

# World Journal of *Gastroenterology*

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## Basic Study

# Effects of hepatitis E virus infection on interferon production *via* ISG15

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**Author contributions:** Wang M and Tian DY designed the research; Wang M, Huang Y, He M and Peng WJ performed the research; Tian DY contributed new reagents; Wang M and Tian DY analyzed the data; Wang M and Tian DY wrote the paper.

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## Abstract

### AIM

To assess the effects of hepatitis E virus (HEV) on the production of type I interferons (IFNs) and determine the underlying mechanisms.

### METHODS

We measured the production of interferon (IFN)-alpha and -beta ( $\alpha/\beta$ ) in genotype 3 HEV-infected C3A cells at different time points (0, 8, 12, 24, 48, 72 and 120 h) by enzyme-linked immunosorbent assay (ELISA). The expression levels of IFN-stimulated gene (ISG)15 in HEV-infected C3A cells at different time points were tested by western blotting. The plasmid-expressing open reading frame 3 (ORF3) or control plasmids (green fluorescent protein-expressing) were transfected into C3A cells, and the levels of IFN- $\alpha/\beta$  and ISG15 were evaluated, respectively. Furthermore, the plasmid-expressing ISG15 or small interfering RNA-inhibiting ISG15 was transfected into infected C3A cells. Then, the production of IFN- $\alpha/\beta$  was also measured by ELISA.

## RESULTS

We showed that genotype 3 HEV could enhance the production of IFN- $\alpha/\beta$  and induce elevation of ISG15 in C3A cells. HEV ORF3 protein could enhance the production of IFN- $\alpha/\beta$  and the expression of ISG15. Additionally, ISG15 silencing enhanced the production of IFN- $\alpha/\beta$ . Overexpression of ISG15 resulted in the reduction of IFN- $\alpha/\beta$ .

## CONCLUSION

HEV may promote production of IFN- $\alpha/\beta$  and expression of ISG15 *via* ORF3 in the early stages, and increased ISG15 subsequently inhibited the production of IFN- $\alpha/\beta$ .

**Key words:** Open reading frame 3; Interferon-stimulated gene 15; Interferon-alpha; Hepatitis E virus; Interferon-beta

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**Core tip:** To date, few studies have investigated the role of ISG15 in hepatitis E virus (HEV) infection and the impact of ISG15 on interferons (IFN) production. This study showed that HEV could inhibit the level of type I IFN through regulating the expression of IFN-stimulated gene (ISG)15, which may attenuate the IFN response, allowing for successful infection of their hosts. The present study enhances the understanding of the interaction between ISG15 and HEV in the host innate immune response, which may provide useful insight for the development of new antiviral drugs and antiviral strategies.

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## INTRODUCTION

Hepatitis E virus (HEV) infection is one of the most common causes of acute hepatitis or sporadic acute hepatitis in many parts of the world<sup>[1]</sup>. At least 20 million HEV infections occur annually, which are mostly self-limiting and have different clinical manifestations, such as asymptomatic, acute liver failure and rare chronicity<sup>[2,3]</sup>. In some cases, HEV infections can cause up to 30% mortality in pregnant women<sup>[4]</sup> and may result in chronic hepatitis in immunocompromised individuals, such as those receiving organ transplants or chemotherapy, and individuals infected with human immunodeficiency virus<sup>[5-7]</sup>. Currently, at least four genotypes exist among HEV strains, of which HEV genotypes 1 and 2 are obligate human pathogens and HEV genotypes 3 and 4 are mostly zoonotic<sup>[8]</sup>. Recently, a study found that a genotype 3 HEV isolated from a chronically infected patient could be adapted to grow in human C3A hepatoma cells<sup>[9]</sup>.

HEV has a 7.2-kb single-stranded, positive-sense RNA genome containing three open reading frames (ORFs)<sup>[10]</sup>. The ORF1 protein comprises the functional domains of virions. ORF2 is the major structural protein of virions and encodes the viral capsid protein. And, ORF3 protein, a small molecule phosphoprotein, is involved in viral particle secretion<sup>[11]</sup>. A recent study found that chronic HEV infections were almost exclusively caused by the zoonotic genotype 3 HEV strains<sup>[12]</sup>.

Type I interferons (IFNs) are key components of innate immunity and known to be the first-line defense against virus infection. IFN exerts many biological effects, such as antiviral and antitumor activity, and immune regulation by activating hundreds of downstream IFN-stimulated genes (ISGs)<sup>[13,14]</sup>. One of the most abundantly IFN-induced proteins is ISG15, which is encoded and expressed as a ubiquitin-like protein. ISG15 is covalently attached to target proteins through a C-terminal LRLRG motif, in a process called ISGylation. ISGylation could alter protein properties directly by addition of ISG15, as well as by reducing the degradation of the target protein by competing with ubiquitin conjugation<sup>[15]</sup>. Previous studies have shown that ISG15 is critical for control of certain viral and bacterial infections and is linked to the process of budding or egress for many RNA virus families<sup>[16-18]</sup>.

As few studies have focused on the role of ISG15 in HEV infection, we investigated whether HEV infection could regulate the expression of ISG15 and the impact of ISG15 on IFN production during HEV infection in the present study, which is of great significance to expand our understanding of the interaction between ISG15 and HEV pathogenesis.

## MATERIALS AND METHODS

### Cells and virus

C3A is a derivative of HepG2 cells (hepatoma cells). C3A cells were obtained from the American Type Culture Collection (ATCC) (Manassas, VA, United States) and cultured in Dulbecco's Modified Eagle's Medium supplemented with 10% fetal bovine serum (Gibco, Thermo Scientific, Waltham, MA, United States), penicillin (100 U/mL) and streptomycin (100  $\mu$ g/mL) at 37 °C with 5% CO<sub>2</sub>. The genotype 3 Kernow-C1 HEV infectious cDNA clone was a kind gift from the National Institute of Diagnostics and Vaccine Development in Infectious Diseases (NIDVD, Xiamen, China). The ATG start codon of ORF3 in Kernow-C1 HEV was altered to GTG to construct an HEV ORF3 mutant, which led to the loss of ORF3 protein expression.

### Reagents and plasmids

Anti-ISG15 antibody was purchased from Signalway Antibody (42509, 1:1000 dilution; Pearland, TX, United States). AntiHEV ORF3-specific antibody was purchased from Abbiotec (250688, 1:1000; San Diego, CA, United States). Primary antibody against  $\beta$ -actin (#BM0627, 1:400 dilution) was purchased from Boster (Wuhan, China). We constructed a plasmid-expressing



**Table 1** Primers used in this study

Name	Primer	Sequence
β-actin	Forward	5'- AGCGAGCATCCCCAAAGTT -3'
	Reverse	5'- GGGCACGAAGGCTCATCATT -3'
ISG15	Forward	5'- GCGCAGATCACCCAGAAGAT -3'
	Reverse	5'- GTTCGTCGCATTGTCCACC -3'
siRNA1	Forward	5'- UGUCGGUGUCAGAGCUGAATT -3'
	Reverse	5'- UUCAGCUCUGACACCGACATT -3'
siRNA2	Forward	5'- GAUGCUGGCGGGCAACGAATT -3'
	Reverse	5'- UUCGUUGCCCGCCAGCAUUCTT -3'
siRNA3	Forward	5'- UGAGCACCGUGUUGAUGAATT -3'
	Reverse	5'- UUCAUGAACACGGUGCUCATT -3'
Negative control	Forward	5'- UUCUCCGAACGUGUCACGUTT -3'
	Reverse	5'- ACGUGACACGUUCGGAGAATT -3'

ORF3 (ORF3) as well as a plasmid-expressing ISG15 (ISG15). ORF3 and ISG15 fragments were cloned into the pEGFP-N1 vector (green fluorescent protein (GFP)-expressing, GenBank accession #55762) by digesting with *Xho* I and *Hind* III to construct the pORF3-EGFP plasmid (ORF3) and pISG15-EGFP plasmid (ISG15). All the clones were confirmed by restriction digestion and DNA sequencing.

#### HEV infection

The cDNA clones were linearized with *Bgl* II, and the genomic RNA was transcribed using the MEGAScript Kit (Ambion, Thermo Scientific). C3A cells cultured in a six-well plate were transfected with capped RNA transcripts from Kernow-C1 HEV infectious cDNA clone using DMR1E-C reagent (Invitrogen, Grand Island, NY, United States) following the manufacturer's protocol. The ratio of RNA to DMR1E-C was 2 µg:8 µL. After 6 h, the medium in the six-well plate was removed and replaced with serum-free medium. The cells and cell culture supernatants were harvested at different time points (0, 8, 12, 24, 48, 72 and 120 h) and used in experiments.

#### Cell transfection

C3A cells were plated in six-well microplates and transfected with a plasmid-expressing ORF3-EGFP (ORF3) or the HEV mutant, leading to the loss of ORF3 protein expression (ΔORF3) using Lipofectamine TM 3000 Transfection Reagent (Invitrogen, San Diego, CA, United States) for 48 h. Then, the cells and cell culture supernatants were harvested at different time points (0, 8, 12, 24, 48, 72 and 120 h) and used in experiments.

#### Small interfering RNA-mediated knockdown

The small interfering RNAs (siRNAs) targeted against ISG15 were purchased from Santa Cruz Biotechnologies (Dallas, TX, United States). C3A cells were transiently transfected with 20 nmol/L of siRNA using the DMR1E-C reagent (Invitrogen) according to the manufacturer's protocol. Primers are shown in Table 1.

#### Enzyme-linked immunosorbent assay

The concentrations of IFN-α and -β (α/β) were evaluated in cell culture supernatants using a human IFN-α/β ELISA kit (Joyee Biotechnics, Shanghai, China).

All samples were tested in triplicate.

#### Western blot analysis

Cells were lysed in RIPA buffer (Beyotime Biotechnology, Shanghai, China), and the protein concentration was determined using an enhanced BCA protein assay kit (Beyotime Biotechnology). Ten micrograms of each protein sample were separated on 12% sodium dodecyl sulfate polyacrylamide gel electrophoresis and subsequently transferred to polyvinylidene difluoride membranes. Samples were then incubated with specific primary antibodies overnight, following incubation with an antimouse secondary antibody solution for 2 h at room temperature. The blotted membranes were exposed to film, and the protein bands were visualized by a scanner system.

#### Real-time reverse transcriptase-polymerase chain reaction

Total RNA from C3A cells was isolated using Trizol (Thermo Scientific) and reverse-transcribed into cDNA using the TaqMan Reverse Transcription Reagents (Thermo Scientific). The mRNA level of ISG15 was quantified by real-time PCR using SYBR master mix (Thermo Scientific). The PCR amplification conditions included 1 cycle of 95 °C for 2 min, and 40 cycles of 95 °C for 10 s, 60 °C for 20 s, and 72 °C for 30 s; the Roche Light Cycler 480 II Real-Time PCR System was used. Threshold cycle (CT) values were determined by RT-PCR and normalized by the housekeeping gene β-actin. The mRNA abundance was calculated using the  $2^{-\Delta\Delta Ct}$  method and expressed as fold-change. Primers are shown in Table 1.

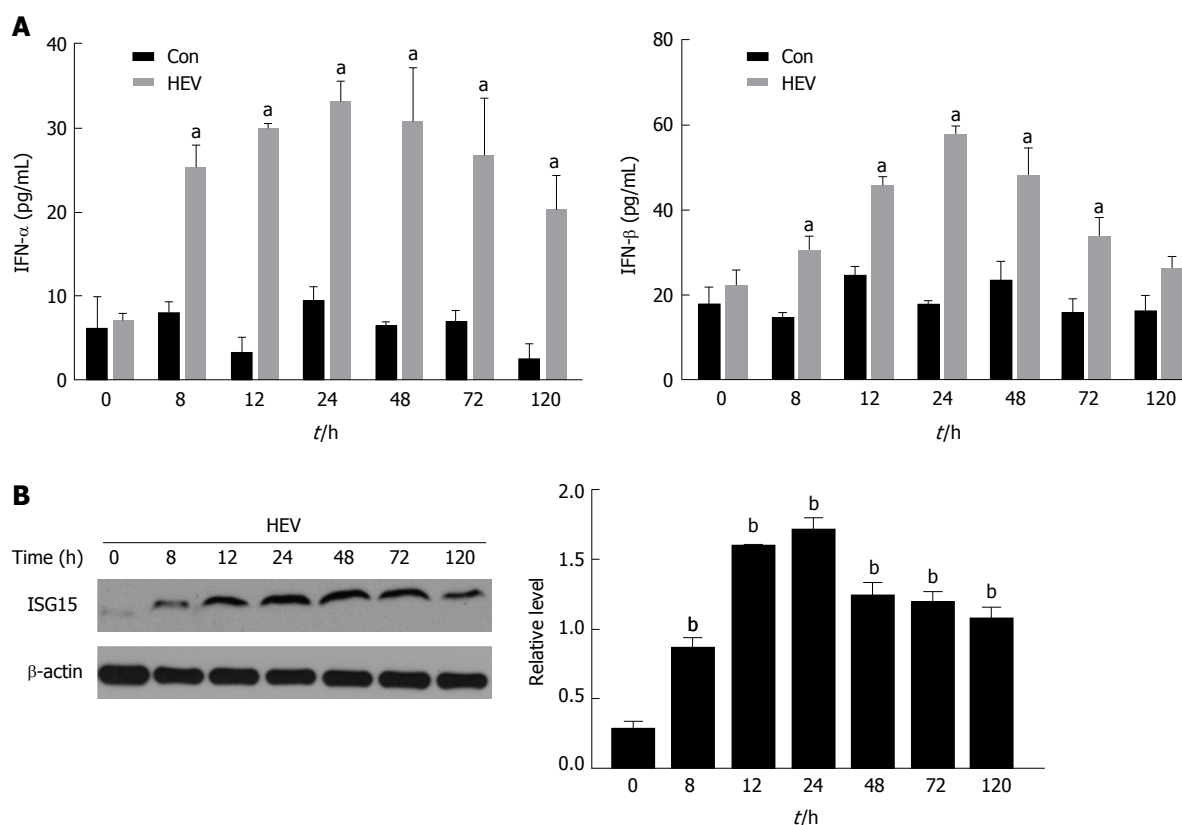
#### Statistical analysis

All the data are presented as mean ± SD. Statistical differences between the samples and the controls were assessed by the Student's *t*-test. *P* < 0.05 was considered statistically significant.

## RESULTS

#### HEV infection enhances the production of IFN-α/β and induces elevation of ISG15 protein in C3A cells

In this study, C3A cells were transfected with HEV RNA.



**Figure 1** Hepatitis E virus infection enhances the production of Interferon- $\alpha$  and - $\beta$  and induces elevation of Interferon-stimulated gene 15 protein in C3A cells. A: The concentrations of IFN- $\alpha$  and  $\beta$  were evaluated in cell culture supernatants at different time points (0, 8, 12, 24, 48, 72 and 120 h) by enzyme-linked immunosorbent assay; B: The expression of ISG15 protein replication was detected at different time points (0, 8, 12, 24, 48, 72 and 120 h) by western blotting. The data are expressed as mean  $\pm$  standard error of the mean. <sup>a</sup> $P < 0.05$ , <sup>b</sup> $P < 0.01$ . HEV: Hepatitis E virus; IFN: Interferon; ISG: IFN-stimulated gene.

To determine whether HEV infection could affect the production of type I IFN, the concentrations of IFN- $\alpha/\beta$  were evaluated in cell culture supernatants at different time points (0, 8, 12, 24, 48, 72 and 120 h) by ELISA. The levels of IFN- $\alpha/\beta$  in the HEV infection group were significantly increased at each time point ( $P < 0.05$ ), and the levels of IFN- $\alpha/\beta$  reached the maximum at 24 h, while no significant changes were observed in the control group (Figure 1A).

Next, we identified whether HEV could induce ISG15. ISG15 protein expression was detected at different time points by western blotting. It showed HEV enhanced the expression of ISG15 ( $P < 0.01$ ) and ISG15 protein expression reach the maximum at 24 h as compared to the control group (Figure 1B).

#### HEV ORF3 protein enhances the production of IFN- $\alpha/\beta$ and the expression of ISG15 protein in C3A cells

To investigate how HEV enhances the production of IFN- $\alpha/\beta$  and induces elevation of ISG15, we constructed a plasmid-expressing ORF3-EGFP (ORF3) and the HEV mutant leading to the loss of ORF3 protein expression ( $\Delta$ ORF3). C3A cells were transfected with ORF3 or  $\Delta$ ORF3 respectively, and the production of IFN- $\alpha/\beta$  and the expression of ISG15 protein were detected. The production of IFN- $\alpha/\beta$  was significantly increased at each time point in the group transfected with ORF3 plasmid (ORF3 group) compared to the ORF3-lacking group ( $\Delta$ ORF3 group) ( $P < 0.05$ ). In addition, the

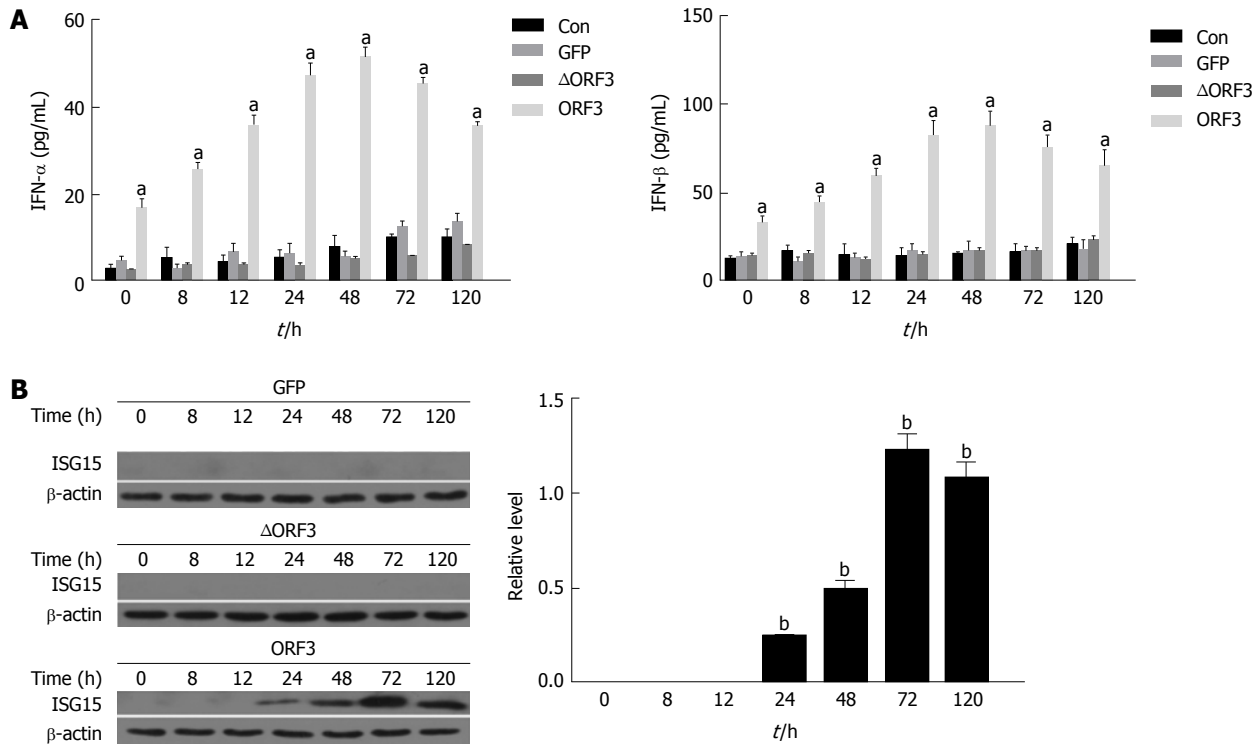
concentrations of IFN- $\alpha/\beta$  increased the maximum at 24 h (Figure 2A). The ORF3 group could induce higher ISG15 protein expression than  $\Delta$ ORF3 group ( $P < 0.01$ ). ISG15 protein was poorly expressed in the early stage, from 0 h to 12 h, and significantly increased at 24 h, reaching the maximum at 72 h (Figure 2B).

#### ISG15 silencing enhances the production of IFN- $\alpha/\beta$ in C3A cells

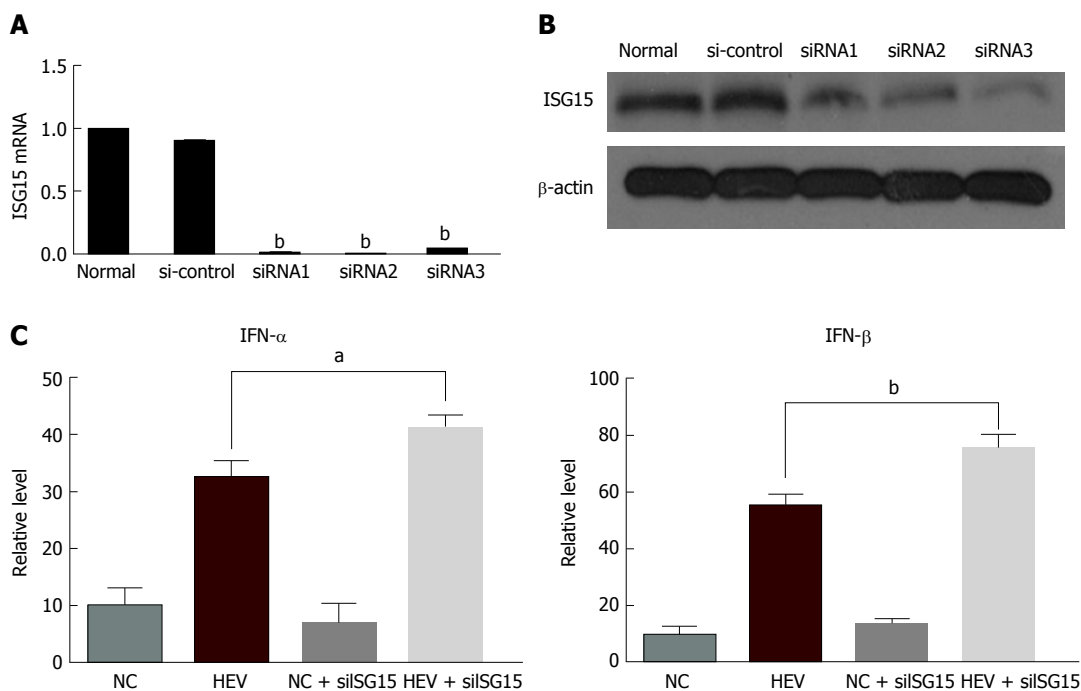
To investigate the levels of IFN- $\alpha/\beta$  when ISG15 was inhibited, we first constructed three siRNAs that could inhibit the expression of ISG15 and then transfected them into C3A cells, respectively. Then, the mRNA and protein levels of ISG15 were detected to determine which siRNA had the best inhibitory effect (Figure 3A and B). The siRNA2 was found to have the best inhibitory effect on ISG15. Next, the siRNA2 was transfected into C3A cells that had been pretreated with HEV RNA. The expression of IFN- $\alpha$  was significantly increased in the HEV-infected C3A cells transfected with siISG15 compared to the HEV-infected cells ( $P < 0.05$ ). Consistently, the expression of IFN- $\beta$  showed the same trend ( $P < 0.01$ ) (Figure 3C).

#### Overexpression of ISG15 results in the reduction of IFN- $\alpha/\beta$ in C3A cells

To investigate the effect of ISG15 overexpression on IFN- $\alpha/\beta$ , the plasmid -expressing ISG15 was constructed and transfected into C3A cells. The level of ISG15 was

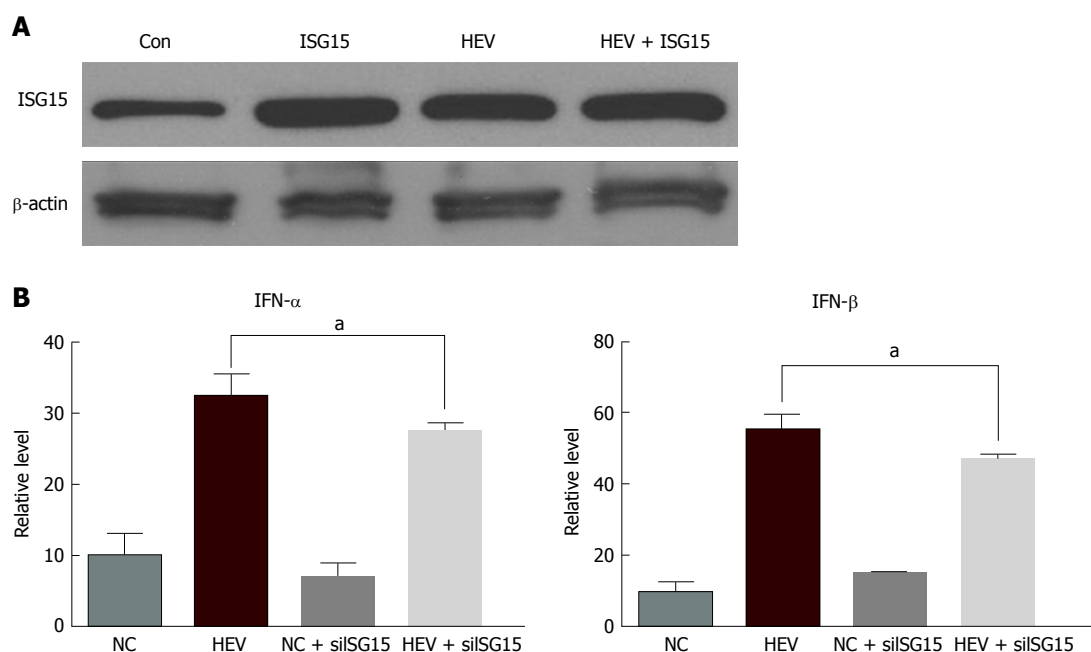


**Figure 2** Hepatitis E virus open reading frame 3 protein enhances the production of Interferon- $\alpha$  and  $\beta$  and the expression of Interferon-stimulated gene 15 protein in C3A cells. C3A cells were transfected with 2  $\mu$ g of either pORF3-EGFP (ORF3) or pEGFP-N1 empty vector (GFP), and the HEV mutant, leading to the loss of ORF3 protein expression ( $\Delta$ ORF3) for 48 h, untreated cells were used as the control (Con). Then, the cells and cell culture supernatants were harvested at different time points (0, 8, 12, 24, 48, 72 and 120 h). A: The concentrations of IFN- $\alpha$  and  $\beta$  were evaluated in cell culture supernatants at different time points (0, 8, 12, 24, 48, 72, 120 h) by enzyme-linked immunosorbent assay; B: The expression of ISG15 protein was detected at different time points (0, 8, 12, 24, 48, 72 and 120 h) by western blotting. The data are expressed as mean  $\pm$  standard error of the mean. <sup>a</sup> $P < 0.05$ , <sup>b</sup> $P < 0.01$ . GFP: Green fluorescent protein; HEV: Hepatitis E virus; IFN: Interferon; ISG: IFN-stimulated gene; ORF: Open reading frame.



**Figure 3** Interferon-stimulated gene 15 silencing enhances the production of Interferon- $\alpha/\beta$  in C3A cells. Three siRNA inhibiting ISG15 (siISG15) and control-siRNA (si-control) were constructed, and C3A cells were cotransfected with 20 nmol/L ISG15-targeted siRNA (siISG15) or control-siRNA (si-control) along with HEV RNA. A: The mRNA level of ISG15 was estimated by reverse transcription-PCR; B: The protein level of ISG15 was estimated by western blotting. Next, the siRNA2 was transfected into C3A cells that had been pretreated with HEV RNA, and the impact of siISG15 on IFN was investigated; C: The level of IFN- $\alpha/\beta$  was tested by enzyme-linked immunosorbent assay. The data are expressed as mean  $\pm$  standard error of the mean. <sup>a</sup> $P < 0.05$ , <sup>b</sup> $P < 0.01$ . HEV: Hepatitis E virus; IFN: Interferon; ISG: IFN-stimulated gene; si: Small interfering.





**Figure 4** Over-expression of Interferon-stimulated gene 15 results in the reduction of Interferon- $\alpha/\beta$  in C3A cells. C3A cells were transfected with 2 mg ISG15-EGFP plasmid (ISG15) along with HEV RNA. A: ISG15 protein expression was detected by western blotting; B: The levels of IFN- $\alpha/\beta$  were detected by enzyme-linked immunosorbent assay. <sup>a</sup> $P < 0.05$ . HEV: Hepatitis E virus; IFN: Interferon; ISG: IFN-stimulated gene.

detected by western blotting. The cells transfected with ISG15 plasmid had a considerably higher ISG15 level than the control cells (Figure 4A). Then, the production of IFN- $\alpha/\beta$  was tested when cells were transfected with ISG15 plasmid and HEV RNA. The level of IFN- $\alpha/\beta$  was decreased when the cells were treated with HEV and ISG15 plasmid ( $P < 0.05$ ) (Figure 4B), which indicated over-expression of ISG15 may result in reduction of IFN- $\alpha/\beta$ .

## DISCUSSION

The innate immune system is the first line of defense against invading pathogens. Type I IFNs, such as IFN- $\alpha/\beta$ , make up part of the innate immunity system and are critical for innate immunity against viral infection. ISG15, a member of the ISGs induced by IFN, has an important role in the regulation of antiviral immunity. In this study, we demonstrated that HEV infection could enhance the production of IFN- $\alpha/\beta$  and induce elevation of ISG15 protein. ORF3 protein may be responsible for the enhancement of IFN- $\alpha/\beta$  and ISG15 by HEV. Furthermore, ISG15 silencing enhanced the production of IFN- $\alpha/\beta$  in C3A cells. Over-expression of ISG15 resulted in the reduction of IFN- $\alpha/\beta$  in C3A cells. Collectively, these results indicated that HEV could induce the production of IFN and ISG15, and the increased ISG15 in turn reduced the level of IFN, which revealed a possible correlation between HEV infection and IFN production by regulating the expression of ISG15.

In our study, we first found that HEV infection could enhance the production of IFN- $\alpha/\beta$  and induce the elevation of ISG15 protein. A previous study demon-

strated that most viruses have the ability to regulate the IFN response during infection<sup>[19]</sup>. When cells are infected by viruses, such as influenza viruses, the activated host innate immune response will lead to secretion of type I IFN. Then, the induced IFN exerts its antiviral activity<sup>[20]</sup>. Over the last decade, numerous studies have reported that ISG15 plays a crucial role against viral infection<sup>[21,22]</sup>. However, few studies have investigated the effects of HEV on ISG15. In the present study, we found that the expression of ISG15 and the IFN level were simultaneously increased during HEV infection, which indicated that HEV could induce ISG15 expression in C3A cells. Recent studies have revealed that ISG15, as an innate immune protein with broad spectrum antiviral activity, continues to accumulate rapidly when the host is infected by a virus<sup>[23]</sup>, which is consistent with the findings in this research.

HEV consists of three ORFs. Genotype 1 HEV strain could inhibit type I IFN induction by ORF1 products<sup>[24]</sup>. However, the ORF3 products of genotype 1 HEV have been reported to enhance the production of IFN<sup>[25]</sup>. Because we found that genotype 3 HEV could induce the level of IFN, we next tested the impact of HEV ORF3 on IFN. We found that HEV ORF3 could increase the levels of IFN- $\alpha/\beta$ , which indicates that HEV ORF3 is responsible for the regulation of IFN. Similarly, we investigated whether HEV ORF3 could affect the expression of ISG15. As we expected, the level of ISG15 was increased by HEV ORF3. Previous studies have shown that ORF3 protein is associated with the egress of HEV<sup>[26]</sup>. As ISG15 is also associated with the process of egress for many RNA virus families<sup>[27]</sup>, we supposed that HEV ORF3 may regulate the egress of HEV through ISG15, which

requires further experiments to confirm.

Furthermore, the possible correlation between enhanced levels of IFN and ISG15 during HEV infection was investigated. ISG15 silencing enhanced the production of IFN- $\alpha/\beta$ , and over-expression of ISG15 resulted in the reduction of IFN- $\alpha/\beta$  in C3A cells, indicating that ISG15 is a negative regulator of IFN. It has been indicated that the absence of intracellular ISG15 in the ISG15-deficient patients prevents the accumulation of USP18, which is a potent negative regulator of IFN- $\alpha/\beta$  signaling, thus resulting in the enhancement and amplification of IFN- $\alpha/\beta$  responses and increase of viral resistance in humans<sup>[15,28]</sup>.

In addition, it was found that ISG15 over-expression inhibited induction of IFN- $\beta$  by HCV infection<sup>[29]</sup>. Evidence revealing high hepatic ISG15 levels was associated with low antiviral IFN-response during the early phase of antiviral therapy<sup>[30]</sup>, supporting the notion that ISG15 is a negative regulator of the IFN system. However, this raises the question of why the levels of IFN- $\alpha/\beta$  could be reduced by HEV infection *via* ISG15. As we know, the IFN production is always enhanced to promote an antiviral state in an infected host, but viruses have evolved many strategies to antagonize the induction of IFNs. As there is little knowledge about how HEV inhibits induction of IFN, the result that the level of IFN could be inhibited by HEV *via* ISG15 will enhance our fundamental knowledge of the mechanisms of induction and evasion of type I IFN responses by HEV.

In conclusion, the findings of the present study showed that HEV could inhibit the level of type I IFN through regulating the expression of ISG15, which may attenuate the IFN response, allowing for successful infection of the host. The present study enhances the understanding of the interaction between ISG15 and HEV in the host innate immune response, which may provide useful insight for the development of new antiviral drugs and antiviral strategies.

## ARTICLE HIGHLIGHTS

### Research background

Hepatitis E virus (HEV) infection is one of the most common causes of acute hepatitis or sporadic acute hepatitis in the world. At least 20 million HEV infections occur annually, which may result in chronic hepatitis in immunocompromised individuals. However, the mechanism of HEV pathogenesis remains obscure.

### Research motivation

Over the last decade, numerous studies have reported that interferon (IFN)-stimulated gene (ISG)15 plays a crucial role against viral infection. However, few studies have investigated the effects of HEV on ISG15.

### Research objectives

In the present study, we investigated whether HEV infection could regulate the expression of ISG15 and the impact of ISG15 on IFN production during HEV infection, which is of great significance to expand our understanding of the interaction between ISG15 and HEV pathogenesis.

### Research methods

In this study, C3A cells were first transfected with genotype 3 HEV RNA. The production of IFN- $\alpha$  and - $\beta$  (IFN- $\alpha/\beta$ ) at different time points (0, 8, 12,

24, 48, 72 and 120 h) were measured by enzyme-linked immunosorbent assay (ELISA). The expression levels of ISG15 in HEV-infected C3A cells at different time points were tested by western blotting. Then, C3A cells were transfected with plasmid-expressing open reading frame 3 (ORF3) or control plasmids, the levels of IFN- $\alpha/\beta$  and ISG15 was evaluated, respectively. Next, the plasmid-expressing ISG15 or small interfering RNA-inhibiting ISG15 was transfected into infected C3A cells. The production of IFN- $\alpha/\beta$  was also measured by ELISA.

### Research results

In this study, we demonstrated that HEV infection could enhance the production of IFN- $\alpha/\beta$  and induce elevation of the ISG15 protein. ORF3 protein may be responsible for the enhancement of IFN- $\alpha/\beta$  and ISG15 by HEV. Furthermore, ISG15 silencing enhanced the production of IFN- $\alpha/\beta$  in C3A cells. Over-expression of ISG15 resulted in the reduction of IFN- $\alpha/\beta$  in C3A cells.

### Research conclusions

The findings of the present study showed that HEV could inhibit the level of type I IFN through regulating the expression of ISG15, a finding which may enhance the understanding of the interaction between ISG15 and HEV in the host innate immune response, and provide useful insight for the development of new antiviral drugs and antiviral strategies.

### Research perspectives

The future research will focus on whether HEV ORF3 could regulate the egress of HEV through ISG15.

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