

World Journal of *Gastroenterology*

World J Gastroenterol 2018 May 14; 24(18): 1925-2046



**REVIEW**

- 1925** Gastrointestinal stromal tumors: A multidisciplinary challenge
Sanchez-Hidalgo JM, Duran-Martinez M, Molero-Payan R, Rufian-Peña S, Arjona-Sanchez A, Casado-Adam A, Cosano-Alvarez A, Briceño-Delgado J
- 1942** New therapeutic options opened by the molecular classification of gastric cancer
Chivu-Economescu M, Matei L, Necula LG, Dragu DL, Bleotu C, Diaconu CC
- 1962** Ambiguous roles of innate lymphoid cells in chronic development of liver diseases
Shen Y, Li J, Wang SQ, Jiang W

MINIREVIEWS

- 1978** Laparoscopic gastrojejunostomy for gastric outlet obstruction in patients with unresectable hepatopancreatobiliary cancers: A personal series and systematic review of the literature
Manuel-Vázquez A, Latorre-Fragua R, Ramiro-Pérez C, López-Marciano A, De la Plaza-Llamas R, Ramia JM
- 1989** Mouse models for investigating the underlying mechanisms of nonalcoholic steatohepatitis-derived hepatocellular carcinoma
Takakura K, Oikawa T, Tomita Y, Mizuno Y, Nakano M, Saeki C, Torisu Y, Saruta M

ORIGINAL ARTICLE**Basic Study**

- 1995** Microbiota modification by probiotic supplementation reduces colitis associated colon cancer in mice
Mendes MC, Paulino DS, Brambilla SR, Camargo JA, Persinoti GF, Carnevalheira JB
- 2009** Ischemia/reperfusion injury in porcine intestine - Viability assessment
Strand-Amundsen RJ, Reims HM, Reinholt FP, Ruud TE, Yang R, Høgetveit JO, Tønnessen TI

Clinical Trials Study

- 2024** Quantitative assessment of hepatic fibrosis in chronic hepatitis B and C: T1 mapping on Gd-EOB-DTPA-enhanced liver magnetic resonance imaging
Pan S, Wang XQ, Guo QY

Observational Study

- 2036** Thiopurines are negatively associated with anthropometric parameters in pediatric Crohn's disease
Gupta N, Lustig RH, Chao C, Vittinghoff E, Andrews H, Leu CS

ABOUT COVER

Editorial board member of *World Journal of Gastroenterology*, Andrew Stewart Day, MD, Professor, Paediatrics Department, University of Otago, Christchurch 8041, New Zealand

AIMS AND SCOPE

World Journal of Gastroenterology (*World J Gastroenterol*, *WJG*, print ISSN 1007-9327, online ISSN 2219-2840, DOI: 10.3748) is a peer-reviewed open access journal. *WJG* was established on October 1, 1995. It is published weekly on the 7th, 14th, 21st, and 28th each month. The *WJG* Editorial Board consists of 642 experts in gastroenterology and hepatology from 59 countries.

The primary task of *WJG* is to rapidly publish high-quality original articles, reviews, and commentaries in the fields of gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, hepatobiliary surgery, gastrointestinal oncology, gastrointestinal radiation oncology, gastrointestinal imaging, gastrointestinal interventional therapy, gastrointestinal infectious diseases, gastrointestinal pharmacology, gastrointestinal pathophysiology, gastrointestinal pathology, evidence-based medicine in gastroenterology, pancreatology, gastrointestinal laboratory medicine, gastrointestinal molecular biology, gastrointestinal immunology, gastrointestinal microbiology, gastrointestinal genetics, gastrointestinal translational medicine, gastrointestinal diagnostics, and gastrointestinal therapeutics. *WJG* is dedicated to become an influential and prestigious journal in gastroenterology and hepatology, to promote the development of above disciplines, and to improve the diagnostic and therapeutic skill and expertise of clinicians.

INDEXING/ABSTRACTING

World Journal of Gastroenterology (*WJG*) is now indexed in Current Contents[®]/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch[®]), Journal Citation Reports[®], Index Medicus, MEDLINE, PubMed, PubMed Central and Directory of Open Access Journals. The 2017 edition of Journal Citation Reports[®] cites the 2016 impact factor for *WJG* as 3.365 (5-year impact factor: 3.176), ranking *WJG* as 29th among 79 journals in gastroenterology and hepatology (quartile in category Q2).

EDITORS FOR THIS ISSUE

Responsible Assistant Editor: *Xiang Li*
Responsible Electronic Editor: *Yan Huang*
Proofing Editor-in-Chief: *Lian-Sheng Ma*

Responsible Science Editor: *Xue-Jiao Wang*
Proofing Editorial Office Director: *Ze-Mao Gong*

NAME OF JOURNAL
World Journal of Gastroenterology

ISSN
ISSN 1007-9327 (print)
ISSN 2219-2840 (online)

LAUNCH DATE
October 1, 1995

FREQUENCY
Weekly

EDITORS-IN-CHIEF
Damian Garcia-Olmo, MD, PhD, Doctor, Professor, Surgeon, Department of Surgery, Universidad Autonoma de Madrid; Department of General Surgery, Fundacion Jimenez Diaz University Hospital, Madrid 28040, Spain

Stephen C Strom, PhD, Professor, Department of Laboratory Medicine, Division of Pathology, Karolinska Institutet, Stockholm 141-86, Sweden

Andrzej S Tarnawski, MD, PhD, DSc (Med), Professor of Medicine, Chief Gastroenterology, VA Long Beach Health Care System, University of California, Irvine, CA, 5901 E. Seventh Str., Long Beach,

CA 90822, United States

EDITORIAL BOARD MEMBERS
All editorial board members resources online at <http://www.wjgnet.com/1007-9327/editorialboard.htm>

EDITORIAL OFFICE
Ze-Mao Gong, Director
World Journal of Gastroenterology
Baishideng Publishing Group Inc
7901 Stoneridge Drive, Suite 501,
Pleasanton, CA 94588, USA
Telephone: +1-925-2238242
Fax: +1-925-2238243
E-mail: editorialoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>

PUBLISHER
Baishideng Publishing Group Inc
7901 Stoneridge Drive, Suite 501,
Pleasanton, CA 94588, USA
Telephone: +1-925-2238242
Fax: +1-925-2238243
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>

PUBLICATION DATE
May 14, 2018

COPYRIGHT
© 2018 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT
All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORS
Full instructions are available online at <http://www.wjgnet.com/bpg/gerinfo/204>

ONLINE SUBMISSION
<http://www.f6publishing.com>

Mouse models for investigating the underlying mechanisms of nonalcoholic steatohepatitis-derived hepatocellular carcinoma

Kazuki Takakura, Tsunekazu Oikawa, Yoichi Tomita, Yusuke Mizuno, Masanori Nakano, Chisato Saeki, Yuichi Torisu, Masayuki Saruta

Kazuki Takakura, Tsunekazu Oikawa, Yoichi Tomita, Yusuke Mizuno, Masanori Nakano, Chisato Saeki, Yuichi Torisu, Masayuki Saruta, Division of Gastroenterology and Hepatology, Department of Internal Medicine, The Jikei University School of Medicine, Tokyo 105-8461, Japan

ORCID number: Kazuki Takakura (0000-0003-1444-3761); Tsunekazu Oikawa (0000-0003-1365-5391); Yoichi Tomita (0000-0001-8674-9837); Yusuke Mizuno (0000-0002-5742-8812); Masanori Nakano (0000-0001-7222-6437); Chisato Saeki (0000-0002-7407-6142); Yuichi Torisu (0000-0002-2349-8855); Masayuki Saruta (0000-0001-8172-3240).

Author contributions: Takakura K and Oikawa T wrote the manuscript; Tomita Y, Mizuno Y, Nakano M and Saeki C critically appraised the manuscript; Torisu Y and Saruta M formatted and edited the final manuscript.

Conflict-of-interest statement: All of the authors declare no potential conflicts of interest relevant to this article.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: Kazuki Takakura, MD, PhD, Division of Gastroenterology and Hepatology, Department of Internal Medicine, The Jikei University School of Medicine, 3-25-8, Nishi-shimbashi, Minato-ku, Tokyo 105-8461, Japan. ktakakura@jikei.ac.jp
Telephone: +81-3-34331111
Fax: +81-3-34350569

Received: March 29, 2018

Peer-review started: April 4, 2018

First decision: April 27, 2018

Revised: May 1, 2018

Accepted: May 6, 2018

Article in press: May 6, 2018

Published online: May 14, 2018

Abstract

As the incidence of hepatocellular carcinoma (HCC) caused by infection with the hepatotropic viruses hepatitis B and hepatitis C decreases, greater attention has become focused on HCC caused by nonalcoholic steatohepatitis (NASH), an advanced form of nonalcoholic fatty liver disease which has shown increasing prevalence in correspondence with the overall increase in metabolic syndrome over the recent decades. Several clinical population studies have shown a positive relationship between NASH and HCC, while also providing initial insights into the underlying mechanisms of HCC development from NASH. Research into the pathological progression of NASH to HCC has advanced by use of several beneficial rodent models. In this review, we summarize the established mouse models for preclinical research of NASH-associated HCC and discuss the underlying hepatic mechanisms of NASH-related tumorigenesis identified to date that could lead to new targets for treatment and prevention.

Key words: Hepatocellular carcinoma; Nonalcoholic steatohepatitis; Nonalcoholic fatty liver disease

© The Author(s) 2018. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: This review provides a brief overview of the molecular mechanisms underlying progression

to hepatocellular carcinoma from nonalcoholic steatohepatitis that have been identified to date using the array of mouse models currently available and popular in the experimental field.

Takakura K, Oikawa T, Tomita Y, Mizuno Y, Nakano M, Saeki C, Torisu Y, Saruta M. Mouse models for investigating the underlying mechanisms of nonalcoholic steatohepatitis-derived hepatocellular carcinoma. *World J Gastroenterol* 2018; 24(18): 1989-1994 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v24/i18/1989.htm> DOI: <http://dx.doi.org/10.3748/wjg.v24.i18.1989>

INTRODUCTION

As Western diet and problems with food satiation have spread across the globe in recent years, there has been a concomitant increase in patients with nonalcoholic fatty liver disease (NAFLD) and its progressive form of nonalcoholic steatohepatitis (NASH). This increase is the result of prevailing metabolic syndrome, including obesity, diabetes and hyperlipidemia^[1-4]. The distinctive characteristic of NAFLD is its diversity of conditions, from simple fatty accumulation in the liver to hepatic injury and inflammation with or without fibrosis^[2,5-7]. The sequential progression to NASH puts the sufferer at risk for irreversible liver cirrhosis and hepatocellular carcinoma (HCC)^[4,7], causing the patient to require more medical attention due to the increased morbidity and mortality^[8]. Indeed, HCC is a leading indication for liver transplantation, especially in developed countries^[9,10].

Compared with the long history of both clinical and laboratory investigations to elucidate the molecular pathogenesis of HCC derived from chronic hepatotropic virus infections, particularly with hepatitis B virus and hepatitis C virus, and from alcoholic liver disease, the pathologic mechanisms of NASH-associated HCC (NASH-HCC) remain largely uninvestigated and unknown. The public health threat associated with the increasing incidence of NASH-HCC^[11], however, highlights the urgent need to gain a more comprehensive and detailed understanding of the mechanisms which mediate NASH-HCC progression. Several experimental mouse models exist for such studies^[12-15] and should be continuously applied to preclinical investigations into the pathogenic pathways of NASH-HCC to advance the subsequent development of methods to manage the modern increasing clinical trend.

Here, we summarize the established mouse models for preclinical research of NASH-HCC progression (Table 1) and discuss the revealed mechanisms and the future prospective of NASH-related tumorigenesis in liver which could lead to new targets for treatment or prevention (Figure 1). Of note, we recognize the existence of other available rodent models which can also be used for assessing the mechanisms of NASH-

HCC; however, we focused this review on the ones which are most representative of metabolic syndrome-associated steatohepatitis and which generate HCC unfaithfully from NASH status within a certain period of time.

CONFIRMED TUMORIGENIC MECHANISMS OF CURRENT NASH-HCC MOUSE MODELS

The established mouse models for preclinical research of NASH-HCC progression are listed below (Table 1).

PTEN null mice

PTEN, a tumor suppressor gene which antagonizes the PI3K/Akt pathway, is mutated in many human cancers, including HCC, and is essential for maintaining homeostasis and preventing oncogenesis in the liver. Decreased *Pten* expression leads to increased tumor grade, advanced stage and poor prognosis. Hepatocyte-specific *Pten* null mice were generated by Horie *et al.*^[12], wherein steatohepatitis emerges at 10 wk old and hepatic tumors at 40-44 wk old. The liver tumors become adenomas in 100% of these mice or HCC in 66% at 74-78 wk old, due to the *Pten* deficiency (*Pten* knock-out, KO) causing lipid accumulation in hepatocytes. In general, these mice have revealed that *Pten* function is crucial for preventing tumorigenesis in liver.

Several other research groups have uncovered different mechanisms of NASH-HCC by using the *Pten* null mouse model. For example, a study of eicosapentaenoic acid (EPA; a typical dietary n-3 polyunsaturated fatty acid contained in fish oil and a reagent for upgrading lipid metabolism^[16]) performed by Ishii *et al.*^[17] showed the effect of EPA on steatohepatitis and tumor formation in *Pten* null mice. The data confirmed that the steatotic change, accumulation of inflammatory cells and presence of ballooning hepatocytes were significantly decreased in the EPA group compared with the control group. In addition, liver adenomas developed in 63% of the control group mice, as compared with 0% of the EPA group mice, by 40 wk of age. HCC developed in 75% of the control group and 13% of the EPA group of the *Pten* KO mice at 76 wk old. In addition, MAPK and Akt, which are both downstream signaling molecules of Ras, were found to be activated in hepatocytes of the *Pten* KO mice, thereby promoting tumorigenesis^[18]. Collectively, these data suggested that EPA alters fatty acid composition in liver and suppresses the development of HCC by inactivating these signaling pathways in *Pten* null mice.

In another study of the *Pten* null mice, reduction of glucose-regulated protein 78 (GRP78; a molecular chaperone elevated in several human cancers, including HCC^[19,20], and which is critical for endoplasmic reticulum folding, stress signaling and PI3K/Akt activation) promoted liver steatosis and liver injury at

Table 1 Mouse models of nonalcoholic steatohepatitis-associated hepatocellular carcinoma

List	Backgrounds	Inducer of NASH/HCC	Carcinogenic duration	HCC occurrence (%)	Ref.
PTEN null mice	Genetic	Spontaneous	40 wk	66 (74-78 wk)	[12,17,18,21,22]
MC4R KO mice	Genetic	HFC diet	1 yr	100	[13,29,31]
STAM mice	DM/HL	Streptozotocin, HFC diet	20 wk	100	[14,32-36]
ALR KO mice	Genetic	Spontaneous	1 yr	60	[15]

HFC: High fat/calorie; DM: Diabetes; HL: Hyperlipidemia; HCC: Hepatocellular carcinoma; NASH: Nonalcoholic steatohepatitis.

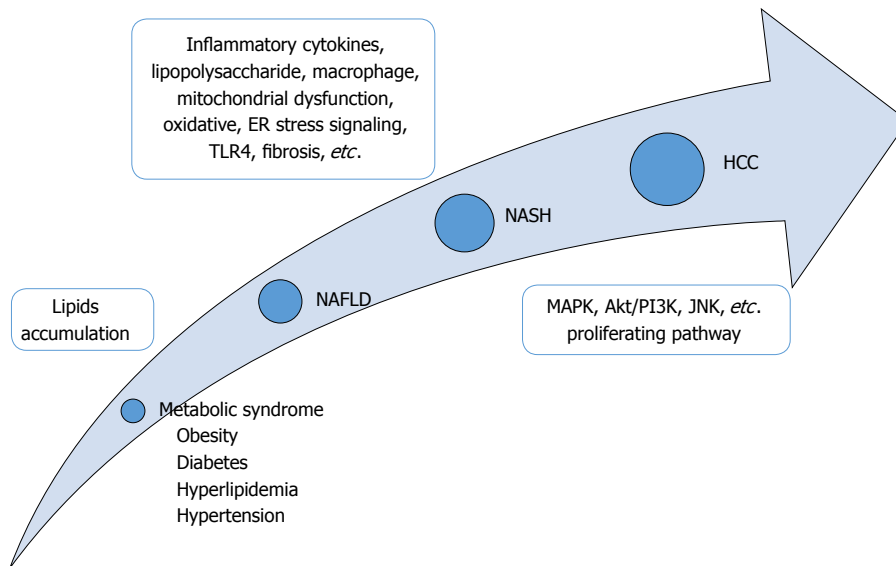


Figure 1 Developmental process of hepatocellular carcinoma via nonalcoholic steatohepatitis. Based on excessive lipids accumulation, several factors such as inflammatory cytokines, oxidative stress or proliferating pathways are involved in the whole process of hepatocellular carcinoma development from nonalcoholic steatohepatitis status via nonalcoholic fatty liver disease. NAFLD: Nonalcoholic fatty liver disease; HCC: Hepatocellular carcinoma; NASH: Nonalcoholic steatohepatitis.

3 mo of age and liver tumors at 6 mo of age^[21]. These effects proceeded HCC or cholangiocarcinoma, which developed at 8-9 mo of age and was accompanied by elevation of p-JNK; in contrast, the GRP78 normal *Pten* null mice never generated tumor lesions in liver, as assessed out to 14 mo of age^[21]. Collectively, these data suggested that JNK might contribute to acceleration of tumorigenesis in liver. Accordingly, these data demonstrated GPR78 as a regulator for *Pten* loss-mediated liver steatosis and tumor progression on the basis of p-JNK elevation.

In a third study of the *Pten* null mice, Miura *et al.*^[22] showed that liver tumors emerged after 36 wk of age, although no liver tumors were found in *Pten* normal mice until 72 wk of age. Toll-like receptor (TLR) 4 expressed on macrophages was found to contribute to the development of steatohepatitis and HCC in *Pten* KO mice. In general, gut-derived materials stimulate the immune system, including the TLRs which recognize bacterial components. TLR4, in particular, senses components of Gram-negative bacteria, including the lipopolysaccharide (LPS)^[23]. In this way, TLRs affect the development of liver diseases. Moreover, macrophages are known to be a major source of proinflammatory cytokines which

facilitate the progression of steatohepatitis^[24,25] and Ly6C is a marker for inflammatory macrophages^[26]. Hepatic macrophages isolated from the *Pten* null mice showed an increased expression of Ly6C. In addition, TLR4 signaling was shown to promote hepatic inflammation as well as subsequent liver tumor growth in the *Pten* null mice. Antibiotic treatment suppressed the tumor growth, in concert with a decreasing LPS level in the portal vein, suggesting that the gut microbiota serves as a source of TLR4 ligand(s) and that the Ly6C-positive macrophages play a role in tumor development in *Pten* null mice. Collectively, these data indicate that gut-derived LPS-induced inflammation via TLR4 on macrophages and TLR4-mediated inflammation result in HCC.

Melanocortin 4 receptor KO mice

Melanocortin 4 receptor (MC4R), a seven-transmembrane G protein-coupled receptor, is involved in regulation of body weight; hence, *MC4R* gene mutation is the major monogenic origin of obesity in human^[27,28]. Feeding of a high-fat diet to MC4R-deficient (MC4R-KO) mice for 20 wk and 1 year leads to NASH and multiple well-differentiated HCC formations in the liver, respectively^[13]. Similar to the findings in *Pten* null mice, Konuma *et al.*^[29]

found that highly-purified EPA treatment of MC4R-KO mice effectively inhibited the development of liver fibrosis without affecting body weight.

According to their previous study, hepatic crown-like structures (hCLSs), a unique histological feature, were found to play a pivotal role in the progression from simple steatosis to NASH^[30], with EPA markedly suppressing hCLS formation and fibrosis *via* prevention of hepatocyte injury. Thus, it was concluded that the beneficial effect of EPA involved the hCLSs. In addition, canagliflozin (CANA, a sodium glucose cotransporter 2 inhibitor and antidiabetic drug) was shown to attenuate NASH-HCC in another study^[31]. Based on the evidence that CANA induces adipose expansion without promoting macrophage augmentation, inflammation or fibrosis and altered glutathione metabolism to reduce oxidative stress in adipose tissue, the authors concluded that the decreased hepatic fat accumulation upon CANA treatment suppresses hepatic inflammation, fibrosis at 20 wk and subsequent NASH-HCC at 52 wk in Western diet-fed MC4R-KO mice.

STAM mice

The STAM mouse model was generated by neonatal male C57BL/6J mice exposure to low-dose streptozotocin at 2 d after birth followed by feeding with a high-fat diet after 4 wk of age^[14]. As a result, NASH developed at 8 wk and HCC at 16–20 wk. This mouse model has specific positive features, such as the average duration of HCC occurrence being within 16–20 wk of age, the number of HCC nodules being over 4 in any single mouse, the basal liver function being relatively preserved and there being no visible metastasis in the entire body^[32]. Moreover, this model has the substantial benefit of its HCC development from NASH being identical to the known progression in human patients, but with the whole process being completed within a relatively short period of time.

By using the STAM model, four studies have uncovered several of the mechanisms underlying NASH-HCC. First, Lau *et al.*^[33] demonstrated that cancer-associated fibroblasts, which regulate liver tumor-initiating cells, are augmented in parallel with increasing human growth factor (HGF) level during fibrosis and that HGF-induced FRA1 activation is related to fibrosis-dependent HCC development. These data suggest that cancer-associated fibroblast-derived, HGF-mediated FRA1 can be a new therapeutic target for NASH-HCC. Second, Fernandes *et al.*^[34] showed that solithromycin, a novel macrolide antibiotic, suppressed NASH, fibrosis and NASH-HCC by modulating the gluconeogenesis pathway, in particular the components of fructose 1, 6-bisphosphatase and glucose-6-phosphatase which are regulated by protein kinase C epsilon. Solithromycin improved the hepatic morphological features, such as the hepatocyte ballooning degeneration, and functions, as evidenced by reduction in NAFLD activity score along with decreased inflammation, fibrosis and HCC progression. This mechanism was ultimately suggested as a candidate

factor of novel treatment of NASH-HCC.

Third, Conti *et al.*^[35] revealed that aberrant expression of hepatic micro (mi)RNAs, such as miR-34a-5p, miR-93-5p, miR-221-3p and miR-222-3p, indicates their mechanistic significance in NASH-HCC tumorigenesis; specifically, 10 over-expressed miRNAs were identified. It is well known that human HCC tumorigenesis is associated with extensive genomic alterations. Therefore, the authors concluded that the altered expression profile of these miRNAs could be a surrogate marker for the initiation and progression of NASH-HCC.

Finally, based on the confirmed finding that NASH-HCC is associated with metabolic alterations in hepatic lipid homeostasis, Pogribny *et al.*^[36] indicated that one of the specific features of NASH-HCC is a significant dysregulation of 1-carbon homeostasis, with decreased expression of key 1-carbon metabolism genes, especially of the S-adenosylhomocysteine hydrolase (*Ahcy*) gene, and increased expression of the S-adenosyl-L-homocysteine (*SAH*) gene. Their results suggest that the inhibition of *Ahcy* expression may be a trigger of SAH elevation and subsequent progression of NASH-HCC.

Augmenter of liver regeneration-KO mice

Augmenter of liver regeneration (ALR), a hepatic growth factor, is widely known as a pleiotropic protein. ALR is critical for mitochondrial function, lipid homeostasis and cell survival. Gandhi *et al.*^[15] generated a liver-specific ALR-L-KO mouse and reported that depletion of hepatic ALR caused steatosis, mitochondrial degeneration and apoptosis of hepatocytes at 2 wk of age. These effects were followed by consecutive cell death, sustained inflammation at 4 wk, fibrosis/cirrhosis at 8 wk and eventually HCC formation (in 60%) at 1 year. Thus, it was theorized that inhibition of ALR synthesis in hepatocytes could lead to mitochondrial dysfunction and cell death, resulting in consecutive NASH and HCC occurrence.

FUTURE PERSPECTIVES FOR THE STUDY OF NASH-HCC BY ANIMAL MODELS

The “two-hit” hypothesis of the underlying mechanism of NASH-HCC involves the excessive accumulation of lipids in liver as the first step, thereby promoting sensitization to LPS, oxidative stress and inflammatory cytokines, representing the second hit^[37–39] (Figure 1). Recently, Tilg and Moschen^[40] proposed a “multiple-hit” hypothesis, in which various factors derived from gut and adipose tissue might take place in parallel during the progression from NAFLD to NASH. However, the definitive mechanisms in the progression from simple fatty liver to NASH and HCC are still under investigation, due to the inherent complexity of the functional combination of several factors. For some time, it was believed that the lack of appropriate animal models which were able to sufficiently reflect the actual

process of human NASH-HCC progression was the main obstacle to such research^[41]. In recent years, however, the situation has changed according to the development and availability of several rodent models. Each model harbors different specific characteristics, including genetic background, obesity status, diet induction, *etc.* Thus, researchers can now evaluate the mechanisms of NASH-HCC related to a specific factor/parameter by using these animal models.

According to the overall analyses of hepatocarcinogenesis in each of the mouse models discussed above, it is the STAM mice that generate HCC unfailingly and most rapidly. The considerable demerit of this mouse model, however, is the obscurity of the original gene of tumorigenesis for HCC due to lack of genetic manipulation and the inclusion of diabetes and hyperlipidemia in the background. Genetic manipulation in mouse models, such as of the PTEN-KO or ALR-KO, is a useful means by which to clarify the role of a specific gene in the molecular foundation of NASH-HCC progression; although, the sequential progression to HCC in these models has a relatively long duration and HCC occurrence is uncertain.

It is still questionable whether or not these available mouse models represent the initiating and/or progression processes of *bona fide* human NASH-HCC. Furthermore, it is noteworthy that among actual NASH patients there are individual differences in degree of fibrosis and timing of tumorigenesis in liver. At the present time, however, it is undoubted that these mouse models are essential for investigating the underlying mechanisms of NASH-HCC. Therefore, the future research targets may move forward towards gaining a more comprehensive NASH-HCC evaluation by using these mouse models.

CONCLUSION

Several mouse models have become available in recent years that support investigation into the underlying mechanisms of NASH-HCC. In response to the growing demand for better management of NASH-HCC, further inquiries are expected by researchers upon selecting an appropriate NASH mouse model according to the specific mechanisms and/or therapeutic targets of interest. After that, we hope to get some breakthrough for new treatment or prevention of NASH-HCC in the near future.

REFERENCES

- 1 **Rinella ME.** Nonalcoholic fatty liver disease: a systematic review. *JAMA* 2015; **313**: 2263-2273 [PMID: 26057287 DOI: 10.1001/jama.2015.5370]
- 2 **Younossi ZM,** Koenig AB, Abdelatif D, Fazel Y, Henry L, Wymer M. Global epidemiology of nonalcoholic fatty liver disease-Metanalytic assessment of prevalence, incidence, and outcomes. *Hepatology* 2016; **64**: 73-84 [PMID: 26707365 DOI: 10.1002/hep.28431]
- 3 **Neuschwander-Tetri BA.** Non-alcoholic fatty liver disease. *BMC Med* 2017; **15**: 45 [PMID: 28241825 DOI: 10.1186/s12916-017-0806-8]
- 4 **Farrell GC,** Larter CZ. Nonalcoholic fatty liver disease: from steatosis to cirrhosis. *Hepatology* 2006; **43**: S99-S112 [PMID: 16447287 DOI: 10.1002/hep.20973]
- 5 **Rozman D.** From nonalcoholic Fatty liver disease to hepatocellular carcinoma: a systems understanding. *Dig Dis Sci* 2014; **59**: 238-241 [PMID: 24385011 DOI: 10.1007/s10620-013-2998-x]
- 6 **Nooureddin M,** Mato JM, Lu SC. Nonalcoholic fatty liver disease: update on pathogenesis, diagnosis, treatment and the role of S-adenosylmethionine. *Exp Biol Med* (Maywood) 2015; **240**: 809-820 [PMID: 25873078 DOI: 10.1177/1535370215579161]
- 7 **Duvnjak M,** Lerotić I, Barsić N, Tomasić V, Virović Jukić L, Velagić V. Pathogenesis and management issues for non-alcoholic fatty liver disease. *World J Gastroenterol* 2007; **13**: 4539-4550 [PMID: 17729403 DOI: 10.3748/wjg.v13.i34.4539]
- 8 **Li L,** Li H, Garzel B, Yang H, Sueyoshi T, Li Q, Shu Y, Zhang J, Hu B, Heyward S, Moeller T, Xie W, Negishi M, Wang H. SLC13A5 is a novel transcriptional target of the pregnane X receptor and sensitizes drug-induced steatosis in human liver. *Mol Pharmacol* 2015; **87**: 674-682 [PMID: 25628225 DOI: 10.1124/mol.114.097287]
- 9 **Wong RJ,** Aguilar M, Cheung R, Perumpail RB, Harrison SA, Younossi ZM, Ahmed A. Nonalcoholic steatohepatitis is the second leading etiology of liver disease among adults awaiting liver transplantation in the United States. *Gastroenterology* 2015; **148**: 547-555 [PMID: 25461851 DOI: 10.1053/j.gastro.2014.11.039]
- 10 **Goldberg D,** Ditch IC, Saeian K, Lalehzari M, Aronsohn A, Gorospe EC, Charlton M. Changes in the Prevalence of Hepatitis C Virus Infection, Nonalcoholic Steatohepatitis, and Alcoholic Liver Disease Among Patients With Cirrhosis or Liver Failure on the Waitlist for Liver Transplantation. *Gastroenterology* 2017; **152**: 1090-1099.e1 [PMID: 28088461 DOI: 10.1053/j.gastro.2017.01.003]
- 11 **Starley BQ,** Calcagno CJ, Harrison SA. Nonalcoholic fatty liver disease and hepatocellular carcinoma: a weighty connection. *Hepatology* 2010; **51**: 1820-1832 [PMID: 20432259 DOI: 10.1002/hep.23594]
- 12 **Horie Y,** Suzuki A, Kataoka E, Sasaki T, Hamada K, Sasaki J, Mizuno K, Hasegawa G, Kishimoto H, Iizuka M, Naito M, Enomoto K, Watanabe S, Mak TW, Nakano T. Hepatocyte-specific Pten deficiency results in steatohepatitis and hepatocellular carcinomas. *J Clin Invest* 2004; **113**: 1774-1783 [PMID: 15199412 DOI: 10.1172/JCI20513]
- 13 **Itoh M,** Suganami T, Nakagawa N, Tanaka M, Yamamoto Y, Kamei Y, Terai S, Sakaida I, Ogawa Y. Melanocortin 4 receptor-deficient mice as a novel mouse model of nonalcoholic steatohepatitis. *Am J Pathol* 2011; **179**: 2454-2463 [PMID: 21906580 DOI: 10.1016/j.ajpath.2011.07.014]
- 14 **Fujii M,** Shibazaki Y, Wakamatsu K, Honda Y, Kawauchi Y, Suzuki K, Arumugam S, Watanabe K, Ichida T, Asakura H, Yoneyama H. A murine model for non-alcoholic steatohepatitis showing evidence of association between diabetes and hepatocellular carcinoma. *Med Mol Morphol* 2013; **46**: 141-152 [PMID: 23430399 DOI: 10.1007/s00795-013-0016-1]
- 15 **Gandhi CR,** Chaillet JR, Nalesnik MA, Kumar S, Dangi A, Demetris AJ, Ferrell R, Wu T, Divanovic S, Stankeiwicz T, Shaffer B, Stolz DB, Harvey SA, Wang J, Starzl TE. Liver-specific deletion of augments liver regeneration accelerates development of steatohepatitis and hepatocellular carcinoma in mice. *Gastroenterology* 2015; **148**: 379-391.e4 [PMID: 25448926 DOI: 10.1053/j.gastro.2014.10.008]
- 16 **Carpentier YA,** Portois L, Malaisse WJ. n-3 fatty acids and the metabolic syndrome. *Am J Clin Nutr* 2006; **83**: 1499S-1504S [PMID: 16841860]
- 17 **Ishii H,** Horie Y, Ohshima S, Anezaki Y, Kinoshita N, Dohmen T, Kataoka E, Sato W, Goto T, Sasaki J, Sasaki T, Watanabe S, Suzuki A, Ohnishi H. Eicosapentaenoic acid ameliorates steatohepatitis and hepatocellular carcinoma in hepatocyte-specific Pten-deficient mice. *J Hepatol* 2009; **50**: 562-571 [PMID: 19162361 DOI: 10.1016/j.jhep.2008.10.031]

- 18 **Shan X**, Czar MJ, Bunnell SC, Liu P, Liu Y, Schwartzberg PL, Wange RL. Deficiency of PTEN in Jurkat T cells causes constitutive localization of Itk to the plasma membrane and hyperresponsiveness to CD3 stimulation. *Mol Cell Biol* 2000; **20**: 6945-6957 [PMID: 10958690]
- 19 **Su R**, Li Z, Li H, Song H, Bao C, Wei J, Cheng L. Grp78 promotes the invasion of hepatocellular carcinoma. *BMC Cancer* 2010; **10**: 20 [PMID: 20082722 DOI: 10.1186/1471-2407-10-20]
- 20 **Luk JM**, Lam CT, Siu AF, Lam BY, Ng IO, Hu MY, Che CM, Fan ST. Proteomic profiling of hepatocellular carcinoma in Chinese cohort reveals heat-shock proteins (Hsp27, Hsp70, GRP78) up-regulation and their associated prognostic values. *Proteomics* 2006; **6**: 1049-1057 [PMID: 16400691 DOI: 10.1002/pmic.200500306]
- 21 **Chen WT**, Zhu G, Pfaffenbach K, Kanel G, Stiles B, Lee AS. GRP78 as a regulator of liver steatosis and cancer progression mediated by loss of the tumor suppressor PTEN. *Oncogene* 2014; **33**: 4997-5005 [PMID: 24141775 DOI: 10.1038/onc.2013.437]
- 22 **Miura K**, Ishioka M, Minami S, Horie Y, Ohshima S, Goto T, Ohnishi H. Toll-like Receptor 4 on Macrophage Promotes the Development of Steatohepatitis-related Hepatocellular Carcinoma in Mice. *J Biol Chem* 2016; **291**: 11504-11517 [PMID: 27022031 DOI: 10.1074/jbc.M115.709048]
- 23 **Kawai T**, Akira S. The role of pattern-recognition receptors in innate immunity: update on Toll-like receptors. *Nat Immunol* 2010; **11**: 373-384 [PMID: 20404851 DOI: 10.1038/ni.1863]
- 24 **Miura K**, Kodama Y, Inokuchi S, Schnabl B, Aoyama T, Ohnishi H, Olefsky JM, Brenner DA, Seki E. Toll-like receptor 9 promotes steatohepatitis by induction of interleukin-1 β in mice. *Gastroenterology* 2010; **139**: 323-34.e7 [PMID: 20347818 DOI: 10.1053/j.gastro.2010.03.052]
- 25 **Miura K**, Yang L, van Rooijen N, Brenner DA, Ohnishi H, Seki E. Toll-like receptor 2 and palmitic acid cooperatively contribute to the development of nonalcoholic steatohepatitis through inflammasome activation in mice. *Hepatology* 2013; **57**: 577-589 [PMID: 22987396 DOI: 10.1002/hep.26081]
- 26 **Tacke F**, Zimmermann HW. Macrophage heterogeneity in liver injury and fibrosis. *J Hepatol* 2014; **60**: 1090-1096 [PMID: 24412603 DOI: 10.1016/j.jhep.2013.12.025]
- 27 **Balthasar N**, Dalgard LT, Lee CE, Yu J, Funahashi H, Williams T, Ferreira M, Tang V, McGovern RA, Kenny CD, Christiansen LM, Edelstein E, Choi B, Boss O, Aschkenasi C, Zhang CY, Mountjoy K, Kishi T, Elmquist JK, Lowell BB. Divergence of melanocortin pathways in the control of food intake and energy expenditure. *Cell* 2005; **123**: 493-505 [PMID: 16269339 DOI: 10.1016/j.cell.2005.08.035]
- 28 **Vaisse C**, Clement K, Durand E, Hercberg S, Guy-Grand B, Froguel P. Melanocortin-4 receptor mutations are a frequent and heterogeneous cause of morbid obesity. *J Clin Invest* 2000; **106**: 253-262 [PMID: 10903341 DOI: 10.1172/JCI9238]
- 29 **Konuma K**, Itoh M, Suganami T, Kanai S, Nakagawa N, Sakai T, Kawano H, Hara M, Kojima S, Izumi Y, Ogawa Y. Eicosapentaenoic acid ameliorates non-alcoholic steatohepatitis in a novel mouse model using melanocortin 4 receptor-deficient mice. *PLoS One* 2015; **10**: e0121528 [PMID: 25816330 DOI: 10.1371/journal.pone.0121528]
- 30 **Itoh M**, Kato H, Suganami T, Konuma K, Marumoto Y, Terai S, Sakugawa H, Kanai S, Hamaguchi M, Fukaiishi T, Aoe S, Akiyoshi K, Komohara Y, Takeya M, Sakaida I, Ogawa Y. Hepatic crown-like structure: a unique histological feature in non-alcoholic steatohepatitis in mice and humans. *PLoS One* 2013; **8**: e82163 [PMID: 24349208 DOI: 10.1371/journal.pone.0082163]
- 31 **Shiba K**, Tsuchiya K, Komiya C, Miyachi Y, Mori K, Shimazu N, Yamaguchi S, Ogasawara N, Katoh M, Itoh M, Suganami T, Ogawa Y. Canagliflozin, an SGLT2 inhibitor, attenuates the development of hepatocellular carcinoma in a mouse model of human NASH. *Sci Rep* 2018; **8**: 2362 [PMID: 29402900 DOI: 10.1038/s41598-018-19658-7]
- 32 **Takakura K**, Koido S, Fujii M, Hashiguchi T, Shibazaki Y, Yoneyama H, Katagi H, Kajihara M, Misawa T, Homma S, Ohkusa T, Tajiri H. Characterization of non-alcoholic steatohepatitis-derived hepatocellular carcinoma as a human stratification model in mice. *Anticancer Res* 2014; **34**: 4849-4855 [PMID: 25202066]
- 33 **Lau EY**, Lo J, Cheng BY, Ma MK, Lee JM, Ng JK, Chai S, Lin CH, Tsang SY, Ma S, Ng IO, Lee TK. Cancer-Associated Fibroblasts Regulate Tumor-Initiating Cell Plasticity in Hepatocellular Carcinoma through c-Met/FRA1/HEY1 Signaling. *Cell Rep* 2016; **15**: 1175-1189 [PMID: 27134167 DOI: 10.1016/j.celrep.2016.04.019]
- 34 **Fernandes P**, Oldach D, Hashiguchi T, Shirakata Y, Yoneyama H, Gholam PM. Solithromycin Diminishes Steatohepatitis by Modulating Gluconeogenesis and Inhibits Tumor Growth in a Diabetic Mouse Model of Non-Alcoholic Steatohepatitis. *J Immunol Infect Inflam Dis* 2016; **14**: 17-19
- 35 **de Conti A**, Ortega JF, Tryndyak V, Dreval K, Moreno FS, Rusyn I, Beland FA, Pogribny IP. MicroRNA deregulation in nonalcoholic steatohepatitis-associated liver carcinogenesis. *Oncotarget* 2017; **8**: 88517-88528 [PMID: 29179453 DOI: 10.18632/oncotarget.19774]
- 36 **Pogribny IP**, Dreval K, Kindrat I, Melnyk S, Jimenez L, de Conti A, Tryndyak V, Pogribna M, Ortega JF, James SJ, Rusyn I, Beland FA. Epigenetically mediated inhibition of S-adenosylhomocysteine hydrolase and the associated dysregulation of l-carbon metabolism in nonalcoholic steatohepatitis and hepatocellular carcinoma. *FASEB J* 2018; **32**: 1591-1601 [PMID: 29127188 DOI: 10.1096/fj.201700866R]
- 37 **Day CP**, James OF. Steatohepatitis: a tale of two "hits"? *Gastroenterology* 1998; **114**: 842-845 [PMID: 9547102]
- 38 **Browning JD**, Horton JD. Molecular mediators of hepatic steatosis and liver injury. *J Clin Invest* 2004; **114**: 147-152 [PMID: 15254578 DOI: 10.1172/JCI22422]
- 39 **Neuschwander-Tetri BA**. Hepatic lipotoxicity and the pathogenesis of nonalcoholic steatohepatitis: the central role of nontriglyceride fatty acid metabolites. *Hepatology* 2010; **52**: 774-788 [PMID: 20683968 DOI: 10.1002/hep.23719]
- 40 **Tilg H**, Moschen AR. Evolution of inflammation in nonalcoholic fatty liver disease: the multiple parallel hits hypothesis. *Hepatology* 2010; **52**: 1836-1846 [PMID: 21038418 DOI: 10.1002/hep.24001]
- 41 **Varela-Rey M**, Embade N, Ariz U, Lu SC, Mato JM, Martinez-Chantar ML. Non-alcoholic steatohepatitis and animal models: understanding the human disease. *Int J Biochem Cell Biol* 2009; **41**: 969-976 [PMID: 19027869 DOI: 10.1016/j.biocel.2008.10.027]

P- Reviewer: Gonzalez-Reimers E, Namisaki T, Peltec A

S- Editor: Gong ZM **L- Editor:** A **E- Editor:** Huang Y





Published by **Baishideng Publishing Group Inc**
7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
Telephone: +1-925-223-8242
Fax: +1-925-223-8243
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>



ISSN 1007-9327

