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***Prospective Study***

**Macrophage inhibitory cytokine-1/growth differentiation factor-15 in premalignant and neoplastic tumours in a high-risk pancreatic cancer cohort**

O’Neill RS *et al*. MIC-1 in PC screening

Robert Sean O’Neill, Sam Emmanuel, David Williams, Alina Stoita

**Robert Sean O’Neill, David Williams, Alina Stoita,** Department of Gastroenterology, St Vincent’s Hospital, Sydney 2010, Australia

**Sam Emmanuel, Alina Stoita,** St Vincent’s Clinical School, Faculty of Medicine, University of New South Wales, Sydney 2010, Australia

**Author contributions:** O’Neill RS contributed to the writing of the manuscript; Emmanuel S provided the statistical analysis; Stoita A and Williams D as senior authors designed the prospective study, performed screening and revised the manuscript.

**Corresponding author: Alina Stoita, FRACP, MBBS, Assistant Lecturer, Doctor,** Department of Gastroenterology, St Vincent’s Hospital Sydney, 390 Victoria Street, Sydney 2010, Australia. alina.stoita@svha.org.au

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**Abstract**

BACKGROUND

Pancreatic cancer (PC) is a leading cause of cancer related mortality worldwide, with poor survival due to late diagnosis. Currently, biomarkers have limited use in early diagnosis of PC. Macrophage inhibitory cytokine-1 or growth differentiation factor-15 (MIC-1/GDF15) has been implicated as a potential serum biomarker in PC and other malignancies.

AIM

To determine the role of MIC-1/GDF15 in detecting pre-malignant pancreatic lesions and neoplastic tumours in an asymptomatic high-risk cohort part of Australian Pancreatic Cancer Screening Program.

METHODS

A feasibility prospective single centre cohort study was performed. Participants recruited for yearly surveillance with endoscopic ultrasound (EUS) had serial fasting blood samples collected before EUS for MIC-1/GDF15, C-reactive protein and carbohydrate antigen 19-9. Patients were stratified into five groups based on EUS findings: normal; pancreatic cysts, branch-duct intraductal papillary mucinous neoplasm; diffuse non-specific abnormalities; and neoplastic tumours. MIC-1/GDF15 serum levels were quantified using ELISA. Participants in whom EUS demonstrated abnormalities but not malignancy were closely followed up with magnetic resonance imaging (MRI) or computed tomography.

RESULTS

One hundred twenty participants were prospectively recruited from 2011-2018. Forty-seven participants (39.2%) had an abnormal EUS and five participants (4.2%) were diagnosed with neoplastic tumours, three by EUS (two pancreatic and one liver) and two by MRI/computed tomography (breast cancer, bladder cancer), which were performed for follow up of abnormal EUS. Baseline serum MIC-1 was a significant predictor of neoplastic tumours on receiver operator characteristic curve analysis [area under curve (AUC) = 0.814, *P* = 0.023]. Baseline serum MIC-1/GDF15 had moderate predictive capacity for branch-duct intraductal papillary mucinous neoplasm (AUC = 0.644) and neoplastic tumours noted on EUS (AUC = 0.793), however this was not significant (*P* = 0.188 and 0.081 respectively). Serial serum MIC-1/GDF15 did not demonstrate a significant percentage change between a normal and abnormal EUS (*P* = 0.213). Median baseline MIC-1/GDF15 was greater in those with neoplastic tumours (Median = 1039.6, interquartile range = 727.0-1977.7) compared to those diagnosed with a benign lesion (Median = 570.1, interquartile range = 460.7-865.2) on EUS and MRI (*P* = 0.012).

CONCLUSION

In this pilot study MIC-1/GDF15 has predictive capacity for neoplastic tumours in asymptomatic individuals with a genetic predisposition for PC. Further imagining may be warranted in patients with abnormal EUS and raised serum MIC-1. Larger multicentric prospective studies are required to further define the role of MIC-1/GDF15 as a serological biomarker in pre-malignant pancreatic lesions and neoplastic tumours.

**Key words:** Growth differentiation factor 15; Cytokines; Pancreatic neoplasms; Digestive system neoplasms; Pancreatic diseases; Biomarkers; Diagnostic screening programs

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**Core tip:** In this prospective cohort study in an asymptomatic population at high risk of developing pancreatic cancer due to a genetic predisposition serum baseline macrophage inhibitory cytokine-1 or growth differentiation factor-15 was shown to be a significant predictor of neoplastic tumours (both pancreatic and extra-pancreatic).

**INTRODUCTION**

Macrophage inhibitory cytokine-1 (MIC-1), also known as growth differentiation factor-15 (GDF-15) is a distant member of the transforming growth factor (TGF-b) superfamily of cytokines, with its original role being identified as a gene expressed in the context of macrophage activation[1,2]. MIC-1/GDF-15 is present in the serum of all individuals with a wide normal range 150-1150 pg/mL[3]. MIC-1/GDF15 has been implicated in regulation of inflammation, metabolism and carcinogenesis, with previous literature demonstrating serum elevation in acute inflammatory conditions, congestive heart failure, renal failure and anti-inflammatory use[4-7]. More recent studies have focused on its role in malignancy, being one of the few secreted proteins induced by p53 activation and its expression was initially postulated to stimulate apoptosis in cancer cells[8-10]. More recently it was suggested that MIC-1/GDF15 directly modulates the biology of tumour progression from initial tumorigenesis to metastasis[11]. In addition to this, MIC-1/GDF15 protein and mRNA was noted to be elevated both in cancer tissue specimens along with peripheral serum samples. MIC-1 has been implicated in colorectal cancer, with serum levels being elevated in patients with premalignant colonic polyps, and subsequently increasing with disease progression, including metastasis, along with predicting disease outcome[12-15]. In addition to this, other studies have identified a potential role of MIC-1/GDF15 in prostate[16], breast[17], pancreatic[18-20], ovarian[21], endometrial[22] and lung cancer[23]. Although the role of MIC-1/GDF15 as a biomarker in malignancy has been explored, there is still ongoing discussion regarding its precise function in malignancy, with researchers hypothesising that MIC-1/GDF15 enhances anti-tumour immunity in the early stages of malignancy, along with stimulating tumour cell spread through promoting tumour angiogenesis as demonstrated in oesophageal squamous cell carcinoma[24].

When analysing the role of MIC-1/GDF-15 in pancreatic cancer (PC), at a molecular level it has been demonstrated to promote pancreatic cell invasion through its interaction with the transcription factor Twist1[25]. In the clinical domain, MIC-1/GDF-15 has been demonstrated to be elevated in the serum of PC patients compared to both healthy controls and those with benign pancreatic tumours, as well as being reported to be beneficial in the diagnosis of pancreatic adenocarcinoma[18,26]. While few individual studies show that MIC-1/GDF-15 is more sensitive than carbohydrate antigen 19-9 (CA19-9) in the diagnosis of PC, a meta-analysis[27] published in 2018 shows that MIC-1 has a comparable diagnostic accuracy to CA19-9 in diagnosis of PC. Further preliminary studies have demonstrated that MIC-1/GDF15 is superior to CA19-9 in differentiating PC from chronic pancreatitisand when used in combination with CA19-9 it improves further the diagnostic accuracy of differentiating PC form chronic pancreatitis and healthy controls[27-29]. A recent meta-analysis published the diagnostic sensitivity and specificity for MIC-1/GDF15 in diagnosing PC as 80% and 85% respectively, with an area under curve (AUC) of 0.894[27]. In addition to this, MIC-1/GDF15 was found to have a positive predictive value of 78.3%, and a negative predictive value of 78.6%[30,31].

In light of the current emerging evidence that advocates for MIC-1/GDF-15 as a potential serological marker of malignancy, the aim of this study was to determine the value of MIC-1/GDF-15 as a serological marker of pancreatic pre-malignant lesions and neoplastic tumours in an asymptomatic high-risk population being screened for pancreatic malignancy in an established PC screening program.

**MATERIALS AND METHODS**

Eligible participants were enrolled in the Australian Pancreatic Cancer Screening study for high-risk individuals performed at St Vincent’s Hospital in Sydney, Australia which had started in 2011. The study was approved by St Vincent’s Hospital Ethics Committee (HREC/10/SVH/33) and uses annual endoscopic ultrasound (EUS) as a screening modality. Asymptomatic individuals with a hereditary predisposition to PC were recruited between May 2011-May 2018 (Inclusion criteria Supplementary file 1). Participants were referred by Australian Family Cancer Clinics, the Australian Familial Pancreatic Cancer Registry, medical practitioners or participants had self-referred. At enrolment participants completed a questionnaire detailing past medical history, smoking and alcohol intake, and basic parameters such as height and weight. Participants were excluded from the study if they had a concurrent diagnosis of active malignancy or were not medically suitable for EUS (renal failure, congestive heart failure, human immunodeficiency virus) thus controlling for conditions that could have influenced MIC-1/GDF-15 level.

MIC-1/GDF15, CA19-9 and C-reactive protein (CRP) levels were determined on a fasting 10 mL blood sample collected from the participants at the time of EUS. CRP levels was used to control for inflammatory conditions that could have increased MIC-1/GDF15 level. When malignancy was detected, EUS fine need aspiration was performed. Participants in whom EUS demonstrated abnormalities but not malignancy were closely followed up with magnetic resonance imaging (MRI) or computed tomography (CT) (if claustrophobic) and repeat EUS in 3-6 mo as per study protocol. MIC-1, CRP and CA19-9 were repeated when a follow up EUS become abnormal.

Statistical analyses were performed using IBM SPSS statistics for Windows (Version 25.0. Armonk, NY). The baseline characteristics of the study population were stratified according to EUS findings: normal EUS, pancreatic cyst, branch-duct intraductal papillary mucinous neoplasm (BD-IPMN), diffuse non-specific abnormalities (*e.g.*, hyperechoic foci, strands, lobularity) and solid neoplastic tumours. Further analysis was then performed on those diagnosed with neoplastic tumours on EUS and subsequent MRI/CT.

Fisher’s exact test (2-tailed) was used to compare categorical characteristics between respective groups. Continuous baseline characteristics including age, body mass index (BMI), number of cigarettes smoked daily, weekly alcohol intake and age of drinking initiation were evaluated for an association with MIC-1/GDF-15 serum levels using Spearman rank correlation. An ANOVA test was used to compare normally distributed continuous variables, whereas a Kruskal-Wallis test was used to compare non-normally distributed continuous variables with two or more samples. Mann-Whitney U test was used to compare non-normally distributed continuous variables. A receiver operating characteristic curve (ROC) of MIC-1/GDF-15 was generated for its ability to determine the presence or absence of pancreatic cyst, BD-IPMN, diffuse non-specific abnormality or neoplastic tumours on EUS using serum levels adjusted for variables shown to either be significantly related to MIC-1/GDF-15 concentrations in this study, or have shown to correlate with MIC-1/GDF15 in previous studies. This included: age, gender, BMI, history of colonic polyps, smoking status, alcohol use, metformin use, past history of cancer, nonsteroidal anti-inflammatory drug (NSAID), and aspirin use. All analyses performed were 2-sided and statistical significance was defined as *P* < 0.05.

**RESULTS**

A total of 120 asymptomatic participants based on the EUS results were stratified as follows; (1) Normal EUS (*n* = 74, 61.2%) as the control group; (2) Pancreatic cyst (*n* = 25, 20.8%); (3) BD-IPMN (*n* = 9, 7.5%); (4) Diffuse non-specific abnormalities (*n* = 9, 7.5%); and (5) Solid neoplastic tumours (*n* = 3, 2.5% which included pancreatic adenocarcinoma, pancreatic neuroendocrine tumour and liver cancer), outlined in Table 1. Two further neoplastic tumours: one breast cancer and a bladder cancer were identified on further imaging (MRI pancreas and CT abdomen) performed for close monitoring of a diffusely abnormal pancreas.

***Study population characteristics***

Table 1 outlines the baseline characteristics of the 120 subjects. The median age of participants diagnosed with BD-IPMN on EUS was higher compared to their counterparts, however this was not statistically significant (*P* = 0.388). There was no significant difference in the number of FDR (*P =* 0.947) or SDR (*P* = 0.432) between groups. The median age of those diagnosed with neoplastic tumours on EUS was higher compared to those with a normal EUS, however this was not statistically significant (*P* = 0.519). Furthermore, those with neoplastic tumours identified on EUS had a higher median number of cigarettes smoked per week (Median = 20) compared to the other groups, however this was not significant (*P* = 0.929). Participants diagnosed with neoplasia on EUS had a higher serum MIC-1/GDF15 [Median = 849.1, interquartile range (IQR) = 604.9-849.1] compared to the other groups however this was not significant (*P* = 0.178) but approached significance when compared to participants with a normal EUS (*P* = 0.061) (Figure 1). Percentage change between serial MIC-1/GDF15 was not significant in those participants who had a normal EUS and subsequent abnormal EUS (tumour, BD-IPMN, cyst, diffuse abnormality) (*P* = 0.213). Median serum CA19-9 was greatest in patients with an EUS indicative of malignancy, this approached significance (*P* = 0.058) when compared to the other groups included in the analysis.

***Correlation of MIC-1/GDF-15 with population variables***

Baseline MIC-1/GDF-15 was significantly correlated with advancing age for the entire cohort (correlation coefficient = 0.602, *P* < 0.01) and age of youngest PC diagnosis (correlation coefficient = 0.223, *P* = 0.015). Increasing BMI did not correlate with increasing serum MIC-1/GDF15 (*P* = 0.548). The number of cigarettes smoked per day, and number of drinks per week did not correlate with increased baseline serum MIC-1/GDF15 values in this population *(P =* 0.138 and *P* = 0.451 respectively).

The total number of both first and second-degree relatives diagnosed with PC had a significant negative correlation with baseline serum MIC-1/GDF15 (correlation coefficient = -0.190, *P* = 0.038). The number of first degree relatives diagnosed with PC did not correlate with baseline serum MIC-1/GDF15 (*P* = 0.238), however the number of second degree relatives diagnosed with PC had a significant negative correlation with baseline serum MIC-1/GDF15 (correlation coefficient = -0.225, *P* = 0.014).

Baseline serum MIC-1/GDF15 did not correlate with gender (*P =* 0.176), BRCA2 status (*P* = 0.097), ethnicity (*P* = 0.570) or Jewish background (*P* = 0.606). Further analysis of dichotomous variables demonstrated that baseline serum MIC-1/GDF-15 was significantly greater in those with a history of cancer (*P* < 0.001), history of diabetes (*P* = 0.001), those taking oral hypoglycaemic medication (*P* = 0.001) and history of coronary artery disease (*P* = 0.005), hypercholesterolaemia (*P* = 0.013) and colon polyps (*P* = 0.005). Serum MIC-1/GDF-15 levels were elevated in those participants taking aspirin regularly (*P* = 0.019) and metformin (*P* = 0.001). Baseline serum MIC-1/GDF15 was not elevated in those with regular NSAID, folate or antidepressant use (*P* = 0.863, 0.928 and 0.172 respectively) in this study population.

***ROC curve for capacity of MIC-1/GDF-15 to identify premalignant lesions on EUS***

Baseline serum MIC-1/GDF-15 was a poor predictor of abnormal EUS in our cohort of asymptomatic high-risk patients as determined using a ROC curve for the capacity for MIC-1/GDF-15 to predict an abnormal EUS. The MIC-1/GDF-15 serum level, when adjusted for aspirin use, alcohol intake per week, smoking status, BMI, NSAID use, history of colonic polyps, gender, metformin use and age had an AUC of 0.576 (95%CI: 0.454-0.698) (*P* = 0.234) (Figure 2A). Similarly, baseline serum MIC-1/GDF-15 could not predict BD-IPMN (AUC = 0.644, 95%CI: 0.414-0.875, *P* = 0.223) (Figure 2B), pancreatic cyst (AUC = 0.347, 95%CI: 0.162-0.532, *P* = 0.131) (Figure 2C) and diffuse abnormalities (AUC = 0.510, 95%CI: 0.254-0.764, *P* = 0.935) (Figure 2D). In those with neoplastic tumours diagnosed on EUS and subsequent biopsy (*n* = 3), the AUC was 0.793, however this was not statistically significant (*P* = 0.081) (Figure 3).

***ROC curve for capacity of MIC-1/GDF-15 to identify neoplastic tumours on EUS and subsequent imaging MRI/CT***

Baseline MIC-1/GDF15 was a significant predictor of neoplastic tumours diagnosed on EUS and MRI/CT (*n* = 5) with an AUC=0.814 (95%CI: 0.657-0.970, *P* = 0.023) (Figure 4). In this asymptomatic cohort three neoplastic tumours were diagnosed on EUS and two other malignancies were diagnosed on further imaging performed to monitor the pancreas (one breast cancer on MRI pancreas and one bladder cancer on CT abdomen). In addition to this, median baseline serum MIC-1/GDF15 in asymptomatic patients found to have neoplastic tumours (Median = 1039.6, IQR = 727.0-1977.7) was significantly greater than benign lesions (Median = 570.1, IQR = 460.7-865.2) (*P* = 0.012) as demonstrated in Figure 5.

**DISCUSSION**

PC is a leading cause of cancer mortality worldwide, with a very poor survival rate due to late diagnosis, primarily due to symptoms presenting at advanced stages of the disease. The prognosis correlates strongly with pathological stage at the time of diagnosis, and despite advances in medicine in the last forty years, the 5-year survival has increased only from 4% to 7%[32]. As a result, efforts are made in detecting PC early at asymptomatic stage and multiple PC screening programs in high risk individuals have been established around the world. These screening programs target individuals with a genetic predisposition for developing PC (people with hereditary cancer syndromes due to known mutations and familial PC). Current screening modalities rely on pancreatic imaging (EUS and MRI) and biomarkers are at research level. Ideally, we need an early sensitive and specific serological marker that can be used as a first line screening tool in a high-risk population and help select cases that need further investigations, such as EUS or MRI. CA19-9 is not sensitive enough to be a marker for early detection of PC, has a specificity of 77%, sensitivity 75%, a positive predictive value of 0.5%-0.9%[33,34] and can be increased in other conditions such as biliary obstruction. Similarly, carcinoembryonic antigen has no utility in early detection of PC with a sensitivity and specificity of 65%[35].

MIC-1/GDF15 has been recently explored as a novel candidate tumour marker for PC with initial results proving to be elevated in the serum of patients with PC compared to healthy controls and those with benign lesions[17].As MIC-1 can be increased in other malignancies, studies report an increase in its diagnostic specificity if CA19-9 is used in combination with MIC-1[28,30]. In addition to this, serum MIC-1/GDF15 has been proven to be more sensitive than CA19-9 in detecting early-stage PC. Importantly, MIC-1 had a sensitivity of 63.1% in detecting patients with CA19-9-negative PC[26].

In this feasibility prospective cohort study in an asymptomatic population at high risk of developing PC undertaking yearly screening with EUS, serum baseline MIC-1/GDF-15 was shown to be a significant predictor of neoplastic tumours (both pancreatic and extra-pancreatic) after ROC curve analysis, with an AUC of 0.814 (*P* = 0.023). In addition, those diagnosed with neoplastic tumours on EUS or MRI/CT had a higher median baseline MIC-1/GDF15 compared to those diagnosed with benign lesions on EUS. Baseline serum MIC-1/GDF-15 had a significant positive correlation with advancing age and age of PC diagnosis in family members. Further analysis of the screening cohort demonstrated that serum MIC-1/GDF-15 was elevated in those with a family history of cancer, history of diabetes, current metformin use and those with previous colonic polyps.

When evaluating the utility of serum baseline MIC-1/GDF15 comparing to EUS results only, using ROC curve analysis, we found that it was best utilised when used in participants who were diagnosed with solid neoplastic tumours or BD-IPMN on EUS, with AUCs of 0.793 and 0.644 respectively, with solid tumours diagnosed on EUS approaching significance despite having only 3 cases. These results demonstrated that MIC-1/GDF15 is elevated in participants with pre-malignant and neoplastic tumours, and seems to bear similar predictive value to prostate-specific antigen testing for prostate cancer and the faecal occult blood test for colonic adenoma[36-39]. Previously Koopmann *et al*[18] were able to demonstrate an AUC for MIC-1/GDF15 of 0.81 for the detection of pancreatic adenocarcinoma, and when used in combination with CA19-9, this increased to 0.87.

Compared with previous studies that evaluate the role of MIC- 1 in patients with known PC or other malignancies our study design is unique. This is a pilot study, the first to the authors knowledge, to evaluate serum MIC-1/GDF15 in an asymptomatic population at high risk of malignancy in an established PC screening program. Based on the inclusion criteria (patients with a genetic predisposition for PC) these participants are at risk of developing other malignancies not just pancreatic, as shown in our cohort where three non-pancreatic malignancies were found at an asymptomatic stage (liver, breast and bladder cancer). This study shows that baseline MIC 1 is elevated in patient with neoplastic tumours and could be potentially used to guide further investigations such as MRI or CT if EUS is negative for PC.

The authors acknowledge that due to the nature of the screening program, the recruitment of asymptomatic high-risk participants is time intensive and the subsequent low incidence of abnormal EUS results and malignant lesions are two limitations of this prospective study. Further larger prospective multi-centre cohort studies are required to further assess the value of MIC-1 in screening for malignancy in this type of cohort.

The authors echo the findings of Wang *et al*[40] who stated that serum MIC-1/GDF15 should be interpreted cautiously due to the potential for a broad range of values in the general population and the need to control for multiple confounding factors, particularly inflammation promoting an elevated MIC-1 serum level. We controlled for conditions that influence MIC-1 levels by using CRP as marker of active inflammation and excluding patients with congestive heart failure, renal failure, human immunodeficiency virus and known malignancy.

Although this study was not able to detect a significant change in serum MIC-1/GDF-15 in participants who had a normal then subsequent abnormal EUS , further studies should endeavour to explore whether percentage change in MIC-1/GDF15 is indicative of tumorigenesis in populations at high risk for developing cancer.

A limitation of the use of MIC-1/GDF15 as a biomarker is a wide normal serum range. Serial monitoring of an individual’s MIC-1/GDF15 serum level would identify those with increasing levels, even those that were within the normal range. It is the aim of this screening program to implement serial serum MIC-1/GDF15 to assess if with a large enough sample size and long-term follow-up, a statistically significant result can be achieved.

Future studies should aim to further evaluate and analyse MIC-1/GDF15 in both the general population and in patients at risk of malignancy due to a genetic predisposition to determine how this serum biomarker can be better applied in the clinical setting with intention to facilitate its progressive implementation regularly in the clinical domain, along with being further assessed in the academic setting[40].

In conclusion, this pilot study, the first of its kind to implement MIC-1/GDF15 as a screening tool in an asymptomatic population with a genetic predisposition of developing PC, provides moderate support to the previous findings that MIC-1/GDF15 is elevated in patients with neoplastic tumours, however the sample size used to assess this was small. In addition, this study highlights that an elevated MIC-1/GDF15 in the context of a negative pancreatic EUS in a high risk of malignancy cohort may warrant further investigation to determine whether an occult malignancy exists.

While population based screening is difficult to implement due to wide range of normal values and its elevation in select disease processes, MIC-1 might be better suited for screening for malignancy in patients with hereditary cancer syndromes where baseline and serial measurement can be used in combination with other validated serological markers to overcome many of these limitations and potentially select patients who require further investigations.

Larger multicentric prospective studies are required to further define the role of MIC-1/GDF15 as a serological biomarker in pre-malignant pancreatic lesions and neoplastic tumours.

**ARTICLE HIGHLIGHTS**

***Research background***

Early detection of pancreatic cancer (PC) is a key priority in order to improve survival. Macrophage inhibitory cytokine-1 or growth differentiation factor-15 (MIC-1/GDF15) is a novel candidate tumour marker for PC with initial results proving to be elevated in the serum of patients with PC compared to healthy controls and those with benign lesions.

***Research motivation***

We need an early sensitive and specific serological marker that can be used as a first line screening tool in patients at risk of PC and help select cases that need further investigations, such as endoscopic ultrasound (EUS) or magnetic resonance imaging. This study evaluates the role of MIC-1 in patients at high risk of developing PC.

***Research objectives***

This is a pilot study to determine the role of MIC-1 in detecting pre-malignant pancreatic lesions and neoplastic tumours in an asymptomatic high-risk cohort part of Australian Pancreatic Cancer Screening Program and correlate with imaging finding.

***Research methods***

Participants recruited for yearly surveillance with EUS had serial fasting blood samples collected for MIC-1, C-reactive protein and carbohydrate antigen 19-9. Patients were stratified into five groups based on EUS findings. MIC-1 serum levels were quantified using ELISA and correlations of MIC-1 with population variables and imaging findings were performed. A receiver operating characteristic curve of MIC-1 was generated for its ability to determine the presence or absence of neoplastic tumours , pancreatic cysts, branch-duct intraductal papillary mucinous neoplasm and diffuse non-specific abnormality using serum levels adjusted for variables shown to either be significantly related to MIC-1/GDF-15 concentrations in this study, or have shown to correlate with MIC-1/GDF15 in previous studies.

***Research results***

One hundred twenty participants were recruited over 8 years. Baseline serum MIC-1 was a significant predictor of neoplastic tumours on receiver operating characteristic curve analysis. Baseline serum MIC-1 had moderate predictive capacity for branch-duct intraductal papillary mucinous neoplasm (AUC = 0.644) and neoplastic tumours noted on EUS (AUC = 0.793), however this was not significant (*P* = 0.188 and 0.081 respectively). Serial serum MIC-1/GDF15 did not demonstrate a significant percentage change between a normal and abnormal EUS. Median baseline MIC-1/GDF15 was greater in those with neoplastic tumours compared to those diagnosed with a benign lesion.

***Research conclusions***

MIC-1 has predictive capacity for neoplastic tumours in asymptomatic individuals with a genetic predisposition for PC. Further imagining may be warranted in patients with raised serum MIC-1 and abnormal EUS.

***Research perspectives***

This pilot study is the first of its kind to implement MIC-1 as a screening tool in an asymptomatic population with a genetic predisposition of developing PC. Our study is a feasibility study and we hope our results will start a new wave of research (larger, multicentric, prospective trials) into investigating the role of this biomarker in early detection of neoplastic tumours to validate our finding and provide further characterisation of this biomarker.

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**Footnotes**

**Institutional review board statement:** The study protocol was reviewed and approved by the Ethics Committees of St Vincent’s Hospital, Sydney, NSW, Australia.

**Clinical trial registration statement:** This is not a clinical trial and therefore does not need to be registered under the clinical trials. Registration applies only to randomised control trials. There is no control arm, nor health intervention, the study looks at a new biomarker in an established pancreatic screening program. Results are not given to the patients and there is no intervention.

**Informed consent statement:** All study participants provided written informed consent prior to study enrolment.

**Conflict-of-interest statement:** The authors declare no competing interests.

**Data sharing statement:** There is no additional data available.

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**Figure Legends**

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**Figure 1** **Boxplot of baseline medium serum macrophage inhibitory cytokine-1** o**r growth differentiation factor-15 levels by group with 95% confidence interval errors bars in participants with a normal endoscopic ultrasound, branched duct intraductal papillary mucinous neoplasm, pancreatic cyst, diffuse abnormality and neoplastic tumours/malignancy detected by endoscopic ultrasound.** BD-IPMN: Branched duct intraductal papillary mucinous neoplasm; EUS: Endoscopic ultrasound; MIC-1/GDF15: Macrophage inhibitory cytokine-1 or growth differentiation factor-15.

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**Figure 2 Receiver operating characteristic curve generated for the capacity of macrophage inhibitory cytokine-1 or growth differentiation factor-15 to predict abnormal endoscopic ultrasound results.** A: Abnormal endoscopic ultrasound (AUC = 0.576, 95%CI: 0.454-0.698, *P* = 0.234); B: Branched duct intraductal papillary mucinous neoplasm (AUC = 0.664, 95%CI: 0.414-0.875, *P* = 0.223); C: Pancreatic cyst (AUC = 0.347, 95%CI: 0.162-0.532, *P* = 0.131); D: Diffuse abnormality (AUC = 0.510, 95%CI: 0.254-0.764, *P* = 0.935). ROC: Receiver operating characteristic.



**Figure 3 Receiver operating characteristic curve generated for the capacity of macrophage inhibitory cytokine-1 or growth differentiation factor-15 to predict solid neoplastic tumours on endoscopy ultrasound (AUC = 0.793, *P =* 0.081, *n* = 3).** ROC: Receiver operating characteristic.

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**Figure 4 Receiver operating characteristic curve generated for the capacity of macrophage inhibitory cytokine-1 or growth differentiation factor-15 to predict solid neoplastic tumours identified on endoscopy ultrasound and magnetic resonance imaging or computed tomography in an asymptomatic population (AUC = 0.814, 95%CI: 0.657-0.970, *P* = 0.023, *n* = 5).** ROC: Receiver operating characteristic.

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**Figure 5 Boxplot of median baseline macrophage inhibitory cytokine-1 or growth differentiation factor-15 in participants diagnosed with benign pancreatic abnormalities (*n* = 42) and solid neoplastic tumours (*n* = 5) lesions on endoscopy ultrasound and magnetic resonance imaging or computed tomography.** MIC-1/GDF15: Macrophage inhibitory cytokine-1 or growth differentiation factor-15.

**Table 1 Characteristics of participants in pancreatic cancer screening program based on endoscopic ultrasound results**

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| **Baseline characteristics** | **Normal EUS (*n* = 74)** | **Pancreatic Cyst (*n* = 25)** | **BD-IPMN (*n* = 9)** | **Diffuse Abnormality (*n* = 9)** | **Neoplastic tumours on EUS (*n* = 3)** | ***P-*value** |
| Age (yr), mean (SD) | 55.0 (9.8) | 57.3 (7.9) | 60.1 (10.0) | 59.3 (8.8) | 57.7 (4.5) | 0.388 |
| Age Quartile, *n* (%) |  |  |  |  |  |  |
| Quartile 1 (35-50) | 23 (31.1) | 5 (20.0) | 1 (11.1) | 2 (22.2) | 0 (0.0) |  |
| Quartile 2 (51-56) | 17 (23.0) | 7 (28.0) | 3 (33.3) | 1 (11.1) | 1 (33.3) |  |
| Quartile 3 (57-63) | 20 (27.0) | 6 (24.0) | 2 (22.2) | 3 (33.3) | 2 (66.7) |  |
| Quartile 4 (64-78) | 14 (18.9) | 7 (28.0) | 3 (33.3) | 3 (33.3) | 0 (0.0) |  |
| BMI, mean (SD) | 27.3 (5.2) | 27.8 (5.4) | 26.8 (4.2) | 31.6 (3.4) | 24.0 (5.2) | 0.117 |
| BMI Quartile, *n* (%) |  |  |  |  |  | 0.0131 |
| Quartile 1 (19.5-23.8) | 18 (24.3) | 6 (24.0) | 4 (44.4) | 0 (0.0) | 2 (66.7) |  |
| Quartile 2 (23.9-27.2) | 22 (29.7) | 8 (32.0) | 0 (0.0) | 0 (0.0) | 0 (0.0) |  |
| Quartile 3 (27.3-30.4) | 18 (24.3) | 6 (24.0) | 2 (22.2) | 3 (33.3) | 1 (33.3) |  |
| Quartile 4 (30.5-46.7) | 16 (21.6) | 5 (20.0) | 3 (33.3) | 6 (66.7) | 0 (0.0) |  |
| Gender, *n* (%) |  |  |  |  |  | 0.362 |
| Female | 51 (68.9) | 18 (72.0) | 5 (55.6) | 4 (44.4) | 1 (33.3) |  |
| Male | 23 (31.1) | 7 (28.0) | 4 (44.4) | 5 (55.6) | 2 (66.7) |  |
| BRCA2 positive, *n* (%) | 10 (13.5) | 7 (28.0) | 0 (0.0) | 3 (33.3) | 2 (66.7) | 0.0321 |
| First Degree Relatives with PC, *n* (%) |  |  |  |  |  | 0.947 |
| 0 | 3 (4.1) | 0 (0.0) | 0 (0.0) | 0 (0.0) | 0 (0.0) |  |
| 1 | 43 (58.1) | 16 (64.0) | 4 (44.4) | 5 (55.6) | 2 (66.7) |  |
| 2 | 21 (28.4) | 5 (20.0) | 5 (55.6) | 4 (44.4) | 1 (33.3) |  |
| 3 | 7 (9.5) | 4 (16.0) | 0 (0.0) | 0 (0.0) | 0 (0.0) |  |
| Second Degree Relative with PC, *n* (%) |  |  |  |  |  | 0.432 |
| 0 | 23 (31.1) | 9 (36.0) | 5 (55.6) | 1 (11.1) | 2 (66.7) |  |
| 1 | 17 (23.0) | 9 (36.0) | 0 (0.0) | 7 (77.8) | 1 (33.3) |  |
| 2 | 20 (27.0) | 3 (12.0) | 2 (22.2) | 1 (11.1) | 0 (0.0) |  |
| 3 | 8 (10.8) | 3 (12.0) | 2 (22.2) | 0 (0.0) | 0 (0.0) |  |
| 4 | 6 (8.1) | 1 (4.0) | 0 (0.0) | 0 (0.0) | 0 (0.0) |  |
| Youngest PC diagnosis, median (IQR)  | 50 (44-64.5) | 60 (46-66) | 65 (45.5-68.5) | 53 (38-70) | 75 (22-75) | 0.519 |
| Ethnicity, *n* (%) |  |  |  |  |  | 0.848 |
| Asian | 1 (1.4) | 1 (4.0) | 0 (0.0) | 0 (0.0) | 0 (0.0) |  |
| Caucasian | 70 (94.6) | 24 (96.0) | 9 (100.0) | 9 (100.0) | 3 (100.0) |  |
| Other | 3 (4.1) | 0 (0.0) | 0 (0.0) | 0 (0.0) | 0 (0.0) |  |
| Jewish Origin, *n* (%) | 5 (6.8) | 7 (28.0) | 1 (11.1) | 1 (11.1) | 0 (0.0) | 0.079 |
| Ashkenazi  | 5 (7.4) | 6 (24.0) | 0 (0.0) | 1 (11.1) | 0 (0.0) | 0.121 |
| Medical History |  |  |  |  |  |  |
| Personal History of cancer, *n* (%) | 13 (17.6) | 5 (20.0) | 3 (33.3) | 4 (44.4) | 1 (33.3) | 0.350 |
| Diabetes, *n* (%) | 4 (5.4) | 1 (4.0) | 1 (11.1) | 2 (22.2) | 0 (0.0) | 0.434 |
| Insulin, *n* (%) | 0 (0.0) | 0 (0.0) | 0 (0.0) | 1 (14.3) | 0 (0.0) | 0.184 |
| Oral hypoglycaemic medication, *n* (%) | 4 (7.4) | 3 (16.7) | 1 (16.7) | 1 (14.3) | 0 (0.0) | 0.840 |
| Smoking status, *n* (%) |  |  |  |  |  | 0.188 |
| Never smoked | 32 (47.8) | 17 (68.0) | 5 (55.6) | 6 (66.7) | 2 (66.7) |  |
| Stopped smoking | 32 (47.8) | 7 (28.0) | 4 (44.4) | 3 (33.3) | 0 (.0) |  |
| Still smoking | 3 (4.5) | 1 (4.0) | 0 (.0) | 0 (.0) | 1 (33.3) |  |
| Cigarettes per day, Median (IQR) | 13.5 (6.0-20.0) | 12.5 (6.3-23.8) | 12.0 (1.0-12.0) | 10.0 (5.0-10.0) | 20.0 (20.0-20.0) | 0.929 |
| Cigarettes per day quartile, *n* (%) |  |  |  |  |  | 0.963 |
| Quartile 1 (1-6) | 11 (30.6) | 2 (25.0) | 1 (33.3) | 1 (33.3) | 0 (0.0) |  |
| Quartile 2 (7-12) | 7 (19.4) | 2 (25.0) | 1 (33.3) | 1 (33.3) | 0 (0.0) |  |
| Quartile 3 (15-20) | 14 (38.9) | 2 (25.0) | 0 (0.0) | 0 (0.0) | 1 (100.0) |  |
| Quartile 4 (25-75) | 4 (11.1) | 2 (25.0) | 1 (33.3) | 1 (33.3) | 0 (0.0) |  |
| Years Smoking, *n* (%) |  |  |  |  |  | 0.629 |
| < 10  | 12 (33.3) | 3 (37.5) | 2 (50.0) | 1 (33.3) | 0 (0.0) |  |
| 11-20 | 11 (30.6) | 3 (37.5) | 0 (0.0) | 1 (33.3) | 0 (0.0) |  |
| 21-30 | 8 (22.2) | 1 (12.5) | 2 (50.0) | 1 (33.3) | 0 (0.0) |  |
| 31-40 | 4 (11.1) | 1 (12.5) | 0 (0.0) | 0 (0.0) | 1 (100.0) |  |
| 41-50 | 1 (2.8) | 0 (0.0) | 0 (0.0) | 0 (0.0) | 0 (0.0) |  |
| > 50 | 12 (33.3) | 3 (37.5) | 2 (50.0) | 1 (33.3) | 0 (0.0) |  |
| Alcohol consumption, *n* (%) |  |  |  |  |  | 0.209 |
| Daily | 19 (25.7) | 7 (28.0) | 0 (0.0) | 3 (33.3) | 2 (66.7) |  |
| Weekly | 14 (18.9) | 5 (20.0) | 1 (11.1) | 2 (22.2) | 0 (0.0) |  |
| Social | 5 (6.8) | 5 (20.0) | 2 (22.2) | 2 (22.2) | 1 (33.3) |  |
| No history of chronic consumption | 36 (48.6) | 8 (32.0) | 6 (66.7) | 2 (22.2) | 0 (0.0) |  |
| Drinks per week, Median (IQR) | 6.0 (3.0-15.0) | 4 (2.0-10.0) | 2.5 (1.0-6.0) | 6.0 (1.0-15.0) | 21.0 (1.0-21.0) | 0.331 |
| Drinks per week quartile, *n* (%) |  |  |  |  |  | 0.328 |
| Quartile 1 (1 - 3) | 16 (25.8) | 6 (31.6) | 5 (62.5) | 2 (28.6) | 1 (33.3) |  |
| Quartile 2 (4 - 6) | 19 (30.6) | 4 (21.1) | 2 (25.0) | 2 (28.6) | 0 (0.0) |  |
| Quartile 3 (7 - 14) | 11 (17.7) | 7 (36.8) | 0 (0.0) | 1 (14.3) | 0 (0.0) |  |
| Quartile 4 (15 - 35) | 16 (25.8) | 2 (10.5) | 1 (12.5) | 2 (28.6) | 2 (66.7) |  |
| Age of first drink, Median (IQR) | 18.0 (17.0-18.0) | 20.0 (18.0-25.0) | 19.0 (18.0-21.0) | 17.0 (15.0-20.0) | 18.0 (15.0-18.0) | 0.0331 |
| Years drinking, *n* (%) |  |  |  |  |  | 0.129 |
| < 10  | 2 (3.4) | 2 (11.8) | 0 (0.0) | 0 (0.0) | 0 (0.0) |  |
| 11-20 | 11 (18.6) | 3 (17.6) | 3 (37.5) | 0 (0.0) | 0 (0.0) |  |
| 21-30 | 13 (22.0) | 6 (35.3) | 0 (0.0) | 2 (28.6) | 1 (33.3) |  |
| 31-40 | 22 (37.3) | 5 (29.4) | 2 (25.0) | 1 (14.3) | 2 (66.7) |  |
| 41-50 | 8 (13.6) | 1 (5.9) | 2 (25.0) | 2 (28.6) | 0 (0.0) |  |
| > 50 | 2 (3.4) | 0 (0.0) | 1 (12.5) | 2 (28.6) | 0 (0.0) |  |
| Biochemistry |  |  |  |  |  |  |
| CRP, Median (IQR) | 1.3 (0.6-2.5) | 1.7 (0.7-4.2) | 1.4 (0.5-1.9) | 0.8 (0.6-4.4) | 0.8 (0.3-0.8) | 0.835 |
| CA19-9, Median (IQR) | 9.0 (6.0-16.0) | 9.0 (7.0-15.8) | 9.0 (5.7-15.0) | 16.0 (8.5-19.5) | 47.0 (22.0-47.0) | 0.058 |
| MIC-1/GDF-15, Median (IQR) | 558.2 (449.6-715.3) | 574.3 (448.5-830.3) | 659.3 (484.2-1077.3) | 553.2 (512.9-967.0) | 849.1 (604.9- 849.1) | 0.178 |

Quartiles were created using the entire cohort, which were split into 4 groups for the appropriate measurements. Percentages for variables such as cigarettes, drinking *etc*. are CUMULATIVE, *i.e.*, ignores variables which did not have a number, presumably because the patient doesn’t drink/smoke. Biochemistry of MIC-1 is at baseline. An ANOVA test was used to compare normally distributed continuous variables, whereas a Kruskal-Wallis test was used to comparing ordinal and non-normally distributed continuous variables. A Fisher’s exact test (2-tailed) was used to compare dichotomous variables. 1Denotes statistical significance. EUS: Endoscopic ultrasound; BD-IPMN: Branch duct intraductal mucinous papillary neoplasia; BMI: Body mass index; PC: Pancreatic cancer; IQR: Interquartile range; CRP: C-reactive protein; CA19-9: Cancer antigen 19-9.