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AIMS AND SCOPE

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Basic Study

Umbilical cord-derived mesenchymal stem cells preconditioned with isorhamnetin: potential therapy for burn wounds

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Abstract

BACKGROUND

Impaired wound healing can be associated with different pathological states. Burn wounds are the most common and detrimental injuries and remain a major health issue worldwide. Mesenchymal stem cells (MSCs) possess the ability to regenerate tissues by secreting factors involved in promoting cell migration, proliferation and differentiation, while suppressing immune reactions. Preconditioning of MSCs with small molecules having cytoprotective properties can enhance the potential of these cells for their use in cell-based therapeutics.

AIM

To enhance the therapeutic potential of MSCs by preconditioning them with isorhamnetin for second degree burn wounds in rats.

METHODS

Human umbilical cord MSCs (hU-MSCs) were isolated and characterized by surface markers, CD105, vimentin and CD90. For preconditioning, hU-MSCs were treated with isorhamnetin after selection of the optimized concentration (5 $\mu\text{mol/L}$) by cytotoxicity analysis. The migration potential of these MSCs was analyzed by the *in vitro* scratch assay. The healing potential of normal, and preconditioned hU-MSCs was compared by transplanting these MSCs in a rat model of a second degree burn wound. Normal, and preconditioned MSCs (IH + MSCs) were transplanted after 72 h of burn injury and observed for 2 wk. Histological and gene expression analyses were performed on day 7 and 14 after cell transplantation to determine complete wound healing.

RESULTS

The scratch assay analysis showed a significant reduction in the scratch area in the case of IH + MSCs compared to the normal untreated MSCs at 24 h, while

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complete closure of the scratch area was observed at 48 h. Histological analysis showed reduced inflammation, completely remodeled epidermis and dermis without scar formation and regeneration of hair follicles in the group that received IH + MSCs. Gene expression analysis was time dependent and more pronounced in the case of IH + MSCs. Interleukin (IL)-1 β , IL-6 and Bcl-2 associated X genes showed significant downregulation, while transforming growth factor β , vascular endothelial growth factor, Bcl-2 and matrix metalloproteinase 9 showed significant upregulation compared to the burn wound, showing increased angiogenesis and reduced inflammation and apoptosis.

CONCLUSION

Preconditioning of hU-MSCs with isorhamnetin decreases wound progression by reducing inflammation, and improving tissue architecture and wound healing. The study outcome is expected to lead to an improved cell-based therapeutic approach for burn wounds.

Key Words: Burn wound; Isorhamnetin; Mesenchymal stem cells; Preconditioning; Wound healing; Small molecule

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Core Tip: In this study, we propose an improved cell-based therapeutic approach using a cytoprotective chemical compound, isorhamnetin to precondition human umbilical cord mesenchymal stem cells (MSCs) for second degree burn wounds. The findings of this study suggest that transplantation of preconditioned MSCs in the rat burn wound model decreases wound progression by the downregulation of inflammatory cytokines and restoration of tissue architecture. The study outcome may lead to an effective cell-based treatment strategy for burn wounds to accelerate wound healing and promote skin regeneration.

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INTRODUCTION

Burn wounds caused by thermal injuries are the most common and devastating injuries, constituting approximately 86% of the burn victims admitted to hospitals. Burn injuries cause high morbidity and mortality with approximately 300000 deaths per year worldwide due to multiple organ dysfunction and abnormalities^[1-4]. These injuries occur by exposure to intolerable heat, particularly hot surfaces, liquids, steam and flames^[5,6]. Burn injuries are classified on the basis of severity and depth of the wound. In the case of superficial burns, heat penetrates only the epidermis and causes skin redness with slight inflammation. Partial thickness burns involve the epidermis and part of the dermis, causing pain and hypersensitivity. If, however, the burn completely damages the dermis, it causes red and white coloration of the tissue and reduces tissue sensitivity. Full thickness burns damage the subcutaneous layer causing white coloration of the tissue with no sensitivity. To improve burn prognosis, efforts are being made to use improved cellular therapy and tissue engineering approaches using biological skin replacements^[7,8].

The process of wound healing is dynamic and involves regeneration of skin tissue after injury. The human body functions in a manner that any disruption to the first line of defense *i.e.*, skin immediately activates a cascade of orchestrated events with three main phases: inflammation, proliferation and remodeling. The inflammatory phase begins as the cytokines and chemokines reinforce different cell types at the wound site by stimulating a signaling cascade, initiating cell migration and proliferation as well as

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elimination of cell debris^[9]. Proliferation serves as the crucial phase of wound healing and occurs as the inflammation subsides. It promotes healing by inducing proliferation of various cell types, including those of the epidermal and endothelial layers and of the connective tissue^[10]. It further leads to the formation of granulation tissue also termed fibroplasia which helps in the reorganization of the dermal matrix^[11]. The final stage of wound healing termed tissue remodeling is responsible for a dynamic change in the tissue framework that continues for a long period as the cells develop matrix structure, which sometimes results in scar formation as the poorly oriented collagen is extensively constituted; however, the wound seems to be healed^[12,13].

Impaired wounds and their aberrant healing are a major healthcare issue worldwide and a liability in patients and their families. Current treatment options used in clinics such as skin grafting are not very efficient and often do not completely promote wound closure. Efforts are being made to develop skin substitutes through bioengineering which is quite costly. Cell therapy has emerged as a promising strategy, with the ability to repair the tissue to the level of its original architecture^[14]. Stem cells have the remarkable potential to differentiate into any cell type which makes them an attractive choice for the treatment of degenerative diseases^[15]. Mesenchymal stem cells (MSCs) represent adult stem cells that belong to the population of non-hematopoietic stem cells originating from the mesodermal germ layer. These are multipotent cells with the ability to differentiate into multiple lineages. MSCs enhance wound healing by moderating the immune response and increasing angiogenesis^[16-18]. Human umbilical cord tissue (Wharton's jelly) serves as a potent and rich source of MSCs. However, these cells can also be isolated from bone marrow, adipose tissues, dental pulp *etc.*, and can be easily expanded in culture^[19-21]. A major obstacle in advancing stem cell therapy into successful clinical trials is their poor survival in the inflammatory environment of the impaired tissue, which reduces the efficiency of these cells^[22]. Therefore, preconditioning strategies can be adopted by treating stem cells with small molecules having cytoprotective properties.

For preconditioning of stem cells, bioactive components of plants that have been traditionally used for the treatment of wounds could be beneficial. Various medicinal plants and plant products have been used in traditional medicine for healing of wounds^[22,23]. The shrub *Hippophae rhamnoides* L commonly known as Sea buckthorn (SBT) which belongs to the family *Elaeagnaceae* has been extensively used for the treatment of skin diseases, asthma and lung disorders. This plant is a good source of a number of bioactive compounds like flavonoids such as quercetin, isorhamnetin, kaempferol *etc.*^[24,25]. A study conducted on the aqueous extract of SBT showed enhanced wound healing due to a significant increase in wound contraction^[26]. Due to its significant medicinal properties and beneficial role in wound healing, we selected isorhamnetin for our study. Isorhamnetin is a methylated derivative of quercetin, known for its anti-microbial and anti-inflammatory properties^[27-29]. It has been used for the treatment of cutaneous wound healing^[30,31]. We propose that this compound could enhance the migration efficiency and survival of MSCs at the wound site and increase their therapeutic potential for effective wound healing.

MATERIALS AND METHODS

Cell culture

Human umbilical cord samples were obtained from Zainab Panjwani Memorial Hospital after approval of the Independent Ethics Committee of International Center for Chemical and Biological Sciences (IEC, ICCBS) with formal consent from the donor. Cord samples (8-10 cm) were collected in sterile phosphate-buffered saline (PBS) and processed in the tissue culture facility using aseptic conditions. Cord samples were dissected and cut into small pieces of 1-3 mm in size. The explants were then cultured in DMEM supplemented with 10% FBS, 100 U penicillin/streptomycin, 1 mmol/L sodium pyruvate and 4 mmol/L L-glutamine and incubated in a CO₂ incubator at 37°C. The medium was changed every third day to remove non-adherent cells. After 10-15 d of culture, the cells were released from the explants and adhered to the flask surface. The cells at this stage were considered passage zero (P0) cells. Once the cells achieved 60%-80% confluence, they were subcultured by adding 1× trypsin-EDTA (0.25%). The cells were centrifuged for 8 min at 1000 rpm and the cell pellet was suspended in fresh DMEM and cultured in two flasks and incubated in a CO₂ incubator at 37°C. These cultures were designated as passage 1 (P1) cells. Further subculturing of the cells was performed in the same manner. All experiments were conducted using P3-P5 cells.

Characterization of human umbilical cord MSCs by immunocytochemistry

Human umbilical cord MSCs (hU-MSCs) were characterized by immunocytochemistry to determine specific surface markers. Approximately, 10000 cells per well were seeded in a 24-well plate and incubated in a CO₂ incubator. Once the cells adhered, the medium was discarded and cells were washed with 1× PBS. The cells were fixed using 4% paraformaldehyde for 10 min. To prevent non-specific binding, blocking solution was added and the cells were incubated for 1 h. Specific monoclonal primary antibodies against CD105, vimentin and CD90 (1:100 dilution) were then added, respectively, and the cells were left overnight at 4°C. Alexa Fluor 546 goat anti-mouse IgG (1:200 dilution) was added for 60 min at 37°C. DAPI solution (0.5 µg/mL) was added for 15 min at 37°C to stain cell nuclei and the cells were observed under a fluorescence microscope (TE2000-S, Nikon, Japan).

Characterization of hU-MSCs by trilineage differentiation

hU-MSCs were characterized for trilineage differentiation as chondrogenic, osteogenic and adipogenic lineages. Approximately, 40000 cells were seeded per well in a 6 well plate and incubated at 37°C in a CO₂ incubator for 24 h. Once the cells achieved 60%-70% confluence, they were washed with 1× PBS and sterile chondro-induction medium was added to each well and the plate was incubated again under the same conditions. The medium was changed every 4th d for approximately 21 d. After 21 d, the induction medium was removed followed by cell fixation with 4% PFA for 15 min. After fixation, the cells were stained with 1% Alcian Blue stain for 60 min followed by washing with 1× PBS. Osteogenic and adipogenic characterization was performed using a similar methodology. Staining was performed using 25% Alizarin Red stain for 50 min and Oil Red O stain for 60 min, respectively. Stained cells were observed under a phase contrast microscope.

Preconditioning of hU-MSCs

For preconditioning, hU-MSCs were treated with isorhamnetin obtained commercially (Leap Chem, China). Calculated volumes for a range of concentrations (2-100 µmol/L) were used to determine the least cytotoxic concentration. The cytotoxicity analysis was performed using the MTT colorimetric assay according to manufacturer's instructions. The optimized concentration (5 µmol/L) of isorhamnetin obtained from the cytotoxicity analysis was used in all subsequent experiments.

Scratch assay

Cell migration potential was analyzed by performing the scratch assay to determine the effect of preconditioning. A T-25 culture flask was seeded with 3 × 10⁶ cells/mL and treated with the optimized concentration (5 µmol/L) of isorhamnetin once the cells attained 80% confluence. After 24 h of treatment, the medium was discarded and the monolayer of cells was gently scratched in a straight line using a 200 µL sterile tip. Cell migration was evaluated by determining the distance covered by the cells, migrating towards the scratch area at time intervals of 24 and 48 h.

Burn wound model

Adult Wistar rats of both sexes weighing 170-200 g were housed in the institutional animal resource facility and maintained at 21 ± 1°C with relative humidity of 55% ± 5% and 12 h light/dark cycle with ad libitum access to food and water. All experiments were performed in accordance with the international guidelines for the care and use of laboratory animals recommended by NIH (NIH Publication No. 85-23, revised 1996) and approved by the Institutional Animal Care and Use Committee (IACUC, animal study protocol number: 2018-0018). The *in vivo* burn wound model was developed following a protocol of consistent burn injury^[32]. Five animals were used for the development of the burn model. Intraperitoneal injections of 60 mg/kg of ketamine hydrochloride (50 mg/mL) and 7 mg/kg of xylazine hydrochloride (100 mg/mL) were given to the rats to induce anesthesia. Hair was removed from the dorsal surface of the skin at the wound site using depilatory cream. The skin was thoroughly cleaned and disinfected with an alcohol swab. A 1.5 cm diameter steel rod weighing 150 g was placed in a beaker containing water and heated on a hot plate until the temperature reached 100°C. The heated rod was kept on the stretched and pulled skin for 10 s without applying any external pressure to form a consistent burn wound. To minimize pain or discomfort, the rats were injected subcutaneously with diclofenac sodium (25 mg/mL) and streptomycin/penicillin (10000 U/mL). The animals were euthanized with a dose of 200 mg/kg body weight of sodium pentobarbital for tissue harvesting.

Transplantation of normal and preconditioned MSCs

Cells were prepared for transplantation after 24 h of treatment while untreated cells were used as the control. Cells were trypsinized and the pellet was suspended in 1× PBS. Localized transplantation of one million cells using a sterile insulin syringe was performed at the wound periphery with multiple injections at different sites after 72 h of burn infliction. Five animals were used for each transplantation group.

Macroscopic visualization

Control and treated burn wounds were macroscopically visualized for rate of skin contraction and detachment of the scab. Images of control and treated burns were captured over a period of 2 wk at defined time points of day 1, 3, 7 and 14 with reference to phases of wound healing.

Histological examination

Histological analysis was performed by harvesting tissue samples on day 1, 3, 7 and 14. About 6 µmol/L thin paraffin sections of embedded tissue blocks were collected on the slides, and were stained using hematoxylin-eosin stain for microscopic analysis.

RNA isolation and cDNA synthesis

For gene expression analysis, tissues were harvested and homogenized in the presence of the One Step-RNA reagent (Bio Basic, Canada). RNA was extracted according to the recommendations provided by the manufacturer. RNA was quantified by measuring the absorbance at 260 nm and the presence of protein contamination was determined by calculating the ratio of absorbance at 260/280 nm. cDNA was synthesized using the RevertAid First Strand cDNA synthesis kit (Fermentas, ThermoFisher Scientific, United States) according to the manufacturer's protocol.

Real-time polymerase chain reaction

Real-time polymerase chain reaction (qPCR) was performed to determine the changes in gene expression. The genes were selected corresponding to the cytokines involved at different phases of wound healing. Amplification was performed using qPCR (CFX96 Touch Real-Time PCR Detection System, Bio-Rad, United States) under the following conditions; denaturation at 95°C for 1 min, annealing at 58°C for 1 min (40 cycles) and extension at 72°C for 1 min. GAPDH was taken as an internal control. The genes analyzed are presented in [Table 1](#).

Statistical analysis

The statistical analysis was performed by One-way ANOVA with the Bonferroni post-hoc test using IBM SPSS Statistics 21. All values were represented as mean ± SE with the number of observations (*n*) as 3-5 and level of confidence as ^a*P* < 0.05; where ^a*P* < 0.05, ^b*P* < 0.01 and ^c*P* < 0.001.

RESULTS

Isolation and characterization of hU-MSCs

Cells released from the explant were termed as P0, which proliferated up to 70% confluence, and formed a monolayer by the 12th d. Later, the cells were subcultured to P1 and left for 12 d to achieve the required confluence for further expansion ([Figure 1](#)). Isolated cells were positive for CD105, vimentin and CD90 surface markers. This confirmed the single and homogenous population of hU-MSCs ([Figure 2A](#)). In the case of trilineage differentiation, cells grown in the selective medium showed mineral deposits after Alizarin Red staining, prominent oil droplets after Oil Red O staining and proteoglycans after Alcian Blue staining, confirming that the cells differentiated into osteocytes, adipocytes and chondrocytes, respectively ([Figure 2B](#)).

Preconditioning of MSCs

The cytotoxicity analysis showed that the percentage of cell survival decreased with increasing concentration of the compound. Less than 5 µmol/L of isorhamnetin was confirmed as non-cytotoxic ([Figure 3A](#)). For preconditioning, hU-MSCs were treated with 5 µmol/L concentration for 24 h. Isorhamnetin-treated MSCs are referred to as preconditioned MSCs (IH + MSCs) in subsequent sections.

Table 1 Gene primer sequences used in this study with their annealing temperatures and product sizes

No.	Genes	Primer sequence (5'-3')	Annealing temperatures (°C)	Product sizes (bp)
1	Interleukin 1 beta (<i>IL-1β</i>)	F: TCATCTTTGAAGAAGAGCCCGT R: GTTCTGTCCATTGAGGTGGAGA	60	170
2	Interleukin 6 (<i>IL-6</i>)	F: GATGGATGCTTCCAAACTGGATA R: TGAATGACTCTGGCTTTGCTTT	58	140
3	Transforming growth factor (<i>TGF-β</i>)	F: TCGCCAGTCCCCGA R: TCGGGCTCCGGGTCA	60	101
4	Vascular endothelial growth factor (<i>VEGF</i>)	F: CCAATTGAGACCCTGGTGGA R: TCCTATGTGCTGGCTTTGGT	60	195
5	Tumor necrosis factor (<i>TNF-α</i>)	F: CCTCTTCTCATTCTGCTCGT R: GATCTGAGTGTGAGGGTCTGG	60	144
6	Bcl-2 associated X (<i>Bax</i>)	F: CTTCTCCGTGTGGCAGC R: CAGACAAACAGCCGCTCC	58	172
7	Apoptosis regulator (<i>Bcl-2</i>)	F: ACCCTGGCATCTTCTCC R: AGAAGTCATCCCCAGCCC	59	172
8	Matrix metalloproteinase 9 (<i>MMP-9</i>)	F: TACCAGCTACTCGAACCAATCA R: AAATAAAAAGGGCCGGTAAGGTG	59	187

F: Forward; R: Reverse.

Scratch assay

Quantitative analysis of the scratch test showed that IH + MSCs migrated towards the scratch better than normal MSCs. The scratch area in IH + MSCs was significantly reduced compared to normal MSCs at 24 h and complete closure of the scratch was observed at 48 h (Figure 3B).

Macroscopic visualization

All groups were macroscopically observed at time points corresponding to the phases of wound healing that is day 1, 4, 7 and 14. Detachment of the scab indicates initiation of wound healing. In the case of the burn wound, the scab was not detached until day 14. Burn wound treated with normal MSCs showed scab detachment at day 14. The wound area was also reduced as compared to the burn wound at the same time point. The IH + MSCs treated burn wound showed scab detachment as well as re-epithelialization of the wound at day 14. Slight hair growth was also observed indicating hair follicle regeneration with complete healing of the wound (Figure 4).

Histological analysis

The histology of burn wound after day 1 of burn injury showed that both the epidermis and dermis were damaged due to inflammation with distorted collagen matrix and dermis. Hair follicles were also inflamed with increased infiltration of inflammatory cells which confirmed second degree burn. At day 7 and 14, angiogenesis and scab formation were observed which indicated progression towards healing but with a deformed tissue framework (Figure 5B). In comparison to the burn control, histological analysis of the MSCs treated wound showed increased angiogenesis and reduced inflammation of the hair follicles and initiation of epidermis formation at day 7. Fourteen days after transplantation, reduced angiogenesis, formation of epidermis and dermis with extracellular matrix deposition and infiltration of inflammatory cells were observed. The IH + MSCs treated burn wound showed significant healing and remodeling without scar formation as compared to the burn control and normal MSCs treated burn wound. At day 7, epidermis formation was observed along with increased angiogenesis and reduced inflammation. It was observed that the dermis formation and collagen deposition were similar to the natural tissue framework. Later, at day 14, complete re-epithelialization was observed with intact collagen, healthy blood vessel formation and completely remodeled tissue

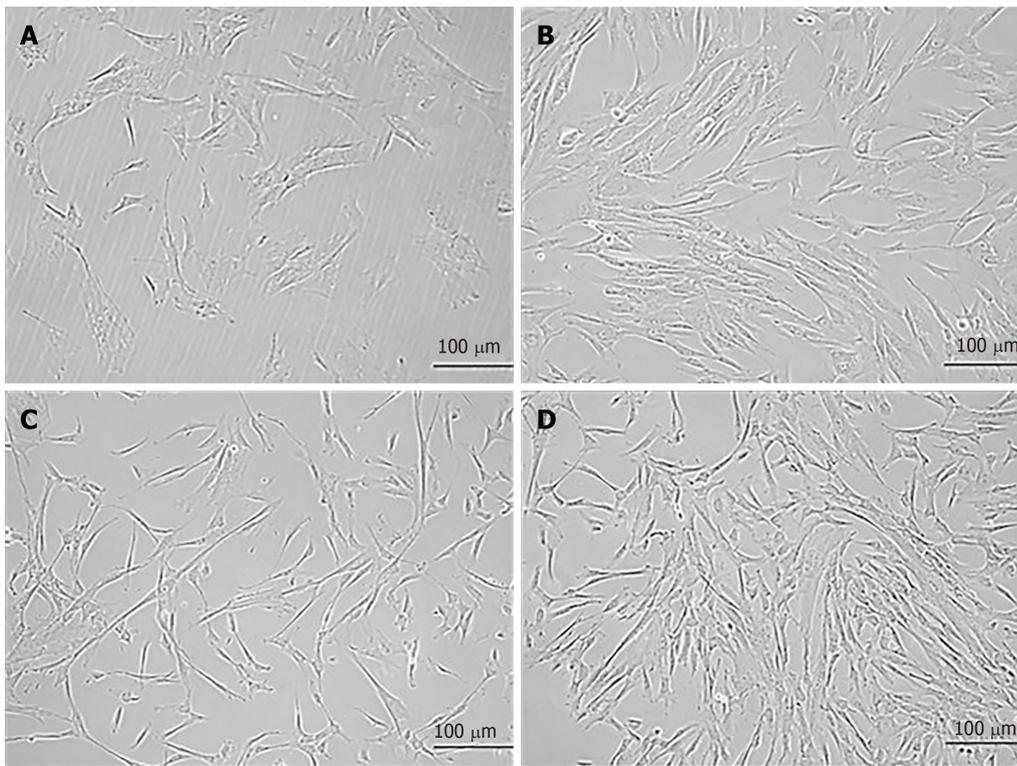


Figure 1 Morphology of mesenchymal stem cells at different passages. Phase contrast images (20 ×) of human umbilical cord-derived mesenchymal stem cells show fibroblast-like morphology at passage 0 (P0) at day 10 and day 12 of culture. Similarly, cells at passage 1 (P1) show fibroblast-like morphology with extensions at day 10 and day 12 following subculture. A: P0 at day 10; B: P0 at day 12; C: P1 at day 10; D: P1 at day 12. Confluent cells are seen in B and D.

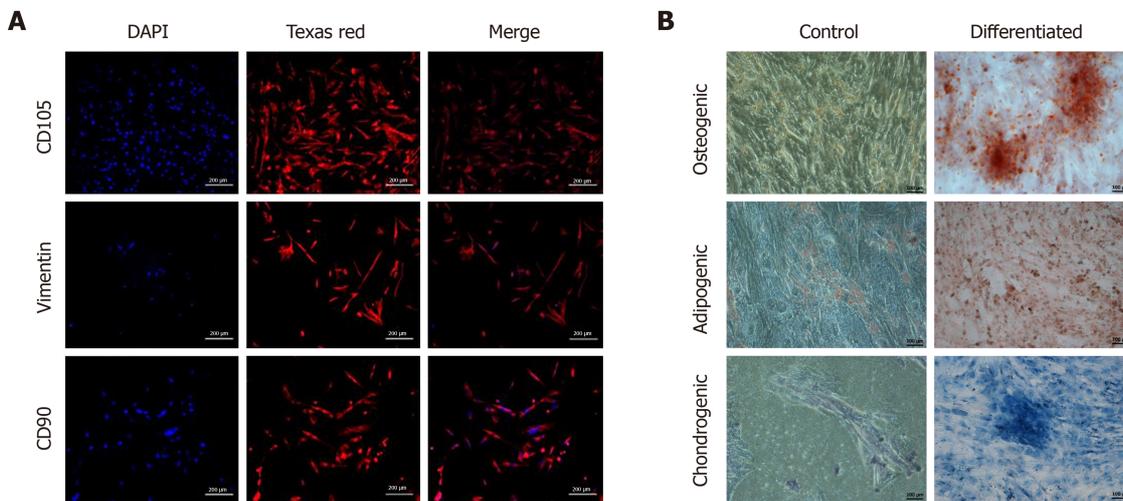


Figure 2 Characterization of mesenchymal stem cells. A: Fluorescent images (10 ×) of isolated mesenchymal stem cells (MSCs) characterized by immunocytochemistry showing positive expression of cell surface markers CD105, vimentin and CD90. Alexa Fluor 546 goat anti-mouse secondary antibody was used for detection. Nuclei were stained with DAPI; B: Phase contrast images (10 ×) of MSCs stained with Alizarin Red stain showing mineral deposits, Oil Red O stain showing oil droplets, and Alcian Blue stain showing proteoglycans, confirming trilineage (osteogenic, adipogenic and chondrogenic) differentiation of MSCs grown in specific differentiation medium, as compared to the control.

framework with no sign of inflammation and scar formation (Figure 5C).

Gene expression analysis

The gene expression analysis of the burn control, normal MSCs, and IH + MSCs treated burn wound showed significant modulation which was time dependent in some cases (Figure 6). The inflammatory genes interleukin (IL)-1 β and IL-6 were significantly downregulated in the case of both normal MSCs and IH + MSCs treated burn wounds as compared to the burn control, but this decrease was observed at day

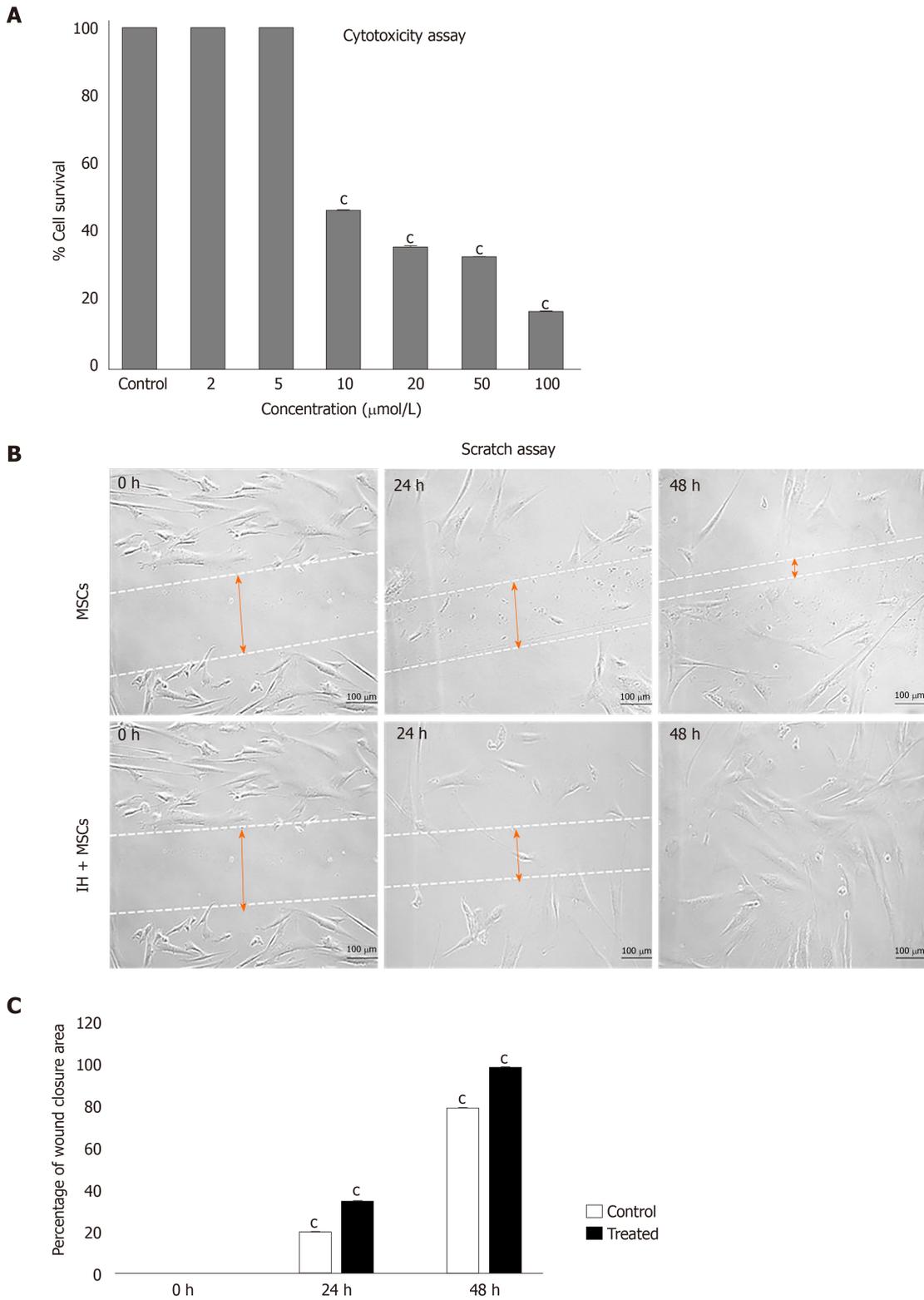


Figure 3 Cytotoxicity analysis and scratch assay. A: MTT assay for cytotoxicity analysis showed a concentration dependent decrease in the viability of human umbilical cord mesenchymal stem cells (MSCs) treated with isorhamnetin, with a significant reduction in the percentage of cell survival at concentrations above 5 µmol/L; B: Scratch assay shows cell migration of normal untreated MSCs and preconditioned MSCs (IH + MSCs) at 0, 24 and 48 h. The scratch area was reduced in IH + MSCs as compared to untreated MSCs after 24 h and complete migration was observed after 48 h. The quantitative analysis showed significant cell migration of isorhamnetin treated MSCs at 24 h and complete migration at 48 h as compared to untreated normal MSCs. For statistical analysis, One way ANOVA was performed, followed by the Bonferroni post hoc test; Data are presented as the mean ± SE (n = 4). *P < 0.001. MSCs: Mesenchymal stem cells; IH + MSCs: Preconditioned mesenchymal stem cells.

14 in the case of IL-1 β . The anti-inflammatory gene, transforming growth factor β (TGF- β) showed significant upregulation in the case of normal MSCs at day 7, and at both time points in the case of IH + MSCs. The angiogenic gene, vascular endothelial growth factor (VEGF) was also upregulated in both treatment groups as compared to the burn control at day 7 and its expression was reduced at day 14. Apoptosis was reduced as shown by the decreased levels of Bcl-2 associated X (Bax) gene in both treatment groups, while anti-apoptotic Bcl-2 gene was upregulated in the case of IH + MSCs at day 7. Matrix metalloproteinase 9 (MMP-9) levels were found to be significantly upregulated in the normal MSCs and IH + MSCs treated burn wounds as compared to the burn control which showed improved tissue architecture.

DISCUSSION

In this study, we used a small molecule, isorhamnetin (a flavonoid), to precondition hU-MSCs, in order to improve the survival and migration potential of MSCs. This could lead to complete wound healing without scar formation, thus enhancing the therapeutic potential of these MSCs. Migration potential corresponds to the rate of wound healing. In the *in vitro* scratch assay, we observed that the preconditioned MSCs showed significant migration potential as compared to normal MSCs after 24 and 48 h. These preconditioned cells migrated towards the scratch resulting in the complete closure of the scratch compared to the normal MSCs. This confirms the improved migration potential of preconditioned MSCs necessary for rapid wound healing.

To analyze the effect of preconditioning *in vivo*, we developed rat burn wound model with a consistent second degree burn. Macroscopic analysis after burn injury showed severe edema with a pale appearance on the burn site. After 24 h, the wound appeared yellowish brown with a well-defined margin of erythema and no signs of blistering, which is in agreement with previous studies^[33,34]. The wound progression led to a thick scab which completely detached at day 21, leaving behind granulation tissue. This indicates the initiation of wound healing through angiogenesis which is responsible for restoring skin structure at the epidermal level^[35]. Macroscopic observation of the wound tissue transplanted with normal MSCs and IH + MSCs showed no apparent change in the wound morphology compared to the burn wound at day 7. Detachment of the scab was observed at day 14 in both groups. In addition, the formation of granulation tissue was observed in normal MSCs, while complete wound closure and re-epithelialization with no granulation tissue and scar formation was seen in the case of IH+MSCs.

Histological analysis of the wound tissues further confirmed our macroscopic findings. Inflammation of the epidermis after 24 h corresponded to the edema and erythema observed macroscopically. Slow progress of the burn wound is indicated by an increase in loose connective tissues and infiltration of lymphocytes and plasma cells after 72 h. Disrupted epidermis and dermis with extreme inflammation and conservation of hypodermis persisted even after 1 wk (day 7), which is in agreement with previous studies^[36,37]. Microscopic analysis of the burn wound at day 14 revealed adequate tissue autolysis with distinct neovascularization, formation of granulation tissue and moderate fibrosis with no signs of remodeling. Normal MSCs and IH + MSCs treated wound tissues revealed significant improvement in wound healing as compared to the burn control. Normal MSCs treated wounds at day 7 showed increased angiogenesis, regeneration of epidermis, and extensive infiltration of inflammatory cells along the epidermal layer and within the lumen of newly formed vessels. Significant fibroblast proliferation was also observed forming granulation tissue under the scab. In contrast, IH + MSCs treated wound tissues showed reduced inflammation, increased neovascularization and granulation tissue formation at day 7, forming the dermis architecture. At day 14, complete scab detachment and regeneration of epidermis along with a keratin layer was observed. Initiation of dermis formation with collagen deposition and an increase in fibroblast proliferation indicates the presence of granulation tissue^[38]. Vascularization was decreased at this stage by stimulating vessel regression. This may progress towards remodeling^[39]. Moreover, the IH + MSCs treated burn wound showed re-epithelialization and remodeling without scar formation, which also supports our macroscopic data.

Modulation of gene expression was time dependent in some cases. Burn wound tissue transplanted with normal, and preconditioned MSCs showed significantly reduced gene expression of both the inflammatory genes, IL-1 β and IL-6, which confirmed the anti-inflammatory effect of the compound particularly at day 14. It is

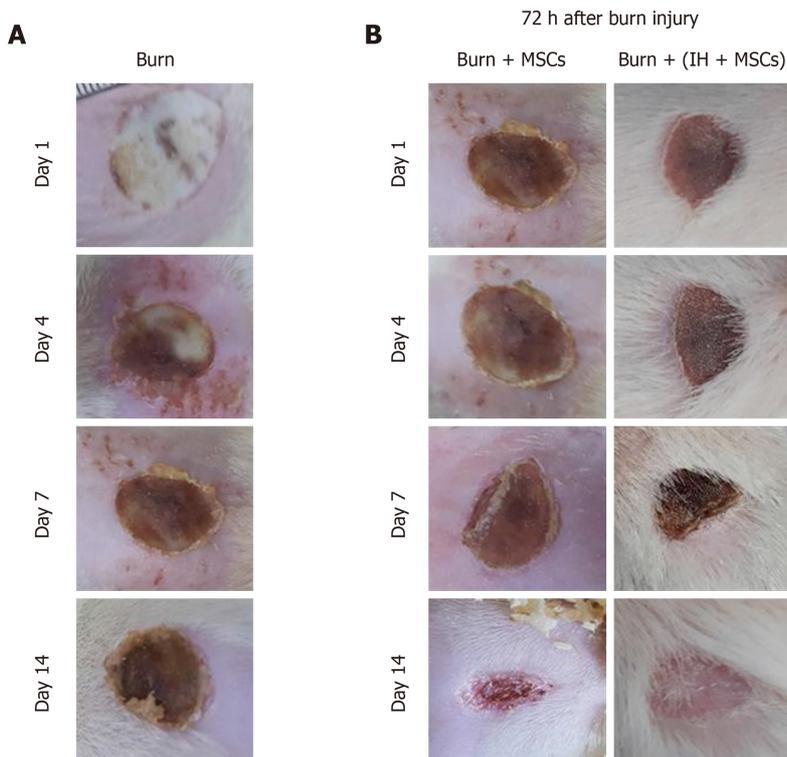


Figure 4 Macroscopic examination of burn wounds. A: Images of burn wound tissue visualized macroscopically at days 1, 4, 7 and 14. Scab formation was observed at day 4 and slight detachment of the scab at day 14; B: Burn wound tissues transplanted with normal mesenchymal stem cells (MSCs) and preconditioned MSCs (IH + MSCs) were visualized 72 h after burn injury *i.e.* at day 1, 4, 7 and 14 following transplantation. At day 4, wound progression and initiation of scab formation were observed in all transplantation groups. At day 7, a thick scab with slight detachment was observed which completely covered the burn site and no sign of infection in the group transplanted with IH + MSCs. At day 14, complete detachment of the scab with slight granulation tissue was observed in the group transplanted with normal MSCs, while complete scab detachment and re-epithelialization were observed along with hair growth in the case of IH + MSCs. MSCs: Mesenchymal stem cells; IH + MSCs: Preconditioned mesenchymal stem cells.

also evident in the case of our histological analysis of the corresponding wound tissues. The expression of TGF- β , an anti-inflammatory cytokine, in the wound bed of both the normal MSCs and IH + MSCs treated groups showed increased levels at day 7, and the level persisted until day 14 in the latter group. It was shown in an earlier study that TGF- β promotes angiogenesis by stimulating the release of IL-1 β and tumor necrosis factor- α ^[40]. As TGF- β expression is essential for dermal wound healing, its increased expression can lead to scar formation following fibrosis^[41]. The expression of VEGF after transplantation of normal MSCs and IH + MSCs was found to be significantly elevated at day 7, which correlated with our histological findings which showed increased angiogenesis. These results are consistent with an earlier study which showed that the time point of VEGF expression is crucial for the progression of wound healing; during proliferation which lasts for about 3-7 d, neovascularization is at its maximum and the VEGF level increases^[42-44]. At day 14, we observed that VEGF expression was reduced in both treatment groups. This provides evidence of tissue maturation and remodeling^[45]. The expression of apoptotic protein Bax was significantly reduced after transplantation of normal MSCs and IH + MSCs at days 7 and 14. Bcl-2, an anti-apoptotic protein, was increased at day 7; however, at day 14, it was downregulated. This is supported by our histological analysis which showed that the tissue framework was regenerated at day 14, leading to re-epithelialization and no scar formation during remodeling. With regard to MMP-9 expression, the IH + MSCs group showed significant upregulation of its expression. MMP-9 is observed in several damaged epithelia including gut and eye along with the skin during wound healing^[46,47]. The elevated expression of MMP-9 suggested an increase in re-epithelialization leading to restoration of the tissue framework similar to that of uninjured tissue.

In burn wounds, inflammation is the major contributor to tissue injury and pathological remodeling. One of the reasons for using isorhamnetin is to reduce the inflammation in burn tissue in combination with MSCs. Previously, it was reported that isorhamnetin possesses anti-inflammatory properties^[48]. It has also been reported to have anti-oxidant properties. During burn wounds, inflammatory cells, neutrophils

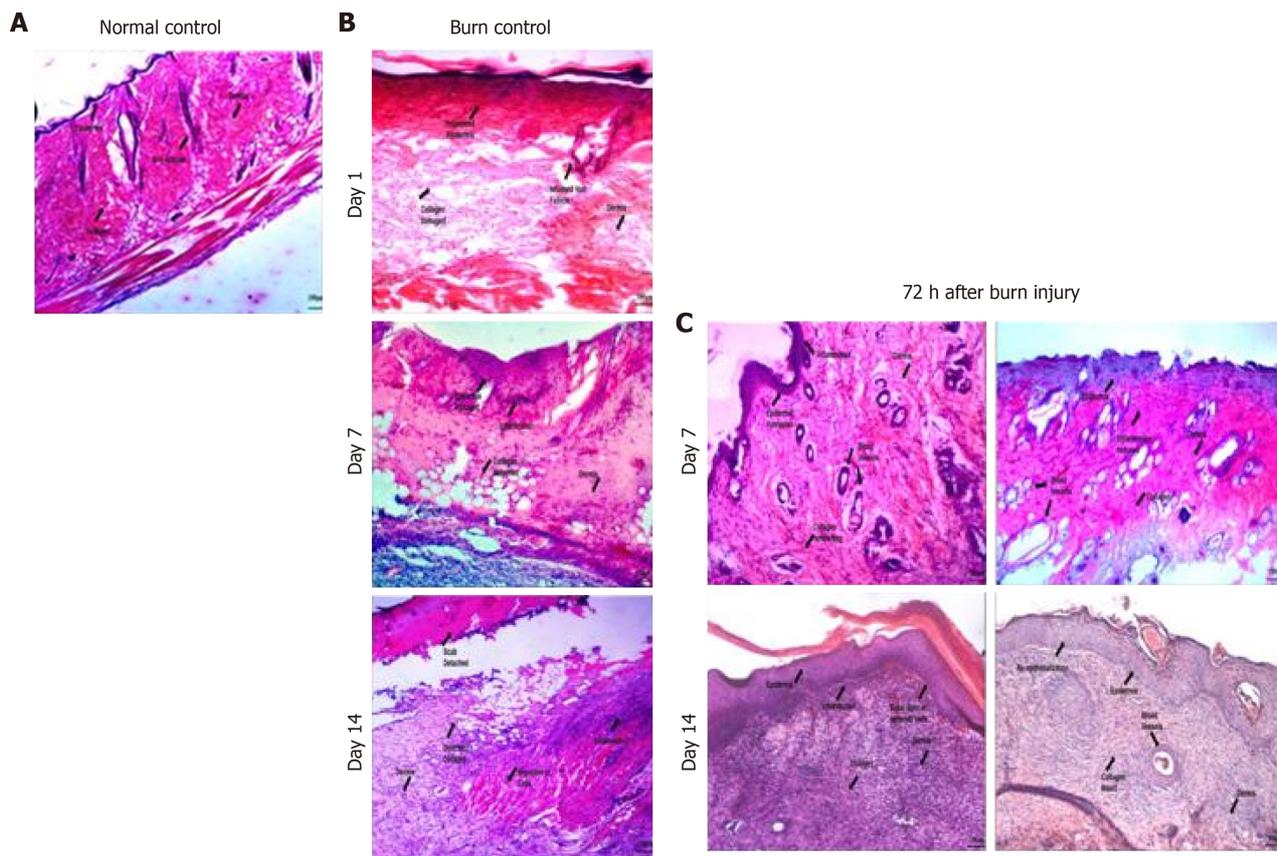


Figure 5 Histological analysis of normal and burn wound tissues. Phase contrast images (20 ×) of histological sections stained with hematoxylin-eosin stain showing normal control (skin with no wound) with normal skin structure composed of the epidermis, dermis and hypodermis along with the muscles, burn wound control at days 1, 7 and 14, and burn wound tissues transplanted with normal mesenchymal stem cells (MSCs) and preconditioned MSCs (IH + MSCs) at days 7 and 14 following 72 h of burn injury. The burn wound at day 1 shows inflamed epidermis and hair follicles, completely distorted dermis and collagen matrix; at day 7, disorganized tissue framework with no distinct layers of skin and the presence of loose connective tissues, while hair follicles show better morphology and reduced inflammation; and at day 14, slight detachment of the scab with deformed collagen and cellular infiltration, loose connective tissue with disorganized dermis and absence of epidermis. Normal MSCs treated burn wound at day 7 shows neovascularization and inflamed epidermis along with collagen deposition, while hair follicles show inflammatory cell infiltration in the periphery with better morphology; and at day 14, epidermis formation with cellular migration and collagen deposition, constructing the dermis. IH + MSCs treated burn wound at day 7 shows increased angiogenesis and reduced inflammation in the epidermis, while hair follicles also show similar morphology to that in normal skin with reduced inflammation; and at day 14, complete re-epithelialization with intact collagen and vascularization, and remodeling of the tissue framework with a distinct layer of epidermis and dermis with no signs of scar. A: Normal control; B: Burn control; C: Seventy two hours (72 h) after burn injury.

and macrophages concentrate at the site of injury, and reactive oxygen species are generated increasing oxidative stress. With its anti-oxidant properties, isorhamnetin has been shown to inhibit hydrogen peroxide-induced reactive oxygen species production^[49]. Results obtained from our *in vivo* experiments correlated well with the anti-inflammatory potential of isorhamnetin, which led to the efficient preconditioning of MSCs for improved wound healing.

CONCLUSION

It is concluded that preconditioning of human umbilical cord-derived MSCs with the anti-inflammatory compound, isorhamnetin, heals the burn wound causing re-epithelialization and remodeling without scar formation. The skin tissue framework was repaired to the level of normal tissue with regeneration of the hair follicles. Preconditioning reduces the inflammatory reaction after burn injury, which not only decreases wound progression by downregulating inflammatory cytokines but also leads to restoring tissue architecture by maintaining the balance in the expression of cytokines in the respective phases of wound healing.

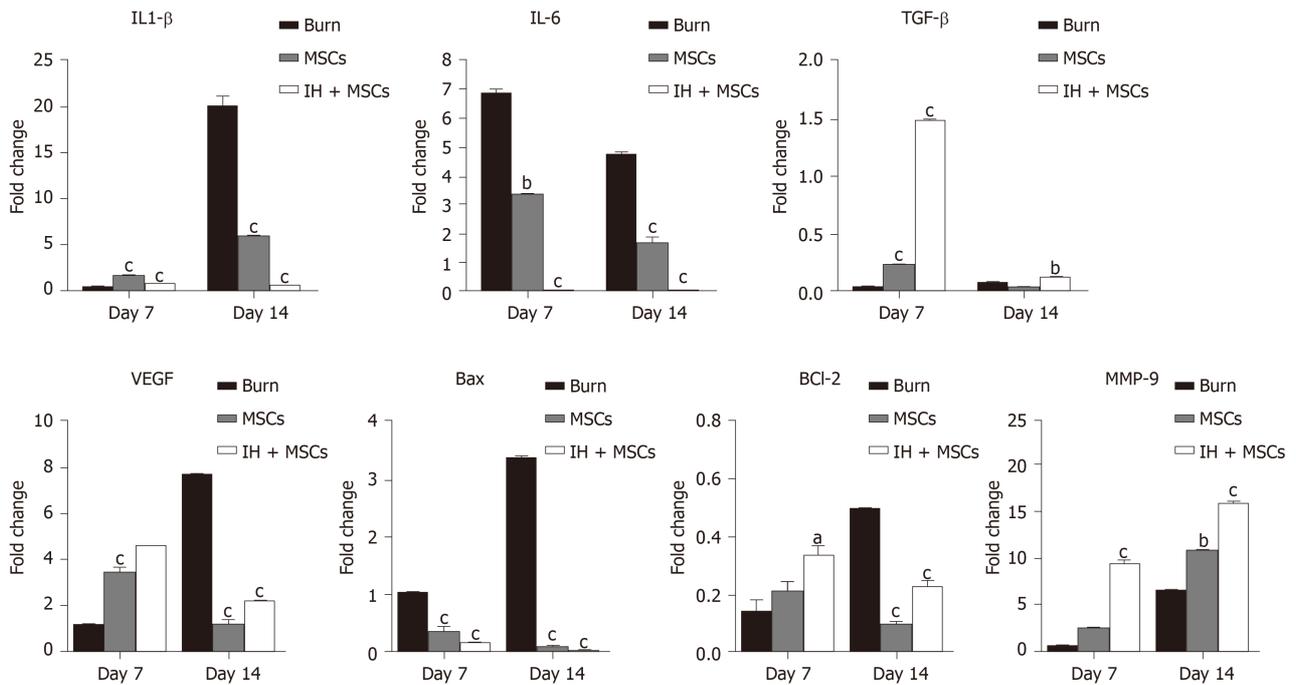


Figure 6 Gene expression analysis of burn wounds. Bar diagrams corresponding to gene expression analysis of interleukin (IL)-1 β , IL-6, transforming growth factor β (TGF- β), vascular endothelial growth factor (VEGF), Bcl-2 associated X (Bax), Bcl-2 and matrix metalloproteinase 9 (MMP-9) are shown in the burn wound control (at days 7 and 14) and experimental groups treated with normal untreated mesenchymal stem cells (MSCs) and preconditioned MSCs (IH + MSCs) at days 7 and 14 following 72 h of burn injury. The pattern of gene expression was time dependent; statistically significant downregulation of IL-1 β (at day 14), IL-6 and Bax (days 7 and 14), and significant upregulation of TGF- β and MMP-9 (days 7 and 14), VEGF and Bcl-2 (day 7) in case of IH + MSCs as compared to burn control. Same pattern was observed in the case of normal MSCs for *IL-1 β* , *IL-6*, *Bax*, *VEGF* and *Bcl-2* genes, while for TGF- β (day 14) and MMP-9 (day 7), the expression levels were non-significant. For statistical analysis, One way ANOVA was performed, followed by the Bonferroni post hoc test. Data are presented as the mean \pm SE ($n = 3$). ^a $P < 0.05$, ^b $P < 0.01$ and ^c $P < 0.001$. IL: Interleukin; TGF- β : Transforming growth factor β ; VEGF: Vascular endothelial growth factor; MMP-9: Matrix metalloproteinase-9.

ARTICLE HIGHLIGHTS

Research background

Impaired wound healing can be associated with different pathological states. Burn wounds are the most common and detrimental injuries that remain a major health issue worldwide. Mesenchymal stem cells (MSCs) possess the ability to regenerate tissues by secreting factors involved in promoting cell migration, proliferation and differentiation, while suppressing immune reactions. Preconditioning of MSCs with small molecules having cytoprotective properties can enhance the potential of these cells for their use in cell-based therapeutics.

Research motivation

Mesenchymal stem cells are a promising source for cell-based treatment to achieve skin regeneration. Preconditioning approaches using treatment with cytoprotective agents can lead to improvement in the wound healing potential of these stem cells and would be a crucial step in defining efficient therapy following burn injuries.

Research objectives

The objective of this study was to enhance the therapeutic potential of human umbilical cord MSCs (hU-MSCs) by preconditioning them with the cytoprotective flavonoid compound, isorhamnetin in a second degree burn wound model.

Research methods

hU-MSCs were isolated and characterized by specific surface markers. hU-MSCs were treated with isorhamnetin to precondition these cells. The migration potential of MSCs was analyzed by the *in vitro* scratch assay, while the healing potential of normal, and preconditioned MSCs was compared by transplanting them into a rat model of a second degree burn wound 72 h after burn injury and observed for 2 wk. Histological and gene expression analyses were performed on day 7 and 14 after cell

transplantation to determine complete wound healing.

Research results

The *in vitro* scratch assay analysis showed that preconditioned hU-MSCs significantly reduced the scratch area at 24 h, and completely closed the scratch area at 48 h as compared to normal MSCs. The preconditioned MSCs showed reduced inflammation, remodeled epidermis and dermis without scar formation and regeneration of hair follicles as observed following histological analysis. Preconditioning of MSCs promoted angiogenesis and remodeling, and decreased apoptosis as observed by gene expression studies of the corresponding burn wounds.

Research conclusions

Preconditioning of hU-MSCs with isorhamnetin decreases wound progression by reducing inflammation, and improving tissue architecture and wound healing. The study outcome is expected to lead to an improved cell-based therapeutic approach for burn wounds.

Research perspectives

The current study will be useful for developing an effective treatment strategy for burn patients based on the synergistic effect of isorhamnetin and MSCs to accelerate wound healing with complete skin regeneration without scar formation.

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