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**Predicting** **dyslipidemia after liver transplantation: a significant role of recipient metabolic inflammation profile**

Huang HT *et al.* Predicting dyslipidemia after LT

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**Abstract**

BACKGROUND

Post-transplant dyslipidemia (PTDL) is a common complication in liver recipients and can cause morbidity and threaten graft function. The crosstalk between metabolic inflammation and dyslipidemia has been recently revealed. However, the role of grafts’ and recipients’ metabolic status in the development of PTDL has not been evaluated.

AIM

To investigate the association of recipients’ metabolic inflammation status with PTDL and construct a predictive model.

METHODS

A total of 396 adult patients who received primary liver transplantation between 2015 and 2017 were enrolled. Metabolomics and cytokines were analyzed using recipients’ pre-transplant peripheral blood in a training set (*n* = 72). An integrated prediction model was established according to the clinical risk factors and metabolic inflammation compounds and further verified in a validation set (*n* = 144).

RESULTS

The serum lipid profile took 3 mo to reach homeostasis after liver transplantation. A total of 278 (70.2%) liver recipients developed PTDL during a follow-up period of 1.78 (1.00, 2.97) years. The PTDL group showed a significantly lower tumor-free survival and overall survival than the non-PTDL group in patients with hepatocellular carcinoma (*n* = 169). The metabolomic analysis showed that metabolic features discriminating between the PTDL and non-PTDL groups were associated with lipid and glucose metabolism-associated pathways. Among metabolites and cytokines differentially expressed between the two groups, interleukin-12 (p70) showed the best diagnostic accuracy and significantly increased the predictive value when it was incorporated into the clinical model in both training and validation sets.

CONCLUSION

Recipients’ pre-transplant serum interleukin-12 (p70) level is associated with the risk of PTDL and has potential clinical value for predicting PTDL.

**Key words:** Dyslipidemia; Liver transplantation; Metabolomics; Cytokines; Predictive model

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**Core tip:** Post-transplant dyslipidemia (PTDL) is a common complication in liver recipients and can cause morbidity and threaten graft function. The crosstalk between metabolic inflammation and dyslipidemia has been recently revealed, however, the role of recipients’ metabolic status in the development of PTDL has not been evaluated. Here, we conducted the first study to explore the association of recipients’ metabolic inflammation with PTDL and further evaluate the diagnostic efficacy of metabolic inflammation compounds. Interleukin-12 (p70) was found to be a valid predictor of PTDL and remarkably improve the predictive ability of the clinical model. **INTRODUCTION**

Dyslipidemia is a common complication after liver transplantation (LT)[1]. Due to the greatly prolonged patient survival after LT in recent years, the prevalence of post-transplant dyslipidemia (PTDL) has been reported to be up to 85%, higher than that in the general population[1,2]. PTDL was closely associated with organ allograft rejection, cardiovascular events, and graft dysfunction, which subsequently led to decreased patients’ survival[3-5]. Therefore, the early screening of patients at a high risk of developing PTDL will give the opportunity for prompt prevention and improving the long-term prognosis.

The underlying mechanism for developing PTDL is still unclear. Several clinical risk factors have been identified, including old age, high body mass index (BMI), renal dysfunction, and immunosuppressive agents[1,6]. Metabolic inflammation, which is initially mediated by innate immune signaling, is increasingly recognized as a driving force in the development of lipid disorders[7]. Of note, cytokines produced during metabolic inflammation have been proved to further worsen lipid disorders[8], which suggested that cytokines could serve as biomarkers for predicting PTDL. Currently, experimental and clinical studies have identified a number of cytokines associated with lipid-induced metabolic diseases, including interleukin (IL)-1, IL-18, IL-6, IL-8, and tumor necrosis factor-α[9,10]. In addition, cytokines can activate a series of metabolic pathways[11] that regulate metabolism and produce metabolites to support the precise changes of immune cell functions[12,13]. These studies further suggested that specific cytokines and metabolites may play roles in the development of PTDL.

In this study, we aimed to investigate the association of pre-transplant metabolite and cytokine profiles with PTDL and establish a predictive model using clinical parameters and metabolic inflammation compounds.

**MATERIALs AND METHODS**

***Patients***

All adult (age ≥ 18 years) patients who received primary LT between January 2015 and December 2017 at the First Affiliated Hospital, Zhejiang University School of Medicine, China were enrolled in this study. The exclusion criteria were multi-organ transplantation, less than a 3-mo follow-up period, death within 3 mo post-LT, and incomplete data. Finally, a total of 396 patients were included. Patients’ characteristics are shown in Table 1. There was no living-donor LT. All patients received standard immunosuppressive protocol as described previously[14]. For hepatocellular carcinoma (HCC) patients, Hangzhou criteria were used as an indication of LT. Hangzhou criteria were defined as: (1) Total tumor diameter no more than 8 cm; and (2) total tumor diameter more than 8 cm, with pathological grade I or II and preoperative alpha fetoprotein level no more than 400 ng/mL[15]. The diagnosis of recurrence was based on imaging appearance. Patients with hepatitis B virus (HBV) infection received standard anti-virus protocol post-LT (low-dose immunoglobulin and nucleoside analogs) as we described before[16]. The follow-up ended on January 1, 2019. Peripheral blood samples from 216 recipients were collected before LT. They were randomly divided (in a 1:2 ratio) into training (*n* = 72) and validation sets (*n* = 144). This study was approved by the Ethics Committee of our hospital according to the Regulations on Human Organ Transplant and national legal requirements. This study conformed to the guidelines of China Ethical Committee and the Declaration of Helsinki. No grafts from prisoners were obtained or used. Written informed consent was obtained from all patients.

***Definition***

Dyslipidemia was defined as total cholesterol (TC) ≥ 240 mg/dL, or triglycerides (TG) ≥ 200 mg/dL, or high-density lipoprotein cholesterol (HDLC) < 40 mg/dL, or a need for using of medication for dyslipidemia[17].

***Metabolomics***

An ultra-performance liquid chromatography-mass spectrometry-based metabolomics analysis was performed as described previously[18]. The raw data were processed using the Compound Discoverer 3.0 (Thermo Fisher) to perform peak alignment, peak picking, and quantization for each metabolite. Partial least squares-discriminant analysis was performed with R pls package. Peaks were then matched with the mzCloud (https://www.mzcloud.org/) and ChemSpider (http://www.chemspider.com/) databases to obtain the accurate qualitative and relative quantitative results. Mummichog enrichment analysis for differential metabolic features was performed using metaboanalyst R package. Mantel tests based on Bray-Curtis distance and Pearson’s correlation analysis were applied to evaluate the correlations between cytokines and metabolic profiles with R ggcor package.

***Human inflammation assay***

Peripheral blood samples from 216 recipients were collected before LT. The cytokine immunoassays were carried out using blood samples collected under sterile conditions in BD vacutainer tubes containing EDTA-K2. Samples were examined for a panel of 37 serum cytokines using Bio-Plex Pro™ Human Inflammation Assays (No. 6625, Bio-Rad, Hercules, CA, United States) according to the standard protocol. The data were analyzed using Bio-Plex Data Pro™ software and Bio-Plex Manager software (Bio-Rad).

***Statistical analysis***

Data analyses were performed using SPSS for Windows version 11.0 (SPSS Inc., Chicago, IL, United States) and MedCalc for Windows version 11.4.2.0 (MedCalc Software, Mariakerke, Belgium). Quantitative variables were presented as the mean ± SD or median (interquartile range) and compared by Student’s *t* test or Mann-Whitney *U* test. Kaplan-Meier analysis was used for survival analysis. Categorical variables were presented as values (percentages) and compared by Pearson’s *χ*2 test or Fisher’s exact test. Risk factors for PTDL were evaluated by logistic regression analysis. Variables with statistical significance (*P* < 0.05) in univariate analysis were transferred to a subsequent stepwise multivariate regression analysis. The cut-off value was selected according to clinical value or median. The establishment of predictive models was described in our previous study[19]. The Hosmer and Lemeshow test was used to assess the models’ goodness-of-fit. The area under the receiver operating characteristic curve (AUC) was calculated to evaluate the predictive ability of PTDL models. The AUCs of different models were compared with MedCalc. A *P* value of < 0.05 was considered statistically significant.

**RESULTS**

***Serum lipid profiles after liver transplantation***

The dynamic changes of serum lipid profiles after LT are shown in Figure 1. TC, TG, LDLC, HDLC, and very low-density lipoprotein cholesterol sharply decreased in the first day following LT and gradually increased thereafter except HDLC (Figure 1A). At 3 mo post-LT, TC, TG, LDLC, and HDLC showed higher levels than preoperative levels, while very low-density lipoprotein cholesterol did not differ significantly compared to the preoperative levels. All lipids displayed relatively steady levels 3 mo post-LT (Figure 1B). The results suggested a 3-mo remodeling of lipid homeostasis in liver recipients.

***Incidence and prognosis of PTDL***

There were 278 (70.2%) patients who developed PTDL during a follow-up period of 1.78 (1.00, 2.97) years. The incidence of PTDL was 54.0% (214/396), 64.1% (238/369), and 69.9% (221/316) at 3 mo, 6 mo, and 1 year post-LT, respectively. The overall patient survival (Figure 2A) and post-transplant complications (Table 2) did not differ significantly between the two groups. However, among 169 HCC patients, the PTDL group showed a significantly lower overall patient survival (*P* = 0.038, Figure 2B) and tumor-free survival (*P* = 0.020, Figure 2C) than the non-PTDL group. There was no significant difference in HCC characteristics between the two groups (Supplementary Table 1). These results indicated that PTDL is associated with an adverse prognosis.

***Clinical risk factors of PTDL***

Donors’ and recipients’ clinical parameters were compared between the PTDL and non-PTDL groups (Table 1). Compared to the non-PTDL group, the PTDL group showed significantly higher recipients’ BMI, higher TG, more gastrointestinal bleeding and ascites cases, but lower HDLC and fewer hepatitis B and HCC cases. In contrast, donor parameters did not differ significantly between the two groups. In logistic analysis, recipient overweight [odds ratio (OR) = 2.705, *P* = 0.002] and hypo-HDLC (OR = 3.090, *P* < 0.001) were independent risk factors for PTDL (Table 3).

***Metabolomic inflammation profile of PTDL***

We performed an ultra-performance liquid chromatography-mass spectrometry based metabolomics approach to map the PTDL-associated metabolic feature. Partial least squares-discriminant analysis was used for quality control (Figure 3A). According to the selection criteria (Variable Importance in the Projection > 1 and *P* < 0.05), there were 30 significant differentially expressed metabolites in positive and negative ion modes between the PTDL (*n* = 45) and non-PTDL groups (*n* = 27) (Figure 3B). Mummichog analysis showed that the metabolic features discriminating between PTDL and non-PTDL group were associated with lipid metabolism-associated pathways (*e.g.*, steroid hormone biosynthesis, glycerophospholipid, ether lipid, and sphingolipid) and glucose metabolism-associated pathways (*e.g.*, glycolysis/gluconeogenesis, pentose phosphate, pentose and glucuronate, fructose and mannose, and galactose) (Figure 3C). In addition, 4/30 metabolites were significantly related to the recipients’ lipid profile (Figure 3D). However, in predicting PTDL, these metabolites presented AUCs of < 0.65.

We further performed a cytokine immunoassay in the same cohort. Compared with the non-PTDL group, the PTDL group showed a significantly lower IL-12 (p70) level and higher IFN-α2 and IFN-β levels (Figure 4A-C). Mantel test suggested a significant correlation between IL-12 (p70) and most identified metabolites (21/30) (Figure 4D), which are enriched in lipid metabolism-associated pathways (*e.g.*, diacylglycerol (20:5/20:5), 1-(11Z-docosenoyl)-glycero-3-phospho-(1'-sn-glycerol), 1-dodecanoyl-2-(5Z,8Z,11Z,14Z,17Z-eicosapentaenoyl)-sn-glycerol, and cholest-5,24-dien-3beta-ol 3-O-beta-D-glucopyranoside. In contrast, these cytokines have no direct correlation with recipients’ pre-transplant metabolic parameters (Figure 4E). In predicting PTDL, IL-12 (p70) presented the best predictive ability among all metabolomic inflammation compounds in the training set (AUC = 0.706) and validation set (AUC = 0.762).

***Predictive models of PTDL***

We constructed predictive models according to the logistic analysis (Table 4). Model 1 included only clinical parameters, whereas model 2 integrated clinical parameters with IL-12 (p70). Both models displayed good fits (*P* > 0.5 to reject model fit). Model 2 presented a significantly larger AUC than model 1 in both training (0.801 *vs* 0.677, *P* = 0.009) and validation sets (0.786 *vs* 0.691, *P* = 0.038) (Figure 5A and B).

**DISCUSSION**

In this study, we demonstrated a dynamic remodeling of lipid homeostasis in liver recipients. We found a longer remodeling time as compared with a previous study in living donor liver transplantation (3 mo *vs* 1 mo)[20], which indicates a longer period for close monitoring and prompt intervention. Furthermore, there was a very high incidence of PTDL in this study (> 70%) and previous ones[6]. We believed that it was due to not only the popularity of the disease but also a lack of proper screening and management of the disease. As a consequence, adverse outcomes such as HCC recurrence would occur. It should be an alarm bell for the transplant society, particularly in China, where there is the biggest HCC burden worldwide and HCC accounts for ~40% of LT indications. In addition, no formal guidelines have been developed to facilitate PTDL management up to now. Statins may be effective with a careful consideration of drug interactions and close patient monitoring[21]. Non-statin pharmacotherapy has also been evaluated in PTDL but only in a limited capacity[21]. Therefore, we appeal that more attention should be paid to PTDL.

We further evaluated the possible risk factors for PTDL and revealed that recipients’ but not grafts’ metabolic status determined the development of PTDL. It was not surprising to find a close correlation between PTDL and pre-transplant lipid profile as well as overweight[22,23]. It was important to determine a link between metabolic inflammation and PTDL. More importantly, most of the metabolites and cytokines that could distinguish PTDL from non-PTDL were not significantly correlated with serum lipid profile, indicating the independent predictive values. Metabolic disorders have been proved to trigger the production of inflammatory compounds, which further worsened the lipid metabolism[7]. As the key medium in this vicious circle, metabolic inflammation compounds have potential predictive abilities for the development of dyslipidemia[24,25]. In this study, we proved that recipients’ initial (pre-transplant) metabolic status was associated with PTDL. Among metabolic inflammation compounds that were differentially expressed between the PTDL and non-PTDL groups, IL-12 (p70) had the best diagnostic ability and remarkably improved the predictive value of the clinical model. In addition, IL-12 (p70) has been suggested to trigger the pathophysiology cascade of metabolomics in our study, although the conclusion needed to be confirmed by further experiments. IL-12 (p70), a member of IL12 cytokine family, was composed of IL-12 (p40) and IL-12 (p35) subunits. Generally, IL-12 (p70) is produced by dendritic cells and influence the development of Th1 cells[26]. Metabolic disorders can directly interfere with dendritic cell responses and decrease the expression IL-12 (p70)[27]. On the other hand, inflammatory cytokines could also regulate lipid metabolism[28]. For instance, IL-12 cytokine family member IL-27 could reduce lipid accumulation and enhance cholesterol efflux[29]. In addition, several other inflammatory cytokines, including IL-6, IL-1β, and tumor necrosis factor-α, have been reported to be associated with dyslipidemia[24]. For instance, in a children cohort, serum levels of IL-10, IL-17, and IL-23 were differentially expressed between dyslipidemia children and healthy individuals[30]. In patients with olanzapine-induced dyslipidemia, IL-1ra was found to be a novel biomarker[31]. However, no significant relevance of these cytokines with PTDL was found in this study.

It was also noteworthy that PTDL might increase the risk of HCC recurrence after LT. In fact, dyslipidemia plays a critical role in the pathogenesis of human malignancies, such as colorectal cancer[32], pancreatic cancer[33], and HCC[34]. Previous studies have proven that, in dyslipidemia, enriched circulating cholesterol could provoke the growth of tumors possibly through its role in maintaining the fluidity and permeability of the plasma membrane[35]. In addition, cholesterol is essential for lipid rafts synthesis, which recruits tumor signal components and initiates different downstream signals[35,36]. Interestingly, IL-12 (p70) may perform a potent anti-cancer role through the recruitment and activation of immune cells, and secretion of other cytokines[37]. A previous study demonstrated that IL-12 (p70) could down-regulate the expression Stat3, which subsequently triggered the differentiation of M1 macrophages and finally inhibited HCC growth[38]. Therefore, low serum IL-12 (P70) levels might be an indicator of both PTDL and HCC recurrence.

In contrast to the recipient, graft characteristics did not significantly correlate with PTDL in this study. In fact, the graft would be the new metabolic center in liver recipients. The crosstalk between graft and recipient occurs immediately after organ implant[39] and both grafts’ and recipients’ phenotype and genotype could affect the development of post-transplant metabolic syndrome[14,19]. We proposed a “central (liver) regulation” of metabolism in some diseases such as post-transplant diabetes[40,41]. We herein hypothesized a “peripheral (*e.g.*, fat, muscle, and the pancreas) regulation” of metabolism in PTDL.

There were limitations in this study. First, this was an HBV- and male-dominant cohort. The results need to be verified in other cohorts with large samples. Second, the follow-up time was not long enough to observe the long-term outcomes such as cardiovascular events, which was reported to be associated with PTDL[42]. In addition, the role of PTDL in HCC recurrence needs to be further studied and the underlying mechanism needs to be explored.

In conclusion, PTDL is very common in liver recipients and associated with a poor prognosis. Recipients’ pre-transplant metabolic inflammation is correlated with the development of PTDL. Among the metabolic inflammation compounds, IL-12 (p70) could be a valid predictor of PTDL and remarkably improve the predictive ability of the clinical model. We recommend prophylactic regulation of metabolic inflammation for the prevention of the development of PTDL in LT candidates, particularly those with metabolic disorders.

**ARTICLE HIGHLIGHTS**

***Research background***

Post-transplant dyslipidemia (PTDL) is a common complication in liver recipients and closely associated with organ allograft rejection, cardiovascular events, and graft dysfunction, which subsequently lead to decreased patient survival. The crosstalk between metabolic inflammation and dyslipidemia has been recently revealed.

***Research motivation***

The role of grafts’ and recipients’ metabolic status in the development of PTDL has not been evaluated.

***Research objectives***

The present study aimed to investigate the association of pre-transplant metabolite and cytokine profiles with PTDL and establish a predictive model using clinical parameters and metabolic inflammation compounds.

***Research methods***

A total of 396 adult patients who received primary liver transplantation between 2015 and 2017 were enrolled. Metabolomics and cytokines were analyzed using recipients’ pre-transplant peripheral blood in a training set (*n* = 72). In addition, an integrated prediction model was established according to the clinical risk factors and metabolic inflammation compounds and further verified in a validation set (*n* = 144).

***Research results***

A 3-month remodeling of lipid homeostasis after liver transplantation was found in liver recipients. There were 278 (70.2%) patients who developed PTDL during a follow-up period of 1.78 (1.00, 2.97) years. The PTDL group showed a significantly lower tumor-free survival and overall survival than the non-PTDL group in patients with hepatocellular carcinoma (*n* = 169). In addition, the metabolomic analysis showed that metabolic features discriminating between the PTDL and non-PTDL groups were associated with lipid and glucose metabolism-associated pathways. Among metabolites and cytokines differentially expressed between the two groups, interleukin (IL)-12 (p70) showed the best diagnostic accuracy and significantly increased the predictive value when it was incorporated into the clinical model in both training and validation sets.

***Research conclusions***

PTDL is very common in liver recipients and associated with a poor prognosis. Among the metabolic inflammation compounds, IL-12 (p70) could be a valid predictor of PTDL and remarkably improve the predictive ability of the clinical model.

***Research perspectives***

Recipients’ pre-transplant serum IL-12 (p70) level is associated with the risk of PTDL and has potential clinical value for predicting PTDL. We recommend prophylactic regulation of metabolic inflammation for the prevention of the development of PTDL in liver transplantation candidates.

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**Footnotes**

**Institutional review board statement:** The study was reviewed and approved by the First Affiliated Hospital, Zhejiang University School of Medicine Institutional Review Board.

**Informed consent statement:** All study participants, or their legal guardian, provided informed written consent prior to study enrollment.

**Conflict-of-interest statement:** The authors declare that there is no conflict of interest to be disclosed.

**Data sharing statement:** The original anonymous dataset is available on request from the corresponding author at lingqi@zju.edu.cn.

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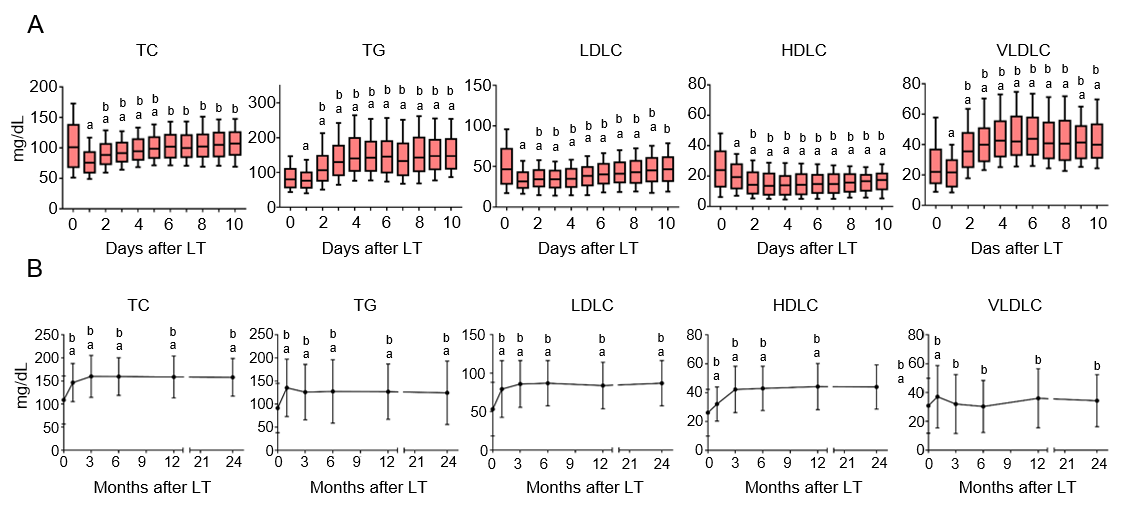
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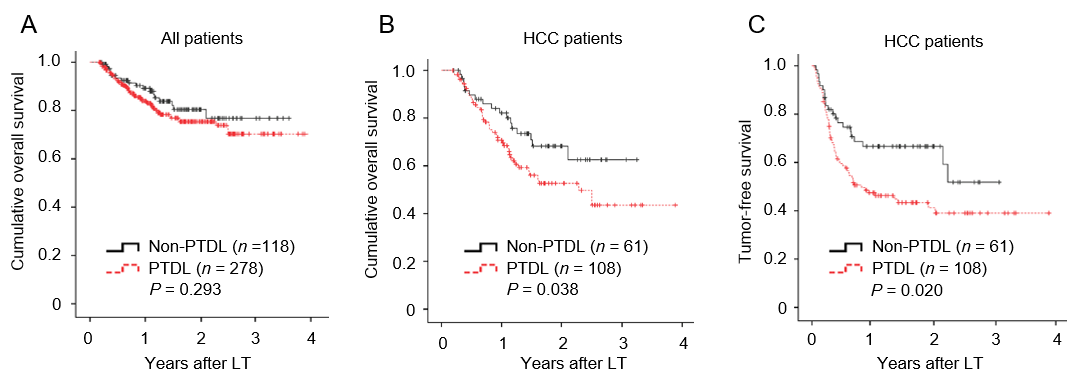
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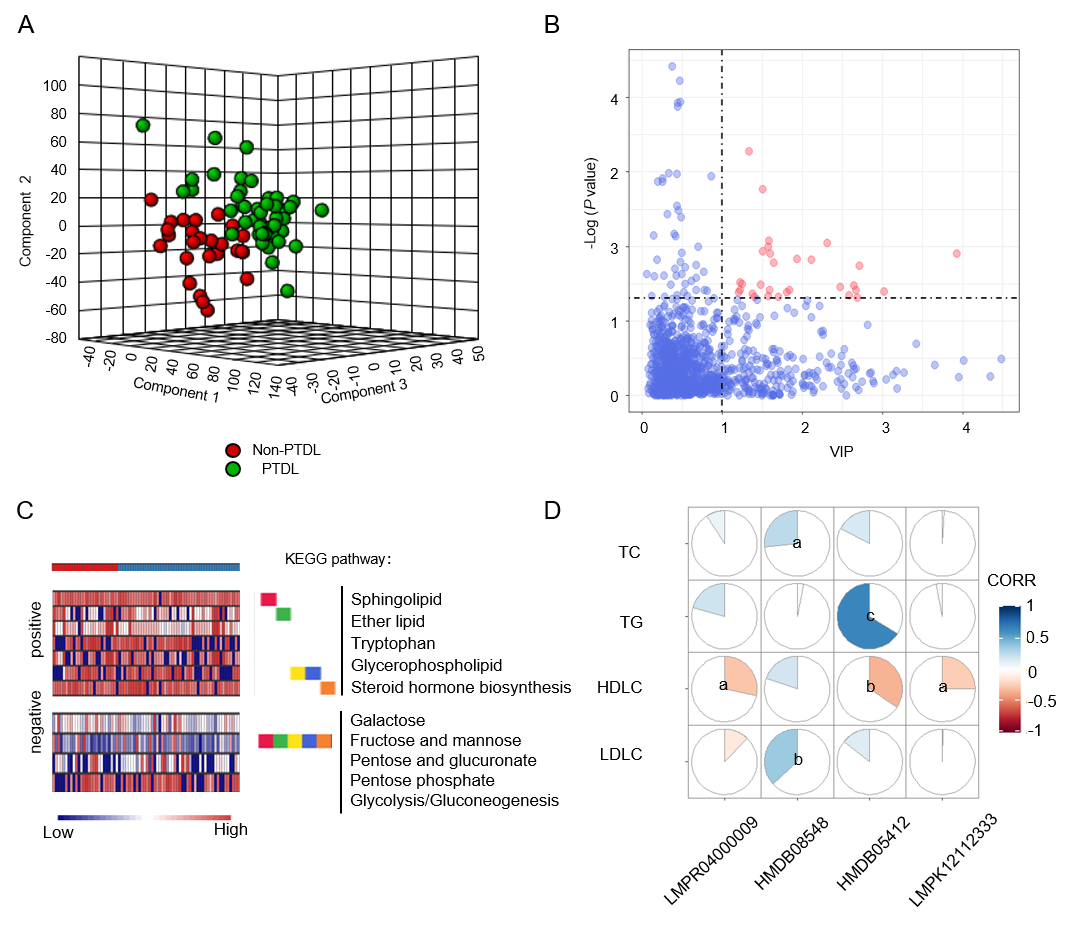
**Figure Legends**

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**Figure 1 Dynamic process of lipid profile after liver transplantation.** A: Dynamic changes of total cholesterol, triglyceride, high-density lipoprotein cholesterol, low-density lipoprotein cholesterol, and very low-density lipoprotein cholesterol during 10 d after liver transplantation; B: Dynamic changes of total cholesterol, triglyceride, high-density lipoprotein cholesterol, low-density lipoprotein cholesterol, and very low-density lipoprotein cholesterol during 24 mo after liver transplantation. Data are expressed as the mean ± SD or median (10-90 percentiles). a*P* < 0.05 *vs* pre-transplant; b*P* < 0.05 *vs* day 1. TC: Total cholesterol; TG: Triglyceride; HDLC: High-density lipoprotein cholesterol; LDLC: Low-density lipoprotein cholesterol; VLDLC: Very low-density lipoprotein cholesterol; LT: Liver transplantation.

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**Figure 2 Impact of post-transplant dyslipidemia on survival.** A: Comparison of overall survival between post-transplant dyslipidemia and non-post-transplant dyslipidemia groups in all patients; B: Comparison of overall survival between post-transplant dyslipidemia and non-post-transplant dyslipidemia groups in hepatocellular carcinoma patients; C: Comparison of tumor-free survival between post-transplant dyslipidemia and non-post-transplant dyslipidemia groups in hepatocellular carcinoma patients. PTDL: Post-transplant dyslipidemia; LT: Liver transplantation; HCC: Hepatocellular carcinoma.

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**Figure 3** **Metabolomic profiles of post-transplant dyslipidemia and non-post-transplant dyslipidemia groups.** A: 3-D scores plot between selected components in partial least squares-discriminant analysis; B: Thirty differentially expressed metabolites (scatter plot; variable importance in the projection > 1, *P* < 0.05) were annotated with red color; C: Concentrated display of the abundant differential features (positive and negative ion modes on the top and bottom, respectively) between non-post-transplant dyslipidemia and post-transplant dyslipidemia groups using Mummichog 2.0; D: Correlation between metabolites and lipid profile in recipients. a*P* < 0.05; b*P* < 0.01; c*P* < 0.001. PTDL: Post-transplant dyslipidemia; VIP: Variable Importance in the Projection; TC: Total cholesterol; TG: Triglycerides; HDLC: High-density lipoprotein cholesterol; LDLC: Low-density lipoprotein cholesterol; CORR: Correlation.

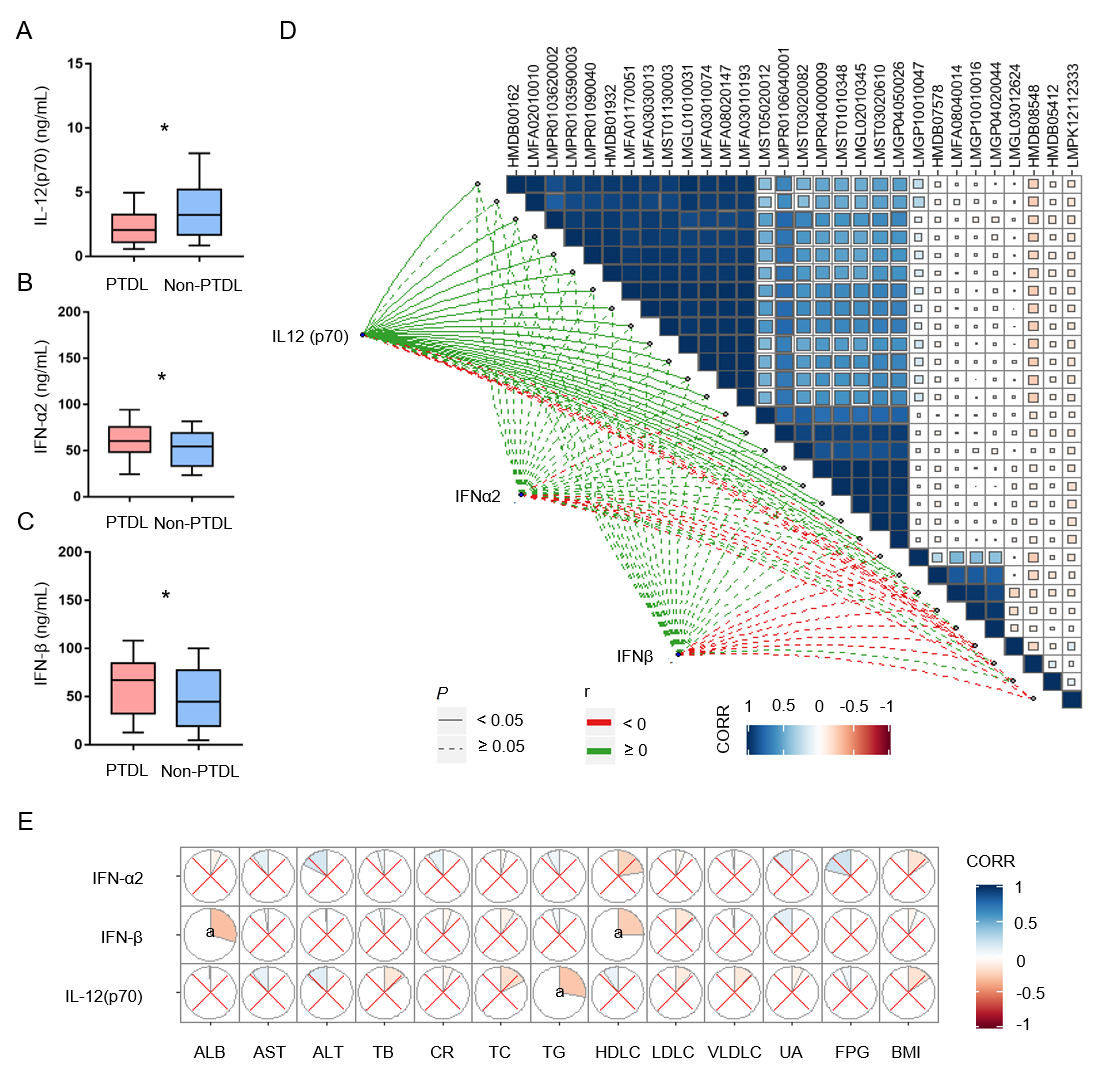
**Figure 4 Peripheral cytokine profile of post-transplant dyslipidemia and non-post-transplant dyslipidemia groups.** A-C: The interleukin (IL)-12 (p70), IFN-α2, and IFN-β levels were significantly differentially expressed between post-transplant dyslipidemia group and non-post-transplant dyslipidemia group; D: Correlations between differential cytokines and metabolic profiles; E: Correlation between differential cytokines and recipients’ pre-transplant clinical metabolic parameters. a*P* < 0.05. PTDL: Post-transplant dyslipidemia; ALB: Albumin; AST: Aspartate aminotransferase; ALT: Alanine aminotransferase; TB: Total bilirubin; CR: Creatinine; TC: Total cholesterol; TG: Triglyceride; HDLC: High-density lipoprotein cholesterol; LDLC: Low density lipoprotein cholesterol; VLDLC: Very low-density lipoprotein cholesterol; UA: Uric acid; FBG: Fasting blood glucose; BMI: Body mass index; CORR: Correlation.

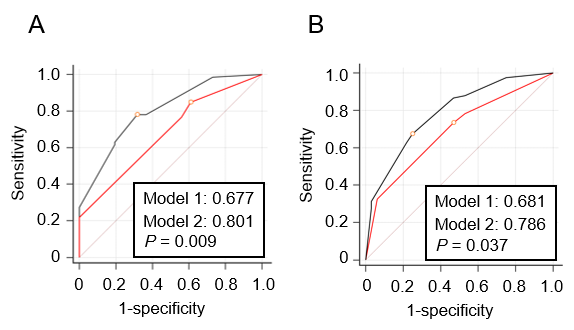
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**Figure 5 Comparison of predictive ability between clinical model (model 1) and integrated model (model 2).** A: Areas under the receiver operating characteristic curves of model 1 and model 2 in training set (*n* = 72); B: Areas under the receiver operating characteristic curves of model 1 and model 2 in validation set (*n* = 144).

**Table 1** **Comparison of perioperative findings between post-transplant dyslipidemia and non-post-transplant dyslipidemia groups**

|  |  |  |  |
| --- | --- | --- | --- |
|  | **PTDL (*n* = 278)** | **Non-PTDL (*n* = 118)** | ***P* value** |
| Recipient factors |  |  |  |
| Age (yr) | 48.2 ± 9.6 | 49.1 ± 9.1 | 0.411 |
| Male, *n* (%) | 237 (85.3) | 102 (86.4) | 0.758 |
| BMI (kg/m2) | 23.2 ± 3.2 | 22.3 ± 2.9 | 0.012 |
| MELD score | 20.9 ± 10.9 | 20.6 ± 11.7 | 0.777 |
| Laboratory value |  |  |  |
| Creatinine (μmol/L) | 65.0 (53.0-83.8) | 66.0 (55.0-78.0) | 0.925 |
| Albumin (g/L) | 34.7 (31.0-38.2) | 35.5 (32.2-39.6) | 0.218 |
| TB (μmol/L) | 63.5 (28.0-335.5) | 58.5 (22.3-205.8) | 0.066 |
| INR | 1.5 (1.3-1.9) | 1.4 (1.2-1.7) | 0.087 |
| AST (U/L) | 66.0 (37.0-128.8) | 71.5 (38.3-125.5) | 0.694 |
| ALT (U/L) | 43 (24.0-93.3) | 40.5 (26.0-125.0) | 0.621 |
| TC (mg/dL) | 100.5 (65.7-139.2) | 104.4 (73.5-135.3) | 0.947 |
| TG (mg/dL) | 88.6 (62.0-115.1) | 70.9 (53.1-97.4) | 0.017 |
| HDLC (mg/dL) | 23.2 (11.6-34.8) | 27.1 (15.5-42.5) | 0.024 |
| LDLC (mg/dL) | 47.6 (27.1-73.5) | 46.4 (30.9-69.6) | 0.676 |
| VLDLC (mg/dL) | 23.2 (15.5-38.7) | 19.3 (11.6-30.9) | 0.052 |
| FBG (mmol/L) | 6.3 (5.3-8.0) | 6.0 (5.3-7.4) | 0.328 |
| Cirrhosis, *n* (%) | 206 (74.1) | 94 (79.7) | 0.238 |
| HCC, *n* (%) | 108 (38.8) | 61 (51.7) | 0.018 |
| HBV status, *n* (%) |  |  |  |
| HBsAg positive | 215 (77.3) | 102 (86.4) | 0.038 |
| HBeAg positive | 81 (29.1) | 27 (22.9) | 0.201 |
| HBV DNA > 1000 copies/mL | 39 (14.0) | 18 (15.3) | 0.751 |
| Pre-LT AVT | 108 (38.8) | 52 (44.1) | 0.333 |
| Comorbidities, *n* (%) |  |  |  |
| Hepatic encephalopathy | 50 (18.0) | 25 (21.2) | 0.457 |
| Hepatorenal syndrome | 22 (7.9) | 10 (8.5) | 0.851 |
| Gastrointestinal bleeding | 57 (20.5) | 14 (11.9) | 0.040 |
| Ascites | 104 (37.4) | 31 (26.3) | 0.032 |
| Donor factors |  |  |  |
| Age (yr) | 38.8 ± 12.3 | 41.3 ± 12.8 | 0.071 |
| Male, *n* (%) | 235 (84.5) | 99 (83.9) | 0.874 |
| Cause of death |  |  |  |
| Trauma | 171 (61.5) | 71 (60.2) | 0.802 |
| CVA | 87 (31.3) | 37 (31.4) | 0.934 |
| Other | 21 (7.6) | 10 (8.5) | 0.755 |
| DBD (*vs* DCD) | 49 (17.6) | 17 (14.4) | 0.432 |
| BMI (kg/m2) | 22.6 ± 2.6 | 23.0 ± 2.7 | 0.169 |
| Graft weight (kg) | 1.4 ± 0.3 | 1.4 ± 0.3 | 0.228 |
| Macrovesicular steatosis | 50 (18.0) | 22 (18.6) | 0.877 |
| Creatinine (μmol/L) | 77.0 (56.5-118.5) | 82.2 (56.1-121.3) | 0.685 |
| Albumin (g/L) | 31.0 (28.0-36.5) | 30.9 (26.7-36.7) | 0.474 |
| TB (μmol/L) | 14.1 (9.1-23.4) | 16.5 (11.1-26.5) | 0.078 |
| AST (U/L) | 52.0 (33.0-87.2) | 51.5 (31.3-94.5) | 0.694 |
| ALT (U/L) | 34.0 (22.0-69.0) | 39.0 (22.5-76.3) | 0.345 |
| Operative factors |  |  |  |
| WIT (min) | 50.7 ± 14.3 | 47.0 ± 10.8 | 0.006 |
| CIT (h) | 8.4 ± 3.1 | 8.7 ± 3.4 | 0.359 |
| Blood loss (L) | 1.0 (0.8-2.0) | 1.0 (0.8-1.5) | 0.117 |
| Immunosuppressant, *n* (%) |  |  |  |
| IL2R antibody | 204 (73.4) | 93 (78.8) | 0.254 |
| Corticosteroid | 156 (56.1) | 58 (49.2) | 0.204 |
| Tacrolimus | 245 (88.1) | 103 (87.3) | 0.814 |

PTDL: Post-transplant dyslipidemia; BMI: Body mass index; MELD: Model for end-stage liver disease; TB: Total bilirubin; INR: International normalized ratio; AST: Aspartate aminotransferase; ALT: Alanine aminotransferase; TC: Total cholesterol; TG: Triglyceride; HDLC: High-density lipoprotein cholesterol; LDLC: Low density lipoprotein cholesterol; VLDLC: Very low-density lipoprotein cholesterol; FBG: Fasting blood glucose; HCC: Hepatocellular carcinoma; HBV: Hepatitis B virus; LT: Liver transplantation; AVT: Antiviral treatment; CVA: Cerebral vascular accident; DBD: Donation after brain death; DCD: Donor after circulatory death; WIT: Warm ischemia time; CIT: Cold ischemia time.

**Table 2 Comparison of post-transplant complications between post-transplant dyslipidemia and non-post-transplant dyslipidemia groups**

|  |  |  |  |
| --- | --- | --- | --- |
|  | **PTDL (*n* = 278)** | **Non-PTDL (*n* = 118)** | ***P* value** |
| Acute kidney injury | 69 (24.8) | 27 (22.9) | 0.681 |
| Early allograft dysfunction | 84 (30.2) | 37 (31.4) | 0.822 |
| Portal vein stenosis | 5 (1.8) | 2 (1.7) | 0.943 |
| CMV infection | 4 (1.4) | 3 (2.5) | 0.730 |
| Bacterial infection | 28 (10.1) | 7 (5.9) | 0.184 |
| Diabetes mellitus | 21 (7.6) | 5 (4.2) | 0.233 |
| Hypertension | 13 (4.7) | 7 (5.9) | 0.602 |
| Cardiovascular diseases | 7 (2.5) | 1 (0.8) | 0.490 |
| Chronic kidney failure | 3 (1.1) | 0 (0) | 0.681 |
| Biliary complications | 40 (14.4) | 15 (12.7) | 0.659 |
| HCC recurrence1 | 60 (55.5) | 21 (34.4) | 0.008 |

1In 169 HCC patients. PTDL: Post-transplant dyslipidemia; CMV: Cytomegalovirus; HCC: Hepatocellular carcinoma.

**Table 3 Logistic regression analysis of variables related to post-transplant dyslipidemia**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
|  | **Univariate** |  | **Multivariate** |  |
|  | **OR (95% CI)** | ***P* value** | **OR (95%CI)** | ***P* value** |
| Overweight | 2.726 (1.494-4.973) | 0.001 | 2.705 (1.457-5.020) | 0.002 |
| Hyper-TG | 7.846 (1.032-59.679) | 0.047 |  |  |
| Hypo-HDLC | 3.116 (1.861-5.217) | < 0.001 | 3.090 (1.828-5.224) | < 0.001 |
| Hepatitis B | 0.489 (0.261-0.915) | 0.025 |  |  |
| Gastrointestinal bleeding | 1.916 (1.021-3.595) | 0.043 |  |  |
| Ascites | 1.677 (1.042-2.701) | 0.033 |  |  |
| HCC | 0.594 (0.385-0.916) | 0.019 |  |  |
| WIT | 2.286 (1.147-4.557) | 0.019 |  |  |

Overweight was defined as body mass index > 25 kg/m2; hyper-triglyceride was defined as triglyceride ≥ 200 mg/dL; hypo-high density lipoprotein cholesterol was defined as high density lipoprotein cholesterol < 40 mg/dL. The warm ischemia time and total intravenous fluids were dichotomized at the median into high and low levels. OR: Odds ratio; CI: Confidence interval; TG: Triglyceride; HDLC: High density lipoprotein cholesterol; HCC: Hepatocellular carcinoma; WIT: Warm ischemia time.

**Table 4 Multivariate logistic regression analysis of risk factors associated with post-transplant dyslipidemia**

|  |  |  |  |
| --- | --- | --- | --- |
|  |  | **Multivariate** | ***P* value** |
| ***β*** | **OR (95%CI)** |
| Model 1 |  |  |  |
| Recipient overweight | 0.991 | 2.693 (1.451-4.998) | 0.002 |
| Recipient hypo-HDLC | 1.134 | 3.107 (1.838-5.251) | < 0.001 |
| Constant | -0.249 |  |  |
| Model 2 | | | |
| Recipient overweight | 1.625 | 3.425 (1.297-9.046) | 0.027 |
| Recipient hypo-HDLC | 1.231 | 5.077 (1.206-21.381) | 0.013 |
| Low IL-12 (p70) level | 1.846 | 6.336 (2.553-15.726) | < 0.001 |
| Constant | -1.494 |  |  |

Overweight was defined as body mass index > 25 kg/m2; hypo-high-density lipoprotein cholesterol was defined as high-density lipoprotein cholesterol < 40 mg/dL; interleukin-12 (p70) level was dichotomized at the median into high and low levels. OR: Odds ratio; CI: Confidence interval; HDLC: High-density lipoprotein cholesterol.