

World Journal of *Stem Cells*

World J Stem Cells 2020 August 26; 12(8): 706-896



THERAPEUTIC AND DIAGNOSTIC GUIDELINES

- 706 Hunting down the dominating subclone of cancer stem cells as a potential new therapeutic target in multiple myeloma: An artificial intelligence perspective
Lee LX, Li SC

OPINION REVIEW

- 721 Off-the-shelf mesenchymal stem cells from human umbilical cord tissue can significantly improve symptoms in COVID-19 patients: An analysis of evidential relations
Pham PV, Vu NB

REVIEW

- 731 Mesenchymal stromal cells as potential immunomodulatory players in severe acute respiratory distress syndrome induced by SARS-CoV-2 infection
Mallis P, Michalopoulos E, Chatzistamatiou T, Stavropoulos-Giokas C
- 752 Practical choice for robust and efficient differentiation of human pluripotent stem cells
Fang M, Liu LP, Zhou H, Li YM, Zheng YW
- 761 Human embryonic stem cells as an *in vitro* model for studying developmental origins of type 2 diabetes
Chen ACH, Lee KF, Yeung WSB, Lee YL
- 776 Autophagy in fate determination of mesenchymal stem cells and bone remodeling
Chen XD, Tan JL, Feng Y, Huang LJ, Zhang M, Cheng B
- 787 Stem cell therapy for Alzheimer's disease
Liu XY, Yang LP, Zhao L
- 803 Exosomes derived from stem cells as an emerging therapeutic strategy for intervertebral disc degeneration
Hu ZL, Li HY, Chang X, Li YY, Liu CH, Gao XX, Zhai Y, Chen YX, Li CQ
- 814 Mesenchymal stem cell-derived exosomes: Toward cell-free therapeutic strategies in regenerative medicine
Ma ZJ, Yang JJ, Lu YB, Liu ZY, Wang XX

ORIGINAL ARTICLE**Basic Study**

- 841 Assessment of tobacco heating system 2.4 on osteogenic differentiation of mesenchymal stem cells and primary human osteoblasts compared to conventional cigarettes
Aspera-Werz RH, Ehnert S, Müller M, Zhu S, Chen T, Weng WD, Jacoby J, Nussler AK

- 857 Human embryonic stem cell-derived mesenchymal stem cells improved premature ovarian failure
Bahrehbar K, Rezazadeh Valojerdi M, Esfandiari F, Fathi R, Hassani SN, Baharvand H

SYSTEMATIC REVIEWS

- 879 Role of mesenchymal stem cell derived extracellular vesicles in autoimmunity: A systematic review
Wang JH, Liu XL, Sun JM, Yang JH, Xu DH, Yan SS

ABOUT COVER

Editorial Board member of *World Journal of Stem Cells*, Dr. Perez-Campo is currently an Associate Professor in the Department of Molecular Biology at the University of Cantabria (Spain). She obtained her degree in Biological Sciences from the University of Salamanca (Spain), where she then went on to complete her PhD in 1999. Dr. Perez-Campo undertook her postdoctoral research at the Paterson Institute for Cancer Research (United Kingdom; currently known as Cancer Research UK Manchester Institute) under the supervision of Prof. Lacaud, where she remained for more than 10 years working in the field of stem cell biology. Upon returning to Spain, she joined the University of Cantabria and focused her research efforts on the molecular mechanisms that control mesenchymal stem cell (MSC) differentiation towards the osteoblastic and adipogenic lineages, and how those mechanisms are altered in osteoporosis. (L-Editor: Filipodia)

AIMS AND SCOPE

The primary aim of *World Journal of Stem Cells* (*WJSC*, *World J Stem Cells*) is to provide scholars and readers from various fields of stem cells with a platform to publish high-quality basic and clinical research articles and communicate their research findings online. *WJSC* publishes articles reporting research results obtained in the field of stem cell biology and regenerative medicine, related to the wide range of stem cells including embryonic stem cells, germline stem cells, tissue-specific stem cells, adult stem cells, mesenchymal stromal cells, induced pluripotent stem cells, embryonal carcinoma stem cells, hemangioblasts, lymphoid progenitor cells, *etc.*

INDEXING/ABSTRACTING

The *WJSC* is now indexed in Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports/Science Edition, Biological Abstracts, BIOSIS Previews, PubMed, and PubMed Central. The 2020 Edition of Journal Citation Reports® cites the 2019 impact factor (IF) for *WJSC* as 3.231; IF without journal self cites: 3.128; Ranking: 18 among 29 journals in cell and tissue engineering; Quartile category: Q3; Ranking: 113 among 195 journals in cell biology; and Quartile category: Q3.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Yan-Xia Xing; Production Department Director: Yun-Xiaojuan Wu; Editorial Office Director: Jin-Lai Wang.

NAME OF JOURNAL

World Journal of Stem Cells

ISSN

ISSN 1948-0210 (online)

LAUNCH DATE

December 31, 2009

FREQUENCY

Monthly

EDITORS-IN-CHIEF

Carlo Ventura

EDITORIAL BOARD MEMBERS

<https://www.wjgnet.com/1948-0210/editorialboard.htm>

PUBLICATION DATE

August 26, 2020

COPYRIGHT

© 2020 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

<https://www.wjgnet.com/bpg/gerinfo/204>

GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjgnet.com/bpg/GerInfo/287>

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjgnet.com/bpg/gerinfo/240>

PUBLICATION ETHICS

<https://www.wjgnet.com/bpg/GerInfo/288>

PUBLICATION MISCONDUCT

<https://www.wjgnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

<https://www.wjgnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjgnet.com/bpg/GerInfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>

Role of mesenchymal stem cell derived extracellular vesicles in autoimmunity: A systematic review

Jing-Hua Wang, Xiao-Ling Liu, Jian-Mei Sun, Jing-Han Yang, Dong-Hua Xu, Shu-Shan Yan

ORCID number: Jian-Hua Wang 0000-0002-8310-8003; Xiao-Ling Liu 0000-0002-7172-575X; Jian-Mei Sun 0000-0001-7305-5588; Jing-Han Yang 0000-0001-7577-2695; Dong-Hua Xu 0000-0002-9146-7858; Shu-Shan Yan 0000-0002-8194-6409.

Author contributions: All authors equally contributed to this work with regard to conception and design of the study, literature analysis, manuscript drafting, critical revision, and editing, and approval of the final version.

Supported by the Shandong Natural Science Foundation, No. ZR2019QH012.

Conflict-of-interest statement: The authors have no potential conflicts of interests to declare.

PRISMA 2009 Checklist statement: The guidelines of the PRISMA 2009 statement have been adopted.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works

Jing-Hua Wang, Jing-Han Yang, Clinical Medicine College, Weifang Medical University, Weifang 261000, Shandong Province, China

Xiao-Ling Liu, Department of Emergency Medicine, Yantai Shan Hospital, Yantai 264001, Shandong Province, China

Jian-Mei Sun, Department of Chemistry, School of Applied Chemistry, Food and Drug, Weifang Engineering Vocational College, Qingzhou 262500, Shandong Province, China

Dong-Hua Xu, Department of Rheumatology of the First Affiliated Hospital, Weifang Medical University, Central Laboratory of the First Affiliated Hospital, Weifang 261000, Shandong Province, China

Shu-Shan Yan, Department of Gastrointestinal and Anal Diseases Surgery of the Affiliated Hospital, Weifang Medical University, Weifang 261000, Shandong Province, China

Corresponding author: Shu-Shan Yan, MD, Doctor, Department of Gastrointestinal and Anal Diseases Surgery, The Affiliated Hospital, Weifang Medical University, Weifang 261000, Shandong Province, China. yanshushan@163.com

Abstract

BACKGROUND

Mesenchymal stem cells (MSCs) have been reported to possess immune regulatory effects in innate and adaptive immune reactions. MSCs can mediate intercellular communications by releasing extracellular vesicles (EVs), which deliver functional molecules to targeted cells. MSC derived EVs (MSC-EVs) confer altering effects on many immune cells, including T lymphocytes, B lymphocytes, natural killer cells, dendritic cells, and macrophages. A large number of studies have suggested that MSC-EVs participate in regulating autoimmunity related diseases. This characteristic of MSC-EVs makes them be potential biomarkers for the diagnosis and treatment of autoimmunity related diseases.

AIM

To verify the potential of MSC-EVs for molecular targeted therapy of autoimmunity related diseases.

METHODS

Literature search was conducted in PubMed to retrieve the articles published between 2010 and 2020 in the English language. The keywords, such as "MSCs,"

on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Received: March 4, 2020

Peer-review started: March 4, 2020

First decision: April 25, 2020

Revised: July 2, 2020

Accepted: July 19, 2020

Article in press: July 19, 2020

Published online: August 26, 2020

P-Reviewer: Pelagalli A, Khan I, Kuo FC

S-Editor: Zhang L

L-Editor: Wang TQ

P-Editor: Li X



“EVs,” “exosome,” “autoimmunity,” “tumor immunity,” and “transplantation immunity,” and Boolean operator “AND” and “NOT” coalesced admirably to be used for searching studies on the specific molecular mechanisms of MSC-EVs in many immune cell types and many autoimmunity related diseases. Studies that did not investigate the molecular mechanisms of MSC-EVs in the occurrence and development of autoimmune diseases were excluded.

RESULTS

A total of 96 articles were chosen for final reference lists. After analyzing those publications, we found that it had been well documented that MSC-EVs have the ability to induce multiple immune cells, like T lymphocytes, B lymphocytes, natural killer cells, dendritic cells, and macrophages, to regulate immune responses in innate immunity and adaptive immunity. Many validated EVs-delivered molecules have been identified as key biomarkers, such as proteins, lipids, and nucleotides. Some EVs-encapsulated functional molecules can serve as promising therapeutic targets particularly for autoimmune disease.

CONCLUSION

MSC-EVs play an equally important part in the differentiation, activation, and proliferation of immune cells, and they may become potential biomarkers for diagnosis and treatment of autoimmunity related diseases.

Key words: Mesenchymal stem cells; Extracellular vesicles; Exosome; Autoimmunity; Tumor immunity; Transplantation immunity

©The Author(s) 2020. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Mesenchymal stem cells (MSCs) have been reported to possess immunomodulatory effects on autoimmune responses. MSCs can mediate intercellular communications by releasing extracellular vesicles (EVs), which deliver functional molecules to targeted cells. MSC derived EVs (MSC-EVs) exert immunomodulatory effects on many immune cells. A large number of studies have suggested that MSC-EVs and the encapsulated bioactive molecules are potential targets for autoimmune disease, cancer, and other diseases. However, there is still a long way for investigating the molecular mechanism of MSC-EVs in autoimmunity. This review will focus on the immunomodulatory function and underlying mechanism of MSC-EVs in autoimmunity related diseases.

Citation: Wang JH, Liu XL, Sun JM, Yang JH, Xu DH, Yan SS. Role of mesenchymal stem cell derived extracellular vesicles in autoimmunity: A systematic review. *World J Stem Cells* 2020; 12(8): 879-896

URL: <https://www.wjgnet.com/1948-0210/full/v12/i8/879.htm>

DOI: <https://dx.doi.org/10.4252/wjsc.v12.i8.879>

INTRODUCTION

Mesenchymal stem cells (MSCs) are a group of common multipotent progenitor cells, which can be found in bone marrow^[1,2], synovium^[3,4], umbilical cord^[5], and adipose tissue^[6]. They are characterized by a multilineage differentiation potential and paracrine function^[7]. There is growing evidence that MSCs exert immunomodulatory effects through their paracrine function^[8], in which multiple small molecules, including extracellular vesicles (EVs), cytokines, chemokines, growth factors, and interleukin (IL), are secreted to the extracellular microenvironment in animal models^[9]. Recently, numerous studies demonstrated that MSCs can be used in clinical therapy for immunomodulation and regenerative medicine *in vivo* and *in vitro*^[10-12]. Despite great improvements in the MSC therapeutic strategies for autoimmune diseases, treatment failures are still common and there is no doubt that it is imperative to carry out more studies to investigate the specific molecular mechanisms. EVs are key components of the paracrine process that play a vital role in intercellular communication by transmitting biological molecules in pathological and physiological

conditions.

EV is newly identified small vesicle wrapped in lipid membranes, which is widely produced by many cells and secreted into the extracellular microenvironment. In 1967, Wolf first discovered EVs and described them as function-free platelet wastes^[13]. EVs can be isolated from various extracellular fluids, like blood, urine, saliva, tear, cerebrospinal fluid, milk, and so on, and various cells, including stem cells^[14-18], primary cells of the immune and nervous system^[19-22], and multiple cancer cell types^[23-25]. Their encapsulated functional molecules can be novel biomarkers and therapeutic targets for many kinds of diseases, for instance, cancer, autoimmune diseases, and neurodegenerative disorders. The role of EVs in immunity and inflammation regulations has been attracting attention during the past few decades. According to diameter, EVs can be divided into three types, including apoptotic bodies, microparticles, and exosomes (Figure 1)^[26,27]. Exosomes are the most common EVs with a diameter of 50-100 nm^[28]. Exosomes were first discovered in sheep reticulocytes, by electron microscopy^[29]. Microparticles, also called microvesicles, are submicronic vesicles with a diameter of 100-1000 nm, which are formed by budding of the cellular membrane after cell stimulation or stress, such as cell activation, apoptosis, and hypoxia. Apoptotic bodies also belong to EVs with a diameter of 50-4000 nm. They are usually released during the stage of cell apoptosis. EVs participate in the intercellular communication by delivering numerous proteins and nucleotides with biological activity, and nucleotides include microRNAs (miRNAs), long non-coding RNAs (lncRNAs), mRNA, and even extra-chromosomal DNA^[30,31]. They play vital roles in regulating inflammation, immune response, vascular reactivity, and tissue repair^[32,33]. During the past few decades, MSC derived EVs (MSC-EVs) have been implicated in regulating inflammation and autoimmunity^[7]. It has been well established that EVs are involved in regulating autoimmune disorders by delivering a large number of bioactive molecules, including cytokines, enzymes, transcription factors, cytokines receptor antagonists, miRNAs, lncRNAs, and circRNAs^[34]. MSCs, as a specific group of cells, are multipotent stem cells characterized by immunomodulatory and self-renew properties^[35,36]. Cosenza *et al.*^[37] have reported the important pathogenic or therapeutic role of MSC-EVs in rheumatic diseases. Therefore, we proposed that MSC-EVs can become potential biotargets for the development of novel molecular targeted drugs in autoimmune related diseases based on the above conclusions. This systematic review will provide in-depth knowledge of biogenesis and functional roles of MSC-EVs, especially exosomes, in autoimmunity.

MATERIALS AND METHODS

Literature search

The key words “MSCs,” “EVs,” “exosome,” “autoimmunity,” “tumor immunity,” and “transplantation immunity” were used to retrieve relevant articles published in English from 2010 to 2020 in PubMed database. Besides, Boolean operator “AND” and “NOT” were combined admirably with those keywords to search the related articles. Reference lists from those articles were reviewed to exclude irrelevant articles. Manuscripts available were reviewed and recognized by using document management tool. All available information was obtained by skimming the abstracts of searched articles. Data were analyzed using descriptive statistics.

All repetitive documents were excluded, and the remainder needed to be restored for reading. Nevertheless, full text retrieval was performed due to many documents with unavailable abstract.

Statistical analysis

This article was a systematic review and no statistical method was used in this article.

RESULTS

Initially, we retrieved 198 records for this review. Then, repetitive and irrelevant documents were excluded, and we retained ultimately 96 high-quality papers with innovative viewpoints for reference lists. The screening process of those documents is showed in Figure 2.

EVs can regulate many immune and inflammatory responses by mediating intercellular communication. Moreover, MSC-EVs have been well documented to

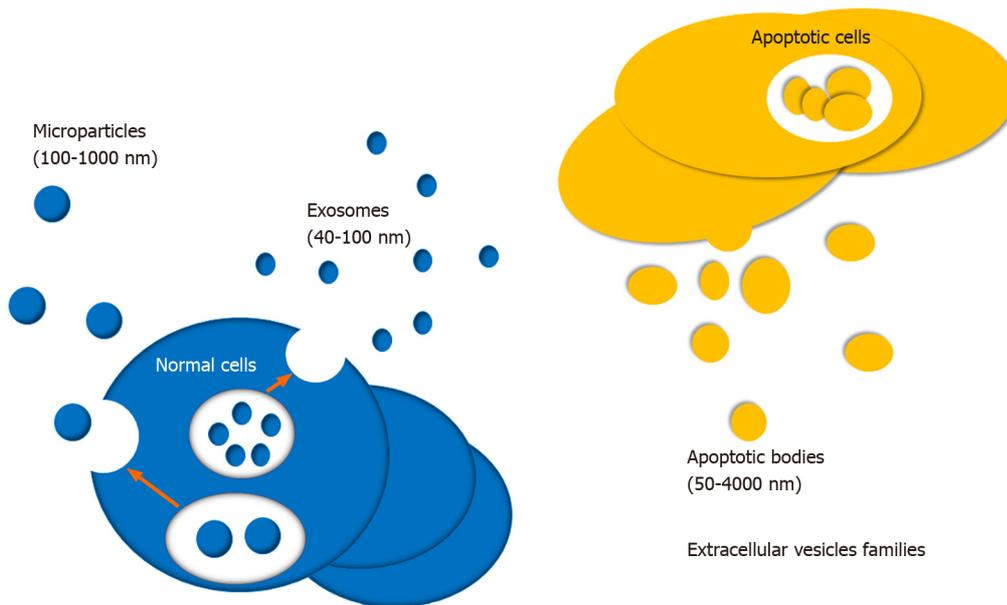


Figure 1 Various kinds of extracellular vesicles. Extracellular vesicles primarily consist of exosomes, apoptotic bodies, and microparticles derived for normal cells or apoptotic cells.

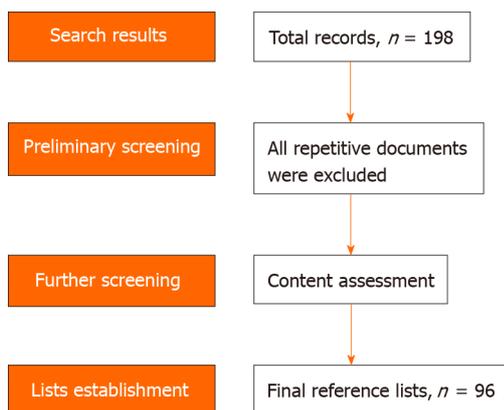


Figure 2 Flowchart for literature retrieval and screening. Flowchart shows that the repeated siftings have brought 198 search records to the 96 articles for final reference lists.

induce multiple immune cells to mediate immune responses in innate immunity and adaptive immunity, namely, they modulate the differentiation, activation, and proliferation of immune cells, like T lymphocytes, B lymphocytes, natural killer cells (NKs), dendritic cells (DCs), and macrophages in the autoimmune system (Figure 3)^[38-41].

There is growing evidence that MSC-EVs serving as a type of signal molecules play major biological roles in the initiation, maintenance, and progression of multiple autoimmune related diseases, such as autoimmune diseases, cancer, and graft-versus-host disease. The features of MSC-EVs immunomodulation and their therapeutic potential in autoimmune related diseases are summarized in Tables 1 and 2.

DISCUSSION

MSC-EVs and T lymphocytes

T lymphocytes are important immune cells in adaptive immunity and play a significant role in the occurrence and development of many autoimmune and inflammatory diseases. MSC derived exosomes and microparticles down-regulate T cell proliferation, and CD4⁺ and CD8⁺ T cell subsets decrease significantly in quantity^[7]. Adipose mesenchymal stem cell (AMSC) derived exosomes depress the

Table 1 Immune modulation of extracellular vesicles in autoimmune related diseases

Disease	EVs	Expression	MSC-EVs source/ Target molecules	Target immune cells	Pathway(s) involved	Ref.
RA	MSC derived exosomal miR-150-5p	Down	Bone marrow derived MSC-EVs/MMP 14 and VEGF	Macrophages	TGF- β pathway	Chen <i>et al</i> ^[80]
	Exosome-encapsulated miR-548a-3p	Down	TLR4	Macrophages	MiR-548a-3p/TLR4/NF- κ B axis	Wang <i>et al</i> ^[81]
	Exosome-encapsulated miR-6089	Down	TLR4	Macrophages	TLR4/NF- κ B signaling pathway	Xu <i>et al</i> ^[82]
	Exosome-derived lncRNA Hotair	Up	MMP-2 and MMP-13	Macrophages	-	Song <i>et al</i> ^[83]
	Exosomal miR-17	Up	TGFBR II	T cells	-	Wang <i>et al</i> ^[84]
	MicroRNA-155	Up	SHIP-1	Macrophages	-	Kurowska-Stolarska <i>et al</i> ^[85]
	MicroRNA-146	Up	-	Macrophages, T cells, B cells	-	Nakasa <i>et al</i> ^[86]
SLE	Exosomal miR-26a	Up	Podocyte proteins, actin family members, and intermediate filaments	Podocytes	-	Ichii <i>et al</i> ^[99]
	Exosomal miRNA-146a	Up	-	-	Interferon- γ pathway	Perez-Hernandez <i>et al</i> ^[100]
pSS	EV derived LCN2	Up	TNF- α	B cells	TNF- α signaling	Aqrabi <i>et al</i> ^[107]
	EV derived APMAP	Up	TNF- α	B cells	TNF- α signaling	Aqrabi <i>et al</i> ^[107]
	EV derived CPNE1	Up	TNF- α	B cells	TNF- α signaling	Aqrabi <i>et al</i> ^[107]
IBD	MSC-EVs	Up	Bone marrow derived MSC-EVs	Macrophages	JAK1/STAT1/STAT6 signaling pathway	Cao <i>et al</i> ^[113]
Breast cancer	Exosomal PD-L1	Down	PD-1	T cells	PD-L1/ PD-1 pathway	Yang <i>et al</i> ^[120]
Lung cancer	EV derived miR-103a	Up	Lung cancer cell derived EVs/PTEN	Macrophages	PI3K/ AKT and STAT3 axis	Hsu <i>et al</i> ^[121]
Pancreatic cancer	Exosomal miR-301a-3p	Up	PTEN	Macrophages	PI3K γ signaling pathway	Wang <i>et al</i> ^[122]
GVHD	MSC-EVs	Up	T cell derived EVs	-	-	Park <i>et al</i> ^[126]

RA: Rheumatoid arthritis; SLE: Systemic lupus erythematosus; pSS: Primary Sjgren's syndrome; IBD: Inflammatory bowel diseases; GVHD: Graft-versus-host disease; MSC: mesenchymal stem cell; EV: Extracellular vesicle; MSC-EV: Mesenchymal stem cell derived extracellular vesicle; MMP: Matrix metalloproteinase; VEGF: Vascular endothelial growth factor; TGFBR II: Transforming growth factor beta receptor II; SHIP-1: Src homology 2-containing inositol phosphatase-1; PD-1: Programmed death-1; PD-L1: PD-1 ligand; LCN2: Neutrophil gelatinase-associated lipocalin; APMAP: Adipocyte plasma membrane-associated protein.

activity of T cells, and up-regulate IL-4, IL-10, and transforming growth factor- β and down-regulate IL-17 and interferon- γ in streptozotocin induced type-1 diabetes mellitus mice, thus deadening the progression of diseases^[42]. MSCs have been extensively reported to decorate the activation of CD4⁺ T cells by some specific T cell effector cytokines or direct contact, down-regulating their immune activity and converting them to a regulatory phenotype (Treg)^[43,44]. Programmed death-1 (PD-1) is a valuable cytokine inducing T cell activity. Research shows that MSCs express and secrete PD-1 ligands (PD-L1 and PD-L2) to regulate T cell dependent immune responses by binding with PD-1^[45], suggesting that MSCs possess immunosuppressive properties *via* the modulation of T cells. AMSCs under stimulation with IFN- γ can secrete a big body of exosomes to the conditioned medium, and importantly, T cells isolated from that medium are significantly inhibited in activity and proliferation^[46]. In a word, MSC-EVs down-regulate the activity and proliferation of T cells to inhibit T dependent autoimmune responses.

Table 2 Therapeutic potential of extracellular vesicles

Disease	EVs	Experimental sample	Therapeutic potential	Ref.
RA	MSC derived exosomal miR-150-5p	Collagen induced arthritis mouse model	MiR-150-5p could reduce joint destruction by inhibiting synoviocyte hyperplasia and angiogenesis	Chen <i>et al</i> ^[80]
	Exosome-encapsulated miR-548a-3p	Macrophage-like cells	MiR-548a-3p could inhibit the proliferation and activation of pTHP-1 cells <i>via</i> the TLR4/NF-κB signaling pathway	Wang <i>et al</i> ^[81]
	Exosome-encapsulated miR-6089	Macrophage-like cells	MiR-6089 could regulate LPS/TLR4-mediated inflammatory response	Xu <i>et al</i> ^[82]
	Exosome-derived lncRNA Hotair	Blood mononuclear cells	Hotair may contribute to the dissolution of bone and cartilage matrix through activation of MMP-2 and MMP-13 in osteoclasts and RA synoviocytes. Hotair is more stable and easily detected in body fluid	Song <i>et al</i> ^[83]
	Exosomal miR-17	Blood mononuclear cells	MiR-17 can suppress regulatory T cell differentiation by inhibiting the expression of TGFBR II	Wang <i>et al</i> ^[84]
	MicroRNA-155	MiR-155-deficient mice	MiR-155-deficient mice are resistant to collagen-induced arthritis, and antigen-specific Th17 cell and autoantibody responses are suppressed markedly to reduce articular inflammation	Kurowska-Stolarska <i>et al</i> ^[85]
	MicroRNA-146	Human RA synovial fibroblasts	MiR-146a is expressed in the superficial and sublining layers of synovial tissue, like synovial fibroblasts, macrophages, T cells, and B cells	Nakasa <i>et al</i> ^[86]
SLE	Exosomal miR-26a	Female B6.MRLc1 and C57BL/6 mice; C57BL/6 (9 mo of age)	Podocytes mainly express miR-26a in mouse kidneys. Glomerular miR-26a expression in B6.MRLc1 mice correlates negatively with the urinary albumin levels and podocyte specific gene expression	Ichii <i>et al</i> ^[89]
	Exosomal miRNA-146a	Urine sample of SLE patients	Up-regulated exosomal miRNA-146a is found in the presence of active lupus nephritis	Perez-Hernandez <i>et al</i> ^[100]
pSS	EV derived LCN2	Saliva and tear samples from pSS patients and healthy controls	EV derived LCN2 is over-expressed in pSS patients	Aqrabi <i>et al</i> ^[107]
	EV derived APMAP	Saliva and tear samples from pSS patients and healthy controls	EV derived APMAP is over-expressed in pSS patients	Aqrabi <i>et al</i> ^[107]
	EV derived CPNE1	Saliva and tear samples from pSS patients and healthy controls	EV derived CPNE1 is over-expressed in pSS patients	Aqrabi <i>et al</i> ^[107]
IBD	MSC-EVs	LPS treated macrophages and an <i>in vivo</i> DSS induced mouse model	EVs promote the up-regulation of pro-inflammatory factors (TNF-α, IL-6, and IL-12) and down-regulation of the anti-inflammatory factor IL-10 in LPS-induced macrophages. EVs promote polarization of M1-like macrophages to an M2-like state	Cao <i>et al</i> ^[113]
Breast cancer	Exosomal PD-L1	MDA-MB-231 (231) human breast cancer cells and 4T1 mouse mammary tumor cells with PD-L1 expression or PD-L1KO	Exosomal PD-L1 bind to PD-1 on T cells to inhibit T cell activation and killing activities	Yang <i>et al</i> ^[120]
Lung cancer	EV derived miR-103a	Human adenocarcinoma cell lines NCI-H1437, NCI-H1792, and NCI-H2087 and human embryonic kidney HEK293 cells	miRNA inhibitor could inhibit effectively miR-103a mediated M2-type polarization, improving the cytokine prolif of tumor infiltration macrophages	Hsu <i>et al</i> ^[121]

Pancreatic cancer	Exosomal miR-301a-3p	Pancreatic cancer blood samples, Pancreatic cancer cell lines PANC-1, BxPC-3 and monocytic cell line THP-1	Pancreatic cells generate miR-301a-3p-rich exosomes in a hypoxic microenvironment, which polarize macrophages to promote malignant behaviors of cancer cells	Wang <i>et al.</i> ^[122]
GVHD	MSC-EVs	Kidney samples from acute cellular rejection	iKEA (integrated kidney exosome analysis) shows a high level of CD3-positive EVs in kidney rejection patients and achieved high detection accuracy (91.1%)	Park <i>et al.</i> ^[126]

RA: Rheumatoid arthritis; SLE: Systemic lupus erythematosus; pSS: Primary Sjgren's syndrome; IBD: Inflammatory bowel diseases; GVHD: Graft-versus-host disease; MSC: mesenchymal stem cell; EV: Extracellular vesicle; MSC-EV: Mesenchymal stem cell derived extracellular vesicle; MMP: Matrix metalloproteinase; VEGF: Vascular endothelial growth factor; TGFBR II: Transforming growth factor beta receptor II; SHIP-1: Src homology 2-containing inositol phosphatase-1; PD-1: Programmed death-1; PD-L1: PD-1 ligand; PD-L1KO: PD-L1 knockout; LCN2: Neutrophil gelatinase-associated lipocalin; APMAP: Adipocyte plasma membrane-associated protein; CPNE1: Copine.

MSC-EVs and B lymphocytes

B lymphocytes are also vital immune cells in adaptive immunity. A growing number of studies suggest that MSCs possess an immunomodulatory effect on B cells, but the molecular mechanisms involved are still mysterious^[47]. Nevertheless, there is little research on the role of MSC-EVs in mediating the regulatory effect of B cells on inflammatory and immune responses. Membrane vesicles derived from MSCs inhibit both B cell proliferation and differentiation in a dose-dependent fashion^[48]. Traggiai E and his colleagues found that MSCs positively influence the proliferation and differentiation of B cells into plasma cells secreting more immunoglobulins^[47]. Thus, MSCs promote downstream immune responses by mediating the conversion of B cells. Systemic lupus erythematosus (SLE) is a typical autoimmune disease characterized by constantly producing various antibodies to counter autologous cells. It is well established that B cells play a critical role in autoimmune responses *via* autoantibodies dependent mechanisms. Therefore, we infer that MSCs mediated cell conversion can boost the inflammatory progression. Therefore, MSCs can serve as a potential therapeutic tool in autoimmune diseases.

MSC-EVs and monocytes

Monocytes are secreted from bone marrow into the circulatory system and transported to target tissue, where they differentiate into mature macrophages^[49]. Macrophages are critical effectors and regulators of the immune system and play a central role in inflammation^[50]. It has been well documented that macrophages can be divided into two subpopulations: The classic M1 and the alternative M2 macrophages under microenvironmental factors. The classical M1 macrophages are induced by TLR ligands and IFN- γ and alternative M2 macrophages are induced by the immune complex IL-4/IL-13^[51,52]. M1 macrophages are characterized by strong microbicidal and tumoricidal activity, which can promote Th1 related inflammatory responses by releasing a range of proinflammatory cytokines, such as IL-6, IL-12, and TNF- α ^[53], whereas M2 macrophages with anti-inflammatory function produce less proinflammatory cytokines and more IL-10 and other anti-inflammatory factors^[54]. In short, both M1 and M2 macrophages contribute to the balance between destruction and repair of tissue in pathological conditions. A study suggested that after coculture

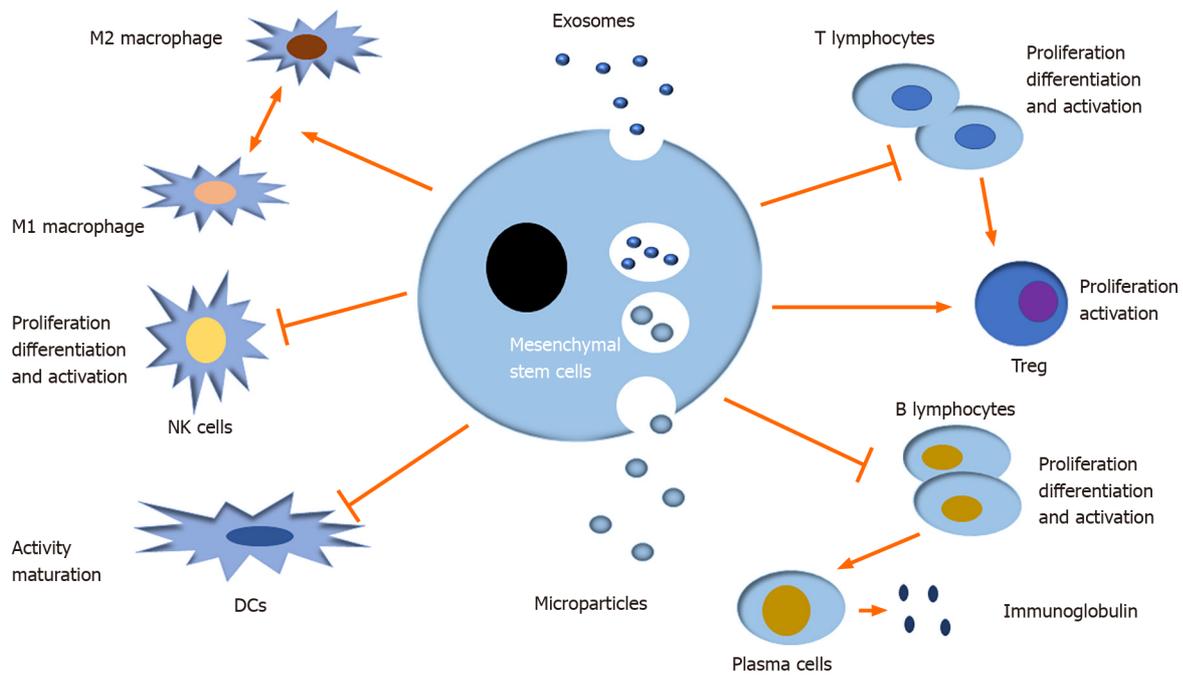


Figure 3 Immunomodulatory effects of mesenchymal stem cell derived extracellular vesicles. Mesenchymal stem cell derived extracellular vesicles (MSC-EVs) exert immunomodulatory effect on innate and adaptive immune reactions mediated by many immune cells, primarily including T lymphocytes, B lymphocytes, natural killer cells, dendritic cells, and macrophages. In brief, MSC-EVs can inhibit the proliferation, differentiation, and activation of T, B, and natural killer cells and the pathogen-presenting function of dendritic cells and macrophages. In addition, macrophage polarization can be shifted under different microenvironments in accompany with MSC-EVs.

of AMSCs with inflammatory cytokines IFN- γ and TNF- α , a higher level of exosomes can be detected in the medium supernatant, which induce M1 differentiate to anti-inflammatory M2 phenotype^[55]. Adipose tissue accumulating constantly in the body leads to obesity and inflammatory responses, which increase the risk of incidence of many chronic diseases, including type 2 diabetes, cardiovascular events, and part of cancers^[56-58]. Previous studies have revealed that the invasion of macrophages and T cells promote the formation of chronic inflammation in white adipose tissues^[59,60]. In high fat diet fed mice, AMSC derived exosomes promote white adipose tissue hypertrophy by inducing M2 macrophage polarization^[61]. A study by Németh *et al*^[62] showed that endotoxin stimulated MSCs induce M2 macrophage polarization to release IL-10 and attenuate sepsis *via* the NF- κ B signal pathway in a mouse model^[62]. MSC-EVs induce the production of M2 macrophages with anti-inflammatory properties to restrain many relevant immune responses.

MSC-EVs and NK cells

NK cells, a vital cell type in the innate immune system, mediate cytotoxic activity and produce certain cytokines and chemokines to mediate antigen presentation, antiviral responses, autoimmune responses, and the occurrence of various autoimmune diseases^[63]. A previous result showed that MSC-EVs injected into periocular tissue depress the transfer of CD161⁺ NK cells, delay the progression of disease, and restore damaged tissue in autoimmune uveitis rat models^[64,65]. Decidua parietalis MSCs release IL-2 to CD69 (NK cell receptor) to stimulate IL-2 dependent NK cells and thus promote the proliferation of activated NK cells^[66]. Thus, decidua parietalis MSCs induce directly the activity of NK cells through IL-2 and CD69. Recent research suggested that fetal liver MSC derived exosomes carrying LAP, TGF β , and TSP1 restrain the proliferation and activation of NK cells *via* TGF β /Smad2/3 signaling^[67]. Although available data show that MSC-EVs depress the activation and proliferation of NK cells, the research on that is limited in quantity and more studies need to be carried out in the future.

MSC-EVs and DCs

DCs, important bone marrow derived APCs, present multiple antigenic peptides (major histocompatibility complex - peptide complexes) to other immune cells, like T cells, and play a key role in bridging innate to adaptive immune systems. Coculture of

DCs with MSC-EVs led to down-regulated cellular surfactants and IL-10, IL-6, and IL-17 and up-regulated the number of regulatory T cells^[68], and the activity and maturation of DCs are apparently restrained. Many studies suggest that these MSC-EVs stimulate immature DCs to release TGF- β and PGE2, and regulate the immunocompetence of T cells in DC and T cell culture medium. Those small molecules mediate autoimmune responses with unclear mechanism. MSCs induce mature DCs to immature status with low immunogenicity and immunoregulatory property. The immature DCs express less immunomodulatory factors Ia, CD11c, CD80, CD86, and CD40, except for increased CD11^[39]. Overall, MSC-EVs down-regulate the immune activity of DCs and T cell dependent adaptive immune responses indirectly. Nevertheless, the research on the interaction between DCs and MSC-EVs is limited, and the exact molecular mechanisms warrant further studies.

MSC-EVs and autoimmune disease

MSC-EVs have been suggested in many kinds of diseases, which can serve as promising strategies for autoimmune disease diagnosis and treatment, such as rheumatoid arthritis (RA), SLE, primary Sjgren's syndrome (pSS), systemic sclerosis, and inflammatory bowel diseases (IBD) due to their vital role in intercellular communications. Nevertheless, the precise molecular mechanism underlying EV regulation in autoimmunity warrants in-depth investigation.

Epidemiological survey and analysis suggest that the incidence of autoimmune diseases has been increasing year by year over the past several decades^[69]. Autoimmune diseases usually influence multiple organs and systems, such as the motor system, respiratory system, digestive system, and circulatory system^[70]. They lead to a heavy burden to public health. It is well known that some autoimmune diseases are genetically susceptible^[71]. Women tend to be affected by some autoimmune diseases, and approximately 90% of patients with autoimmune disease are female^[72]. Currently, glucocorticoids and immunosuppressive drugs are still the most frequently used non-specific therapeutic agents. That traditional therapeutic strategy causes many adverse reactions, such as opportunistic infections and metabolic abnormalities, and the development of biological molecular targeted drugs to cause slower disease progression is a priority. Accumulating data reveal the biological features of MSCs in relieving immune cell-driven systemic inflammatory responses to down-regulate immune responses, such as autoimmune diseases^[73], and MSC-EVs are a significant regulator^[74]. The current knowledge of EVs in autoimmune diseases will be discussed in detail in the following text.

MSC-EVs and RA

RA is one of the most common chronic and systemic autoimmune diseases involving multiple systems, which is characterized by the destruction of synovial joints. The representative clinical manifestations are redness, swelling, and pain of distal joints, especially small joints of hands and feet^[75]. Many researchers have suggested that the occurrence of RA is caused by many complex factors, such as genetic factors and environmental factors^[76,77]. Dysregulation of immune responses occupies a necessary position in RA.

Increasing data have revealed EVs as critical regulators in the pathogenesis of RA by delivering specific functional molecules to targeted cells. Previously, the effectiveness of MSC therapies has been elucidated in cartilage repair in both animal studies^[78] and human clinical trials^[79]. Previous studies have revealed that EVs generated by MSCs play a critical role in protecting against cartilage destruction and enhancing cartilage regeneration. Particularly, exosomal noncoding RNAs (ncRNAs), including miRNAs and lncRNAs, have been implicated in regulating inflammation and immune response. MSC derived exosomal miR-150-5p down-regulated inflammatory responses and reduced joint destruction and vasculitis by targeting matrix metalloproteinase 14 (MMP14) and vascular endothelial growth factor in a collagen-induced arthritis mouse model, which is considered as a potential therapeutic biomarker for RA^[80]. We have previously demonstrated the important role of exosome-encapsulated miR-6089 and miR-548a-3p in affecting macrophage-mediated inflammatory response in RA^[81,82]. Exosome-derived lncRNA Hotair affected the migration of activated macrophages and significantly decreased the levels of MMP-2 and MMP-13, suggesting that it is a potential biomarker for RA^[83]. A study by Wang *et al.*^[84] has shown that exosomal miR-17 inhibits regulatory T cells by targeting TGFBR II in RA^[84]. Besides, exosomes-encapsulated miR-155 and miR-146a produced by DCs can serve as important regulators in immune response and inflammatory response in RA^[85-87]. It has been shown that the expression of exosomal amyloid A is positively correlated with anti-CCP antibody and CRP, suggesting a vital role of exosomal

protein in predicating the disease activity of RA patients^[88]. Taken together, exosomal ncRNAs play critical roles in regulating immune and inflammatory cells and thus participate in the occurrence and development of RA. Nevertheless, more studies are warranted to explore the molecular mechanisms of those exosomes harboring ncRNAs in the pathogenesis of RA.

MSC-EVs and SLE

SLE is a systemic autoimmune disease with various autoantibodies, which usually affects multi-organ systems due to enhanced inflammation and complex autoimmune disorders^[89,90]. It has been well established that SLE is caused by the abundant activation of T and B lymphocytes, elevated pro-inflammatory cytokines, sedimentation of immune complex substance, and finally multiple organ damage, while the kidney is the most commonly involved organ in SLE and lupus nephritis (LN) is often caused^[91]. EVs are significant regulators in mediating cell-to-cell communications involved in inflammation and immune regulations. Mounting evidence has suggested that EV delivered nucleic acids, proteins, autoantigens, cytokines, and surface receptors can serve as significant regulators in SLE^[92,93].

Microvesicles purified from SLE patients have been identified to contain higher concentrations of immunoglobulins and complements^[94,95]. Circulating exosomes from patients with SLE have been shown to induce a proinflammatory immune response, which is characterized by high levels of TNF- α , IL-1 β , IL-6, and other inflammatory mediators^[93]. The study by Asami *et al*^[96] supports that MSCs may confer immunosuppressive effects in SLE^[96]. Previously published studies have elucidated that the EVs produced from MSCs, can also contribute to immunosuppressive function in SLE^[97]. Accordingly, EVs can be used as drug carriers because they are less immunogenic. Umbilical cord derived MSCs have been used in the treatment of SLE patients, which shows good tolerance and few adverse events associated with transplantation^[98]. Therefore, MSCs and MSC-EVs can effectively control the active SLE and be used as a therapeutic strategy, particularly for the treatment of refractory SLE. Ichii and the colleagues have found that exosomal miR-26a is positively associated with urinary protein level, which suggests that exosomal miR-26a in urine of LN patients can be used as a potential biomarker for predicting podocyte injury^[99]. In addition, Perez-Hernandez *et al*^[100] have shown that urinary exosomal miRNA-146a is significantly up-regulated in active LN patients^[100]. Therefore, testing urinary exosomal miRNA can be a non-invasive method for the detection and monitoring of LN. Nevertheless, the specific molecular mechanism of EVs in regulating autoimmunity in SLE is still unclear, which warrants further investigation by more future studies.

MSC-EVs and pSS

pSS is a systemic autoimmune disease that is characterized by chronic lymphocyte infiltration in the exocrine glands, primarily the lacrimal and salivary glands^[101,102]. The primary target organs are the lacrimal and salivary glands, and dry eyes and dry mouth are often caused^[103]. EVs purified from saliva^[104,105] and tear fluid^[106,107] have been identified to be potential biomarkers for the diagnosis and treatment of pSS in previous studies. Those differentially expressed proteins isolated from EVs of saliva and tear fluid from patients with pSS can contribute to pSS by regulating TNF- α signaling and B cell survival, including neutrophil gelatinase-associated lipocalin, adipocyte plasma membrane-associated protein, and copine^[107]. The increase of platelet-derived microvesicles, soluble CD40 ligand (sCD40L), and soluble P-selectin (sCD62P) in pSS patients reflects platelet activation, which can serve as disease biomarkers^[108]. Currently, studies on MSC-EV mediated immune responses in pSS are rare. More studies are needed to elucidate the role and underlying mechanisms of EVs in pSS.

MSC-EVs and IBD

IBD is a common digestive disease characterized by chronic, relapsing gastrointestinal tract inflammatory reactions, including two main forms, Crohn's disease and ulcerative colitis^[109,110]. To the best of our knowledge, the pathogenic mechanisms and pathogenesis of IBD are complicated, and many factors contribute to the occurrence of this disease, like autoimmune disorder, genetics, and environment^[111]. Macrophages have been seen as important immune cells inducing IBD^[112]. Experimental studies showed that inflammatory responses are significantly restrained by inducing the production of M2 macrophages in the dextran sulphate sodium induced mouse model of colitis^[113]. Moreover, higher levels of immunosuppressive factors (IL-10 and TGF- β)

were observed in mice treated with MSC-EVs, promoting repair and regeneration of damaged epithelial cells^[113]. Studies have confirmed that MSC-EVs down-regulate the production of IL-1 β , NO, and IL-18 by depressing NF- κ B and iNOS-driven signaling in 2,4,6-trinitrobenzene sulfonic acid induced colitis^[114,115]. Therefore, MSC-EVs, as an important regulator, can suppress inflammatory responses and promote injured tissue repair. That delineates the potential of MSC-EVs as biomarkers for IBD treatment.

MSC-EVs and tumor immunity

Tumor immunity is critical in the processes of immune response, immune escape, and immune surveillance in cancer^[116,117]. Previous research findings show that EVs play a critical role in anti-tumor immune reaction and inflammatory response during carcinogenesis and cancer progression^[118]. In the last decade, exosomes have attracted more and more attention in cancer immunity, particularly as tumor suppressors^[119]. Some bioactive factors encapsulated in EVs promote immune and inflammatory responses and thus lead to tumorigenesis, while some exert immune suppressive effects by inducing Tregs and M1 polarization.

A previous study has demonstrated the specific binding capacity of exosomal PD-L1 to its receptor PD-1 to depress the anti-tumor effect of T cells in breast cancer^[120]. MSCs also express and release PD-L1 to regulate T cell activity, and thus both MSCs and exosomes possess immunosuppressive effect^[45]. Besides, it has been documented that EVs play a critical in anti-tumor immune response by regulating macrophages polarization. It has been found that EVs-delivering miR-103a contributes to lung cancer by targeting PTEN and inducing M2 polarization^[121]. Similarly, exosomal miR-301a-3p purified from pancreatic cancer cells was found to induce M2 macrophage polarization *via* the PTEN/PI3K γ signaling pathway^[122]. Taken together, EVs, particularly MSC-EVs, exert immunomodulatory effects on cancer and mediate intercellular communications between cancer cells and immune cells through EVs harboring bioactive molecules, including proteins and ncRNAs.

MSC-EVs and transplantation immunity

Kidney transplantation is the current preferred treatment for end stage renal disease. However, the long-term survival rate of the transplanted kidney is still low because the transplanted recipients often suffer from acute or chronic rejection for a long period of time^[123], which finally leads to graft-versus-host disease. Biopsy is still the gold standard for the diagnosis of rejection of kidney transplantation^[124,125], but it is risky and traumatic. EVs in urine can be a potential biomarker for monitoring kidney transplant rejection^[126]. T cells infiltrate the renal tubule during acute inflammatory response, which is a major cause for transplanted renal damage. MSC-EVs possess potential of inhibition of T cell activity and proliferation and thus EVs tend to gather in damaged renal tissues and are more likely to enter the urine. Consequently, using urine for detecting rejection of kidney transplantation is more likely to operate and promising. In addition, a previous report has showed that MSC derived exosomes provide a novel and effective clinical treatment for graft-versus-host disease^[127]. Nonetheless, the role of MSC-EVs in transplantation immunity needs to be further investigated in the future.

Conclusions and prospects

MSC-EVs are a hot topic in current molecular biology. Accumulated data have implicated their immunomodulatory effects on many immune cells, including T cells, B cells, macrophages, NK cells, and DCs. Increasing studies have confirmed that MSC-EVs can serve as regulators in the pathogenesis of autoimmune related diseases. In particular, MSC-EVs and the encapsulated bioactive molecules are potential targets for the diagnosis and treatment of autoimmune disease, cancer, and other diseases. MSC-EVs can serve as new medicines in the suppression of inflammatory responses. Increasing experimental results show that application of MSC-EVs can effectively inhibit immune reactions and promote the survival and regeneration of injured cells. However, there is still a long way for investigating the therapeutic strategy for autoimmunity related diseases based on MSC-EVs. More in-depth research is warranted in the future, particularly regarding the molecular mechanism of MSC-EVs in autoimmunity.

ARTICLE HIGHLIGHTS

Research background

Mesenchymal stem cells (MSCs) have been reported to possess immune regulatory effects in innate and adaptive immune reactions. MSCs can mediate intercellular communications by releasing extracellular vesicles (EVs), which deliver functional molecules to targeted cells. MSC derived EVs (MSC-EVs) confer altering effects on many immune cells, including T lymphocytes, B lymphocytes, natural killer cells, dendritic cells, and macrophages. A large number of studies have suggested that MSC-EVs participate in regulating autoimmunity related diseases. This characteristic of MSC-EVs makes them be potential biomarkers for the diagnosis and treatment of autoimmunity related diseases.

Research motivation

This article describes and focuses on the identification, characteristics, immunomodulatory function, and underlying mechanism of MSC-EVs in autoimmunity related diseases. Understanding the immunomodulation effects of MSC-EVs better will help us to investigate the pathogenesis of diseases and develop novel targeted medicines.

Research objectives

The immune modulation of MSC-EVs play a key role in disease initiation, maintenance, and progression. This article provides a new direction for us to understand the precise mechanisms of action of autoimmunity related diseases, which will promote the improvement of therapeutic regimen.

Research methods

Literature search was conducted in PubMed to retrieve articles published between 2010 and 2020 in the English language. The keywords, such as “MSCs,” “EVs,” “autoimmune responses,” “immune cells,” and “autoimmunity related diseases,” and Boolean operator “AND” and “NOT” coalesced admirably to be used for searching *in vitro* studies on the specific molecular mechanisms of MSC-EVs in many immune cell types and many autoimmunity related diseases. Studies that did not investigate the molecular mechanisms of MSC-EVs in the occurrence and development of autoimmune diseases were excluded.

Research results

A large number of articles were retrieved and their abstracts were skimmed. When analyzing the publications, we found that it has been well documented that MSC-EVs have the ability to induce multiple immune cells, like T lymphocytes, B lymphocytes, natural killer cells, dendritic cells, and macrophages, to regulate immune responses in innate immunity and adaptive immunity. Many validated EVs-delivered molecules have been identified as key biomarkers, such as proteins, lipids, and nucleotides. Some EVs-encapsulated functional molecules can serve as promising therapeutic targets particularly for autoimmune disease.

Research conclusions

MSC-EVs play an important part in the differentiation, activation, and proliferation of immune cells, and they may become potential biomarkers for the diagnosis and treatment of autoimmunity related diseases.

Research perspectives

MSC-EVs can serve as regulators in the pathogenesis of autoimmune related diseases. In particular, MSC-EVs and the encapsulated bioactive molecules are potential targets for the diagnosis and treatment of autoimmune disease, cancer, and other diseases. However, there is still a long way for investigating the therapeutic strategy for autoimmunity related diseases based on MSC-EVs. More in-depth research is warranted in the future, particularly regarding the molecular mechanism of MSC-EVs in autoimmunity

REFERENCES

- 1 **da Silva Meirelles L**, Chagastelles PC, Nardi NB. Mesenchymal stem cells reside in virtually all post-natal organs and tissues. *J Cell Sci* 2006; **119**: 2204-2213 [PMID: 16684817 DOI: 10.1242/jcs.02932]
- 2 **Pittenger MF**, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, Moorman MA, Simonetti DW, Craig S, Marshak DR. Multilineage potential of adult human mesenchymal stem cells. *Science* 1999; **284**: 143-147 [PMID: 10102814 DOI: 10.1126/science.284.5411.143]
- 3 **De Bari C**, Dell'Accio F, Tylzanowski P, Luyten FP. Multipotent mesenchymal stem cells from adult human synovial membrane. *Arthritis Rheum* 2001; **44**: 1928-1942 [PMID: 11508446 DOI: 10.1002/1529-0131(200108)44:8<1928::AID-ART331>3.0.CO;2-P]
- 4 **De Bari C**, Dell'Accio F, Vandenabeele F, Vermeesch JR, Raymackers JM, Luyten FP. Skeletal muscle repair by adult human mesenchymal stem cells from synovial membrane. *J Cell Biol* 2003; **160**: 909-918 [PMID: 12629053 DOI: 10.1083/jcb.200212064]
- 5 **Ding DC**, Chang YH, Shyu WC, Lin SZ. Human umbilical cord mesenchymal stem cells: a new era for stem cell therapy. *Cell Transplant* 2015; **24**: 339-347 [PMID: 25622293 DOI: 10.3727/096368915X686841]
- 6 **Zuk PA**, Zhu M, Ashjian P, De Ugarte DA, Huang JI, Mizuno H, Alfonso ZC, Fraser JK, Benhaim P, Hedrick MH. Human adipose tissue is a source of multipotent stem cells. *Mol Biol Cell* 2002; **13**: 4279-4295 [PMID: 12475952 DOI: 10.1091/mbc.e02-02-0105]
- 7 **Cosenza S**, Toupet K, Maumus M, Luz-Crawford P, Blanc-Brude O, Jorgensen C, Noël D. Mesenchymal stem cells-derived exosomes are more immunosuppressive than microparticles in inflammatory arthritis. *Theranostics* 2018; **8**: 1399-1410 [PMID: 29507629 DOI: 10.7150/thno.21072]
- 8 **Liang X**, Ding Y, Zhang Y, Tse HF, Lian Q. Paracrine mechanisms of mesenchymal stem cell-based therapy: current status and perspectives. *Cell Transplant* 2014; **23**: 1045-1059 [PMID: 23676629 DOI: 10.3727/096368913X667709]
- 9 **Prockop DJ**, Oh JY. Mesenchymal stem/stromal cells (MSCs): role as guardians of inflammation. *Mol Ther* 2012; **20**: 14-20 [PMID: 22008910 DOI: 10.1038/mt.2011.211]
- 10 **Galipeau J**, Sensébé L. Mesenchymal Stromal Cells: Clinical Challenges and Therapeutic Opportunities. *Cell Stem Cell* 2018; **22**: 824-833 [PMID: 29859173 DOI: 10.1016/j.stem.2018.05.004]
- 11 **Squillaro T**, Peluso G, Galderisi U. Clinical Trials With Mesenchymal Stem Cells: An Update. *Cell Transplant* 2016; **25**: 829-848 [PMID: 26423725 DOI: 10.3727/096368915X689622]
- 12 **Trento C**, Bernardo ME, Nagler A, Kuçi S, Bornhäuser M, Köhl U, Strunk D, Galleu A, Sanchez-Guijo F, Gaipa G, Introna M, Bukauskas A, Le Blanc K, Apperley J, Roelofs H, Van Campenhout A, Beguin Y, Kuball J, Lazzari L, Avanzini MA, Fibbe W, Chabannon C, Bonini C, Dazzi F. Manufacturing Mesenchymal Stromal Cells for the Treatment of Graft-versus-Host Disease: A Survey among Centers Affiliated with the European Society for Blood and Marrow Transplantation. *Biol Blood Marrow Transplant* 2018; **24**: 2365-2370 [PMID: 30031938 DOI: 10.1016/j.bbmt.2018.07.015]
- 13 **Wolf P**. The nature and significance of platelet products in human plasma. *Br J Haematol* 1967; **13**: 269-288 [PMID: 6025241 DOI: 10.1111/j.1365-2141.1967.tb08741.x]
- 14 **Ratajczak J**, Miekus K, Kucia M, Zhang J, Reca R, Dvorak P, Ratajczak MZ. Embryonic stem cell-derived microvesicles reprogram hematopoietic progenitors: evidence for horizontal transfer of mRNA and protein delivery. *Leukemia* 2006; **20**: 847-856 [PMID: 16453000 DOI: 10.1038/sj.leu.2404132]
- 15 **Camussi G**, Deregibus MC, Bruno S, Grange C, Fonsato V, Tetta C. Exosome/microvesicle-mediated epigenetic reprogramming of cells. *Am J Cancer Res* 2011; **1**: 98-110 [PMID: 21969178]
- 16 **Lai RC**, Chen TS, Lim SK. Mesenchymal stem cell exosome: a novel stem cell-based therapy for cardiovascular disease. *Regen Med* 2011; **6**: 481-492 [PMID: 21749206 DOI: 10.2217/rme.11.35]
- 17 **Timmers L**, Lim SK, Hoefler IE, Arslan F, Lai RC, van Oorschot AA, Goumans MJ, Strijder C, Sze SK, Choo A, Piek JJ, Doevendans PA, Pasterkamp G, de Kleijn DP. Human mesenchymal stem cell-conditioned medium improves cardiac function following myocardial infarction. *Stem Cell Res* 2011; **6**: 206-214 [PMID: 21419744 DOI: 10.1016/j.scr.2011.01.001]
- 18 **Timmers L**, Lim SK, Arslan F, Armstrong JS, Hoefler IE, Doevendans PA, Piek JJ, El Oakley RM, Choo A, Lee CN, Pasterkamp G, de Kleijn DP. Reduction of myocardial infarct size by human mesenchymal stem cell conditioned medium. *Stem Cell Res* 2007; **1**: 129-137 [PMID: 19383393 DOI: 10.1016/j.scr.2008.02.002]
- 19 **Chavez-Muñoz C**, Morse J, Kilani R, Ghahary A. Primary human keratinocytes externalize stratifin protein via exosomes. *J Cell Biochem* 2008; **104**: 2165-2173 [PMID: 18452139 DOI: 10.1002/jcb.21774]
- 20 **Fauré J**, Lachenal G, Court M, Hirrlinger J, Chatellard-Causse C, Blot B, Grange J, Schoehn G, Goldberg Y, Boyer V, Kirchhoff F, Raposo G, Garin J, Sadoul R. Exosomes are released by cultured cortical neurones. *Mol Cell Neurosci* 2006; **31**: 642-648 [PMID: 16446100 DOI: 10.1016/j.mcn.2005.12.003]
- 21 **Guescini M**, Genedani S, Stocchi V, Agnati LF. Astrocytes and Glioblastoma cells release exosomes carrying mtDNA. *J Neural Transm (Vienna)* 2010; **117**: 1-4 [PMID: 19680595 DOI: 10.1007/s00702-009-0288-8]
- 22 **Potolicchio I**, Carven GJ, Xu X, Stipp C, Riese RJ, Stern LJ, Santambrogio L. Proteomic analysis of microglia-derived exosomes: metabolic role of the aminopeptidase CD13 in neuropeptide catabolism. *J Immunol* 2005; **175**: 2237-2243 [PMID: 16081791 DOI: 10.4049/jimmunol.175.4.2237]
- 23 **Al-Nedawi K**, Meehan B, Micallef J, Lhotak V, May L, Guha A, Rak J. Intercellular transfer of the oncogenic receptor EGFRvIII by microvesicles derived from tumour cells. *Nat Cell Biol* 2008; **10**: 619-624 [PMID: 18425114 DOI: 10.1038/ncb1725]
- 24 **Skog J**, Würdinger T, van Rijn S, Meijer DH, Gainche L, Sena-Esteves M, Curry WT Jr, Carter BS, Krichevsky AM, Breakefield XO. Glioblastoma microvesicles transport RNA and proteins that promote tumour growth and provide diagnostic biomarkers. *Nat Cell Biol* 2008; **10**: 1470-1476 [PMID: 19011622 DOI: 10.1038/ncb1800]
- 25 **Al-Nedawi K**, Meehan B, Kerbel RS, Allison AC, Rak J. Endothelial expression of autocrine VEGF upon the uptake of tumor-derived microvesicles containing oncogenic EGFR. *Proc Natl Acad Sci USA* 2009; **106**:

- 3794-3799 [PMID: [19234131](#) DOI: [10.1073/pnas.0804543106](#)]
- 26 **Raposo G**, Stoorvogel W. Extracellular vesicles: exosomes, microvesicles, and friends. *J Cell Biol* 2013; **200**: 373-383 [PMID: [23420871](#) DOI: [10.1083/jcb.201211138](#)]
- 27 **Dignat-George F**, Boulanger CM. The many faces of endothelial microparticles. *Arterioscler Thromb Vasc Biol* 2011; **31**: 27-33 [PMID: [21160065](#) DOI: [10.1161/ATVBAHA.110.218123](#)]
- 28 **Choi DS**, Kim DK, Kim YK, Gho YS. Proteomics of extracellular vesicles: Exosomes and ectosomes. *Mass Spectrom Rev* 2015; **34**: 474-490 [PMID: [24421117](#) DOI: [10.1002/mas.21420](#)]
- 29 **Pan BT**, Teng K, Wu C, Adam M, Johnstone RM. Electron microscopic evidence for externalization of the transferrin receptor in vesicular form in sheep reticulocytes. *J Cell Biol* 1985; **101**: 942-948 [PMID: [2993317](#) DOI: [10.1083/jcb.101.3.942](#)]
- 30 **Yáñez-Mó M**, Siljander PR, Andreu Z, Zavec AB, Borràs FE, Buzas EI, Buzas K, Casal E, Cappello F, Carvalho J, Colás E, Cordeiro-da Silva A, Fais S, Falcon-Perez JM, Ghoobrial IM, Giebel B, Gimona M, Graner M, Gursel I, Gursel M, Heegaard NH, Hendrix A, Kierulf P, Kokubun K, Kosanovic M, Kralj-Iglic V, Krämer-Albers EM, Laitinen S, Lässer C, Lener T, Ligeti E, Liné A, Lipps G, Lorente A, Lötvall J, Manček-Keber M, Marcilla A, Mittelbrunn M, Nazarenko I, Nolte-t Hoen EN, Nyman TA, O'Driscoll L, Olivan M, Oliveira C, Pállinger É, Del Portillo HA, Reventós J, Rigau M, Rohde E, Sammar M, Sánchez-Madrid F, Santarém N, Schallmoser K, Ostenfeld MS, Stoorvogel W, Stukelj R, Van der Grein SG, Vasconcelos MH, Wauben MH, De Wever O. Biological properties of extracellular vesicles and their physiological functions. *J Extracell Vesicles* 2015; **4**: 27066 [PMID: [25979354](#) DOI: [10.3402/jev.v4.27066](#)]
- 31 **Robbins PD**, Morelli AE. Regulation of immune responses by extracellular vesicles. *Nat Rev Immunol* 2014; **14**: 195-208 [PMID: [24566916](#) DOI: [10.1038/nri3622](#)]
- 32 **Distler JH**, Huber LC, Gay S, Distler O, Pisetsky DS. Microparticles as mediators of cellular cross-talk in inflammatory disease. *Autoimmunity* 2006; **39**: 683-690 [PMID: [17178565](#) DOI: [10.1080/08916930601061538](#)]
- 33 **Turpin D**, Truchetet ME, Faustin B, Augusto JF, Contin-Bordes C, Brisson A, Blanco P, Duffau P. Role of extracellular vesicles in autoimmune diseases. *Autoimmun Rev* 2016; **15**: 174-183 [PMID: [26554931](#) DOI: [10.1016/j.autrev.2015.11.004](#)]
- 34 **Toñño-Vian M**, Guillén MI, Alcaraz MJ. Extracellular vesicles: A new therapeutic strategy for joint conditions. *Biochem Pharmacol* 2018; **153**: 134-146 [PMID: [29427625](#) DOI: [10.1016/j.bcp.2018.02.004](#)]
- 35 **Maumus M**, Jorgensen C, Noël D. Mesenchymal stem cells in regenerative medicine applied to rheumatic diseases: role of secretome and exosomes. *Biochimie* 2013; **95**: 2229-2234 [PMID: [23685070](#) DOI: [10.1016/j.biochi.2013.04.017](#)]
- 36 **Ansboro S**, Roelofs AJ, De Bari C. Mesenchymal stem cells for the management of rheumatoid arthritis: immune modulation, repair or both? *Curr Opin Rheumatol* 2017; **29**: 201-207 [PMID: [27941390](#) DOI: [10.1097/BOR.0000000000000370](#)]
- 37 **Cosenza S**, Ruiz M, Maumus M, Jorgensen C, Noël D. Pathogenic or Therapeutic Extracellular Vesicles in Rheumatic Diseases: Role of Mesenchymal Stem Cell-Derived Vesicles. *Int J Mol Sci* 2017; **18** [PMID: [28441721](#) DOI: [10.3390/ijms18040889](#)]
- 38 **Asari S**, Itakura S, Ferreri K, Liu CP, Kuroda Y, Kandeel F, Mullen Y. Mesenchymal stem cells suppress B-cell terminal differentiation. *Exp Hematol* 2009; **37**: 604-615 [PMID: [19375651](#) DOI: [10.1016/j.exphem.2009.01.005](#)]
- 39 **Zhang B**, Liu R, Shi D, Liu X, Chen Y, Dou X, Zhu X, Lu C, Liang W, Liao L, Zenke M, Zhao RC. Mesenchymal stem cells induce mature dendritic cells into a novel Jagged-2-dependent regulatory dendritic cell population. *Blood* 2009; **113**: 46-57 [PMID: [18832657](#) DOI: [10.1182/blood-2008-04-154138](#)]
- 40 **Prigione I**, Benvenuto F, Bocca P, Battistini L, Uccelli A, Pistoia V. Reciprocal interactions between human mesenchymal stem cells and gammadelta T cells or invariant natural killer T cells. *Stem Cells* 2009; **27**: 693-702 [PMID: [19096038](#) DOI: [10.1634/stemcells.2008-0687](#)]
- 41 **Ren G**, Zhang L, Zhao X, Xu G, Zhang Y, Roberts AI, Zhao RC, Shi Y. Mesenchymal stem cell-mediated immunosuppression occurs via concerted action of chemokines and nitric oxide. *Cell Stem Cell* 2008; **2**: 141-150 [PMID: [18371435](#) DOI: [10.1016/j.stem.2007.11.014](#)]
- 42 **Nojehdehi S**, Soudi S, Hesampour A, Rasouli S, Soleimani M, Hashemi SM. Immunomodulatory effects of mesenchymal stem cell-derived exosomes on experimental type-1 autoimmune diabetes. *J Cell Biochem* 2018; **119**: 9433-9443 [PMID: [30074271](#) DOI: [10.1002/jcb.27260](#)]
- 43 **Melief SM**, Schrama E, Brugman MH, Tiemessen MM, Hoogduijn MJ, Fibbe WE, Roelofs H. Multipotent stromal cells induce human regulatory T cells through a novel pathway involving skewing of monocytes toward anti-inflammatory macrophages. *Stem Cells* 2013; **31**: 1980-1991 [PMID: [23712682](#) DOI: [10.1002/stem.1432](#)]
- 44 **Mougiakakos D**, Jitschin R, Johansson CC, Okita R, Kiessling R, Le Blanc K. The impact of inflammatory licensing on heme oxygenase-1-mediated induction of regulatory T cells by human mesenchymal stem cells. *Blood* 2011; **117**: 4826-4835 [PMID: [21389316](#) DOI: [10.1182/blood-2010-12-324038](#)]
- 45 **Davies LC**, Heldring N, Kadri N, Le Blanc K. Mesenchymal Stromal Cell Secretion of Programmed Death-1 Ligands Regulates T Cell Mediated Immunosuppression. *Stem Cells* 2017; **35**: 766-776 [PMID: [27671847](#) DOI: [10.1002/stem.2509](#)]
- 46 **Serejo TRT**, Silva-Carvalho AÉ, Braga LDCF, Neves FAR, Pereira RW, Carvalho JL, Saldanha-Araujo F. Assessment of the Immunosuppressive Potential of INF- γ Licensed Adipose Mesenchymal Stem Cells, Their Secretome and Extracellular Vesicles. *Cells* 2019; **8** [PMID: [30621275](#) DOI: [10.3390/cells8010022](#)]
- 47 **Traggiai E**, Volpi S, Schena F, Gattorno M, Ferlito F, Moretta L, Martini A. Bone marrow-derived mesenchymal stem cells induce both polyclonal expansion and differentiation of B cells isolated from healthy donors and systemic lupus erythematosus patients. *Stem Cells* 2008; **26**: 562-569 [PMID: [18024418](#) DOI: [10.1634/stemcells.2007-0528](#)]
- 48 **Budoni M**, Fierabracci A, Luciano R, Petrini S, Di Ciommo V, Muraca M. The immunosuppressive effect of mesenchymal stromal cells on B lymphocytes is mediated by membrane vesicles. *Cell Transplant* 2013; **22**: 369-379 [PMID: [23433427](#) DOI: [10.3727/096368911X582769](#)]
- 49 **Bolego C**, Cignarella A, Staels B, Chinetti-Gbaguidi G. Macrophage function and polarization in cardiovascular disease: a role of estrogen signaling? *Arterioscler Thromb Vasc Biol* 2013; **33**: 1127-1134

- [PMID: 23640494 DOI: 10.1161/ATVBAHA.113.301328]
- 50 **Gordon S**, Martinez FO. Alternative activation of macrophages: mechanism and functions. *Immunity* 2010; **32**: 593-604 [PMID: 20510870 DOI: 10.1016/j.immuni.2010.05.007]
- 51 **Zhou D**, Huang C, Lin Z, Zhan S, Kong L, Fang C, Li J. Macrophage polarization and function with emphasis on the evolving roles of coordinated regulation of cellular signaling pathways. *Cell Signal* 2014; **26**: 192-197 [PMID: 24219909 DOI: 10.1016/j.cellsig.2013.11.004]
- 52 **Sica A**, Mantovani A. Macrophage plasticity and polarization: in vivo veritas. *J Clin Invest* 2012; **122**: 787-795 [PMID: 22378047 DOI: 10.1172/JCI59643]
- 53 **Mantovani A**, Sica A, Sozzani S, Allavena P, Vecchi A, Locati M. The chemokine system in diverse forms of macrophage activation and polarization. *Trends Immunol* 2004; **25**: 677-686 [PMID: 15530839 DOI: 10.1016/j.it.2004.09.015]
- 54 **Martinez FO**. Regulators of macrophage activation. *Eur J Immunol* 2011; **41**: 1531-1534 [PMID: 21607943 DOI: 10.1002/eji.201141670]
- 55 **Domenis R**, Cifù A, Quaglia S, Pistis C, Moretti M, Vicario A, Parodi PC, Fabris M, Niazzi KR, Soon-Shiong P, Curcio F. Pro inflammatory stimuli enhance the immunosuppressive functions of adipose mesenchymal stem cells-derived exosomes. *Sci Rep* 2018; **8**: 13325 [PMID: 30190615 DOI: 10.1038/s41598-018-31707-9]
- 56 **Hossain P**, Kavar B, El Nahas M. Obesity and diabetes in the developing world—a growing challenge. *N Engl J Med* 2007; **356**: 213-215 [PMID: 17229948 DOI: 10.1056/NEJMp068177]
- 57 **Hotamisligil GS**. Inflammation and metabolic disorders. *Nature* 2006; **444**: 860-867 [PMID: 17167474 DOI: 10.1038/nature05485]
- 58 **Xu H**, Barnes GT, Yang Q, Tan G, Yang D, Chou CJ, Sole J, Nichols A, Ross JS, Tartaglia LA, Chen H. Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. *J Clin Invest* 2003; **112**: 1821-1830 [PMID: 14679177 DOI: 10.1172/JCI19451]
- 59 **Weisberg SP**, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AW Jr. Obesity is associated with macrophage accumulation in adipose tissue. *J Clin Invest* 2003; **112**: 1796-1808 [PMID: 14679176 DOI: 10.1172/JCI19246]
- 60 **Wu H**, Ghosh S, Perrard XD, Feng L, Garcia GE, Perrard JL, Sweeney JF, Peterson LE, Chan L, Smith CW, Ballantyne CM. T-cell accumulation and regulated on activation, normal T cell expressed and secreted upregulation in adipose tissue in obesity. *Circulation* 2007; **115**: 1029-1038 [PMID: 17296858 DOI: 10.1161/CIRCULATIONAHA.106.638379]
- 61 **Zhao H**, Shang Q, Pan Z, Bai Y, Li Z, Zhang H, Zhang Q, Guo C, Zhang L, Wang Q. Exosomes From Adipose-Derived Stem Cells Attenuate Adipose Inflammation and Obesity Through Polarizing M2 Macrophages and Beiging in White Adipose Tissue. *Diabetes* 2018; **67**: 235-247 [PMID: 29133512 DOI: 10.2337/db17-0356]
- 62 **Németh K**, Leelahavanichkul A, Yuen PS, Mayer B, Parmelee A, Doi K, Robey PG, Leelahavanichkul K, Koller BH, Brown JM, Hu X, Jelinek I, Star RA, Mezey E. Bone marrow stromal cells attenuate sepsis via prostaglandin E(2)-dependent reprogramming of host macrophages to increase their interleukin-10 production. *Nat Med* 2009; **15**: 42-49 [PMID: 19098906 DOI: 10.1038/nm.1905]
- 63 **Biron CA**, Nguyen KB, Pien GC, Cousens LP, Salazar-Mather TP. Natural killer cells in antiviral defense: function and regulation by innate cytokines. *Annu Rev Immunol* 1999; **17**: 189-220 [PMID: 10358757 DOI: 10.1146/annurev.immunol.17.1.189]
- 64 **Bai L**, Shao H, Wang H, Zhang Z, Su C, Dong L, Yu B, Chen X, Li X, Zhang X. Effects of Mesenchymal Stem Cell-Derived Exosomes on Experimental Autoimmune Uveitis. *Sci Rep* 2017; **7**: 4323 [PMID: 28659587 DOI: 10.1038/s41598-017-04559-y]
- 65 **Shigemoto-Kuroda T**, Oh JY, Kim DK, Jeong HJ, Park SY, Lee HJ, Park JW, Kim TW, An SY, Prockop DJ, Lee RH. MSC-derived Extracellular Vesicles Attenuate Immune Responses in Two Autoimmune Murine Models: Type 1 Diabetes and Uveoretinitis. *Stem Cell Reports* 2017; **8**: 1214-1225 [PMID: 28494937 DOI: 10.1016/j.stemcr.2017.04.008]
- 66 **Abumaree MH**, Bahattab E, Alsadoun A, Al Dosaimani A, Abomaray FM, Khatlani T, Kalionis B, El-Muzaini MF, Alawad AO, AlAskar AS. Characterization of the interaction between human decidua parietalis mesenchymal stem/stromal cells and natural killer cells. *Stem Cell Res Ther* 2018; **9**: 102 [PMID: 29650045 DOI: 10.1186/s13287-018-0844-y]
- 67 **Fan Y**, Herr F, Vernochet A, Mennesson B, Oberlin E, Durrbach A. Human Fetal Liver Mesenchymal Stem Cell-Derived Exosomes Impair Natural Killer Cell Function. *Stem Cells Dev* 2019; **28**: 44-55 [PMID: 30328799 DOI: 10.1089/scd.2018.0015]
- 68 **Favaro E**, Carpanetto A, Caorsi C, Giovarelli M, Angelini C, Cavallo-Perin P, Tetta C, Camussi G, Zanone MM. Human mesenchymal stem cells and derived extracellular vesicles induce regulatory dendritic cells in type 1 diabetic patients. *Diabetologia* 2016; **59**: 325-333 [PMID: 26592240 DOI: 10.1007/s00125-015-3808-0]
- 69 **Ji J**, Sundquist J, Sundquist K. Gender-specific incidence of autoimmune diseases from national registers. *J Autoimmun* 2016; **69**: 102-106 [PMID: 26994904 DOI: 10.1016/j.jaut.2016.03.003]
- 70 **Cooper GS**, Bynum ML, Somers EC. Recent insights in the epidemiology of autoimmune diseases: improved prevalence estimates and understanding of clustering of diseases. *J Autoimmun* 2009; **33**: 197-207 [PMID: 19819109 DOI: 10.1016/j.jaut.2009.09.008]
- 71 **Gershwin LJ**. Current and Newly Emerging Autoimmune Diseases. *Vet Clin North Am Small Anim Pract* 2018; **48**: 323-338 [PMID: 29248206 DOI: 10.1016/j.cvsm.2017.10.010]
- 72 **Autoimmune disease and the environment**. *Environ Health Perspect* 1998; **106**: A592-A593 [PMID: 10048952 DOI: 10.1289/ehp.106-1533238]
- 73 **Wang Y**, Chen X, Cao W, Shi Y. Plasticity of mesenchymal stem cells in immunomodulation: pathological and therapeutic implications. *Nat Immunol* 2014; **15**: 1009-1016 [PMID: 25329189 DOI: 10.1038/ni.3002]
- 74 **Xu H**, Jia S, Xu H. Potential therapeutic applications of exosomes in different autoimmune diseases. *Clin Immunol* 2019; **205**: 116-124 [PMID: 31228581 DOI: 10.1016/j.clim.2019.06.006]
- 75 **Lee DM**, Weinblatt ME. Rheumatoid arthritis. *Lancet* 2001; **358**: 903-911 [PMID: 11567728 DOI: 10.1016/S0140-6736(01)06075-5]

- 76 **MacGregor AJ**, Snieder H, Rigby AS, Koskenvuo M, Kaprio J, Aho K, Silman AJ. Characterizing the quantitative genetic contribution to rheumatoid arthritis using data from twins. *Arthritis Rheum* 2000; **43**: 30-37 [PMID: 10643697 DOI: 10.1002/1529-0131(200001)43:1<30::AID-ANR5>3.0.CO;2-B]
- 77 **Padyukov L**, Silva C, Stolt P, Alfredsson L, Klareskog L. A gene-environment interaction between smoking and shared epitope genes in HLA-DR provides a high risk of seropositive rheumatoid arthritis. *Arthritis Rheum* 2004; **50**: 3085-3092 [PMID: 15476204 DOI: 10.1002/art.20553]
- 78 **Guo W**, Zheng X, Zhang W, Chen M, Wang Z, Hao C, Huang J, Yuan Z, Zhang Y, Wang M, Peng J, Wang A, Wang Y, Sui X, Xu W, Liu S, Lu S, Guo Q. Mesenchymal Stem Cells in Oriented PLGA/ACECM Composite Scaffolds Enhance Structure-Specific Regeneration of Hyaline Cartilage in a Rabbit Model. *Stem Cells Int* 2018; **2018**: 6542198 [PMID: 29666653 DOI: 10.1155/2018/6542198]
- 79 **Kamei N**, Ochi M, Adachi N, Ishikawa M, Yanada S, Levin LS, Kamei G, Kobayashi T. The safety and efficacy of magnetic targeting using autologous mesenchymal stem cells for cartilage repair. *Knee Surg Sports Traumatol Arthrosc* 2018; **26**: 3626-3635 [PMID: 29549388 DOI: 10.1007/s00167-018-4898-2]
- 80 **Chen Z**, Wang H, Xia Y, Yan F, Lu Y. Therapeutic Potential of Mesenchymal Cell-Derived miRNA-150-5p-Expressing Exosomes in Rheumatoid Arthritis Mediated by the Modulation of MMP14 and VEGF. *J Immunol* 2018; **201**: 2472-2482 [PMID: 30224512 DOI: 10.4049/jimmunol.1800304]
- 81 **Wang Y**, Zheng F, Gao G, Yan S, Zhang L, Wang L, Cai X, Wang X, Xu D, Wang J. MiR-548a-3p regulates inflammatory response via TLR4/NF- κ B signaling pathway in rheumatoid arthritis. *J Cell Biochem* 2018 [PMID: 29315763 DOI: 10.1002/jcb.26659]
- 82 **Xu D**, Song M, Chai C, Wang J, Jin C, Wang X, Cheng M, Yan S. Exosome-encapsulated miR-6089 regulates inflammatory response via targeting TLR4. *J Cell Physiol* 2019; **234**: 1502-1511 [PMID: 30132861 DOI: 10.1002/jcp.27014]
- 83 **Song J**, Kim D, Han J, Kim Y, Lee M, Jin EJ. PBMC and exosome-derived Hotair is a critical regulator and potent marker for rheumatoid arthritis. *Clin Exp Med* 2015; **15**: 121-126 [PMID: 24722995 DOI: 10.1007/s10238-013-0271-4]
- 84 **Wang L**, Wang C, Jia X, Yu J. Circulating Exosomal miR-17 Inhibits the Induction of Regulatory T Cells via Suppressing TGFBR II Expression in Rheumatoid Arthritis. *Cell Physiol Biochem* 2018; **50**: 1754-1763 [PMID: 30384383 DOI: 10.1159/000494793]
- 85 **Kurowska-Stolarska M**, Alivernini S, Ballantine LE, Asquith DL, Millar NL, Gilchrist DS, Reilly J, Ierna M, Fraser AR, Stolarski B, McSharry C, Hueber AJ, Baxter D, Hunter J, Gay S, Liew FY, McInnes IB. MicroRNA-155 as a proinflammatory regulator in clinical and experimental arthritis. *Proc Natl Acad Sci* 2011; **108**: 11193-11198 [PMID: 21690378 DOI: 10.1073/pnas.1019536108]
- 86 **Nakasa T**, Miyaki S, Okubo A, Hashimoto M, Nishida K, Ochi M, Asahara H. Expression of microRNA-146 in rheumatoid arthritis synovial tissue. *Arthritis Rheum* 2008; **58**: 1284-1292 [PMID: 18438844 DOI: 10.1002/art.23429]
- 87 **Taganov KD**, Boldin MP, Chang KJ, Baltimore D. NF-kappaB-dependent induction of microRNA miR-146, an inhibitor targeted to signaling proteins of innate immune responses. *Proc Natl Acad Sci USA* 2006; **103**: 12481-12486 [PMID: 16885212 DOI: 10.1073/pnas.0605298103]
- 88 **Yoo J**, Lee SK, Lim M, Sheen D, Choi EH, Kim SA. Exosomal amyloid A and lymphatic vessel endothelial hyaluronic acid receptor-1 proteins are associated with disease activity in rheumatoid arthritis. *Arthritis Res Ther* 2017; **19**: 119 [PMID: 28569211 DOI: 10.1186/s13075-017-1334-9]
- 89 **Al-Shobaili HA**, Al Robaee AA, Alzolibani AA, Rasheed Z. Antibodies against 4-hydroxy-2-nonenal modified epitopes recognized chromatin and its oxidized forms: role of chromatin, oxidized forms of chromatin and 4-hydroxy-2-nonenal modified epitopes in the etiopathogenesis of SLE. *Dis Markers* 2012; **33**: 19-34 [PMID: 22710866 DOI: 10.3233/DMA-2012-0900]
- 90 **Colasanti T**, Maselli A, Conti F, Sanchez M, Alessandri C, Barbati C, Vacirca D, Tinari A, Chiarotti F, Giovannetti A, Franconi F, Valesini G, Malorni W, Pierdominici M, Ortona E. Autoantibodies to estrogen receptor α interfere with T lymphocyte homeostasis and are associated with disease activity in systemic lupus erythematosus. *Arthritis Rheum* 2012; **64**: 778-787 [PMID: 21968947 DOI: 10.1002/art.33400]
- 91 **Rahman A**, Isenberg DA. Systemic lupus erythematosus. *N Engl J Med* 2008; **358**: 929-939 [PMID: 18305268 DOI: 10.1056/NEJMra071297]
- 92 **Perez-Hernandez J**, Redon J, Cortes R. Extracellular Vesicles as Therapeutic Agents in Systemic Lupus Erythematosus. *Int J Mol Sci* 2017; **18** [PMID: 28350323 DOI: 10.3390/ijms18040717]
- 93 **Lee JY**, Park JK, Lee EY, Lee EB, Song YW. Circulating exosomes from patients with systemic lupus erythematosus induce a proinflammatory immune response. *Arthritis Res Ther* 2016; **18**: 264 [PMID: 27852323 DOI: 10.1186/s13075-016-1159-y]
- 94 **Nielsen CT**, Østergaard O, Stener L, Iversen LV, Truedsson L, Gullstrand B, Jacobsen S, Heegaard NH. Increased IgG on cell-derived plasma microparticles in systemic lupus erythematosus is associated with autoantibodies and complement activation. *Arthritis Rheum* 2012; **64**: 1227-1236 [PMID: 22238051 DOI: 10.1002/art.34381]
- 95 **Østergaard O**, Nielsen CT, Iversen LV, Tanassi JT, Knudsen S, Jacobsen S, Heegaard NH. Unique protein signature of circulating microparticles in systemic lupus erythematosus. *Arthritis Rheum* 2013; **65**: 2680-2690 [PMID: 23817959 DOI: 10.1002/art.38065]
- 96 **Asami T**, Ishii M, Fujii H, Namkoong H, Tasaka S, Matsushita K, Ishii K, Yagi K, Fujiwara H, Funatsu Y, Hasegawa N, Betsuyaku T. Modulation of murine macrophage TLR7/8-mediated cytokine expression by mesenchymal stem cell-conditioned medium. *Mediators Inflamm* 2013; **2013**: 264260 [PMID: 24191131 DOI: 10.1155/2013/264260]
- 97 **Lai RC**, Arslan F, Lee MM, Sze NS, Choo A, Chen TS, Salto-Tellez M, Timmers L, Lee CN, El Oakley RM, Pasterkamp G, de Kleijn DP, Lim SK. Exosome secreted by MSC reduces myocardial ischemia/reperfusion injury. *Stem Cell Res* 2010; **4**: 214-222 [PMID: 20138817 DOI: 10.1016/j.scr.2009.12.003]
- 98 **Wang D**, Li J, Zhang Y, Zhang M, Chen J, Li X, Hu X, Jiang S, Shi S, Sun L. Umbilical cord mesenchymal stem cell transplantation in active and refractory systemic lupus erythematosus: a multicenter clinical study. *Arthritis Res Ther* 2014; **16**: R79 [PMID: 24661633 DOI: 10.1186/ar4520]

- 99 **Ichii O**, Otsuka-Kanazawa S, Horino T, Kimura J, Nakamura T, Matsumoto M, Toi M, Kon Y. Decreased miR-26a expression correlates with the progression of podocyte injury in autoimmune glomerulonephritis. *PLoS One* 2014; **9**: e110383 [PMID: 25329154 DOI: 10.1371/journal.pone.0110383]
- 100 **Perez-Hernandez J**, Forner MJ, Pinto C, Chaves FJ, Cortes R, Redon J. Increased Urinary Exosomal MicroRNAs in Patients with Systemic Lupus Erythematosus. *PLoS One* 2015; **10**: e0138618 [PMID: 26390437 DOI: 10.1371/journal.pone.0138618]
- 101 **Jonsson R**, Bolstad AI, Brokstad KA, Brun JG. Sjögren's syndrome--a plethora of clinical and immunological phenotypes with a complex genetic background. *Ann N Y Acad Sci* 2007; **1108**: 433-447 [PMID: 17894008 DOI: 10.1196/annals.1422.046]
- 102 **Jonsson R**, Vogelsang P, Volchenkov R, Espinosa A, Wahren-Herlenius M, Appel S. The complexity of Sjögren's syndrome: novel aspects on pathogenesis. *Immunol Lett* 2011; **141**: 1-9 [PMID: 21777618 DOI: 10.1016/j.imlet.2011.06.007]
- 103 **Ramos-Casals M**, Brito-Zerón P, Sisó-Almirall A, Bosch X, Tzioufas AG. Topical and systemic medications for the treatment of primary Sjögren's syndrome. *Nat Rev Rheumatol* 2012; **8**: 399-411 [PMID: 22549247 DOI: 10.1038/nrrheum.2012.53]
- 104 **Hu S**, Vissink A, Arellano M, Roozendaal C, Zhou H, Kallenberg CG, Wong DT. Identification of autoantibody biomarkers for primary Sjögren's syndrome using protein microarrays. *Proteomics* 2011; **11**: 1499-1507 [PMID: 21413148 DOI: 10.1002/pmic.201000206]
- 105 **Delaleu N**, Mydel P, Kwee I, Brun JG, Jonsson MV, Jonsson R. High fidelity between saliva proteomics and the biologic state of salivary glands defines biomarker signatures for primary Sjögren's syndrome. *Arthritis Rheumatol* 2015; **67**: 1084-1095 [PMID: 25545990 DOI: 10.1002/art.39015]
- 106 **Tomosugi N**, Kitagawa K, Takahashi N, Sugai S, Ishikawa I. Diagnostic potential of tear proteomic patterns in Sjögren's syndrome. *J Proteome Res* 2005; **4**: 820-825 [PMID: 15952728 DOI: 10.1021/pr0497576]
- 107 **Aqrabi LA**, Galtung HK, Vestad B, Øvstebø R, Thiede B, Rusthen S, Young A, Guerreiro EM, Utheim TP, Chen X, Utheim ØA, Palm Ø, Jensen JL. Identification of potential saliva and tear biomarkers in primary Sjögren's syndrome, utilising the extraction of extracellular vesicles and proteomics analysis. *Arthritis Res Ther* 2017; **19**: 14 [PMID: 28122643 DOI: 10.1186/s13075-017-1228-x]
- 108 **Sellam J**, Proulle V, Jünger A, Ittah M, Miceli Richard C, Gottenberg JE, Toti F, Benessiano J, Gay S, Freyssinet JM, Mariette X. Increased levels of circulating microparticles in primary Sjögren's syndrome, systemic lupus erythematosus and rheumatoid arthritis and relation with disease activity. *Arthritis Res Ther* 2009; **11**: R156 [PMID: 19832990 DOI: 10.1186/ar2833]
- 109 **Corridoni D**, Arseneau KO, Cominelli F. Inflammatory bowel disease. *Immunol Lett* 2014; **161**: 231-235 [PMID: 24938525 DOI: 10.1016/j.imlet.2014.04.004]
- 110 **Hodson R**. Inflammatory bowel disease. *Nature* 2016; **540**: S97 [PMID: 28002398 DOI: 10.1038/540S97a]
- 111 **Baumgart DC**, Carding SR. Inflammatory bowel disease: cause and immunobiology. *Lancet* 2007; **369**: 1627-1640 [PMID: 17499605 DOI: 10.1016/S0140-6736(07)60750-8]
- 112 **Lee SH**, Kwon JE, Cho ML. Immunological pathogenesis of inflammatory bowel disease. *Intest Res* 2018; **16**: 26-42 [PMID: 29422795 DOI: 10.5217/ir.2018.16.1.26]
- 113 **Cao L**, Xu H, Wang G, Liu M, Tian D, Yuan Z. Extracellular vesicles derived from bone marrow mesenchymal stem cells attenuate dextran sodium sulfate-induced ulcerative colitis by promoting M2 macrophage polarization. *Int Immunopharmacol* 2019; **72**: 264-274 [PMID: 31005036 DOI: 10.1016/j.intimp.2019.04.020]
- 114 **Yang J**, Liu XX, Fan H, Tang Q, Shou ZX, Zuo DM, Zou Z, Xu M, Chen QY, Peng Y, Deng SJ, Liu YJ. Extracellular Vesicles Derived from Bone Marrow Mesenchymal Stem Cells Protect against Experimental Colitis via Attenuating Colon Inflammation, Oxidative Stress and Apoptosis. *PLoS One* 2015; **10**: e0140551 [PMID: 26469068 DOI: 10.1371/journal.pone.0140551]
- 115 **Wu H**, Fan H, Shou Z, Xu M, Chen Q, Ai C, Dong Y, Liu Y, Nan Z, Wang Y, Yu T, Liu X. Extracellular vesicles containing miR-146a attenuate experimental colitis by targeting TRAF6 and IRAK1. *Int Immunopharmacol* 2019; **68**: 204-212 [PMID: 30654310 DOI: 10.1016/j.intimp.2018.12.043]
- 116 **de Visser KE**, Eichten A, Coussens LM. Paradoxical roles of the immune system during cancer development. *Nat Rev Cancer* 2006; **6**: 24-37 [PMID: 16397525 DOI: 10.1038/nrc1782]
- 117 **Ostrand-Rosenberg S**. Immune surveillance: a balance between protumor and antitumor immunity. *Curr Opin Genet Dev* 2008; **18**: 11-18 [PMID: 18308558 DOI: 10.1016/j.gde.2007.12.007]
- 118 **Greening DW**, Gopal SK, Xu R, Simpson RJ, Chen W. Exosomes and their roles in immune regulation and cancer. *Semin Cell Dev Biol* 2015; **40**: 72-81 [PMID: 25724562 DOI: 10.1016/j.semedb.2015.02.009]
- 119 **Zhang HG**, Grizzle WE. Exosomes and cancer: a newly described pathway of immune suppression. *Clin Cancer Res* 2011; **17**: 959-964 [PMID: 21224375 DOI: 10.1158/1078-0432.CCR-10-1489]
- 120 **Yang Y**, Li CW, Chan LC, Wei Y, Hsu JM, Xia W, Cha JH, Hou J, Hsu JL, Sun L, Hung MC. Exosomal PD-L1 harbors active defense function to suppress T cell killing of breast cancer cells and promote tumor growth. *Cell Res* 2018; **28**: 862-864 [PMID: 29959401 DOI: 10.1038/s41422-018-0060-4]
- 121 **Hsu YL**, Hung JY, Chang WA, Jian SF, Lin YS, Pan YC, Wu CY, Kuo PL. Hypoxic Lung-Cancer-Derived Extracellular Vesicle MicroRNA-103a Increases the Oncogenic Effects of Macrophages by Targeting PTEN. *Mol Ther* 2018; **26**: 568-581 [PMID: 29292163 DOI: 10.1016/j.ymthe.2017.11.016]
- 122 **Wang X**, Luo G, Zhang K, Cao J, Huang C, Jiang T, Liu B, Su L, Qiu Z. Hypoxic Tumor-Derived Exosomal miR-301a Mediates M2 Macrophage Polarization via PTEN/PI3Kγ to Promote Pancreatic Cancer Metastasis. *Cancer Res* 2018; **78**: 4586-4598 [PMID: 29880482 DOI: 10.1158/0008-5472.CAN-17-3841]
- 123 **Srinivas TR**, Meier-Kriesche HU. Minimizing immunosuppression, an alternative approach to reducing side effects: objectives and interim result. *Clin J Am Soc Nephrol* 2008; **3** Suppl 2: S101-S116 [PMID: 18308998 DOI: 10.2215/CJN.03510807]
- 124 **Halawa A**. The early diagnosis of acute renal graft dysfunction: a challenge we face. The role of novel biomarkers. *Ann Transplant* 2011; **16**: 90-98 [PMID: 21436782]
- 125 **Williams WW**, Taheri D, Tolkoff-Rubin N, Colvin RB. Clinical role of the renal transplant biopsy. *Nat Rev Nephrol* 2012; **8**: 110-121 [PMID: 22231130 DOI: 10.1038/nrneph.2011.213]

- 126 **Park J**, Lin HY, Assaker JP, Jeong S, Huang CH, Kurdi T, Lee K, Fraser K, Min C, Eskandari S, Routray S, Tannous B, Abdi R, Riella L, Chandraker A, Castro CM, Weissleder R, Lee H, Azzi JR. Integrated Kidney Exosome Analysis for the Detection of Kidney Transplant Rejection. *ACS Nano* 2017; **11**: 11041-11046 [PMID: 29053921 DOI: 10.1021/acsnano.7b05083]
- 127 **Kordelas L**, Rebmann V, Ludwig AK, Radtke S, Ruesing J, Doeppner TR, Epple M, Horn PA, Beelen DW, Giebel B. MSC-derived exosomes: a novel tool to treat therapy-refractory graft-versus-host disease. *Leukemia* 2014; **28**: 970-973 [PMID: 24445866 DOI: 10.1038/leu.2014.41]



Published by **Baishideng Publishing Group Inc**
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: <https://www.f6publishing.com/helpdesk>

<https://www.wjgnet.com>

