

# World Journal of *Gastroenterology*

*World J Gastroenterol* 2018 July 7; 24(25): 2647-2784



### REVIEW

- 2647 Role of microRNAs in the main molecular pathways of hepatocellular carcinoma  
*Vasuri F, Visani M, Acquaviva G, Brand T, Fiorentino M, Pession A, Tallini G, D'Errico A, de Biase D*
- 2661 Apoptosis and non-alcoholic fatty liver diseases  
*Kanda T, Matsuoka S, Yamazaki M, Shibata T, Nirei K, Takahashi H, Kaneko T, Fujisawa M, Higuchi T, Nakamura H, Matsumoto N, Yamagami H, Ogawa M, Imazu H, Kuroda K, Moriyama M*
- 2673 Diets, functional foods, and nutraceuticals as alternative therapies for inflammatory bowel disease: Present status and future trends  
*Mijan MA, Lim BO*

### MINIREVIEWS

- 2686 Advances in immuno-oncology biomarkers for gastroesophageal cancer: Programmed death ligand 1, microsatellite instability, and beyond  
*Lin EM, Gong J, Klempner SJ, Chao J*
- 2698 Minimally invasive donor hepatectomy, are we ready for prime time?  
*Au KP, Chok KS*

### ORIGINAL ARTICLE

#### Basic Study

- 2710 Intra-individual comparison of therapeutic responses to vascular disrupting agent CA4P between rodent primary and secondary liver cancers  
*Liu YW, De Keyser F, Feng YB, Chen F, Song SL, Swinnen J, Bormans G, Oyen R, Huang G, Ni YC*

#### Retrospective Study

- 2722 Gastric cancer in Alaska Native people: A cancer health disparity  
*Martinson HA, Shelby NJ, Alberts SR, Olnes MJ*

#### Observational Study

- 2733 Transforming growth factor- $\beta$  and peripheral regulatory cells are negatively correlated with the overall survival of hepatocellular carcinoma  
*An Y, Gao S, Zhao WC, Qiu BA, Xia NX, Zhang PJ, Fan ZP*

### SYSTEMATIC REVIEWS

- 2741 Current global trends in the incidence of pediatric-onset inflammatory bowel disease  
*Sýkora J, Pomahačová R, Kreslová M, Cvalínová D, Štych P, Schwarz J*

### META-ANALYSIS

- 2764 Systematic review and meta-analysis on the association of tuberculosis in Crohn's disease patients treated with tumor necrosis factor- $\alpha$  inhibitors (Anti-TNF $\alpha$ )  
*Cao BL, Qasem A, Sharp RC, Abdelli LS, Naser SA*

### CASE REPORT

- 2776 Liposarcoma of the stomach: Report of two cases and review of the literature  
*Kang WZ, Xue LY, Wang GQ, Ma FH, Feng XL, Guo L, Li Y, Li WK, Tian YT*

**ABOUT COVER**

Editorial board member of *World Journal of Gastroenterology*, Dar-In Tai, MD, PhD, Attending Doctor, Chief Doctor, Professor, Department of Gastroenterology and Hepatology, Chang Gung Memorial Hospital, Taipei 105, Taiwan

**AIMS AND SCOPE**

*World Journal of Gastroenterology* (*World J Gastroenterol*, *WJG*, print ISSN 1007-9327, online ISSN 2219-2840, DOI: 10.3748) is a peer-reviewed open access journal. *WJG* was established on October 1, 1995. It is published weekly on the 7<sup>th</sup>, 14<sup>th</sup>, 21<sup>st</sup>, and 28<sup>th</sup> each month. The *WJG* Editorial Board consists of 642 experts in gastroenterology and hepatology from 59 countries.

The primary task of *WJG* is to rapidly publish high-quality original articles, reviews, and commentaries in the fields of gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, hepatobiliary surgery, gastrointestinal oncology, gastrointestinal radiation oncology, gastrointestinal imaging, gastrointestinal interventional therapy, gastrointestinal infectious diseases, gastrointestinal pharmacology, gastrointestinal pathophysiology, gastrointestinal pathology, evidence-based medicine in gastroenterology, pancreatology, gastrointestinal laboratory medicine, gastrointestinal molecular biology, gastrointestinal immunology, gastrointestinal microbiology, gastrointestinal genetics, gastrointestinal translational medicine, gastrointestinal diagnostics, and gastrointestinal therapeutics. *WJG* is dedicated to become an influential and prestigious journal in gastroenterology and hepatology, to promote the development of above disciplines, and to improve the diagnostic and therapeutic skill and expertise of clinicians.

**INDEXING/ABSTRACTING**

*World Journal of Gastroenterology* (*WJG*) is now indexed in Current Contents<sup>®</sup>/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch<sup>®</sup>), Journal Citation Reports<sup>®</sup>, Index Medicus, MEDLINE, PubMed, PubMed Central and Directory of Open Access Journals. The 2018 edition of Journal Citation Reports<sup>®</sup> cites the 2017 impact factor for *WJG* as 3.300 (5-year impact factor: 3.387), ranking *WJG* as 35<sup>th</sup> among 80 journals in gastroenterology and hepatology (quartile in category Q2).

**EDITORS FOR THIS ISSUE**

**Responsible Assistant Editor:** *Xiang Li*  
**Responsible Electronic Editor:** *Yan Huang*  
**Proofing Editor-in-Chief:** *Lian-Sheng Ma*

**Responsible Science Editor:** *Xue-Jiao Wang*  
**Proofing Editorial Office Director:** *Ze-Mao Gong*

**NAME OF JOURNAL**  
*World Journal of Gastroenterology*

**ISSN**  
 ISSN 1007-9327 (print)  
 ISSN 2219-2840 (online)

**LAUNCH DATE**  
 October 1, 1995

**FREQUENCY**  
 Weekly

**EDITORS-IN-CHIEF**  
**Damian Garcia-Olmo, MD, PhD, Doctor, Professor, Surgeon**, Department of Surgery, Universidad Autonoma de Madrid; Department of General Surgery, Fundacion Jimenez Diaz University Hospital, Madrid 28040, Spain

**Stephen C Strom, PhD, Professor**, Department of Laboratory Medicine, Division of Pathology, Karolinska Institutet, Stockholm 141-86, Sweden

**Andrzej S Tarnawski, MD, PhD, DSc (Med), Professor of Medicine, Chief Gastroenterology**, VA Long Beach Health Care System, University of California, Irvine, CA, 5901 E. Seventh Str., Long Beach,

CA 90822, United States

**EDITORIAL BOARD MEMBERS**  
 All editorial board members resources online at <http://www.wjgnet.com/1007-9327/editorialboard.htm>

**EDITORIAL OFFICE**  
 Ze-Mao Gong, Director  
*World Journal of Gastroenterology*  
 Baishideng Publishing Group Inc  
 7901 Stoneridge Drive, Suite 501,  
 Pleasanton, CA 94588, USA  
 Telephone: +1-925-2238242  
 Fax: +1-925-2238243  
 E-mail: [editorialoffice@wjgnet.com](mailto:editorialoffice@wjgnet.com)  
 Help Desk: <http://www.f6publishing.com/helpdesk>  
<http://www.wjgnet.com>

**PUBLISHER**  
 Baishideng Publishing Group Inc  
 7901 Stoneridge Drive, Suite 501,  
 Pleasanton, CA 94588, USA  
 Telephone: +1-925-2238242  
 Fax: +1-925-2238243  
 E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
 Help Desk: <http://www.f6publishing.com/helpdesk>  
<http://www.wjgnet.com>

**PUBLICATION DATE**  
 July 7, 2018

**COPYRIGHT**  
 © 2018 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

**SPECIAL STATEMENT**  
 All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

**INSTRUCTIONS TO AUTHORS**  
 Full instructions are available online at <http://www.wjgnet.com/bpg/gerinfo/204>

**ONLINE SUBMISSION**  
<http://www.f6publishing.com>

## Diets, functional foods, and nutraceuticals as alternative therapies for inflammatory bowel disease: Present status and future trends

Mohammad Al Mijan, Beong Ou Lim

Mohammad Al Mijan, Beong Ou Lim, Department of Integrated Biosciences, College of Biomedical & Health Science, Konkuk University, Chungju 380-701, South Korea

ORCID number: Mohammad Al Mijan (0000-0003-3530-4699); Beong Ou Lim (0000-0002-9618-5956).

Author contributions: Mijan MA prepared the manuscript; Lim BO provided overall guidance and supervision in writing the article.

Conflict-of-interest statement: The authors declare no conflict-of-interests.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: Beong Ou Lim, PhD, Professor, Department of Integrated Biosciences, College of Biomedical & Health Science, Konkuk University, 322 Danwol-dong, Chungju-shi, Chungbuk-do 380-701, South Korea. [beongou@kku.ac.kr](mailto:beongou@kku.ac.kr)

Telephone: +82-43- 8403570

Fax: +82-43- 8563572

Received: April 17, 2018

Peer-review started: April 19, 2018

First decision: May 9, 2018

Revised: May 19, 2018

Accepted: June 9, 2018

Article in press: June 9, 2018

Published online: July 7, 2018

### Abstract

Inflammatory bowel disease (IBD) is a serious health concern among western societies. The disease is also on the rise in some East Asian countries and in Australia. Health professionals and dietitians around the world are facing an unprecedented challenge to prevent and control the increasing prevalence of IBD. The current therapeutic strategy that includes drugs and biological treatments is inefficient and are associated with adverse health consequences. In this context, the use of natural products is gaining worldwide attention. *In vivo* studies and clinical evidence suggest that well-planned dietary regimens with specific nutrients can alleviate gastrointestinal inflammation by modulating inflammatory cytokines, such as tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ), interleukin 1 (IL-1), IL-6, IL-1 $\beta$ , and IL-10. Alternatively, the avoidance of high-fat and high-carbohydrate diets is regarded as an effective tool to eliminate the causes of IBD. Many functional foods and bioactive components have received attention for showing strong therapeutic effects against IBD. Both animal and human studies suggest that bioactive functional foods can ameliorate IBD by downregulating the pro-inflammatory signaling pathways, such as nuclear factor  $\kappa$ B, STAT1, STAT6, and pro-inflammatory cytokines, including IL-1 $\beta$ , IL-4, IL-6, COX-2, TNF- $\alpha$ , and interferon  $\gamma$ . Therefore, functional foods and diets have the potential to alleviate IBD by modulating the underlying pathogenic mechanisms. Future comprehensive studies are needed to corroborate the potential roles of functional foods and diets in the prevention and control of IBD.

**Key words:** Inflammatory bowel disease; Colitis; Diets; Functional foods; Bioactive compounds; Inflammatory cytokines; Alternative therapy

© The Author(s) 2018. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** Diets and functional foods are two of the most potential alternative therapies for inflammatory bowel disease (IBD). Dietary supplementation of probiotics and non-starch polysaccharides demonstrated strong therapeutic actions on IBD. Likewise, functional foods have received more attention than ever as alternative therapies for IBD. Plant-derived extracts and bioactive compounds exhibited anti-inflammatory actions against IBD. Both diets and functional foods have a very important role to play in the near future. We have discussed the roles of both diets and functional foods in IBD management.

Mijan MA, Lim BO. Diets, functional foods, and nutraceuticals as alternative therapies for inflammatory bowel disease: Present status and future trends. *World J Gastroenterol* 2018; 24(25): 2673-2685 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v24/i25/2673.htm> DOI: <http://dx.doi.org/10.3748/wjg.v24.i25.2673>

## INTRODUCTION

Inflammatory bowel disease (IBD) is a chronic gastrointestinal disorder characterized by relapsing inflammation and severe mucosal damage in the intestine. There are two common forms of IBD, namely, ulcerative colitis (UC) and Crohn's disease (CD), which are generally associated with diarrhea, nausea, abdominal pain, fatigue, rectal bleeding, weight loss, anxiety, *etc*<sup>[1]</sup>. Currently, IBD is one of the most prevalent gastrointestinal diseases among the developed nations in the West, affecting nearly 1.6 million people in the United States and 2.5-3.0 million people in northern Europe<sup>[2,3]</sup>. Although IBD is mostly prevalent in North America and Europe, the adoption of western dietary habits and lifestyle has led to, countries like- China, South Korea, and Australia witnessing a significant rise in the incidence of IBD<sup>[4]</sup>.

The exact etiology of IBD has yet to be defined, but it is believed that genetic susceptibility, environment, immunoregulatory dysfunction, intestinal microbiota, nutrition, and lifestyle are the key players in the pathogenesis of IBD<sup>[5]</sup>. Activation of macrophages and an uncontrolled production of pro- and anti-inflammatory cytokines, including tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ), interleukins, and interferon  $\gamma$  (IFN- $\gamma$ ) in the intestinal mucosa, mediate the inflammation by inducing inflammatory pathways<sup>[6]</sup>. Conventional therapies based on steroidal and non-steroidal drugs and biological agents are inefficient in treating IBD and are often associated with adverse side effects. As a consequence, tremendous research attention is now being focused on finding alternative therapies based on plants and other natural products.

There is a growing consensus that diet and nutrition play a critical role in the etiopathogenesis of IBD, and

hence dietary therapy has a great implication on the treatment of IBD<sup>[7]</sup>. Recent research evidence suggests that the supplementation of fruits and vegetables, probiotic bacteria, dietary fibers, and fat-soluble vitamins can substantially reduce the symptoms of IBD through their anti-inflammatory functions<sup>[7-11]</sup>. In contrast, as the high-fat and high-carbohydrate foods are supposedly involved in the etiology of IBD, eliminating these foods from the diet could be an essential tool in the management of IBD<sup>[12]</sup>. Bioactive natural compounds and functional foods have been a major focus of research throughout the last decade as potential therapies for IBD, and many research groups have demonstrated positive and outstanding results. Plant-derived extracts, antioxidants, phytochemicals, polyunsaturated fatty acids, and dietary peptides have demonstrated strong anti-inflammatory effects against IBD due to their modulatory actions on pro- and anti-inflammatory cytokines and signaling pathways<sup>[13-18]</sup>. Ongoing and future research is expected to provide more evidence and explanations regarding the use of diets and functional foods to control IBD. It appears that the alternative therapies based on diets and functional foods will be the future of IBD management. The present study therefore provides an overview on the current status and the future direction of the use of diets, functional foods, and bioactive compounds against IBD.

## CURRENT STATUS AND TREATMENTS OF IBD

The chronic and recurrent inflammation of the gastrointestinal tract associated with IBD, represented by UC and CD accompanies several gastrointestinal and systemic disorders and mental illnesses<sup>[19]</sup>. Although the two forms of IBD share some common features, they are regarded as separate entities, as they possess distinct histopathological and symptomatic characteristics. UC is generally defined as a mucosal or submucosal inflammation of mainly the rectum and occasionally of the colonic area. The common symptoms of UC include abdominal pain, diarrhea, malnutrition, rectal pain and bleeding. CD is regarded as a transmural inflammation of the ileum and colon, though it can affect any part of the gastrointestinal tract and form granulomas, fistulas, and strictures in the intestine. Patients diagnosed with CD often have abdominal pain, diarrhea, fever, loss of appetite and weight, anemia, and intermittent anal fissures.

Although the exact etiology of IBD has yet to be specified, a number of factors including- diet, immunity, environment, heredity, and microbiota, contribute to the development of IBD<sup>[20]</sup>. A complex interaction of environmental, genetic, microbial, and immunological factors might cause the activation of the mucosal immune response and the release of numerous cytokines<sup>[21]</sup>. Cytokines are cell signaling molecules generated predominantly by immune cells that have specific roles in the communication and interaction between cells and

**Table 1** Overview of the conventional therapies for inflammatory bowel disease

Therapeutic agent	Active compound	Mode of action	Ref.
Aminosalicylates (ASA)	5-ASA	Decreases MPO activity, inhibits $\beta$ -catenin activation Inhibits the generation and activity of IL-1 $\beta$ , IL-4, IL-5, IL-8, granulocyte-macrophage colony stimulating factor, and TNF- $\alpha$	[23]
Corticosteroids	Corticosteroids		[24]
Immunosuppressants	Azathioprine 6-mercaptopurine Cyclosporine A Tacrolimus Methotrexaten	Clinical remission Mucosal healing	[25]
Antibiotics	Metronidazole Ciprofloxacin	Decrease disease activity index Maintain remission	[26]
Biological therapy	Infliximab Adalimumab Certolizumab	Neutralizes TNF- $\alpha$ Reduces inflammation	[27]

TNF- $\alpha$ : Tumor necrosis factor  $\alpha$ ; IL: Interleukin.

the onset of local and systemic inflammation. Under normal conditions, the intestinal mucosa can maintain the balance between pro-inflammatory cytokines, such as TNF- $\alpha$ , IFN- $\gamma$ , interleukin (IL)-1, IL-6, and IL-12 and anti-inflammatory cytokines, which includes IL-4, IL-10, and IL-11. In IBD patients, the intestinal homeostasis and the fine balance between pro- and anti-inflammatory cytokines is disrupted, causing an increased number and activities of pro-inflammatory cytokines in the mucosa, leading to tissue damage and inflammation. Furthermore, the weakened epithelial barrier function and the increased intestinal permeability in IBD subjects facilitate mucosal inflammation<sup>[22]</sup>.

Currently, there is no effective therapy available that can completely cure IBD. Current therapeutic options are incapable of targeting the underlying pathogenic mechanisms of IBD; instead, they are specifically designed to instigate and maintain the remission of the disease and help mitigate complications in patients<sup>[1]</sup>. Aminosalicylates and corticosteroids are considered first-line therapy for IBD (Table 1). Both of these drugs have shown efficacies in ameliorating the severity and the symptoms of IBD through their abilities to down-regulate the pro-inflammatory cytokines and signaling pathways<sup>[23,24]</sup>. Immunosuppressive agents, including azathioprine, 6-mercaptopurine, cyclosporine A, and antibiotics, which are mostly used as adjunct therapies, can decrease intestinal inflammation by suppressing the mucosal immune response<sup>[25,26]</sup> (Table 1). A more recent and innovative approach is called "biological therapy," where monoclonal antibodies, such as infliximab and adalimumab, are applied to downregulate the immune response pathways<sup>[27]</sup>.

Despite providing some symptomatic and temporary relief, current drug therapies are described as inadequate with serious side effects<sup>[28]</sup>. Biological therapies, which are currently a mainstay for of IBD treatment, are expensive and associated with adverse health effects.

Therefore, the development of alternative IBD therapies using natural products that are highly effective, safe, and inexpensive is in great demand.

## DIETS AND DIETARY INTERVENTIONS FOR IBD

Diets comprise the usual food and drink that a person regularly consume. Diets, among other factors, play a crucial role in the etiology of IBD. Dietary interventions in the form of either providing specific nutrients or dietary restrictions are regarded as effective tools in treating IBD. Currently, due to the lack of adequate data and research evidence, health professionals and dieticians often find it difficult to recommend dietary strategies for IBD patients. However, recent research outcomes are providing evidence that many nutrients and food elements can cure IBD symptoms; hence a dietary plan based on proper nutrients could be an effective therapeutic strategy against IBD.

### Probiotics

Probiotics are described as live microorganisms that benefit humans by promoting gut health and the immune system upon ingestion in an acceptable amount. Numerous possible mechanisms through which probiotic bacteria exert their beneficial effects have been proposed. Probiotics can reduce harmful microorganisms and maintain the microbial balance inside the gut by blocking the site of adhesion, competing for nutrients, and killing pathogenic microorganisms<sup>[29]</sup>. Production of short-chain fatty acids (SCFA) and butyrate by probiotic bacteria lowers the pH level in the colon and limits the growth of pathogens<sup>[30]</sup>. In addition, probiotic bacteria can function as anti-inflammatory agents by modulating the NF- $\kappa$ B signaling pathway, inflammatory cytokines, and the regulatory T cell response<sup>[31]</sup>. Two of

the most widely studied genera that have been proven effective in alleviating gastrointestinal inflammation are *Lactobacillus* and *Bifidobacteria*. Lee *et al.*<sup>[32]</sup> reported that *Lactobacillus suntoryeus* suppressed toll-like receptor (TLR)-4 linked NF- $\kappa$ B and IL-6 expression in TNBS-induced colitis (Table 2). In a mouse model of IBD induced by *E. coli* 0111 LPS, soy milk fermented with *Lactococcus lactis* subsp. *lactis* S-SU2 prevented colonic shortening and spleen enlargement, and repaired epithelial damage<sup>[33]</sup>. *Lactobacillus paracasei* LS2 isolated from kimchi decreased the number of neutrophils (CD11b<sup>+</sup>Gr-1<sup>+</sup>) and, macrophages (CD11b<sup>+</sup> F4/80<sup>+</sup>), and decreased TNF- $\alpha$  and IFN- $\gamma$  expression in DSS-induced UC<sup>[34]</sup>. An oral administration of *Lactococcus lactis* NZ9000 (NZ-HO) secreting an anti-inflammatory substance called recombinant mouse heme oxygenase (mHO-1) to mice decreased the disease activity index (DAI), increased the production of IL-10, and suppressed IL-1 $\alpha$  and IL-6 expression<sup>[35]</sup>. A study by Yokota *et al.*<sup>[36]</sup> revealed that supplying drinking water containing *Lactobacillus plantarum* AN1 isolated from fermented fish to an IBD mouse model increased the indigenous population of lactic acid bacteria in the colon, and their synergistic effects reversed colonic shortening, spleen enlargement, and colonic tissue damage significantly. Several other strains of *Lactobacillus plantarum* exhibited therapeutic effects on gastrointestinal inflammation through their modulatory functions against inflammatory cytokines<sup>[37]</sup>. A recent study indicated that *Lactobacillus sakei* attenuated the clinical symptoms and histological damage by suppressing inflammatory mediators, such as NF- $\kappa$ B, STAT1, and TL4<sup>[38]</sup>. A combined therapy consisting of *Lactobacillus casei*, butyrate, and *Pistacia atlantica* significantly improved histological scores and reduced MPO activity in a rat model of IBD<sup>[39]</sup>.

The antimicrobial and anti-inflammatory effects of *Bifidobacteria* are also well-known, and this probiotic genus has a wide application against gastrointestinal inflammation. Reportedly, *Bifidobacterium adolescentis* IM38 alleviated inflammation by downregulating NF- $\kappa$ B expression and lipopolysaccharide production in high-fat diet-induced ulcerative colitis in mice<sup>[40]</sup>. An *in vitro* and *in vivo* study suggested that *Bifidobacteria bifidum* 231 enhanced the IL-10 production in IEC-6 cell lines and improved the macroscopic and histological conditions in TNBS-induced colitis<sup>[41]</sup>. *Bifidobacterium longum* CCM7952 strengthened the epithelial barrier function and reduced clinical symptoms in experimental colitis<sup>[31]</sup>.

### Non-starch polysaccharides

Non-starch polysaccharides (NPS), classified as dietary fiber and prebiotics, are obtained from various natural sources that have been studied extensively as therapeutics against inflammation and other immune-related problems. All of the major components of NPS, including cellulose, glucomannan, glucan, pectin, inulin, and oligosaccharides have exhibited anti-inflammatory and immunomodulatory functions<sup>[42]</sup>. It has been

suggested that most of the NPS components reach the large intestine intact, where they are fermented by probiotic and useful bacteria to exert their anti-inflammatory functions<sup>[43]</sup>.

Konjac glucomannan is a plant-derived polysaccharide that has been used to treat gastrointestinal inflammatory disorders. For example, supplementation with konjac glucomannan hydrolysate for fourteen days to IBD patients resulted in improved bowel movement, fecal consistency, reduced abdominal pain, and a better lifestyle<sup>[44]</sup> (Table 2).  $\beta$ -Glucan was orally administered to an animal model of IBD, which resulted in improved fecal output and reduced colorectal distension<sup>[45]</sup>. In another study, oat  $\beta$ -glucan reduced the levels of MPO, NO, and MDA, and suppressed the expression of IL-1 $\beta$ , IL-6, and iNOS in a DSS-induced colitis model in mice<sup>[46]</sup>. Bacterial  $\beta$ -(1,3)-glucan prevented IBD in mice by recovering regulatory T cells (Tregs) and the defects of natural killer (NK) cells and by suppressing the excessive production of IgA<sup>[47,48]</sup>. Azuma *et al.*<sup>[49,50]</sup> reported that cellulose nanofibers obtained from seaweed and pear reversed colonic shortening, reduced colonic damage and, suppressed NF- $\kappa$ B expression and MPO activity in colitic mice.

Prebiotics are non-digestible polysaccharides, which generally includes oligosaccharides and inulin that act as nutrients for the native gut microbiota that offer health benefits to the host<sup>[30]</sup>. Providing fructooligosaccharides to colitic mice resulted in increased lactic acid bacteria in the gut and decreased pro-inflammatory cytokines such as, IFN- $\gamma$ , IL-17, and TNF- $\alpha$ <sup>[51]</sup>. In a DSS-induced colitis mouse model, goat milk oligosaccharides reduced the colonic tissue damage and increased the favorable microbial population in the intestine<sup>[52]</sup>. Štofilová *et al.*<sup>[53]</sup> demonstrated that prebiotic inulin together with *Lactobacillus plantarum* LS/07 CCM7766 symbiotically improved colonic and jejunal tissue damage by down-regulating IL-2, IL-6, IL-17, TNF- $\alpha$ , COX-2, and NF- $\kappa$ B expression in an N, N-dimethylhydrazin-induced colitis model in rats.

### Vitamins

Fat soluble vitamins such as Vitamin A and D have protective roles against the pathogenesis of IBD. Accumulating evidence suggests that patients with IBD are frequently diagnosed with low levels of fat soluble vitamins, and therefore specific supplementations of those vitamins are often recommended<sup>[5]</sup>. Gubatan *et al.*<sup>[54]</sup> recently reported that a low level of vitamin D in the patients with UC at the time of remission increased the risk of clinical relapse of IBD. Vitamin D deficiency in UC patients has been found to be associated with mucosal inflammation and disease activity<sup>[55]</sup>. Therefore, vitamin D supplementation can result in positive outcomes in IBD patients. It has been demonstrated that vitamin D, by downregulating pro-inflammatory cytokines, IL-6, IL-21, TNF- $\alpha$ , and IFN- $\gamma$  and by stabilizing the intestinal barrier, can contribute to the amelioration of IBD symptoms<sup>[56]</sup>. According to Zhu

**Table 2** Role of nutrients and diets against inflammatory bowel disease

Base material	Main compounds/agents	Mode of action	Ref.
Probiotics			
Lactic acid bacteria	<i>Lactobacillus suntoryeus</i>	Inhibited the activation of TLR-4-linked NF-κB activation	[32]
	<i>Lactococcus lactis</i> subsp. <i>lactis</i> S-SU2	Prevented the colonic shortening, lowering of liver and thymus weights, and spleen enlargement	[33]
	<i>Lactobacillus paracasei</i> LS2 (from kimchi)	Increased IL-10	[34]
	<i>Lactococcus lactis</i> NZ9000 (NZ-HO)	Reduced TNF-α, IFN-γ, IL-1β and MPO activity	[35]
	<i>Lactobacillus plantarum</i> AN1	Reduced CD11b+ F4/80+ and CD11b+ Gr-1+	[36]
	<i>Lactobacillus sakei</i> K040706	Increased IL-10; reduced IL-1α and IL-6	[38]
Bifidobacteria	<i>Bifidobacterium bifidum</i> 231	Ameliorated the atrophy of colon length, mucosal damage, and spleen enlargement	[41]
	<i>Bifidobacterium longum</i> CCM7952	Reduced the expression of iNOS, TNF-α, IL-1β, and IL-6	[31]
		Suppressed NF-κB, STAT3, and TLR4 expression	
		Increased IL-10; Decreased IL-1β	
		Engaged TLR2; Contained NOD2	
		Improved epithelial barrier	
Dietary fibers and prebiotics			
Konjac glucomannan	Konjac glucomannan hydrolysate	Reduced bowel movement, diarrhea, blood in feces, abdominal pain, and flatulence	[44]
Glucan	β-(1,3-1,6)-d-glucan	Improved fecal output	[45]
	Oat β-glucan	Reduced visceral pain	[46]
		Lowered MPO, NO, and MDA	
		Inhibited the expressions of TNF-α, IL-1β, IL-6 and iNOS	
	Glucan from mushroom ( <i>Pleurotus pulmonarius</i> )	Reduced histological damage	[47]
	Bacterial β-(1,3)-glucan	Reduced the expression of IL-1β	[48]
		Reversed Treg reduction	
Nanofiber	Cellulose nanofiber from seaweed	Decreased NK cell defects and IgA production	[49]
Prebiotics	Cellulose nanofiber from pear	Improved intestinal tissue injury	[50]
	Fructooligosaccharides	Suppressed the activation of NF-κB	[51]
	Goat milk oligosaccharide	Suppressed colon atrophy	[52]
	Inulin	Suppressed the activation of NF-κB	[53]
		Decreased IFN-γ, IL-17, and TNF-α levels	
		Increased LAB population	
		Decreased inflammation	
		Improved mucosal damage	
		Decreased TNFα, COX-2, IL-2, and IL-6	
Vitamins	1α,25-dihydroxyvitamin D3	Suppressed TNF-α	[57]
		Enhanced IL-10 production	
	1,25-dihydroxyvitamin D3	Reduced IFN-γ	[58]
	Vitamin D3	Increased CD4+ T cells and IL-6	[59]
		Protected mitochondria	
	Vitamin A	Inhibited nuclear respiratory factor (NFR)-1 and mitochondrial transcription factor A (TFAM)	[60]

TLR: Toll-like receptor; TNF-α: Tumor necrosis factor α; IFN-γ: Interferon γ; IL: Interleukin; NF-κB: Nuclear factor κB.

*et al.*<sup>[57]</sup> (2005), 1α,25-dihydroxyvitamin D3 together with calcium substantially reduced TNF-α expression and relieved the symptoms of IBD in IL-10 knockout mice. In human CD4+ cells, 1,25-dihydroxyvitamin D3 increased the level of IL-10 and inhibited the proliferation of T cells<sup>[58]</sup>. Patients suffering from Crohn's disease have shown an increased level of IL-6 after vitamin D3 treatment<sup>[59]</sup>. The protective effects of vitamin A have been investigated in a TNBS-induced colitis mouse

model, and after 21 days of treatment, vitamin A substantially increased the proliferation of mitochondrial transcription factors NFR-1 and TFAM and prevented intestinal tissue damage<sup>[60]</sup>.

### Specific carbohydrate diets and FODMAP

The specific carbohydrate diet (SCD) was first designed and mentioned by Sydney Haas in 1914 to cure celiac disease<sup>[20]</sup>. The term became popularized when Elaine

Gottschall published his book entitled "Breaking the Vicious Cycle: intestinal health through diet" in 2012 about how he and his daughter were cured from IBD by strictly following this diet plan<sup>[61]</sup>. The SCD plan allows the intake of only monosaccharides and the complete avoidance of disaccharides and polysaccharides, because they remain undigested and unabsorbed in the digestive tract and lead to the overproduction of yeast and bacteria, which eventually causes intestinal injury. In a clinical study, children with Crohn's disease under this dietary plan for 12 and 52 wk had remarkably reduced mucosal damage and improved clinical symptoms<sup>[62]</sup>. FODMAPs are specific carbohydrate foods that contain mono-, di-, and oligosaccharides and polyols<sup>[63]</sup>. These carbohydrates are poorly digested, but easily fermented by the colonic bacteria, leading to bloating, abdominal cramping and discomfort, and diarrhea, which are also associated with IBD<sup>[64]</sup>. As a consequence, scientists developed the idea of eliminating FODMAPs from the diet for IBD patients as a cure. The efficacies of FODMAP diets have been demonstrated through human studies, and patients who adhered to FODMAP elimination have experienced fewer abdominal disorders and better quality of life<sup>[65,66]</sup>.

## FUNCTIONAL FOODS AND NUTRACEUTICALS FOR IBD

Functional foods are any fresh or processed foods that provide health benefits and have disease prevention activities beyond their basic nutritional value. Nutraceuticals are foods or food supplements that deliver concentrated form of bioactive substances with medicinal properties. Although functional foods have been used as traditional medicines to treat chronic diseases for several centuries, it is modern scientific discoveries that are establishing the health benefits of functional foods and natural bioactive compounds providing the underlying mechanisms of their actions. Potential roles of functional foods against IBD have been broadly studied over the last decade, and overwhelming research evidence suggests that plant extracts, polyphenols, fatty acids, and amino acids can attenuate IBD symptoms by interfering with inflammatory pathways<sup>[6,67]</sup> (Table 3).

### Plant and fruit extracts

The history of using plant extracts as alternative therapies for boosting the immune system and treating chronic inflammatory disorders dates back to ancient times. The anti-inflammatory activities of the plant extracts derive mainly from their abilities to modulate inflammatory cytokines. Several plant-derived extracts have strong therapeutic effects against IBD, and there is a growing interest in developing an effective IBD therapy based on plant extracts. *Coriolus versicolor*, mostly grown in China is a medicinal mushroom that has well-known health benefits. This mushroom contains polysaccharides,

including krestin, lignin, and glucan. Our investigation on the effect of *Coriolus versicolor* extract (CVE) on UC in mice demonstrated that CVE could relieve the symptoms of colitis by decreasing the level of IgE in the serum and lymph nodes and, by suppressing the expression of TNF- $\alpha$ , IFN- $\gamma$ , IL-4, IL-1 $\beta$  and IL-6<sup>[68]</sup>. *Cordyceps militaris*, a folk medicinal mushroom found in East Asia, also has proven health benefits against inflammation, most likely due to its polysaccharide contents<sup>[69]</sup>. A study by Han *et al.*<sup>[70]</sup> reported that *Cordyceps militaris* prevented epithelial damage, inflammatory cell migration, and colonic shortening by decreasing TNF- $\alpha$  and iNOS levels. Interestingly, recent studies claim that the mushrooms grown on germinated cereal grains are rich in antioxidants and other bioactives and possess potent antioxidative and anti-inflammatory functions<sup>[71,72]</sup>. The chaga mushroom grown on germinated brown rice suppressed the expression of COX-2, TNF- $\alpha$ , IL-4, STAT1 and STAT6, and reduced the levels of IgE and IgA<sup>[73]</sup>. Additionally, *Ganoderma lucidum* grown on germinated rice reduced the inflammation in colitic mice by downregulating NF- $\kappa$ B and MAPK pathways<sup>[74]</sup>.

Fruit extracts, due to their potential nutraceutical properties, have been investigated as therapeutic agents to treat IBD. A *Prunus mume* mixture in a DSS-induced colitis mouse model ameliorated inflammation by decreasing inflammatory cytokines and the immune response<sup>[75]</sup>. Pomegranate (*Punica granatum*), a tropical fruit rich in polyphenols especially ellagitannins and ellagic acid, has antioxidant and anti-inflammatory properties<sup>[76]</sup>. Pomegranate extracts, in TNBS-induced colitis rats, decreased TNF- $\alpha$ , MAPK phosphorylation, and NF- $\kappa$ B translocation<sup>[77]</sup>. Berry fruits are known for being rich in bioactive compounds and their therapeutic actions against numerous health problems<sup>[78]</sup>. The oral administration of blueberry extracts to colitic mice alleviated inflammation by a three-fold mechanism: antioxidation, inhibition of NF- $\kappa$ B translocation, and suppression of inflammatory cytokines<sup>[79]</sup>. The application of *Aronia melanocarpa* Elliot, also known as black chokeberry, relieved colitis symptoms in mice through its antioxidative and anti-inflammatory activities<sup>[80,81]</sup>. Ginger, which has been traditionally used as a spice and a natural remedy, has shown anti-inflammatory properties. Ginger extract, when administered to colitic mice, ameliorated colonic inflammation by downregulating NF- $\kappa$ B and IL-1 $\beta$  expression<sup>[82]</sup>.

Marine foods and extracts have lately received attention as bioactive substances for IBD. The ethanol extract from *Halictis discus hannai* Ino remarkably decreased mucosal tissue damage and lowered the expressions of IL-4, IFN- $\gamma$ , STAT1, and STAT6 in a mouse model of colitis<sup>[83]</sup>. Green algae extract also exhibits strong remedial effects against DSS-induced colitis<sup>[84]</sup>.

### Phytochemicals

Phytochemicals perform important bioactive functions against oxidative and inflammatory disorders. Plant-

**Table 3** Role of natural extracts and phytochemicals against inflammatory bowel disease

Base material	Main compound/agent	Mode of action	Ref.
Extracts Mushroom	<i>Coriolus versicolor</i> extract	Reduced TNF- $\alpha$ , IL-1 $\beta$ and IL-6 Reduced STAT1 and STAT6	[68]
	<i>Cordiceps militaris</i> extract	Decreased epithelial damage Suppressed iNOS and TNF- $\alpha$ mRNA expression	[70]
	<i>Inonotus obliquus</i> extract	Suppressed TNF- $\alpha$ , COX-2, and IFN- $\gamma$	[73]
	<i>Ganoderma lucidum</i> extract	Inhibited MAPK phosphorylation and NF- $\kappa$ B activation Decreased histological score	[74]
Fruit extracts <i>Prunus mume</i>	<i>Prunus mume</i> extract	Suppressed mucosal damage, TNF- $\alpha$ , and iNOS expressions	[75]
	Pomegranate extract (ellagitannins and ellagic acid)	Decreased the expression of TNF- $\alpha$ , COX-2, IL-4, and STAT6	[77]
Pomegranate	Cranberry fruit/extract Blueberry extract	Prevented the translocation of NF- $\kappa$ B	[78]
		Modulated NF- $\kappa$ B and IL-1 $\beta$ signaling Attenuated colon shortening	[79]
Cranberry	<i>Averrhoa bilimbi</i>	Suppressed pro-inflammatory cytokines	[80]
		Prevented oxidation Inhibited pro-inflammatory mediators	[81]
<i>Aronia melanocarpa</i>	<i>Aronia melanocarpa</i> juice	Reduced NF- $\kappa$ B translocation	[82]
Ginger	Ginger extract (zingerone)	Decreased mucosal injury	[82]
Marine food	<i>Haliotis discus hannai</i> Ino extract	Decrease the level of pro-inflammatory cytokines	[83]
		Improved colonic damage Decreased TBARS concentration Suppressed NF- $\kappa$ B and IL-1 $\beta$	[84]
	Green algae extract	Suppressed colonic tissue damage Downregulated IFN- $\gamma$ and IL-4 Ameliorated colonic tissue damage	
	Decreased pro-inflammatory cytokines		
Phytochemicals	Apple polyphenols	Reduced COX-2 and TNF- $\alpha$ Recovered transglutaminase protein	[85]
	Resveratrol	Suppressed NF- $\kappa$ B and TNF- $\alpha$ Reduced clinical score	[86]
	Cardamonin	Reduced histopathological damage Reduced iNOS, NF- $\kappa$ B, TNF- $\alpha$ , COX-2, and caspase-3	[18]
	Ginsenoside Rg1	Suppressed IL-1 $\beta$ and TNF- $\alpha$ Reduced colonic damage and DAI Improved colon shortening and DAI	[90]
	Sulforaphane	Suppressed STAT3 expression	[91]
	Curcumin	Reduced TNF- $\alpha$ , IL-1 $\beta$ , and MPO Attenuated morphological damage	[92]

TLR: Toll-like receptor; TNF- $\alpha$ : Tumor necrosis factor  $\alpha$ ; IFN- $\gamma$ : Interferon  $\gamma$ ; IL: Interleukin; NF- $\kappa$ B: Nuclear factor  $\kappa$ B.

derived bioactive compounds can repress inflammation by inhibiting oxidative damage and interacting with the immune system. In a previous study, apple polyphenol extract reduced mucosal inflammation by reversing transglutaminase depletion in a TNBS-induced colitis rat model<sup>[85]</sup>. Resveratrol is an important polyphenol found abundantly in peanut, berries, and red grapes. This polyphenol exhibits versatile biological functions that are generally attributed to its modulating actions against oxidative processes and inflammatory pathways<sup>[86]</sup>. A randomized controlled trial conducted by Samsami-Kor *et al.*<sup>[87]</sup> revealed that patients with UC supplemented with resveratrol had lower inflammation and decreased levels of TNF- $\alpha$  and NF- $\kappa$ B compared with the placebo group. A component from Chinese traditional medicine

called cardamonin was administered to rats with acetic acid-induced colitis, and at the end of the trial, the rats had a reduced DAI score and improved histopathological conditions<sup>[18]</sup>. Cardamonin supplementation also reduced MDA and MPO activities, and NF- $\kappa$ B, TNF- $\alpha$ , and COX-2 expression. Previous reports suggest that NLRP12, a NOD-like receptor, can attenuate colonic inflammation by downregulating inflammatory cytokines and promoting the growth of useful bacteria in the gut<sup>[88,89]</sup>. Zhu *et al.*<sup>[90]</sup> reported that Ginsenoside Rg1, a red ginseng compound, inhibited the inflammatory response and colonic damage by upregulating NLRP12 in mice with UC. A broccoli-derived isothiocyanate compound sulforaphane, exhibited anti-colitic activities by preventing colonic atrophy and increasing the expression of the Nrf2-dependent gene in

mice<sup>[91]</sup>. Curcumin, which is isolated from turmeric, has medicinal application in some Eastern Asian countries, and it is one of the most studied phytochemicals against ulcerative colitis. A study conducted on colitic mice found that curcumin supplementation could improve the histopathological score in the colon by suppressing the activity and the DNA-binding ability of STAT3, and by reducing TNF- $\alpha$  and IL-1 $\beta$  expression<sup>[92]</sup>. A combined therapy with curcumin, green tea polyphenol, and selenium exhibited outstanding results with decreased inflammatory symptoms and DAI both in human subjects with colitis and in DSS- and TNBS-induced colitic mice<sup>[93]</sup>.

### Fatty acids

Polyunsaturated fatty acids (PUFAs) are important pharmaconutrients that can exert therapeutic functions to control inflammatory disorders by modulating the immune response. Therapeutic effects of PUFAs against IBD have been demonstrated over the years, and growing evidence suggests that supplementing with PUFAs through the diet could be an interesting strategy for managing IBD<sup>[94]</sup>. The role of omega-3 fatty acids, including EPA and DHA, has been investigated in rats. The results indicate that EPA and DHA combined with olive oil and quercitrin reduced the levels of iNOS, COX-2, TNF- $\alpha$ , LTB<sub>4</sub>, and IL-1 $\beta$  in colitic rats<sup>[95]</sup>. It is assumed that EPA and DHA-derived metabolites, namely, protectin, resolvin, and maresin are the factors responsible for the anti-inflammatory functions<sup>[96]</sup>. An adjunct therapy of omega-3 PUFAs with  $\epsilon$ -5-ASA showed that the dual therapy was more effective in downregulating NF- $\kappa$ B and inducing PPAR $\gamma$  in a rat model of colitis than a higher concentration of 5-ASA alone<sup>[97]</sup>. Administering EPA together with arachidonic acid (AA) to colitic mice resulted in decreased TNF- $\alpha$  and IL-6 and increased PPAR $\gamma$ <sup>[98]</sup>. The protective role of conjugated linoleic acid (CLA) on IBD was investigated by Bassaganya-Riera and Hontecillas (2006), and they concluded that CLA could efficiently delay the onset of colitis and decrease the severity of inflammation by influencing PPAR $\gamma$  expression<sup>[99]</sup>. Alpha linoleic acid from sage oil significantly lowered the inflammatory damage in experimental colitis by decreasing the levels of IL-6, COX-2, and TNF- $\alpha$ <sup>[100]</sup>.

Short chain fatty acids (SCFAs) including acetate, propionate, and butyrate have exhibited therapeutic benefits for colitis. Butyrate limits the immune response and modulates the inflammatory mediators to alleviate mucosal inflammation<sup>[101]</sup>. In a previous study, butyrate supplementation to colitic rats maintained the integrity of the colonic mucosa by enhancing the production of regulatory T cells (Tregs) in blood and the plasma levels of IL-10 and IL-12<sup>[102]</sup>. Segain *et al.*<sup>[103]</sup> reported that butyrate inhibited the NF- $\kappa$ B activation and degraded I $\kappa$ B $\alpha$  level in a rat model of colitis. Other SCFAs, including acetate and propionate also showed preventive activity against IBD in mice by inhibiting the expression of

immune-related genes and inflammatory mediators, such as NF- $\kappa$ B and IL-6<sup>[104]</sup>.

### Bioactive peptides

Dietary peptides have displayed bioactive functions against several illnesses, including chronic inflammation, diabetes, hypertension, and oxidation<sup>[105]</sup>. Therefore bioactive peptides have the potential to be used as an alternative therapy for IBD and other chronic inflammatory disorders. According to Hou *et al.*<sup>[106]</sup>, treatment with alanyl-glutamine in a mouse model of colitis suppressed Th-17 cytokines and macrophage migration to the peritoneal cavity, indicating a reduction in the inflammatory response. Propionyl-L-carnitine, an essential factor of transporting fatty acids in mitochondria reduced mucosal inflammation through antioxidative effects in TNBS-induced colitis<sup>[107]</sup>. Bovine glycomacropeptide resulted in decreased mucosal damage in the colon, decreased MPO activity, and increased IL-10 in lymphocyte-driven colitis<sup>[108]</sup>. In a study by Azuma *et al.*<sup>[109]</sup>, fish scale gelatin peptide demonstrated anti-inflammatory functions in ulcerative colitis through its inhibitory actions against the activation of NF- $\kappa$ B and the accumulation of monocyte chemoattractant protein-1 (MCP-1) in serum. Bioactive peptides isolated from salmon also showed anti-inflammatory functions in experimental colitis in mice<sup>[110]</sup>.

---

## FUTURE TRENDS

The inefficiency of current drug therapies along with the increasing prevalence of IBD from the West towards East Asian and other westernized countries and its recent globalization have triggered a significant amount of research aiming to develop alternative therapies based on natural substances that are highly effective and safe. A coordinated effort based on identifying and solving the environmental and dietary risk factors for IBD will be a priority in the future<sup>[111]</sup>. As diet is one of the key etiological factors of the disease, a multifaceted dietary intervention involving the elimination of certain foods and the inclusion of food components that can target the underlying causes of IBD is immensely needed. Manipulation of the gut ecosystem with probiotic bacteria is an interesting topic of research in the management of IBD. However, current data for the recommendation of probiotics for chronic metabolic illnesses are still insufficient. Recently, "designer probiotics" has drawn attention as an innovative approach, where genetically engineered bacteria with specific functionalities are administered to patients<sup>[112]</sup>. Promising results were found when recombinant *Bifidobacteria* were used as carriers for alpha-melanocyte and manganese superoxide dismutase in experimental colitis<sup>[113,114]</sup>. In recent years, "specific targeting" which allows nutrients or bioactive compounds to reach and target the specific site of inflammation to exert their effects, has become a trending topic of research for IBD control and will

be of tremendous importance in future research<sup>[115]</sup>. Development of novel cell models that can simulate the GI tract is considered to be a futuristic model of research in the quest for natural alternative therapies for IBD<sup>[116]</sup>. More importantly, the complete and precise understanding of the pathogenic mechanisms of IBD is necessary, as it will help researchers find suitable and efficacious treatments for IBD using available and prospective natural therapeutic agents. Therefore, future research studies will be centered upon the development of more effective therapeutic strategies for IBD based on health functional materials that will be capable of reaching the target site and exerting their functions to control the underlying pathogenic mechanisms of IBD.

## CONCLUSION

Diets and functional foods have emerged as promising alternatives for the prevention and treatment of IBD during the past decade. While diets and dietary habits are key modulating factors involved in the pathogenesis of IBD, several food components such as dietary fibers, probiotics, non-starch polysaccharides, and fat soluble vitamins, have been effective in ameliorating gastrointestinal inflammation. Functional foods and bioactive compounds, including plant-derived extracts, phytochemicals, antioxidants, omega-3 fatty acids, and dietary peptides, have exhibited strong anti-inflammatory effects against IBD both in animal models and human subjects. Functional foods can modulate inflammatory cytokines and can interact with the immune system to produce anti-inflammatory functions against IBD. Therefore, diets and functional foods will play a significant role to control IBD in near future. At the same time, regular food intake, well-managed lifestyle, rest, and medication would require enough attention for the efficient management of IBD.

## ACKNOWLEDGMENTS

This work was supported by the Korea Institute of Planning and Evaluation for Technology in Food, Agriculture, Forestry and Fisheries (IPET) through the High Value-added Food Technology Development Program, funded by the Ministry of Agriculture, Food and Rural Affairs (MAFRA; 117073-3).

## REFERENCES

- 1 **Pithadia AB**, Jain S. Treatment of inflammatory bowel disease (IBD). *Pharmacol Rep* 2011; **63**: 629-642 [PMID: 21857074]
- 2 **Burisch J**, Jess T, Martinato M, Lakatos PL; ECCO -EpiCom. The burden of inflammatory bowel disease in Europe. *J Crohns Colitis* 2013; **7**: 322-337 [PMID: 23395397 DOI: 10.1016/j.crohns.2013.01.010]
- 3 **Bhandari S**, Larson ME, Kumar N, Stein D. Association of Inflammatory Bowel Disease (IBD) with Depressive Symptoms in the United States Population and Independent Predictors of Depressive Symptoms in an IBD Population: A NHANES Study. *Gut Liver* 2017; **11**: 512-519 [PMID: 28395506 DOI: 10.5009/gnl16347]
- 4 **Ng SC**. Epidemiology of inflammatory bowel disease: focus on Asia. *Best Pract Res Clin Gastroenterol* 2014; **28**: 363-372 [PMID: 24913377 DOI: 10.1016/j.bpg.2014.04.003]
- 5 **Uranga JA**, López-Miranda V, Lombó F, Abalo R. Food, nutrients and nutraceuticals affecting the course of inflammatory bowel disease. *Pharmacol Rep* 2016; **68**: 816-826 [PMID: 27267792 DOI: 10.1016/j.pharep.2016.05.002]
- 6 **Hur SJ**, Kang SH, Jung HS, Kim SC, Jeon HS, Kim IH, Lee JD. Review of natural products actions on cytokines in inflammatory bowel disease. *Nutr Res* 2012; **32**: 801-816 [PMID: 23176791 DOI: 10.1016/j.nutres.2012.09.013]
- 7 **Durchschein F**, Petritsch W, Hammer HF. Diet therapy for inflammatory bowel diseases: The established and the new. *World J Gastroenterol* 2016; **22**: 2179-2194 [PMID: 26900283 DOI: 10.3748/wjg.v22.i7.2179]
- 8 **Paturi G**, Mandimika T, Butts CA, Zhu S, Roy NC, McNabb WC, Ansell J. Influence of dietary blueberry and broccoli on cecal microbiota activity and colon morphology in mdr1a(-/-) mice, a model of inflammatory bowel diseases. *Nutrition* 2012; **28**: 324-330 [PMID: 22113065 DOI: 10.1016/j.nut.2011.07.018]
- 9 **Hamer HM**, Jonkers DM, Vanhoutvin SA, Troost FJ, Rijkers G, de Bruïne A, Bast A, Venema K, Brummer RJ. Effect of butyrate enemas on inflammation and antioxidant status in the colonic mucosa of patients with ulcerative colitis in remission. *Clin Nutr* 2010; **29**: 738-744 [PMID: 20471725 DOI: 10.1016/j.clnu.2010.04.002]
- 10 **Le Leu RK**, Young GP, Hu Y, Winter J, Conlon MA. Dietary red meat aggravates dextran sulfate sodium-induced colitis in mice whereas resistant starch attenuates inflammation. *Dig Dis Sci* 2013; **58**: 3475-3482 [PMID: 23990000 DOI: 10.1007/s10620-013-2844-1]
- 11 **Di Luccia B**, Manzo N, Baccigalupi L, Calabrò V, Crescenzi E, Ricca E, Pollice A. Lactobacillus gasseri SF1183 affects intestinal epithelial cell survival and growth. *PLoS One* 2013; **8**: e69102 [PMID: 23894414 DOI: 10.1016/j.jff.2017.12.049]
- 12 **Lim HS**, Kim SK, Hong SJ. Food Elimination Diet and Nutritional Deficiency in Patients with Inflammatory Bowel Disease. *Clin Nutr Res* 2018; **7**: 48-55 [PMID: 29423389 DOI: 10.7762/cnr.2018.7.1.48]
- 13 **Farombi EO**, Adedara IA, Ajayi BO, Ayepola OR, Egbeme EE. Kolaviron, a natural antioxidant and anti-inflammatory phytochemical prevents dextran sulphate sodium-induced colitis in rats. *Basic Clin Pharmacol Toxicol* 2013; **113**: 49-55 [PMID: 23336970 DOI: 10.1111/bcpt.12050]
- 14 **Sobczak M**, Zakrzewski PK, Cygankiewicz AI, Mokrowiecka A, Chen C, Sałaga M, Małacka-Panas E, Kordek R, Krajewska WM, Fichna J. Anti-inflammatory action of a novel orally available peptide 317 in mouse models of inflammatory bowel diseases. *Pharmacol Rep* 2014; **66**: 741-750 [PMID: 25149976 DOI: 10.1016/j.pharep.2014.03.007]
- 15 **Chen P**, Zhou X, Zhang L, Shan M, Bao B, Cao Y, Kang A, Ding A. Anti-inflammatory effects of Huangqin tang extract in mice on ulcerative colitis. *J Ethnopharmacol* 2015; **162**: 207-214 [PMID: 25576893 DOI: 10.1016/j.jep.2014.12.039]
- 16 **Zhang H**, Liu R, Tsao R. Anthocyanin-rich phenolic extracts of purple root vegetables inhibit pro-inflammatory cytokines induced by H<sub>2</sub>O<sub>2</sub> and enhance antioxidant enzyme activities in Caco-2 cells. *J Funct Foods* 2016; **22**: 363-375 [DOI: 10.1016/j.jff.2016.01.004]
- 17 **Hwang YH**, Kim DG, Li W, Yang HJ, Yim NH, Ma JY. Anti-inflammatory effects of Forsythia suspensa in dextran sulfate sodium-induced colitis. *J Ethnopharmacol* 2017; **206**: 73-77 [PMID: 28502906 DOI: 10.1016/j.jep.2017.05.011]
- 18 **Ali AA**, Abd Al Haleem EN, Khaleel SA, Sallam AS. Protective effect of cardamonin against acetic acid-induced ulcerative colitis in rats. *Pharmacol Rep* 2017; **69**: 268-275 [PMID: 28129600 DOI: 10.1016/j.pharep.2016.11.002]
- 19 **Kirsner JB**. Historical origins of current IBD concepts. *World J Gastroenterol* 2001; **7**: 175-184 [PMID: 11819757 DOI: 10.3748/

- wjg.v7.i2.175]
- 20 **Knight-Sepulveda K**, Kais S, Santaolalla R, Abreu MT. Diet and Inflammatory Bowel Disease. *Gastroenterol Hepatol* (NY) 2015; **11**: 511-520 [PMID: 27118948]
  - 21 **Ardizzone S**, Bianchi Porro G. Biologic therapy for inflammatory bowel disease. *Drugs* 2005; **65**: 2253-2286 [PMID: 16266194 DOI: 10.2165/00003495-200565160-00002]
  - 22 **Zhao L**, Wu H, Zhao A, Lu H, Sun W, Ma C, Yang Y, Xin X, Zou H, Qiu M, Jia W. The in vivo and in vitro study of polysaccharides from a two-herb formula on ulcerative colitis and potential mechanism of action. *J Ethnopharmacol* 2014; **153**: 151-159 [PMID: 24548752 DOI: 10.1016/j.jep.2014.02.008]
  - 23 **Horváth K**, Varga C, Berkó A, Pósa A, László F, Whittle BJ. The involvement of heme oxygenase-1 activity in the therapeutic actions of 5-aminosalicylic acid in rat colitis. *Eur J Pharmacol* 2008; **581**: 315-323 [PMID: 18215658 DOI: 10.1016/j.ejphar.2007.12.004]
  - 24 **Ito K**, Chung KF, Adcock IM. Update on glucocorticoid action and resistance. *J Allergy Clin Immunol* 2006; **117**: 522-543 [PMID: 16522450 DOI: 10.1016/j.jaci.2006.01.032]
  - 25 **Renna S**, Cottone M, Orlando A. Optimization of the treatment with immunosuppressants and biologics in inflammatory bowel disease. *World J Gastroenterol* 2014; **20**: 9675-9690 [PMID: 25110407 DOI: 10.3748/wjg.v20.i29.9675]
  - 26 **Nitzan O**, Elias M, Peretz A, Saliba W. Role of antibiotics for treatment of inflammatory bowel disease. *World J Gastroenterol* 2016; **22**: 1078-1087 [PMID: 26811648 DOI: 10.3748/wjg.v22.i3.1078]
  - 27 **Crowe JS**, Roberts KJ, Carlton TM, Maggiore L, Cubitt MF, Clare S, Harcourt K, Reckless J, MacDonald TT, Ray KP, Vossenkämper A, West MR. Preclinical Development of a Novel, Orally-Administered Anti-Tumour Necrosis Factor Domain Antibody for the Treatment of Inflammatory Bowel Disease. *Sci Rep* 2018; **8**: 4941 [PMID: 29563546 DOI: 10.1038/s41598-018-23277-7]
  - 28 **Moura FA**, de Andrade KQ, dos Santos JC, Araújo OR, Goulart MO. Antioxidant therapy for treatment of inflammatory bowel disease: Does it work? *Redox Biol* 2015; **6**: 617-639 [PMID: 26520808 DOI: 10.1016/j.redox.2015.10.006]
  - 29 **Currò D**, Ianiro G, Pecere S, Bibbò S, Cammarota G. Probiotics, fibre and herbal medicinal products for functional and inflammatory bowel disorders. *Br J Pharmacol* 2017; **174**: 1426-1449 [PMID: 27696378 DOI: 10.1111/bph.13632]
  - 30 **Derikx LA**, Dieleman LA, Hoentjen F. Probiotics and prebiotics in ulcerative colitis. *Best Pract Res Clin Gastroenterol* 2016; **30**: 55-71 [PMID: 27048897 DOI: 10.1016/j.bpg.2016.02.005]
  - 31 **Srutkova D**, Schwarzer M, Hudcovic T, Zakostelska Z, Drab V, Spanova A, Rittich B, Kozakova H, Schabussova I. Bifidobacterium longum CCM 7952 Promotes Epithelial Barrier Function and Prevents Acute DSS-Induced Colitis in Strictly Strain-Specific Manner. *PLoS One* 2015; **10**: e0134050 [PMID: 26218526 DOI: 10.1371/journal.pone.0134050]
  - 32 **Lee JH**, Lee B, Lee HS, Bae EA, Lee H, Ahn YT, Lim KS, Huh CS, Kim DH. Lactobacillus suntoryeus inhibits pro-inflammatory cytokine expression and TLR-4-linked NF-kappaB activation in experimental colitis. *Int J Colorectal Dis* 2009; **24**: 231-237 [PMID: 19050899 DOI: 10.1007/s00384-008-0618-6]
  - 33 **Kawahara M**, Nemoto M, Nakata T, Kondo S, Takahashi H, Kimura B, Kuda T. Anti-inflammatory properties of fermented soy milk with Lactococcus lactis subsp. lactis S-SU2 in murine macrophage RAW264.7 cells and DSS-induced IBD model mice. *Int Immunopharmacol* 2015; **26**: 295-303 [PMID: 25887264 DOI: 10.1016/j.intimp.2015.04.004]
  - 34 **Park JS**, Joe I, Rhee PD, Jeong CS, Jeong G. A lactic acid bacterium isolated from kimchi ameliorates intestinal inflammation in DSS-induced colitis. *J Microbiol* 2017; **55**: 304-310 [PMID: 28124779 DOI: 10.1007/s12275-017-6447-y]
  - 35 **Shigemori S**, Watanabe T, Kudoh K, Ihara M, Nigar S, Yamamoto Y, Suda Y, Sato T, Kitazawa H, Shimosato T. Oral delivery of Lactococcus lactis that secretes bioactive heme oxygenase-1 alleviates development of acute colitis in mice. *Microb Cell Fact* 2015; **14**: 189 [PMID: 26608030 DOI: 10.1186/s12934-015-0378-2]
  - 36 **Yokota Y**, Shikano A, Kuda T, Takei M, Takahashi H, Kimura B. Lactobacillus plantarum AN1 cells increase caecal L. reuteri in an ICR mouse model of dextran sodium sulphate-induced inflammatory bowel disease. *Int Immunopharmacol* 2018; **56**: 119-127 [PMID: 29414641 DOI: 10.1016/j.intimp.2018.01.020]
  - 37 **Le B**, Yang SH. Efficacy of Lactobacillus plantarum in prevention of inflammatory bowel disease. *Toxicol Rep* 2018; **5**: 314-317 [DOI: 10.1016/j.toxrep.2018.02.007]
  - 38 **Seo S**, Shin J-S, Lee WS, Rhee YK, Cho CW, Hong H-D, Lee K-T. Anti-colitis effect of Lactobacillus sakei K040706 via suppression of inflammatory responses in the dextran sulfate sodium-induced colitis mice model. *J Funct Food* 2017; **29**: 256-268 [DOI: 10.1016/j.jff.2016.12.045]
  - 39 **Gholami M**, Ghasemi-Niri SF, Maqbool F, Baeri M, Memariani Z, Pousti I, Abdollahi M. Experimental and Pathological study of Pistacia atlantica, butyrate, Lactobacillus casei and their combination on rat ulcerative colitis model. *Pathol Res Pract* 2016; **212**: 500-508 [PMID: 26972417 DOI: 10.1016/j.prp.2016.02.024]
  - 40 **Lim SM**, Kim DH. Bifidobacterium adolescentis IM38 ameliorates high-fat diet-induced colitis in mice by inhibiting NF-κB activation and lipopolysaccharide production by gut microbiota. *Nutr Res* 2017; **41**: 86-96 [PMID: 28479226 DOI: 10.1016/j.nutres.2017.04.003]
  - 41 **Satish Kumar CS**, Kondal Reddy K, Boobalan G, Gopala Reddy A, Sudha Rani Chowdhary CH, Vinoth A, Jayakanth K, Srinivasa Rao G. Immunomodulatory effects of Bifidobacterium bifidum 231 on trinitrobenzenesulfonic acid-induced ulcerative colitis in rats. *Res Vet Sci* 2017; **110**: 40-46 [PMID: 28159236 DOI: 10.1016/j.rvsc.2016.10.010]
  - 42 **Liu X**, Wu Y, Li F, Zhang D. Dietary fiber intake reduces risk of inflammatory bowel disease: result from a meta-analysis. *Nutr Res* 2015; **35**: 753-758 [PMID: 26126709 DOI: 10.1016/j.nutres.2015.05.021]
  - 43 **Nie Y**, Lin Q, Luo F. Effects of Non-Starch Polysaccharides on Inflammatory Bowel Disease. *Int J Mol Sci* 2017; **18**: pii: E1372 [PMID: 28654020 DOI: 10.3390/ijms18071372]
  - 44 **Suwannaporn P**, Thep Wong K, Tester R, Al-Ghazzewi F, Piggott J, Shen N, Chen Z, Chen F, Yang J, Zhang D, Tang M. Tolerance and nutritional therapy of dietary fibre from konjac glucomannan hydrolysates for patients with inflammatory bowel disease (IBD). *Bioact Carbohydr Diet Fiber* 2013; **2**: 93-98 [DOI: 10.1016/j.bcdf.2013.09.005]
  - 45 **Asano T**, Tanaka K, Suemasu S, Ishihara T, Tahara K, Suzuki T, Suzuki H, Fukudo S, Mizushima T. Effects of β-(1,3-1,6)-D-glucan on irritable bowel syndrome-related colonic hypersensitivity. *Biochem Biophys Res Commun* 2012; **420**: 444-449 [PMID: 22430139 DOI: 10.1016/j.bbrc.2012.03.015]
  - 46 **Liu B**, Lin Q, Yang T, Zeng L, Shi L, Chen Y, Luo F. Oat β-glucan ameliorates dextran sulfate sodium (DSS)-induced ulcerative colitis in mice. *Food Funct* 2015; **6**: 3454-3463 [PMID: 26292622 DOI: 10.1039/c5fo00563a]
  - 47 **Lavi I**, Levinson D, Peri I, Nimri L, Hadar Y, Schwartz B. Orally administered glucans from the edible mushroom Pleurotus pulmonarius reduce acute inflammation in dextran sulfate sodium-induced experimental colitis. *Br J Nutr* 2010; **103**: 393-402 [PMID: 19772681 DOI: 10.1017/S0007114509991760]
  - 48 **Lee KH**, Park M, Ji KY, Lee HY, Jang JH, Yoon IJ, Oh SS, Kim SM, Jeong YH, Yun CH, Kim MK, Lee IY, Choi HR, Ko KS, Kang HS. Bacterial β-(1,3)-glucan prevents DSS-induced IBD by restoring the reduced population of regulatory T cells. *Immunobiology* 2014; **219**: 802-812 [PMID: 25092569 DOI: 10.1016/j.imbio.2014.07.003]
  - 49 **Azuma K**, Osaki T, Ifuku F, Saimoto H, Morimoto M, Takashima O, Tsuka T, Imagawa T, Okamoto Y, Minami S. Suppressive effects of cellulose nanofibers—made from adlay and seaweed—on colon inflammation in an inflammatory bowel-disease model. *Bioact Carbohydr Diet Fiber* 2013; **3**: 65-72 [DOI: 10.1016/j.bcdf.2013.09.006]

- 50 **Azuma K**, Osaki T, Ifuku F, Saimoto H, Morimoto M, Takashima O, Tsuka T, Imagawa T, Okamoto Y, Minami S. Anti-inflammatory effects of cellulose nanofiber made from pear in inflammatory bowel disease model. *Bioact Carbohydr Diet Fiber* 2014; **3**: 1-10 [DOI: 10.1016/j.bcdf.2013.11.001]
- 51 **Capitán-Cañadas F**, Ocón B, Aranda CJ, Anzola A, Suárez MD, Zarzuelo A, de Medina FS, Martínez-Augustín O. Fructooligosaccharides exert intestinal anti-inflammatory activity in the CD4+ CD62L+ T cell transfer model of colitis in C57BL/6J mice. *Eur J Nutr* 2016; **55**: 1445-1454 [PMID: 26154776 DOI: 10.1007/s00394-015-0962-6]
- 52 **Lara-Villoslada F**, Debras E, Nieto A, Concha A, Gálvez J, López-Huertas E, Boza J, Obled C, Xaus J. Oligosaccharides isolated from goat milk reduce intestinal inflammation in a rat model of dextran sodium sulfate-induced colitis. *Clin Nutr* 2006; **25**: 477-488 [PMID: 16375993 DOI: 10.1016/j.clnu.2005.11.004]
- 53 **Štofilová J**, Szabadosová V, Hřčková G, Salaj R, Bertková I, Hijová E, Strojný L, Bomba A. Co-administration of a probiotic strain *Lactobacillus plantarum* LS/07 CCM7766 with prebiotic inulin alleviates the intestinal inflammation in rats exposed to N,N-dimethylhydrazine. *Int Immunopharmacol* 2015; **24**: 361-368 [PMID: 25536541 DOI: 10.1016/j.intimp.2014.12.022]
- 54 **Gubatan J**, Mitsuhashi S, Zenlea T, Rosenberg L, Robson S, Moss AC. Low Serum Vitamin D During Remission Increases Risk of Clinical Relapse in Patients With Ulcerative Colitis. *Clin Gastroenterol Hepatol* 2017; **15**: 240-246.e1 [PMID: 27266980 DOI: 10.1016/j.cgh.2016.05.035]
- 55 **Meckel K**, Li YC, Lim J, Kocherginsky M, Weber C, Almoghrabi A, Chen X, Kaboff A, Sadiq F, Hanauer SB, Cohen RD, Kwon J, Rubin DT, Hanan I, Sakuraba A, Yen E, Bissonnette M, Pekow J. Serum 25-hydroxyvitamin D concentration is inversely associated with mucosal inflammation in patients with ulcerative colitis. *Am J Clin Nutr* 2016; **104**: 113-120 [PMID: 27281309 DOI: 10.3945/ajcn.115.123786]
- 56 **Chen SW**, Wang PY, Zhu J, Chen GW, Zhang JL, Chen ZY, Zuo S, Liu YC, Pan YS. Protective effect of 1,25-dihydroxyvitamin D3 on lipopolysaccharide-induced intestinal epithelial tight junction injury in caco-2 cell monolayers. *Inflammation* 2015; **38**: 375-383 [PMID: 25344656 DOI: 10.1007/s10753-014-0041-9]
- 57 **Zhu Y**, Mahon BD, Froicu M, Cantorna MT. Calcium and 1 alpha,25-dihydroxyvitamin D3 target the TNF-alpha pathway to suppress experimental inflammatory bowel disease. *Eur J Immunol* 2005; **35**: 217-224 [PMID: 15593122 DOI: 10.1002/eji.200425491]
- 58 **Bartels LE**, Jørgensen SP, Agnholt J, Kelsen J, Hvas CL, Dahlerup JF. 1,25-dihydroxyvitamin D3 and dexamethasone increase interleukin-10 production in CD4+ T cells from patients with Crohn's disease. *Int Immunopharmacol* 2007; **7**: 1755-1764 [PMID: 17996686 DOI: 10.1016/j.intimp.2007.09.016]
- 59 **Bendix-Struve M**, Bartels LE, Agnholt J, Dige A, Jørgensen SP, Dahlerup JF. Vitamin D3 treatment of Crohn's disease patients increases stimulated T cell IL-6 production and proliferation. *Aliment Pharmacol Ther* 2010; **32**: 1364-1372 [PMID: 21050239 DOI: 10.1111/j.1365-2036.2010.04463.x]
- 60 **Reifen R**, Levy E, Berkovich Z, Tirosh O. Vitamin A exerts its anti-inflammatory activities in colitis through preservation of mitochondrial activity. *Nutrition* 2015; **31**: 1402-1407 [PMID: 26429662 DOI: 10.1016/j.nut.2015.05.011]
- 61 **Gottschall E**. Breaking the Vicious Cycle: Intestinal Health Through Diet. Balti—more, Canada: Kirkton Press; 2012
- 62 **Cohen SA**, Gold BD, Oliva S, Lewis J, Stallworth A, Koch B, Eshee L, Mason D. Clinical and mucosal improvement with specific carbohydrate diet in pediatric Crohn disease. *J Pediatr Gastroenterol Nutr* 2014; **59**: 516-521 [PMID: 24897165 DOI: 10.1097/MPG.0000000000000449]
- 63 **Marion-Letellier R**, Savoye G, Ghosh S. IBD: In Food We Trust. *J Crohns Colitis* 2016; **10**: 1351-1361 [PMID: 27194533 DOI: 10.1093/ecco-jcc/jjw106]
- 64 **Marcason W**. What is the FODMAP diet? *J Acad Nutr Diet* 2012; **112**: 1696 [PMID: 23017576]
- 65 **Geary RB**, Irving PM, Barrett JS, Nathan DM, Shepherd SJ, Gibson PR. Reduction of dietary poorly absorbed short-chain carbohydrates (FODMAPs) improves abdominal symptoms in patients with inflammatory bowel disease—a pilot study. *J Crohns Colitis* 2009; **3**: 8-14 [PMID: 21172242 DOI: 10.1016/j.crohns.2008.09.004]
- 66 **Pedersen N**, Ankersen DV, Felding M, Wachmann H, Végh Z, Molzen L, Burisch J, Andersen JR, Munkholm P. Low-FODMAP diet reduces irritable bowel symptoms in patients with inflammatory bowel disease. *World J Gastroenterol* 2017; **23**: 3356-3366 [PMID: 28566897 DOI: 10.3748/wjg.v23.i18.3356]
- 67 **Larussa T**, Imeneo M, Luzza F. Potential role of nutraceutical compounds in inflammatory bowel disease. *World J Gastroenterol* 2017; **23**: 2483-2492 [PMID: 28465632 DOI: 10.3748/wjg.v23.i14.2483]
- 68 **Lim BO**. *Coriolus versicolor* suppresses inflammatory bowel disease by inhibiting the expression of STAT1 and STAT6 associated with IFN- $\gamma$  and IL-4 expression. *Phytother Res* 2011; **25**: 1257-1261 [PMID: 21796702 DOI: 10.1002/ptr.3378]
- 69 **Won SY**, Park EH. Anti-inflammatory and related pharmacological activities of cultured mycelia and fruiting bodies of *Cordyceps militaris*. *J Ethnopharmacol* 2005; **96**: 555-561 [PMID: 15619578 DOI: 10.1016/j.jep.2004.10.009]
- 70 **Han ES**, Oh JY, Park HJ. *Cordyceps militaris* extract suppresses dextran sodium sulfate-induced acute colitis in mice and production of inflammatory mediators from macrophages and mast cells. *J Ethnopharmacol* 2011; **134**: 703-710 [PMID: 21277968 DOI: 10.1016/j.jep.2011.01.022]
- 71 **Jeon TI**, Hwang SG, Lim BO, Park DK. Extracts of *Pheillus linteus* grown on germinated brown rice suppress liver damage induced by carbon tetrachloride in rats. *Biotechnol Lett* 2003; **25**: 2093-2096 [PMID: 14969415]
- 72 **Debnath T**, Park SR, Kim DH, Jo JE, Lim BO. Anti-oxidant and anti-inflammatory activities of *Inonotus obliquus* and germinated brown rice extracts. *Molecules* 2013; **18**: 9293-9304 [PMID: 23917116 DOI: 10.3390/molecules18089293]
- 73 **Debnath T**, Hasnat MA, Pervin M, Lee SY, Park SR, Kim DH, Kweon HJ, Kim JM, Lim BO. Chaga Mushroom (*Inonotus obliquus*) Grown on Germinated Brown Rice Suppresses Inflammation Associated with Colitis in Mice. *Food Sci Biotechnol* 2012; **21**: 1235-1241 [DOI: 10.1007/s10068-012-0162-6]
- 74 **Hasnat MA**, Pervin M, Cha KM, Kim SK, Lim BO. Anti-inflammatory activity on mice of extract of *Ganoderma lucidum* grown on rice via modulation of MAPK and NF- $\kappa$ B pathways. *Phytochemistry* 2015; **114**: 125-136 [PMID: 25457483 DOI: 10.1016/j.phytochem.2014.10.019]
- 75 **Jin HL**, Lee BR, Lim KJ, Debnath T, Shin HM, Lim BO. Anti-inflammatory Effects of Prunus mume Mixture in Colitis Induced by Dextran Sodium Sulfate. *Kor J Medicinal Crop Sci* 2011; **19**: 16-23 [DOI: 10.7783/KJMCS.2011.19.1.016]
- 76 **Kamali M**, Tavakoli H, Khodadoost M, Daghighzadeh H, Kamalinejad M, Gachkar L, Mansourian M, Adibi P. Efficacy of the *Punica granatum* peels aqueous extract for symptom management in ulcerative colitis patients. A randomized, placebo-controlled, clinical trial. *Complement Ther Clin Pract* 2015; **21**: 141-146 [PMID: 26256131 DOI: 10.1016/j.ctcp.2015.03.001]
- 77 **Rosillo MA**, Sánchez-Hidalgo M, Cárdeno A, Aparicio-Soto M, Sánchez-Fidalgo S, Villegas I, de la Lastra CA. Dietary supplementation of an ellagic acid-enriched pomegranate extract attenuates chronic colonic inflammation in rats. *Pharmacol Res* 2012; **66**: 235-242 [PMID: 22677088 DOI: 10.1016/j.phrs.2012.05.006]
- 78 **Xiao X**, Kim J, Sun Q, Kim D, Park CS, Lu TS, Park Y. Preventive effects of cranberry products on experimental colitis induced by dextran sulphate sodium in mice. *Food Chem* 2015; **167**: 438-446 [PMID: 25149009 DOI: 10.1016/j.foodchem.2014.07.006]
- 79 **Pervin M**, Hasnat MA, Lim JH, Lee YM, Kim EO, Um BH, Lim BO. Preventive and therapeutic effects of blueberry (*Vaccinium corymbosum*) extract against DSS-induced ulcerative colitis by regulation of antioxidant and inflammatory mediators. *J Nutr Biochem* 2016; **28**: 103-113 [PMID: 26878787 DOI: 10.1016/

- j.jnutbio.2015.10.006]
- 80 **Suluvooy JK**, Sakthivel KM, Guruvayoorappan C, Berlin Grace VM. Protective effect of Averrhoa bilimbi L. fruit extract on ulcerative colitis in wistar rats via regulation of inflammatory mediators and cytokines. *Biomed Pharmacother* 2017; **91**: 1113-1121 [PMID: 28531922 DOI: 10.1016/j.biopha.2017.05.057]
  - 81 **Valcheva-Kuzmanova S**, Kuzmanov A, Kuzmanova V, Tzaneva M. Aronia melanocarpa fruit juice ameliorates the symptoms of inflammatory bowel disease in TNBS-induced colitis in rats. *Food Chem Toxicol* 2018; **113**: 33-39 [PMID: 29331733 DOI: 10.1016/j.fct.2018.01.011]
  - 82 **Hsiang CY**, Lo HY, Huang HC, Li CC, Wu SL, Ho TY. Ginger extract and zingerone ameliorated trinitrobenzene sulphonic acid-induced colitis in mice via modulation of nuclear factor- $\kappa$ B activity and interleukin-1 $\beta$  signalling pathway. *Food Chem* 2013; **136**: 170-177 [PMID: 23017409 DOI: 10.1016/j.foodchem.2012.07.124]
  - 83 **Debnath T**, Mijan MA, Kim DH, Jo JE, Kim YO, Lee JJ, Han JP, Lim BO. Anti-inflammatory effects of *Haliotis discus hannai* Ino on dextran sulfate sodium-induced colitis in mice. *J Food Biochem* 2015; **39**: 209-217 [DOI: 10.1111/jfbc.12118]
  - 84 **Bitencourt MA**, Silva HM, Abilio GM, Miranda GE, Moura AM, Araújo-Júnior JX, Silveira EJ, Santos BV, Souto JT. Anti-inflammatory effects of methanolic extract of green algae *Caulerpa mexicana* in a murine model of ulcerative colitis. *Rev Brasileira Farmacog* 2015; **25**: 677-682 [DOI: 10.1016/j.bjp.2015.10.001]
  - 85 **D'Argenio G**, Mazzone G, Tuccillo C, Ribecco MT, Graziani G, Gravina AG, Caserta S, Guido S, Fogliano V, Caporaso N, Romano M. Apple polyphenols extract (APE) improves colon damage in a rat model of colitis. *Dig Liver Dis* 2012; **44**: 555-562 [PMID: 22381211 DOI: 10.1016/j.dld.2012.01.009]
  - 86 **Martín AR**, Villegas I, Sánchez-Hidalgo M, de la Lastra CA. The effects of resveratrol, a phytoalexin derived from red wines, on chronic inflammation induced in an experimentally induced colitis model. *Br J Pharmacol* 2006; **147**: 873-885 [PMID: 16474422 DOI: 10.1038/sj.bjp.0706469]
  - 87 **Samsami-Kor M**, Daryani NE, Asl PR, Hekmatdoost A. Anti-inflammatory Effects of Resveratrol in Patients with Ulcerative Colitis: A Randomized, Double-Blind, Placebo-controlled Pilot Study. *Arch Med Res* 2015; **46**: 280-285 [PMID: 26002728 DOI: 10.1016/j.arcmed.2015.05.005]
  - 88 **Chen L**, Wilson JE, Koenigsnecht MJ, Chou WC, Montgomery SA, Truax AD, Brickey WJ, Packey CD, Maharshak N, Matsushima GK, Plevy SE, Young VB, Sartor RB, Ting JP. NLRP12 attenuates colon inflammation by maintaining colonic microbial diversity and promoting protective commensal bacterial growth. *Nat Immunol* 2017; **18**: 541-551 [PMID: 28288099 DOI: 10.1038/ni.3690]
  - 89 **Zaki MH**, Vogel P, Malireddi RK, Body-Malapel M, Anand PK, Bertin J, Green DR, Lamkanfi M, Kanneganti TD. The NOD-like receptor NLRP12 attenuates colon inflammation and tumorigenesis. *Cancer Cell* 2011; **20**: 649-660 [PMID: 22094258 DOI: 10.1016/j.ccr.2011.10.022]
  - 90 **Zhu G**, Wang H, Wang T, Shi F. Ginsenoside Rg1 attenuates the inflammatory response in DSS-induced mice colitis. *Int Immunopharmacol* 2017; **50**: 1-5 [PMID: 28605639 DOI: 10.1016/j.intimp.2017.06.002]
  - 91 **Wagner AE**, Will O, Sturm C, Lipinski S, Rosenstiel P, Rimbach G. DSS-induced acute colitis in C57BL/6 mice is mitigated by sulfuraphane pre-treatment. *J Nutr Biochem* 2013; **24**: 2085-2091 [PMID: 24231100 DOI: 10.1016/j.jnutbio.2013.07.009]
  - 92 **Liu L**, Liu YL, Liu GX, Chen X, Yang K, Yang YX, Xie Q, Gan HK, Huang XL, Gan HT. Curcumin ameliorates dextran sulfate sodium-induced experimental colitis by blocking STAT3 signaling pathway. *Int Immunopharmacol* 2013; **17**: 314-320 [PMID: 23856612 DOI: 10.1016/j.intimp.2013.06.020]
  - 93 **Shapira S**, Leshno A, Katz D, Maharshak N, Hevroni G, Jean-David M, Kraus S, Galazan L, Aroch I, Kazanov D, Hallack A, Becker S, Umanski M, Moshkowitz M, Dotan I, Arber N. Of mice and men: a novel dietary supplement for the treatment of ulcerative colitis. *Therap Adv Gastroenterol* 2017; **11**: 1756283X17741864 [PMID: 29383023 DOI: 10.1177/1756283X17741864]
  - 94 **Uchiyama K**, Nakamura M, Odahara S, Koido S, Katahira K, Shiraishi H, Ohkusa T, Fujise K, Tajiri H. N-3 polyunsaturated fatty acid diet therapy for patients with inflammatory bowel disease. *Inflamm Bowel Dis* 2010; **16**: 1696-1707 [PMID: 20222122 DOI: 10.1002/ibd.21251]
  - 95 **Camuesco D**, Comalada M, Concha A, Nieto A, Sierra S, Xaus J, Zarzuelo A, Gálvez J. Intestinal anti-inflammatory activity of combined quercitrin and dietary olive oil supplemented with fish oil, rich in EPA and DHA (n-3) polyunsaturated fatty acids, in rats with DSS-induced colitis. *Clin Nutr* 2006; **25**: 466-476 [PMID: 16698151 DOI: 10.1016/j.clnu.2005.12.009]
  - 96 **Schwanke RC**, Marcon R, Bento AF, Calixto JB. EPA- and DHA-derived resolvins' actions in inflammatory bowel disease. *Eur J Pharmacol* 2016; **785**: 156-164 [PMID: 26325092 DOI: 10.1016/j.ejphar.2015.08.050]
  - 97 **Mbodji K**, Charpentier C, Guérin C, Querec C, Bole-Feysot C, Aziz M, Savoye G, Déchelotte P, Marion-Letellier R. Adjunct therapy of n-3 fatty acids to 5-ASA ameliorates inflammatory score and decreases NF- $\kappa$ B in rats with TNBS-induced colitis. *J Nutr Biochem* 2013; **24**: 700-705 [PMID: 22841543 DOI: 10.1016/j.jnutbio.2012.03.022]
  - 98 **Roy N**, Barnett M, Knoch B, Dommels Y, McNabb W. Nutrigenomics applied to an animal model of Inflammatory Bowel Diseases: transcriptomic analysis of the effects of eicosapentaenoic acid- and arachidonic acid-enriched diets. *Mutat Res* 2007; **622**: 103-116 [PMID: 17574631 DOI: 10.1016/j.mrfmmm.2007.04.003]
  - 99 **Bassaganya-Riera J**, Hontecillas R. CLA and n-3 PUFA differentially modulate clinical activity and colonic PPAR-responsive gene expression in a pig model of experimental IBD. *Clin Nutr* 2006; **25**: 454-465 [PMID: 16698153 DOI: 10.1016/j.clnu.2005.12.008]
  - 100 **Reifen R**, Karlinsky A, Stark AH, Berkovich Z, Nyska A.  $\alpha$ -Linolenic acid (ALA) is an anti-inflammatory agent in inflammatory bowel disease. *J Nutr Biochem* 2015; **26**: 1632-1640 [PMID: 26350254 DOI: 10.1016/j.jnutbio.2015.08.006]
  - 101 **Cushing K**, Alvarado DM, Ciorba MA. Butyrate and Mucosal Inflammation: New Scientific Evidence Supports Clinical Observation. *Clin Transl Gastroenterol* 2015; **6**: e108 [PMID: 26312412 DOI: 10.1038/ctg.2015.34]
  - 102 **Zhang M**, Zhou Q, Dorfman RG, Huang X, Fan T, Zhang H, Zhang J, Yu C. Butyrate inhibits interleukin-17 and generates Tregs to ameliorate colorectal colitis in rats. *BMC Gastroenterol* 2016; **16**: 84 [PMID: 27473867 DOI: 10.1186/s12876-016-0500-x]
  - 103 **Segain JP**, Raingeard de la Blétière D, Bourreille A, Leray V, Gervois N, Rosales C, Ferrier L, Bonnet C, Blottière HM, Galmiche JP. Butyrate inhibits inflammatory responses through NF $\kappa$ B inhibition: implications for Crohn's disease. *Gut* 2000; **47**: 397-403 [PMID: 10940278]
  - 104 **Tedelind S**, Westberg F, Kjerrulf M, Vidal A. Anti-inflammatory properties of the short-chain fatty acids acetate and propionate: a study with relevance to inflammatory bowel disease. *World J Gastroenterol* 2007; **13**: 2826-2832 [PMID: 17569118 DOI: 10.3748/wjg.v13.i20.2826]
  - 105 **Chakrabarti S**, Jahandideh F, Wu J. Food-derived bioactive peptides on inflammation and oxidative stress. *Biomed Res Int* 2014; **2014**: 608979 [PMID: 24527452 DOI: 10.1155/2014/608979]
  - 106 **Hou YC**, Liu JJ, Pai MH, Tsou SS, Yeh SL. Alanyl-glutamine administration suppresses Th17 and reduces inflammatory reaction in dextran sulfate sodium-induced acute colitis. *Int Immunopharmacol* 2013; **17**: 1-8 [PMID: 23721689 DOI: 10.1016/j.intimp.2013.05.004]
  - 107 **Scioli MG**, Stasi MA, Passeri D, Doldo E, Costanza G, Camerini R, Fociani P, Arcuri G, Lombardo K, Pace S, Borsini F, Orlandi A. Propionyl-L-Carnitine is Efficacious in Ulcerative Colitis Through its Action on the Immune Function and Microvasculature. *Clin Transl Gastroenterol* 2014; **5**: e55 [PMID: 24646507 DOI: 10.1038/ctg.2014.4]
  - 108 **Ortega-González M**, Capitán-Cañadas F, Requena P, Ocoñ B, Romero-Calvo I, Aranda C, Suárez MD, Zarzuelo A, Sánchez de Medina F, Martínez-Augustín O. Validation of bovine

- glycomacropeptide as an intestinal anti-inflammatory nutraceutical in the lymphocyte-transfer model of colitis. *Br J Nutr* 2014; **111**: 1202-1212 [PMID: 24229852 DOI: 10.1017/S0007114513003590]
- 109 **Azuma K**, Osaki T, Tsuka T, Imagawa T, Okamoto Y, and Minami S. Effects of fish scale collagen peptide on an experimental ulcerative colitis mouse model. *Pharm Nutr* 2014; **2**: 161-168 [DOI: 10.1016/j.phanu.2014.10.001]
- 110 **Grimstad T**, Bjørndal B, Cacabelos D, Aasprong OG, Omdal R, Svardal A, Bohov P, Pamplona R, Portero-Otin M, Berge RK, Hausken T. A salmon peptide diet alleviates experimental colitis as compared with fish oil. *J Nutr Sci* 2013; **2**: e2 [PMID: 25191568 DOI: 10.1017/jns.2012.23]
- 111 **Kaplan GG**, Ng SC. Understanding and Preventing the Global Increase of Inflammatory Bowel Disease. *Gastroenterology* 2017; **152**: 313-321.e2 [PMID: 27793607 DOI: 10.1053/j.gastro.2016.10.020]
- 112 **Singh B**, Mal G, Marotta F. Designer Probiotics: Paving the Way to Living Therapeutics. *Trends Biotechnol* 2017; **35**: 679-682 [PMID: 28483159 DOI: 10.1016/j.tibtech.2017.04.001]
- 113 **Wei P**, Yang Y, Liu Z, Huang J, Gong Y, Sun H. Oral Bifidobacterium longum expressing alpha-melanocyte-stimulating hormone to fight experimental colitis. *Drug Deliv* 2016; **23**: 2058-2064 [PMID: 26673899 DOI: 10.3109/10717544.2015.1122672]
- 114 **Liu M**, Li S, Zhang Q, Xu Z, Wang J, Sun H. Oral engineered Bifidobacterium longum expressing rhMnSOD to suppress experimental colitis. *Int Immunopharmacol* 2018; **57**: 25-32 [PMID: 29455070 DOI: 10.1016/j.intimp.2018.02.004]
- 115 **Romach E**, Uni Z, Friedman M, Aizenberg I, Berkovich Z, Reifen R. A new mode of probiotic therapy: Specific targeting. *J Funct Foods* 2015; **16**: 386-392 [DOI: 10.1016/j.jff.2015.04.029]
- 116 **Cencic A**, Chingwaru W. The role of functional foods, nutraceuticals, and food supplements in intestinal health. *Nutrients* 2010; **2**: 611-625 [PMID: 22254045 DOI: 10.3390/nu2060611]

**P- Reviewer:** Eleftheriadis NP, Guan YS, Ierardi E  
**S- Editor:** Gong ZM **L- Editor:** A **E- Editor:** Huang Y





Published by **Baishideng Publishing Group Inc**  
7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA  
Telephone: +1-925-223-8242  
Fax: +1-925-223-8243  
E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
Help Desk: <http://www.f6publishing.com/helpdesk>  
<http://www.wjgnet.com>



ISSN 1007-9327

