

# World Journal of *Gastroenterology*

*World J Gastroenterol* 2018 November 14; 24(42): 4721-4834



**EDITORIAL**

- 4721 Increased susceptibility of aging gastric mucosa to injury and delayed healing: Clinical implications  
*Tarnawski AS, Ahluwalia A*

**REVIEW**

- 4728 Liver as a target of human immunodeficiency virus infection  
*Ganesan M, Poluektova LY, Kharbanda KK, Osna NA*

**MINIREVIEWS**

- 4738 CXC family of chemokines as prognostic or predictive biomarkers and possible drug targets in colorectal cancer  
*Cabrero-de las Heras S, Martínez-Balibrea E*
- 4750 Gut microbiota in common elderly diseases affecting activities of daily living  
*Shimizu Y*

**ORIGINAL ARTICLE****Basic Study**

- 4759 Yiguanjian decoction enhances fetal liver stem/progenitor cell-mediated repair of liver cirrhosis through regulation of macrophage activation state  
*Xu Y, Fan WW, Xu W, Jiang SL, Chen GF, Liu C, Chen JM, Zhang H, Liu P, Mu YP*
- 4773 Ubiquitin-like modifier activating enzyme 2 promotes cell migration and invasion through Wnt/ $\beta$ -catenin signaling in gastric cancer  
*Li J, Sun X, He P, Liu WQ, Zou YB, Wang Q, Meng XW*

**Case Control Study**

- 4787 Mode of delivery by an ulcerative colitis mother in a case of twins: Immunological differences in cord blood and placenta  
*Dunsmore G, Koleva P, Sutton RT, Ambrosio L, Huang V, Elahi S*

**Retrospective Cohort Study**

- 4798 Increased end-stage renal disease risk in patients with inflammatory bowel disease: A nationwide population-based study  
*Park S, Chun J, Han KD, Soh H, Choi K, Kim JH, Lee J, Lee C, Im JP, Kim JS*

**Retrospective Study**

- 4809 Prediction of colorectal tumor grade and invasion depth through narrow-band imaging scoring  
*Maeyama Y, Mitsuyama K, Noda T, Nagata S, Nagata T, Yoshioka S, Yoshida H, Mukasa M, Sumie H, Kawano H, Akiba J, Araki Y, Kakuma T, Tsuruta O, Torimura T*

**SYSTEMATIC REVIEWS**

- 4821 Burden and outcomes for complex perianal fistulas in Crohn's disease: Systematic review  
*Panes J, Reinisch W, Rupniewska E, Khan S, Fornis J, Khalid JM, Bojic D, Patel H*

**ABOUT COVER**

Editorial board member of *World Journal of Gastroenterology*, Kazuaki Inoue, MD, PhD, Associate Professor, Department of Internal Medicine, Division of Gastroenterology, Showa University Fujigaoka Hospital, Yokohama 227-8501, Japan

**AIMS AND SCOPE**

*World Journal of Gastroenterology* (*World J Gastroenterol*, *WJG*, print ISSN 1007-9327, online ISSN 2219-2840, DOI: 10.3748) is a peer-reviewed open access journal. *WJG* was established on October 1, 1995. It is published weekly on the 7<sup>th</sup>, 14<sup>th</sup>, 21<sup>st</sup>, and 28<sup>th</sup> each month. The *WJG* Editorial Board consists of 642 experts in gastroenterology and hepatology from 59 countries.

The primary task of *WJG* is to rapidly publish high-quality original articles, reviews, and commentaries in the fields of gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, hepatobiliary surgery, gastrointestinal oncology, gastrointestinal radiation oncology, gastrointestinal imaging, gastrointestinal interventional therapy, gastrointestinal infectious diseases, gastrointestinal pharmacology, gastrointestinal pathophysiology, gastrointestinal pathology, evidence-based medicine in gastroenterology, pancreatology, gastrointestinal laboratory medicine, gastrointestinal molecular biology, gastrointestinal immunology, gastrointestinal microbiology, gastrointestinal genetics, gastrointestinal translational medicine, gastrointestinal diagnostics, and gastrointestinal therapeutics. *WJG* is dedicated to become an influential and prestigious journal in gastroenterology and hepatology, to promote the development of above disciplines, and to improve the diagnostic and therapeutic skill and expertise of clinicians.

**INDEXING/ABSTRACTING**

*World Journal of Gastroenterology* (*WJG*) is now indexed in Current Contents®/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports®, Index Medicus, MEDLINE, PubMed, PubMed Central and Directory of Open Access Journals. The 2018 edition of Journal Citation Reports® cites the 2017 impact factor for *WJG* as 3.300 (5-year impact factor: 3.387), ranking *WJG* as 35<sup>th</sup> among 80 journals in gastroenterology and hepatology (quartile in category Q2).

**EDITORS FOR THIS ISSUE**

**Responsible Assistant Editor:** Xiang Li  
**Responsible Electronic Editor:** Yan Huang  
**Proofing Editor-in-Chief:** Lian-Sheng Ma

**Responsible Science Editor:** Rao-Yu Ma  
**Proofing Editorial Office Director:** Ze-Mao Gong

**NAME OF JOURNAL**  
*World Journal of Gastroenterology*

**ISSN**  
ISSN 1007-9327 (print)  
ISSN 2219-2840 (online)

**LAUNCH DATE**  
October 1, 1995

**FREQUENCY**  
Weekly

**EDITORS-IN-CHIEF**  
**Andrzej S Tarnawski, MD, PhD, DSc (Med),**  
**Professor of Medicine, Chief Gastroenterology, VA**  
Long Beach Health Care System, University of California, Irvine, CA, 5901 E. Seventh Str, Long Beach, CA 90822, United States

**EDITORIAL BOARD MEMBERS**  
All editorial board members resources online at <http://www.wjgnet.com/1007-9327/editorialboard.htm>

**EDITORIAL OFFICE**  
Ze-Mao Gong, Director  
*World Journal of Gastroenterology*  
Baishideng Publishing Group Inc  
7901 Stoneridge Drive, Suite 501,  
Pleasanton, CA 94588, USA  
Telephone: +1-925-2238242  
Fax: +1-925-2238243  
E-mail: [editorialoffice@wjgnet.com](mailto:editorialoffice@wjgnet.com)  
Help Desk: <http://www.f6publishing.com/helpdesk>  
<http://www.wjgnet.com>

**PUBLISHER**  
Baishideng Publishing Group Inc  
7901 Stoneridge Drive, Suite 501,  
Pleasanton, CA 94588, USA  
Telephone: +1-925-2238242  
Fax: +1-925-2238243  
E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
Help Desk: <http://www.f6publishing.com/helpdesk>  
<http://www.wjgnet.com>

**PUBLICATION DATE**  
November 14, 2018

**COPYRIGHT**  
© 2018 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

**SPECIAL STATEMENT**  
All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

**INSTRUCTIONS TO AUTHORS**  
Full instructions are available online at <http://www.wjgnet.com/bpg/gerinfo/204>

**ONLINE SUBMISSION**  
<http://www.f6publishing.com>

## Gut microbiota in common elderly diseases affecting activities of daily living

Yukihiro Shimizu

Yukihiro Shimizu, Gastroenterology center, Nanto Municipal Hospital, Toyama 932-0221, Japan

ORCID number: Yukihiro Shimizu (0000-0002-8216-9269).

Author contributions: Shimizu Y designed the review, summarized the literatures and wrote the manuscript.

Conflict-of-interest statement: No conflict-of interest to declare.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: Yukihiro Shimizu, MD, PhD, Director, Doctor, Gastroenterology Center, Nanto Municipal Hospital, 939 Inami, Nanto City, Toyama 932-0221, Japan. [rsf14240@nifty.com](mailto:rsf14240@nifty.com)  
Telephone: +81-763-821475  
Fax: +81-763-821853

Received: August 27, 2018

Peer-review started: August 27, 2018

First decision: October 9, 2018

Revised: October 13, 2018

Accepted: October 21, 2018

Article in press: October 21, 2018

Published online: November 14, 2018

### Abstract

Gut microbiota are involved in the development or prevention of various diseases such as type 2 diabetes, fatty liver, and malignancy such as colorectal cancer, breast cancer and hepatocellular carcinoma. Alzheimer's

disease, osteoporosis, sarcopenia, atherosclerotic stroke and cardiovascular disease are major diseases associated with decreased activities of daily living (ADL), especially in elderly people. Recent analyses have revealed the importance of gut microbiota in the control of these diseases. The composition or diversity of these microbiota is different between patients with these conditions and healthy controls, and administration of probiotics or prebiotics has been shown effective in the treatment of these diseases. Gut microbiota may affect distant organs through mechanisms that include regulating the absorption of nutrients and/or the production of microbial metabolites, regulating and interacting with the systemic immune system, and translocating bacteria/bacterial products through disrupted mucosal barriers. Thus, the gut microbiota may be important regulators in the development of diseases that affect ADL. Although adequate exercise and proper diet are important for preventing these diseases, their combination with interventions that manipulate the composition and/or diversity of gut microbiota could be a promising strategy for maintaining health condition and preserving ADL. This review thus summarizes current understanding of the role of gut microbiota in the development or prevention of diseases closely associated with the maintenance of ADL.

**Key words:** Probiotics; Prebiotics; Alzheimer's disease; Osteoporosis; Sarcopenia; Gut microbiota; Cardiovascular disease; Cerebrovascular disease

© The Author(s) 2018. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** Gut microbiota involved in the development or prevention of various diseases. The composition of gut microbiota is altered by conditions such as type 2 diabetes, fatty liver, and malignancy such as colorectal cancer, breast cancer and hepatocellular carcinoma. Moreover, gut microbiota have been associated with Alzheimer's disease, osteoporosis, sarcopenia, atherosclerotic stroke

and cardiovascular disease. All of these diseases are major causes of decreased activities of daily living (ADL). This review summarizes current understanding of the role of gut microbiota in the development or prevention of diseases closely associated with the maintenance of ADL.

Shimizu Y. Gut microbiota in common elderly diseases affecting activities of daily living. *World J Gastroenterol* 2018; 24(42): 4750-4758 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v24/i42/4750.htm> DOI: <http://dx.doi.org/10.3748/wjg.v24.i42.4750>

## INTRODUCTION

More than  $> 10^{14}$  microorganisms live in the human gut, with most residing in the ileum and colon. *Firmicutes* and *Bacteroidetes* are the major phyla, followed by *Proteobacteria*, *Actinobacteria*, and *Faecalibacteria*<sup>[1]</sup>. The functions and role of these bacteria in maintaining homeostasis of energy, metabolism, and immunity have been extensively analyzed. Moreover, dysbiosis in the gut has been shown to be associated with various diseases, including obesity, type 2 diabetes, fatty liver, autism, inflammatory bowel diseases and malignancies such as colorectal cancer, breast cancer and hepatocellular carcinoma<sup>[1]</sup>. The repertoire of gut microbiota is thought to be formed within one to three years after birth, but may be altered by foods, stress, treatment with antibiotics and aging.

Several aging-related diseases are causes of decreased activities of daily living (ADL). These diseases, including Alzheimer's disease (AD), osteoporosis, sarcopenia, atherosclerotic stroke and cardiovascular disease, are major causes of being bedridden in Japan<sup>[2]</sup>. Because the population of elderly people has been rapidly increasing, methods are urgently needed to prevent and treat these disorders.

Gut microbiota have been shown to interact with the nervous<sup>[3-7]</sup>, musculoskeletal<sup>[8-11]</sup> and cardiovascular<sup>[12]</sup> systems, suggesting that manipulation of gut microbiota may be a key strategy for the prevention or treatment of disorders associated with reduced ADL. There are three possible mechanisms by which gut microbiota can affect distant organs. The first is by regulating the absorption of nutrients from food or the production of microbial metabolites such as short-chain fatty acids (SCFA) or vitamins; the second is by regulating of the systemic immune system; and the third is by translocating bacteria/bacterial products through disrupted mucosal barriers (Figure 1)<sup>[8]</sup>. This review summarizes current knowledge of the role of gut microbiota in the regulation of the development of AD, osteoporosis, sarcopenia, and cerebro and cardiovascular diseases.

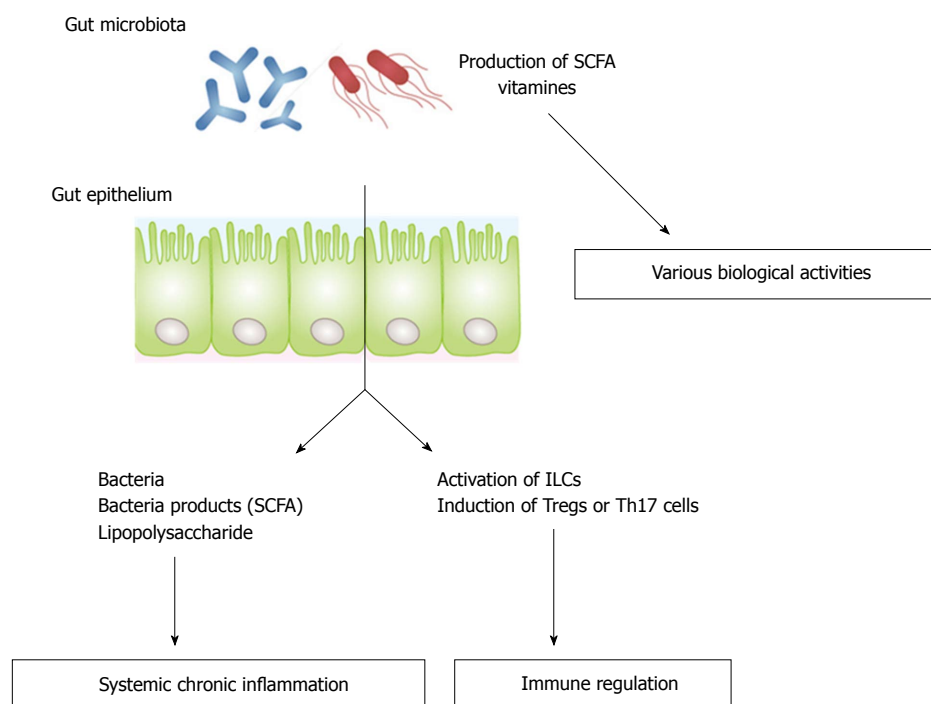
## AD

AD is a neurodegenerative disease and the major cause

of cognitive decline in elderly people. Pathologically, AD is thought to result from Amyloid (A) deposits in the brain. However, neuroinflammatory reactions caused by A deposits, lipopolysaccharide (LPS), bacterial products, or neuroactive molecules may be more important for the development of AD than A deposits itself<sup>[13]</sup>. A microbiota-gut-brain axis has been identified. This axis allows gut microbiota to affect the physiology and pathology of the brain. Intestinal mucosal barriers are occasionally disrupted by pathogenetic bacteria and their products such as LPS, leading to an increase in the permeability of the blood-brain barrier. Thus, bacteria and/or bacterial products from the gut could cause chronic inflammatory responses in the brain, with these responses, in turn, inducing the formation of neurodegenerative lesions of the brain, such as those observed in AD<sup>[14]</sup>. Signaling pathways through the gut-brain axis may also participate in AD<sup>[15]</sup>. Moreover, the immune system may be an important regulator of gut-brain interactions. Gut microbes have been found to influence the maturation and function of microglia, the most abundant type of immune cells residing in the brain<sup>[16]</sup>. Gut microbes have also been shown to modulate activity of astrocytes, which, although they are not brain-resident immune cells, possess immune-related functions such antigen presentation or cytokine production<sup>[17]</sup>. Moreover, microbes modulate the activation and maturation of peripheral immune cells. All of these activities of gut microbiota may be associated with neuroinflammation, brain injury, and autoimmunity<sup>[16]</sup>. Conversely, the brain could regulate the gut through neurologic, immunologic, and hormonal messages<sup>[4]</sup>.

Germ-free (GF) mice have provided important information on the role of the gut microbiota in the homeostasis of brain function. GF mice have been reported to show anxiety-like behavior, which may be due to upregulated expression of postsynaptic density protein 95<sup>[18]</sup>. GF mice were also shown to be deficit in spatial or working memory<sup>[19]</sup>. These data suggest that gut microbiota play a pivotal role in healthy functioning of the brain. However, gut microbiota have also been found to play a negative role. For example, experiments in a mouse model of AD found that A $\beta$  deposition in the brain was lower in GF-A $\beta$  precursor protein APP transgenic mice than in conventionally-raised transgenic APPPS1 mice. In addition, colonization of GF-APPPS1 mice with microbiota showed increased levels of A deposit in the brain, indicating that gut microbiota are involved in A deposits in the brain<sup>[20]</sup>. These seemingly inconsistent results suggest that the composition and/or balance of gut flora may determine the effect of gut microbiota on brain physiology and pathology.

Gut microbiota produce several substances, including SCFA, serotonin, and LPS, which act as important mediators not only of physiological functions but also for pathogenetic agents. Major SCFAs are butyrate, acetate and propionate, with each molecule having specific functions in the gut and/or brain.



**Figure 1** Three possible mechanisms by which gut microbiota can affect distant organs. SCFA: Short chain fatty acids; ILCs: Innate lymphoid cells; Th17: T helper 17.

Butylate was shown to have protective properties in the brain<sup>[21]</sup>, and administration of sodium butylate to a mouse model of AD improved learning and memory function<sup>[22]</sup>. Recently, four cultivable butylate-producing bacteria were isolated from fecal samples from Japanese AD patients, all of which are considered to be unique operational taxonomic units. The analysis for biochemical mechanism of butylate production from those bacteria may contribute the novel approach to stimulate the butylate production in the gut, possibly leading to the improvement of the memory function in patients with AD<sup>[23]</sup>.

Gut microbiota have been shown to produce more than 90% of whole body serotonin, an important metabolite that regulates cognitive ability. Moreover, exogenous serotonin was shown effective in reducing A deposits in the brain<sup>[7]</sup>. GF mice show decreased levels of serum serotonin, indicating that the gut microbiota is major sources of serum serotonin. To date, however, there has been no direct evidence showing that serotonin produced in the gut passes the blood-brain barrier in physiological conditions. Thus, the actual role of gut-derived serotonin in brain function remains unclear. Other gut microbiota metabolites associated with AD include mannitol, succinic acid and 3,4-dihydroxybenzeneacetic acid, all of which are related to AD cognitive decline or susceptibility to AD<sup>[5]</sup>.

No specific gut microbe has been associated to date with AD development. Rather, a recent analysis revealed that reduced diversity of microbes is associated with AD<sup>[6]</sup>. A similar negative correlation between gut microbiota diversity and susceptibility to AD development

was observed in a comparative analysis of gut microbes in subjects from developed and developing countries. Microbial diversity is lower, and AD prevalence higher, in developed than in developing countries<sup>[24]</sup>.

The contribution of gut microbiota to cognitive function was indirectly suggested by the effect of probiotics on AD. Probiotics are microorganisms that maintain or restore beneficial bacteria and maintain a balance between "good" and "bad" bacteria in the gut. Probiotics containing *Lactobacillus helveticus* R0052 and *Bifidobacterium longum* R0175 were found to alleviate cognitive behavioral responses to external stimuli in healthy volunteers<sup>[25]</sup>. Recently, administration of *Bifidobacterium breve* strain A1 to AD model mice was shown to reverse the cognitive dysfunction<sup>[26]</sup>. These findings indicate that changes in the gut microbiota could sufficiently improve cognitive function, and suggest that probiotics may have possible therapeutic potential in patients with AD.

## OSTEOPOROSIS

Osteoporosis is a bone disease characterized by reductions in bone density and quality, and is a major cause of vertebral compression and femoral bone fractures. These fractures often reduce patient ADL or quality of life and may result in patients being bed-ridden. The pathogenesis of the disease is complex, but common processes include elevated bone reabsorption, decreased bone formation, and increased inflammatory reactions in bone.

Links between gut microbiota and bone have been

elucidated mostly from studies in mice. Compared with normal mice, GM mice showed increased trabecular bone volume/tissue volume, with these parameters becoming normalized after colonization<sup>[9]</sup>. The numbers of CD4<sup>+</sup> T cells and osteoclast precursor cells (CD11b<sup>+</sup>/Gr1<sup>-</sup>), and the expression of mRNA encoding the osteolytic cytokine TNF- $\alpha$  are reduced in bone marrow of GM mice, with all of these factors leading to reduced osteoclastogenesis<sup>[9]</sup>.

Gut microbiota were recently shown to increase serum concentrations of insulin-like growth factor-1 (IGF-1), which promotes bone formation and remodeling<sup>[27]</sup>. Depletion of microbiota by the administration of broad-spectrum antibiotics reduced serum IGF-1 levels, a reduction reversed by SCFA supplementation, suggesting that SCFA production by microbiota through the fermentation of dietary fiber may induce expression of IGF-1. These reports indicate that gut microbiota regulate bone metabolism, although their effects may differ according to genetic background, age, and/or sex. The role of gut microbiota in bone metabolism can also be analyzed by assessing the effects of probiotics or prebiotics on bone density. In mouse models, the effects of gut microbiota have been analyzed in male and ovariectomized female mice, the latter being a model of a postmenopausal, estrogen-deficient state. Probiotics such as *Lactobacillus* strains have been shown to increase bone mass along with changes in gut microbiota<sup>[28-32]</sup>. For example, *Lactobacillus ruteri* administered to ovariectomized mice was shown to protect these mice from bone loss, possibly by reducing expression of Trap 5 and receptor activator of NF- $\kappa$ B ligand, which are markers of osteoclast activation and bone resorption, leading to a reduction in osteoclastogenesis. Oral administration of *Bifidobacterium longum* for 16 wk to ovariectomized rats was found to upregulate bone formation, as shown by increases in serum osteocalcin concentrations and osteoblasts, and to downregulate bone absorption, as shown by decreased serum C-terminal telopeptide concentrations and osteoclasts, leading to an increase in bone mass density<sup>[33]</sup>. These effects were mediated by upregulation of the *Bem-2* and *Sparc* genes, the former being a key gene for osteoblast differentiation and the latter being a gene involved in bone calcification.

In addition, several reports have described the effects of prebiotics on bone density. Prebiotics are non-digestible food ingredients that promote the growth of beneficial microorganisms in the gut. Major prebiotics include non-digestible oligosaccharides, such as fructo-oligosaccharides (FOS), galacto-oligosaccharides (GOS), xylo-oligosaccharides, and inulin, with FOS and GOS shown to increase *bifidobacteria* in the intestine<sup>[34,35]</sup>. Administration of GOS or inulin to healthy male rats was shown to increase calcium absorption from the intestine, leading to an increase in bone mineral density<sup>[36,37]</sup>. FOS and inulin were also shown to enhance intestinal calcium absorption in gastrectomized male rats<sup>[38,39]</sup>

and ovariectomized female rats<sup>[40]</sup>. FOS and GOS were shown to increase bone strength by enhancing calcium absorption<sup>[34,35,41]</sup>. The main mechanism by which prebiotics increase calcium absorption from the intestine may involve the production of SCFA by microbiota through fermentation of the prebiotics, with SCFA shown to directly stimulate calcium absorption by gut epithelium<sup>[35]</sup>. Collectively, supplementation with probiotics or prebiotics may be a potential therapeutic intervention for the prevention or treatment of osteoporosis in humans. However, there is limited evidence showing that prebiotics are effective in patients with postmenopausal or senile osteoporosis, although one study reported that supplementation with calcium and FOS had beneficial therapeutic effects on women with postmenopausal osteoporosis<sup>[42]</sup>.

## SARCOPENIA

Sarcopenia is a condition in which muscle mass reduction is accompanied by reduced muscle performance. Causes of muscle weakness include aging, lack of physical exercise, malnutrition, and complications of malignancy. Mitochondrial dysfunction, accompanied by down-regulation of mitochondrial gene expression, has been observed in muscles of pre-frail elderly individuals<sup>[43]</sup>, suggesting that activation of mitochondrial biogenesis may be important in preventing sarcopenia in frail elderly individuals.

Recent evidence has shown that gut microbiota influence skeletal muscle. Microbial metabolites from the intestines have been shown to act as nutrients or metabolic modulators in muscles. These metabolites include folate, vitamin B12, and tryptophan; bacteria involved in the production of each nutrient or metabolite have been identified<sup>[12]</sup>. Possible effects are thought to include biosynthesis of amino acids, DNA synthesis or methylation, prevention of oxidative stress or endothelial damage, and stimulation of anabolism or cell proliferation *via* IGF-1 synthesis<sup>[44-47]</sup>. Other metabolites include SCFAs, which target muscle mitochondria, leading to the promotion of mitochondrial biogenesis through binding to fatty acid receptors 2 and 3 in skeletal muscle cells<sup>[48,49]</sup>. Among the SCFAs, butyrate has been shown to have the most prominent effect on skeletal muscles. Butyrate may help prevent muscle loss and maintain muscle mass through anti-inflammatory effects and activation of regulatory pathways, resulting in increased ATP production and suppression of muscle protein catabolism and apoptosis<sup>[50,51]</sup>. Another microbial metabolite, urolithin A, was recently shown to preserve the biogenesis of skeletal muscle cell mitochondria and to improve exercise capacity in mice and rats<sup>[52]</sup>. Moreover, a randomized, double-blind, placebo-controlled trial demonstrated that oral intake of urolithin A modulated muscle and mitochondrial biomarkers<sup>[12]</sup>.

The composition of gut microbiota characteristic of

sarcopenic patients has not been determined. However, the effects of probiotics on sarcopenia have been analyzed. Supplementation with *Lactobacillus* species was found to prevent muscle mass reduction in mice with acute leukemia<sup>[53]</sup>. In humans, treatment of elderly patients with prebiotics containing FOS and inulin for 13 wk improved muscle function, as shown by decreases in exhaustion and improved handgrip strength<sup>[54]</sup>. Taken together, these findings indicate that muscle mass and functions are closely associated with the composition of gut microbiota.

## ATHEROSCLEROTIC CARDIOVASCULAR DISEASE AND STROKE

Atherosclerosis is a cause of cardio- and cerebrovascular diseases, which are often lethal or linked to serious conditions leading to decreased quality of life. The composition of gut microbiota in patients with atherosclerotic diseases has been reported altered. For example, an analysis of gut microbiota in patients with symptomatic atherosclerosis and healthy controls showed that *Collinsella* was reduced in the patients, while *Roseburia* and *Eubacterium* were enriched in healthy persons<sup>[55]</sup>. In contrast, an analysis of gut microbiota in patients with atherosclerotic cardiovascular disease and healthy subjects found that *Enterobacteriaceae* and *Streptococcus spp.* were enriched in the patients, with the streptococci latter being oral bacteria shown to positively correlate with diastolic and systolic blood pressure<sup>[56]</sup>. Another report from Japan revealed that bacteria of the order *Lactobacillales*, such as *Lactobacillus*, *Streptococcus*, and *Enterococcus*, were significantly increased, while the phylum *Bacteroidetes* (*Bacteroides* and *Prevotella*) was significantly decreased, in patients with cardiovascular disease compared with healthy controls. Patients with atherosclerotic stroke or transient ischemic attack showed dysbiosis of gut microbiota, with an increase in opportunistic bacteria such as *Enterobacter*, *Megasphaera*, *Oscillibacter* and *Desulfovibrio* and a decrease in commensal bacteria such as *Bacteroides*, *Prevotella* and *Fecalibacterium*, with these changes correlating with disease severity<sup>[57]</sup>. Transplantation of gut microbiota in mice was found to result in the transmission of susceptibility to atherosclerosis, directly demonstrating the participation of gut microbiota in atherosclerosis<sup>[58]</sup>.

Several mechanisms have been proposed by which gut microbiota regulate the development of atherosclerotic disease. For example, microbiota may enhance lipid metabolism. GM mice showed decreased lipolysis<sup>[59]</sup> and *Bifidobacteria* was shown to reduce cholesterol levels<sup>[60]</sup>. Another mechanism is associated with the ability of healthy gut microbiota to maintain gut permeability by reinforcing tight junctions of intestinal epithelium. Gut dysbiosis disrupts these tight junctions, increasing intestinal permeability and increasing the

absorption of endotoxin, especially LPS, from the gut. These endotoxins can then the systemic circulation, causing chronic systemic inflammation, including inflammation of the cardiovascular system.

In addition to gut microbiota playing a protective role in the development of cardiovascular disease, gut microbiota may also play a promotional role in the development of this disease. Gut microbiota produce trimethylamine (TMA) from dietary choline and L-carnitine, with TMA subsequently oxidized in the liver, forming trimethylamine-N-oxides (TMAOs). TMAOs have been shown to induce macrophage foam cell formation and plaque formation in the aorta and coronary arteries<sup>[61]</sup>, with high levels of TMAO in patients showing a positive correlation with the incidence of death from cardiovascular disease or myocardial infarction<sup>[62]</sup>.

Hypertension is a major risk factor for atherosclerosis and has been associated with gut dysbiosis in animal models<sup>[12]</sup>. Administration of a combination of antibiotics has been shown to lower blood pressure in treatment-resistant patients with hypertension<sup>[63]</sup>. Moreover, hypertension, often observed in obese pregnant women, was shown to be associated with alterations in the composition of gut microbiota, with an abundance of the butyrate-producing genus *Odoribacter* associated with lower blood pressure<sup>[64]</sup>. These findings indicate that gut microbiota regulate blood pressure in humans, perhaps mediated by an interaction between SCFA and G-protein-coupled receptors, including Gpr41 and Olfr78<sup>[65]</sup>.

A recent report using a mouse model of experimental transient cerebral ischemia induced by occlusion of middle cerebral artery showed that the composition of microbiota in young mice changed to that similar to uninjured aged mice after stroke, and transplantation of fecal gavage from young mice into those mice demonstrated beneficial effect on the outcome of stroke with less infarct size and less mortality. One possible mechanism of the effect could be due to increased SCFA production from youthful microbiota, which was accompanied by lower inflammatory cytokine levels in the plasma. The report clearly indicate that gut microbiota could modify the outcome of stroke, and intervention with youthful microbiota may have a therapeutic potential for the disease<sup>[66]</sup>.

Probiotics have also been reported to affect blood pressure. Administration of *Lactobacillus plantarum* was shown to reduce the severity of myocardial infarction in a rat model<sup>[67]</sup>, an effect thought to be associated with intestinal microbial metabolites<sup>[68]</sup>. In addition, administration of *Lactobacillus rhamnosus* GR-1 induced significant attenuation of left ventricular hypertrophy and improved systolic and diastolic left ventricular function in a rat experimental model of myocardial infarction<sup>[69]</sup>. Collectively, these data suggest a therapeutic strategy with modulation of gut microbiota can be used to prevent or treat cerebro- and cardiovascular diseases caused by atherosclerosis.

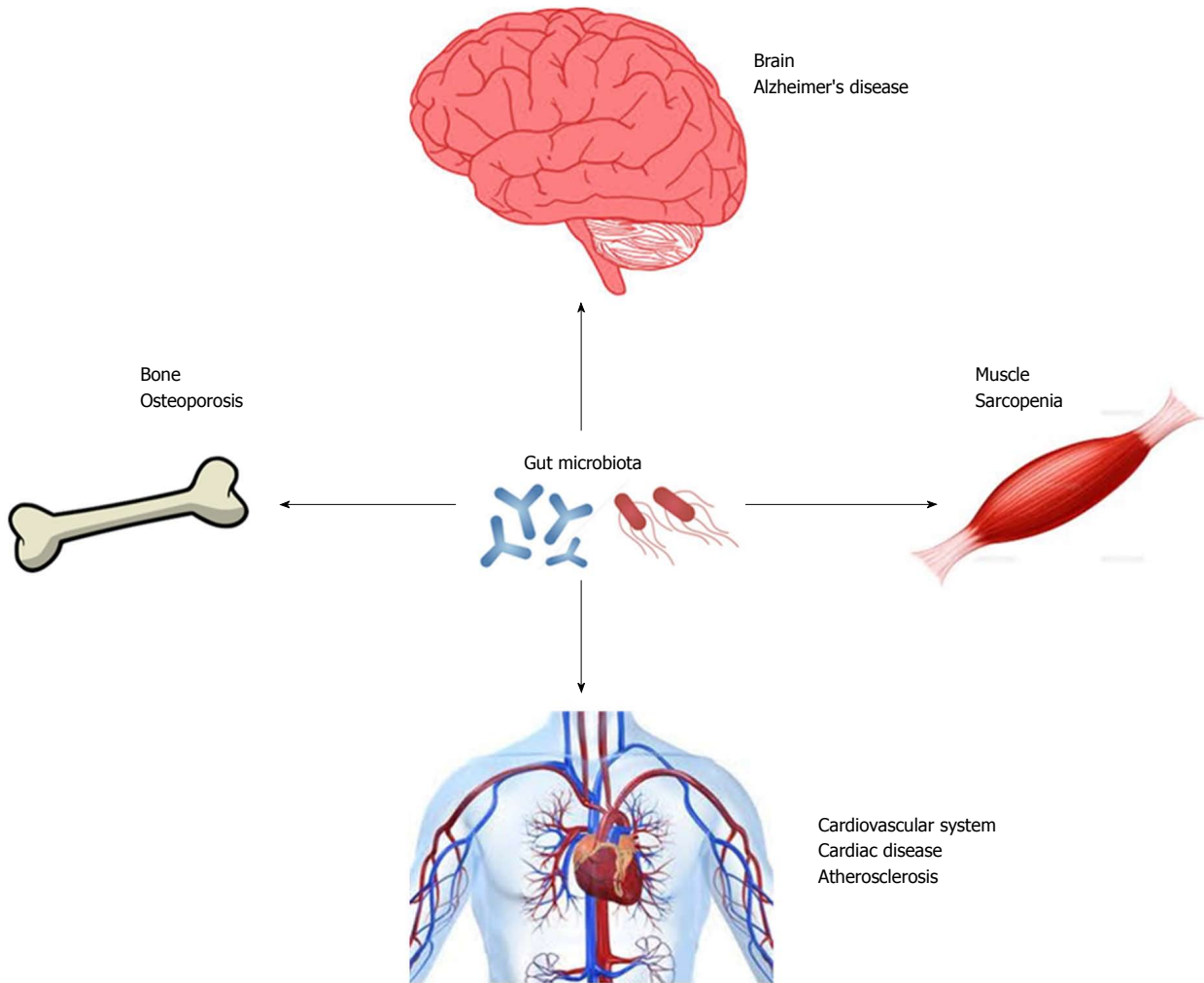


Figure 2 Gut microbiota may be important regulators in development of diseases that may affect activities of daily living.

## CONCLUSION

Gut microbiota may be important regulators in the development of diseases that affect ADL (Figure 2). Although, adequate exercise and a proper diet are important for the prevention of these diseases, the combination of these lifestyle interventions with methods that manipulate the composition and/or diversity of gut microbiota may be a promising strategy for maintenance of healthy conditions with preserved ADL.

## REFERENCES

- 1 Minemura M, Shimizu Y. Gut microbiota and liver diseases. *World J Gastroenterol* 2015; **21**: 1691-1702 [PMID: 25684933 DOI: 10.3748/wjg.v21.i6.1691]
- 2 Yoshino H, Sakurai T, Hasegawa K, Yokono K. Causes of decreased activity of daily life in elderly patients who need daily living care. *Geriatr Gerontol Int* 2011; **11**: 297-303 [PMID: 21272178 DOI: 10.1111/j.1447-0594.2010.00683.x]
- 3 Sochocka M, Donskow-Lysoniewska K, Diniz BS, Kurpas D, Brzozowska E, Leszek J. The Gut Microbiome Alterations and Inflammation-Driven Pathogenesis of Alzheimer's Disease-a Critical Review. *Mol Neurobiol* 2018 [PMID: 29936690 DOI: 10.1007/s12035-018-1188-4]
- 4 Choi J, Hur TY, Hong Y. Influence of Altered Gut Microbiota Composition on Aging and Aging-Related Diseases. *J Lifestyle Med* 2018; **8**: 1-7 [PMID: 29581954 DOI: 10.15280/jlm.2018.8.1.1]
- 5 Xu R, Wang Q. Towards understanding brain-gut-microbiome connections in Alzheimer's disease. *BMC Syst Biol* 2016; **10** Suppl 3: 63 [PMID: 27585440 DOI: 10.1186/s12918-016-0307-y]
- 6 Vogt NM, Kerby RL, Dill-McFarland KA, Harding SJ, Merluzzi AP, Johnson SC, Carlsson CM, Asthana S, Zetterberg H, Blennow K, Bendlin BB, Rey FE. Gut microbiome alterations in Alzheimer's disease. *Sci Rep* 2017; **7**: 13537 [PMID: 29051531 DOI: 10.1038/s41598-017-13601-y]
- 7 Hu X, Wang T, Jin F. Alzheimer's disease and gut microbiota. *Sci China Life Sci* 2016; **59**: 1006-1023 [PMID: 27566465 DOI: 10.1007/s11427-016-5083-9]
- 8 Hernandez CJ, Guss JD, Luna M, Goldring SR. Links Between the Microbiome and Bone. *J Bone Miner Res* 2016; **31**: 1638-1646 [PMID: 27317164 DOI: 10.1002/jbmr.2887]
- 9 Sjögren K, Engdahl C, Henning P, Lerner UH, Tremaroli V, Lagerquist MK, Bäckhed F, Ohlsson C. The gut microbiota regulates bone mass in mice. *J Bone Miner Res* 2012; **27**: 1357-1367 [PMID: 22407806 DOI: 10.1002/jbmr.1588]
- 10 Steves CJ, Bird S, Williams FM, Spector TD. The Microbiome and Musculoskeletal Conditions of Aging: A Review of Evidence for Impact and Potential Therapeutics. *J Bone Miner Res* 2016; **31**: 261-269 [PMID: 26676797 DOI: 10.1002/jbmr.2765]
- 11 Ticinesi A, Lauretani F, Milani C, Nouvenne A, Tana C, Del Rio D, Maggio M, Ventura M, Meschi T. Aging Gut Microbiota at the

- Cross-Road between Nutrition, Physical Frailty, and Sarcopenia: Is There a Gut-Muscle Axis? *Nutrients* 2017; **9** [PMID: 29189738 DOI: 10.3390/nu9121303]
- 12 **Tang WH**, Kitai T, Hazen SL. Gut Microbiota in Cardiovascular Health and Disease. *Circ Res* 2017; **120**: 1183-1196 [PMID: 28360349 DOI: 10.1161/CIRCRESAHA.117.309715]
  - 13 **Bronzuoli MR**, Iacomino A, Steardo L, Scuderi C. Targeting neuroinflammation in Alzheimer's disease. *J Inflamm Res* 2016; **9**: 199-208 [PMID: 27843334 DOI: 10.2147/JIR.S86958]
  - 14 **Feng Q**, Chen WD, Wang YD. Gut Microbiota: An Integral Moderator in Health and Disease. *Front Microbiol* 2018; **9**: 151 [PMID: 29515527 DOI: 10.3389/fmicb.2018.00151]
  - 15 **Köhler CA**, Maes M, Slyepchenko A, Berk M, Solmi M, Lanctôt KL, Carvalho AF. The Gut-Brain Axis, Including the Microbiome, Leaky Gut and Bacterial Translocation: Mechanisms and Pathophysiological Role in Alzheimer's Disease. *Curr Pharm Des* 2016; **22**: 6152-6166 [PMID: 27604604 DOI: 10.2174/1381612822666160907093807]
  - 16 **Fung TC**, Olson CA, Hsiao EY. Interactions between the microbiota, immune and nervous systems in health and disease. *Nat Neurosci* 2017; **20**: 145-155 [PMID: 28092661 DOI: 10.1038/nn.4476]
  - 17 **Dong Y**, Benveniste EN. Immune function of astrocytes. *Glia* 2001; **36**: 180-190 [PMID: 11596126 DOI: 10.1002/glia.1107]
  - 18 **Diaz Heijtz R**, Wang S, Anuar F, Qian Y, Björkholm B, Samuelsson A, Hibberd ML, Forssberg H, Pettersson S. Normal gut microbiota modulates brain development and behavior. *Proc Natl Acad Sci USA* 2011; **108**: 3047-3052 [PMID: 21282636 DOI: 10.1073/pnas.1010529108]
  - 19 **Gareau MG**, Wine E, Rodrigues DM, Cho JH, Whary MT, Philpott DJ, Macqueen G, Sherman PM. Bacterial infection causes stress-induced memory dysfunction in mice. *Gut* 2011; **60**: 307-317 [PMID: 20966022 DOI: 10.1136/gut.2009.202515]
  - 20 **Harach T**, Marungruang N, Duthilleul N, Cheatham V, Mc Coy KD, Frisoni G, Neher JJ, Fåk F, Jucker M, Lasser T, Bolmont T. Reduction of Abeta amyloid pathology in APPPS1 transgenic mice in the absence of gut microbiota. *Sci Rep* 2017; **7**: 41802 [PMID: 28176819 DOI: 10.1038/srep41802]
  - 21 **Bourassa MW**, Alim I, Bultman SJ, Ratan RR. Butyrate, neuroepigenetics and the gut microbiome: Can a high fiber diet improve brain health? *Neurosci Lett* 2016; **625**: 56-63 [PMID: 26868600 DOI: 10.1016/j.neulet.2016.02.009]
  - 22 **Govindarajan N**, Agis-Balboa RC, Walter J, Sananbenesi F, Fischer A. Sodium butyrate improves memory function in an Alzheimer's disease mouse model when administered at an advanced stage of disease progression. *J Alzheimers Dis* 2011; **26**: 187-197 [PMID: 21593570 DOI: 10.3233/JAD-2011-110080]
  - 23 **Nguyen TTT**, Fujimura Y, Mimura I, Fujii Y, Nguyen NL, Arakawa K, Morita H. Cultivable butyrate-producing bacteria of elderly Japanese diagnosed with Alzheimer's disease. *J Microbiol* 2018; **56**: 760-771 [PMID: 30136260 DOI: 10.1007/s12275-018-8297-7]
  - 24 **Fox M**, Knapp LA, Andrews PW, Fincher CL. Hygiene and the world distribution of Alzheimer's disease: Epidemiological evidence for a relationship between microbial environment and age-adjusted disease burden. *Evol Med Public Health* 2013; **2013**: 173-186 [PMID: 24481197 DOI: 10.1093/emph/eot015]
  - 25 **Messaoudi M**, Violle N, Bisson JF, Desor D, Javelot H, Rougeot C. Beneficial psychological effects of a probiotic formulation (*Lactobacillus helveticus* R0052 and *Bifidobacterium longum* R0175) in healthy human volunteers. *Gut Microbes* 2011; **2**: 256-261 [PMID: 21983070 DOI: 10.4161/gmic.2.4.16108]
  - 26 **Kobayashi Y**, Sugahara H, Shimada K, Mitsuyama E, Kuhara T, Yasuoka A, Kondo T, Abe K, Xiao JZ. Therapeutic potential of *Bifidobacterium breve* strain A1 for preventing cognitive impairment in Alzheimer's disease. *Sci Rep* 2017; **7**: 13510 [PMID: 29044140 DOI: 10.1038/s41598-017-13368-2]
  - 27 **Yan J**, Herzog JW, Tsang K, Brennan CA, Bower MA, Garrett WS, Sartor BR, Aliprantis AO, Charles JF. Gut microbiota induce IGF-1 and promote bone formation and growth. *Proc Natl Acad Sci USA* 2016; **113**: E7554-E7563 [PMID: 27821775 DOI: 10.1073/pnas.1607235113]
  - 28 **McCabe LR**, Irwin R, Schaefer L, Britton RA. Probiotic use decreases intestinal inflammation and increases bone density in healthy male but not female mice. *J Cell Physiol* 2013; **228**: 1793-1798 [PMID: 23389860 DOI: 10.1002/jcp.24340]
  - 29 **Chiang SS**, Pan TM. Antiosteoporotic effects of *Lactobacillus*-fermented soy skim milk on bone mineral density and the microstructure of femoral bone in ovariectomized mice. *J Agric Food Chem* 2011; **59**: 7734-7742 [PMID: 21668014 DOI: 10.1021/jf2013716]
  - 30 **Rodrigues FC**, Castro AS, Rodrigues VC, Fernandes SA, Fontes EA, de Oliveira TT, Martino HS, de Lucena Fortes Ferreira CL. Yacon flour and *Bifidobacterium longum* modulate bone health in rats. *J Med Food* 2012; **15**: 664-670 [PMID: 22510044 DOI: 10.1089/jmf.2011.0296]
  - 31 **Britton RA**, Irwin R, Quach D, Schaefer L, Zhang J, Lee T, Parameswaran N, McCabe LR. Probiotic *L. reuteri* treatment prevents bone loss in a menopausal ovariectomized mouse model. *J Cell Physiol* 2014; **229**: 1822-1830 [PMID: 24677054 DOI: 10.1002/jcp.24636]
  - 32 **Ohlsson C**, Engdahl C, Fåk F, Andersson A, Windahl SH, Farman HH, Movérare-Skrtic S, Islander U, Sjögren K. Probiotics protect mice from ovariectomy-induced cortical bone loss. *PLoS One* 2014; **9**: e92368 [PMID: 24637895 DOI: 10.1371/journal.pone.0092368]
  - 33 **Parvaneh K**, Ebrahimi M, Sabran MR, Karimi G, Hwei AN, Abdul-Majeed S, Ahmad Z, Ibrahim Z, Jamaluddin R. Probiotics (*Bifidobacterium longum*) Increase Bone Mass Density and Upregulate Sparc and Bmp-2 Genes in Rats with Bone Loss Resulting from Ovariectomy. *Biomed Res Int* 2015; **2015**: 897639 [PMID: 26366421 DOI: 10.1155/2015/897639]
  - 34 **Weaver CM**, Martin BR, Nakatsu CH, Armstrong AP, Clavijo A, McCabe LD, McCabe GP, Duignan S, Schoterman MH, van den Heuvel EG. Galactooligosaccharides improve mineral absorption and bone properties in growing rats through gut fermentation. *J Agric Food Chem* 2011; **59**: 6501-6510 [PMID: 21553845 DOI: 10.1021/jf2009777]
  - 35 **Weaver CM**. Diet, gut microbiome, and bone health. *Curr Osteoporos Rep* 2015; **13**: 125-130 [PMID: 25616772 DOI: 10.1007/s11914-015-0257-0]
  - 36 **Chonan O**, Watanuki M. Effect of galactooligosaccharides on calcium absorption in rats. *J Nutr Sci Vitaminol (Tokyo)* 1995; **41**: 95-104 [PMID: 7616330 DOI: 10.3177/jnsv.41.95]
  - 37 **Roberfroid MB**, Cumps J, Devogelaer JP. Dietary chicory inulin increases whole-body bone mineral density in growing male rats. *J Nutr* 2002; **132**: 3599-3602 [PMID: 12468594 DOI: 10.1093/jn/132.12.3599]
  - 38 **Ohta A**, Motohashi Y, Sakai K, Hirayama M, Adachi T, Sakuma K. Dietary fructooligosaccharides increase calcium absorption and levels of mucosal calbindin-D9k in the large intestine of gastrectomized rats. *Scand J Gastroenterol* 1998; **33**: 1062-1068 [PMID: 9829361 DOI: 10.1080/003655298750026769]
  - 39 **Shiga K**, Nishimukai M, Tomita F, Hara H. Ingestion of difructose anhydride III, a non-digestible disaccharide, improves postgastroctomy osteopenia in rats. *Scand J Gastroenterol* 2006; **41**: 1165-1173 [PMID: 16990201 DOI: 10.1080/00365520600575753]
  - 40 **Zafar TA**, Weaver CM, Zhao Y, Martin BR, Wastney ME. Nondigestible oligosaccharides increase calcium absorption and suppress bone resorption in ovariectomized rats. *J Nutr* 2004; **134**: 399-402 [PMID: 14747679 DOI: 10.1093/jn/134.2.399]
  - 41 **Mathey J**, Puel C, Kati-Coulibaly S, Bennetau-Pelissero C, Davicco MJ, Lebecque P, Horcajada MN, Coxam V. Fructooligosaccharides maximize bone-sparing effects of soy isoflavone-enriched diet in the ovariectomized rat. *Calcif Tissue Int* 2004; **75**: 169-179 [PMID: 15164148 DOI: 10.1007/s00223-004-0128-7]
  - 42 **Slevin MM**, Allsopp PJ, Magee PJ, Bonham MP, Naughton VR, Strain JJ, Duffy ME, Wallace JM, Mc Sorley EM. Supplementation with calcium and short-chain fructo-oligosaccharides affects

- markers of bone turnover but not bone mineral density in postmenopausal women. *J Nutr* 2014; **144**: 297-304 [PMID: 24453130 DOI: 10.3945/jn.113.188144]
- 43 **St-Jean-Pelletier F**, Pion CH, Leduc-Gaudet JP, Sgarioni N, Zovilé I, Barbat-Artigas S, Reynaud O, Alkaterji F, Lemieux FC, Grenon A, Gaudreau P, Hepple RT, Chevalier S, Belanger M, Morais JA, Aubertin-Leheudre M, Gouspillou G. The impact of ageing, physical activity, and pre-frailty on skeletal muscle phenotype, mitochondrial content, and intramyocellular lipids in men. *J Cachexia Sarcopenia Muscle* 2017; **8**: 213-228 [PMID: 27897402 DOI: 10.1002/jcsm.12139]
  - 44 **LeBlanc JG**, Milani C, de Giori GS, Sesma F, van Sinderen D, Ventura M. Bacteria as vitamin suppliers to their host: a gut microbiota perspective. *Curr Opin Biotechnol* 2013; **24**: 160-168 [PMID: 22940212 DOI: 10.1016/j.copbio.2012.08.005]
  - 45 **Kuo HK**, Liao KC, Leveille SG, Bean JF, Yen CJ, Chen JH, Yu YH, Tai TY. Relationship of homocysteine levels to quadriceps strength, gait speed, and late-life disability in older adults. *J Gerontol A Biol Sci Med Sci* 2007; **62**: 434-439 [PMID: 17452739 DOI: 10.1093/gerona/62.4.434]
  - 46 **Lin R**, Liu W, Piao M, Zhu H. A review of the relationship between the gut microbiota and amino acid metabolism. *Amino Acids* 2017; **49**: 2083-2090 [PMID: 28932911 DOI: 10.1007/s00726-017-2493-3]
  - 47 **Dukes A**, Davis C, El Refaey M, Upadhyay S, Mork S, Arounleut P, Johnson MH, Hill WD, Isaacs CM, Hamrick MW. The aromatic amino acid tryptophan stimulates skeletal muscle IGF1/p70s6k/mTOR signaling in vivo and the expression of myogenic genes in vitro. *Nutrition* 2015; **31**: 1018-1024 [PMID: 26059377 DOI: 10.1016/j.nut.2015.02.011]
  - 48 **den Besten G**, Lange K, Havinga R, van Dijk TH, Gerding A, van Eunen K, Müller M, Groen AK, Hooiveld GJ, Bakker BM, Reijngoud DJ. Gut-derived short-chain fatty acids are vividly assimilated into host carbohydrates and lipids. *Am J Physiol Gastrointest Liver Physiol* 2013; **305**: G900-G910 [PMID: 24136789 DOI: 10.1152/ajpgi.00265.2013]
  - 49 **den Besten G**, van Eunen K, Groen AK, Venema K, Reijngoud DJ, Bakker BM. The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. *J Lipid Res* 2013; **54**: 2325-2340 [PMID: 23821742 DOI: 10.1194/jlr.R036012]
  - 50 **Walsh ME**, Bhattacharya A, Sataranatarajan K, Qaisar R, Sloane L, Rahman MM, Kinter M, Van Remmen H. The histone deacetylase inhibitor butyrate improves metabolism and reduces muscle atrophy during aging. *Aging Cell* 2015; **14**: 957-970 [PMID: 26290460 DOI: 10.1111/acer.12387]
  - 51 **den Besten G**, Gerding A, van Dijk TH, Ciapaite J, Bleeker A, van Eunen K, Havinga R, Groen AK, Reijngoud DJ, Bakker BM. Protection against the Metabolic Syndrome by Guar Gum-Derived Short-Chain Fatty Acids Depends on Peroxisome Proliferator-Activated Receptor  $\gamma$  and Glucagon-Like Peptide-1. *PLoS One* 2015; **10**: e0136364 [PMID: 26292284 DOI: 10.1371/journal.pone.0136364]
  - 52 **Ryu D**, Mouchiroud L, Andreux PA, Katsyuba E, Moullan N, Nicolet-Dit-Félix AA, Williams EG, Jha P, Lo Sasso G, Huzard D, Aebischer P, Sandi C, Rinsch C, Auwerx J. Urolithin A induces mitophagy and prolongs lifespan in *C. elegans* and increases muscle function in rodents. *Nat Med* 2016; **22**: 879-888 [PMID: 27400265 DOI: 10.1038/nm.4132]
  - 53 **Bindels LB**, Beck R, Schakman O, Martin JC, De Backer F, Sohet FM, Dewulf EM, Pachikian BD, Neyrinck AM, Thissen JP, Verrax J, Calderon PB, Pot B, Granelle C, Cani PD, Scott KP, Delzenne NM. Restoring specific lactobacilli levels decreases inflammation and muscle atrophy markers in an acute leukemia mouse model. *PLoS One* 2012; **7**: e37971 [PMID: 22761662 DOI: 10.1371/journal.pone.0037971]
  - 54 **Buigues C**, Fernández-Garrido J, Pruimboom L, Hoogland AJ, Navarro-Martínez R, Martínez-Martínez M, Verdejo Y, Mascarós MC, Peris C, Cauli O. Effect of a Prebiotic Formulation on Frailty Syndrome: A Randomized, Double-Blind Clinical Trial. *Int J Mol Sci* 2016; **17** [PMID: 27314331 DOI: 10.3390/ijms17060932]
  - 55 **Karlsson FH**, Fåk F, Nookaew I, Tremaroli V, Fagerberg B, Petranovic D, Bäckhed F, Nielsen J. Symptomatic atherosclerosis is associated with an altered gut metagenome. *Nat Commun* 2012; **3**: 1245 [PMID: 23212374 DOI: 10.1038/ncomms2266]
  - 56 **Jie Z**, Xia H, Zhong SL, Feng Q, Li S, Liang S, Zhong H, Liu Z, Gao Y, Zhao H, Zhang D, Su Z, Fang Z, Lan Z, Li J, Xiao L, Li J, Li R, Li X, Li F, Ren H, Huang Y, Peng Y, Li G, Wen B, Dong B, Chen JY, Geng QS, Zhang ZW, Yang H, Wang J, Wang J, Zhang X, Madsen L, Brix S, Ning G, Xu X, Liu X, Hou Y, Jia H, He K, Kristiansen K. The gut microbiome in atherosclerotic cardiovascular disease. *Nat Commun* 2017; **8**: 845 [PMID: 29018189 DOI: 10.1038/s41467-017-00900-1]
  - 57 **Yin J**, Liao SX, He Y, Wang S, Xia GH, Liu FT, Zhu JJ, You C, Chen Q, Zhou L, Pan SY, Zhou HW. Dysbiosis of Gut Microbiota With Reduced Trimethylamine-N-Oxide Level in Patients With Large-Artery Atherosclerotic Stroke or Transient Ischemic Attack. *J Am Heart Assoc* 2015; **4** [PMID: 26597155 DOI: 10.1161/JAHA.115.002699]
  - 58 **Gregory JC**, Buffa JA, Org E, Wang Z, Levison BS, Zhu W, Wagner MA, Bennett BJ, Li L, DiDonato JA, Lusis AJ, Hazen SL. Transmission of atherosclerosis susceptibility with gut microbial transplantation. *J Biol Chem* 2015; **290**: 5647-5660 [PMID: 25550161 DOI: 10.1074/jbc.M114.618249]
  - 59 **Bäckhed F**, Ding H, Wang T, Hooper LV, Koh GY, Nagy A, Semenkovich CF, Gordon JI. The gut microbiota as an environmental factor that regulates fat storage. *Proc Natl Acad Sci USA* 2004; **101**: 15718-15723 [PMID: 15505215 DOI: 10.1073/pnas.0407076101]
  - 60 **Tahri K**, Crociani J, Ballongue J, Schneider F. Effects of three strains of bifidobacteria on cholesterol. *Lett Appl Microbiol* 1995; **21**: 149-151 [PMID: 7576497 DOI: 10.1111/j.1472-765X.1995.tb01028.x]
  - 61 **Wang Z**, Klipfell E, Bennett BJ, Koeth R, Levison BS, Dugar B, Feldstein AE, Britt EB, Fu X, Chung YM, Wu Y, Schauer P, Smith JD, Allayee H, Tang WH, DiDonato JA, Lusis AJ, Hazen SL. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature* 2011; **472**: 57-63 [PMID: 21475195 DOI: 10.1038/nature09922]
  - 62 **Tang WH**, Wang Z, Levison BS, Koeth RA, Britt EB, Fu X, Wu Y, Hazen SL. Intestinal microbial metabolism of phosphatidylcholine and cardiovascular risk. *N Engl J Med* 2013; **368**: 1575-1584 [PMID: 23614584 DOI: 10.1056/NEJMoa1109400]
  - 63 **Qi Y**, Aranda JM, Rodriguez V, Raizada MK, Pepine CJ. Impact of antibiotics on arterial blood pressure in a patient with resistant hypertension - A case report. *Int J Cardiol* 2015; **201**: 157-158 [PMID: 26301638 DOI: 10.1016/j.ijcard.2015.07.078]
  - 64 **Gomez-Arango LF**, Barrett HL, McIntyre HD, Callaway LK, Morrison M, Dekker Nitert M; SPRING Trial Group. Increased Systolic and Diastolic Blood Pressure Is Associated With Altered Gut Microbiota Composition and Butyrate Production in Early Pregnancy. *Hypertension* 2016; **68**: 974-981 [PMID: 27528065 DOI: 10.1161/HYPERTENSIONAHA.116.07910]
  - 65 **Pluznick JL**. Microbial Short-Chain Fatty Acids and Blood Pressure Regulation. *Curr Hypertens Rep* 2017; **19**: 25 [PMID: 28315048 DOI: 10.1007/s11906-017-0722-5]
  - 66 **Spychala MS**, Venna VR, Jandzinski M, Doran SJ, Durgan DJ, Ganesh BP, Ajami NJ, Putluri N, Graf J, Bryan RM, McCullough LD. Age-related changes in the gut microbiota influence systemic inflammation and stroke outcome. *Ann Neurol* 2018; **84**: 23-36 [PMID: 29733457 DOI: 10.1002/ana.25250]
  - 67 **Lam V**, Su J, Koprowski S, Hsu A, Tweddell JS, Rafiee P, Gross GJ, Salzman NH, Baker JE. Intestinal microbiota determine severity of myocardial infarction in rats. *FASEB J* 2012; **26**: 1727-1735 [PMID: 22247331 DOI: 10.1096/fj.11-197921]
  - 68 **Lam V**, Su J, Hsu A, Gross GJ, Salzman NH, Baker JE. Intestinal Microbial Metabolites Are Linked to Severity of Myocardial

Shimizu Y. Role of gut microbiota in healthy condition

Infarction in Rats. *PLoS One* 2016; **11**: e0160840 [PMID: 27505423 DOI: 10.1371/journal.pone.0160840]

- 69 **Gan XT**, Ettinger G, Huang CX, Burton JP, Haist JV, Rajapurohitam V, Sidaway JE, Martin G, Gloor GB, Swann JR, Reid G, Karmazyn

M. Probiotic administration attenuates myocardial hypertrophy and heart failure after myocardial infarction in the rat. *Circ Heart Fail* 2014; **7**: 491-499 [PMID: 24625365 DOI: 10.1161/CIRCHEARTFAILURE.113.000978]

**P- Reviewer:** Das U, Yu JS **S- Editor:** Wang XJ **L- Editor:** A  
**E- Editor:** Huang Y





Published by **Baishideng Publishing Group Inc**  
7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA  
Telephone: +1-925-223-8242  
Fax: +1-925-223-8243  
E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
Help Desk: <http://www.f6publishing.com/helpdesk>  
<http://www.wjgnet.com>



ISSN 1007-9327

