

World Journal of *Clinical Oncology*

World J Clin Oncol 2020 September 24; 11(9): 679-760



REVIEW

- 679 Powerful quantifiers for cancer transcriptomics
Jacobas DA
- 705 Healthcare delivery interventions to reduce cancer disparities worldwide
Dickerson JC, Ragavan MV, Parikh DA, Patel MI

MINIREVIEWS

- 723 Circulating tumor DNA: Where are we now? A mini review of the literature
Galarza Fortuna GM, Dvir K

ORIGINAL ARTICLE**Basic Study**

- 732 Diacerein treatment prevents colitis-associated cancer in mice
Paulino DSM, Mendes MCS, Camargo JA, Brambilla SR, Wood Dos Santos T, Ribeiro ML, Carvalheira JBC

Retrospective Cohort Study

- 747 Comparison of efficacy between adjuvant chemotherapy and chemoradiation therapy for pancreatic cancer: AJCC stage-based approach
You MS, Ryu JK, Huh G, Chun JW, Paik WH, Lee SH, Kim YT

ABOUT COVER

Editor-in-Chief of *World Journal of Clinical Oncology*, Professor Hiten Patel is an expert in the field of Urology and a United Kingdom-trained surgeon who provides both general and specialist urological care. Professor Patel has co-led the implementation of minimal access surgery, the fast-track/enhanced recovery after surgery (ERAS) program, and robotic surgery within several units, including the Section of Laparoscopy and Robotic Surgery at the University of Rochester Medical Centre (New York, United States), Guy's and St Thomas' NHS Foundation Trust (London, England) and University Hospital North Norway (Tromsø, Norway). He is passionate about applying simulation training to help educate teams within the field of surgery, surgical techniques, new technologies, and enhanced recovery. (L-Editor: Filipodia)

AIMS AND SCOPE

The primary aim of *World Journal of Clinical Oncology* (*WJCO*, *World J Clin Oncol*) is to provide scholars and readers from various fields of oncology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJCO mainly publishes articles reporting research results and findings obtained in the field of oncology and covering a wide range of topics including art of oncology, biology of neoplasia, breast cancer, cancer prevention and control, cancer-related complications, diagnosis in oncology, gastrointestinal cancer, genetic testing for cancer, gynecologic cancer, head and neck cancer, hematologic malignancy, lung cancer, melanoma, molecular oncology, neurooncology, palliative and supportive care, pediatric oncology, surgical oncology, translational oncology, and urologic oncology.

INDEXING/ABSTRACTING

The *WJCO* is now abstracted and indexed in PubMed, PubMed Central, Emerging Sources Citation Index (Web of Science), China National Knowledge Infrastructure (CNKI), China Science and Technology Journal Database (CSTJ), and Superstar Journals Database.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: *Li-Li Wang*; Production Department Director: *Yun-Xiaojuan Wu*; Editorial Office Director: *Jia-Ping Yan*.

NAME OF JOURNAL

World Journal of Clinical Oncology

ISSN

ISSN 2218-4333 (online)

LAUNCH DATE

November 10, 2010

FREQUENCY

Monthly

EDITORS-IN-CHIEF

Hiten RH Patel

EDITORIAL BOARD MEMBERS

<https://www.wjgnet.com/2218-4333/editorialboard.htm>

PUBLICATION DATE

September 24, 2020

COPYRIGHT

© 2020 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

<https://www.wjgnet.com/bpg/gerinfo/204>

GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjgnet.com/bpg/GerInfo/287>

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjgnet.com/bpg/gerinfo/240>

PUBLICATION ETHICS

<https://www.wjgnet.com/bpg/GerInfo/288>

PUBLICATION MISCONDUCT

<https://www.wjgnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

<https://www.wjgnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjgnet.com/bpg/GerInfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>

Circulating tumor DNA: Where are we now? A mini review of the literature

Gliceida Maria Galarza Fortuna, Kathrin Dvir

ORCID number: Gliceida Maria Galarza Fortuna 0000-0002-1084-953X; Kathrin Dvir 0000-0002-3866-9290.

Author contributions: Galarza Fortuna GM and Dvir K were in charge of conceptualization, data curation, methodology, project administration, and writing the original draft and the definite version of the manuscript.

Conflict-of-interest statement: Galarza Fortuna GM and Dvir K have no conflict of interest to disclose.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Unsolicited manuscript

Received: April 29, 2020

Gliceida Maria Galarza Fortuna, Kathrin Dvir, Internal Medicine, Mount Sinai Medical Center, Miami Beach, FL 33140, United States

Corresponding author: Gliceida Maria Galarza Fortuna, MD, Internal Medicine, Mount Sinai Medical Center, 4300 Alton Road, Miami Beach, FL 33140, United States.
gliceida.galarza@gmail.com

Abstract

For many years tissue biopsy has been the primary procedure to establish cancer diagnosis and determine further treatment and prognosis. However, this method has multiple drawbacks, including, to mention some, being an invasive procedure carrying significant risk for fragile patients and allowing only for a “snapshot” of the tumor biology in time. The process of liquid biopsy allows for a minimally invasive procedure that provides molecular information about underlying cancer by analyzing circulating tumor DNA (ctDNA) *via* next-generation sequencing technology and circulating tumor cells. This paper focuses on describing the basis of ctDNA and its current utilities.

Key Words: Circulating tumor DNA; Liquid biopsy; Molecular profiling; Cancer diagnosis; Cancer screening; Cancer treatment

©The Author(s) 2020. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: This report provides an updated review of the clinical utilities of liquid biopsy for the screening, diagnosis, prognosis and treatment of cancer.

Citation: Galarza Fortuna GM, Dvir K. Circulating tumor DNA: Where are we now? A mini review of the literature. *World J Clin Oncol* 2020; 11(9): 723-731

URL: <https://www.wjnet.com/2218-4333/full/v11/i9/723.htm>

DOI: <https://dx.doi.org/10.5306/wjco.v11.i9.723>

INTRODUCTION

Understanding tumor genetic make-up is more important now than ever given the

Peer-review started: April 29, 2020**First decision:** July 25, 2020**Revised:** July 31, 2020**Accepted:** September 2, 2020**Article in press:** September 2, 2020**Published online:** September 24, 2020**P-Reviewer:** Jeong KY, Scaggiante B**S-Editor:** Huang P**L-Editor:** A**P-Editor:** Wang LL

vast array of available targeted therapies. Traditionally, a biopsy was the only existing approach to understanding tissue histological composition and its genetic environment. However, this approach allows a merely static analysis of a tumor at a given time and a given location, while cancer is a rather dynamic entity undergoing continual alterations. The concept that a tumor can harvest different genetic material, which can be identified by next-generation sequencing (NGS), has been extensively studied and validated; this discordance can occur both within the primary tumor and between primary and metastatic lesions^[1] and is partly because a tumor is comprised of different cell clones^[2]. For example, Geyer *et al.*^[3] demonstrated the presence of intra-tumor heterogeneity in breast cancer, as evidenced by the presence of overexpressed *HER2* mutation only in some regions of a primary tumor. This lack of uniformity within a tumor's genetic environment and spatial heterogeneity is a therapeutic challenge. A single biopsy would not signify an accurate and complete assessment of a tumor's genetic composition. Hence, a need has risen for more comprehensive techniques, which would yield a better characterization of tumor composition and its driver mutations.

HOW IS CIRCULATING TUMOR DNA DETECTED?

Circulating tumor cells (CTCs) have been observed in patients' bloodstream. CTCs are believed to reach a patient's plasma by migration from the principal or metastatic tumor site secondary to tumor invasion, shedding or after the tumor site experiences mechanical stress after surgery^[4]. Analysis of both CTC and circulating tumor DNA (ctDNA) is the backbone of the development of liquid biopsy.

In 1948, a group of French scientists detected free DNA fragments circulating in the plasma^[5]. Several successive studies have been conducted to pursue the mechanism in which DNA fragments are released into the serum from the cells in their healthy, inflamed, or diseased states. To date, the consensus hypothesis is that the DNA enters the circulation through passive and active mechanisms^[6]. The passive release of DNA fragments into the circulation is thought to be secondary to cell death, both through the apoptosis and necrosis pathways. In contrast, the active secretion of DNA fragments into the bloodstream has yet to be understood entirely^[2]. Some studies propose that tumor cells release micro-vesicles (exons) containing fragments of double-stranded DNA (ctDNA); however, this theory is still not universally accepted^[7]. Even though circulating cell-free DNA (cfDNA) is detectable in healthy individuals, its concentration is significantly increased in cancer patients^[8]. ctDNA is also released to the bloodstream by the above-described mechanisms from primary and metastatic tumor sites.

Two main approaches are utilized for the detection of ctDNA - a targeted, and an untargeted approach. The targeted approach can detect previously determined genetic mutations, such as specific driver mutations that frequently occur in individual tumors and toward which targeted therapy has been developed^[9]. On the other hand, the untargeted approach does not need any prior knowledge of the genetic mutations associated with the tumor under study.

PCR techniques are the backbone of all strategies for the detection of ctDNA in the targeted approach. Several PCR techniques such as digital PCR (dPCR), emulsion PCR (ePCR), and BEAMing (beads, emulsion, amplification, and magnetics) - which is a high-throughput droplet-based dPCR, have been developed for the detection of determining DNA mutations^[10]. Nunes *et al.*^[11] used quantitative methylation-specific PCR to determine *APC*, *HOXA9*, *RARB2*, and *RASSF1A* promoter methylation levels. Their study was able to detect high *HOXA9* methylation levels in patients with squamous cell lung cancer with 82.2% accuracy. Correspondingly, Klotten *et al.*^[12] identified *KRAS* mutation in patients with colorectal cancer using Intplex allele-specific PCR (Intplex PCR). In this study, ctDNA analysis using the Intplex technique had a 70% specificity and 50% sensitivity when comparing serum samples with the tissue sample. Furthermore, the concordance with their match tissue sample was 66%.

NGS, on the other hand, is capable of analyzing millions of DNA sequences and then compare it with a pre-determined genome or produce a *de novo* sequence assembly^[13]. Thompson *et al.*^[14] detected approximately 275 alterations in 45 genes on patients, of which 86% were previously diagnosed with lung cancer. *Epidermal growth factor receptor (EGFR)* mutation was the most common mutation in their study. When compared to tissue DNA analysis, their concordance was 79%. Another important finding of this study was that patients who had failed treatment, as evidenced by disease progression while on therapy, ctDNA analysis showed newly emerged

resistance mutations when tissue analysis was not feasible.

Given the presence of both circulating cell-free DNA from healthy tissue and ctDNA, the isolation of ctDNA continues to be a diagnostic challenge, as only approximately 0.01% of all circulating DNA is tumor-derived^[15]. This limitation has been overcome by the recent development of “ultra-sensitive” assays that allow differentiating ctDNA from cfDNA, which are being used not only for the detection of genetic mutations but also for the early detection of disease recurrence and monitoring for therapy response^[7]. One example of an ultrasensitive assay is the Cancer Personalized Profiling by Deep Sequencing (CAPP-Seq); this technology consists of a capture-based ctDNA detection method which can detect most of the main types of mutations: Copy number alterations, rearrangement, indels, and single nucleotide variants, by the evaluation of large segments of the genome utilizing enriched genomic regions that have been selected before sequencing^[16,17]. This method allows for the detection of various mutations, increasing the sensitivity of the test, compared to other NGS based assays, and aids the evaluation of intratumor heterogeneity^[17]. This technological advancement has led to the development of liquid biopsy, which provides a genetic characterization of tumors from blood, bronchial alveolar lavage samples, or colony-stimulating factor samples. This technology brings many clinical utilities, especially in patients with solid tumors that are not amenable to repeat biopsies, including the measurement of disease burden and detection of emerging mutations, among others.

LIQUID BIOPSY AS A PROGNOSTIC MARKER

As already mentioned, cfDNA levels are higher in the disease state when compared to healthy individuals^[8]. Hence, several studies have tried to establish a correlation between cfDNA and CTC levels and disease prognosis. Lee *et al*^[18] studied the relationship between CTC in patients with epithelial ovarian cancer and disease prognosis; they found that newly diagnosed patients with CTC > 3 had a significantly shorter progression-free survival. This marker was also determined to be a poor prognostic factor in multivariate Analysis (HR = 1.3; 95%CI: 3.08-32.149).

Likewise, Ito *et al*^[19] studied a group of patients with metastatic breast cancer treated with the microtubule-depolymerizing agent eribulin and determined the presence of CTC. They further classified the CTC into mesenchymal and epithelial depending on their vimentin or pan-cytokeratin positivity. Patients with a high number of CTC (≥ 3) had a significantly shorter overall survival ($P = 0.037$). No difference was observed on the sub-analysis of mesenchymal and epithelial CTC.

Furthermore, some genetic alterations detected through ctDNA analysis are associated with increased survival. Cheng *et al*^[20] established that the detection of *ERBB2* exon 17 mutation and *K-Ras G12V* mutation in a cohort of patients with metastatic pancreatic cancer was associated with a statistically significant increase in overall survival (P values = 0.016 and 0.015; respectively). Another important finding of this research is an increased observed rate of *BRCA2* mutations in patients with metastatic pancreatic cancer; prior data showed a 5% mutation rate in these patients; this study showed an 11.7% mutation rate. *BRCA2* mutations in pancreatic cancer have been associated with an improved response to cisplatin-base chemotherapy. These observations have led to a new possible pharmacological approach to a disease that often carries a dismal survival.

Correspondingly, Xu *et al*^[21] developed and validated a combined prognosis score (cp-score) using eight methylation markers found on ctDNA in addition to clinical, demographic, and the American Joint Committee on Cancer (AJCC) stage. In their research among 377 hepatocellular carcinomas (HCC) samples, a cp-score ≤ 0.24 was determined to be a low risk while a cp-score > 0.24 was classified as high risk, with a statistically significant median survival ($P < 0.0001$). This research showed that cp-score, in combination with Tumor-Node-Metastasis staging, increased the prognostic prediction accuracy for patients with HCC.

LIQUID BIOPSY AND THERAPEUTIC GUIDANCE

Therapy selection is among the most clinically relevant current utilization of liquid biopsies; this holds especially true for tumors that are difficult to biopsy or for patients that are too fragile to undergo surgical exploration^[22]. Moreover, this approach also provides the means to monitor tumor evolution and the development of therapy

resistance.

Shu *et al*^[23] studied ctDNA by targeted NGS-based gene mutation profiling in a total of 605 cancer patients with 29 different tumor types. In their study, the most frequently observed mutated tumor suppressor genes were *TP53*, *APC* and *DNMT3A*, while the most commonly mutated oncogenes were *EGFR*, *KRAS*, and *PIK3CA*; 35.3% of the detected mutations were clinically-actionable, and 66% of those mutated genes have FDA approved targeted therapy or have therapy undergoing current clinical trials. This fact makes ctDNA evaluation a helpful tool to identify molecular mutations to which targeted therapies are available and thus guide management, which would, after that, influences survival.

Correspondingly, microsatellite instability (MSI) is a biomarker used to predict response to immune checkpoint blockade for cervical, cholangiocarcinoma, colorectal, endometrial, esophageal, esophagogastric, gastric, ovarian, pancreatic and prostate cancer^[24]. Willis *et al*^[24] validated MSI detection using a plasma-based genotyping panel, Guardant360. This method was able to detect MSI with an accuracy of 98.4% and a positive predictive value of 95% compared to tissue-based testing. Moreover, in their study they were able to follow up 16 patients with metastatic gastric cancer treated with either pembrolizumab or nivolumab after not achieving remission with the standard chemotherapy regimen; 10 of the 16 patients achieved complete (3/16) or partial response (7/16) while three patients were reported to have stable disease, resulting in an objective response rate of 63% and a disease control rate of 81%. Immune checkpoint inhibitors are gaining FDA approval rapidly in advanced cancer, and the detection of MSI provides the clinical oncologist with a novel, fast and non-invasive tool to predict response to therapy that is especially beneficial in this patient population given their poor clinical and performance status, which limits the possibility of more traditional testing such as biopsies.

MONITORING TREATMENT RESISTANCE WITH CTDNA

In lung cancer, namely non-small cell lung cancer, the presence of a somatic mutation in the *EGFR-L858R* and exon 19 deletion- has been noted to predict the response to EGFR tyrosine kinase inhibitors (TKIs) such as erlotinib, gefitinib, and afatinib. Therefore, several PCR based platforms have been used to detect this mutation in plasma^[25,26]. However, this response is often limited to the first 10-12 mo of treatment; when this is lost given the development of acquired resistance mutations^[27].

Tumor resistance to either targeted therapy or chemotherapy follows one of two pathways, a pharmacological or biological resistance. Pharmacological resistance infers the progression of disease in the setting of inadequate drug exposure against a targeted protein. On the other hand, biological resistance consists of changes in the drug target, such as the development of a secondary *EGFR* mutation such as *T790M*, *D761Y*, and *L747S* mutations. This biological mechanism, in part, depends on the initial biological heterogeneity of the tumor. Directed therapy to specific oncogenes leads to increase gene copies of the sub-clones present in the tumor as a response to the selection pressure. The selected sub-clones often lead to the emergence of a structural change of the targeted protein – alterations in the drug target – or leads to the development of a bypass track that feeds the tumor even in the presence of the inhibition on the initial target^[26].

Therefore, monitoring ctDNA to detect secondary mutations' emergence could provide a tool to adjust therapy before the development of clinical signs of disease progression. The JP-CLEAR trial analyzed the plasma of 121 patients with advance or postoperative recurrent non-small cell lung cancer on first or second-generation TKI therapy without known disease progression. Their plasma was monitor every 1-2 mo with a PCR based method developed to detect sensitizing *EGFR* mutations and the *T790M* resistance mutation. In their study, 33.3% of the patients developed *T790M* mutation and disease progression while on first-line therapy with TKIs, proving ctDNA monitoring to be a useful tool to assess the emergence of secondary mutations associated with treatment resistance^[28].

Similarly, colorectal cancer with wild type *RAS* and *BRAF* genes is known for responding to cetuximab treatment, while *HER2* amplification has been associated with treatment resistance^[29]. Liu *et al*^[29] used a ddPCR based method to detect *HER2* amplification on a total of 36 patients with wild type *RAS/BRAF* metastatic colorectal cancer undergoing therapy with cetuximab. Their *HER2* status was determined at the time of disease progression. Of the 36 patients with documented disease progression while on cetuximab treatment, five were found to have *HER2* amplification at the time

of progression. During the study, plasma ctDNA for *HER2* status was carried out, and *HER2* levels were able to predict radiological progression with a lead time of 2 mo. This idea again proves that monitoring plasma ctDNA while on therapy can be an essential tool in detecting disease progression before any apparent changes in radiographic studies.

CTDNA AS A SCREENING TOOL

To date, ctDNA has not yet made its way onto becoming a screening tool. This technology holds significant potential to become a vital screening strategy, leading to earlier diagnosis. However, ctDNA must overcome many barriers before being applied as a screening strategy. One of the most critical limiting factors is the low concentration of ctDNA in asymptomatic patients^[30]. The level of ctDNA in the healthy population has been determined to range from 1-10 ng/mL^[31]. Given its low concentration, approximately 150-300 mL of blood would be required to reach a 95% sensitivity of a screening test for breast cancer^[30,32]. Moreover, given that healthy cells contribute to cfDNA in plasma, introducing ctDNA as a screening tool could lead to an increased rate of false positives^[30].

DISCUSSION

The introduction of liquid biopsy into clinical practice provided a novel tool for the detection, monitoring, and characterization of malignancy. It allows the detection of tumor circulating DNA *via* peripheral blood sampling, analyzing its genomic compositing and specific targetable genomic alterations, thereby enhancing the delivery of personalized medicine *via* targeted treatments. Liquid biopsy can be utilized to monitor the response to therapy and act as a surveillance modality in detecting disease progression. Moreover, its non-invasive nature renders a feasible solution for patients who are not candidates for surgical intervention or tissue sampling. Despite its many advantages, widely acknowledged by the scientific community, liquid biopsy has essential disadvantages, namely test sensitivity, which reaches 85% in certain malignancies^[33] and high cost.

Since its introduction, liquid biopsy has been the focus of research and has been incorporated into many clinical trials' protocols worldwide. Many of the currently registered trials researching liquid biopsy and ctDNA in clinicaltrials.gov (Table 1) are aiming to characterize the following three topics: (1) The detection of targetable genetic alterations in patients who are not amenable to surgical biopsies; (2) Monitoring response to therapy and detecting minimal residual disease; and (3) Identifying the development of *de-novo* mutations after administration of specific treatments. Commonly used modalities to detect circulating tumor DNA extracted *via* liquid biopsy are NGS, BEAMing, and PCR. While these modalities can identify mutant alleles in as little as 2% frequency^[6], all three have significant limitations. NGS requires high-quality DNA fragments for the analysis, which is often difficult to detect through liquid biopsy; it necessitates trained bioinformaticians to analyze the data, and it is associated with a high cost with variable insurance coverage^[6]. The BEAMing modality provides an even higher mutation detection ability while carrying a lower cost when compared to PCR and NGS. However, it requires a previously established DNA template to target the genomic area of interest, therefore limiting its mutations detection repertoire^[34]. Similarly, the different PCR modalities (*i.e.*, dPCR and ePCR), also require pre-established genomic templates, thereby limiting its array of detection to specific targetable mutations rather than allowing a broad screening for alternated tumor genome^[35].

In summary, liquid biopsy is useful in detecting ctDNA in most types of cancer at both early and advanced stages of cancer. Studies have shown ctDNA often precedes the radiological and clinical signs of disease by as early as six months^[36]. Its ability to detect tumor genome *via* non-invasive technique holds an immense potential to be utilized as a screening method for cancer detection on appropriate individuals.

CONCLUSION

Multiple commercial ctDNA analysis platforms are currently available and are widely

Table 1 Current active and completed phase II-III clinical trials utilizing liquid biopsy technology

Study title	Cancer type	Phase	Clinical aspect being investigated	Fluid analyzed	Target(s) and study objectives	Identifier
TAGRISSO (Osimertinib) in NSCLC patients in whom <i>T790</i> mutations are detected by liquid biopsy using BALF, plasma or pleural effusion	Lung cancer (NSCLC)	Phase II	Therapeutic guidance	BALF, plasma or pleural effusion.	<i>EGFR T790M</i> detection	NCT03394118
Post-surgical liquid biopsy-guided treatment of stage III and high-risk stage II colon cancer patients: The PEGASUS trial	Colon cancer	Phase II	Therapeutic guidance	Plasma	ctDNA detection	NCT04259944
Liquid biopsy as a tool to evaluate resistance to first and third lines (AZD9291) (<i>EGFR</i>) (TKIs) in <i>EGFR</i> mutant NSCLC	Lung cancer (NSCLC)	Phase II	Clonal evolution – disease monitoring/therapy resistance	Plasma	<i>EGFR, T790M, C797S, L858R, del 19 EGFR, KRAS /NRAS, BRAF, PI3K</i>	NCT02771314
Liquid biopsy in monitoring the neoadjuvant chemotherapy and operation in gastric cancer	Gastric cancer	Phase II	Disease monitoring/progression/future directed therapy development	Plasma	Compare CTC, ctDNA and ctDNA detection to tumor markers level (CEA, CA 19-9 and CA72-4) and imaging findings to assess post-surgical prognosis	NCT03957564
Clinical utility of liquid biopsy in brigatinib ALK+ patients (CUBIK)	Lung cancer (NSCLC)	Phase II	Disease monitoring/clonal evolution.	Plasma	<i>EML4-ALK</i> rearrangement detection	NCT04223596
Olmotinib trial in <i>T790M</i> (+) NSCLC patients detected by liquid biopsy using BALF extracellular vesicular DNA	Lung cancer (NSCLC)	Phase II	Therapeutic guidance- treatment	BALF	<i>EGFR T790M</i> detection	NCT03228277
Nalirinox neo-pancreas <i>RAS</i> mut ctDNA study	Pancreatic cancer	Phase II	Disease monitoring/clonal evolution.	Plasma	<i>KRAS</i> detection	NCT04010552
Mechanisms of resistance in <i>EGFR</i> mutated nonpretreated advanced lung cancer receiving OSimertinib (MELROSE)	Lung cancer (NSCLC)	Phase II	Disease monitoring/clonal evolution/mechanisms of resistance	Plasma	<i>EGFR T790M</i> detection	NCT03865511
PRIMUS002: Looking at 2 neo-adjuvant treatment regimens for resectable and borderline resectable pancreatic cancer (PRIMUS002)	Pancreatic cancer	Phase II	Therapy response	Plasma	Liquid biopsy analyzed using ctDNA at different time-points to see if these can be used to define patient subgroups	NCT04176952
Intermittent or continuous panitumumab plus FOLFIRI for <i>RAS/B-RAF</i> wild-type metastatic colorectal cancer (IMPROVE)	Colon cancer	Phase II	Disease monitoring/clonal evolution/mechanisms of resistance	Plasma	<i>RAS</i> and <i>BRAF</i> detection	NCT04425239
Osimertinib treatment on <i>EGFR T790M</i> plasma positive NSCLC patients (APPLE)	Lung cancer (NSCLC)	Phase II	Disease monitoring/progression	Plasma	<i>EGFR T790M</i> detection	NCT02856893
Treatment of metastatic bladder cancer at the time of biochemical relapse following radical cystectomy (TOMBOLA)	Bladder cancer	Phase II	Disease monitoring/prognosis	Plasma	ctDNA levels to assess metastatic disease	NCT04138628
Deferred cytoreductive nephrectomy in synchronous metastatic renal cell carcinoma: The NORDIC-SUN-Trial (NORDIC-SUN)	Renal cell carcinoma	Phase III	Future translational research	Plasma	ctDNA quantity and NGS to establish mutations.	NCT03977571
Study of the molecular features of postmenopausal women with HR+ HER2-negative aBC on first-line treatment with ribociclib and letrozole and, in patients with a <i>PIK3CA</i> mutation, on second-line treatment with alpelisib plus fulvestrant (BioItaLEE)	Breast cancer	Phase III	Disease progression/clonal evolution/therapy response	Plasma	<i>PIK3CA</i> detection	NCT03439046
Second-line FOLFIRI + panitumumab in subjects with wild	Colorectal cancer	Phase II	Disease progression/clonal evolution	Plasma	<i>RAS/BRAF</i> detection	NCT03751176

NSCLC: Non-small cell lung cancers; BALF: Bronchoalveolar lavage fluid; EGFR: Epidermal growth factor receptor; TKIs: Tyrosine kinase inhibitors; ctDNA: Circulating tumor DNA; CEA: Carcinoembryonic antigen; CTC: Circulating tumor cells; NGS: Next-generation sequencing.

used in clinical oncology. This mini-review aims to describe the fundamental technical aspects of liquid biopsies and their clinical implications. Since its introduction, liquid biopsies have been incorporated into clinical protocols and clinical trials worldwide. The current registered active phase 2-4 clinical trials have been summarized (Table 1). Most of these studies aim to utilize liquid biopsy for: (1) The detection of known mutations that have targeted therapy in patients who are not amenable for surgical biopsies; (2) Monitoring disease progression and therapeutic response – minimal residual disease; and (3) Detection the development of *de novo* mutations after specific therapies. The introduction of liquid biopsy into clinical practice provides a novel tool for the diagnosis, future development of personalized/targeted treatments, and monitoring of multiple malignancies. Furthermore, it gives the clinician with a new understanding of tumors' biology by providing a more comprehensive map of the genetic origin of cancerous processes. With the increased use of liquid biopsies, new associations between driver mutations and specific tumors will become more apparent, which could lead to the development of more targeted therapies and even the utilization of this method for screening purposes before the development of diseased states.

ACKNOWLEDGEMENTS

The authors would like to acknowledge Dr. Mike Cusnir, for his contribution and support in this project.

REFERENCES

- 1 **McGranahan N**, Swanton C. Clonal Heterogeneity and Tumor Evolution: Past, Present, and the Future. *Cell* 2017; **168**: 613-628 [PMID: 28187284 DOI: 10.1016/j.cell.2017.01.018]
- 2 **Schwarzenbach H**, Hoon DS, Pantel K. Cell-free nucleic acids as biomarkers in cancer patients. *Nat Rev Cancer* 2011; **11**: 426-437 [PMID: 21562580 DOI: 10.1038/nrc3066]
- 3 **Geyer FC**, Weigelt B, Natrajan R, Lambros MB, de Biase D, Vatcheva R, Savage K, Mackay A, Ashworth A, Reis-Filho JS. Molecular analysis reveals a genetic basis for the phenotypic diversity of metaplastic breast carcinomas. *J Pathol* 2010; **220**: 562-573 [PMID: 20099298 DOI: 10.1002/path.2675]
- 4 **Luo W**, Rao M, Qu J, Luo D. Applications of liquid biopsy in lung cancer-diagnosis, prognosis prediction, and disease monitoring. *Am J Transl Res* 2018; **10**: 3911-3923 [PMID: 30662639]
- 5 **Mandel P**, Metais P. Les acides nucléiques du plasma sanguin chez l'homme. *C R Seances Soc Biol Fil. C R Seances Soc Biol Fil* 1948; **142**: 241-243 [PMID: 18875018]
- 6 **Crowley E**, Di Nicolantonio F, Loupakis F, Bardelli A. Liquid biopsy: monitoring cancer-genetics in the blood. *Nat Rev Clin Oncol* 2013; **10**: 472-484 [PMID: 23836314 DOI: 10.1038/nrclinonc.2013.110]
- 7 **Alix-Panabières C**, Pantel K. Clinical Applications of Circulating Tumor Cells and Circulating Tumor DNA as Liquid Biopsy. *Cancer Discov* 2016; **6**: 479-491 [PMID: 26969689 DOI: 10.1158/2159-8290.CD-15-1483]
- 8 **Alix-Panabières C**, Schwarzenbach H, Pantel K. Circulating tumor cells and circulating tumor DNA. *Annu Rev Med* 2012; **63**: 199-215 [PMID: 22053740 DOI: 10.1146/annurev-med-062310-094219]
- 9 **Heitzer E**, Ulz P, Geigl JB. Circulating tumor DNA as a liquid biopsy for cancer. *Clin Chem* 2015; **61**: 112-123 [PMID: 25388429 DOI: 10.1373/clinchem.2014.222679]
- 10 **Postel M**, Roosen A, Laurent-Puig P, Taly V, Wang-Renault SF. Droplet-based digital PCR and next generation sequencing for monitoring circulating tumor DNA: a cancer diagnostic perspective. *Expert Rev Mol Diagn* 2018; **18**: 7-17 [PMID: 29115895 DOI: 10.1080/14737159.2018.1400384]
- 11 **Nunes SP**, Diniz F, Moreira-Barbosa C, Constâncio V, Silva AV, Oliveira J, Soares M, Paulino S, Cunha AL, Rodrigues J, Antunes L, Henrique R, Jerónimo C. Subtyping Lung Cancer Using DNA Methylation in Liquid Biopsies. *J Clin Med* 2019; **8** [PMID: 31546933 DOI: 10.3390/jcm8091500]
- 12 **Kloten V**, Rüchel N, Bröchle NO, Gasthaus J, Freudenmacher N, Steib F, Mijnes J, Eschenbruch J, Binnebösel M, Knüchel R, Dahl E. Liquid biopsy in colon cancer: comparison of different circulating DNA extraction systems following absolute quantification of KRAS mutations using Intplex allele-specific PCR. *Oncotarget* 2017; **8**: 86253-86263 [PMID: 29156792 DOI: 10.18632/oncotarget.21134]
- 13 **Elazezy M**, Joosse SA. Techniques of using circulating tumor DNA as a liquid biopsy component in cancer management. *Comput Struct Biotechnol J* 2018; **16**: 370-378 [PMID: 30364656 DOI: 10.1016/j.csbj.2018.10.002]
- 14 **Thompson JC**, Yee SS, Troxel AB, Savitch SL, Fan R, Balli D, Lieberman DB, Morrissette JD, Evans TL,

- Baum J, Aggarwal C, Kosteva JA, Alley E, Ciunci C, Cohen RB, Bagley S, Stonehouse-Lee S, Sherry VE, Gilbert E, Langer C, Vachani A, Carpenter EL. Detection of Therapeutically Targetable Driver and Resistance Mutations in Lung Cancer Patients by Next-Generation Sequencing of Cell-Free Circulating Tumor DNA. *Clin Cancer Res* 2016; **22**: 5772-5782 [PMID: 27601595 DOI: 10.1158/1078-0432.CCR-16-1231]
- 15 **Cheng F**, Su L, Qian C. Circulating tumor DNA: a promising biomarker in the liquid biopsy of cancer. *Oncotarget* 2016; **7**: 48832-48841 [PMID: 27223063 DOI: 10.18632/oncotarget.9453]
- 16 **Newman AM**, Bratman SV, To J, Wynne JF, Eclov NC, Modlin LA, Liu CL, Neal JW, Wakelee HA, Merritt RE, Shrager JB, Loo BW Jr, Alizadeh AA, Diehn M. An ultrasensitive method for quantitating circulating tumor DNA with broad patient coverage. *Nat Med* 2014; **20**: 548-554 [PMID: 24705333 DOI: 10.1038/nm.3519]
- 17 **Bratman SV**, Newman AM, Alizadeh AA, Diehn M. Potential clinical utility of ultrasensitive circulating tumor DNA detection with CAPP-Seq. *Expert Rev Mol Diagn* 2015; **15**: 715-719 [PMID: 25773944 DOI: 10.1586/14737159.2015.1019476]
- 18 **Lee M**, Kim EJ, Cho Y, Kim S, Chung HH, Park NH, Song YS. Predictive value of circulating tumor cells (CTCs) captured by microfluidic device in patients with epithelial ovarian cancer. *Gynecol Oncol* 2017; **145**: 361-365 [PMID: 28274569 DOI: 10.1016/j.ygyno.2017.02.042]
- 19 **Ito M**, Horimoto Y, Tokuda E, Murakami F, Uomori T, Himuro T, Nakai K, Orihata G, Iijima K, Saito M. Impact of circulating tumour cells on survival of eribulin-treated patients with metastatic breast cancer. *Med Oncol* 2019; **36**: 89 [PMID: 31520329 DOI: 10.1007/s12032-019-1314-9]
- 20 **Cheng H**, Liu C, Jiang J, Luo G, Lu Y, Jin K, Guo M, Zhang Z, Xu J, Liu L, Ni Q, Yu X. Analysis of ctDNA to predict prognosis and monitor treatment responses in metastatic pancreatic cancer patients. *Int J Cancer* 2017; **140**: 2344-2350 [PMID: 28205231 DOI: 10.1002/ijc.30650]
- 21 **Xu RH**, Wei W, Krawczyk M, Wang W, Luo H, Flagg K, Yi S, Shi W, Quan Q, Li K, Zheng L, Zhang H, Caughey BA, Zhao Q, Hou J, Zhang R, Xu Y, Cai H, Li G, Hou R, Zhong Z, Lin D, Fu X, Zhu J, Duan Y, Yu M, Ying B, Zhang W, Wang J, Zhang E, Zhang C, Li O, Guo R, Carter H, Zhu JK, Hao X, Zhang K. Circulating tumour DNA methylation markers for diagnosis and prognosis of hepatocellular carcinoma. *Nat Mater* 2017; **16**: 1155-1161 [PMID: 29035356 DOI: 10.1038/nmat4997]
- 22 **Corcoran RB**, Chabner BA. Application of Cell-free DNA Analysis to Cancer Treatment. *N Engl J Med* 2018; **379**: 1754-1765 [PMID: 30380390 DOI: 10.1056/NEJMr1706174]
- 23 **Shu Y**, Wu X, Tong X, Wang X, Chang Z, Mao Y, Chen X, Sun J, Wang Z, Hong Z, Zhu L, Zhu C, Chen J, Liang Y, Shao H, Shao YW. Circulating Tumor DNA Mutation Profiling by Targeted Next Generation Sequencing Provides Guidance for Personalized Treatments in Multiple Cancer Types. *Sci Rep* 2017; **7**: 583 [PMID: 28373672 DOI: 10.1038/s41598-017-00520-1]
- 24 **Willis J**, Lefterova MI, Artyomenko A, Kasi PM, Nakamura Y, Mody K, Catenacci DVT, Fakih M, Barbacioru C, Zhao J, Sikora M, Fairclough SR, Lee H, Kim KM, Kim ST, Kim J, Gavino D, Benavides M, Peled N, Nguyen T, Cusnir M, Eskander RN, Azzi G, Yoshino T, Banks KC, Raymond VM, Lanman RB, Chudova DI, Talasz A, Kopetz S, Lee J, Odegaard JI. Validation of Microsatellite Instability Detection Using a Comprehensive Plasma-Based Genotyping Panel. *Clin Cancer Res* 2019; **25**: 7035-7045 [PMID: 31383735 DOI: 10.1158/1078-0432.CCR-19-1324]
- 25 **Liang Z**, Cheng Y, Chen Y, Hu Y, Liu WP, Lu Y, Wang J, Wang Y, Wu G, Ying JM, Zhang HL, Zhang XC, Wu YL. EGFR T790M ctDNA testing platforms and their role as companion diagnostics: Correlation with clinical outcomes to EGFR-TKIs. *Cancer Lett* 2017; **403**: 186-194 [PMID: 28642172 DOI: 10.1016/j.canlet.2017.06.008]
- 26 **Camidge DR**, Pao W, Sequist LV. Acquired resistance to TKIs in solid tumours: learning from lung cancer. *Nat Rev Clin Oncol* 2014; **11**: 473-481 [PMID: 24981256 DOI: 10.1038/nrclinonc.2014.104]
- 27 **Del Re M**, Crucitta S, Gianfilippo G, Passaro A, Petrini I, Restante G, Michelucci A, Fogli S, de Marinis F, Porta C, Chella A, Danesi R. Understanding the Mechanisms of Resistance in EGFR-Positive NSCLC: From Tissue to Liquid Biopsy to Guide Treatment Strategy. *Int J Mol Sci* 2019; **20** [PMID: 31416192 DOI: 10.3390/ijms20163951]
- 28 **Usui K**, Yokoyama T, Naka G, Ishida H, Kishi K, Uemura K, Ohashi Y, Kunitoh H. Plasma ctDNA monitoring during epidermal growth factor receptor (EGFR)-tyrosine kinase inhibitor treatment in patients with EGFR-mutant non-small cell lung cancer (JP-CLEAR trial). *Jpn J Clin Oncol* 2019; **49**: 554-558 [PMID: 30809659 DOI: 10.1093/jcco/hyz023]
- 29 **Liu R**, Zhao X, Guo W, Huang M, Qiu L, Zhang W, Zhang Z, Li W, Zhu X, Chen Z. Dynamic monitoring of HER2 amplification in circulating DNA of patients with metastatic colorectal cancer treated with cetuximab. *Clin Transl Oncol* 2020; **22**: 928-934 [PMID: 31571151 DOI: 10.1007/s12094-019-02215-7]
- 30 **Chen M**, Zhao H. Next-generation sequencing in liquid biopsy: cancer screening and early detection. *Hum Genomics* 2019; **13**: 34 [PMID: 31370908 DOI: 10.1186/s40246-019-0220-8]
- 31 **Wan JCM**, Massie C, Garcia-Corbacho J, Mouliere F, Brenton JD, Caldas C, Pacey S, Baird R, Rosenfeld N. Liquid biopsies come of age: towards implementation of circulating tumour DNA. *Nat Rev Cancer* 2017; **17**: 223-238 [PMID: 28233803 DOI: 10.1038/nrc.2017.7]
- 32 **Tangvarasittichai O**, Jaiwang W, Tangvarasittichai S. The plasma DNA concentration as a potential breast cancer screening marker. *Indian J Clin Biochem* 2015; **30**: 55-58 [PMID: 25646041 DOI: 10.1007/s12291-013-0407-z]
- 33 **Revelo AE**, Martin A, Velasquez R, Kulandaisamy PC, Bustamante J, Keshishyan S, Otterson G. Liquid biopsy for lung cancers: an update on recent developments. *Ann Transl Med* 2019; **7**: 349 [PMID: 31516895 DOI: 10.21037/atm.2019.03.28]
- 34 **O'Leary B**, Hrebien S, Beaney M, Fribbens C, Garcia-Murillas I, Jiang J, Li Y, Huang Bartlett C, André F, Loibl S, Loi S, Cristofanilli M, Turner NC. Comparison of BEAMing and Droplet Digital PCR for Circulating Tumor DNA Analysis. *Clin Chem* 2019; **65**: 1405-1413 [PMID: 31551314 DOI: 10.1373/clinchem.2019.305805]
- 35 **Sato Y**, Matoba R, Kato K. Recent Advances in Liquid Biopsy in Precision Oncology Research. *Biol Pharm Bull* 2019; **42**: 337-342 [PMID: 30828064 DOI: 10.1248/bpb.b18-00804]
- 36 **Jamal-Hanjani M**, Wilson GA, McGranahan N, Birkbak NJ, Watkins TBK, Veeriah S, Shafi S, Johnson

DH, Mitter R, Rosenthal R, Salm M, Horswell S, Escudero M, Matthews N, Rowan A, Chambers T, Moore DA, Turajlic S, Xu H, Lee SM, Forster MD, Ahmad T, Hiley CT, Abbosh C, Falzon M, Borg E, Marafioti T, Lawrence D, Hayward M, Kolvekar S, Panagiotopoulos N, Janes SM, Thakrar R, Ahmed A, Blackhall F, Summers Y, Shah R, Joseph L, Quinn AM, Crosbie PA, Naidu B, Middleton G, Langman G, Trotter S, Nicolson M, Remmen H, Kerr K, Chetty M, Gomersall L, Fennell DA, Nakas A, Rathinam S, Anand G, Khan S, Russell P, Ezhil V, Ismail B, Irvin-Sellers M, Prakash V, Lester JF, Kornaszewska M, Attanoos R, Adams H, Davies H, Dentro S, Tanieri P, O'Sullivan B, Lowe HL, Hartley JA, Iles N, Bell H, Ngai Y, Shaw JA, Herrero J, Szallasi Z, Schwarz RF, Stewart A, Quezada SA, Le Quesne J, Van Loo P, Dive C, Hackshaw A, Swanton C; TRACERx Consortium. Tracking the Evolution of Non-Small-Cell Lung Cancer. *N Engl J Med* 2017; **376**: 2109-2121 [PMID: 28445112 DOI: 10.1056/NEJMoa1616288]



Published by **Baishideng Publishing Group Inc**
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: <https://www.f6publishing.com/helpdesk>

<https://www.wjgnet.com>

