stroke, hypertension, prevention and epidemiology, dyslipidemia and metabolic disorders, cardiac imaging, pediatrics, nursing, and health promotion. Priority publication will be given to articles concerning diagnosis and treatment of cardiology diseases. The following aspects are covered: Clinical diagnosis, laboratory diagnosis, differential diagnosis, imaging tests, pathological diagnosis, molecular biological diagnosis, immunological diagnosis, genetic diagnosis, functional diagnostics, and physical diagnosis; and comprehensive therapy, drug therapy, surgical therapy, interventional treatment, minimally invasive therapy, and robot-assisted therapy.

We encourage authors to submit their manuscripts to *WJC*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

INDEXING/ABSTRACTING

World Journal of Cardiology (*WJC*) is now abstracted and indexed in Emerging Sources Citation Index (Web of Science), PubMed, PubMed Central, Scopus, China National Knowledge Infrastructure (CNKI), and Superstar Journals Database.

EDITORS FOR THIS ISSUE

Responsible Assistant Editor: *Xiang Li*
Responsible Electronic Editor: *Yun-Xiao Jian Wu*
Proofing Editor-in-Chief: *Lian-Sheng Ma*

Responsible Science Editor: *Ying Dou*
Proofing Editorial Office Director: *Jin-Lei Wang*

NAME OF JOURNAL

World Journal of Cardiology

ISSN

ISSN 1949-8462 (online)

LAUNCH DATE

December 31, 2009

FREQUENCY

Monthly

EDITORIAL BOARD MEMBERS

All editorial board members resources online at <http://www.wjgnet.com/1949-8462/editorialboard.htm>

EDITORIAL OFFICE

Jin-Lei Wang, Director
World Journal of Cardiology
Baishideng Publishing Group Inc

7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA

Telephone: +1-925-2238242

Fax: +1-925-2238243

E-mail: editorialoffice@wjgnet.com

Help Desk: <http://www.f6publishing.com/helpdesk>

<http://www.wjgnet.com>

PUBLISHER

Baishideng Publishing Group Inc

7901 Stoneridge Drive, Suite 501,

Pleasanton, CA 94588, USA

Telephone: +1-925-2238242

Fax: +1-925-2238243

E-mail: bpgooffice@wjgnet.com

Help Desk: <http://www.f6publishing.com/helpdesk>

<http://www.wjgnet.com>

PUBLICATION DATE

September 26, 2018

COPYRIGHT

© 2018 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT

All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORS

<http://www.wjgnet.com/bpg/gerinfo/204>

ONLINE SUBMISSION

<http://www.f6publishing.com>

Myocardial reperfusion injury and oxidative stress: Therapeutic opportunities

Jaime González-Montero, Roberto Brito, Abraham IJ Gajardo, Ramón Rodrigo

Jaime González-Montero, Roberto Brito, Abraham IJ Gajardo, Ramón Rodrigo, Molecular and Clinical Pharmacology Program, Institute of Biomedical Sciences, Faculty of Medicine, University of Chile, Santiago 70058, Chile

Roberto Brito, Abraham IJ Gajardo, Internal Medicine Department, University of Chile, Clinical Hospital, Santiago 70058, Chile

ORCID number: Jaime González-Montero (0000-0003-0324-2948); Roberto Brito (0000-0001-8205-1984); Abraham IJ Gajardo (0000-0002-6387-3779); Ramón Rodrigo (0000-0003-1724-571X).

Author contributions: All authors equally contributed to this paper with conception and design of the study, literature review and analysis, drafting and critical revision and editing, and final approval of the final version.

Supported by FONDEF grant No. ID15110285.

Conflict-of-interest statement: No potential conflicts of interest.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: Dr. Ramón Rodrigo. Molecular and Clinical Pharmacology Program, Institute of Biomedical Sciences, Faculty of Medicine, University of Chile, Independencia 1027, Santiago 70058, Chile. rrodrigo@med.uchile.cl
Telephone: +56-2-29786126
Fax: +56-2-29786126

Received: March 27, 2018

Peer-review started: March 28, 2018

First decision: April 11, 2018

Revised: April 28, 2018

Accepted: May 9, 2018

Article in press: May 10, 2018

Published online: September 26, 2018

Abstract

Acute myocardial infarction (AMI) is the leading cause of death worldwide. Its associated mortality, morbidity and complications have significantly decreased with the development of interventional cardiology and percutaneous coronary angioplasty (PCA) treatment, which quickly and effectively restore the blood flow to the area previously subjected to ischemia. Paradoxically, the restoration of blood flow to the ischemic zone leads to a massive production of reactive oxygen species (ROS) which generate rapid and severe damage to biomolecules, generating a phenomenon called myocardial reperfusion injury (MRI). In the clinical setting, MRI is associated with multiple complications such as lethal reperfusion, no-reflow, myocardial stunning, and reperfusion arrhythmias. Despite significant advances in the understanding of the mechanisms accounting for the myocardial ischemia reperfusion injury, it remains an unsolved problem. Although promising results have been obtained in experimental studies (mainly in animal models), these benefits have not been translated into clinical settings. Thus, clinical trials have failed to find benefits from any therapy to prevent MRI. There is major evidence with respect to the contribution of oxidative stress to MRI in cardiovascular diseases. The lack of consistency between basic studies and clinical trials is not solely based on the diversity inherent in epidemiology but is also a result of the methodological weaknesses of some studies. It is quite possible that pharmacological issues, such as doses, active ingredients, bioavailability, routes of administration, co-therapies, startup time of the drug intervention,

and its continuity may also have some responsibility for the lack of consistency between different studies. Furthermore, the administration of high ascorbate doses prior to reperfusion appears to be a safe and rational therapy against the development of oxidative damage associated with myocardial reperfusion. In addition, the association with N-acetylcysteine (a glutathione donor) and deferoxamine (an iron chelator) could improve the antioxidant cardioprotection by ascorbate, making it even more effective in preventing myocardial reperfusion damage associated with PCA following AMI.

Key words: Acute myocardial infarction; Reperfusion injury; Oxidative stress; Ascorbate; N-acetylcysteine; Deferoxamine

© The Author(s) 2018. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Acute myocardial infarction is the leading cause of death in the world. At least half of the resulting myocardial damage is associated with myocardial reperfusion. Myocardial reperfusion injury is associated with reactive oxygen species production and iron mobilization. Treatment with antioxidants such as ascorbate, N-acetylcysteine, and an iron chelator such as deferoxamine, could prevent the development of this damage.

González-Montero J, Brito R, Gajardo AIJ, Rodrigo R. Myocardial reperfusion injury and oxidative stress: Therapeutic opportunities. *World J Cardiol* 2018; 10(9): 74-86 Available from: URL: <http://www.wjgnet.com/1949-8462/full/v10/i9/74.htm> DOI: <http://dx.doi.org/10.4330/wjc.v10.i9.74>

INTRODUCTION

Acute myocardial infarction (AMI) is the leading cause of death worldwide, and it is associated with high morbidity and mortality. The AMI complications have significantly decreased with the development of interventional cardiology and percutaneous coronary angioplasty (PCA) treatment, which quickly and effectively restore the blood flow to the area previously subjected to ischemia^[1]. Paradoxically, the restoration of blood flow to the ischemic zone leads to a massive production of reactive oxygen species (ROS), which generate rapid and severe damage to biomolecules, in a phenomenon called myocardial reperfusion injury (MRI)^[2,3]. Sources of ROS in reperfusion include the predominant contribution of NADPH oxidases, which are present in many cell types in myocardial tissue. Other sources are xanthine oxidase, uncoupled eNOS and the mitochondrion^[4]. In the clinical setting, MRI is associated with multiple complications such as lethal reperfusion, no-reflow phenomenon, myocardial stunning, and reperfusion arrhythmias (Figure 1).

Despite significant advances in the understanding

of the mechanisms accounting for MRI, it remains an unsolved problem. Although promising results have been obtained in experimental studies (mainly in animal models) these benefits have not been translated into clinical settings. Clinical trials have failed to find benefits from any therapy to prevent MRI, demonstrating a clear dissociation between the bench and the bedside^[5].

Prevention of MRI in the clinical setting has intrinsic difficulties in its approach. First, any therapy oriented to MRI prevention must be administered prior to myocardial reperfusion (in other words, prior to PCA). In addition, it should be applied in doses high enough to counterbalance the rapid and massive ROS production following reperfusion. Moreover, there are many different visions regarding the best biomarker to define MRI in patients, and so clinical trials express their results with different outcomes (such as clinical outcomes, serum cardiac biomarkers, echocardiographic parameters, cardiac magnetic resonance, among many others) which makes the analyses even more difficult. All these elements have made it difficult to develop an effective therapy to prevent MRI in AMI patients. The present review focuses on the cellular and molecular mechanisms of oxidative-stress induced MRI during AMI, and the key points to develop an appropriate strategy to reduce oxidative damage derived from myocardial reperfusion.

PATHOPHYSIOLOGY

MRI is a clinical problem associated with procedures such as thrombolysis, angioplasty, and coronary bypass surgery, which are commonly used to re-establish the blood flow and minimize the damage to the heart due to severe myocardial ischemia^[3]. There are three main hypotheses which have been proposed to explain the pathogenesis of ischemia reperfusion (IR) injury: oxidative stress, iron mobilization, and Ca²⁺-overload^[6,7]. All of these mechanisms are most likely related, but it is not known whether they operate simultaneously or one precedes the other (Figure 2).

Oxidative stress

The level of myocardial tissue oxygenation increases following restoration of blood flow, which is initiated with a burst of ROS generation^[8]; these ROS are the major initiators of myocardial damage in MRI^[3]. Increased ROS production is mainly due to the activation of xanthine oxidase in endothelial cells, mitochondrial electron transport chain reactions in cardiomyocytes, and NADPH oxidase in inflammatory cells^[9] (Figure 1).

Oxidative stress occurs when there is an imbalance between the generation of ROS and the antioxidant defense systems in the body so that the latter becomes overwhelmed^[10]. ROS include hydrogen peroxide (H₂O₂), the superoxide radical anion, the hydroxyl radical (OH[•]), and peroxynitrite anion (ONOO⁻), and they have all been shown to increase with reperfusion^[11] (Figure 2). As a result of lipid peroxidation, oxidation of DNA and proteins and membrane damage may take place.

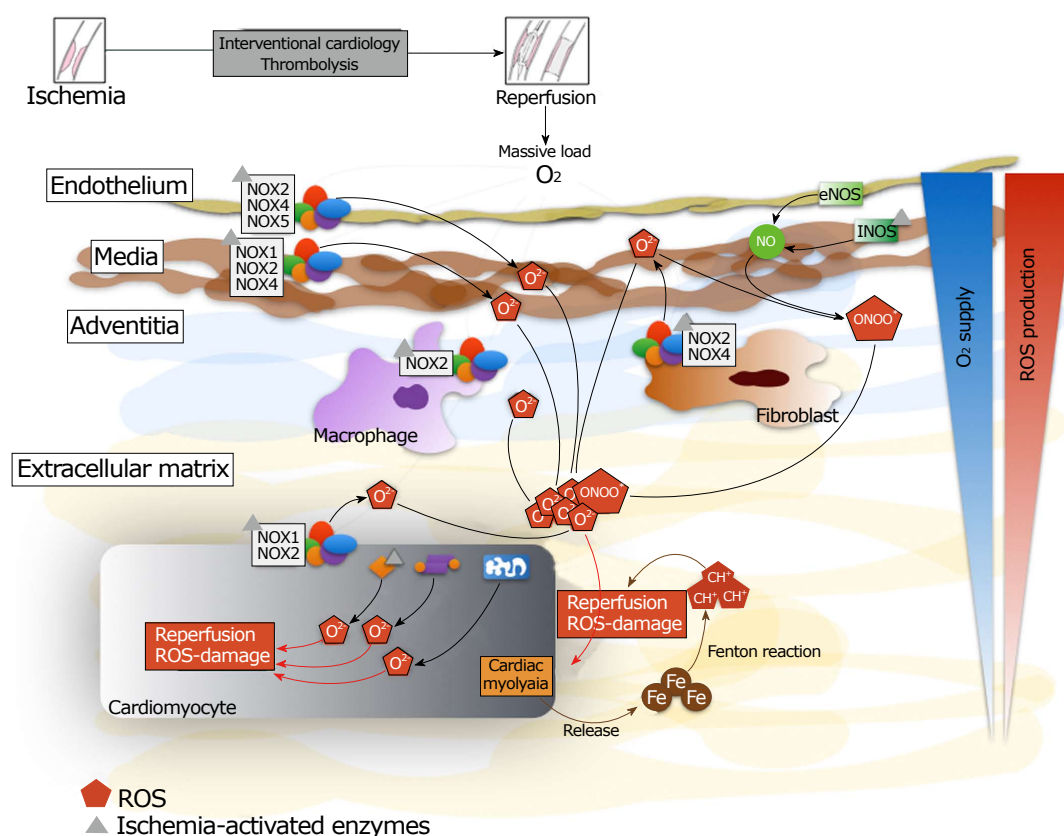


Figure 1 Generation of reactive oxygen species and mobilization of iron after myocardial reperfusion. There is a massive production of reactive oxygen species and iron mobilization by the different cellular types of the myocardial tissue. The iron reacts with superoxide anion to produce hydroxyl radical by the Fenton reaction. Inside cardiomyocyte, there is intracellular production of reactive oxygen species through NADPH oxidase, eNOS uncoupled, xanthine oxidase and mitochondrion. NOX: NADPH oxidase; ROS: Reactive oxygen species; Fe: Iron; eNOS: Endothelial nitric oxide synthases.

This leads to alterations in membrane permeability and to modifications of protein structure and functional changes^[12].

ROS sources: In pathophysiological conditions, there are many sources of ROS in myocardial tissue. The most important sources are NADPH oxidases (NOX), uncoupled eNOS, xanthine oxidases and the mitochondrion. NOX catalyzes the one electron reduction of O_2 to generate super-oxide radical anion ($O_2^{\cdot-}$), using NADPH as the source of electrons. This enzyme is largely present in the activated neutrophil, wherein it generates large amounts of toxic $O_2^{\cdot-}$ and other ROS important in bactericidal function^[13]. Pathogenic roles of NOX-derived ROS are also verified in human IR injury *in vivo*^[14]. It was recently reported that in isolated perfused murine hearts that NOX1 and/or NOX2 gene knock-out significantly attenuated MRI (by up to 50% of the final infarct size)^[15], thus demonstrating the crucial importance of this enzyme in MRI.

The NO synthases (NOS) are a family of enzymes that convert the amino acid L-arginine to L-citrulline and NO. Endothelial NOS (eNOS) plays a major role in the regulation of vascular function. The eNOS may become

uncoupled in the absence of the NOS substrate L-arginine or the cofactor BH₄. Uncoupled eNOS results in the production of $O_2^{\cdot-}$ instead of NO^[16-18]. This perpetuates a vicious cycle because peroxynitrite, the reaction product of superoxide and NO, leads to further eNOS uncoupling^[19]. Furthermore, eNOS uncoupling may play a major role in MRI by increasing ROS production and limiting NO availability^[20].

Xanthine oxidase is predominantly present in the vascular endothelium in the normal heart and generates $O_2^{\cdot-}$, H_2O_2 , and OH^{\cdot} as byproducts of its normal metabolic action^[21]. Under pathological conditions, such as tissue ischemia, xanthine dehydrogenase can be converted to Xanthine oxidase. In IR this enzyme catalyzes the formation of uric acid with the coproduction of $O_2^{\cdot-}$ ^[22]. Superoxide release results in the recruitment and activation of neutrophils and their adherence to endothelial cells, which stimulates the formation of xanthine oxidase in the endothelium, with further $O_2^{\cdot-}$ production^[23].

Mitochondria are cellular organelles involved in energy production, so any injury that they may suffer could cause impairment of cellular energy that could lead, depending on the intensity of the injury, to apoptosis or different levels of cellular damage. During ischemia, due

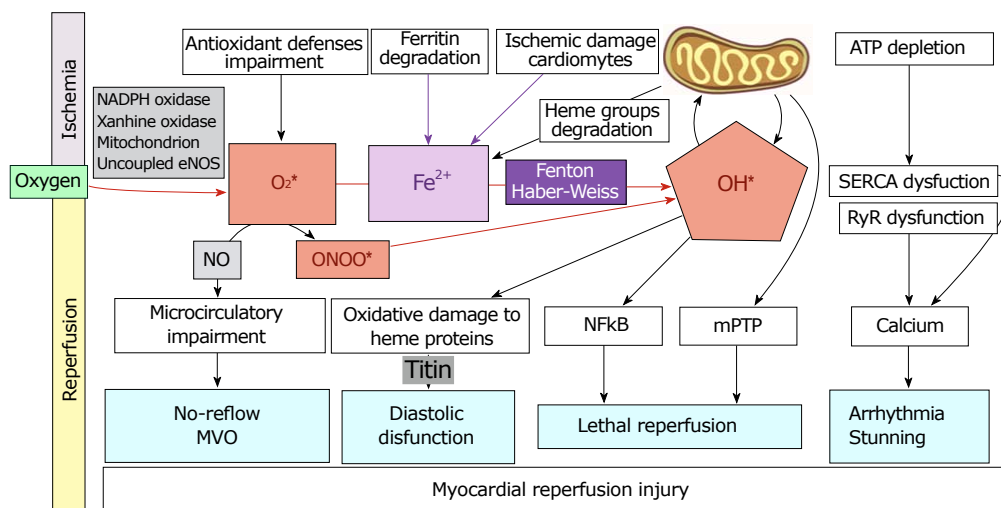


Figure 2 Role of reactive oxygen species and iron mobilization in myocardial reperfusion injury and its clinical implications. MVO: Microvascular obstruction; ONOO⁻: Peroxynitrite; NO: Nitric oxide; OH[•]: Radical hydroxyl; Fe: Iron; RyR: Ryanodine receptor channel; SERCA: Sarco/endoplasmic reticulum Ca²⁺-ATPase.

to the lack of oxygen, the electron transport chain cannot function correctly and therefore ROS are produced at high levels. Additionally, ROS may cause oxidative damage of mitochondrial DNA, impairing mitochondrial function. This damage performs a positive feedback on ROS production that, at the same time, perpetuates mitochondrial damage and ROS synthesis. Oxidative injury to the mitochondrial membrane can also occur, resulting in membrane depolarization and the uncoupling of oxidative phosphorylation, with altered cellular respiration^[24]. This can ultimately lead to mitochondrial damage, with release of cytochrome c, activation of caspases, and apoptosis^[25].

RNS sources: The ROS are not solely responsible for free radical damage. Reactive nitrogen species (RNS), mainly peroxynitrite anions (ONOO⁻), also generate RNS-damage, thus producing nitrosative stress. Peroxynitrite results from the interaction between NO and the superoxide anion^[4], and NO is synthesized mainly by nitric oxide synthases which have two isoforms in the cardiomyocyte: endothelial (eNOS) and inducible (iNOS). Oxidative and nitrosative damage causes the uncoupling of both NOS isoforms, resulting in the enhanced synthesis of O₂^{•-}^[4].

Evidence supports the view that nitrosative stress plays an important role in the pathogenesis of MRI. While NO itself is not harmful, some of the reaction products (mainly OH[•]) resulting from high ONOO⁻ formation in the cell are highly cytotoxic substances^[26]. The production of O₂^{•-} is increased during reperfusion, which interacts with NO and leads to the formation of ONOO⁻, thus triggering the previously described phenomenon^[27]. Peroxynitrite not only causes structural damage by attacking macromolecules, but it also leads to myocardial functional impairment^[28]. The general view about the mechanisms that lead to nitrosative stress is that IR

can induce iNOS expression and that the resulting high concentrations of NO can lead to cardiac injury^[26]. The drop in NO concentration occurring during cardiac IR plays an important role in triggering the transcription nuclear factor kappaB (NF-κB) leading to activation and successive induction of iNOS expression during the reperfusion phase^[29-31]. Figure 1 shows a diagram of ROS and RNS sources in myocardial tissue.

Iron mobilization

It has been postulated that iron homeostasis could play an important role in the development of MRI in the cardiomyocytes^[32,33]. Free iron is deleterious for cells; thus generally it is bound to proteins forming complexes^[34]. During ischemia, iron metabolism is impaired, and it is released as free iron. This catalytic free iron can generate ROS through the Fenton reaction, catalyzing the production of ·OH from H₂O₂ and O₂^{•-}^[35]. It has been reported that susceptibility to injury from H₂O₂ in rat hearts is associated with the magnitude of the intracellular low molecular weight iron pool^[36]. Some metals with redox properties have a well-documented role in the development of MRI^[37,38]. Following reperfusion, both iron and copper are released to the coronary circulation^[32] which can contribute to ROS generation (Figure 2). In patients with thalassemia the iron overload is related to arrhythmias and congestive heart failure, which is the main cause of death among these patients^[39]. Iron chelation therapy has significantly improved the survival of patients with thalassemia^[40], because iron chelators are effective and safe drugs to treat the iron poisoning^[41].

Calcium homeostasis

Oxidative stress modifies phospholipids and proteins leading to lipid peroxidation and thiol-group oxidation; these changes are considered to alter membrane

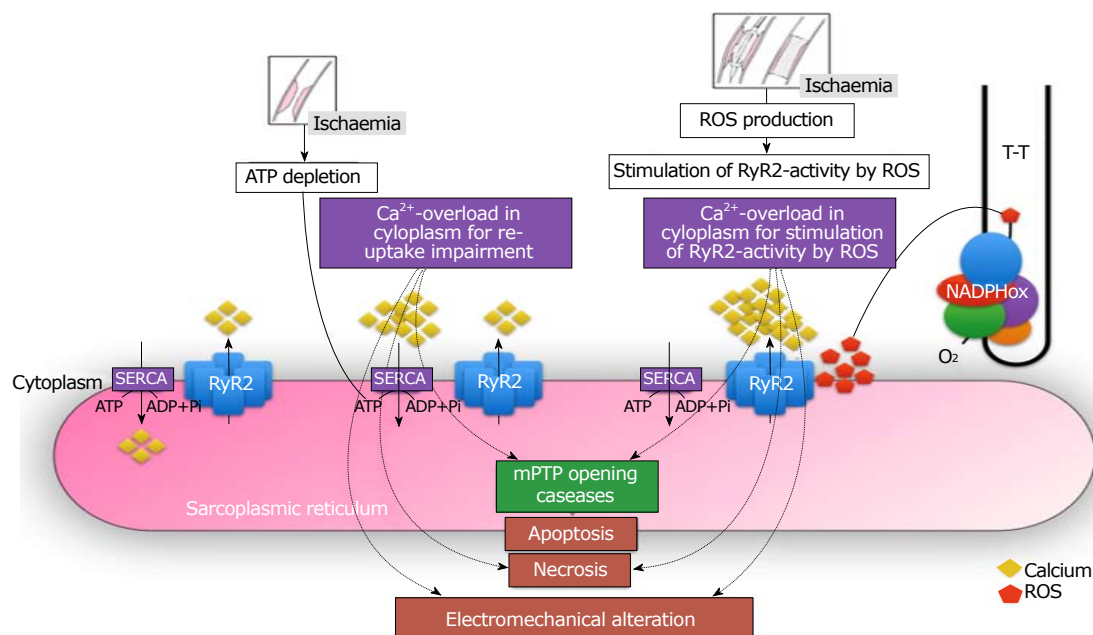


Figure 3 Central role of calcium in the electro-mechanical dissociation of cardiomyocyte after myocardial reperfusion. RyR: Ryanodine receptor channel; SERCA: Sarco / endoplasmic reticulum Ca^{2+} -ATPase; mPTP: Mitochondrial permeability transition pore; Ca: Calcium; ROS: Reactive oxygen species.

permeability and configuration in addition to producing functional modifications of various cellular proteins^[42]. Oxidative stress may result in cellular defects including a depression in the sarcolemma Ca^{2+} -pump ATPase that leads to a decreased Ca^{2+} -efflux, and a depression in (Na + K)-ATPase activity that, in turn, leads to an increased Ca^{2+} -influx^[43]. Oxidative stress has also been reported to depress the sarcoplasmic reticulum Ca^{2+} -pump ATPase (SERCA) and thus inhibit Ca^{2+} sequestration from the cytoplasm in cardiomyocytes^[44]. The depression in Ca^{2+} -regulatory mechanism by ROS ultimately results in intracellular Ca^{2+} ($[\text{Ca}^{2+}]_i$) overload and cell death. In addition, an increase in $[\text{Ca}^{2+}]_i$ during ischemia induces the conversion of xanthine dehydrogenase to xanthine oxidase and subsequently results in increased production of $\text{O}_2^{\bullet-}$ ^[44].

Recently it has been shown that the function of the channel ryanodine receptor (RyR) is controlled by ROS^[45]. It has been demonstrated that NADPH oxidase and the RyR channel could be located adjacent to each other in the T-tubules of cardiomyocytes^[46]. Thus, the increase in ROS production after myocardial reperfusion could lead to an increase in RyR channel function, resulting in an intracellular calcium overload, thereby causing activation of pro-apoptotic intracellular pathways, necrosis, and electromechanical alteration. All these mechanisms are summarized in Figure 3.

Redox-sensitive signaling pathways: Not only do ROS exert their actions by directly modifying organic molecules, but ROS are also involved in the regulation of the expression of several genes^[47]. NF- κ B and AP-1, both of which can experience ROS-mediated activation, stimulate the transcription of several protein mediators,

for example, proinflammatory cytokines that activate several cell death pathways^[48]. The role of cytokines, chemokines, leukocytes, and acute-phase proteins such as high-sensitivity C-reactive protein in the pathogenesis of MRI has been reported in several studies^[49,50]. Oxidative stress, ROS and inflammation are linked in a way that is very difficult to dissect. These phenomena have important molecular bridges that are activated in the presence of ROS^[51], leading to the activation of multiple mechanisms that cause heart tissue remodeling and therefore enhance the susceptibility to rhythm disorders. Among those molecules, the most studied has been the transcriptional factor NF- κ B, a factor that responds to changes of the cellular oxidative state, ischemia-reperfusion, and inflammatory molecules^[52]. When NF- κ B is activated, for example in the presence of ROS by phosphorylation of its inhibitory cofactor (Ik-B), it bonds to a DNA response element and promotes the transcription of genes involved in inflammatory and pro-fibrotic response, for example IL-6, which transforms growth factors TGF- β and TNF- α ^[53]. Those molecules act in various tissues, but particularly in the heart, producing extracellular matrix remodeling and fibrosis (structural remodeling), which changes the electrophysiological properties of the heart. Several studies have associated NF- κ B activation with cardiac dysfunction, ventricular hypertrophy, and maladaptive cardiac growth^[54] (Figure 2).

Exposure to low-to-moderate ROS levels should trigger a survival response and reinforce ROS scavengers of the antioxidant defense system to elicit a cardioprotective effect for myocardial reperfusion. The molecular mechanism responsible for this adaptive change involves enhanced antioxidant activity achieved

by up-regulating several housekeeping genes partly under the control of Nrf2 (nuclear factor-erythroid 2-related factor 2); Nrf2 is normally sequestered in the cytosol by Keap1^[55]. Upon oxidative stimulation, Nrf2 oxidizes or covalently modifies Keap1 thiol groups, which dissociate from Keap1 and undergo nuclear translocation. In the nucleus, Nrf2 binds to antioxidant response elements in target gene promoters^[56], which increase the expression of antioxidant enzymes. It has been demonstrated that the constitutive levels/activities of a number of important antioxidants and phase 2 enzymes, such as CAT, GSH-Px, glutathione reductase, glutathione transferase, NADPH-quinone oxidoreductase 1, and heme oxygenase-1 in primary cardiomyocytes are dependent on Nrf2 status. In addition, Nrf2 diminishes the susceptibility of cardiomyocytes to injury elicited by oxidants and electrophilic species^[57], making the Nrf2 signaling pathway an important mechanism for myocardial cytoprotection. It is of interest to note that ROS levels could be responsible for the activation of NF- κ B and/or Nrf2 pathways.

Clinical implications: Myocardial damage caused by ischemia-reperfusion events are mainly associated with four clinical conditions: lethal reperfusion, myocardial stunning, no-reflow phenomenon, and reperfusion arrhythmias (Figure 2).

Lethal reperfusion is a paradoxical type of MRI caused by the restoration of coronary blood flow after an ischemic episode. It is defined as the death of cardiomyocytes that were viable immediately before myocardial reperfusion. Its main manifestation is as an increased infarct size due to reperfusion, a condition mainly associated with AMI^[3]. In the late fifties, it was suggested that myocardial reperfusion contributes part of the histological damage associated with ischemia-reperfusion models. However, for decades it was very complex to determine the precise evolution of necrosis along the transition from ischemia to reperfusion in myocardial tissue^[58]. Nowadays, the harmful effects of myocardial reperfusion damage, also known as lethal reperfusion injury, are considered to involve myocardial cell death derived from the restoration of blood flow subsequent to an ischemic process, and to act through mechanisms strongly associated with oxidative stress^[3].

Reperfusion arrhythmias clinically represent a major comorbidity of AMI with an 88.7% occurrence rate in certain small clinical trials with continuous monitoring^[59]. In addition, postoperative atrial fibrillation (POAF), the most common reperfusion arrhythmia associated with cardiac surgeries, has an incidence ranging between 20%-40%^[60]. Myocardial stunning, despite being a reversible damage, is the cause of an impaired ventricular function that leads to increased morbidity. It is derived from a short-term ischemia-reperfusion process that was first reported in the early 1930s^[61]. Myocardial stunning is present to a greater or lesser extent in all survivors of AMI. In the late 1980s evidence began to appear suggesting an important role of oxidative stress

in the development of myocardial stunning, proposing that the main injury pathway could be an altered calcium homeostasis associated with sarcoplasmic reticulum damage^[62]. More recently, clinical studies have strengthened this hypothesis^[63], and it has been reported in animal models that interventions aimed to improve antioxidant defenses attenuate myocardial stunning^[64,65].

The no-reflow phenomenon is an impaired myocardial perfusion of a specific segment of the coronary system that is not associated with an angiographic occlusion of the respective vessel^[66]. Vascular and endothelial damage can occur after the reperfusion of previously blocked coronary circulation. It can be exhibited as a microvascular dysfunction after restoring the flow during either angioplasty or thrombolysis, thus leading to the development of the no-reflow phenomenon^[67]. The presence of coronary microvascular dysfunction and this phenomenon are associated with larger infarct size, lower left ventricular ejection fraction, adverse left ventricular remodeling in the remote stage of myocardial infarction, and increased incidences of heart failure and death, compared with patients without no-reflow phenomenon^[68]. Some studies using animal models showed that antioxidant strategies are able to reduce this phenomenon^[69-71], and this data is consistent with a small clinical trial finding that antioxidant depletion is associated with no-reflow phenomenon in AMI^[72]. In addition, recent research in rabbits shows that the suppression of the oxidative stress-sensitive transcription factor NF- κ B, a key mediator of inflammation in cardiovascular systems, reduces myocardial no-reflow phenomenon^[73].

Recently, our group has reported major clinical benefits with the use of antioxidants in pathologies associated with myocardial reperfusion, such as POAF and AMI. With regard to POAF, we documented a significant decrease in the incidence of this arrhythmia in patients undergoing cardiac surgery with extracorporeal circulation after administration of ascorbate, alpha-tocopherol, and omega-3 polyunsaturated fatty acids, which was accompanied by a significant decrease in oxidative stress biomarkers in auricular tissue and peripheral blood^[60].

ROLE OF ANTIOXIDANTS

Despite a molecular basis and *in vitro* evidence supporting the use of antioxidants to prevent MRI, clinical evidence continues to be controversial. In the clinical setting, impaired micro-circulatory reperfusion was improved by ascorbate infusion in patients undergoing elective PCA^[74]. Similar results were recently reported by our group^[75]. These results suggest a positive role of antioxidants in counteracting the deleterious effects of oxidative stress on microvascular function. On the other hand, the ROS scavenger edaravone when administered to patients with AMI immediately prior to reperfusion, significantly reduced infarct size and

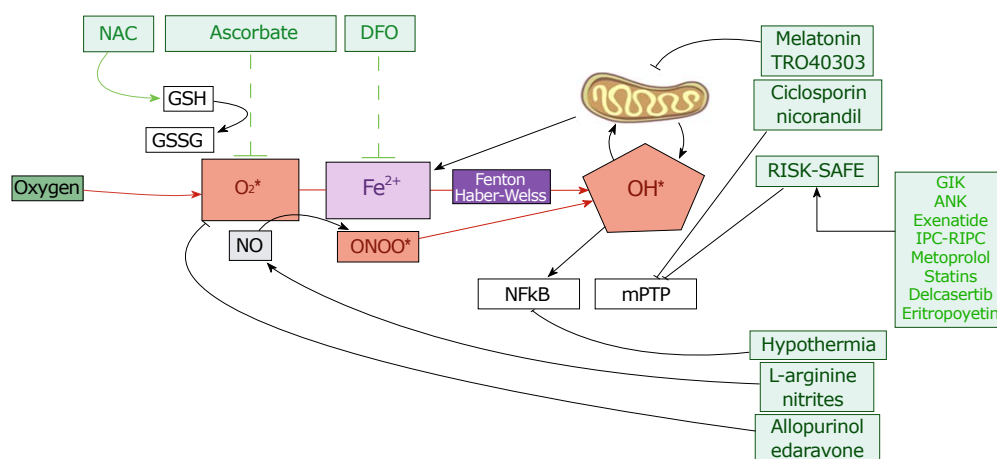


Figure 4 Experimental, pharmacological and clinical approaches to prevent myocardial reperfusion injury at cellular level. RISK: Reperfusion injury salvage kinase pathway; SAFE: Survivor activating factor enhancement pathway; GSH: Reduced glutathione; GSSG: Oxidized glutathione; NAC: N-acetylcysteine; DFO: deferoxamine; ONOO*: Peroxynitrite; NO: Nitric oxide; OH*: Radical hydroxyl; mPTP: Mitochondrial permeability transition pore.

reperfusion arrhythmias^[76,77]. Also some experimental studies reported that the use of deferoxamine (DFO) and N-acetylcysteine (NAC) could improve microvascular dysfunction^[78,79].

Carotenoids represent another potential pharmacological alternative in the management of MRI^[80]. Carotenoids are a widely distributed group of fat-soluble pigments which exert antioxidant, anti-inflammatory, and antiproliferative properties^[81]. Several experimental data support potential role of carotenoids in this pathological condition: Tong *et al.*^[81] demonstrated that pretreatment with lycopene reduced cardiomyocyte death induced by ischemia/reoxygenation *in vitro*, and also reduced myocardial infarct size in an *in vivo* model of AMI^[82]. Another carotenoid, crocetin, protected against myocardial reperfusion injury *in vivo* by inhibiting ROS production, reducing eNOS expression and myocardium apoptosis^[82]. All-trans retinoic acid presented also protective activity against reperfusion injury both *in vitro* and *in vivo*, probably by down-regulating MAPK signaling^[84]. Despite the fact that carotenoids have been useful in preventing MRI in experimental studies and have arisen as a promising pharmacological alternative, further clinical studies and randomized clinical trials are required.

In the following paragraphs we will discuss a new hypothesis for the prevention of MRI through the combined use of ascorbate, NAC, and DFO prior to reperfusion in order to strengthen antioxidant defense systems and so prevent oxidative damage (Figure 4).

Ascorbate

The basis of this hypothesis is to achieve high plasma levels of ascorbate prior to reperfusion in order to strengthen the antioxidant defense system of myocardial tissue. Thus, when oxygen suddenly arrives to the previously ischemia-damaged myocardial tissue—which is the primary substrate for the production of the highly reactive superoxide anion radical—ascorbate may

efficiently reduce ROS and prevent oxidative damage^[5,8]. To support this hypothesis, we will discuss the main actions of this antioxidant and its pharmacokinetic properties.

Ascorbate is an essential antioxidant that performs its roles in different cell locations by acting in water-soluble components^[85,86]. The most studied mechanism in which ascorbate acts is partly based on its ability to directly reduce ROS^[87-89]. Besides its ROS scavenger actions, ascorbate exerts a complex modulation of numerous enzymes involved in ROS production, endothelial dysfunction, platelet aggregation, and smooth muscle cell tone^[90-92]. The four most important mechanisms in which ascorbate modulates the endothelial function are NADPH down-regulation, and the up-regulation of eNOS, phospholipase A2, and antioxidant enzymes. NADPH oxidase, the most important superoxide source in the cardiovascular system, can be directly down-regulated by ascorbate^[91,92]. The mechanism behind this effect has not been completely elucidated. It has been reported that ascorbate could be involved in the transcriptional and post-transcriptional modulation of NADPH oxidase^[89,93] as well as in its synthesis^[94]. In the presence of oxidative stress, eNOS is mostly in its uncoupled form which leads to endothelial dysfunction. In this context, ascorbate has been shown to increase eNOS activity, by preventing the oxidation of tetrahydrobiopterin and by inhibiting the p47phox subunit expression^[95]. Therefore, ascorbate increases NO synthesis, reduces ROS formation and contributes to vascular tone regulation^[96-98]. In relation to the up-regulation of antioxidant enzymes, some studies have demonstrated a positive correlation between antioxidant vitamin and antioxidant enzyme activity, particularly SOD. The mechanisms underlying these findings are not well explained, but it is plausible to hypothesize the existence of transcriptional and post-transcriptional events involved in the up-regulation of those antioxidant enzymes^[92].

Table 1 Clinical trials

	Study details	Country	n Intervention		Main findings	Ref.
AA	Ascorbate previous to elective coronary angioplasty	Italy	28	28	Decrease in oxidative stress and improves reperfusion parameters	[74]
	Ascorbate previous to primary coronary angioplasty in patients with AMI	Chile	53	46	Improve ventricular function and reperfusion No differences in infarct size	[75]
NAC	N-acetylcysteine previous and after primary coronary angioplasty in patients with AMI	Germany	126	126	Decrease in oxidative stress No differences in infarct size	[105]
	N-acetylcysteine and nitroglycerine previous to primary coronary angioplasty in patients with AMI	Australia	67	65	Decrease in infarct size and cardiac damage biomarkers	[116]
DFO	Deferoxamine previous and after coronary angioplasty in patients with AMI	Australia	28	32	Decrease in oxidative stress No differences in infarct size	[114]

Main clinical studies that have used ascorbate, N-acetylcysteine or deferoxamine to prevent reperfusion injury in patients affected by acute myocardial infarction and treated with coronary angioplasty. AA: Ascorbate; NAC: N-acetylcysteine; DFO: Deferoxamine; IR: Ischemia reperfusion; AMI: Acute myocardial infarction.

Ascorbate counteracts and prevents the oxidation of lipids, proteins, and DNA, subsequently protecting their structure and biological function. Together with glutathione, ascorbate constitutes a primary line of defense against ROS^[99]. Ascorbate, in aqueous compartments, can recycle α -tocopherol in membranes by reducing the α -tocopheroxyl radical back to α -tocopherol^[100]. Accordingly, ascorbate has been shown to recycle α -tocopherol in lipid bilayers^[101] and erythrocytes^[95].

Ascorbate scavenging is concentration-dependent and requires intravenous administration. This is necessary because ascorbate concentration in plasma is tightly controlled and an excess of ascorbate is excreted as a function of dosage. In fact, even with supplementation approaching maximally tolerated doses, ascorbate plasma concentrations are always < 250 μ mol/L. By contrast, intravenously injected ascorbate can safely lead to concentrations of 25-30 mmol/L^[102]. It is of interest to mention that intra-arterial administration of high doses of ascorbate has been demonstrated to abolish both *in vivo* and *in vitro* effects of the superoxide anion with respect to the impairment of vascular endothelial function in patients with essential hypertension^[103]. Unfortunately, oral doses are not enough to scavenge superoxide anions, thus a beneficial effect should not be expected.

Our group recently developed a randomized clinical trial in patients with AMI undergoing PCA, where massive doses of ascorbate (or placebo) were administered prior to PCA. Patients treated with ascorbate prior to myocardial reperfusion showed a better recovery of ejection fraction at 2-3 mo (measured by cardiac magnetic resonance) and significantly higher myocardial perfusion after PCA (*TIMI*-myocardial perfusion grade) than placebo patients, with no differences in infarct size^[75] (Table 1).

N-acetyl-L-cysteine

Ascorbate consumes glutathione (GSH) to exert its antioxidant activity. High doses of ascorbate might be

associated with a decrease in cellular GSH reserves^[5]. For this reason, N-acetyl-L-cysteine (NAC) - a known GSH-donor-may also have synergistic effects with high doses of ascorbate. In the following paragraphs, we will discuss the potential role of NAC in preventing MRI.

Despite numerous studies and a prolonged track record of clinical trials, the effects of NAC are clouded in controversy and its pharmacological mechanism has not yet been fully clarified. However, there is plenty of evidence regarding its mechanism of action. First of all, NAC's main feature, and also the most studied one, is its capacity to act as a precursor for synthesis of GSH, thus replenishing GSH that has become depleted through the use of this peptide in detoxification routes^[104]. However, it is vital to think of NAC as a pro-drug, because actions that are driven by this drug are dependent on its successful conversion to the antioxidant and detoxifying agent, GSH. Another frequently mentioned property of NAC is its intrinsic antioxidant activity. Nevertheless, the evidence regarding the antioxidant potential of NAC suggests that it does not have a noteworthy direct antioxidant activity^[105].

NAC acts indirectly through chelation of metal ions such as catalytic iron^[106,107] giving it the capability of mediating Fenton's reaction, thus ameliorating the possibility of the formation of hydroxyl radicals. This property is due to the fact that NAC forms conjugates with some metals. However, the importance of this mechanism in driving any protective effects compared to intracellular GSH replenishing is still unclear. Current evidence agrees on the capability of NAC to act as an inhibitor of NF- κ B^[108], a transcription factor that plays a critical role in inflammation, immunity, cell proliferation, differentiation, and survival. In conclusion, molecular mechanisms by which NAC exerts its diverse effects are complex and still unclear. Although it has been shown that NAC interacts with numerous biochemical pathways, its main mechanism involves serving as a precursor of cysteine and replenishing cellular GSH levels^[104].

NAC has been widely used in different experimental and clinical settings to counteract oxidative stress. It has been demonstrated that NAC in combination with nitroglycerin and streptokinase is associated with significantly less oxidative stress and improved preservation of left ventricular function^[109]. However, it has also been reported that a high-dose of NAC prior to PCA, although it reduces oxidative stress, does not provide an additional advantage in the prevention of MRI^[110]. Additionally, an interesting study published in 2006 shows that administration of NAC in combination with streptokinase significantly diminishes oxidative stress and improves left ventricular function in patients with AMI^[111]. A recent study using a rat model of myocardial ischemia-reperfusion injury demonstrates that treatment with continuous infusion of NAC (150 mg/kg per hour) starting 30 min before occlusion and lasting for 2 h (or until 1 h after the start of reperfusion) produces a significant limitation of the infarct and allows the recovery of the decreased total glutathione when compared to control^[112]. Recently has been published the NACIAM trial by Pasupathy *et al.*^[113], that demonstrated a protective effect with the use of high doses of NAC in combination with a nitric oxide donor in patients with AMI (Table 1). This important study shows that NAC has a powerful protective effect when used in combination and previous to myocardial reperfusion. In summary, due to the known antioxidant and cardioprotective effect and its role as GSH-donor, it is plausible to suggest that NAC might have a synergistic effect with high doses of ascorbate and deferoxamine to prevent MRI.

Deferoxamine

Given the known role of iron in the lethal reperfusion, iron chelators have been tested to ameliorate this injury. One of the most frequently used drugs for this purpose is DFO. The first reports of its use to improve cardiac function in myocardium iron overload by directly removing iron from the myocardium^[114] date from 1980s^[115]. In animal models of AMI, the use of DFO has exhibited positive results. Some studies performed in dogs reported a decrease in the infarct size when they used DFO during the reperfusion, suggesting that iron-catalyzed production of ROS contributes to cardiomyocyte necrosis in the setting of MRI^[116,117]. Studies have described improved recovery of myocardial function after ischemia, by using iron chelation^[36,118]. The results obtained from animal models of MRI have suggested the use of iron chelators in the human model with partial results to date. Paraskevaidis *et al.*^[119] suggested DFO infusion was able to reduce myocardial stunning after elective coronary artery bypass grafting and to improve long-term ejection fraction. In a recent clinical study, Chan *et al.*^[120] randomized patients with STEMI to intravenous deferoxamine before coronary angioplasty and then for 12 h vs placebo (Table 1). The serum iron levels and lipid peroxidation biomarkers were reduced in the DFO-group without differences in the infarct size. The role of iron and ascorbate in the MRI

has become of increasing interest in the last few years. It has been demonstrated that the combined use of DFO and ascorbate prevent reperfusion arrhythmias^[121].

As has been previously discussed, cumulated evidence from both experimental and clinical studies leads us to support the view that a novel combined antioxidant strategy could limit MRI and its consequences. This novel hypothesis is based on the combined use of antioxidants prior to the reperfusion therapy in order to limit the oxidative challenge during reperfusion. The key points of this novel intervention are: (1) To achieve high plasma concentrations of ascorbate through massive intravenous doses to counteract the ROS and RNS production; (2) the use of NAC to prevent GSH depletion; and (3) the use of DFO to diminish the catalytic free iron levels in order to prevent the ROS production by the Fenton reaction.

Accordingly, in our laboratory recent studies of the murine Langendorff model have been conducted to determine the effect of antioxidants in MRI. We are now studying the effect of ascorbate, NAC, and DFO used alone and in association. Under these conditions, we expect a lower vulnerability of the myocardial tissue to the reperfusion injury associated with oxidative stress. This protective effect could be expressed by a lower infarct size, reduced post-reperfusion arrhythmias and myocardial stunning occurrence, and improved microvascular function. Finally, at present, there is no evidence available from any trial that has applied this antioxidant protocol to diminish MRI. Table 1 shows a summary of the main clinical studies that have used antioxidants to prevent MRI in patients with AMI.

CONCLUSION

There is major evidence with respect to the contribution of oxidative stress to MRI in cardiovascular diseases. Despite the many significant advances in the understanding of the mechanisms of MRI, it remains an unsolved problem. There is a lack of consistency between basic studies and clinical trials aimed to reduce MRI through antioxidant therapies. Although promising results have been obtained in experimental studies (mainly in animal models), these benefits have not been translated into clinical settings. It is noteworthy that the administration of high ascorbate doses prior to reperfusion and also NAC administration appear to be safe and rational therapies against the development of oxidative damage associated with myocardial reperfusion. Furthermore, ascorbate association with NAC and DFO could improve the beneficial effect of ascorbate, making it even more effective in preventing myocardial reperfusion damage associated with PCA following AMI.

REFERENCES

- 1 Roe MT, Halabi AR, Mehta RH, Chen AY, Newby LK, Harrington RA, Smith SC Jr, Ohman EM, Gibler WB, Peterson ED.

- Documented traditional cardiovascular risk factors and mortality in non-ST-segment elevation myocardial infarction. *Am Heart J* 2007; **153**: 507-514 [PMID: 17383286 DOI: 10.1016/j.ahj.2006.12.018]
- 2 **Vanden Hoek TL**, Li C, Shao Z, Schumacker PT, Becker LB. Significant levels of oxidants are generated by isolated cardiomyocytes during ischemia prior to reperfusion. *J Mol Cell Cardiol* 1997; **29**: 2571-2583 [PMID: 9299379 DOI: 10.1006/jmcc.1997.0497]
 - 3 **Yellon DM**, Hausenloy DJ. Myocardial reperfusion injury. *N Engl J Med* 2007; **357**: 1121-1135 [PMID: 17855673 DOI: 10.1056/NEJMra071667]
 - 4 **Rodrigo R**. Oxidative stress and antioxidants: their role in human disease. Nova Biomedical Books, 2009 [cited 2018 Mar 26]: 358
 - 5 **Rodrigo R**, Libuy M, Feliú F, Hasson D. Molecular basis of cardioprotective effect of antioxidant vitamins in myocardial infarction. *Biomed Res Int* 2013; **2013**: 437613 [PMID: 23936799 DOI: 10.1155/2013/437613]
 - 6 **Ferrari R**. The role of mitochondria in ischemic heart disease. *J Cardiovasc Pharmacol* 1996; **28** Suppl 1: S1-10 [PMID: 8891865 DOI: 10.1097/00005344-199600003-00002]
 - 7 **Jahangiri A**, Leifert WR, Kind KL, McMurchie EJ. Dietary fish oil alters cardiomyocyte Ca²⁺ dynamics and antioxidant status. *Free Radic Biol Med* 2006; **40**: 1592-1602 [PMID: 16632119 DOI: 10.1016/j.freeradbiomed.2005.12.026]
 - 8 **Rodrigo R**, Prieto JC, Castillo R. Cardioprotection against ischaemia/reperfusion by vitamins C and E plus n-3 fatty acids: molecular mechanisms and potential clinical applications. *Clin Sci (Lond)* 2013; **124**: 1-15 [PMID: 22963444 DOI: 10.1042/CS20110663]
 - 9 **Chamiec T**, Herbaczyńska-Cedro K, Ceremużyński L. Effects of antioxidant vitamins C and E on signal-averaged electrocardiogram in acute myocardial infarction. *Am J Cardiol* 1996; **77**: 237-241 [PMID: 8607400 DOI: 10.1016/S0002-9149(97)89385-X]
 - 10 **Juránek I**, Bezek S. Controversy of free radical hypothesis: reactive oxygen species—cause or consequence of tissue injury? *Gen Physiol Biophys* 2005; **24**: 263-278 [PMID: 16308423]
 - 11 **Eaton P**, Clements-Jewery H. Peroxynitrite: *in vivo* cardioprotectant or arrhythmogen? *Br J Pharmacol* 2008; **155**: 972-973 [PMID: 18806818 DOI: 10.1038/bjp.2008.372]
 - 12 **Zimmerman JJ**. Defining the role of oxyradicals in the pathogenesis of sepsis. *Crit Care Med* 1995; **23**: 616-617 [PMID: 7712748 DOI: 10.1097/00003246-199504000-00003]
 - 13 **Brandes RP**, Kreuzer J. Vascular NADPH oxidases: molecular mechanisms of activation. *Cardiovasc Res* 2005; **65**: 16-27 [PMID: 15621030 DOI: 10.1016/j.cardiores.2004.08.007]
 - 14 **Loukogeorgakis SP**, van den Berg MJ, Sofat R, Nitsch D, Charakida M, Haiyee B, de Groot E, MacAllister RJ, Kuijpers TW, Deanfield JE. Role of NADPH oxidase in endothelial ischemia/reperfusion injury in humans. *Circulation* 2010; **121**: 2310-2316 [PMID: 20479156 DOI: 10.1161/CIRCULATIONAHA.108.814731]
 - 15 **Braunersreuther V**, Montecucco F, Asrih M, Pelli G, Galan K, Frias M, Burger F, Quinderé AL, Montessuit C, Krause KH, Mach F, Jaquet V. Role of NADPH oxidase isoforms NOX1, NOX2 and NOX4 in myocardial ischemia/reperfusion injury. *J Mol Cell Cardiol* 2013; **64**: 99-107 [PMID: 24051369 DOI: 10.1016/j.jmcc.2013.09.007]
 - 16 **Vásquez-Vivar J**, Kalyanaraman B, Martásek P, Hogg N, Masters BS, Karoui H, Tordo P, Pritchard KA Jr. Superoxide generation by endothelial nitric oxide synthase: the influence of cofactors. *Proc Natl Acad Sci USA* 1998; **95**: 9220-9225 [PMID: 9689061 DOI: 10.1073/pnas.95.16.9220]
 - 17 **Xia Y**, Zweier JL. Direct measurement of nitric oxide generation from nitric oxide synthase. *Proc Natl Acad Sci USA* 1997; **94**: 12705-12710 [PMID: 9356514 DOI: 10.1073/pnas.94.23.12705]
 - 18 **Chalupsky K**, Cai H. Endothelial dihydrofolate reductase: critical for nitric oxide bioavailability and role in angiotensin II uncoupling of endothelial nitric oxide synthase. *Proc Natl Acad Sci USA* 2005; **102**: 9056-9061 [PMID: 15941833 DOI: 10.1073/pnas.0409594102]
 - 19 **Schulz E**, Jansen T, Wenzel P, Daiber A, Münzel T. Nitric oxide, tetrahydrobiopterin, oxidative stress, and endothelial dysfunction in hypertension. *Antioxid Redox Signal* 2008; **10**: 1115-1126 [PMID: 18321209 DOI: 10.1089/ars.2007.1989]
 - 20 **Landmesser U**, Dikalov S, Price SR, McCann L, Fukai T, Holland SM, Mitch WE, Harrison DG. Oxidation of tetrahydrobiopterin leads to uncoupling of endothelial cell nitric oxide synthase in hypertension. *J Clin Invest* 2003; **111**: 1201-1209 [PMID: 12697739 DOI: 10.1172/JCI14172]
 - 21 **Friedl HP**, Smith DJ, Till GO, Thomson PD, Louis DS, Ward PA. Ischemia-reperfusion in humans. Appearance of xanthine oxidase activity. *Am J Pathol* 1990; **136**: 491-495 [PMID: 2316621]
 - 22 **Granger DN**. Role of xanthine oxidase and granulocytes in ischemia-reperfusion injury. *Am J Physiol* 1988; **255**: H1269-H1275 [PMID: 3059826 DOI: 10.1152/ajpheart.1988.255.6.H1269]
 - 23 **Wang X**, Han M, Bao J, Tu W, Dai Z. A superoxide anion biosensor based on direct electron transfer of superoxide dismutase on sodium alginate sol-gel film and its application to monitoring of living cells. *Anal Chim Acta* 2012; **717**: 61-66 [PMID: 22304816 DOI: 10.1016/j.aca.2011.12.045]
 - 24 **Nathan AT**, Singer M. The oxygen trail: tissue oxygenation. *Br Med Bull* 1999; **55**: 96-108 [PMID: 10695081 DOI: 10.1258/0007142991902312]
 - 25 **Macdonald J**, Galley HF, Webster NR. Oxidative stress and gene expression in sepsis. *Br J Anaesth* 2003; **90**: 221-232 [PMID: 12538380 DOI: 10.1093/bja/aeg034]
 - 26 **Darra E**, Rungtatscher A, Carcereri de Prati A, Podesser BK, Faggian G, Scarabelli T, Mazzucco A, Hallström S, Suzuki H. Dual modulation of nitric oxide production in the heart during ischemia/reperfusion injury and inflammation. *Thromb Haemost* 2010; **104**: 200-206 [PMID: 20508903 DOI: 10.1160/TH09-08-0554]
 - 27 **Rodrigo R**, Vinay J, Castillo R, Cereceda M, Asenjo R, Zamorano J, Araya J, Castillo-Koch R, Espinoza J, Larraín E. Use of vitamins C and E as a prophylactic therapy to prevent postoperative atrial fibrillation. *Int J Cardiol* 2010; **138**: 221-228 [PMID: 19446899 DOI: 10.1016/j.ijcard.2009.04.043]
 - 28 **Ferdinandy P**, Danial H, Ambrus I, Rothery RA, Schulz R. Peroxynitrite is a major contributor to cytokine-induced myocardial contractile failure. *Circ Res* 2000; **87**: 241-247 [PMID: 10926876 DOI: 10.1161/01.RES.87.3.241]
 - 29 **Kanno S**, Lee PC, Zhang Y, Ho C, Griffith BP, Shears LL 2nd, Billiar TR. Attenuation of myocardial ischemia/reperfusion injury by superinduction of inducible nitric oxide synthase. *Circulation* 2000; **101**: 2742-2748 [PMID: 10851213 DOI: 10.1161/01.CIR.101.23.2742]
 - 30 **Suzuki H**, Colasanti M. Cross-talk between constitutive and inducible nitric oxide synthases. *Circulation* 2001; **103**: E81-E81 [PMID: 11294818 DOI: 10.1161/01.CIR.103.14.e81]
 - 31 **Kitamoto S**, Egashira K, Kataoka C, Koyanagi M, Katoh M, Shimokawa H, Morishita R, Kaneda Y, Sueishi K, Takeshita A. Increased activity of nuclear factor-kappaB participates in cardiovascular remodeling induced by chronic inhibition of nitric oxide synthesis in rats. *Circulation* 2000; **102**: 806-812 [PMID: 10942751 DOI: 10.1161/01.CIR.102.7.806]
 - 32 **Chevion M**, Jiang Y, Har-El R, Berenshtein E, Uretzky G, Kitrossky N. Copper and iron are mobilized following myocardial ischemia: possible predictive criteria for tissue injury. *Proc Natl Acad Sci USA* 1993; **90**: 1102-1106 [PMID: 8430081 DOI: 10.1073/pnas.90.3.1102]
 - 33 **Korkmaz S**, Barnucz E, Loganathan S, Li S, Radovits T, Hegedus P, Zubarevich A, Hirschberg K, Weymann A, Puskás LG, Özsvári B, Faragó N, Kanizsai I, Fábán G, Gyuris M, Merkely B, Karck M, Szabó C, Szabó G. Q50, an iron-chelating and zinc-complexing agent, improves cardiac function in rat models of ischemia/reperfusion-induced myocardial injury. *Circ J* 2013; **77**: 1817-1826 [PMID: 23575364 DOI: 10.1253/circj.CJ-12-1162]
 - 34 **Esposito BP**, Breuer W, Sirankapracha P, Pootrakul P, Hershko C, Cabantchik ZI. Labile plasma iron in iron overload: redox activity and susceptibility to chelation. *Blood* 2003; **102**: 2670-2677 [PMID: 12805056 DOI: 10.1182/blood-2003-03-0807]
 - 35 **Merkofer M**, Kissner R, Hider RC, Brunk UT, Koppenol WH. Fenton chemistry and iron chelation under physiologically relevant conditions: Electrochemistry and kinetics. *Chem Res Toxicol* 2006;

- 19: 1263-1269 [PMID: 17040095 DOI: 10.1021/tx060101w]
- 36 **Voogd A**, Sluiter W, Koster JF. The increased susceptibility to hydrogen peroxide of the (post-)ischemic rat heart is associated with the magnitude of the low molecular weight iron pool. *Free Radic Biol Med* 1994; **16**: 453-458 [PMID: 8005530 DOI: 10.1016/0891-5849(94)90122-8]
- 37 **Pucheu S**, Coudray C, Tresallet N, Favier A, de Leiris J. Effect of iron overload in the isolated ischemic and reperfused rat heart. *Cardiovasc Drugs Ther* 1993; **7**: 701-711 [PMID: 8241014 DOI: 10.1007/BF00877824]
- 38 **Tang WH**, Wu S, Wong TM, Chung SK, Chung SS. Polyol pathway mediates iron-induced oxidative injury in ischemic-reperfused rat heart. *Free Radic Biol Med* 2008; **45**: 602-610 [PMID: 18549825 DOI: 10.1016/j.freeradbiomed.2008.05.003]
- 39 **Zurlo MG**, De Stefano P, Borgna-Pignatti C, Di Palma A, Piga A, Melevendi C, Di Gregorio F, Burattini MG, Terzoli S. Survival and causes of death in thalassaemia major. *Lancet* 1989; **2**: 27-30 [PMID: 2567801 DOI: 10.1016/S0140-6736(89)90264-X]
- 40 **Xia S**, Zhang W, Huang L, Jiang H. Comparative efficacy and safety of deferoxamine, deferiprone and deferasirox on severe thalassemia: a meta-analysis of 16 randomized controlled trials. *PLoS One* 2013; **8**: e82662 [PMID: 24376563 DOI: 10.1371/journal.pone.0082662]
- 41 **Hoffbrand AV**, Taher A, Cappellini MD. How I treat transfusional iron overload. *Blood* 2012; **120**: 3657-3669 [PMID: 22919029 DOI: 10.1182/blood-2012-05-370098]
- 42 **Hool LC**. The L-type Ca(2+) channel as a potential mediator of pathology during alterations in cellular redox state. *Heart Lung Circ* 2009; **18**: 3-10 [PMID: 19119068 DOI: 10.1016/j.hlc.2008.11.004]
- 43 **Dixon IM**, Hata T, Dhalla NS. Sarcolemmal Na(+)-K(+)-ATPase activity in congestive heart failure due to myocardial infarction. *Am J Physiol* 1992; **262**: C664-C671 [PMID: 1312780 DOI: 10.1152/ajpcell.1992.262.3.C664]
- 44 **Sasaki M**, Joh T. Oxidative stress and ischemia-reperfusion injury in gastrointestinal tract and antioxidant, protective agents. *J Clin Biochem Nutr* 2007; **40**: 1-12 [PMID: 18437208 DOI: 10.3164/jcbn.40.1]
- 45 **Donoso P**, Sanchez G, Bull R, Hidalgo C. Modulation of cardiac ryanodine receptor activity by ROS and RNS. *Front Biosci* (Landmark Ed) 2011; **16**: 553-567 [PMID: 21196188 DOI: 10.2741/3705]
- 46 **Prosser BL**, Ward CW, Lederer WJ. X-ROS signaling: rapid mechano-chemo transduction in heart. *Science* 2011; **333**: 1440-1445 [PMID: 21903813 DOI: 10.1126/science]
- 47 **Kim YH**, Lim DS, Lee JH, Shim WJ, Ro YM, Park GH, Becker KG, Cho-Chung YS, Kim MK. Gene expression profiling of oxidative stress on atrial fibrillation in humans. *Exp Mol Med* 2003; **35**: 336-349 [PMID: 14646586 DOI: 10.1038/emmm.2003.45]
- 48 **Bowie A**, O'Neill LA. Oxidative stress and nuclear factor-kappaB activation: a reassessment of the evidence in the light of recent discoveries. *Biochem Pharmacol* 2000; **59**: 13-23 [PMID: 10605930 DOI: 10.1016/S0006-2952(99)00296-8]
- 49 **Chung MK**, Martin DO, Sprecher D, Wazni O, Kanderian A, Carnes CA, Bauer JA, Tchou PJ, Niebauer MJ, Natale A, Van Wagoner DR. C-reactive protein elevation in patients with atrial arrhythmias: inflammatory mechanisms and persistence of atrial fibrillation. *Circulation* 2001; **104**: 2886-2891 [PMID: 11739301 DOI: 10.1161/hc4901.101760]
- 50 **Lamm G**, Auer J, Weber T, Berent R, Ng C, Eber B. Postoperative white blood cell count predicts atrial fibrillation after cardiac surgery. *J Cardiothorac Vasc Anesth* 2006; **20**: 51-56 [PMID: 16458214 DOI: 10.1053/j.jvca.2005.03.026]
- 51 **Pavlović D**, Đorđević V, Kocić G. A "cross-talk" between oxidative stress and REDOX cell signaling. *Med Biol* 2002; **9**: 131-137
- 52 **Chandra J**, Samali A, Orrenius S. Triggering and modulation of apoptosis by oxidative stress. *Free Radic Biol Med* 2000; **29**: 323-333 [PMID: 11035261 DOI: 10.1016/S0891-5849(00)00302-6]
- 53 **Liakopoulos OJ**, Schmitto JD, Kazmaier S, Bräuer A, Quintel M, Schoendube FA, Dörge H. Cardiopulmonary and systemic effects of methylprednisolone in patients undergoing cardiac surgery. *Ann Thorac Surg* 2007; **84**: 110-8; discussion 118-9 [PMID: 17588396 DOI: 10.1016/j.athoracsur.2007.01.003]
- 54 **Opie LH**, Commerford PJ, Gersh BJ, Pfeffer MA. Controversies in ventricular remodelling. *Lancet* 2006; **367**: 356-367 [PMID: 16443044 DOI: 10.1016/S0140-6736(06)68074-4]
- 55 **Rodrigo R**, Libuy M, Feliú F, Hasson D. Oxidative stress-related biomarkers in essential hypertension and ischemia-reperfusion myocardial damage. *Dis Markers* 2013; **35**: 773-790 [PMID: 24347798 DOI: 10.1155/2013/974358]
- 56 **Kobayashi M**, Yamamoto M. Molecular mechanisms activating the Nrf2-Keap1 pathway of antioxidant gene regulation. *Antioxid Redox Signal* 2005; **7**: 385-394 [PMID: 15706085 DOI: 10.1089/ars.2005.7.385]
- 57 **Zhu H**, Jia Z, Misra BR, Zhang L, Cao Z, Yamamoto M, Trush MA, Misra HP, Li Y. Nuclear factor E2-related factor 2-dependent myocardial cytoprotection against oxidative and electrophilic stress. *Cardiovasc Toxicol* 2008; **8**: 71-85 [PMID: 18463988 DOI: 10.1007/s12012-008-9016-0]
- 58 **Piper HM**, García-Dorado D, Ovize M. A fresh look at reperfusion injury. *Cardiovasc Res* 1998; **38**: 291-300 [PMID: 9709390 DOI: 10.1016/S0008-6363(98)00033-9]
- 59 **Tatli E**, Alicik G, Buturak A, Yilmaztepe M, Aktoz M. Arrhythmias following revascularization procedures in the course of acute myocardial infarction: are they indicators of reperfusion or ongoing ischemia? *ScientificWorldJournal* 2013; **2013**: 160380 [PMID: 23431252 DOI: 10.1155/2013/160380]
- 60 **Rodrigo R**, Korantzopoulos P, Cereceda M, Asenjo R, Zamorano J, Villalabertia E, Baeza C, Aguayo R, Castillo R, Carrasco R, Gormaz JG. A randomized controlled trial to prevent post-operative atrial fibrillation by antioxidant reinforcement. *J Am Coll Cardiol* 2013; **62**: 1457-1465 [PMID: 23916928 DOI: 10.1016/j.jacc.2013.07.014]
- 61 **Tennant R**, Wiggers CJ. The effects of coronary occlusion on myocardial contraction. *Am J Physiol Content. American Physiological Society* 1935; **112**: 351-361 [DOI: 10.1152/ajplegacy.1935.112.2.351]
- 62 **Bolli R**. Mechanism of myocardial "stunning". *Circulation* 1990; **82**: 723-738 [PMID: 2203553 DOI: 10.1161/01.CIR.82.3.723]
- 63 **Laky D**, Parascan L, Căndeia V. Myocardial stunning. Morphological studies in acute experimental ischemia and intraoperative myocardial biopsies. *Rom J Morphol Embryol* 2008; **49**: 153-158 [PMID: 18516320]
- 64 **Kals J**, Starkopf J, Zilmer M, Pruler T, Pulges K, Hallaste M, Kals M, Pulges A, Soomets U. Antioxidant UPF1 attenuates myocardial stunning in isolated rat hearts. *Int J Cardiol* 2008; **125**: 133-135 [PMID: 17395289 DOI: 10.1016/j.ijcard.2007.01.032]
- 65 **Crystal GJ**, Malik G, Yoon SH, Kim SJ. Isoflurane late preconditioning against myocardial stunning is associated with enhanced antioxidant defenses. *Acta Anaesthesiol Scand* 2012; **56**: 39-47 [PMID: 22103751 DOI: 10.1111/j.1399-6576.2011.02583.x]
- 66 **Kloner RA**, Ganote CE, Jennings RB. The "no-reflow" phenomenon after temporary coronary occlusion in the dog. *J Clin Invest* 1974; **54**: 1496-1508 [PMID: 4140198 DOI: 10.1172/JCI107898]
- 67 **Movahed MR**, Butman SM. The pathogenesis and treatment of no-reflow occurring during percutaneous coronary intervention. *Cardiovasc Revasc Med* 2008; **9**: 56-61 [PMID: 18206640 DOI: 10.1016/j.carrev.2007.08.005]
- 68 **Bouletti C**, Mewton N, Germain S. The no-reflow phenomenon: State of the art. *Arch Cardiovasc Dis* 2015; **108**: 661-674 [PMID: 26616729 DOI: 10.1016/j.acvd.2015.09.006]
- 69 **Oyanagui Y**, Sato S. Superoxide dismutases and anti-oxidants protected mice from no-reflow and necrotic damage induced by ischemia. *Free Radic Res Commun* 1993; **18**: 147-157 [PMID: 8319925 DOI: 10.3109/10715769309147488]
- 70 **Shimizu M**, Sjöquist PO, Wang QD, Rydén L. Effects of the angiotensin AT1 receptor blocker candesartan on myocardial ischemic/reperfusion injury. *J Am Soc Nephrol* 1999; **10 Suppl 11**: S137-S142 [PMID: 9892154]
- 71 **Molyneux CA**, Glyn MC, Ward BJ. Oxidative stress and cardiac microvascular structure in ischemia and reperfusion: the protective effect of antioxidant vitamins. *Microvasc Res* 2002; **64**: 265-277 [PMID: 12204651 DOI: 10.1006/mvre.2002.2419]
- 72 **Matsumoto H**, Inoue N, Takaoka H, Hata K, Shinke T, Yoshikawa R, Masai H, Watanabe S, Ozawa T, Yokoyama M. Depletion of

- antioxidants is associated with no-reflow phenomenon in acute myocardial infarction. *Clin Cardiol* 2004; **27**: 466-470 [PMID: 15346844 DOI: 10.1002/clc.4960270809]
- 73 **Zeng M**, Yan H, Chen Y, Zhao HJ, Lv Y, Liu C, Zhou P, Zhao B. Suppression of NF- κ B reduces myocardial no-reflow. *PLoS One* 2012; **7**: e47306 [PMID: 23056624 DOI: 10.1371/journal.pone.0047306]
 - 74 **Basili S**, Tanzilli G, Mangieri E, Raparelli V, Di Santo S, Pignatelli P, Violi F. Intravenous ascorbic acid infusion improves myocardial perfusion grade during elective percutaneous coronary intervention: relationship with oxidative stress markers. *JACC Cardiovasc Interv* 2010; **3**: 221-229 [PMID: 20170881 DOI: 10.1016/j.jcin.2009.10.025]
 - 75 **Ramos C**, Brito R, González-Montero J, Valls N, Gormaz JG, Prieto JC, Aguayo R, Puentes Á, Noriega V, Pereira N, Palavecino T, Rodrigo R. Effects of a novel ascorbate-based protocol on infarct size and ventricle function in acute myocardial infarction patients undergoing percutaneous coronary angioplasty. *Arch Med Sci* 2017; **13**: 558-567 [PMID: 28507569 DOI: 10.5114/aoms.2016.59713]
 - 76 **Valls N**, Gormaz JG, Aguayo R, González J, Brito R, Hasson D, Libuy M, Ramos C, Carrasco R, Prieto JC, Dussaillant G, Puentes Á, Noriega V, Rodrigo R. Amelioration of persistent left ventricular function impairment through increased plasma ascorbate levels following myocardial infarction. *Redox Rep* 2016; **21**: 75-83 [PMID: 26066587 DOI: 10.1179/1351000215Y.0000000018]
 - 77 **Tsujita K**, Shimomura H, Kaikita K, Kawano H, Hokamaki J, Nagayoshi Y, Yamashita T, Fukuda M, Nakamura Y, Sakamoto T, Yoshimura M, Ogawa H. Long-term efficacy of edaravone in patients with acute myocardial infarction. *Circ J* 2006; **70**: 832-837 [PMID: 16799234 DOI: 10.1253/circj.70.832]
 - 78 **Defraigne JO**, Pincemail J, Detry O, Franssen C, Meurisse M, Limet R. Preservation of cortical microcirculation after kidney ischemia-reperfusion: value of an iron chelator. *Ann Vasc Surg* 1994; **8**: 457-467 [PMID: 7811583 DOI: 10.1007/BF02133066]
 - 79 **Brunet J**, Boily MJ, Cordeau S, Des Rosiers C. Effects of N-acetylcysteine in the rat heart reperfused after low-flow ischemia: evidence for a direct scavenging of hydroxyl radicals and a nitric oxide-dependent increase in coronary flow. *Free Radic Biol Med* 1995; **19**: 627-638 [PMID: 8529922 DOI: 10.1016/0891-5849(95)00077-B]
 - 80 **Ciccone MM**, Cortese F, Gesualdo M, Carbonara S, Zito A, Ricci G, De Pascalis F, Scicchitano P, Riccioni G. Dietary intake of carotenoids and their antioxidant and anti-inflammatory effects in cardiovascular care. *Mediators Inflamm* 2013; **2013**: 782137 [PMID: 24489447 DOI: 10.1155/2013/782137]
 - 81 **Tong C**, Peng C, Wang L, Zhang L, Yang X, Xu P, Li J, Delplancke T, Zhang H, Qi H. Intravenous Administration of Lycopene, a Tomato Extract, Protects against Myocardial Ischemia-Reperfusion Injury. *Nutrients* 2016; **8**: 138 [PMID: 26950150 DOI: 10.3390/nu8030138]
 - 82 **Wang Y**, Chung SJ, McCullough ML, Song WO, Fernandez ML, Koo SI, Chun OK. Dietary carotenoids are associated with cardiovascular disease risk biomarkers mediated by serum carotenoid concentrations. *J Nutr* 2014; **144**: 1067-1074 [PMID: 24744306 DOI: 10.3945/jn.113.184317]
 - 83 **Wang Y**, Sun J, Liu C, Fang C. Protective effects of crocetin pretreatment on myocardial injury in an ischemia/reperfusion rat model. *Eur J Pharmacol* 2014; **741**: 290-296 [PMID: 25176181 DOI: 10.1016/j.ejphar.2014.07.052]
 - 84 **Zhu Z**, Zhu J, Zhao X, Yang K, Lu L, Zhang F, Shen W, Zhang R. All-Trans Retinoic Acid Ameliorates Myocardial Ischemia/Reperfusion Injury by Reducing Cardiomyocyte Apoptosis. *PLoS One* 2015; **10**: e0133414 [PMID: 26186635 DOI: 10.1371/journal.pone.0133414]
 - 85 **Levine M**, Rumsey SC, Daruwala R, Park JB, Wang Y. Criteria and recommendations for vitamin C intake. *JAMA* 1999; **281**: 1415-1423 [PMID: 10217058 DOI: 10.1001/jama.281.15.1415]
 - 86 **Wang X**, Quinn PJ. The location and function of vitamin E in membranes (review). *Mol Membr Biol* 2000; **17**: 143-156 [PMID: 11128973 DOI: 10.1080/09687680010000311]
 - 87 **Heller R**, Werner-Felmayer G, Werner ER. Alpha-Tocopherol and endothelial nitric oxide synthesis. *Ann N Y Acad Sci* 2004; **1031**: 74-85 [PMID: 15753135 DOI: 10.1196/annals.1331.007]
 - 88 **Heller R**, Werner-Felmayer G, Werner ER. Antioxidants and endothelial nitric oxide synthesis. *Eur J Clin Pharmacol. Springer-Verlag* 2006; **62** (S1): 21-28 [DOI: 10.1007/s00228-005-0009-7]
 - 89 **Gille L**, Staniek K, Nohl H. Effects of tocopheryl quinone on the heart: model experiments with xanthine oxidase, heart mitochondria, and isolated perfused rat hearts. *Free Radic Biol Med* 2001; **30**: 865-876 [PMID: 11295529 DOI: 10.1016/S0891-5849(01)00475-0]
 - 90 **Ramlawi B**, Otu H, Mieno S, Boodhwani M, Sodha NR, Clements RT, Bianchi C, Sellke FW. Oxidative stress and atrial fibrillation after cardiac surgery: a case-control study. *Ann Thorac Surg* 2007; **84**: 1166-1172; discussion 1172-1173 [PMID: 17888965 DOI: 10.1016/j.athoracsur.2007.04.12]
 - 91 **Newaz MA**, Yousefipour Z, Nawal NN. Modulation of nitric oxide synthase activity in brain, liver, and blood vessels of spontaneously hypertensive rats by ascorbic acid: protection from free radical injury. *Clin Exp Hypertens* 2005; **27**: 497-508 [PMID: 16081342 DOI: 10.1081/CEH-200067681]
 - 92 **Guney M**, Oral B, Demirin H, Karahan N, Mungan T, Delibas N. Protective effects of vitamins C and E against endometrial damage and oxidative stress in fluoride intoxication. *Clin Exp Pharmacol Physiol* 2007; **34**: 467-474 [PMID: 17439417 DOI: 10.1111/j.1440-1681.2007.04596.x]
 - 93 **Gille L**, Gregor W, Staniek K, Nohl H. Redox-interaction of alpha-tocopheryl quinone with isolated mitochondrial cytochrome bc1 complex. *Biochem Pharmacol* 2004; **68**: 373-381 [PMID: 15194009 DOI: 10.1016/j.bcp.2004.03.031]
 - 94 **Ulker S**, McKeown PP, Bayraktutan U. Vitamins reverse endothelial dysfunction through regulation of eNOS and NAD(P)H oxidase activities. *Hypertension* 2003; **41**: 534-539 [PMID: 12623955 DOI: 10.1161/01.HYP.0000057421.28533.37]
 - 95 **May JM**, Qu ZC, Mendiratta S. Protection and recycling of alpha-tocopherol in human erythrocytes by intracellular ascorbic acid. *Arch Biochem Biophys* 1998; **349**: 281-289 [PMID: 9448716 DOI: 10.1006/abbi.1997.0473]
 - 96 **Taddei S**, Virdis A, Ghiadoni L, Salvetti A. Endothelial dysfunction in hypertension: fact or fancy? *J Cardiovasc Pharmacol* 1998; **32** Suppl 3: S41-S47 [PMID: 9883747]
 - 97 **Newaz MA**, Nawal NN, Rohaizan CH, Muslim N, Gapor A. alpha-Tocopherol increased nitric oxide synthase activity in blood vessels of spontaneously hypertensive rats. *Am J Hypertens* 1999; **12**: 839-844 [PMID: 10480480 DOI: 10.1016/S0895-7061(99)00022-9]
 - 98 **Wu F**, Schuster DP, Tymi K, Wilson JX. Ascorbate inhibits NADPH oxidase subunit p47phox expression in microvascular endothelial cells. *Free Radic Biol Med* 2007; **42**: 124-131 [PMID: 17157199 DOI: 10.1016/j.freeradbiomed.2006.10.033]
 - 99 **Gao F**, Yao CL, Gao E, Mo QZ, Yan WL, McLaughlin R, Lopez BL, Christopher TA, Ma XL. Enhancement of glutathione cardioprotection by ascorbic acid in myocardial reperfusion injury. *J Pharmacol Exp Ther* 2002; **301**: 543-550 [PMID: 11961055 DOI: 10.1124/jpet.301.2.543]
 - 100 **Packer JE**, Slater TF, Willson RL. Direct observation of a free radical interaction between vitamin E and vitamin C. *Nature* 1979; **278**: 737-738 [PMID: 431730 DOI: 10.1038/278737a0]
 - 101 **Niki E**, Noguchi N, Tsuchihashi H, Gotoh N. Interaction among vitamin C, vitamin E, and beta-carotene. *Am J Clin Nutr* 1995; **62**: 1322S-1326S [PMID: 7495227 DOI: 10.1093/ajcn/62.6.1322S]
 - 102 **Levine M**, Padayatty SJ, Espey MG. Vitamin C: a concentration-function approach yields pharmacology and therapeutic discoveries. *Adv Nutr* 2011; **2**: 78-88 [PMID: 22332036 DOI: 10.3945/an.110.000109]
 - 103 **Schneider MP**, Delles C, Schmidt BM, Oehmer S, Schwarz TK, Schmieder RE, John S. Superoxide scavenging effects of N-acetylcysteine and vitamin C in subjects with essential hypertension. *Am J Hypertens* 2005; **18**: 1111-1117 [PMID: 16109326 DOI: 10.1016/j.amjhyper.2005.02.006]
 - 104 **Rushworth GF**, Megson IL. Existing and potential therapeutic uses for N-acetylcysteine: the need for conversion to intracellular glutathione for antioxidant benefits. *Pharmacol Ther* 2014; **141**: 150-159 [PMID: 24080471 DOI: 10.1016/j.pharmthera.2013.09.006]
 - 105 **Winterbourn CC**, Metodiewa D. Reactivity of biologically

- important thiol compounds with superoxide and hydrogen peroxide. *Free Radic Biol Med* 1999; **27**: 322-328 [PMID: 10468205 DOI: 10.1016/S0891-5849(99)00051-9]
- 106 **Lodge JK**, Traber MG, Packer L. Thiol chelation of Cu²⁺ by dihydrolipoic acid prevents human low density lipoprotein peroxidation. *Free Radic Biol Med* 1998; **25**: 287-297 [PMID: 9680174 DOI: 10.1016/S0891-5849(98)00048-3]
 - 107 **Joshi D**, Mittal DK, Shrivastava S, Shukla S. Protective role of thiol chelators against dimethylmercury induced toxicity in male rats. *Bull Environ Contam Toxicol* 2010; **84**: 613-617 [PMID: 20401649 DOI: 10.1007/s00128-010-9982-3]
 - 108 **Lu Y**, Qin W, Shen T, Dou L, Man Y, Wang S, Xiao C, Li J. The antioxidant N-acetylcysteine promotes atherosclerotic plaque stabilization through suppression of RAGE, MMPs and NF- κ B in ApoE-deficient mice. *J Atheroscler Thromb* 2011; **18**: 998-1008 [PMID: 21873804 DOI: 10.5551/jat.8870]
 - 109 **Arstall MA**, Yang J, Stafford I, Betts WH, Horowitz JD. N-acetylcysteine in combination with nitroglycerin and streptokinase for the treatment of evolving acute myocardial infarction. Safety and biochemical effects. *Circulation* 1995; **92**: 2855-2862 [PMID: 7586252 DOI: 10.1161/01.CIR.92.10.2855]
 - 110 **Thiele H**, Hildebrand L, Schirdewahn C, Eitel I, Adams V, Fuernau G, Erbs S, Linke A, Diederich KW, Nowak M, Desch S, Gutberlet M, Schuler G. Impact of high-dose N-acetylcysteine versus placebo on contrast-induced nephropathy and myocardial reperfusion injury in unselected patients with ST-segment elevation myocardial infarction undergoing primary percutaneous coronary intervention. The LIPSIA-N-ACC (Prospective, Single-Blind, Placebo-Controlled, Randomized Leipzig Immediate Percutaneous Coronary Intervention Acute Myocardial Infarction N-ACC) Trial. *J Am Coll Cardiol* 2010; **55**: 2201-2209 [PMID: 20466200 DOI: 10.1016/j.jacc.2009.08.091]
 - 111 **Yesilbursa D**, Serdar A, Senturk T, Serdar Z, Sağ S, Cordan J. Effect of N-acetylcysteine on oxidative stress and ventricular function in patients with myocardial infarction. *Heart Vessels* 2006; **21**: 33-37 [PMID: 16440146 DOI: 10.1007/s00380-005-0854-4]
 - 112 **Abe M**, Takiguchi Y, Ichimaru S, Tsuchiya K, Wada K. Comparison of the protective effect of N-acetylcysteine by different treatments on rat myocardial ischemia-reperfusion injury. *J Pharmacol Sci* 2008; **106**: 571-577 [PMID: 18385540 DOI: 10.1254/jphs.FP0071664]
 - 113 **Pasupathy S**, Tavella R, Grover S, Raman B, Procter NEK, Du YT, Mahadavan G, Stafford I, Heresztyn T, Holmes A, Zeitz C, Arstall M, Selvanayagam J, Horowitz JD, Beltrame JF. Early Use of N-acetylcysteine With Nitrate Therapy in Patients Undergoing Primary Percutaneous Coronary Intervention for ST-Segment-Elevation Myocardial Infarction Reduces Myocardial Infarct Size (the NACIAM Trial [N-acetylcysteine in Acute Myocardial Infarction]). *Circulation* 2017; **136**: 894-903 [PMID: 28634219 DOI: 10.1161/CIRCULATIONAHA.117.027575]
 - 114 **Kolnagou A**, Kleanthous M, Kontoghiorghes GJ. Reduction of body iron stores to normal range levels in thalassaemia by using a deferiprone/deferioxamine combination and their maintenance thereafter by deferiprone monotherapy. *Eur J Haematol* 2010; **85**: 430-438 [PMID: 20662901 DOI: 10.1111/j.1600-0609.2010.01499.x]
 - 115 **Freeman AP**, Giles RW, Berdoukas VA, Walsh WF, Choy D, Murray PC. Early left ventricular dysfunction and chelation therapy in thalassemia major. *Ann Intern Med* 1983; **99**: 450-454 [PMID: 6625375 DOI: 10.7326/0003-4819-99-4-450]
 - 116 **Reddy BR**, Kloner RA, Przyklenk K. Early treatment with deferoxamine limits myocardial ischemic/reperfusion injury. *Free Radic Biol Med* 1989; **7**: 45-52 [PMID: 2753395 DOI: 10.1016/0891-5849(89)90099-3]
 - 117 **Lesnfsky EJ**, Repine JE, Horwitz LD. Deferoxamine pretreatment reduces canine infarct size and oxidative injury. *J Pharmacol Exp Ther* 1990; **253**: 1103-1109 [PMID: 2359019]
 - 118 **Williams RE**, Zweier JL, Flaherty JT. Treatment with deferoxamine during ischemia improves functional and metabolic recovery and reduces reperfusion-induced oxygen radical generation in rabbit hearts. *Circulation* 1991; **83**: 1006-1014 [PMID: 1847847 DOI: 10.1161/01.CIR.83.3.1006]
 - 119 **Paraskevaidis IA**, Iliodromitis EK, Vlahakos D, Tsiapras DP, Nikolaidis A, Marathias A, Michalis A, Kremastinos DT. Deferoxamine infusion during coronary artery bypass grafting ameliorates lipid peroxidation and protects the myocardium against reperfusion injury: immediate and long-term significance. *Eur Heart J* 2005; **26**: 263-270 [PMID: 15618054 DOI: 10.1093/eurheartj/ehi028]
 - 120 **Chan W**, Taylor AJ, Ellims AH, Lefkovits L, Wong C, Kingwell BA, Natoli A, Croft KD, Mori T, Kaye DM, Dart AM, Duffy SJ. Effect of iron chelation on myocardial infarct size and oxidative stress in ST-elevation-myocardial infarction. *Circ Cardiovasc Interv* 2012; **5**: 270-278 [PMID: 22496085 DOI: 10.1161/CIRCINTERVENTIONS.111.966226]
 - 121 **Karahaliou A**, Katsouras C, Koulouras V, Nikas D, Niokou D, Papadopoulos G, Nakos G, Sideris D, Michalis L. Ventricular arrhythmias and antioxidative medication: experimental study. *Hellenic J Cardiol* 2008; **49**: 320-328 [PMID: 18846922]

P- Reviewer: Ciccon MM, Sun CK, Schoenhagen P **S- Editor:** Ji FF
L- Editor: A **E- Editor:** Wu YXJ





Published by **Baishideng Publishing Group Inc**
7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
Telephone: +1-925-223-8242
Fax: +1-925-223-8243
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>

