

World Journal of *Gastroenterology*

World J Gastroenterol 2020 April 7; 26(13): 1382-1545



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INDEXING/ABSTRACTING

The *WJG* is now indexed in Current Contents®/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports®, Index Medicus, MEDLINE, PubMed, PubMed Central, and Scopus. The 2019 edition of Journal Citation Report® cites the 2018 impact factor for *WJG* as 3.411 (5-year impact factor: 3.579), ranking *WJG* as 35th among 84 journals in gastroenterology and hepatology (quartile in category Q2). CiteScore (2018): 3.43.

RESPONSIBLE EDITORS FOR THIS ISSUE

Responsible Electronic Editor: *Yu-Jie Ma*
 Proofing Production Department Director: *Xiang Li*

NAME OF JOURNAL

World Journal of Gastroenterology

ISSN

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

LAUNCH DATE

October 1, 1995

FREQUENCY

Weekly

EDITORS-IN-CHIEF

Subrata Ghosh, Andrzej S Tarnawski

EDITORIAL BOARD MEMBERS

<http://www.wjgnet.com/1007-9327/editorialboard.htm>

EDITORIAL OFFICE

Ze-Mao Gong, Director

PUBLICATION DATE

April 7, 2020

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<https://www.wjgnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

<https://www.wjgnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjgnet.com/bpg/GerInfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>

Basic Study

Calpain-2 activity promotes aberrant endoplasmic reticulum stress-related apoptosis in hepatocytes

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Author contributions: Xie RJ, Han B and Zheng L performed the experiments; Hu XX, Cai S, Chen YS, Yang Y and Yang T analyzed the data; Xie RJ and Yang Q designed the study; Xie RJ wrote the manuscript; all authors approved the final manuscript. Xie RJ and Hu XX contributed equally to this work.

Supported by the National Natural Science Foundation of China, No. 81560105; the Department of Science and Technology of Guizhou Province, No. LH (2014) 7074.

Institutional animal care and use committee statement: All animal experiments conformed to the internationally accepted principles for the care and use of laboratory animals.

Conflict-of-interest statement: The authors declare no competing interests.

Data sharing statement: The datasets in the present study are available upon reasonable request.

ARRIVE guidelines statement: The authors have read the ARRIVE guidelines, and the manuscript was prepared and revised

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Abstract

BACKGROUND

Calpain-2 is a Ca²⁺-dependent cysteine protease, and high calpain-2 activity can enhance apoptosis mediated by multiple triggers.

AIM

To investigate whether calpain-2 can modulate aberrant endoplasmic reticulum (ER) stress-related apoptosis in rat hepatocyte BRL-3A cells.

METHODS

BRL-3A cells were treated with varying doses of dithiothreitol (DTT), and their viability and apoptosis were quantified by 3-[4, 5-dimethyl-2-thiazolyl]-2, 5-diphenyl-2-H-tetrazolium bromide and flow cytometry. The expression of ER stress- and apoptosis-related proteins was detected by Western blot analysis. The protease activity of calpain-2 was determined using a fluorescent substrate, *N*-succinyl-Leu-Leu-Val-Tyr-AMC. Intracellular Ca²⁺ content, and ER and calpain-2 co-localization were characterized by fluorescent microscopy. The impact of calpain-2 silencing by specific small interfering RNA on caspase-12 activation and apoptosis of BRL-3A cells was quantified.

RESULTS

DTT exhibited dose-dependent cytotoxicity against BRL-3A cells and treatment with 2 mmol/L DTT triggered BRL-3A cell apoptosis. DTT treatment significantly upregulated 78 kDa glucose-regulated protein, activating transcription factor 4, C/EBP-homologous protein expression by >2-fold, and

according to the ARRIVE guidelines.

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Manuscript source: Unsolicited manuscript

Received: December 2, 2019

Peer-review started: December 2, 2019

First decision: December 12, 2019

Revised: February 20, 2020

Accepted: March 9, 2020

Article in press: March 9, 2020

Published online: April 7, 2020

P-Reviewer: Aureliano M, Abdalnour-Nakhoul SM, El-Bendary M, Marickar F

S-Editor: Zhang L

L-Editor: Filipodia

E-Editor: Zhang YL



enhanced PRKR-like ER kinase phosphorylation, caspase-12 and caspase-3 cleavage in BRL-3A cells in a trend of time-dependence. DTT treatment also significantly increased intracellular Ca²⁺ content, calpain-2 expression, and activity by >2-fold in BRL-3A cells. Furthermore, immunofluorescence revealed that DTT treatment promoted the ER accumulation of calpain-2. Moreover, calpain-2 silencing to decrease calpain-2 expression by 85% significantly mitigated DTT-enhanced calpain-2 expression, caspase-12 cleavage, and apoptosis in BRL-3A cells.

CONCLUSION

The data indicated that Ca²⁺-dependent calpain-2 activity promoted the aberrant ER stress-related apoptosis of rat hepatocytes by activating caspase-12 in the ER.

Key words: Calcium; Calpain-2; Caspase-12; Endoplasmic reticulum stress; Apoptosis; Hepatocyte

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Core tip: Hepatocyte apoptosis is associated with many liver diseases. During the process of apoptosis, calpain-2 can cleave several apoptosis-related proteins. However, the regulatory mechanisms by which calpain-2 regulates the endoplasmic reticulum (ER) stress-mediated hepatocyte apoptosis remain unclear. In this study, the effect of calpain-2 on ER stress-mediated hepatocyte apoptosis and the underlying regulatory mechanisms was investigated. Our data indicate that calpain-2 is crucial for the aberrant ER stress-induced apoptosis of hepatocytes and may be a novel therapeutic target for liver diseases.

Citation: Xie RJ, Hu XX, Zheng L, Cai S, Chen YS, Yang Y, Yang T, Han B, Yang Q. Calpain-2 activity promotes aberrant endoplasmic reticulum stress-related apoptosis in hepatocytes. *World J Gastroenterol* 2020; 26(13): 1450-1462

URL: <https://www.wjgnet.com/1007-9327/full/v26/i13/1450.htm>

DOI: <https://dx.doi.org/10.3748/wjg.v26.i13.1450>

INTRODUCTION

Hepatocyte apoptosis participates in the pathogenesis of many types of liver diseases. Actually, apoptotic hepatocytes are observed in the liver of patients with acute liver injury, non-alcoholic fatty liver diseases, hepatic fibrosis, and liver cirrhosis^[1-5]. However, the regulatory mechanisms underlying hepatocyte apoptosis remains elusive. It is notable that aberrant endoplasmic reticulum (ER) stress can trigger hepatocyte apoptosis^[6,7]. While the ER is physiologically responsible for the control of protein proper folding and function, many factors such as the unfolded protein response, ER overload response and others can disturb ER function, leading to ER stress^[8,9]. During the process of ER stress, the ER chaperon glucose-regulated protein (GRP78) changes its binding from ER transmembrane protein PRKR-like endoplasmic reticulum kinase (PERK) to unfolded/misfolded proteins to release PERK^[10]. The released PERK is subjected to self-phosphorylation, which promotes the expression of transcription factors of activating transcription factor 4 (ATF4) and C/EBP-homologous protein (CHOP) in cells^[11]. The upregulated ATF4 and CHOP can enter the nucleus to regulate the transcription of related genes^[12]. Such compensative responses may reduce the accumulation of unfolded proteins, leading to the re-establishment of cellular homeostasis. However, if the ER stress cannot be alleviated, aberrant ER stress can trigger cell apoptosis, particularly by activating caspase-12^[13-15]. However, how aberrant ER stress induces caspase-12 activation to trigger cell apoptosis has not been clarified in hepatocytes.

Calpain-2 is a Ca²⁺-dependent cysteine protease that can cleave its protein substrates. Calpain-2 can regulate cell cycle, differentiation, and apoptosis^[16]. Previous studies have revealed that calpains promote ER stress-related apoptosis by activating caspase-12^[17-19]. Actually, our previous studies indicated that ER stress occurred in hepatocytes in a rat model of carbon tetrachloride-induced hepatic fibrosis, which was associated with increased calpain-2 and caspase-12 expression and hepatocyte apoptosis^[20,21]. However, it is unclear whether calpain-2 can modulate caspase-12

activation and ER stress-related apoptosis in hepatocytes.

This study explored the importance of calpain-2 in regulating caspase-12 activation and dithiothreitol (DTT)-induced ER stress-related apoptosis in hepatocytes. The results indicated that calpain-2 activity was crucial for DTT-induced ER stress-related hepatocyte apoptosis by activating caspase-12 *in vitro*.

MATERIALS AND METHODS

Special reagents included rat non-tumor BRL-3A cells (Number: KCB92013YJ, Kunming Cell Bank of Chinese Academy of Sciences, China), fetal bovine serum, Dulbecco's modified Eagle's medium (DMEM; GIBCO, New York, NY, United States), Acrylamide, bisacrylamide, ammonium peroxydisulfate, glycine, Tri-hydroxymethyl aminomethane, 3-[4, 5-dimethyl-2-thiazolyl]-2, 5-diphenyl-2-H- tetrazolium bromide (MTT), Tween 20, DTT, dimethyl sulfoxide (Genview, League City, TX, United States), *N*-succinyl-Leu-Leu-Val-Tyr-AMC, antibodies against GRP78, PERK, ATF4, CHOP, and caspase-12 (Abcam, Cambridge, United Kingdom), antibodies against p-PERK (Affinity Biosciences, Cincinnati, OH, United States), calpain-2 and β -actin (Cell Signaling Technologies, Danvers, MA, United States), caspase-3 and secondary antibodies (Boster Biological Engineering, Wuhan, China), polyvinylidene difluoride membranes and enhanced chemiluminescence kit (Millipore, Burlington, MA, United States). Calpain-2-specific and control small interfering RNAs (siRNAs) and siRNA dilution buffer were produced by Santa Cruz Biotechnology (Santa Cruz, CA, United States), and the Annexin V-FITC Detection Kit, ER-tracker Red, and Fluo-3 AM were obtained from Beyotime Institute of Biotechnology (Nanjing, China).

Cell culture and treatment

Rat non-tumor hepatocyte BRL-3A cells were cultured in high-glucose DMEM supplemented with 10% fetal bovine serum, 100 units/mL penicillin, and 100 μ g/mL streptomycin at 37 °C in a humidified atmosphere of 5% CO₂. When the cells reached 80% confluency, the cells were treated with DTT, an inhibitor of disulfide bond formation, to induce ER stress.

MTT assay

We examined the impact of DTT on the BRL-3A cell viability by MTT. Briefly, BRL-3A cells (5×10^3 cells/well) were treated in triplicate with 0-10 mmol/L of DTT for 24 h. During the last 4-h culture, the cells were treated with 0.5 mg/mL MTT reagent. The resulting formazan in individual wells was dissolved with 150 μ L dimethyl sulfoxide, and the absorbance at 570 nm/L in individual wells was measured using a microplate reader.

Flow cytometry

The effect of DTT on the apoptosis of BRL-3A cells was quantified by flow cytometry using the Annexin V/propidium (PI) kit per the manufacturer's protocol. Briefly, BRL-3A cells (5×10^5 cells/flask) were treated in triplicate with vehicle or 2.0 mmol/L DTT, a sub-toxic dose, for varying time periods^[22]. After being washed twice with phosphate-buffered saline, the cells were stained with FITC-Annexin-V and PI. The percentages of apoptotic cells in individual groups were quantified by flow cytometry in a FACSCalibur™ (BD Biosciences, Franklin Lakes, NJ, United States).

Western blotting

After treatment with 2.0 mmol/L DTT for varying time periods, BRL-3A cells were lysed in a RIPA buffer and centrifuged, followed by quantifying the protein concentrations. The cell lysates (50 μ g/lane) were separated by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) on 10%-12% gels and transferred onto polyvinylidene difluoride membranes. After being blocked in 5% non-fat dry milk in Tris-buffered saline with Tween-20, the membranes were probed overnight at 4°C with primary antibodies against GRP78 (1:1500), PERK (1:1500), p-PERK (1:1500), ATF4 (1:1500), CHOP (1:1500), calpain-2 (1:1000), caspase-12 (1:1000), cleaved caspase-3 (1:500), and β -actin (1:1000). The bound antibodies were detected with horseradish peroxidase-conjugated secondary antibodies and visualized using enhanced chemiluminescence reagents. The signal intensity was measured using Bio-Rad imaging system (Bio-Rad, Hercules, CA, United States) and analyzed by Quantity One software (Bio-Rad).

Microscopy analysis of intracellular Ca²⁺ content

The intracellular Ca²⁺ content in BRL-3A cells was detected by microscopy after staining with a Ca²⁺-sensitive fluorescent dye, Fluo-3 AM^[23]. In brief, BRL-3A cells

were treated in triplicate with 2.0 mmol/L DTT for 0, 6, 12, and 24 h. After being washed with Hank's balanced salt solution, the cells were stained with Fluo-3 AM at 37 °C for 45 min. The morphology and fluorescent signals in individual wells of cells were examined under a light and fluorescent microscope (FV1000; Olympus, Tokyo, Japan).

Calpain activity assay

We measured cellular calpain enzymatic activity using a fluorescence substrate *N*-succinyl-Leu-Leu-Val-Tyr-AMC, as previously described^[24]. In brief, individual cell lysates were reacted at 37°C with 40 μmol/L *N*-succinyl-Leu-Leu-Val-Tyr-AMC for 1 h, and the fluorescent signals were measured using a fluorescence plate reader.

Immunofluorescent microscopy

BRL-3A cells were cultured on coverslips overnight, and treated with 2.0 mmol/L DTT for 0, 6, 12, and 24 h. The cells were stained with ER-tracker Red (ER fluorescent dye)^[25,26], fixed in 2% paraformaldehyde, and permeabilized with 0.2% Triton X-100. Subsequently, the cells were stained with anti-calpain-2 and Alexa Fluor 488-conjugated secondary antibody. We photoimaged the fluorescent signals under a fluorescence microscope (IX71; Olympus).

Calpain-2 siRNA transfection

BRL-3A cells (5×10^5 cells/flask) were grown in antibiotic-free medium overnight and transfected with control or calpain-2 specific siRNA for 48 h. The efficacy of calpain-2 silencing was determined by Western blotting. Subsequently, the different groups of cells were treated in triplicate with 2.0 mmol/L DTT for 24 h and used for analysis of apoptosis, calpain-2, and cleaved caspase-12 expression.

Statistical analysis

Data are present as the mean \pm SD. We compared the different groups of data by one-way analysis of variance and post hoc least significant difference test using SPSS 13.0 software. Statistical significance was defined at a *P* value < 0.05.

RESULTS

DTT exhibits cytotoxicity against BRL-3A cells in a dose-dependent manner

To determine the toxicity of DTT, BRL-3A cells were treated with 0-10 mmol/L DTT for 24 h and their viability was measured by MTT (Figure 1A). DTT had dose-dependent cytotoxicity against BRL-3A cells and the median lethal dose (LD50) value for BRL-3A cells was about 4 mmol/L in our experimental condition. Longitudinal analysis displayed that treatment with 2 mmol/L DTT for 6 h significantly increased the percentages of apoptotic BRL-3A cells by about 3-fold (*P* < 0.01), and treatment for a longer period did not significantly increase the frequency of apoptotic BRL-3A cells (Figure 1B and C). Hence, DTT exhibited dose-dependent cytotoxicity against BRL-3A cells by increasing their apoptosis *in vitro*.

DTT induces ER stress in BRL-3A cells

DTT can induce ER stress in many types of cells^[27,28]. To understand the role of DTT in decreasing viability, BRL-3A cells were treated with 2.0 mmol/L DTT for varying time periods and their ER stress-related proteins were quantified by Western blot (Figure 2). DTT treatment for 6 h upregulated GRP78, ATF4, and CHOP expression and PERK phosphorylation by about 2-fold in BRL-3A cells and treatment with DTT for a longer period further increased its effects. The effects of DTT on upregulating GRP78 expression and PERK phosphorylation appeared to have a trend of time-dependence. Thus, DTT induced ER stress in BRL-3A cells *in vitro*.

DTT enhances caspase-12 and caspase-3 activation and calpain-2 activity in BRL-3A cells

Aberrant ER stress can promote sensitive cell apoptosis by inducing caspase-12 and caspase-3 activation, and calpain-2 participates in the process of ER-stress-related apoptosis^[18,19,25]. To understand the consequence of ER stress induced by DTT, the relative levels of caspase-3, and caspase-12 cleavage, and calpain-2 activity were quantified. Treatment with 2 mmol/L DTT significantly induced time-dependent caspase-12 cleavage and increased the levels of cleaved caspase-3 in BRL-3A cells by 2-2.5 fold (Figure 3A and B). Similarly, DTT treatment significantly upregulated calpain-2 expression by more than 2-fold and enhanced calpain-2 activity in a trend of time-dependence in BRL-3A cells (Figure 3C and D). Given that calpain-2 activity is Ca²⁺-dependent, we further quantified intracellular Ca²⁺ content in each group of cells

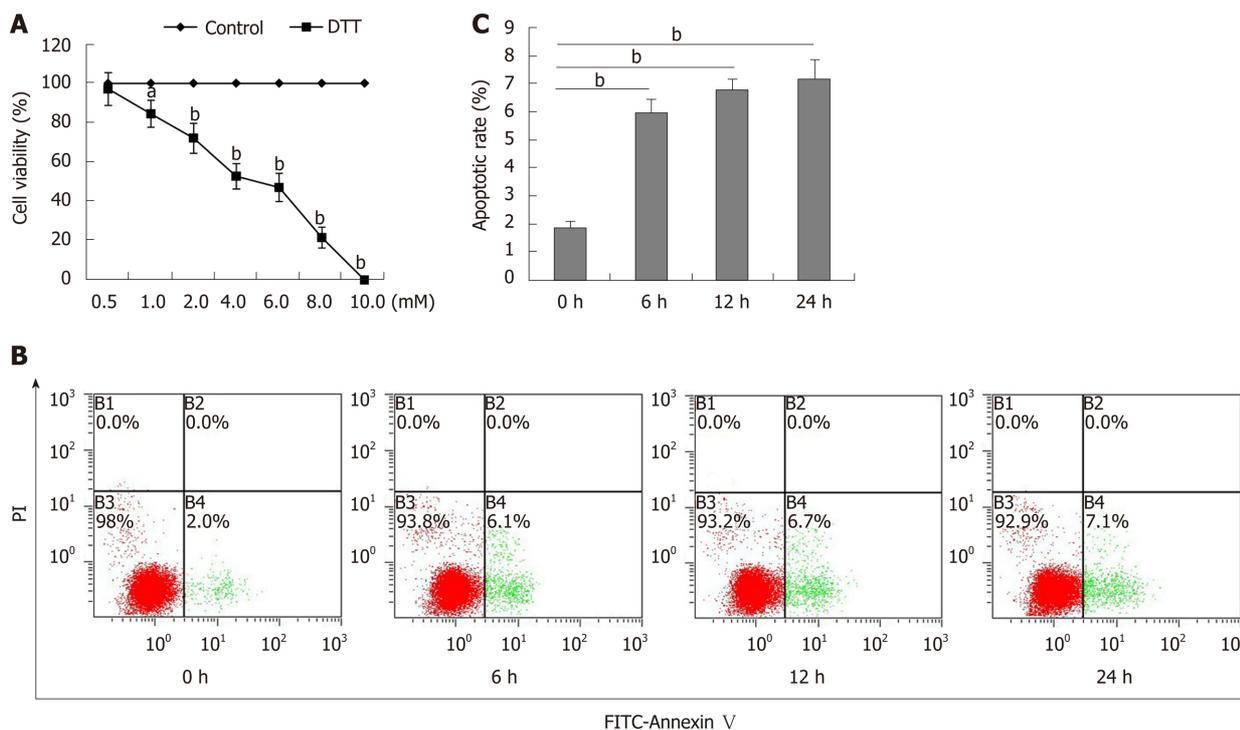


Figure 1 Dithiothreitol exhibits dose-dependent cytotoxicity against BRL-3A cells. **A**: BRL-3A cells were treated in triplicate with, or without, the indicated doses of dithiothreitol (DTT) for 24 h and their viability was examined by MTT. The percentages (y) of cell viability were calculated by the equation, $y = -9.3654x + 95.744$ with a correlation ($R^2 = 0.9781$), here x represents DTT concentration. Accordingly, the DTT $LD_{50} = 4.88$ mM; **B**, **C**: Subsequently, BRL-3A cells were treated with 2.0 mmol/L DTT for varying periods and stained with FITC-Annexin-V and PI. The percentages of mechanically damaged (B1 quadrant), healthy (B3), early (B4) and late apoptotic and necrotic (B2) cells were analyzed by flow cytometry. Data are representative flow cytometry charts or expressed as the mean \pm SD of each group from three separate experiments. ^a $P < 0.05$, ^b $P < 0.01$.

by microscopy. After staining with Fluo-3 AM, we observed that DTT treatment time dependently increased the content of clustery Ca^{2+} by 2-3 fold in BRL-3A cells at 12 and 24 h post treatment (Figure 4). To identify additional evidence to demonstrate the importance of calpain-2 activity, we stained the different groups of cells with fluorescent anti-calpain-2 and ER-tracker Red, and observed the co-localization of red ER and green calpain-2 signals by fluorescent microscopy. As shown in Figure 5, there were obviously increased merged yellow signals in the DTT-treated cells, particularly at the later time point, while there was little merged signal in the cells without DTT treatment. Collectively, such data indicated that DTT-induced aberrant ER stress promoted the apoptosis of BRL-3A cells by activating caspase-12 and increasing calpain-2 activity.

Calpain-2 silencing mitigates DTT-induced caspase-12 activation and apoptosis in BRL-3A cells

Finally, we tested whether calpain-2 silencing could modulate the DTT-induced caspase-12 activation and apoptosis of BRL-3A cells. We found that transfection with calpain-2 specific siRNA, but not the control, significantly decreased calpain-2 expression by about 85%, demonstrating the efficacy of calpain-2 silencing ($P < 0.01$; Figure 6A). Calpain-2 silencing also significantly mitigated the DTT-upregulated calpain-2 expression near 80% ($P < 0.01$; Figure 6B). Although calpain-2 silencing did not alter the DTT-upregulated caspase-12 expression, it significantly reduced the caspase-12 cleavage near 63% in BRL-3A cells, relative to that in the control cells ($P < 0.01$; Figure 6C). More importantly, calpain-2 silencing significantly decreased the percentage of DTT-induced apoptosis of BRL-3A cells by 50% ($P < 0.05$; Figure 6D and E). Together, the significantly decreased caspase-12 activation and cell apoptosis indicated that calpain-2 activity was crucial for DTT-induced ER stress-related caspase-12 activation and apoptosis in BRL-3A cells.

DISCUSSION

Many factors can induce ER stress including chemical agents such as tunicamycin,

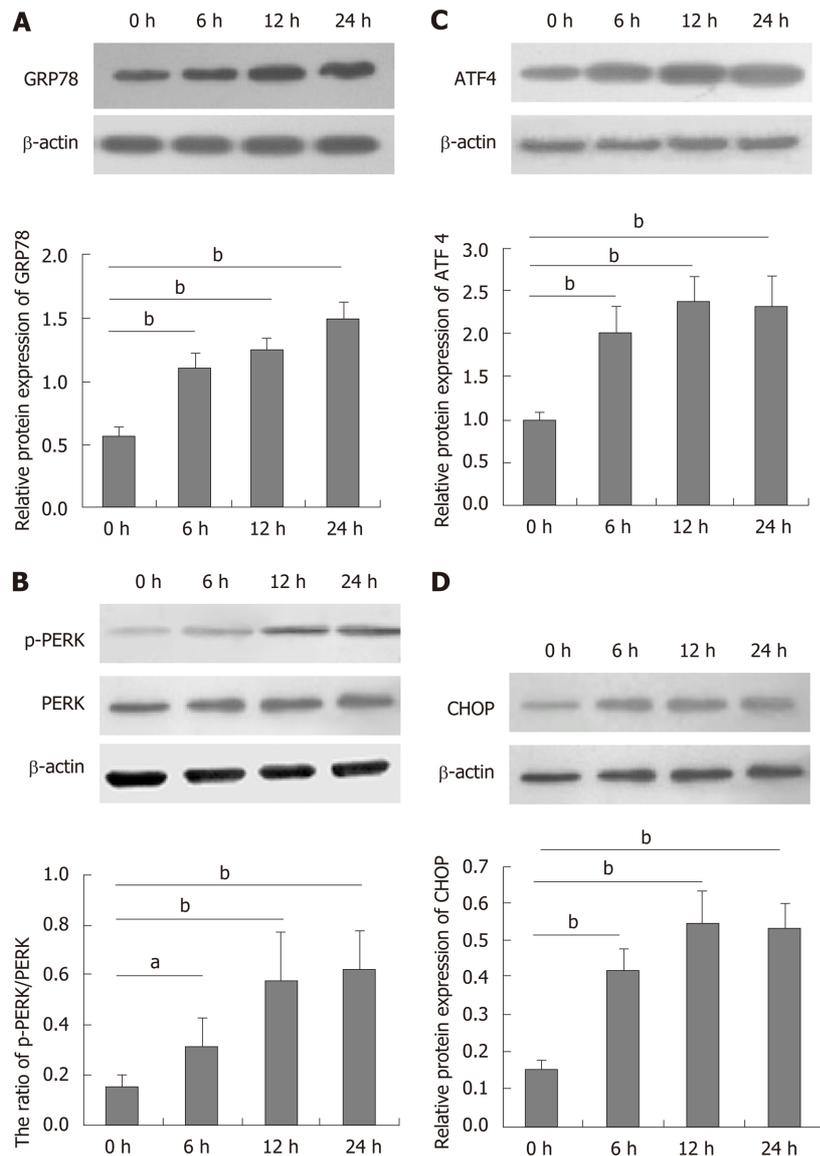


Figure 2 Dithiothreitol induces endoplasmic reticulum stress in BRL-3A cells. BRL-3A cells were treated in triplicate with, or without, 2.0 mmol/L dithiothreitol for the indicated time periods, and the relative levels of glucose-regulated protein 78, activating transcription factor 4 and C/EBP-homologous protein to β -actin expression and PERK phosphorylation in each group of cells were quantified by Western blot. Data are representative images or expressed as the mean \pm SD of each group of cells from three separate experiments. A: Relative glucose-regulated protein 78 expression; B: Relative PERK phosphorylation; C: Relative activating transcription factor 4 expression; D: Relative C/EBP-homologous protein expression. ^a $P < 0.05$, ^b $P < 0.01$.

thapsigargin, and DTT^[27,29]. In this study, we used a sub-toxic 2.0 mmol/L DTT to induce ER stress in BRL-3A cells, based on a previous report^[22]. It is well known that DTT can inhibit the formation of disulfide bonds and result in the accumulation of unfolded proteins in the ER, leading to ER stress.

Aberrant ER stress-induced apoptosis of hepatocytes participates in the pathogenesis of several types of liver diseases^[30-32]. In this study, we found that DTT had strong cytotoxicity against rat hepatocyte BRL-3A cells and its cytotoxicity was dose-dependent. Furthermore, treatment with DTT induced ER stress in BRL-3A cells by significantly upregulating GRP78, ATF4, and CHOP expression and PERK phosphorylation. More importantly, DTT treatment significantly increased the frequency of apoptotic BRL-3A cells, supporting that aberrant ER stress promotes the apoptosis of hepatocytes^[33,34]. Given that hepatocyte apoptosis participates in the pathogenesis of several types of liver diseases, inhibition of ER stress may be valuable for protecting hepatocytes from apoptosis.

ER stress-related apoptosis is independent of mitochondria and death receptors, and is thought to be mediated by activating caspase-12, an ER-anchored caspase^[35,36]. A previous study has reported that the caspase-12 gene is mutated in most human

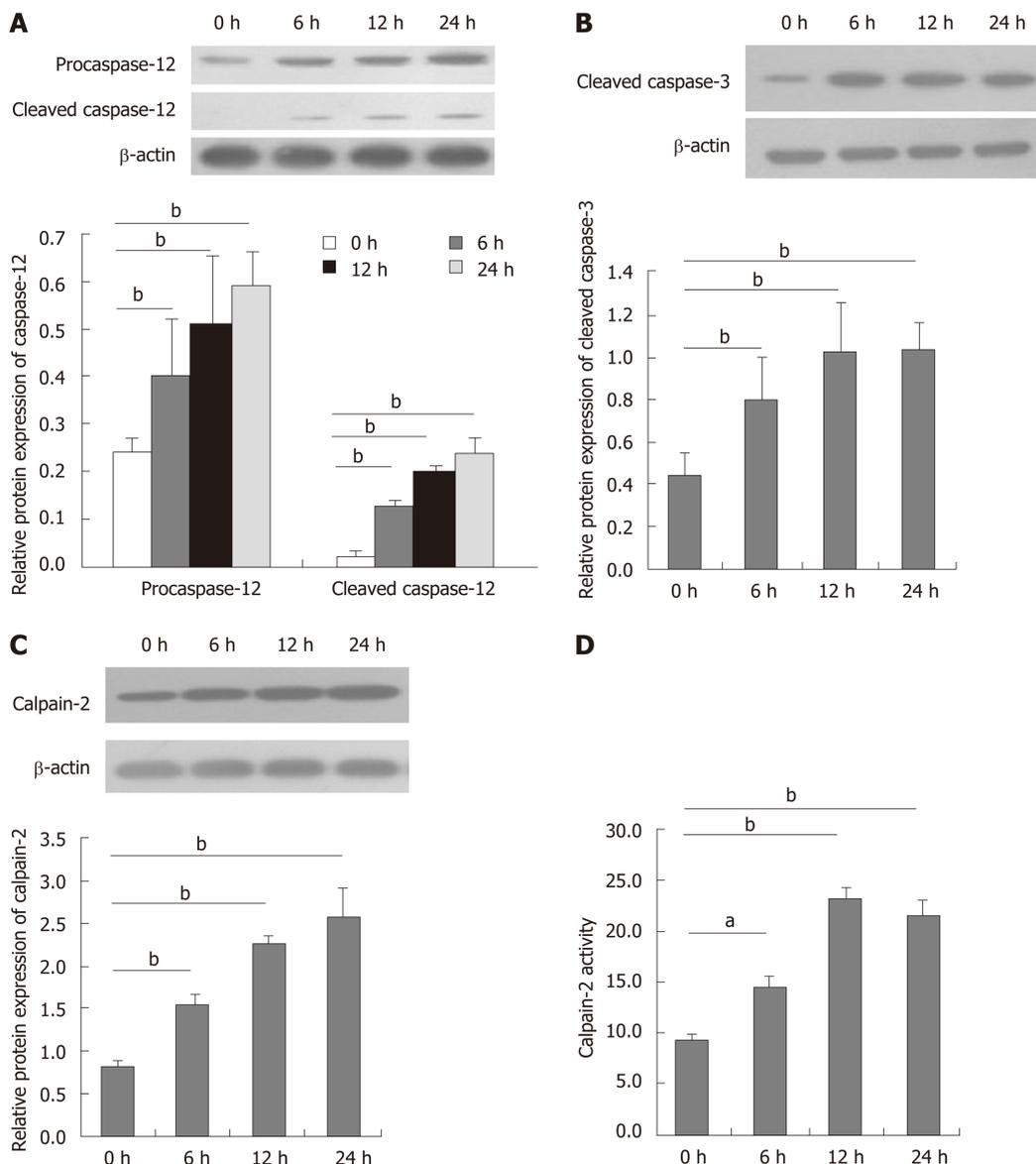


Figure 3 Dithiothreitol induces caspase-12 and caspase-3 activation and increases calpain-2 activity in BRL-3A cells. After treatment with 2.0 mmol/L dithiothreitol for varying periods, the relative levels of caspase-12, cleaved caspase-12 and cleaved caspase-3 as well as calpain-2 expression in each group of BRL-3A cells were quantified by Western blotting. Furthermore, the activity of calpain-2 in each group of cells was measured. Data are representative images or expressed as the mean \pm SD of each group of cells from three separate experiments. A: Caspase-12 activation; B: Caspase-3 activation; C: Calpain-2 expression; D: Calpain-2 activity. ^a*P* < 0.05, ^b*P* < 0.01.

hepatocyte lines^[23], so we chose rat non-tumor hepatocyte BRL-3A cells as a cell model in this study. Similar to other caspases, procaspase-12 has to be activated by cleaving a short inhibitory peptide, and cleaved caspase-12 can activate the downstream effector caspase-3, leading to apoptosis^[37]. Actually, caspase-12^{-/-} cells are resistant to ER stress-induced apoptosis^[38]. In this study, we found that DTT treatment significantly induced caspase-12 and caspase-3 cleavage in BRL-3A cells. Such data indicated that activated caspase-12 and caspase-3 contributed to the ER stress-related apoptosis of BRL-3A cells, although the precise mechanisms underlying caspase-12 activation in hepatocytes remain incompletely understood. A recent study revealed that enhanced calpain-2 activity is crucial for caspase-12 activation^[39]. Calpain-2 is a Ca²⁺-dependent cysteine protease, and its activity and function depend on the concentrations of intracellular Ca²⁺^[40]. As the ER holds the major pool of Ca²⁺, ER stress can promote Ca²⁺ efflux from the ER, which may be responsible for enhancing calpain-2 activity and its ER accumulation^[41]. Interestingly, we found that DTT treatment significantly increased intracellular Ca²⁺ content and calpain-2 activity in BRL-3A cells. Furthermore, DTT treatment promoted the accumulation of calpain-2 in the ER of BRL-3A cells. More importantly, calpain-2 silencing not only significantly

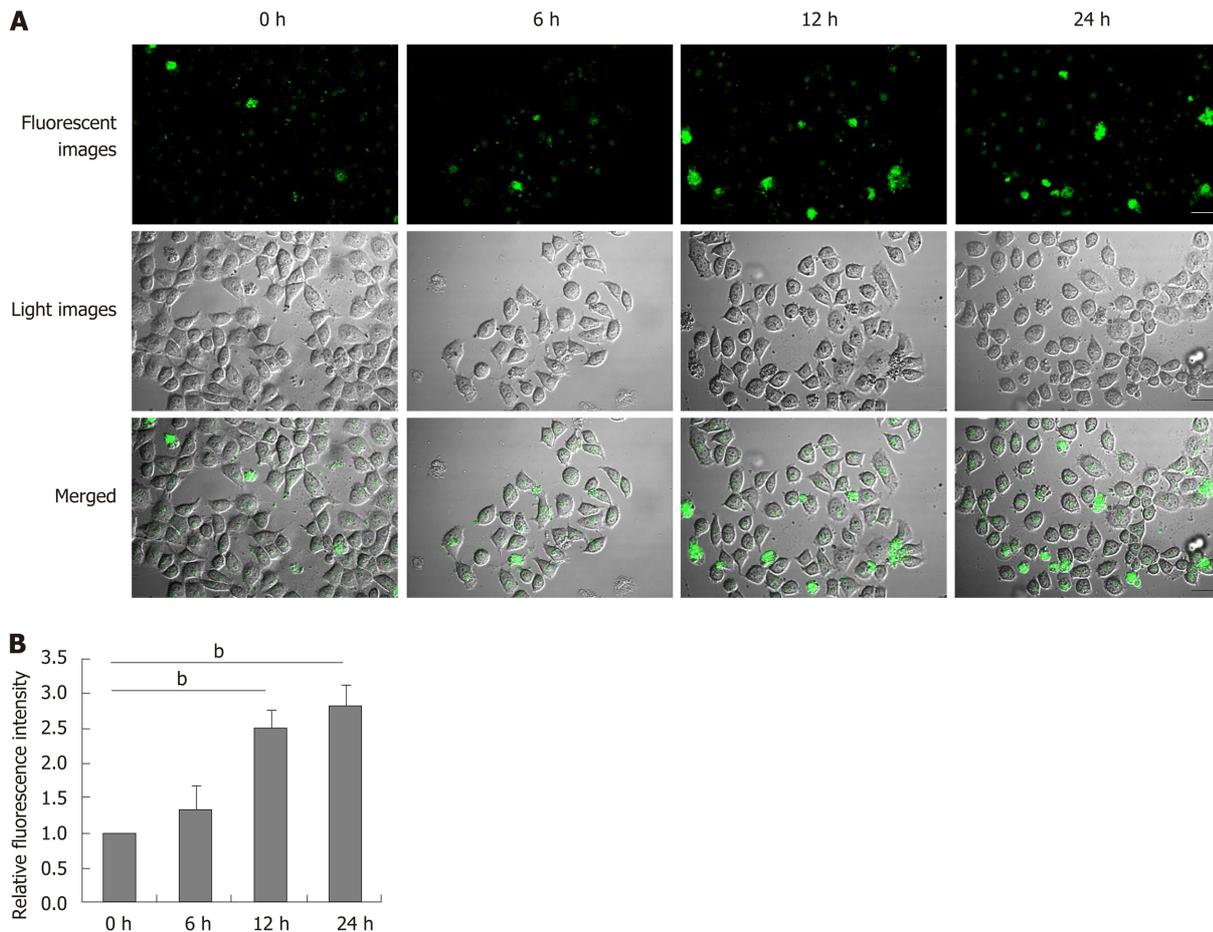


Figure 4 Dithiothreitol increases the levels of intracellular Ca^{2+} in BRL-3A cells. After treatment with 2.0 mmol/L dithiothreitol for varying time periods, the cells were labeled by Fluo-3 AM. The cell morphology and fluorescent signals in cells were observed by microscopy. A: Representative images of Fluo-3 fluorescence and morphology in BRL-3A cells following dithiothreitol treatment (Scale bars: 25 μm); B: Quantification of fluorescence intensity in BRL-3A cells. Data are representative images (magnification 200 \times) or expressed as the mean \pm SD of each group of cells from three separate experiments. ^b $P < 0.01$.

mitigated DTT-upregulated calpain-2 expression and caspase-12 activation, but also decreased the DTT-triggered apoptosis of BRL-3A cells. Hence, the DTT-induced ER stress increased intracellular Ca^{2+} content and calpain-2 expression, leading to calpain-2 activation, which cleaved caspase-12 to trigger apoptosis of BRL-3A cells. In our recent preliminary studies, we found that treatment with Z-LLY-fmk, a specific inhibitor of calpain, significantly mitigated the DTT-induced hepatocyte apoptosis by more than 90% (Chen *et al.*^[11], unpublished data). Such novel findings may provide new evidence to demonstrate that the Ca^{2+} -dependent calpain-2 activity is crucial for promoting the ER stress-related apoptosis in hepatocytes.

In conclusion, our data indicated that DTT exhibited dose-dependent cytotoxicity against rat hepatocytes, and induced ER stress and apoptosis in BRL-3A cells. Evidently, DTT treatment significantly upregulated GRP78, ATF4, and CHOP expression and PERK phosphorylation; increased intracellular Ca^{2+} content and calpain-2 activity; and induced caspase-12 and caspase-3 activation in BRL-3A cells. Furthermore, calpain-2 silencing significantly mitigated DTT-upregulated calpain-2 expression and DTT-induced caspase-12 activation as well as apoptosis of BRL-3A cells. The results indicated that enhanced calpain-2 activity promoted aberrant ER stress-mediated apoptosis of hepatocytes. Our data suggest that ER stress may induce Ca^{2+} release from the ER and lead to the recruitment and activation of calpain-2 in the ER, where calpain-2 activates caspase-12 and caspase-3 and triggers apoptosis in hepatocytes. Therefore, ER stress may be a novel therapeutic target and our findings may provide new evidence to demonstrate the importance of Ca^{2+} -dependent calpain-2 in caspase-12 activation and ER stress-related apoptosis in hepatocytes.

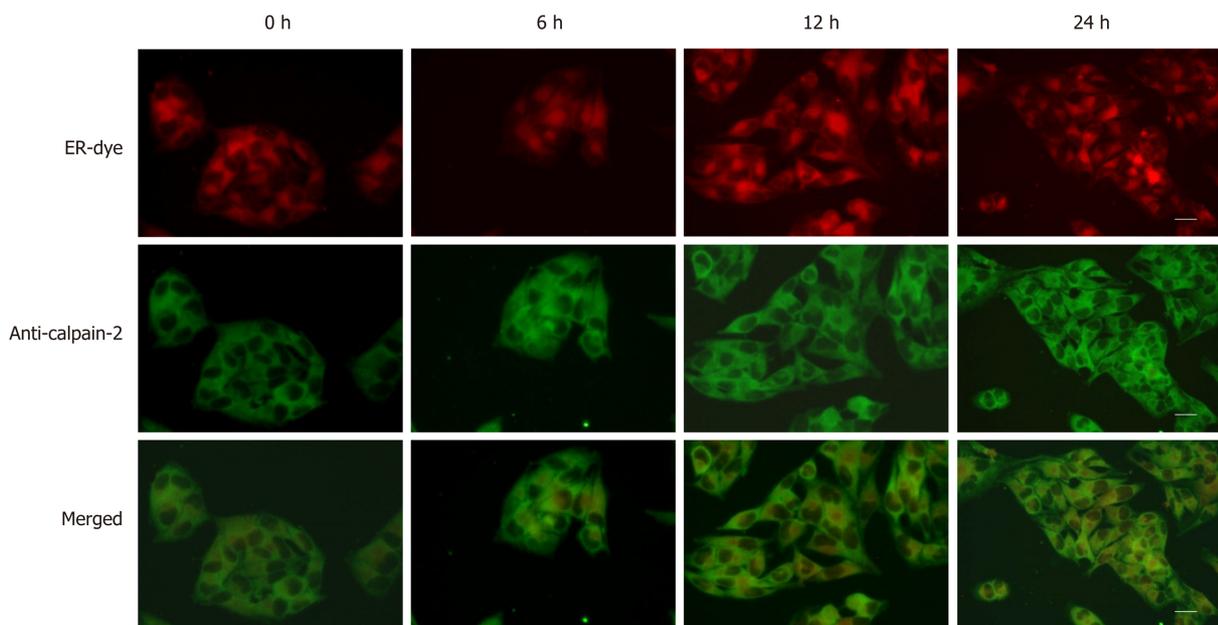
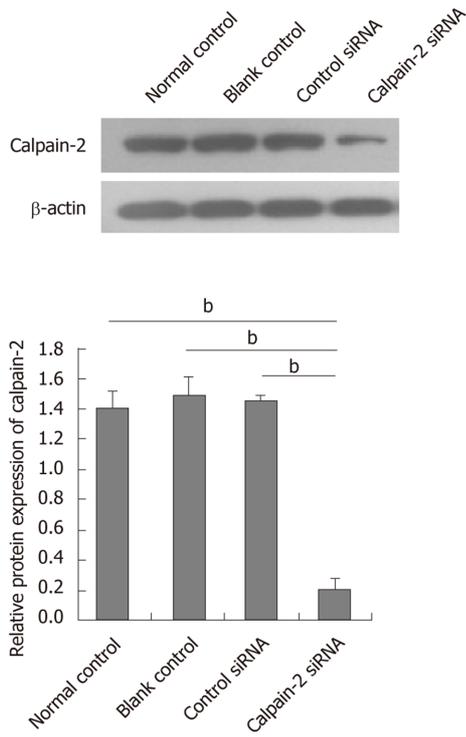
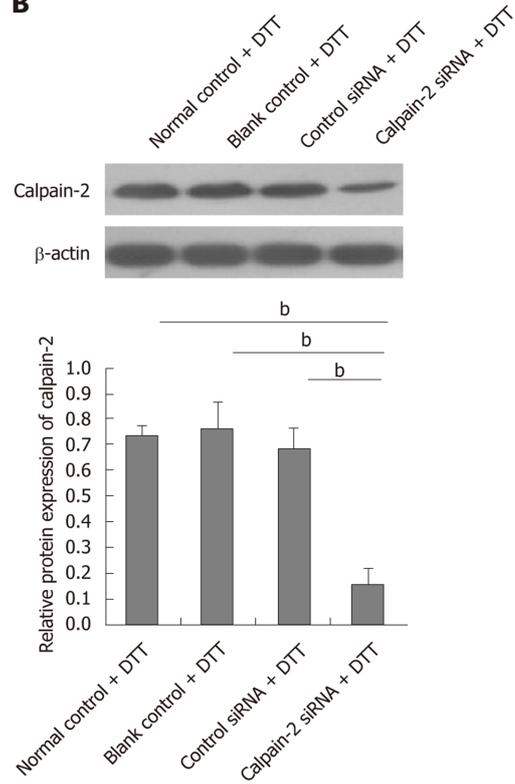


Figure 5 Dithiothreitol promotes the accumulation of calpain-2 in the endoplasmic reticulum of BRL-3A cells. After treatment with 2.0 mmol/L dithiothreitol for varying time periods, the cells were labeled with endoplasmic reticulum-tracker red dye, fixed, permeabilized, followed by staining with fluorescent-anti-calpain-2 (green). The cells were examined by fluorescent microscopy, scale bars, 25 μ m. Data are representative images (magnification 200 \times) of each group of cells from three separate experiments.

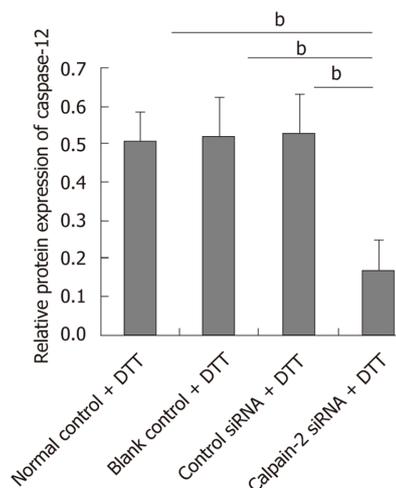
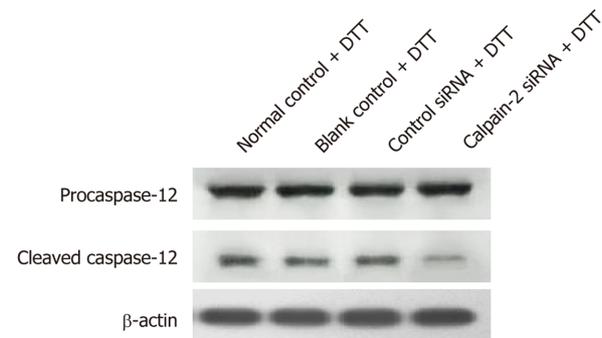
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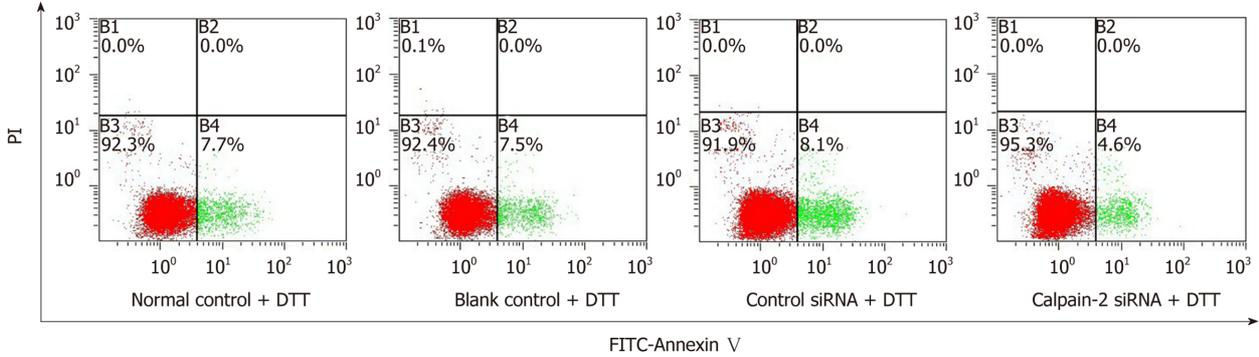
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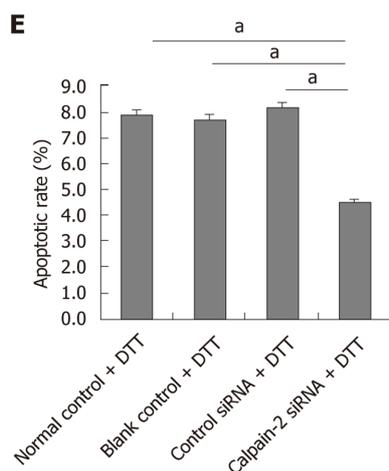


Figure 6 Calpain-2 silencing mitigates dithiothreitol-upregulated calpain-2 expression, caspase-12 activation and apoptosis of BRL-3A cells. BRL-3A cells were transfected with, or without, control or calpain-2 specific siRNA for 48 h and treated with, or without, dithiothreitol (DTT). The relative levels of calpain-2 expression and caspase-12 activation were quantified by Western blot. The percentages of apoptotic cells were quantified by flow cytometry. Data are representative images or expressed as the mean \pm SD of each group of cells from three separate experiments. A: Calpain-2 silencing; B: Calpain-2 silencing mitigates DTT-upregulated calpain-2 expression; C: Calpain-2 silencing decreases DTT-induced caspase-12 activation; D, E: Calpain-2 silencing mitigates DTT-triggered apoptosis of BRL-3A cells. ^a $P < 0.05$, ^b $P < 0.01$.

ARTICLE HIGHLIGHTS

Research background

Endoplasmic reticulum (ER) stress-mediated hepatocyte apoptosis is associated with many liver diseases, however, the underlying mechanisms remain unknown. Calpain-2 is a Ca^{2+} -dependent cysteine protease, which can cleave several apoptosis-related proteins and mediate the process of apoptosis. Our data indicate that calpain-2 is crucial for the aberrant ER stress-induced apoptosis of hepatocytes and may be a novel therapeutic target for liver diseases.

Research motivation

Calpain-2 protease is involved in multiple signaling pathways mediating apoptotic processes, including the mitochondrial pathway. However, the regulatory mechanisms by which calpain-2 regulates ER stress-mediated hepatocyte apoptosis remain unclear. Our study further assessed its potential as a therapeutic target for inhibiting hepatocyte apoptosis.

Research objectives

In this study, the effect of calpain-2 on ER stress-mediated hepatocyte apoptosis and the underlying regulatory mechanisms were investigated. Our data indicates that ER stress may be a novel therapeutic target, and our findings provide new evidence to demonstrate the importance of Ca^{2+} -dependent calpain-2 in caspase-12 activation and ER stress-related apoptosis in hepatocytes.

Research methods

Our study employed Western blotting, flow cytometry, confocal microscopy, and calpain-2 small interfering RNA to elucidate the mechanism of calpain-2-mediated activation of caspase-12, a key molecule in ER stress-related apoptosis. These research methods are relatively mature technically, thus ensuring the reliability of the results from this study.

Research results

The ER stress inducer dithiothreitol can significantly increase intracellular Ca^{2+} content, calpain-2 expression and activity in hepatocytes and promote caspase-12 cleavage and apoptosis in hepatocytes. Moreover, calpain-2 silencing can mitigate dithiothreitol-enhanced calpain-2 expression, caspase-12 cleavage, and apoptosis in hepatocytes. These results indicated that enhanced calpain-2 activity promoted aberrant ER stress-mediated apoptosis of hepatocytes. In future studies, we will further investigate whether calpain-2 can mediate hepatocyte death through other mechanisms, such as regulating autophagy.

Research conclusions

Ca^{2+} -dependent calpain-2 activity promoted the aberrant ER stress-related apoptosis of rat hepatocytes by activating caspase-12 in the ER. ER stress may be a novel therapeutic target and our findings may provide new evidence to demonstrate the importance of Ca^{2+} -dependent calpain-2 in caspase-12 activation and ER stress-related apoptosis in hepatocytes. ER stress may induce Ca^{2+} release from the ER and lead to the recruitment and activation of calpain-2 in the ER, where, calpain-2 activates caspase-12 and caspase-3, and triggers apoptosis in hepatocytes. Calpain-2 activity plays an important role in aberrant ER stress-related apoptosis in hepatocytes.

Inhibition of calpain-2 expression and activity is expected to be an effective way to alleviate hepatocyte apoptosis and liver injury.

Research perspectives

Because calpain-2 is a protease, in addition to focusing on its expression, studies should also focus on its activity. In our recent preliminary studies, we found that treatment with Z-LLY-fmk, a specific inhibitor of calpain activity, significantly mitigated DTT-induced hepatocyte apoptosis by more than 90%.

ACKNOWLEDGEMENTS

We thank Hua Pei and Jinxingyi Wang for their technical advice in using flow cytometry and microscope in the Basic Medical Science Research Center of Guizhou Medical University. We would like to thank Dr. Tengxiang Chen for his comments on this manuscript.

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