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**Systemic adverse effects and toxicities associated with immunotherapy: A review**

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**Abstract**

Immunotherapy is rapidly evolving secondary to the advent of newer immunotherapeutic agents and increasing approval of the current agents by the United States Food and Drug Administration to treat a wide spectrum of cancers. Immunotherapeutic agents have gained immense popularity due to their tumor-specific action. Immunotherapy is slowly transforming into a separate therapeutic entity, and the fifth pillar of management for cancers alongside surgery, radiotherapy, chemotherapy, and targeted therapy. However, like any therapeutic entity it has its own adverse effects. With the increasing use of immunotherapeutic agents, it is vital for physicians to acquaint themselves with these adverse effects. The aim of this review is to investigate the common systemic adverse effects and toxicities associated with the use of different classes of immunotherapeutic agents. We provide an overview of potential adverse effects and toxicities associated with different classes of immunotherapeutic agents organized by organ systems, as well as an extensive discussion of the current recommendations for treatment and clinical trial data. As we continue to see increasing usage of these agents in clinical practice, it is vital for physicians to familiarize themselves with these effects.

**Key Words:** Immunotherapy; Cancer; Malignancy; Adverse effects; Immune checkpoint inhibitor; Monoclonal antibody

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**Core Tip:** While immunotherapy is widely becoming a popular therapeutic treatment modality for a broad range of malignancies, it also has been known to cause a number of adverse effects. These effects have been found to be prevalent in a large number of organ systems. Clinicians should be aware of the various adverse effects associated with these novel therapeutic agents, as well as recommendations for management of these side effects in order to facilitate improved outcomes for patients undergoing these therapies.

**INTRODUCTION**

Immunotherapy has become an increasingly popular therapeutic option for patients suffering from a variety of malignancies over the past several years. The development of these therapies began nearly two decades ago, when Leach *et al*[[1](#_ENREF_1" \o "Leach, 1996 #3)] discovered that inhibition of cytotoxic T-lymphocyte associated protein 4 (CTLA-4) *in vivo* led to the rejection of tumor cells. Furthermore, in 2004, Chen described the role of programmed cell death protein 1 (PD-1) and programmed death-ligand 1 (PD-L1) in the suppression of T-cell responses in the tumor microenvironment[[2](#_ENREF_2" \o "Chen, 2004 #4)]. These discoveries have sparked extensive research in the field of immunotherapy, ultimately leading to the approval of ipilimumab (CTLA-4 inhibitor) by the United States Food and Drug Administration (FDA) for the treatment of melanoma in 2011[[3](#_ENREF_3)]. Since then, immunotherapy has attracted immense attention as a therapeutic modality. The idea behind the utilization of these agents is based upon the premise of using the body’s own immune system to attack and destroy cancerous cells[[4](#_ENREF_4" \o "Li, 2018 #2)]. Currently, immunotherapeutic agents have been approved for a wide variety of malignancies, including but not limited to melanoma, non-small cell lung cancer, colorectal cancer, bladder cancer, breast cancer, and head and neck cancers[[5](#_ENREF_5" \o ",  #5)]. Although the success of these therapies has been well documented, the literature does report significant side effects closely associated with their use, known as immune related adverse effects (irAE). In this review, we provide a system-based overview of the documented adverse effects associated with the use of immunotherapy. It is essential for care providers to know these systemic adverse effects and toxicities when prescribing these agents for the treatment of cancers.

**CLASSES OF IMMUNOTHERAPY AGENTS AND MECHANISMS**

Immunotherapy works by stimulating the body’s own immune system to target cancerous cells. With a better understanding of the workings of the immune system and the identification of new molecular targets, the classes of agents continue to grow. Currently, immunotherapeutic agents can be classified into 5 major types including[[6](#_ENREF_6" \o "Yaguchi, 2016 #12),[7](#_ENREF_7)]: Immune checkpoint inhibitors (ICI); Adoptive cell therapy; Monoclonal antibodies; Treatment vaccines; Immune system modulators.

The National Institute of Health along with the approval of immunotherapeutic agents from the FDA have led to the recommendations for use of these agents in different types of cancers as summarized in Table 1. The mechanism of action of the 4 most frequently used immunotherapeutic agents is described as follows (see Table 2 for an overview of individual immunotherapeutic agents and their proposed mechanisms): (1) ICIs work through therapeutic targeting of the checkpoint molecules CTLA-4, PD-1, and PD-L1, which are normally utilized by healthy cells to promote self-tolerance and inhibit destruction by autoreactive T-cells[8]. However, tumor cells often utilize these proteins to avoid T-cell mediated destruction. By utilization of the ICIs, T-cells may have a greater ability to recognize tumor cells as foreign entities and promote cell lysis; (2) T-Cell transfer therapy is a treatment method in which a patient’s T cells are collected and modified in a laboratory setting in order to improve their ability to bind and kill cancer cells. By collecting activated T-cells from cancer tissue, or genetically engineering T-cells, transfer therapy allows for passive immunization against cancers[[9](#_ENREF_9)]; (3) monoclonal antibodies are engineered to be antigen specific, often tumor-specific, and mediate the destruction of tumor cells *via* antibody-dependent cell-mediated cytotoxicity, promotion of phagocytosis or complement activation (referred to as complement-dependent cytotoxicity), or vascular and stromal ablation. A multitude of monoclonal antibodies have been developed to target various endogenous molecules[[10](#_ENREF_10)]; and (4) therapeutic cancer vaccines deliver concentrated antigen to both HLA class I and II molecules of antigen presenting cells, promoting both CD4 and CD8 T-cell responses which help to generate an anti-tumor response[[11](#_ENREF_11)].

**SYSTEM BASED CONSEQUENCES OF IMMUNOTHERAPEUTIC AGENTS**

The following sections, organized by organ system, will describe and detail the potential adverse effects and toxicities associated with the use of currently approved immunotherapeutic agents. The systems specific adverse effects described are presented in the context of following systems: cardiovascular, dermatologic, endocrine, gastrointestinal, neurologic, and pulmonary. In addition, we will provide an overview of some of the adverse events unique to chimeric antigen receptor (CAR) T-cell therapy.

**CARDIOVASCULAR**

The cardiovascular consequences of immunotherapeutic agents are being increasingly documented in medical literature, especially with the use of ICIs. The reported adverse effects can be further classified into two specific subtypes including structural, such as pericarditis and myocarditis, and arrhythmogenic, such as sick sinus syndrome. Cardiotoxicity has been most frequently reported with the use of ICIs and CAR T-cell therapy.

***Structural consequences***

The structural consequences may manifest in several ways. Patients may be asymptomatic, but laboratory investigations may report elevated markers of myocardial necrosis[[12](#_ENREF_12" \o "Jagielska, 2020 #41)]. Furthermore, they can present with systolic dysfunction, symptoms of myocarditis, pericarditis, pericardial effusions, cardiac tamponade, or stress induced cardiomyopathies[[12](#_ENREF_12" \o "Jagielska, 2020 #41)]. Structural cardiotoxicity secondary to the use of immunotherapy may be life-threatening and high suspicion for early recognition is imperative to prevent adverse outcomes.

The cause of cardiotoxicity due to immunotherapy is still largely under investigation. For immune checkpoint inhibitors, the presence of similar T-cell receptor sequences in tumor and in cardiac muscle is presumed to be the reason for myocardial and pericardial inflammation[[13](#_ENREF_13" \o "Ganatra, 2019 #38)]. Furthermore, this can be confirmed with histopathological findings of fibrinous exudates with lymphocyte, plasma cell, and macrophage infiltration in patients with pericarditis[[14](#_ENREF_14" \o "Ala, 2019 #39)]. Additionally, histopathological findings of endomyocardial CD8+ T-cell cells on post-mortem biopsy of those who had myocarditis secondary to immune checkpoint inhibitors also supports this theory[[15](#_ENREF_15" \o "Yang, 2018 #40)]. With respect to CAR T-Cell therapy, the mechanism of action of cardiotoxicity is presumed to be secondary to a cytokine storm generated by the reprogrammed T-cells leading to cardiovascular collapse[[13](#_ENREF_13" \o "Ganatra, 2019 #38)]. Furthermore, like immune checkpoint inhibitors, common antigens between cardiac tissues and tumor cells, specifically the protein titin, has been hypothesized as a mechanism of action of cardiotoxicity from CAR T-cell therapy[[13](#_ENREF_13" \o "Ganatra, 2019 #38)].

The literature also reports structural consequences with the use of monoclonal antibody therapy. Patient’s with non-small-cell lung cancer treated with pembrolizumab, a PD-1 inhibitor, were found to have immune-mediated pericarditis[[16](#_ENREF_16" \o "Dhenin, 2019 #63)]. Additionally, trastuzumab, a HER2 receptor blocker used in breast cancer treatment, is commonly known to cause cardiotoxicity. While the exact mechanism of cardiotoxicity is unknown, studies have shown that the structural changes are different from those found with anthracyclines[[17](#_ENREF_17" \o "Ewer, 2015 #64)]. The most current hypotheses focuses on trastuzumab’s interference in the integral HER2 signaling cascade resulting in the accumulation of toxic metabolites leading to cardiotoxicity. However, the exact molecular contributor is still unknown[[18](#_ENREF_18" \o "Mohan, 2018 #62)].

Treatment for the cardiotoxicity is centered around treating the over-active T-cell response with therapies such as high dose intravenous steroids, immunosuppressants, plasmapheresis, and intravenous immunoglobulins[[14](#_ENREF_14" \o "Ala, 2019 #39)]. Furthermore, cessation of therapy should be considered with persistent cardiac symptoms longer than three months, if immunosuppressive therapy will need to be ongoing, or if there is evidence of life-threatening side-effects[[14](#_ENREF_14" \o "Ala, 2019 #39)].

***Rhythm consequences***

Arrhythmias that have been well documented in the setting of immunotherapy use include atrial fibrillation, supraventricular arrhythmias, ventricular arrhythmias, and heart blocks[[15](#_ENREF_15" \o "Yang, 2018 #40)].

The cause of rhythm consequences secondary to the use of immunotherapy is also still largely under investigation with even less knowledge about the mechanism when compared to structural consequences. If an arrhythmia presents in the setting of immunotherapy use, it is most often a result of persistent myocarditis that disrupts cardiac conduction[[15](#_ENREF_15" \o "Yang, 2018 #40)]. However, post-mortem histopathologic analysis has demonstrated lymphocyte infiltration in both the sinoatrial node and the atrioventricular node, which provides strong evidence for a T-cell mediated insult similar to the mechanism of action of structural cardiotoxicity[[15](#_ENREF_15" \o "Yang, 2018 #40)].

Patients who develop arrhythmias in the setting of immunotherapy should be treated with antiarrhythmics[[15](#_ENREF_15" \o "Yang, 2018 #40)]. If the arrhythmia persists, an evaluation for a permanent pacemaker should be considered[[15](#_ENREF_15" \o "Yang, 2018 #40)]. Similarly to structural consequences, cessation of immunotherapy should be considered with persistent cardiotoxic side effects and evidence of life-threatening consequences[[14](#_ENREF_14" \o "Ala, 2019 #39)].

**DERMATOLOGICAL**

Dermatologic toxicities are extremely common with immunotherapy, most commonly resulting in a maculopapular rash and pruritis[[19](#_ENREF_19" \o "Si, 2020 #65)]. However, lichen dermatitis, psoriasis, bullous pemphigoid, as well as life-threatening adverse events like erythema multiforme or Stevens-Johnson syndrome (SJS) have also been reported[[19](#_ENREF_19" \o "Si, 2020 #65)].

The exact mechanism of causation of these lesions is not well understood, but several hypotheses have been put forth. Antibodies directed against melanoma-associated antigens can cross-react with the antigens produced by normal melanocytes. The presence of a Vitiligo-like lesion has been associated with higher objective tumor response rates.

Monoclonal antibodies are also frequently linked to dermatological adverse events. Rituximab, a CD20 antagonist, has been associated with morbilliform skin eruptions or even SJS, with the former requiring steroid therapy, while the latter requires immediate discontinuation of the agent[[20](#_ENREF_20" \o "Pasadyn, 2020 #66)]. Anakinra, an IL-1 antagonist, is associated with new-onset psoriasis as well as injection-site reactions. Tocilizumab, an IL-6 receptor antagonist, may rarely present with skin rash[[20](#_ENREF_20" \o "Pasadyn, 2020 #66)]. Ipilimumab, a CTLA-4 inhibitor used in the treatment of melanoma, has a diverse range of dermatologic adverse effects associated with its use such as rashes, folliculitis, mucositis, rosacea, SJS and vitiligo[[20](#_ENREF_20" \o "Pasadyn, 2020 #66)].

It is highly recommended for patients to undergo a dermatologic exam prior to starting immunotherapy to establish a baseline[[19](#_ENREF_19" \o "Si, 2020 #65)]. Depending on the severity of the condition, topical or systemic corticosteroids can be considered. However, prompt discontinuation is necessary in the event of severe reactions such as SJS[[20](#_ENREF_20" \o "Pasadyn, 2020 #66)]. Early detection and prompt management of these reactions is necessary as they profoundly diminish quality of life.

**ENDOCRINE**

Endocrine irAEs are also well documented, with the most common being acute hypophysitis and thyroid dysfunction[[21](#_ENREF_21" \o "Puzanov, 2017 #8)]. Additionally, adrenal insufficiency, pancreatic dysfunction resulting in worsening glycemic control with subsequent diabetic ketoacidosis (DKA) or hyperosmolar hyperglycemic state, and hypoparathyroidism have also been reported[[21](#_ENREF_21" \o "Puzanov, 2017 #8)]. These adverse events are most commonly associated with ICI therapy. Conversely, oncolytic viruses, adoptive T-cell transfer, and cancer vaccines may rarely cause endocrinological dysfunction[[22](#_ENREF_22" \o "Chalan, 2018 #67)]. Currently, there are no specific recommendations for the screening of patients on ICI therapy for endocrine dysfunction, thereby making establishing a diagnosis of endocrine irAE quite challenging. Therefore, it is of critical importance that care providers monitor these patients closely to avoid morbidity and mortality secondary to the use of these therapeutic agents. The specific endocrine system related side effects, including proposed mechanism, presentation, and management strategies are discussed in detail in the following sections.

***Thyroid***

As discussed, thyroid dysfunction is one of the most common irAEs secondary to the use of ICI therapy. The mechanism of the development of thyroid dysfunction is currently being studied; however, there are several theories that have been proposed as the possible cause. First, it is assumed that increased immune system activation secondary to the use of ICIs may lead to an enhanced cytotoxic immune reaction that damages healthy tissue[[23](#_ENREF_23" \o "Jannin, 2019 #46)]. Second, it has been recognized that patients suffering from autoimmune damage associated with ICIs have an increase in the HLA-DR expression on CD14+CD16+ monocytes[[23](#_ENREF_23" \o "Jannin, 2019 #46)]. Some investigators have hypothesized that this overexpression may lead to excess monocytic activation, which infiltrate the thyroid gland, recognize self-antigens and induce subsequent cytotoxic damage to the normal cells[[23](#_ENREF_23" \o "Jannin, 2019 #46)]. However, it is not entirely clear whether the monocytes are directly responsible for the cytotoxic damage, or whether the damage is mediated by monocyte-activated autoreactive CD8+ T-cells. It has been well recognized that the incidence of autoimmune thyroid dysfunction occurs at much higher rate with PD-1/PD-L1 inhibitors as compared to CTLA-4 inhibitors. It is believed that this observation could be because healthy thyroid tissue has an increased expression of PD-L1, or because intra-thyroid lymphocytes have lower CTLA-4 expression[[23](#_ENREF_23" \o "Jannin, 2019 #46)].

Thyroid dysfunction could lead to features of hypo- or hyperthyroidism in these patients. Symptoms of hypothyroidism include fatigue, cold intolerance, weight gain, constipation, and hair loss, among many others. These patients may present with elevated Thyroid stimulating hormone (TSH) and low levels of T4. According to a retrospective cohort study by Ma *et al*[[24](#_ENREF_24" \o "Ma, 2019 #47)], the median time to onset of hypothyroidism in patients receiving pembrolizumab (PD-1 inhibitor) was found to be 84 d (range: 43-544; *P* = 0.333). For those receiving nivolumab (PD-1 inhibitor) and ipilimumab (CTLA-4 inhibitor) + nivolumab (PD-1 inhibitor), the median time to onset was 84 d (range: 14–154; *P* = 0.002) and 62 d (range: 21–141; *P* = 0.057) respectively[[24](#_ENREF_24" \o "Ma, 2019 #47)].

Patients may also present with thyrotoxicosis, often secondary to thyroiditis or Graves’ disease[[21](#_ENREF_21" \o "Puzanov, 2017 #8)]. These patients will have typical features of hyperthyroidism such as weight loss, palpitations, tremors, diarrhea, and heat intolerance. Furthermore, many patients may commonly be asymptomatic, with the hyperthyroid state uncovered on routine laboratory investigations[[21](#_ENREF_21" \o "Puzanov, 2017 #8)]. Ma *et al*[[24](#_ENREF_24)] found that the median time to onset of thyrotoxicosis in patients treated with pembrolizumab (PD-1 inhibitor) was 44 d (range: 19-447; *P* = 0.593), while time for those receiving nivolumab (PD-1 inhibitor) was 56 d (range: 13-126; *P* = 0.004) and ipilimumab (CTLA-4 inhibitor) + nivolumab (PD-1 inhibitor) was 21 d (range: 7-64; *P* = 0.158). It is abundantly evident from these studies that the use of combination therapy leads to a more rapid symptom onset.

The treatment of thyroid dysfunction in these patients varies and depends on several factors such as whether the patient is suffering from hypo- *vs* hyperthyroidism, the severity of symptoms, and whether additional intervention such as hospitalization or surgery are indicated. The Society for Immunotherapy of Cancer has put forth guidelines for patients with hypothyroidism. These guidelines recommend that patients with severe symptoms with limitations of activities of daily living should discontinue ICI therapy, and should start standard thyroid replacement therapy[[21](#_ENREF_21" \o "Puzanov, 2017 #8)]. Additionally, TSH and free T4 should be repeated after 6-8 wk of therapy to allow for titration of the dose. Resolution of symptoms is not needed prior to restarting ICI therapy, and these patients may resume ICI therapy once symptoms are no longer limiting self-care activities of daily living[[21](#_ENREF_21" \o "Puzanov, 2017 #8)]. For patients with hyperthyroidism, the recommendation for discontinuation of ICIs is similar. In patients with severe symptoms that limit self-care activities of daily living, ICI should be discontinued immediately. Additionally, patients may benefit from beta-blockers for symptomatic improvement of hyperthyroid symptoms[[21](#_ENREF_21" \o "Puzanov, 2017 #8)]. Of note, corticosteroids are not required for patients with thyroid irAEs.

***Pancreas***

Another frequently effected organ is the pancreas, as patients on ICIs have been reported to have diabetes as a consequence of the therapy[[25](#_ENREF_25" \o "Tan, 2019 #48)]. These patients frequently present with symptoms consistent with DKA and may be subsequently diagnosed with new onset type 1 diabetes mellitus. However there have also been reports of insidious worsening of type 2 diabetes mellitus presenting as ketoacidosis as well[[25](#_ENREF_25" \o "Tan, 2019 #48),[26](#_ENREF_26)]. The pathophysiology of the development of diabetes in these patients is not well understood, but it is postulated that it may be due to autoreactive CD8+ T cell activation against pancreatic beta cells as a consequence of the blockade of inhibitory pathways, such as the PD-1/PD-L1 pathway[[27](#_ENREF_27" \o "Clotman, 2018 #50)]. Patients may present with symptoms typical of hyperglycemia or ketoacidosis, including polyuria, polydipsia, weight loss, nausea, and vomiting. In a review of cases reported by Tan *et al*[[25](#_ENREF_25" \o "Tan, 2019 #48)], it was found that the median onset of clinical presentation was 7.5 wk after starting ICI therapy, although cases have been described as early as one week or as late as one year after initiation. While more than two-thirds of the patients presented in DKA, only about 50% had positive auto-antibodies (anti-GAD, anti-insulin, anti-islet cell A, anti-zinc transport 8)[[25](#_ENREF_25" \o "Tan, 2019 #48)].

Patients presenting with features of DKA should discontinue ICI therapy immediately and should be managed according to standard guidelines for DKA management[[21](#_ENREF_21" \o "Puzanov, 2017 #8)]. If patients do not have DKA, but fasting glucose is in the 250-500 mg/dL range, ICI should also be held and the patient should be treated with insulin with a goal of fasting glucose level below 160 mg/dL[[21](#_ENREF_21" \o "Puzanov, 2017 #8)].All patients with new onset diabetes secondary to the use of ICIs should receive an endocrinology consultation.

***Pituitary/adrenal insufficiency***

Pituitary dysfunction and adrenal insufficiency have also been well documented irAEs of ICI therapy. In a meta-analysis conducted by Lu *et al*[[28](#_ENREF_28" \o "Lu, 2019 #51)], the incidence of adrenal insufficiency was found to be 2.43% (95%CI: 1.73%-3.22%), while the rate of hypophysitis was reported to be 3.25% (95%CI: 2.15%-4.51%). Hypophysitis most frequently occurs secondary to CTLA-4 inhibitor therapy[[28](#_ENREF_28" \o "Lu, 2019 #51)]. The specific mechanism is not well understood; however, multiple theories are currently being investigated. First, similar to all irAEs, it is believed that inhibition of self-tolerance leads to an autoimmune response against pituitary endocrine cells[[29](#_ENREF_29" \o "Wehbeh, 2019 #52)]. It has also been shown that CTLA-4 expression is greater on endocrine cells of the pituitary gland, especially prolactin and TSH secreting cells, which could possibly explain as to why both pituitary and thyroid dysfunction are the most common irAEs, and why symptoms occur more frequently in patients receiving CTLA-4 inhibitor therapy[[29](#_ENREF_29)]. Patients may present with symptoms of headache, vision changes, and fatigue[[21](#_ENREF_21" \o "Puzanov, 2017 #8)]. If patients subsequently develop adrenal insufficiency secondary to pituitary dysfunction, symptoms may include abdominal pain, vomiting, weakness, fatigue, hypotension, weight loss, and features of adrenal crisis. Additionally, the loss of function of other anterior pituitary hormones (FSH, LH, ACTH, TSH, Prolactin, GH) may lead to presenting symptoms consistent with the effects of the deficient hormone. Literature reports the median onset of clinical presentation to be 12 wk (range 3-76 wk) from initiation of ICI therapy.

Management of the pituitary dysfunction is focused on the replacement of the deficient hormone. In patients with adrenal insufficiency, steroids should be started to replace the corticosteroid deficit[[21](#_ENREF_21" \o "Puzanov, 2017 #8)]. ICI therapy should be held in patients with moderate symptoms requiring local or non-invasive intervention, who have some limitation of instrumental activities of daily living[[21](#_ENREF_21" \o "Puzanov, 2017 #8)]. ICI may be resumed when an initial work up is completed and the hormone replacement has begun[[21](#_ENREF_21" \o "Puzanov, 2017 #8)].

**GASTROINTESTINAL**

The literature reports abundant gastrointestinal (GI) adverse effects associated with the use of immunotherapeutic agents. These have been reported across every class of immunotherapeutic agent, except for therapeutic cancer vaccines. In contrast to other cytotoxic therapies, cancer vaccines have reported minimal toxicities in a majority of clinical trials[[30](#_ENREF_30" \o "Guo, 2013 #33)]. However these therapies are largely still in the development phase and adverse events will most likely be elucidated in the future.

The side effects associated with ICIs tend to be mild and transient with watery, non-bloody diarrhea being the most commonly reported[[31](#_ENREF_31" \o "Assarzadegan, 2018 #19)]. Colitis, presenting with bloody diarrhea and fever, has also been reported. Typically, these adverse effects are more frequent with anti-CTLA-4 and anti-PD-1 agents[[32](#_ENREF_32" \o "Marin-Acevedo, 2018 #20)]. Mouth ulcers, anal lesions like fistulas, abscesses or fissures, and other extra-intestinal manifestations may also occur[[33](#_ENREF_33" \o "Marthey, 2016 #21)]. Some patients may also have asymptomatic elevation in the liver function tests[[34](#_ENREF_34" \o "Morgan, 2010 #27)]. Although the upper GI tract is rarely involved, some reported adverse effects include gastritis, esophagitis and aphthous ulcers[[32](#_ENREF_32" \o "Marin-Acevedo, 2018 #20)].

The management of GI adverse effects as a result of ICI therapy is stratified based on the severity of the diarrhea or colitis. They are graded from 1-5 based on the severity, number of stools per day, and the need for hospitalization or additional intervention[[35](#_ENREF_35),[36](#_ENREF_36)].Treatment typically ranges from supportive (antidiarrheals, fluids/electrolytes) to hospitalization (IV corticosteroids and discontinuation of immunotherapy). Life threatening enterocolitis with perforation requires immediate surgical intervention.

T-cell therapy is associated with on-target adverse effects which are usually not life-threatening but may be treatment limiting. These on-target adverse effects are expected as the engineered T-cells may share specific target antigens with different organ systems with the most common T-cell adverse event being colitis[[37](#_ENREF_37" \o "Tey, 2014 #26)]. The decision to discontinue T-cell therapy should be individualized to the patient and based on the severity of illness.

Monoclonal antibody agents have also been associated with diverse GI toxicities. Anti-angiogenic agents such as bevacizumab (VEGF inhibitor) and aflibercept (VEGF inhibitor) have been associated with dose-dependent GI perforations[[38](#_ENREF_38" \o "Qi, 2014 #29)]. It is hypothesized that this may be due to ischemia secondary to limitation of blood flow to the GI tract[[39](#_ENREF_39" \o "Qi, 2014 #30)]. The management of GI perforation includes prompt surgical intervention, bowel rest, and initiation of intravenous antibiotics. It has been reported that EGFR inhibitors such as cetuximab and panitumumab have a 66% increased risk of diarrhea compared to chemotherapy alone[[40](#_ENREF_40" \o "Miroddi, 2015 #31)]. Hypomagnesemia is another common adverse effect of EGFR inhibitor therapy. It is dependent on the duration of the therapy and could possibly be the result of chronic diarrhea[[41](#_ENREF_41" \o "Arora, 2017 #28)]. The management of diarrhea consists of bowel rest, hydration, electrolyte repletion, and the use of anti-motility agents. Hospitalization may be necessary in patients with severe dehydration. Finally, Trastuzumab (HER2/neu inhibitor) has been found to be associated with a higher prevalence of diarrhea compared to chemotherapy alone[[42](#_ENREF_42" \o "Bang, 2010 #32)].

**NEUROLOGICAL**

Although rare, neurological irAEs are well reported side effects of ICI therapy, occurring in 1% of patients receiving monotherapy and 2%-3% of patients receiving a combination of ICIs[[43](#_ENREF_43" \o "Reynolds, 2019 #42)]. Median onset of symptom occurrence is 6 wk; however, symptoms have been shown to occur as early as after the first dose and as late as at the 68th week after initiation of therapy[[43](#_ENREF_43" \o "Reynolds, 2019 #42)]. Neurologic events are more commonly reported with CTLA-4 inhibitor therapy[[44](#_ENREF_44" \o "Dalakas, 2018 #43)].

Numerous neurologic irAE have been reported. These include inflammatory myopathies, myasthenia gravis, vasculitis, small fiber sensory type neuropathies, immune-mediated neuropathies, cranial mononeuropathies, aseptic meningitis, autoimmune encephalitis, and multiple sclerosis (both exacerbation and new onset CNS demyelinating disease)[[44](#_ENREF_44" \o "Dalakas, 2018 #43)]. The presentation is largely dependent on the type of irAE. The mechanism for neurological symptoms is most likely due to the increased T-cell and macrophage activation secondary to immune checkpoint inhibition, leading to autoimmune damage to parts of the nervous system[[45](#_ENREF_45" \o "Manousakis, 2013 #44)]. Furthermore, it has been suggested that the aberrant T-cell activation and release of inflammatory cytokines may lead to sub-perineural edema and inflammation of the endoneurial microvessels, which is in part responsible for the nervous system dysfunction[[46](#_ENREF_46" \o "Choi, 2020 #45)]. The treatment of the neurological adverse effects depends on the severity of the symptoms.

For any neurologic irAE, ICI therapy should be paused immediately, with considerations for possible permanent discontinuation of therapy if symptoms are severe (limiting ability to perform self-care activities of daily living) or progressive[[21](#_ENREF_21" \o "Puzanov, 2017 #8)]. Moreover, patients with moderate symptoms or greater, based on limitation of daily activities, should be started on methylprednisolone (0.5 to 1.0 mg/kg/d for moderate; 1-2 mg/kg/d for severe) after a central nervous system infection has been ruled out as a cause of the neurologic dysfunction[[21](#_ENREF_21" \o "Puzanov, 2017 #8)]. Additionally, in patients with involvement of the respiratory muscles, admission to the intensive care unit for close observation and ventilator therapy may be warranted[[46](#_ENREF_46" \o "Choi, 2020 #45)]. The decision to restart the patient on immunotherapy should be a joint decision between the patient, oncologist and the neurology consultant.

Monoclonal antibodies and cancer vaccines have not demonstrated consistent evidence of causing adverse neurological consequences. However, additional research is required to elucidate whether this is due to a lack of evidence or the true absence of adverse effects.

**PULMONARY**

The most frequently cited pulmonary adverse effects of immunotherapy are interstitial lung disease and concomitant pneumonitis. Ipilimumab, a CTLA-4 inhibitor, has been shown to be a common agent responsible for pulmonary adverse effects[[47](#_ENREF_48" \o "Delaunay, 2019 #58)]. Most clinical trials reported a 3%-5% incidence of pneumonitis following treatment with immunotherapy. Studies by Delaunay *et al*[[47](#_ENREF_48" \o "Delaunay, 2019 #58)] and Naidoo *et al*[[48](#_ENREF_49)] reported that the incidence was higher in current or ex-smokers. A recent metanalysis by Jiang *et al*[49] investigated fatal immune-related adverse events by checkpoint inhibitors, with ten out of twelve trials evaluating CTLA-4 inhibitors. Based on the analysis, CTLA-4 inhibitors had a higher risk of fatal adverse events of pulmonary toxicity, with an overall risk of 20.5% (OR = 3.48, *P* = 0.10). A subgroup analysis was also performed according to the different dosing regimens and it was reported that the occurrence of toxicity with a 10 mg/kg dose was greater when compared to a 3 mg/kg dose (dose-dependent toxicity)[49]. The study by Nishino *et al*[[50](#_ENREF_51" \o "Nishino, 2016 #54)] reviewed ten different nivolumab (PD-1 inhibitor) trials alone or in combination with other immune checkpoint inhibitors for advanced cancers such as melanoma, lung cancer, and lymphoma, and identified pneumonitis in 11.8% patients. Radiological findings of sarcoid-like granulomatosis has also been reported in a patient receiving both anti-CTLA4 and anti-PD-1 agents. Dose-related adverse effects are less frequently encountered with the use of PD-1 inhibitors, and most toxicities are similar at doses ranging from 0.3 to 10 mg/kg.

The preferred initial modality for identifying features of pneumonitis is computed tomography scan. Pneumonitis can be classified as grade 1 with subclinical radiographic changes, grade 2 with mild dyspnea and cough, grade 3 and 4 with severe and life-threatening symptoms, and radiographic findings suggestive of acute interstitial pneumonia, acute respiratory distress syndrome, and/or cryptogenic organizing pneumonia[[51](#_ENREF_52" \o "Chuzi, 2017 #69)]. The base of the lungs are most frequently involved, followed by the middle and upper regions; however, most often it may be mixed and multifocal. Bronchoalveolar lavage may be helpful in patients with cellular interstitial pneumonitis in that the fluid may show lymphocytosis, however there have been relatively few reports describing this finding and further research is needed[[53](#_ENREF_52" \o "Chuzi, 2017 #69)].

The first step in management of all cases is the discontinuation of the immunotherapy, and initiation of corticosteroids[51]. The average duration of corticosteroid therapy also varies, but in most cases the median time is about 6.1 wk. Grade 1 Pneumonitis can be treated by interrupting immunotherapy, whereas grade 2 responds to oral corticosteroids. Grade 3 and 4 may need IV steroids, with some cases requiring infliximab, mycophenolate, tocilizumab, and IL-1 targeted agents[51].

Restarting immunotherapy in these patients is a challenge as pneumonitis can reoccur at any time; hence, close follow-up is recommended. Reinitiating ICIs can lead to recurrence of Pneumonitis in up to 33% of cases[[52](#_ENREF_53" \o "Abou Alaiwi, 2020 #70)]. A flare of Pneumonitis can also occur during the steroid taper phase, and manifest similar to the initial episode. Data shows that steroids are responsive in subsequent flares as well as recurrence. Additionally, another challenging scenario is the relationship between radiotherapy and chemotherapy in aiding the development of side effects when used both prior to the initiation of immunotherapy as well as in combination with it[[52](#_ENREF_53" \o "Abou Alaiwi, 2020 #70)].

**TOXICITY RELATED TO CAR T-CELL THERAPY**

CAR T-cell therapy is a type of T-cell transfer therapy currently approved for the treatment of several hematologic malignancies, including acute lymphoblastic leukemia and diffuse large B-cell lymphoma[53]. The mechanism of action involves the creation of CAR T-cells directed against the CD19 protein, which is present on B-cells[53]. While this treatment modality has proven effective in destroying chemotherapy-resistant tumor cells, it does not come without its own adverse events. It is thought that the irAEs resulting from CD19-directed CAR T cells are due to the fact that these modified T cells maintain their native receptors, and have the potential to expand rapidly once activated *via* interaction with the CD19 antigen[54]. These cells then have the potential to damage normal tissues expressing non CD19 antigens[54]. Several irAEs have been reported, including cytokine release syndrome (CRS), neurotoxicity, and hematologic abnormalities such as B cell aplasia, hypogammaglobulinemia, anemia, or thrombocytopenia[55].

CRS is an overwhelming release of cytokines, especially IL-1 and IL-6, that occurs as a result of CAR T-cell therapy. According to a review by Long *et al*[55], it may occur in up to 90% of patients. The symptoms of CRS vary greatly, and can involve a number of different organ systems. One of the most common symptoms is fever, and initial symptoms may mimic an acute infectious process[56]. In addition, there are several life-threatening complications that have been reported, including cardiac dysfunction, acute respiratory distress syndrome, neurologic toxicities, renal failure, hepatic failure, and hypercoagulability[55,56].

Laboratory findings in CRS are variable. Inflammatory markers such as C-reactive protein are often elevated[57]. In addition, patients may present with leukocytosis or leukopenia, anemia or thrombocytopenia, and elevated levels of D-dimer or ferritin[57]. Elevated levels of inflammatory cytokines, such as interferon gamma, IL-6, IL-10, or IL-2 may be present[57]. It should be noted that elevated levels of IL-6 are often found in CAR T-cell therapy induced CRS[57].

The National Cancer Institute Common Terminology Criteria for Adverse Events (CTCAE v5.0) has established a grading system to assess the severity of CRS[58]. The grading criteria for CRS according to the CTCAE v5.0 can be classified as follows[58]: Grade 1: Fever with or without constitutional symptoms; Grade 2: Hypotension responding to fluids; hypoxia responding to < 40% O2; Grade 3: Hypotension managed with one pressor; hypoxia requiring ≥ 40% O2; Grade 4: Life-threatening consequences; urgent intervention indicated; Grade 5: Death.

The management of CRS is centered around preventing life-threatening effects of the syndrome while allowing for the CAR T-cells to continue their anti-tumor activity[56]. For patients with grade 1 CRS, supportive care may be most appropriate treatment option[56]. Patients with grade 2 CRS may be treated conservatively, however for patients with extensive comorbidities and those of advanced age, treatment with Tocilizumab (IL-6 receptor inhibitor) and corticosteroids should be considered[56]. Finally, patients with grade 3 or 4 CRS should be admitted to the intensive care unit and treated with Tocilizumab (IL-6 receptor inhibitor) and corticosteroids. Tocilizumab has been evaluated and has been found to have considerable efficacy as a treatment option for CAR T-cell induced CRS[56,57]. Lee *et al*[56] recommend giving tocilizumab at a dose of 4 mg/kg over the course of one hour in adults. They also recommend repeating the dose if there are no signs of clinical improvement over 24 h to 48 h[56].

Neurotoxicity is another common adverse effect secondary to CAR T-cell therapy. According to a review by Gust *et al*[59], the incidence of neurotoxicity in patients undergoing CAR T-cell therapy varies between 2% to 70% of patients. The clinical syndrome of CAR T-cell induced neurotoxicity is often referred to as immune effector cell-associated neurotoxicity syndrome (ICANS)[59]. The most well known risk factors for developing ICANS are systemic cytokine release in addition to the severity of CRS[59]. Symptoms normally begin within the first 7 d following CAR T-cell infusion[59]. Symptoms include cognitive impairment, tremor, headache, altered levels of consciousness, seizures, and even coma[59]. The feared complication is cerebral edema, which is often fatal[59]. Currently, there is no evidence that blockade of a single inflammatory mediator is beneficial in ICANS. Therefore, it is recommended that corticosteroids be used as first line treatment due to their ability to reduce cerebral edema as well as to induce an immunosuppressive effect that decreases the secretion of pro-inflammatory cytokines[59].

**CONCLUSION**

Immunotherapy, a rapidly evolving therapeutic entity, has proven to be a worthwhile addition to the arsenal of therapeutic modalities care providers have against cancers. Advancement in the field is highly dependent on a better understanding of the immune system and the advent of newer agents. However, like any other therapeutic modality, it has its own set of adverse effects and toxicities. It is essential that physicians familiarize themselves with the adverse effects and toxicities associated with these agents as their role in management of cancers continues to rise. Additionally, proper patient education on the adverse effects of these agents along with the process of joint decision making between the provider and physician will hopefully result in a decrease in unfavorable outcomes. It is also worth noting that immunotherapy is still in its infancy phase, and as newer, more specific therapies emerge secondary to a better understanding of the workings of the immune system, the ratio of benefits to risks may increase significantly.

**REFERENCES**

1 **Leach DR**, Krummel MF, Allison JP. Enhancement of antitumor immunity by CTLA-4 blockade. *Science* 1996; **271**: 1734-1736 [PMID: 8596936 DOI: 10.1126/science.271.5256.1734]

2 **Chen L**. Co-inhibitory molecules of the B7-CD28 family in the control of T-cell immunity. *Nat Rev Immunol* 2004; **4**: 336-347 [PMID: 15122199 DOI: 10.1038/nri1349]

3 **Yang Y**. Cancer immunotherapy: harnessing the immune system to battle cancer. *J Clin Invest* 2015; **125**: 3335-3337 [PMID: 26325031 DOI: 10.1172/JCI83871]

4 **Li X**, Shao C, Shi Y, Han W. Lessons learned from the blockade of immune checkpoints in cancer immunotherapy. *J Hematol Oncol* 2018; **11**: 31 [PMID: 29482595 DOI: 10.1186/s13045-018-0578-4]

5 **Cancer Research Institute**. FDA Approval Timeline of Active Immunotherapies. [cited 13 January 2021]. Available from: https://www.cancerresearch.org/scientists/immuno-oncology-landscape/fda-approval-timeline-of-active-immunotherapies

6 **Yaguchi T**, Kawakami Y. Cancer-induced heterogeneous immunosuppressive tumor microenvironments and their personalized modulation. *Int Immunol* 2016; **28**: 393-399 [PMID: 27401477 DOI: 10.1093/intimm/dxw030]

7 **National Cancer Institute**. Cancer Immunotherapy. [cited 13 January 2021]. Available from: https://www.cancer.gov/about-cancer/treatment/types/immunotherapy

8 **van den Bulk J**, Verdegaal EM, de Miranda NF. Cancer immunotherapy: broadening the scope of targetable tumours. *Open Biol* 2018; **8** [PMID: 29875199 DOI: 10.1098/rsob.180037]

9 **Zhao Q**, Yu J, Meng X. A good start of immunotherapy in esophageal cancer. *Cancer Med* 2019; **8**: 4519-4526 [PMID: 31231980 DOI: 10.1002/cam4.2336]

10 **Scott AM**, Allison JP, Wolchok JD. Monoclonal antibodies in cancer therapy. *Cancer Immun* 2012; **12**: 14 [PMID: 22896759]

11 **Melief CJ**, van Hall T, Arens R, Ossendorp F, van der Burg SH. Therapeutic cancer vaccines. *J Clin Invest* 2015; **125**: 3401-3412 [PMID: 26214521 DOI: 10.1172/JCI80009]

12 **Jagielska B**, Ozdowska P, Gepner K, Kubala S, Siedlecki JA, Sarnowski TJ, Sarnowska E. Cardiotoxicity danger in immunotherapy. *IUBMB Life* 2020; **72**: 1160-1167 [PMID: 32359132 DOI: 10.1002/iub.2299]

13 **Ganatra S**, Parikh R, Neilan TG. Cardiotoxicity of Immune Therapy. *Cardiol Clin* 2019; **37**: 385-397 [PMID: 31587780 DOI: 10.1016/j.ccl.2019.07.008]

14 **Ala CK**, Klein AL, Moslehi JJ. Cancer Treatment-Associated Pericardial Disease: Epidemiology, Clinical Presentation, Diagnosis, and Management. *Curr Cardiol Rep* 2019; **21**: 156 [PMID: 31768769 DOI: 10.1007/s11886-019-1225-6]

15 **Yang S**, Asnani A. Cardiotoxicities associated with immune checkpoint inhibitors. *Curr Probl Cancer* 2018; **42**: 422-432 [PMID: 30173878 DOI: 10.1016/j.currproblcancer.2018.07.002]

16 **Dhenin A**, Samartzi V, Lejeune S, Seront E. Cascade of immunologic adverse events related to pembrolizumab treatment. *BMJ Case Rep* 2019; **12** [PMID: 31167767 DOI: 10.1136/bcr-2018-229149]

17 **Ewer MS**, Ewer SM. Cardiotoxicity of anticancer treatments. *Nat Rev Cardiol* 2015; **12**: 547-558 [PMID: 25962976 DOI: 10.1038/nrcardio.2015.65]

18 **Mohan N**, Jiang J, Dokmanovic M, Wu WJ. Trastuzumab-mediated cardiotoxicity: current understanding, challenges, and frontiers. *Antib Ther* 2018; **1**: 13-17 [PMID: 30215054 DOI: 10.1093/abt/tby003]

19 **Si X**, He C, Zhang L, Liu X, Li Y, Wang H, Guo X, Zhou J, Duan L, Wang M, Zhang L. Management of immune checkpoint inhibitor-related dermatologic adverse events. *Thorac Cancer* 2020; **11**: 488-492 [PMID: 31814310 DOI: 10.1111/1759-7714.13275]

20 **Pasadyn SR**, Knabel D, Fernandez AP, Warren CB. Cutaneous adverse effects of biologic medications. *Cleve Clin J Med* 2020; **87**: 288-299 [PMID: 32357984 DOI: 10.3949/ccjm.87a.19119]

21 **Puzanov I**, Diab A, Abdallah K, Bingham CO 3rd, Brogdon C, Dadu R, Hamad L, Kim S, Lacouture ME, LeBoeuf NR, Lenihan D, Onofrei C, Shannon V, Sharma R, Silk AW, Skondra D, Suarez-Almazor ME, Wang Y, Wiley K, Kaufman HL, Ernstoff MS; Society for Immunotherapy of Cancer Toxicity Management Working Group. Managing toxicities associated with immune checkpoint inhibitors: consensus recommendations from the Society for Immunotherapy of Cancer (SITC) Toxicity Management Working Group. *J Immunother Cancer* 2017; **5**: 95 [PMID: 29162153 DOI: 10.1186/s40425-017-0300-z]

22 **Chalan P**, Di Dalmazi G, Pani F, De Remigis A, Corsello A, Caturegli P. Thyroid dysfunctions secondary to cancer immunotherapy. *J Endocrinol Invest* 2018; **41**: 625-638 [PMID: 29238906 DOI: 10.1007/s40618-017-0778-8]

23 **Jannin A**, Penel N, Ladsous M, Vantyghem MC, Do Cao C. Tyrosine kinase inhibitors and immune checkpoint inhibitors-induced thyroid disorders. *Crit Rev Oncol Hematol* 2019; **141**: 23-35 [PMID: 31202955 DOI: 10.1016/j.critrevonc.2019.05.015]

24 **Ma C**, Hodi FS, Giobbie-Hurder A, Wang X, Zhou J, Zhang A, Zhou Y, Mao F, Angell TE, Andrews CP, Hu J, Barroso-Sousa R, Kaiser UB, Tolaney SM, Min L. The Impact of High-Dose Glucocorticoids on the Outcome of Immune-Checkpoint Inhibitor-Related Thyroid Disorders. *Cancer Immunol Res* 2019; **7**: 1214-1220 [PMID: 31088848 DOI: 10.1158/2326-6066.CIR-18-0613]

25 **Tan MH**, Iyengar R, Mizokami-Stout K, Yentz S, MacEachern MP, Shen LY, Redman B, Gianchandani R. Spectrum of immune checkpoint inhibitors-induced endocrinopathies in cancer patients: a scoping review of case reports. *Clin Diabetes Endocrinol* 2019; **5**: 1 [PMID: 30693099 DOI: 10.1186/s40842-018-0073-4]

26 **Kichloo A**, Albosta MS, McMahon S, Movsesian K, Wani F, Jamal SM, Aljadah M, Singh J. Pembrolizumab-Induced Diabetes Mellitus Presenting as Diabetic Ketoacidosis in a Patient With Metastatic Colonic Adenocarcinoma. *J Investig Med High Impact Case Rep* 2020; **8**: 2324709620951339 [PMID: 32830561 DOI: 10.1177/2324709620951339]

27 **Clotman K**, Janssens K, Specenier P, Weets I, De Block CEM. Programmed Cell Death-1 Inhibitor-Induced Type 1 Diabetes Mellitus. *J Clin Endocrinol Metab* 2018; **103**: 3144-3154 [PMID: 29955867 DOI: 10.1210/jc.2018-00728]

28 **Lu J**, Li L, Lan Y, Liang Y, Meng H. Immune checkpoint inhibitor-associated pituitary-adrenal dysfunction: A systematic review and meta-analysis. *Cancer Med* 2019; **8**: 7503-7515 [PMID: 31679184 DOI: 10.1002/cam4.2661]

29 **Wehbeh L**, Alreddawi S, Salvatori R. Hypophysitis in the era of immune checkpoint inhibitors and immunoglobulin G4-related disease. *Expert Rev Endocrinol Metab* 2019; **14**: 167-178 [PMID: 30939947 DOI: 10.1080/17446651.2019.1598260]

30 **Guo C**, Manjili MH, Subjeck JR, Sarkar D, Fisher PB, Wang XY. Therapeutic cancer vaccines: past, present, and future. *Adv Cancer Res* 2013; **119**: 421-475 [PMID: 23870514 DOI: 10.1016/B978-0-12-407190-2.00007-1]

31 **Assarzadegan N**, Montgomery E, Anders RA. Immune checkpoint inhibitor colitis: the flip side of the wonder drugs. *Virchows Arch* 2018; **472**: 125-133 [PMID: 29143108 DOI: 10.1007/s00428-017-2267-z]

32 **Marin-Acevedo JA**, Harris DM, Burton MC. Immunotherapy-Induced Colitis: An Emerging Problem for the Hospitalist. *J Hosp Med* 2018; **13**: 413-418 [PMID: 29419822 DOI: 10.12788/jhm.2925]

33 **Marthey L**, Mateus C, Mussini C, Nachury M, Nancey S, Grange F, Zallot C, Peyrin-Biroulet L, Rahier JF, Bourdier de Beauregard M, Mortier L, Coutzac C, Soularue E, Lanoy E, Kapel N, Planchard D, Chaput N, Robert C, Carbonnel F. Cancer Immunotherapy with Anti-CTLA-4 Monoclonal Antibodies Induces an Inflammatory Bowel Disease. *J Crohns Colitis* 2016; **10**: 395-401 [PMID: 26783344 DOI: 10.1093/ecco-jcc/jjv227]

34 **Morgan RA**, Yang JC, Kitano M, Dudley ME, Laurencot CM, Rosenberg SA. Case report of a serious adverse event following the administration of T cells transduced with a chimeric antigen receptor recognizing ERBB2. *Mol Ther* 2010; **18**: 843-851 [PMID: 20179677 DOI: 10.1038/mt.2010.24]

35 **Rocha M**, Correia de Sousa J, Salgado M, Araújo A, Pedroto I. Management of Gastrointestinal Toxicity from Immune Checkpoint Inhibitor. *GE Port J Gastroenterol* 2019; **26**: 268-274 [PMID: 31328141 DOI: 10.1159/000494569]

36 **Samaan MA**, Pavlidis P, Papa S, Powell N, Irving PM. Gastrointestinal toxicity of immune checkpoint inhibitors: from mechanisms to management. *Nat Rev Gastroenterol Hepatol* 2018; **15**: 222-234 [PMID: 29512649 DOI: 10.1038/nrgastro.2018.14]

37 **Tey SK**. Adoptive T-cell therapy: adverse events and safety switches. *Clin Transl Immunology* 2014; **3**: e17 [PMID: 25505965 DOI: 10.1038/cti.2014.11]

38 **Qi WX**, Shen Z, Tang LN, Yao Y. Bevacizumab increases the risk of gastrointestinal perforation in cancer patients: a meta-analysis with a focus on different subgroups. *Eur J Clin Pharmacol* 2014; **70**: 893-906 [PMID: 24858820 DOI: 10.1007/s00228-014-1687-9]

39 **Qi WX**, Shen F, Qing Z, Xiao-Mao G. Risk of gastrointestinal perforation in cancer patients treated with aflibercept: a systematic review and meta-analysis. *Tumour Biol* 2014; **35**: 10715-10722 [PMID: 25070487 DOI: 10.1007/s13277-014-2369-z]

40 **Miroddi M**, Sterrantino C, Simonelli I, Ciminata G, Phillips RS, Calapai G. Risk of grade 3-4 diarrhea and mucositis in colorectal cancer patients receiving anti-EGFR monoclonal antibodies regimens: A meta-analysis of 18 randomized controlled clinical trials. *Crit Rev Oncol Hematol* 2015; **96**: 355-371 [PMID: 26160607 DOI: 10.1016/j.critrevonc.2015.06.004]

41 **Arora N**, Gupta A, Singh PP. Biological agents in gastrointestinal cancers: adverse effects and their management. *J Gastrointest Oncol* 2017; **8**: 485-498 [PMID: 28736636 DOI: 10.21037/jgo.2017.01.07]

42 **Bang YJ**, Van Cutsem E, Feyereislova A, Chung HC, Shen L, Sawaki A, Lordick F, Ohtsu A, Omuro Y, Satoh T, Aprile G, Kulikov E, Hill J, Lehle M, Rüschoff J, Kang YK; ToGA Trial Investigators. Trastuzumab in combination with chemotherapy versus chemotherapy alone for treatment of HER2-positive advanced gastric or gastro-oesophageal junction cancer (ToGA): a phase 3, open-label, randomised controlled trial. *Lancet* 2010; **376**: 687-697 [PMID: 20728210 DOI: 10.1016/S0140-6736(10)61121-X]

43 **Reynolds KL**, Guidon AC. Diagnosis and Management of Immune Checkpoint Inhibitor-Associated Neurologic Toxicity: Illustrative Case and Review of the Literature. *Oncologist* 2019; **24**: 435-443 [PMID: 30482825 DOI: 10.1634/theoncologist.2018-0359]

44 **Dalakas MC**. Neurological complications of immune checkpoint inhibitors: what happens when you 'take the brakes off' the immune system. *Ther Adv Neurol Disord* 2018; **11**: 1756286418799864 [PMID: 30245744 DOI: 10.1177/1756286418799864]

45 **Manousakis G**, Koch J, Sommerville RB, El-Dokla A, Harms MB, Al-Lozi MT, Schmidt RE, Pestronk A. Multifocal radiculoneuropathy during ipilimumab treatment of melanoma. *Muscle Nerve* 2013; **48**: 440-444 [PMID: 23447136 DOI: 10.1002/mus.23830]

46 **Choi J**, Lee SY. Clinical Characteristics and Treatment of Immune-Related Adverse Events of Immune Checkpoint Inhibitors. *Immune Netw* 2020; **20**: e9 [PMID: 32158597 DOI: 10.4110/in.2020.20.e9]

47 **Delaunay M**, Prévot G, Collot S, Guilleminault L, Didier A, Mazières J. Management of pulmonary toxicity associated with immune checkpoint inhibitors. *Eur Respir Rev* 2019; **28** [PMID: 31694838 DOI: 10.1183/16000617.0012-2019]

48 **Naidoo J**, Page DB, Wolchok JD. Immune modulation for cancer therapy. *Br J Cancer* 2014; **111**: 2214-2219 [PMID: 25211661 DOI: 10.1038/bjc.2014.348]

49 **Jiang Y**, Zhang N, Pang H, Gao X, Zhang H. Risk and incidence of fatal adverse events associated with immune checkpoint inhibitors: a systematic review and meta-analysis. *Ther Clin Risk Manag* 2019; **15**: 293-302 [PMID: 30858709 DOI: 10.2147/TCRM.S191022]

50 **Nishino M**, Ramaiya NH, Awad MM, Sholl LM, Maattala JA, Taibi M, Hatabu H, Ott PA, Armand PF, Hodi FS. PD-1 Inhibitor-Related Pneumonitis in Advanced Cancer Patients: Radiographic Patterns and Clinical Course. *Clin Cancer Res* 2016; **22**: 6051-6060 [PMID: 27535979 DOI: 10.1158/1078-0432.CCR-16-1320]

51 **Chuzi S**, Tavora F, Cruz M, Costa R, Chae YK, Carneiro BA, Giles FJ. Clinical features, diagnostic challenges, and management strategies in checkpoint inhibitor-related pneumonitis. *Cancer Manag Res* 2017; **9**: 207-213 [PMID: 28652812 DOI: 10.2147/CMAR.S136818]

52 **Abou Alaiwi S**, Xie W, Nassar AH, Dudani S, Martini D, Bakouny Z, Steinharter JA, Nuzzo PV, Flippot R, Martinez-Chanza N, Wei X, McGregor BA, Kaymakcalan MD, Heng DYC, Bilen MA, Choueiri TK, Harshman LC. Safety and efficacy of restarting immune checkpoint inhibitors after clinically significant immune-related adverse events in metastatic renal cell carcinoma. *J Immunother Cancer* 2020; **8** [PMID: 32066646 DOI: 10.1136/jitc-2019-000144]

53 **Feins S**, Kong W, Williams EF, Milone MC, Fraietta JA. An introduction to chimeric antigen receptor (CAR) T-cell immunotherapy for human cancer. *Am J Hematol* 2019; **94**: S3-S9 [PMID: 30680780 DOI: 10.1002/ajh.25418]

54 **Kambhampati S**, Gray L, Fakhri B, Lo M, Vu K, Arora S, Kaplan L, Ai WZ, Andreadis C. Immune-related Adverse Events Associated With Checkpoint Inhibition in the Setting of CAR T Cell Therapy: A Case Series. *Clin Lymphoma Myeloma Leuk* 2020; **20**: e118-e123 [PMID: 31948859 DOI: 10.1016/j.clml.2019.12.014]

55 **Long B**, Brém E, Koyfman A. Oncologic Emergencies: Immune-Based Cancer Therapies and Complications. *West J Emerg Med* 2020; **21**: 566-580 [PMID: 32421502 DOI: 10.5811/westjem.2020.1.45898]

56 **Lee DW**, Gardner R, Porter DL, Louis CU, Ahmed N, Jensen M, Grupp SA, Mackall CL. Current concepts in the diagnosis and management of cytokine release syndrome. *Blood* 2014; **124**: 188-195 [PMID: 24876563 DOI: 10.1182/blood-2014-05-552729]

57 **Fajgenbaum DC**, June CH. Cytokine Storm. *N Engl J Med* 2020; **383**: 2255-2273 [PMID: 33264547 DOI: 10.1056/NEJMra2026131]

58 **National Cancer Institute Division of Cancer Treatment and Diagnosis**. Common Terminology Criteria for Adverse Events. [cited 13 January 2021]. Available from: https://ctep.cancer.gov/protocolDevelopment/electronic\_applications/docs/CTCAE\_v5\_Quick\_Reference\_5x7.pdf

59 **Gust J**, Ponce R, Liles WC, Garden GA, Turtle CJ. Cytokines in CAR T Cell-Associated Neurotoxicity. *Front Immunol* 2020; **11**: 577027 [PMID: 33391257 DOI: 10.3389/fimmu.2020.577027]

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**Table 1 Immunotherapy agents approved for treatment by the United States Food and Drug Administration in various types of cancers**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Organ system | Cancer | Tumor marker | Staging at initiation of immunotherapy | Immunotherapy |
| Brain and central nervous system | GBM | IDH mutations, 1p19q deletion, MGMT promoter methylation, EGFRVIII amplification | Recurrent GBM | Bevacizumab |
| Pediatric neuroblastoma | Homovanillic acid, Vanillylmandelic acid, LDH, NSE | High risk patients with partial response to first line agents | Dinutuximab |
| Head and neck | Head and neck SCC | CEA, SSC-Ag | Recurrent/metastatic SSC which progressed during/after platinum-based chemotherapy | Pembrolizumab |
| Breast | Breast cancer | BRCA1, BRCA2, CA 15-3, CA27.29, ER positive, PR positive, HER2/neu | HER2/neu positive | Trastuzumab |
| Unresectable locally advanced or metastatic triple-negative, PD-L1-positive breast cancer | Atezolizumab |
| Gastrointestinal | SCC of the Esophagus | CEA, CA 19-9, SSC-Ag | Locally advanced or metastatic SSC, that progressed after treatment with one or more lines of standard therapy | Pembrolizumab |
| Gastric and gastroesophageal junction cancer | CEA, CA 19-9, CA 72-4, DPD | Advanced cancer, that progressed despite two or more lines of standard treatment | Pembrolizumab |
| Gastric cancer | CEA, CA 19-9, CA 72-4, DPD | Advanced cancer | Nivolumab (only approved in Japan) |
| Pancreatic cancer | CA 19-9, DPD, CEA: not frequently used | Advanced cancer with high microsatellite instability or high tumor mutational burden | Pembrolizumab |
| Colorectal cancer | CEA, KRAS, BRAF V600, CA 19-9: not frequently used, DPD, MSU, dMMR | Progressive CRC after Fluoropyrimidine Oxaliplatin and irinotecan treatment regimen | Pembrolizumab |
| Hematological | CLL | Beta-2-microglobulin, chromosome 17p deletion | Relapsed CLL, in combination with Fludrabine and cyclophosphamide | Ofatumumab |
| Combination with Chlorambucil | Obinutuzumab |
| B-cell precursor ALL | BCR-ABL | Relapsed/refractory Ph-negative B-cell Precursor ALL | Blinatumomab |
| Follicular lymphoma | Beta-2-microglobulin | Relapse/refractory | Obinutuzumab |
| Hodgkin’s lymphoma | CD20 | Non-responsive to therapy or relapse after > 3 therapies | Pembrolizumab |
| Non-Hodgkin’s lymphoma | CD20 | Relapsed/progressing after autologous Hematopoietic stem cell transfer + Brentuximab Vedotrin or after 3 systemic therapies | Nivolumab |
| Multiple myeloma | Beta-2-microglobulin, immunoglobulins | (1) Initial treatment in combination with dexamethasone, in patients eligible for autologous HSCT and maintenance therapy after autologous HSCT; (2) After > 3 therapies or non-responsive to proteasome inhibitor and immunomodulatory drug; (3) Combination with lenalidomide + dexamethasone or bortezomib + dexamethasone after > 1 therapy; (4) Combination with revlimid + dexamethasone after 1-3 therapies | (1) Linalidomide; (2) Daratumab; (3) Daratumab; (4) Elotuzumab |
| Respiratory | NSCLC | ALK gene, BRAF V600, KRAS, PD-L1, ROS1 | Metastatic cancer expressing PD-L1 and progressing during/after platinum-based chemotherapy or with EGFR or ALK mutations | Pembrolizumab |
| In combination with pemetrexed and carboplatin, with or without PD-L1 expression | Pembrolizumab |
| Progressing during/after platinum-based chemotherapy | Nivolumab |
| Progressing cancer while using approved therapy for the mutation in metastatic cancer expressing PD-L1 and progressing during/after platinum-based chemotherapy or with EGFR or ALK mutations | Atezolizumab |
| Squamous NSCLC | First line in combination with gemcitabine + cisplatin | Necitumumab |
| Renal | Renal cell carcinoma |  | Advanced RCC after antiangiogenic therapy | Nivolumab |
| Skin | Melanoma | BRAF V600 | Unresectable cutaneous, subcutaneous or nodular lesions in relapsing melanoma after surgical resection | Talimogene Iaherparepvec (vaccine) |
| Unresectable/metastatic | Pembrolizumab |
| Unresectable/metastatic without BRAFV600 mutation | Nivolumab, Ipilimumab |
| Adjuvant therapy for stage III | Ipilimumab |
| Urinary | Bladder cancer | Bladder tumor antigen, chromosome 3, 7, 17, 9p21 mutation, FGFR2, FGFR3 mutation | Locally advanced or metastatic bladder cancer, progressed during/after platinum-based chemotherapy or within 12 mo neoadjuvant/adjuvant treatment | Avelumab, Durvalumab, Pembrolizumab, Nivolumab, Atezolizumab |
| Locally advanced or metastatic cancer with patient ineligible for platinum-based chemotherapy | Pembrolizumab, Atezolizumab |

GBM: Glioblastoma multiforme; SCC: Squamous cell carcinoma; SSC-Ag: Squamous cell carcinoma antigen; CLL: Chronic lymphocytic leukemia; ALL: Acute lymphoblastic leukemia; HSCT: Hematopoietic stem cell transfer; NSCLC: Non small-cell lung carcinoma.

**Table 2 Overview of some of the common immunotherapeutic agents and their proposed mechanisms**

|  |  |
| --- | --- |
| **Agent** | **Mechanism of action** |
| Ipilimumab | CTLA-4 inhibitor |
| Nivolumab | PD-1 inhibitor |
| Pembrolizumab | PD-1 inhibitor |
| Atezolizumab | PD-L1 inhibitor |
| Avelumab | PD-L1 inhibitor |
| Durvalumab | PD-L1 inhibitor |
| Talimogene Iaherparepvec | Cancer vaccine (directly destroys cancer cells, upregulates production of GM-CSF) |
| Necitumumab | EGFR inhibitor |
| Bevacizumab | VEGF inhibitor |
| Elotuzumab (anti-SLAMF7 monoclonal antibody) | Anti-SLAMF7 monoclonal antibody |
| Daratumumab (anti-CD38 monoclonal antibody) | Anti-CD38 monoclonal antibody |
| Lenalidomide | Immunomodulatory agent |
| Obinutuzumab | CD20 inhibitor |
| Ofatumumab | CD20 inhibitor |
| Blinatumomab | Bispecific T-cell engager |
| Trastuzumab | HER2/neu inhibitor |
| Dinutuximab | GD2-binding monoclonal antibody |

CTLA-4: Cytotoxic T-lymphocyte associated protein 4; PD-1: Programmed cell death protein 1; PD-L1: Programmed death-ligand 1.