

World Journal of *Gastroenterology*

World J Gastroenterol 2020 May 14; 26(18): 2126-2275



OPINION REVIEW

- 2126 Annexin A2 promotion of hepatocellular carcinoma tumorigenesis *via* the immune microenvironment
Qiu LW, Liu YF, Cao XQ, Wang Y, Cui XH, Ye X, Huang SW, Xie HJ, Zhang HJ

REVIEW

- 2138 Medical management of metabolic and cardiovascular complications after liver transplantation
Becchetti C, Dirchwolf M, Banz V, Dufour JF
- 2155 Natural products that target macrophages in treating non-alcoholic steatohepatitis
Li CL, Zhou WJ, Ji G, Zhang L

MINIREVIEWS

- 2166 Hepatitis B virus recurrence after liver transplantation: An old tale or a clear and present danger?
Lenci I, Milana M, Grassi G, Manzia TM, Gazia C, Tisone G, Angelico R, Baiocchi L
- 2177 New role for ceramide in hypoxia and insulin resistance
Xia QS, Lu FE, Wu F, Huang ZY, Dong H, Xu LJ, Gong J
- 2187 Role of gut microbiota on intestinal barrier function in acute pancreatitis
Li XY, He C, Zhu Y, Lu NH

ORIGINAL ARTICLE**Basic Study**

- 2194 Genetic association analysis of *CLEC5A* and *CLEC7A* gene single-nucleotide polymorphisms and Crohn's disease
Elleisy N, Rohde S, Huth A, Gittel N, Glass Á, Möller S, Lamprecht G, Schäffler H, Jaster R
- 2203 Folic acid attenuates high-fat diet-induced steatohepatitis *via* deacetylase SIRT1-dependent restoration of PPAR α
Xin FZ, Zhao ZH, Zhang RN, Pan Q, Gong ZZ, Sun C, Fan JG

Retrospective Study

- 2221 Prognostic significance of hepatic encephalopathy in patients with cirrhosis treated with current standards of care
Bohra A, Worland T, Hui S, Terbah R, Farrell A, Robertson M
- 2232 Optimal proximal resection margin distance for gastrectomy in advanced gastric cancer
Kim A, Kim BS, Yook JH, Kim BS

META-ANALYSIS

- 2247** Computed tomography *vs* liver stiffness measurement and magnetic resonance imaging in evaluating esophageal varices in cirrhotic patients: A systematic review and meta-analysis
Li Y, Li L, Weng HL, Liebe R, Ding HG

CASE REPORT

- 2268** Ectopic hepatocellular carcinoma mimicking a retroperitoneal tumor: A case report
Adachi Y, Hayashi H, Yusa T, Takematsu T, Matsumura K, Higashi T, Yamamura K, Yamao T, Imai K, Yamashita Y, Baba H

ABOUT COVER

Associate Editor of *World Journal of Gastroenterology*, Mark De Ridder, MD, PhD, Chairman, Full Professor, Department of Radiotherapy, UZ Brussel, Vrije Universiteit Brussel, Brussels 1090, Belgium

AIMS AND SCOPE

The primary aim of *World Journal of Gastroenterology* (*WJG*, *World J Gastroenterol*) is to provide scholars and readers from various fields of gastroenterology and hepatology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJG mainly publishes articles reporting research results and findings obtained in the field of gastroenterology and hepatology and covering a wide range of topics including gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, gastrointestinal oncology, and pediatric gastroenterology.

INDEXING/ABSTRACTING

The *WJG* is now indexed in Current Contents®/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports®, Index Medicus, MEDLINE, PubMed, PubMed Central, and Scopus. The 2019 edition of Journal Citation Report® cites the 2018 impact factor for *WJG* as 3.411 (5-year impact factor: 3.579), ranking *WJG* as 35th among 84 journals in gastroenterology and hepatology (quartile in category Q2). CiteScore (2018): 3.43.

RESPONSIBLE EDITORS FOR THIS ISSUE

Responsible Electronic Editor: *Yan-Liang Zhang*
 Proofing Production Department Director: *Yun-Xiaojuan Wu*
 Responsible Editorial Office Director: *Ze-Mao Gong*

NAME OF JOURNAL

World Journal of Gastroenterology

ISSN

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

LAUNCH DATE

October 1, 1995

FREQUENCY

Weekly

EDITORS-IN-CHIEF

Subrata Ghosh, Andrzej S Tarnawski

EDITORIAL BOARD MEMBERS

<http://www.wjgnet.com/1007-9327/editorialboard.htm>

PUBLICATION DATE

May 14, 2020

COPYRIGHT

© 2020 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

<https://www.wjgnet.com/bpg/gerinfo/204>

GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjgnet.com/bpg/GerInfo/287>

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjgnet.com/bpg/gerinfo/240>

PUBLICATION ETHICS

<https://www.wjgnet.com/bpg/GerInfo/288>

PUBLICATION MISCONDUCT

<https://www.wjgnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

<https://www.wjgnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjgnet.com/bpg/GerInfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>

New role for ceramide in hypoxia and insulin resistance

Qing-Song Xia, Fu-Er Lu, Fan Wu, Zhao-Yi Huang, Hui Dong, Li-Jun Xu, Jing Gong

ORCID number: Qing-Song Xia (0000-0002-8948-2573); Fu-Er Lu (0000-0003-3372-4024); Fan Wu (0000-0003-3938-493X); Zhao-Yi Huang (0000-0003-4234-3872); Hui Dong (0000-0003-4771-0846); Li-Jun Xu (0000-0002-2013-9419); Jing Gong (0000-0003-0757-212).

Author contributions: Xia QS and Wu F performed the literature search and drafted the manuscript; Huang ZY designed the figures; Dong H and Xu LJ summarized the literature; Gong J and Lu FE supervised and approved the final version of the manuscript.

Supported by the National Natural Science Foundation of China, No. 81904158; TCM Modernization Research of National Key Research and Development Program, No. 2018YFC1704202.

Conflict-of-interest statement:

There is no conflict of interest associated with any of the senior author or other coauthors who contributed their efforts in this manuscript.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited

Qing-Song Xia, Fu-Er Lu, Fan Wu, Zhao-Yi Huang, Hui Dong, Li-Jun Xu, Jing Gong, Department of Integrated Traditional Chinese and Western Medicine, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan 430030, Hubei Province, China

Corresponding author: Jing Gong, MD, Doctor, Department of Integrated Traditional Chinese and Western Medicine, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, No. 1095, Jiefang Road, Wuhan 430030, Hubei Province, China. jgongtcm@126.com

Abstract

Ceramides are significant metabolic products of sphingolipids in lipid metabolism and are associated with insulin resistance and hepatic steatosis. In chronic inflammatory pathological conditions, hypoxia occurs, the metabolism of ceramide changes, and insulin resistance arises. Hypoxia-inducible factors (HIFs) are a family of transcription factors activated by hypoxia. In hypoxic adipocytes, HIF-1 α upregulates *pla2g16* (a novel HIF-1 α target gene) gene expression to activate the NLRP3 inflammasome pathway and stimulate insulin resistance, and adipocyte-specific *Hif1a* knockout can ameliorate homocysteine-induced insulin resistance in mice. The study on the HIF-2 α – NEU3 – ceramide pathway also reveals the role of ceramide in hypoxia and insulin resistance in obese mice. Under obesity-induced intestinal hypoxia, HIF-2 α increases the production of ceramide by promoting the expression of the gene *Neu3* encoding sialidase 3, which is a key enzyme in ceramide synthesis, resulting in insulin resistance in high-fat diet-induced obese mice. Moreover, genetic and pathophysiologic inhibition of the HIF-2 α – NEU3 – ceramide pathway can alleviate insulin resistance, suggesting that these could be potential drug targets for the treatment of metabolic diseases. Herein, the effects of hypoxia and ceramide, especially in the intestine, on metabolic diseases are summarized.

Key words: Ceramide; Intestinal hypoxia; Insulin resistance; Diabetes mellitus; Hypoxia-inducible factors; Obesity

©The Author(s) 2020. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Hypoxia is an essential risk factor that promotes insulin resistance in a variety of tissues, such as adipocytes, intestines, and the liver. In hypoxic adipocytes, hypoxia-inducible factor-1 α upregulates *pla2g16* gene expression to activate the NLRP3 inflammasome pathway, leading to insulin resistance. In obese animals or people, increased ceramide further results in insulin resistance under hypoxia. In intestinal epithelial cells, hypoxia-inducible factor-2 α is activated and accumulates under hypoxia

manuscript

Received: December 30, 2019**Peer-review started:** December 30, 2019**First decision:** March 27, 2020**Revised:** April 8, 2020**Accepted:** April 24, 2020**Article in press:** April 24, 2020**Published online:** May 14, 2020**P-Reviewer:** Balaban YH,
Tomizawa M**S-Editor:** Zhang L**L-Editor:** Wang TQ**E-Editor:** Zhang YL

in high-fat diet-fed mice, which upregulates the target gene *Neu3*, accelerating the process of insulin resistance.

Citation: Xia QS, Lu FE, Wu F, Huang ZY, Dong H, Xu LJ, Gong J. New role for ceramide in hypoxia and insulin resistance. *World J Gastroenterol* 2020; 26(18): 2177-2186

URL: <https://www.wjnet.com/1007-9327/full/v26/i18/2177.htm>

DOI: <https://dx.doi.org/10.3748/wjg.v26.i18.2177>

INTRODUCTION

Diabetes mellitus is caused by abnormal secretion or utilization of insulin, resulting in disorders of carbohydrate, protein, and fat metabolism. Hyperglycemia is the primary symptom that can induce visual lesions and impair the kidney, heart, brain, and other organs. Diabetes is characterized by high morbidity and mortality, which brings serious economic and medical burdens to modern society. According to the latest report of the International Diabetes Federation, there were 463 million patients with diabetes in the world in 2019, which is expected to reach 700 million in 2045 at a growth rate of 51%. According to the Global Burden of Disease Study 2013, in 2013, globally, 1.47 million people died because of diabetes and its complications^[1]. In 2019, the International Diabetes Federation estimates that 4.2 million people worldwide died from diabetes every year, which was one of the three major causes of noncommunicable diseases worldwide^[2].

Insulin resistance is another essential clinical feature of diabetes mellitus. Both weight gain and obesity are important risk factors for metabolic diseases such as type 2 diabetes mellitus (T2DM) and nonalcoholic fatty liver disease (NAFLD)^[3]. It is universally known that low-grade inflammation, abnormal glucose and lipid metabolism, endoplasmic reticulum stress, and oxidative stress are involved in insulin resistance^[4]. Recently, it was reported that the regulator of hypoxia [hypoxia-inducible factor (HIF)] and the corresponding changes in lipid metabolism, especially ceramide, promote the progression of insulin resistance and NAFLD^[5].

HYPOXIA-INDUCIBLE FACTOR IN HYPOXIA

Normoxia refers to physiological oxygen levels (PO_2) in normal tissue in a healthy state, but the oxygen content of different tissues varies in the physiological state, creating a wider range of oxygen levels (range from 13 kPa in the pulmonary vein to 2.7 kPa in the interstitial spaces), such as the intestinal mucosa PO_2 being significantly lower than that of the lung mucosa^[6]. Hypoxia refers to the phenomenon of insufficient oxygen in tissues or blood relative to physiological conditions.

HIF is a pivotal intracellular transcriptional regulator in response to hypoxia in metazoan development, physiology, and disease pathogenesis^[7]. Most species that breathe oxygen express the highly conserved transcription complex HIF-1^[8]. HIF-1, a heterodimer composed of an alpha and a beta subunit, belongs to the Per-Arnt-Sim (PAS) subfamily of the basic helix-loop-helix (bHLH) family of transcription factors. The structure of HIF consists of the following three parts: An N-terminal basic helix-loop-helix domain for deoxyribonucleic acid binding, a central region PAS domain that facilitates heterodimerization, and a C-terminus for recruitment of transcriptional coregulatory proteins^[9]. There are six members of the human HIF family: HIF-1 α , HIF-1 β , HIF-2 α , HIF-2 β , HIF-3 α , and HIF-3 β . Many cells express HIF-1 α and HIF-2 α , especially intestinal epithelial cells^[10].

There are two major regulatory mechanisms under normoxia. One way is the degradation of HIF α protein. Hydroxylated by the prolyl hydroxylase domain, HIF α binds to the E3 ubiquitin ligase complex containing the von Hippel-Lindau disease tumor suppressor protein, resulting in expeditious degradation of HIF α . The other way is suppression of transcriptional activity. After hydroxylation by HIF α asparaginyl residue with factor inhibiting HIF1 enzyme, the interaction of HIF α with the transcriptional coactivator cAMP-response element binding protein-binding protein and histone acetyltransferase p300 is incapacitated, thus impeding transcription. However, in hypoxia, HIF α subunits remain stabilized and are not hydroxylated by prolyl hydroxylase domain and factor inhibiting HIF1, which are O_2 -dependent oxygenases, resulting in the accumulation of HIF α and the upregulation of

target gene expression^[5,11].

HYPOXIA AND INSULIN RESISTANCE

Metabolic syndrome is a clustering of central obesity, insulin resistance, dysglycemia, and a proatherogenic plasma lipid profile, which are associated with the risk of developing cardiovascular disease and T2DM and are presumably caused by chronic inflammation^[12,13]. Low-grade inflammation was due to hypoxia, lipids and metabolites, reactive oxygen species, and endoplasmic reticulum stress^[4]. At the onset of obesity, resident M2 macrophages contribute to tissue and vascular remodeling to help adipocytes accommodate the new environment of overnutrition to protect adipose tissue from hypoxia and ischemia. Owing to the imbalance between M1 macrophages and M2 macrophages (decrease in protective M2 macrophages and increase in deleterious M1 macrophages), obesity promotes the development of hypoxia in adipose tissue. Moreover, M1 macrophages generate reactive oxygen species and nitrogen monoxide (NO), which influence endothelial cells, compromising the angiogenesis needed to confront hypoxia^[14]. In hypoxic tissues, for example, adipose tissue, chronic low-grade inflammation enhances the expression of HIF-1 α , which stimulates inflammatory genes to amplify the “meta-inflammatory” reaction, leading to insulin resistance^[15,16].

Increased uncoupled respiration by saturated fatty acids binding to adenosine diphosphate/adenosine-triphosphate translocase 2 (ANT2) is the original event in hypoxic adipocytes. A mass of free fatty acids provokes an ANT2-dependent increase in uncoupled mitochondrial respiration and oxygen consumption in obese/high-fat diet (HFD) mice, which stimulates the production of HIF-1 α and relative hypoxia in adipocytes. Increased HIF-1 α stimulates NO production by inducing iNOS expression. Then, insulin resistance could emerge by NO nitrosylation of the insulin-signaling molecule protein kinase B (Akt/PKB), which suppresses Akt-phosphorylated activation^[17]. In addition, abundant HIF-1 α increases lactate production in hypoxic adipocytes, giving rise to higher fasting blood glucose levels and accumulation of basal hepatic glucose^[18]. Simultaneously, the activation of ANT2 plays a crucial role in furthering adipose inflammation and fibrosis and metabolic dysfunction. Nevertheless, adipocyte-specific ANT2 knockout seems to be effective in preventing inflammation and fibrosis in adipose tissue and improving glucose tolerance and insulin sensitivity in mice^[19,20].

At length, two pathways dominate the accumulation of HIF-1 α to generate insulin resistance in hypoxic adipocytes: The JAK-signal transducer and activator of transcription 3 (STAT3) signaling pathway and the phospholipase A2 group 16-lysophosphatidylcholine pathway. In the JAK-STAT3 signaling pathway, stabilization and accumulation of HIF-1 α enhance the expression of suppressor of cytokine signaling 3 in the nucleus. Suppressor of cytokine signaling 3 protein phosphorylates STAT3, which downregulates the expression of adiponectin (encoded by *ADIPOQ*)^[21,22]. In the phospholipase A2 group 16-lysophosphatidylcholine pathway, HIF-1 α mediates homocysteine-induced adipose *pla2g16* (a novel HIF-1 α target gene) gene expression to elevate lysophosphatidylcholine (lyso-PC), which acts as a second signal activator in homocysteine-induced activation of the NLRP3 inflammasome pathway. Lysophosphatidylcholine (lyso-PC) not only further activates NLRP3 inflammasomes in adipocytes but also stimulates adipose tissue macrophage NLRP3 inflammasomes in a paracrine manner to induce insulin resistance^[23] (Figure 1).

CERAMIDE AND INSULIN RESISTANCE

Ceramides, a family of waxy lipid molecules that are composed of sphingosine and fatty acid, are important pathogenic lipids in obesity-related disorders. Starting with saturated fatty acids and palmitate intake, *de novo* synthesis of ceramide undergoes four major steps. This begins with the condensation of palmitate and serine to form 3-keto-dihydrosphingosine. This reaction is catalyzed by the enzyme serine palmitoyl transferase and is the rate-limiting step of the pathway. In turn, 3-keto-dihydrosphingosine is reduced to dihydrosphingosine, followed by acylation through (dihydro) ceramide synthase to produce dihydroceramide. Then, ceramide synthesis is catalyzed by dihydroceramide desaturase^[24]. Ceramide is also produced through the sphingomyelinase and salvage pathways. *Via* hydrolysis of sphingomyelin, which is catalyzed by the enzyme sphingomyelinase, ceramide can be generated. In addition, the salvage pathway reutilizes long-chain sphingoid bases to form ceramide through the action of ceramide synthase^[25] (Figure 2).

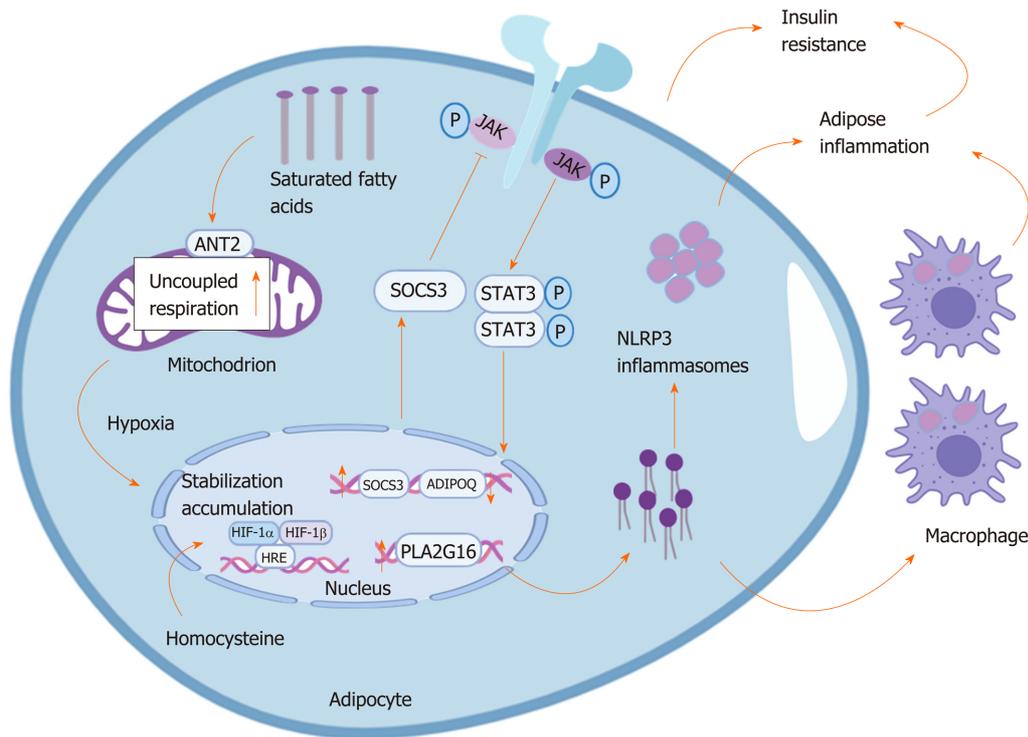


Figure 1 Accumulation of hypoxia-inducible factor 1α induces insulin resistance in hypoxic adipocytes. Owing to excess saturated fatty acids binding to adenosine diphosphate/adenosine-triphosphate translocase 2 in mitochondria, which increases uncoupled respiration leading to hypoxia in adipose tissue, the stabilized and accumulated hypoxia-inducible factor 1α (HIF-1α) further regulates relative target genes. On the one hand, HIF-1α induces expression of suppressor of cytokine signalling 3, which in turn activates signal transducer and activator of transcription 3, and dimerized signal transducer and activator of transcription 3 enters the cell nucleus and inhibits the transcription of ADIPOQ, resulting in insulin resistance. On the other hand, HIF-1α up-regulates the expression of pla2g16 to increase the level of lyso-PC, which in turn activates NLRP3 inflammasomes and stimulates NLRP3 inflammasomes in macrophages of adipose tissue, promoting insulin resistance. ANT2: Adenosine diphosphate/adenosine-triphosphate translocase 2; HIF-1α: Hypoxia-inducible factor 1α; SOCS3: Suppressor of cytokine signalling 3; JAK: Janus kinase; STAT3: Signal transducer and activator of transcription 3; HRE: Hypoxia-inducible factor regulating element; P: Phosphate; lyso-PC: Lyso-phosphatidylcholine.

In obese rodents, the production of ceramide increased compared with that in lean controls^[26], especially glucosylceramide^[27]. Similarly, studies performed in insulin-resistant human subjects demonstrate aberrant ceramide accumulation^[28]. In subjects with T2DM, investigators observed elevations in serum ceramide compared with healthy control subjects^[29]. It was reported that exercising training improved insulin sensitivity in obese and T2DM patients, and decreased the level of plasma ceramide especially C16:0 and C14:0. C16:0 was reduced from 2.5 nmol/mL to 1.75 nmol/mL and C14:0 reduced from 0.213 nmol/mL to 0.185 nmol/mL^[30]. In another clinical trial, it was found that in the group treated with berberine, the weight, body mass index, and ceramide of patients with T2DM significantly decreased compared with the lifestyle intervention group^[31]. However, due to the small sample size and limitations of ceramide detection methods, there is no consistent clinical data on the specific ceramide concentration in obese or diabetic patients.

Risk factors that associate with obesity, such as saturated fatty acids and inflammatory cytokines, selectively promote sphingolipid synthesis enzymes. Moreover, lipidomic profiling reveals the relationship between sphingolipid levels and metabolic diseases, and sphingolipid is shown to be involved in insulin resistance, pancreatic beta cell failure, cardiomyopathy, and vascular dysfunction in *in vivo* and *in vitro* studies^[32,33]. Adiponectin modulates ceramide by controlling its rate of degradation^[34].

Mechanism of ceramide synthesis affecting insulin resistance

Ceramide is produced in response to almost all stress stimuli, including those associated with obesity (*e.g.*, chemotherapy, inflammatory agonists, and saturated fatty acids). Aberrant accumulation of ceramide may lead to the activation of several signals, which may impair normal cellular function, especially insulin^[34,35]. How does ceramide synthesis affect insulin resistance in metabolic disease? By blocking translocation of the glucose transporter 4 through the inhibition of Akt/PKB activation, ceramides inhibit insulin-stimulated glucose uptake and glycogen

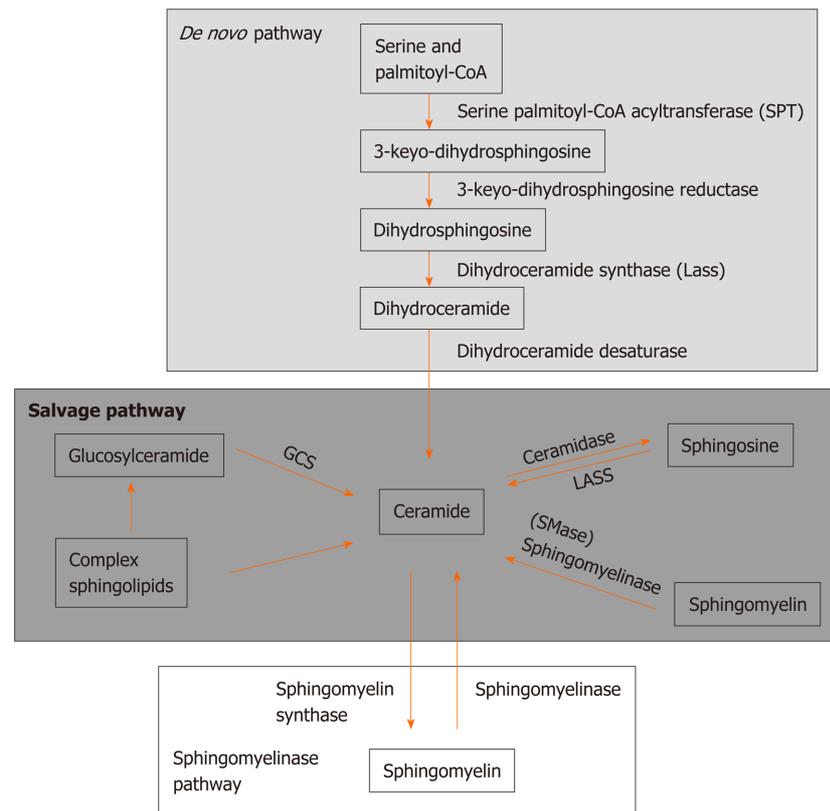


Figure 2 Ceramide synthesis pathways. Ceramides are synthesized through three ways, namely, *de novo* pathway, salvage pathway, and sphingomyelinase pathway. The *de novo* synthesis of ceramide commences with the condensation of serine and palmitate via action of serine palmitoyl-coenzyme A acyltransferase, followed by the continuous action of 3-keto-dihydrosphingosine reductase, dihydroceramide synthases, and dihydroceramide desaturase. In the sphingomyelinase pathway, ceramide can be produced from hydrolysis of sphingomyelin through the action of either acid or neutral sphingomyelinase, and ceramide also can synthesize sphingomyelin through the action of sphingomyelin synthase. The salvage pathway is more complex than the other two pathways. Glucosylceramide, complex sphingolipids, sphingosine, and sphingomyelin can generate ceramide from the action of diverse enzymes such as glucosylceramide synthase, LASS, and sphingomyelinase. SPT: Serine palmitoyl-coenzyme A acyltransferase; SMase: Sphingomyelinase; GCS: Glucosylceramide synthase.

synthesis in adipocytes and isolated skeletal muscle^[36,37]. Ceramides block the activation of Akt/PKB through two key regulatory mechanisms. First, ceramide activates the atypical protein kinase C isoform protein kinase C ζ and stabilizes interactions between Akt/PKB and protein kinase C ζ by recruiting the enzymes to detergent-resistant membrane fractions^[38]. The enzyme's PH domain of Akt/PKB reduces its affinity for phosphoinositides, resulting in inactivation of Akt/PKB and preventing the translocation of Akt/PKB to the plasma membrane^[39,40]. The second mechanism is that activation of protein phosphatase 2A (PPA2, the primary phosphatase responsible for dephosphorylating Akt/PKB) dephosphorylates Akt/PKB. The effects of ceramide on Akt/PKB can be prevented by adding okadaic acid or overexpressing the SV40 small T antigen to inhibit PPA2^[41] (Figure 3).

HYPOXIA AND CERAMIDE

HIF α is stabilized and activated under hypoxic conditions^[42]. It seems that hypoxia may enhance the level of ceramide in the majority of tissues. Hypoxia leads to ceramide upregulation in NT-2 neuronal precursor cells due to the actions of acid sphingomyelinase and ceramide synthase (LASS-5) to a large extent^[43]. In addition, in resistant pulmonary arteries, hypoxia induces increased ceramide and reactive oxygen species^[44]. Hypoxia activates neutral sphingomyelinases (nSMases), which are key enzymes in ceramide synthesis, enhancing the production of ceramide and the subsequent ceramide-triggered activation of protein kinase C ζ , which is an early and essential event in the signaling cascade of acute hypoxic pulmonary arteries. Inhibition of nSMase (GW4869) can prevent p47^{phox} phosphorylation induced by

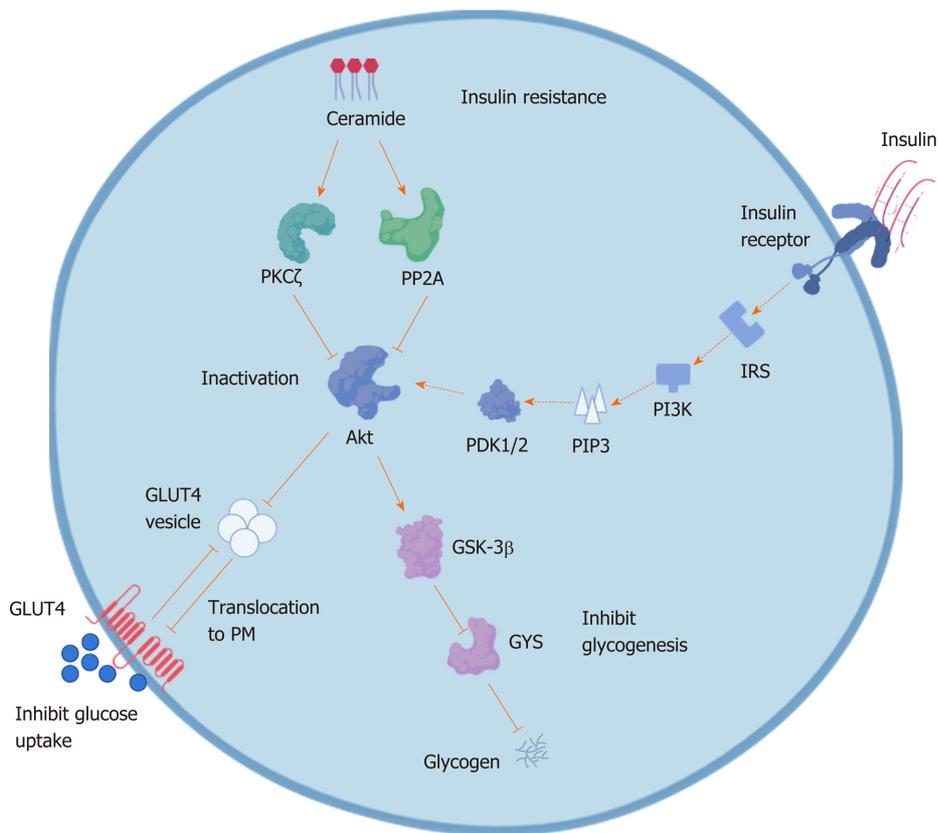


Figure 3 The mechanism of ceramide inducing insulin resistance. Ceramide inactivates protein kinase B (Akt) through stimulating the activity of protein kinase C ζ isoform and protein phosphatase 2A which phosphorylates and inhibits the translocation of Akt. The inactivation of Akt prevents from translocation of glucosetransporter4 vesicle to plasma membrane, resulting in inhibiting glucose uptake. Simultaneously, inactivated Akt in turn activates glycogen synthase 3, leading to inactivation of glycogen synthase and thus inhibition of glycogen synthesis and resulting in insulin resistance. PKB: Protein kinase B; PKC ζ : Protein kinase C ζ ; PP2A: Protein phosphatase 2A; GLUT4: Glucosetransporter4; PM: Plasma membrane; GSK-3: Glycogen synthase 3; GYS: Glycogen synthase; IRS: Insulin receptor substrate; PI-3K: Phosphoinositide 3-kinase; PIP3: Phosphatidylinositol-3,4,5-trisphosphate; PDK1/2: 3-phosphoinositide-dependent protein kinase 1/2.

hypoxia^[45]. Likewise, palmitoyltransferase (SPT) and glucosylceramide synthase (GCS) are the pivotal enzymes of ceramide synthesis, which may regulate the cellular level of ceramide, deciding the fate of the cell exposed to hypoxia. The hypoxia-induced increase in ceramide is partially attributed to the transcriptional upregulation of SPT2. Specific siRNA of SPT2 or GCS can reduce ceramide^[46]. Therefore, ceramide synthase inhibitors may be an efficient way to restrain ceramide synthase against hypoxic injury^[47].

RELATIONSHIP BETWEEN INTESTINAL HYPOXIA AND CERAMIDE INCREASES WITH INSULIN RESISTANCE

Intestinal mucosal barrier and hypoxia

The intestine is one passageway that communicates between the environment and the external environment of the human body and plays an essential role in the absorption of nutrients and protection from chemical and physical injury. The function of absorption and protection is benefited by the intestinal mucosal barrier, which involves the external physical barrier and internal functional immune barrier^[48]. The physical barrier mainly consists of cells and extracellular components, and the cellular components comprise intestinal epithelial cells and the inherent layer. Intestinal epithelial cells consist of absorption (absorbent intestinal cells and M cells) and secretion lines (Pan cells, cup cells, tuft cells, and intestinal endocrine cells)^[49], and the inherent layer includes dendritic cells, macrophages, epithelial lymphocytes, regulatory T cells, and B lymphocytes^[50]. The functional immune barrier consists of the chemical barrier (antimicrobial peptides, digestive secretions, cytokines, inflammatory mediators, *etc.*), intestinal microbiota barrier, and immune function barrier^[51]. The barrier functions of the intestinal mucous membrane are regulated by the availability of oxygen^[52].

Intestinal tissue oxygen has several characteristics. First, the intestinal epithelium is located between the underlying mucosa with high oxygen content and the anaerobic lumen of the intestine, forming a cliffy oxygen gradient under physiological conditions^[53]. Second, slight changes in blood flow can cause a significant variation in intestinal oxygen, such as an increase in blood flow volume after feeding (5% of total blood flow increased to 30%), which leads to a change in blood flow of the intestinal mucosa being the reason for the distinct change in local oxygen levels^[54]. Intestinal epithelial cells have better adaptability and regulation of hypoxia than other tissues, and physiologic hypoxia might be an adaptive regulation mechanism for the steep oxygen gradient^[55].

Intestinal hypoxia is divided into physiological hypoxia and pathological hypoxia. Physiological hypoxia refers to a relatively low PO₂ state present in mucosal epithelial cells even at baseline levels because the intestinal mucosa has a wealth of blood vessels, even if a slight reduction in blood flow can lead to a greater reduction in oxygen transport to the intestinal epithelial cells^[52,56]. Pathological intestinal hypoxia widely exists in cancer, acute lung injury, inflammatory bowel disease and metabolic diseases^[5,54,57]. Intestinal hypoxia is usually associated with the destruction of the intestinal mucosal barrier, such as that occurs in inflammatory intestinal diseases from the reduction in blood supply due to inflammatory immersion, edema, and vasoconstriction, leading to limited oxygen transport of the intestinal epithelium and aggregation of polymorphonuclear cells. At the same time, a large number of neutrophils rapidly deplete local oxygen through respiratory action, leading to hypoxia in the intestines^[54]. The HIF-2 α –NEU3–ceramide pathway may explain the relationship between intestinal hypoxia and insulin resistance.

The HIF-2 α –NEU3–ceramide pathway

In intestinal epithelial cells, HIF-2 α is activated and accumulates by hypoxia in HFD mice, which upregulates the target gene *Neu3* encoding sialidase 3. Sialidase 3 hydrolyses gangliosides to form ceramides in the salvage pathway^[25]. The HIF-2 α –NEU3–ceramide pathway can promote the development of metabolic diseases, such as NAFLD, obesity, and insulin resistance. Ceramides are synthesized through three different pathways: *De novo* pathway, sphingomyelinase (SMase) pathway, and salvage pathway^[58]. Increased levels of ceramide cause obesity, insulin resistance, and hepatic steatosis owing to upregulation of fatty acid synthesis. Nevertheless, the target genes of the three ceramide synthesis pathways, including *Degs2*, *Smpd1*, *Smpd3*, *Smpd4*, *Enpp7*, *Neu3*, *Glb1*, and *Gba2*, were substantially downregulated, further resulting in the reduction in ceramide in intestine-specific HIF-2 α ablation mice, which significantly ameliorates HFD-induced obesity and hepatic steatosis and improves insulin sensitivity in mice. In addition, treatment with a pharmacological specific inhibitor of HIF-2 α (PT2385) or inhibitor of NEU3 (N-acetyl-2,3-didehydro-N-acetyl-neuraminic acid, DANA, or naringin) lessens serum levels of ceramides, reduces obesity and fatty liver, and enhances insulin sensitivity^[33,59].

The components of the intestinal barrier are abundant. HIF-1 α derived from intestinal epithelial cells is important for intestinal intraepithelial lymphocytes and intestinal flora homeostasis. Whether other mechanisms are involved in insulin resistance under hypoxia requires more research to confirm.

CONCLUSION

The global incidence of T2DM has obviously increased in recent decades with economic development and lifestyle changes, especially in developed countries. Chronic inflammation, hypoxia, and the metabolism of ceramide are closely related to insulin resistance. Many studies have shown that HIF α regulates insulin resistance, for example, in adipocyte-specific *Hif1a* knockout mice, homocysteine-induced insulin resistance is ameliorated, the NLRP3 inflammasome is inhibited, and the production of ceramide is decreased^[23]. Meanwhile, intestine-specific *Hif2a* ablation mice show improved HFD-induced insulin resistance^[33].

Ceramide is a significant metabolic product of sphingolipids and contributes to insulin resistance and hepatic steatosis^[60]. Under hypoxia, HIF-2 α can induce ceramide in adipocytes and intestines, resulting in insulin resistance in HFD-induced obesity mice. As a result of a cliffy oxygen gradient in intestinal tissue and inflammatory changes in the intestinal mucosal barrier, hypoxia occurs in the intestine. Intestinal hypoxia may lead to HFD-induced insulin resistance. A study on the HIF-2 α –NEU3–ceramide pathway revealed the role of ceramide in hypoxia and insulin resistance in obese mice.

In summary, hypoxia is a key feature of the progression of metabolic disease and

HIF signaling, which can strongly influence metabolic disease by both genetic and pathophysiological inhibition. Recent discoveries have identified exciting effects of pharmacologic inhibitors of HIF-2 α or inhibitors of key enzymes (sialidase 3, NEU3) in ceramide synthesis. This may become a promising approach to the treatment of metabolic diseases, including insulin resistance and NAFLD.

REFERENCES

- GBD 2013 Mortality and Causes of Death Collaborators.** Global, regional, and national age-sex specific all-cause and cause-specific mortality for 240 causes of death, 1990-2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet* 2015; **385**: 117-171 [PMID: 25530442 DOI: 10.1016/S0140-6736(14)61682-2]
- Federation ID.** IDF Diabetes Atlas, 9th edition 2019. Brussels, Belgium. Available from: <https://www.diabetesatlas.org/en/sections/proven-and-effective-actions.html>
- Shoelson SE, Herrero L, Naaz A.** Obesity, inflammation, and insulin resistance. *Gastroenterology* 2007; **132**: 2169-2180 [PMID: 17498510 DOI: 10.1053/j.gastro.2007.03.059]
- Dali-Youcef N, Mecili M, Ricci R, Andr s E.** Metabolic inflammation: connecting obesity and insulin resistance. *Ann Med* 2013; **45**: 242-253 [PMID: 22834949 DOI: 10.3109/07853890.2012.705015]
- Gonzalez FJ, Xie C, Jiang C.** The role of hypoxia-inducible factors in metabolic diseases. *Nat Rev Endocrinol* 2018; **15**: 21-32 [PMID: 30275460 DOI: 10.1038/s41574-018-0096-z]
- Fagundes RR, Taylor CT.** Determinants of hypoxia-inducible factor activity in the intestinal mucosa. *J Appl Physiol* (1985) 2017; **123**: 1328-1334 [PMID: 28408694 DOI: 10.1152/jappphysiol.00203.2017]
- Semenza GL.** Life with oxygen. *Science* 2007; **318**: 62-64 [PMID: 17916722 DOI: 10.1126/science.1147949]
- Wang GL, Jiang BH, Rue EA, Semenza GL.** Hypoxia-inducible factor 1 is a basic-helix-loop-helix-PAS heterodimer regulated by cellular O₂ tension. *Proc Natl Acad Sci USA* 1995; **92**: 5510-5514 [PMID: 7539918 DOI: 10.1073/pnas.92.12.5510]
- Yang J, Zhang L, Erbel PJ, Gardner KH, Ding K, Garcia JA, Bruick RK.** Functions of the Per/ARNT/Sim domains of the hypoxia-inducible factor. *J Biol Chem* 2005; **280**: 36047-36054 [PMID: 16129688 DOI: 10.1074/jbc.M501755200]
- Mastrogiannaki M, Matak P, Keith B, Simon MC, Vaulont S, Peyssonnaud C.** HIF-2 α , but not HIF-1 α , promotes iron absorption in mice. *J Clin Invest* 2009; **119**: 1159-1166 [PMID: 19352007 DOI: 10.1172/JCI38499]
- Jaakkola P, Mole DR, Tian YM, Wilson MI, Gielbert J, Gaskell SJ, von Kriegsheim A, Hebestreit HF, Mukherji M, Schofield CJ, Maxwell PH, Pugh CW, Ratcliffe PJ.** Targeting of HIF- α to the von Hippel-Lindau ubiquitylation complex by O₂-regulated prolyl hydroxylation. *Science* 2001; **292**: 468-472 [PMID: 11292861 DOI: 10.1126/science.1059796]
- Ye J, McGuinness OP.** Inflammation during obesity is not all bad: evidence from animal and human studies. *Am J Physiol Endocrinol Metab* 2013; **304**: E466-E477 [PMID: 23269411 DOI: 10.1152/ajpendo.00266.2012]
- van der Pouw Kraan TC, Chen WJ, Bunck MC, van Raalte DH, van der Zijl NJ, van Genugten RE, van Bloemendaal L, Baggen JM, Sern  EH, Diamant M, Horrevoets AJ.** Metabolic changes in type 2 diabetes are reflected in peripheral blood cells, revealing aberrant cytotoxicity, a viral signature, and hypoxia inducible factor activity. *BMC Med Genomics* 2015; **8**: 20 [PMID: 25956355 DOI: 10.1186/s12920-015-0096-y]
- Rutkowski JM, Davis KE, Scherer PE.** Mechanisms of obesity and related pathologies: the macro- and microcirculation of adipose tissue. *FEBS J* 2009; **276**: 5738-5746 [PMID: 19754873 DOI: 10.1111/j.1742-4658.2009.07303.x]
- Schenk S, Saberi M, Olefsky JM.** Insulin sensitivity: modulation by nutrients and inflammation. *J Clin Invest* 2008; **118**: 2992-3002 [PMID: 18769626 DOI: 10.1172/JCI34260]
- Halberg N, Khan T, Trujillo ME, Wernstedt-Asterholm I, Attie AD, Sherwani S, Wang ZV, Landskroner-Eiger S, Dineen S, Magalang UJ, Brekken RA, Scherer PE.** Hypoxia-inducible factor 1 α induces fibrosis and insulin resistance in white adipose tissue. *Mol Cell Biol* 2009; **29**: 4467-4483 [PMID: 19546236 DOI: 10.1128/MCB.00192-09]
- Yasukawa T, Tokunaga E, Ota H, Sugita H, Martyn JA, Kaneki M.** S-nitrosylation-dependent inactivation of Akt/protein kinase B in insulin resistance. *J Biol Chem* 2005; **280**: 7511-7518 [PMID: 15632167 DOI: 10.1074/jbc.M411871200]
- Lee YS, Kim JW, Osborne O, Oh DY, Sasik R, Schenk S, Chen A, Chung H, Murphy A, Watkins SM, Quehenberger O, Johnson RS, Olefsky JM.** Increased adipocyte O₂ consumption triggers HIF-1 α , causing inflammation and insulin resistance in obesity. *Cell* 2014; **157**: 1339-1352 [PMID: 24906151 DOI: 10.1016/j.cell.2014.05.012]
- Leake I.** ANT2 mediates hypoxia and inflammation in obesity. *Nat Rev Endocrinol* 2019; **15**: 64 [PMID: 30531919 DOI: 10.1038/s41574-018-0140-z]
- Seo JB, Riopel M, Cabrales P, Huh JY, Bandyopadhyay GK, Andreyev AY, Murphy AN, Beeman SC, Smith GI, Klein S, Lee YS, Olefsky JM.** Knockdown of *Ant2* Reduces Adipocyte Hypoxia and Improves Insulin Resistance in Obesity. *Nat Metab* 2019; **1**: 86-97 [PMID: 31528845 DOI: 10.1038/s42255-018-0003-x]
- Jiang C, Kim JH, Li F, Qu A, Gavrilova O, Shah YM, Gonzalez FJ.** Hypoxia-inducible factor 1 α regulates a SOCS3-STAT3-adiponectin signal transduction pathway in adipocytes. *J Biol Chem* 2013; **288**: 3844-3857 [PMID: 23255598 DOI: 10.1074/jbc.M112.426338]
- Kanatani Y, Usui I, Ishizuka K, Bukhari A, Fujisaka S, Urakaze M, Haruta T, Kishimoto T, Naka T, Kobayashi M.** Effects of pioglitazone on suppressor of cytokine signaling 3 expression: potential mechanisms for its effects on insulin sensitivity and adiponectin expression. *Diabetes* 2007; **56**: 795-803 [PMID: 17327450 DOI: 10.2337/db06-1039]
- Zhang SY, Dong YQ, Wang P, Zhang X, Yan Y, Sun L, Liu B, Zhang D, Zhang H, Liu H, Kong W, Hu G, Shah YM, Gonzalez FJ, Wang X, Jiang C.** Adipocyte-derived Lysophosphatidylcholine Activates Adipocyte and Adipose Tissue Macrophage Nod-Like Receptor Protein 3 Inflammasomes Mediating Homocysteine-Induced Insulin Resistance. *EBioMedicine* 2018; **31**: 202-216 [PMID: 29735414 DOI: 10.1016/j.ebiom.2018.05.012]

- 10.1016/j.ebiom.2018.04.022]
- 24 **Hannun YA**, Obeid LM. Principles of bioactive lipid signalling: lessons from sphingolipids. *Nat Rev Mol Cell Biol* 2008; **9**: 139-150 [PMID: 18216770 DOI: 10.1038/nrm2329]
- 25 **Kitatani K**, Idkowiak-Baldys J, Hannun YA. The sphingolipid salvage pathway in ceramide metabolism and signaling. *Cell Signal* 2008; **20**: 1010-1018 [PMID: 18191382 DOI: 10.1016/j.cellsig.2007.12.006]
- 26 **Yetukuri L**, Katajamaa M, Medina-Gomez G, Seppänen-Laakso T, Vidal-Puig A, Oresic M. Bioinformatics strategies for lipidomics analysis: characterization of obesity related hepatic steatosis. *BMC Syst Biol* 2007; **1**: 12 [PMID: 17408502 DOI: 10.1186/1752-0509-1-12]
- 27 **Bijl N**, Scheij S, Houten S, Boot RG, Groen AK, Aerts JM. The glucosylceramide synthase inhibitor N-(5-adamantane-1-yl-methoxy-pentyl)-deoxynojirimycin induces sterol regulatory element-binding protein-regulated gene expression and cholesterol synthesis in HepG2 cells. *J Pharmacol Exp Ther* 2008; **326**: 849-855 [PMID: 18550691 DOI: 10.1124/jpet.108.139394]
- 28 **Adams JM 2nd**, Pratipanawat T, Berria R, Wang E, DeFronzo RA, Sullards MC, Mandarino LJ. Ceramide content is increased in skeletal muscle from obese insulin-resistant humans. *Diabetes* 2004; **53**: 25-31 [PMID: 14693694 DOI: 10.2337/diabetes.53.1.25]
- 29 **Górska M**, Dobrzyń A, Baranowski M. Concentrations of sphingosine and sphinganine in plasma of patients with type 2 diabetes. *Med Sci Monit* 2005; **11**: CR35-CR38 [PMID: 15614193]
- 30 **Kasumov T**, Solomon TP, Hwang C, Huang H, Haus JM, Zhang R, Kirwan JP. Improved insulin sensitivity after exercise training is linked to reduced plasma C14:0 ceramide in obesity and type 2 diabetes. *Obesity (Silver Spring)* 2015; **23**: 1414-1421 [PMID: 25966363 DOI: 10.1002/oby.21117]
- 31 **Chang X**, Wang Z, Zhang J, Yan H, Bian H, Xia M, Lin H, Jiang J, Gao X. Lipid profiling of the therapeutic effects of berberine in patients with nonalcoholic fatty liver disease. *J Transl Med* 2016; **14**: 266 [PMID: 27629750 DOI: 10.1186/s12967-016-0982-x]
- 32 **Holland WL**, Summers SA. Sphingolipids, insulin resistance, and metabolic disease: new insights from in vivo manipulation of sphingolipid metabolism. *Endocr Rev* 2008; **29**: 381-402 [PMID: 18451260 DOI: 10.1210/er.2007-0025]
- 33 **Xie C**, Yagai T, Luo Y, Liang X, Chen T, Wang Q, Sun D, Zhao J, Ramakrishnan SK, Sun L, Jiang C, Xue X, Tian Y, Krausz KW, Patterson AD, Shah YM, Wu Y, Jiang C, Gonzalez FJ. Activation of intestinal hypoxia-inducible factor 2 α during obesity contributes to hepatic steatosis. *Nat Med* 2017; **23**: 1298-1308 [PMID: 29035368 DOI: 10.1038/nm.4412]
- 34 **Summers SA**, Goodpaster BH. CrossTalk proposal: Intramyocellular ceramide accumulation does modulate insulin resistance. *J Physiol* 2016; **594**: 3167-3170 [PMID: 26996141 DOI: 10.1113/JP271676]
- 35 **Chavez JA**, Summers SA. A ceramide-centric view of insulin resistance. *Cell Metab* 2012; **15**: 585-594 [PMID: 22560211 DOI: 10.1016/j.cmet.2012.04.002]
- 36 **Hofmann K**, Dixit VM. Ceramide in apoptosis--does it really matter? *Trends Biochem Sci* 1998; **23**: 374-377 [PMID: 9810222 DOI: 10.1016/s0968-0004(98)01289-4]
- 37 **Summers SA**, Garza LA, Zhou H, Birnbaum MJ. Regulation of insulin-stimulated glucose transporter GLUT4 translocation and Akt kinase activity by ceramide. *Mol Cell Biol* 1998; **18**: 5457-5464 [PMID: 9710629 DOI: 10.1128/mcb.18.9.5457]
- 38 **Fox TE**, Houck KL, O'Neill SM, Nagarajan M, Stover TC, Pomianowski PT, Unal O, Yun JK, Naides SJ, Kester M. Ceramide recruits and activates protein kinase C zeta (PKC zeta) within structured membrane microdomains. *J Biol Chem* 2007; **282**: 12450-12457 [PMID: 17308302 DOI: 10.1074/jbc.M700082200]
- 39 **Stratford S**, DeWald DB, Summers SA. Ceramide dissociates 3'-phosphoinositide production from pleckstrin homology domain translocation. *Biochem J* 2001; **354**: 359-368 [PMID: 11171115 DOI: 10.1042/0264-6021:3540359]
- 40 **Powell DJ**, Hajdud E, Kular G, Hundal HS. Ceramide disables 3-phosphoinositide binding to the pleckstrin homology domain of protein kinase B (PKB)/Akt by a PKCzeta-dependent mechanism. *Mol Cell Biol* 2003; **23**: 7794-7808 [PMID: 14560023 DOI: 10.1128/mcb.23.21.7794-7808.2003]
- 41 **Chavez JA**, Knotts TA, Wang LP, Li G, Dobrowsky RT, Florant GL, Summers SA. A role for ceramide, but not diacylglycerol, in the antagonism of insulin signal transduction by saturated fatty acids. *J Biol Chem* 2003; **278**: 10297-10303 [PMID: 12525490 DOI: 10.1074/jbc.M212307200]
- 42 **Ivan M**, Kondo K, Yang H, Kim W, Valiando J, Ohh M, Salic A, Asara JM, Lane WS, Kaelin WG. HIF α targeted for VHL-mediated destruction by proline hydroxylation: implications for O $_2$ sensing. *Science* 2001; **292**: 464-468 [PMID: 11292862 DOI: 10.1126/science.1059817]
- 43 **Jin J**, Hou Q, Mullen TD, Zeidan YH, Bielawski J, Kravka JM, Bielawska A, Obeid LM, Hannun YA, Hsu YT. Ceramide generated by sphingomyelin hydrolysis and the salvage pathway is involved in hypoxia/reoxygenation-induced Bax redistribution to mitochondria in NT-2 cells. *J Biol Chem* 2008; **283**: 26509-26517 [PMID: 18676372 DOI: 10.1074/jbc.M801597200]
- 44 **Moreno L**, Moral-Sanz J, Morales-Cano D, Barreira B, Moreno E, Ferrarini A, Pandolfi R, Ruperez FJ, Cortijo J, Sanchez-Luna M, Villamor E, Perez-Vizcaino F, Cogolludo A. Ceramide mediates acute oxygen sensing in vascular tissues. *Antioxid Redox Signal* 2014; **20**: 1-14 [PMID: 23725018 DOI: 10.1089/ars.2012.4752]
- 45 **Frazziano G**, Moreno L, Moral-Sanz J, Menendez C, Escolano L, Gonzalez C, Villamor E, Alvarez-Sala JL, Cogolludo AL, Perez-Vizcaino F. Neutral sphingomyelinase, NADPH oxidase and reactive oxygen species. Role in acute hypoxic pulmonary vasoconstriction. *J Cell Physiol* 2011; **226**: 2633-2640 [PMID: 21792922 DOI: 10.1002/jcp.22611]
- 46 **Kang MS**, Ahn KH, Kim SK, Jeon HJ, Ji JE, Choi JM, Jung KM, Jung SY, Kim DK. Hypoxia-induced neuronal apoptosis is mediated by de novo synthesis of ceramide through activation of serine palmitoyltransferase. *Cell Signal* 2010; **22**: 610-618 [PMID: 19932170 DOI: 10.1016/j.cellsig.2009.11.015]
- 47 **Crowder CM**. Cell biology. Ceramides--friend or foe in hypoxia? *Science* 2009; **324**: 343-344 [PMID: 19372418 DOI: 10.1126/science.1173278]
- 48 **Bischoff SC**, Barbara G, Buurman W, Ockhuizen T, Schulzke JD, Serino M, Tilg H, Watson A, Wells JM. Intestinal permeability--a new target for disease prevention and therapy. *BMC Gastroenterol* 2014; **14**: 189 [PMID: 25407511 DOI: 10.1186/s12876-014-0189-7]
- 49 **Gehart H**, Clevers H. Tales from the crypt: new insights into intestinal stem cells. *Nat Rev Gastroenterol Hepatol* 2019; **16**: 19-34 [PMID: 30429586 DOI: 10.1038/s41575-018-0081-y]
- 50 **Rescigno M**. The intestinal epithelial barrier in the control of homeostasis and immunity. *Trends Immunol* 2011; **32**: 256-264 [PMID: 21565554 DOI: 10.1016/j.it.2011.04.003]
- 51 **Cummings JH**, Antoine JM, Azpiroz F, Bourdet-Sicard R, Brandtzaeg P, Calder PC, Gibson GR, Guarner F, Isolauri E, Pannemans D, Shortt C, Tuijelaars S, Watzl B. PASSCLAIM--gut health and immunity. *Eur*

- J Nutr* 2004; **43** Suppl 2: II118-II173 [PMID: 15221356 DOI: 10.1007/s00394-004-1205-4]
- 52 **Colgan SP**, Taylor CT. Hypoxia: an alarm signal during intestinal inflammation. *Nat Rev Gastroenterol Hepatol* 2010; **7**: 281-287 [PMID: 20368740 DOI: 10.1038/nrgastro.2010.39]
- 53 **Zeitouni NE**, Chotikatum S, von Kückritz-Blickwede M, Naim HY. The impact of hypoxia on intestinal epithelial cell functions: consequences for invasion by bacterial pathogens. *Mol Cell Pediatr* 2016; **3**: 14 [PMID: 27002817 DOI: 10.1186/s40348-016-0041-y]
- 54 **Shehade H**, Oldenhove G, Moser M. Hypoxia in the intestine or solid tumors: a beneficial or deleterious alarm signal? *Eur J Immunol* 2014; **44**: 2550-2557 [PMID: 25043839 DOI: 10.1002/eji.201444719]
- 55 **Furuta GT**, Turner JR, Taylor CT, Hershberg RM, Comerford K, Narravula S, Podolsky DK, Colgan SP. Hypoxia-inducible factor 1-dependent induction of intestinal trefoil factor protects barrier function during hypoxia. *J Exp Med* 2001; **193**: 1027-1034 [PMID: 11342587 DOI: 10.1084/jem.193.9.1027]
- 56 **Glover LE**, Lee JS, Colgan SP. Oxygen metabolism and barrier regulation in the intestinal mucosa. *J Clin Invest* 2016; **126**: 3680-3688 [PMID: 27500494 DOI: 10.1172/JCI84429]
- 57 **Wu G**, Xu G, Chen DW, Gao WX, Xiong JQ, Shen HY, Gao YQ. Hypoxia Exacerbates Inflammatory Acute Lung Injury via the Toll-Like Receptor 4 Signaling Pathway. *Front Immunol* 2018; **9**: 1667 [PMID: 30083155 DOI: 10.3389/fimmu.2018.01667]
- 58 **Pagadala M**, Kasumov T, McCullough AJ, Zein NN, Kirwan JP. Role of ceramides in nonalcoholic fatty liver disease. *Trends Endocrinol Metab* 2012; **23**: 365-371 [PMID: 22609053 DOI: 10.1016/j.tem.2012.04.005]
- 59 **Miyagi T**, Wada T, Yamaguchi K, Hata K, Shiozaki K. Plasma membrane-associated sialidase as a crucial regulator of transmembrane signalling. *J Biochem* 2008; **144**: 279-285 [PMID: 18632803 DOI: 10.1093/jb/mvn089]
- 60 **Holland WL**, Brozinick JT, Wang LP, Hawkins ED, Sargent KM, Liu Y, Narra K, Hoehn KL, Knotts TA, Siesky A, Nelson DH, Karathanasis SK, Fontenot GK, Birnbaum MJ, Summers SA. Inhibition of ceramide synthesis ameliorates glucocorticoid-, saturated-fat-, and obesity-induced insulin resistance. *Cell Metab* 2007; **5**: 167-179 [PMID: 17339025 DOI: 10.1016/j.cmet.2007.01.002]



Published By Baishideng Publishing Group Inc
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA
Telephone: +1-925-3991568
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>

