World Journal of *Hepatology*

World J Hepatol 2021 April 27; 13(4): 393-521





Published by Baishideng Publishing Group Inc

World Journal of Hepatology

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INDEXING/ABSTRACTING

The WJH is now abstracted and indexed in PubMed, PubMed Central, Emerging Sources Citation Index (Web of Science), Scopus, China National Knowledge Infrastructure (CNKI), China Science and Technology Journal Database (CSTJ), and Superstar Journals Database. The WJH's CiteScore for 2019 is 5.8 and Scopus CiteScore rank 2019: Hepatology is 22/61.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Li-Li Wang, Production Department Director: Xiang Li, Editorial Office Director: Xiang Li.

NAME OF JOURNAL	INSTRUCTIONS TO AUTHORS
World Journal of Hepatology	https://www.wignet.com/bpg/gerinfo/204
ISSN	GUIDELINES FOR ETHICS DOCUMENTS
ISSN 1948-5182 (online)	https://www.wjgnet.com/bpg/GerInfo/287
LAUNCH DATE	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH
October 31, 2009	https://www.wignet.com/bpg/gerinfo/240
FREQUENCY	PUBLICATION ETHICS
Monthly	https://www.wjgnet.com/bpg/GerInfo/288
EDITORS-IN-CHIEF	PUBLICATION MISCONDUCT
Nikolaos Pyrsopoulos, Ke-Qin Hu, Koo Jeong Kang	https://www.wjgnet.com/bpg/gerinfo/208
EDITORIAL BOARD MEMBERS	ARTICLE PROCESSING CHARGE
https://www.wjgnet.com/1948-5182/editorialboard.htm	https://www.wjgnet.com/bpg/gerinfo/242
PUBLICATION DATE	STEPS FOR SUBMITTING MANUSCRIPTS
April 27, 2021	https://www.wjgnet.com/bpg/GerInfo/239
COPYRIGHT	ONLINE SUBMISSION
© 2021 Baishideng Publishing Group Inc	https://www.f6publishing.com

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World Journal of Hepatology

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World J Hepatol 2021 April 27; 13(4): 433-455

DOI: 10.4254/wjh.v13.i4.433

ISSN 1948-5182 (online)

ORIGINAL ARTICLE

Clinical and Translational Research Bile acid indices as biomarkers for liver diseases I: Diagnostic markers

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Author contributions: Alamoudi JA is the primary researcher, collected and analyzed data, wrote the manuscript, prepared figures and formatted manuscript for publication; Li W and Gautam N helped in the liquid chromatography-tandem mass spectrometry sample analysis; Meza J supervised, reviewed, and approved all statistical analysis and provided intellectual input and feedback on manuscript; Olivera M and Mukherjee S helped in recruiting and consenting patients and sample collection as well as experimental design; Alnouti Y is the primary investigator who was responsible for the experimental design and supervising all aspects of this project and manuscript preparation.

Supported by University of Nebraska Medical Center-Clinical Research Center and Great Plains

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Abstract

BACKGROUND

Hepatobiliary diseases result in the accumulation of toxic bile acids (BA) in the liver, blood, and other tissues which may contribute to an unfavorable prognosis.

AIM

To discover and validate diagnostic biomarkers of cholestatic liver diseases based on the urinary BA profile.

METHODS

We analyzed urine samples by liquid chromatography-tandem mass spectrometry and compared the urinary BA profile between 300 patients with hepatobiliary diseases vs 103 healthy controls by statistical analysis. The BA profile was characterized using BA indices, which quantifies the composition, metabolism, hydrophilicity, and toxicity of the BA profile. BA indices have much lower interand intra-individual variability compared to absolute concentrations of BA. In addition, BA indices demonstrate high area under the receiver operating



Health Research Consortium; and Society of American Gastrointestinal and Endoscopic Surgeons, No. 36-5360-2186-001.

Institutional review board

statement: The study was reviewed and approved by the University of Nebraska Medical Center Institutional Review Board (approval No. 487-10-EP).

Clinical trial registration statement:

This study is registered at ClinicalTrials.gov. The registration identification number is NCT01200082

Informed consent statement: All

study participants, or their legal guardian, provided informed written consent prior to study enrollment.

Conflict-of-interest statement: The authors declare that there is no conflict of interests in this study.

Data sharing statement: Technical appendix, statistical code, and data set available from the corresponding author at yalnouti@unmc.edu. Participants gave informed consent for data sharing.

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Manuscript source: Unsolicited manuscript

Specialty type: Gastroenterology and hepatology

Country/Territory of origin: United States

Peer-review report's scientific

characteristic curves, and changes of BA indices are associated with the risk of having a liver disease, which demonstrates their use as diagnostic biomarkers for cholestatic liver diseases.

RESULTS

Total and individual BA concentrations were higher in all patients. The percentage of secondary BA (lithocholic acid and deoxycholic acid) was significantly lower, while the percentage of primary BA (chenodeoxycholic acid, cholic acid, and hyocholic acid) was markedly higher in patients compared to controls. In addition, the percentage of taurine-amidation was higher in patients than controls. The increase in the non-12a-OH BA was more profound than 12a-OH BA (cholic acid and deoxycholic acid) causing a decrease in the 12α -OH/ non-12α-OH ratio in patients. This trend was stronger in patients with more advanced liver diseases as reflected by the model for end-stage liver disease score and the presence of hepatic decompensation. The percentage of sulfation was also higher in patients with more severe forms of liver diseases.

CONCLUSION

BA indices have much lower inter- and intra-individual variability compared to absolute BA concentrations and changes of BA indices are associated with the risk of developing liver diseases.

Key Words: Hepatobiliary diseases; Bile acids; Bile acid indices; Diagnosis; Biomarker; Liver diseases

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Core Tip: We have developed the concept of "bile acids (BA) indices" based on the detailed quantitative analysis of the urinary BA profile in patients with cholestatic liver diseases. We demonstrated the use of BA indices as diagnostic biomarkers for cholestatic liver diseases. BA indices had much lower inter- and intra-individual variability compared to absolute concentrations of the total and individual BA. In addition, BA indices demonstrated high area under the receiver operating characteristic curves, and changes of BA indices were associated with the risk of having a liver disease as determined by the logistic regression analysis.

Citation: Alamoudi JA, Li W, Gautam N, Olivera M, Meza J, Mukherjee S, Alnouti Y. Bile acid indices as biomarkers for liver diseases I: Diagnostic markers. World J Hepatol 2021; 13(4): 433-455

URL: https://www.wjgnet.com/1948-5182/full/v13/i4/433.htm **DOI:** https://dx.doi.org/10.4254/wjh.v13.i4.433

INTRODUCTION

Bile acids (BA) have many physiological functions such as cholesterol absorption and elimination, fat absorption, and maintenance of healthy microbiome^[1,2]. BA are also signaling molecules/hormones, which are involved in the regulation of their own homeostasis, thyroid hormone signaling, glucose and lipid metabolism, energy expenditure, and cellular immunity^[2-5]. Conversely, certain BA are also cytotoxic at high concentrations and have deleterious effects on hepatocytes and cholangiocytes, which play a major role in liver injury during various liver diseases^[5-8].

Cholestatic liver diseases are associated with a reduction in bile flow due to impairment of bile flow or defects in bile production^[9]. This causes accumulation of BA in the liver, which spills out into the systemic circulation, extrahepatic tissues, and eventually into urine. Numerous clinical and preclinical studies have shown up to a 100-fold increase in BA concentrations in the blood and urine during various liver diseases^[8,10-13]. Elevated BA concentrations were shown to correlate with the progression of damages to the liver and bile duct in cholestatic rats, rabbits, and in humans^[14-18].



quality classification

Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C, C Grade D (Fair): 0 Grade E (Poor): 0

Received: January 5, 2021 Peer-review started: January 5, 2021 First decision: January 25, 2021 Revised: February 11, 2021 Accepted: March 22, 2021 Article in press: March 22, 2021

Published online: April 27, 2021

P-Reviewer: Nakajima A S-Editor: Gao CC L-Editor: A P-Editor: Li JH



Biomarkers currently used in the clinic for the diagnosis and prognosis of liver diseases are primarily serum liver enzymes such as aspartate aminotransferase (AST) and alanine aminotransferase (ALT) as well as bilirubin^[19,20]. However, they are not specific to the liver or bile duct injuries, may increase in non-hepatobiliary diseases, and require severe cell injury at advanced disease stages before their blood levels increase^[19,20]. BA were extensively investigated for decades as biomarkers for numerous hepatobiliary diseases^[13,21-23]. However, these efforts never translated into the clinic, with the few exception of limited use in the diagnosis of intrahepatic cholestasis of pregnancy and biliary atresia in infants. This could be attributed to the marked differences in the physiological and pathological properties of the different individual BA. For example, detailed profiling of the more toxic and relevant individual BA rather than total BA concentration may better correlate with the liver condition during hepatobiliary diseases^[10,12,24]. Also, the extreme inter-and intraindividual variability of total and individual BA concentrations due to many factors such as food ingestion and diurnal variation, makes it challenging to determine the normal baseline ranges^[25,26].

We have developed the concept of "BA indices", which are ratios calculated from the absolute concentrations of individual BA and their metabolites (Table 1). These ratios provide comprehensive quantification of the composition, metabolism, hydrophilicity, formation of secondary BA, and toxicity of the BA profile^[9,26]. BA indices have much lower variability than the absolute BA concentrations used to calculate them. Indeed, we have demonstrated that BA indices offered numerous advantages over absolute total and individual BA concentrations including low interand intra-individual variability and were resistant to covariate influences such as age, gender, body mass index (BMI), food consumption, and moderate alcohol consumption^[9,26].

We have expanded on our previous pilot study, where we have recruited 300 patients with liver diseases and 103 control subjects over a period of 7 years. This study includes a series of two papers. In this article, we have shown the utility of BA indices as diagnosing markers for liver diseases by compared the urinary BA profile between healthy controls and patients and between patients with different severity levels of liver disease. In the 2nd article, we have built a survival model, the Bile Acid Score (BAS), to predict the prognosis of liver diseases using significant BA indices identified in this article.

MATERIALS AND METHODS

Study participants

For controls, 103 healthy subjects (32 male and 71 female) without liver diseases between the ages of 19 and 65 years were recruited by the Clinical Research Center at the University of Nebraska Medical Center (UNMC) (Omaha, NE, United States). The registry URL was (https://www.clinicaltrials.gov/ct2/show/NCT01200082?term=aln outi&draw=2&rank=1). The clinical trial number was NCT01200082. Inclusion criteria for the healthy controls included normal liver functions, as verified by ALT < 50 U/L, AST < 56 U/L, gamma-glutamyl transferase < 78 U/L, absence of diabetes, and no- or moderate alcohol drinking^[27] The study was approved by Institutional Review Board at UNMC and written informed consents were provided for all participating subjects. Thirty milliliters urine samples were collected from controls at fasting conditions in the first visit, and 1, 2, and 4 wk thereafter.

Patients diagnosed with one or multi-hepatobiliary conditions due to chronic hepatitis C (n = 71), hepatitis B (n = 15), alcoholic liver disease/alcoholic cirrhosis (n = 15) 117), primary biliary cholangitis (PBC) (n = 12), primary sclerosing cholangitis (n = 17), autoimmune hepatitis (n = 27), alpha-1-antitrypsin deficiency (n = 6), nonalcoholic fatty liver disease/nonalcoholic steatohepatitis (n = 56), carcinoma (n = 26), cryptogenic cirrhosis (n = 11), polycystic liver disease (n = 5), elevated liver function test (LFT) (n = 22), and unknown etiology (n = 5), were enrolled in the hepatology clinic in UNMC. A total of 300 patients (157 male and 143 female) between the ages of 19 years and 83 years were recruited. Thirty milliliters of urine samples were collected on their first and follow-up visits to the hepatology clinic. All urine samples were stored in -80 °C until analyzed by liquid chromatography-tandem mass spectrometry (LC-MS/MS). Patients were divided into three disease-severity groups based on their model for end-stage liver disease (MELD) score: low-MELD (6-15 score), medium-MELD (16-25), and high-MELD (26-40). High MELD group was not included while performing the statistical analysis, because there were only four subjects in that group.



Table 1 List of bile acid indices							
Composition	Hepatic metabolism	Hydrophilicity	CYP8B1 activity	Intestinal contribution			
Concentration of individual BA	Total sulfated	Total mono-OH	Total 12α-OH	Total primary			
% of individual BA	Total G-amidated	Total di-OH	Total non-12α-OH	Total secondary			
	Total T-amidated	Total tri-OH	12α-OH/non-12α-OH	Primary/secondary			
	% Sulfation	% Mono-OH	CA/CDCA	% Primary			
	% Amidation	% Di-OH	% 12а-ОН	% Secondary			
	% G-amidation	% Tri-OH	% Non-12a-OH				
	% T-amidation						

BA: Bile acids; G: Glycine; T: Taurine; CDCA: Chenodeoxycholic acid; CA: Cholic acid.

In addition, patients were also categorized according to hepatic decompensation (presence or history of encephalopathy, bleeding varices, ascites, or jaundice)^[28].

Non-BA parameters

AST, ALT, albumin, and serum creatinine were measured using the Beckman Coulter reagents (Beckman Coulter, Inc, Brea, California). Protime and international normalized ratio (INR) were measured using STANeoplastine "CI PLUS 10" reagent kit (Diagnostica Stago Inc, Parsippany, New Jersey). Total bilirubin in serum was analyzed using QuantiChromTM Bilirubin assay kit (BioAssay Systems, Hayward, CA, United States). AST/ALT ratio and AST/platelet ratio index (APRI) were calculated.

BA quantification by LC-MS/MS

Urine samples were extracted using solid phase extraction as described previously^[9,26,29,30]. BA concentrations were quantified by LC-MS/MS, as we described previously^[31].

Calculation of BA indices

In addition to the absolute concentration of individual and total BA, the BA profile in urine was characterized using "BA indices" (Table 1), and as we have described previously^[9,26,30,31]. BA indices describe the composition, hydrophilicity, formation of 12a-OH BA by CYP8B1, metabolism, and formation of secondary BA by intestinal bacteria. The composition indices were calculated as the ratio of the concentration of individual BA in all of their forms (sulfated, unsulfated amidated, and unamidated) to the total concentration of BA. The percentages of mono-OH BA: [lithocholic acid (LCA)], di-OH BA: [ursodeoxycholic acid (UDCA), murideoxycholic acid (MDCA), chenodeoxycholic acid (CDCA), hyodeoxycholic acid (HDCA), and deoxycholic acid (DCA)], and tri-OH BA: [cholic acid (CA), muricholic acid (MCA), and hyocholic acid (HCA)] were calculated as the ratio of the concentration of the sum of the respective BA in all their forms to the total concentration of BA.

The 12a-OH BA are formed by CYP8B1 in the liver and include DCA, CA, nor-DCA, and 3-dehydroCA. Therefore, CYP8B1 activity can be measured by the ratio of 12a-OH BA to the remaining of all other BA (non-12α-OH BA). Another marker for CYP8B1 is the ratio of CA to CDCA because CA is formed by the 12a hydroxylation of CDCA. In the same way, the ratio of 12a-OH (DCA, CA, nor-DCA, and 3-dehydroCA in all of their forms) to non-12a-OH (CDCA, HDCA, LCA, UDCA, MDCA, HCA, MCA, 12oxo-CDCA, 6-oxo-LCA, 7-oxo-LCA, 12-oxo-LCA, isoLCA, isoDCA in all of their forms) was calculated.

BA are metabolized primarily by sulfation, glycine (G), and taurine (T) amidation in the liver. The percentage of individual BA sulfation was calculated as a ratio of the concentration of sulfated BA, in both the amidated and unamidated forms, to the total individual BA concentration in all their forms (amidated, unamidated, sulfated, and unsulfated). In both the sulfated and unsulfated forms, the percentage of individual BA amidation have been calculated as the ratio of the concentration of amidated BA, to the total concentration of individual BA in all of their forms (amidated, unamidated, sulfated, and unsulfated). Additionally, percentages of amidation were divided into the percentages of BA existing as G or as T amidates.

The ratio of primary (CA, CDCA, MCA and HCA in all of their forms) to secondary BA (DCA, LCA, UDCA, HDCA, MDCA, Nor-DCA, 12-oxo-CDCA, 3-dehydroCA, 6-oxo-LCA, 7-oxo-LCA, 12-oxo-LCA, isoLCA, and isoDCA in all of their forms) was calculated.

Statistical analysis

Independent sample-*t*-test and Mann-Whitney test were used to study the demographic differences between controls and patients because the sample size was > $30^{[32]}$. Independent sample-*t*-test was used for continuous variables and Mann-Whitney test was used for categorical variables. The demographic variables were (age, BMI, gender, and race). Subjects were divided into four age groups (19-29, 30-41, 42-53, 54-83 years), and the variable age was studied as both a continuous and a categorical variable. Subjects were also divided into three BMI groups (normal: BMI < 25, overweight: BMI 25-29.9, and obese: BMI ≥ 30) and the effect of BMI was studied as both a continuous and a categorical variable. Also, subjects were divided into five race groups (White, Black, Asian, Hispanic, others), and the variable race was studied as a categorical variable.

Urine samples were collected from controls and patients on their first visit and follow-up visits. Mixed effects models were used to compare patients *vs* controls and the demographic variables were included as covariates. Statistically significant covariates were returned to the mixed effects models as interaction terms with the primary group, *i.e.*, patients *vs* control.

BA indices were compared between controls, low-MELD (patients), and medium-MELD (patients) groups using mixed effects models followed by pairwise comparisons using Bonferroni's adjustment if the P value was < 0.05. BA indices were compared between compensated and decompensated patients using mixed effects models. Mixed effects models were also used to determine the association between non-BA parameters including (AST, ALT, bilirubin, MELD score, AST/ALT, creatinine, INR, APRI, protime, and albumin) and BA indices. Receiver operating characteristic curve (ROC) analyses were used to determine cut-off values of BA as markers for the diagnosis of liver diseases with optimum sensitivity and specificity. The areas under the ROC curve (AUC) values were compared between urinary BA profiles and non-BA parameters. The mixed effects models were used to compare BA indices with AUC > 0.7 between controls and the patients with specific disease subtypes described in the "Study Participants" section (same patients can belong to different disease groups). Polycystic liver disease and unknown etiology subtypes were not included in the comparison between the disease subtypes because they had < six subjects.

Univariate logistic regression analysis was used to determine the association between BA concentrations and indices and the likelihood of developing a liver disease. From logistic regression analysis, the odds ratios (ORs) were calculated for a 10% and 20% change from the mean value of BA indices in the healthy controls.

P value of 0.05 was considered significant for all the statistical tests described above. All statistical analysis was performed using the Statistical Product and Service Solutions (SPSS) software, version 25 (IBM corporation, Armonk, NY, United States).

RESULTS

Demographics

Table 2 shows a summary of the demographics of both patients and controls participants. We enrolled 103 controls (32 males and 71 females) and 300 patients (157 males and 143 females), who were treated for cholestatic liver diseases in UNMC, over the period from November of 2011 to December of 2018. To compare the demographics between the two groups, age and BMI covariates were compared as both continuous and categorical variables using *t*-test, and Mann-Whitney test, respectively. While gender and race were compared as categorical variables using Mann-Whitney test. Age, gender, and BMI were significantly different between control and patients (*P* value < 0.05), while race was not different. Therefore, the statistically significant demographic variables (age, BMI, and gender) were included as covariates in the mixed effects models to compare BA indices between patients and controls.

Table 2 Demographics		
	Controls	Patients
n	103	300
Gender ¹		
Male, female	32, 71	157, 143
Age (yr) ¹		
mean ± SE	44.3 ± 0.64	52.1± 0.54
19-29	17	11
30-41	28	40
42-53	30	92
54-83	28	157
BMI ¹		
mean ± SE	27.5 ± 0.28	30.9 ± 0.32
Normal BMI < 25	30	69
Overweight BMI = 25-29.9	45	104
Obese BMI ≥ 30	28	127
Race		
White	88	247
Black	7	14
Asian	7	13
Hispanic	1	8
Others	0	18

¹Significant difference between controls and patients (P < 0.05). BMI: Body mass index.

Differences in BA between patients vs controls are not due to differences in demographics

Because some of the covariates (age, BMI, and gender) were significantly different between the two groups (Table 2), we reran the univariate mixed effect analysis with these covariates (multivariate analysis). First, association between these covariates and BA indices was identified, and then the covariates with significant association with BA indices were incorporated in the multivariate mixed effect analyses as interaction terms with the group (patients and controls). We did not find any difference in the association between covariates and BA indices between the two groups except for the % primary and % secondary BA with gender (Supplementary Table 1).

BA profiles in controls vs patients

Table 3 shows the absolute concentrations of major urinary BA in controls and patients. Table 4 compares representative absolute BA concentrations and indices between controls and patients. Supplementary Table 2 shows the full list of BA concentrations and indices. Total BA was 5.9-fold higher in patients compared with controls. All individual BA concentrations were also higher in patients, except MDCA, but to different extents. The highest increase was in UDCA (11.9-fold), while the lowest increase was for DCA and HDCA (1.6-fold). The percentage of UDCA, CDCA, MCA, CA, and HCA were higher (1.2-1.6-fold), while the percentage of LCA, DCA, HDCA, and MDCA were lower (0.5-0.8-fold) in patients *vs* controls.

Unamidated, G-amidated, and T-amidated BA which were 3.3-, 5.9-, and 9.4-fold higher in patients than controls. Therefore, the overall % amidation and % G-amidation did not change or slightly decreased in patients, whereas % T-amidation increased from 8.0% in controls to 10.8% in patients. Similarly, the concentrations of both sulfated and unsulfated were approximately 6-fold higher in patient; so that the

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Table 3 Absolute concentrations of major bile acids in controls and patients						
DA	Unamidated	G-BA	T-BA	Total		
ВА	mean ± SE, µmol/L					
Controls						
Unsulfated BA						
LCA	0.000 ± 0.00	0.000 ± 0.00	0.000 ± 0.00	0.001 ± 0.00		
UDCA	0.004 ± 0.00	0.033 ± 0.00	0.002 ± 0.00	0.038 ± 0.00		
CDCA	0.003 ± 0.00	0.008 ± 0.00	0.002 ± 0.00	0.013 ± 0.00		
DCA	0.022 ± 0.00	0.011 ± 0.00	0.002 ± 0.00	0.035 ± 0.00		
HDCA	0.01 ± 0.00	0.00 ± 0.00	ND	0.007 ± 0.00		
MDCA	0.060 ± 0.01	ND	ND	0.058 ± 0.01		
CA	0.179 ± 0.03	0.067 ± 0.00	0.009 ± 0.00	0.255 ± 0.03		
MCA	0.028 ± 0.00	0.287 ± 0.02	0.041 ± 0.00	0.356 ± 0.02		
НСА	0.008 ± 0.00	0.016 ± 0.00	0.001 ± 0.00	0.026 ± 0.00		
Other BA ¹	0.160 ± 0.01	-	-	0.160 ± 0.01		
Total unsulfated	0.464 ± 0.04	0.422 ± 0.02	0.057 ± 0.00	0.943 ± 0.05		
Sulfated BA						
LCA	0.010 ± 0.00	0.780 ± 0.04	0.220 ± 0.01	1.010 ± 0.05		
UDCA	0.450 ± 0.02	1.040 ± 0.05	0.030 ± 0.00	1.520 ± 0.07		
CDCA	0.070 ± 0.01	2.380 ± 0.13	0.060 ± 0.00	2.510 ± 0.13		
DCA	0.010 ± 0.00	2.900 ± 0.14	0.220 ± 0.02	3.130 ± 0.16		
CA	0.004 ± 0.00	0.056 ± 0.01	0.126 ± 0.01	0.190 ± 0.01		
Total sulfated	0.535 ± 0.03	7.170 ± 0.28	0.650 ± 0.03	8.350 ± 0.31		
Overall total	1.000 ± 0.05	7.590 ± 0.29	0.710 ± 0.03	9.300 ± 0.33		
Patients						
Unsulfated BA						
LCA	0.004 ± 0.00	0.001 ± 0.00	0.0001 ± 0.00	0.005 ± 0.00		
UDCA	0.079 ± 0.03	0.410 ± 0.17	0.012 ± 0.00	0.500 ± 0.21		
CDCA	0.020 ± 0.00	0.090 ± 0.01	0.100 ± 0.02	0.210 ± 0.03		
DCA	0.040 ± 0.00	0.040 ± 0.00	0.010 ± 0.00	0.090 ± 0.01		
HDCA	0.010 ± 0.00	0.00 ± 0.00	ND	0.010 ± 0.00		
MDCA	0.050 ± 0.01	ND	ND	0.050 ± 0.01		
CA	0.240 ± 0.03	0.550 ± 0.07	0.320 ± 0.08	1.120 ± 0.14		
MCA	0.120 ± 0.02	1.940 ± 0.29	0.730 ± 0.09	2.790 ± 0.34		
НСА	0.010 ± 0.00	0.170 ± 0.02	0.090 ± 0.02	0.270 ± 0.04		
Other BA ¹	0.860 ± 0.13	-	-	0.860 ± 0.13		
Total	0.460 ± 0.04	0.42 ± 0.02	0.06 ± 0.00	5.910 ± 0.57		
Sulfated BA						
LCA	0.030 ± 0.01	2.230 ± 0.20	0.650 ± 0.06	2.910 ± 0.24		
UDCA	1.560 ± 0.23	15.30 ± 2.68	1.230 ± 0.27	18.10 ± 3.08		
CDCA	0.190 ± 0.03	18.70 ± 1.79	1.910 ± 0.38	20.80 ± 2.07		
DCA	0.040 ± 0.01	4.280 ± 0.54	0.520 ± 0.07	4.840 ± 0.58		
СА	0.080 ± 0.01	0.910 ± 0.13	1.030 ± 0.21	2.010 ± 0.31		

Total	1.900 ± 0.24	41.40 ± 4.12	5.340 ± 0.74	48.70 ± 4.77
Overall total	3.330 ± 0.33	44.60 ± 4.46	6.610 ± 0.85	54.60 ± 5.20

¹Other bile acids: Nor-deoxycholic acid, 12-oxo-chenodeoxycholic acid, 3-dehydrocholic acid, 6-oxo-lithocholic acid, 7-oxo-lithocholic acid, 12-oxo-lithocholic acid, isolithocholic acid, and isodeoxycholic acid.

ND: Not detected; -: Not quantified; BA: Bile acids; G: Glycine; T: Taurine; CDCA: Chenodeoxycholic acid; CA: Cholic acid; LCA: Lithocholic acid; UDCA: Ursodeoxycholic acid; DCA: Deoxycholic acid; HDCA: Hyodeoxycholic acid; MDCA: Murideoxycholic acid; MCA: Muricholic acid; HCA: Hyocholic acid; MDCA: Murideoxycholic acid; MDCA: Murideoxy

% sulfation of BA was unchanged.

The absolute concentrations of mono-, di-, and tri-OH BA were also higher in patients compared with controls, but the % mono-OH decreased (0.8-fold), di-OH remained unchanged, and % tri-OH increased (1.4-fold) due to increasing % CA (1.2-fold), % MCA (1.6-fold), and % HCA (1.5-fold).

Total 12 α -OH and non-12 α -OH BA were 2.3-fold and 8.2-fold higher in patients, so that the ratio of 12 α -OH/ non-12 α -OH and the % 12 α -OH decreased (approximately 0.5-fold), while % non-12 α -OH BA increased (1.2-fold).

Total primary and secondary BA were 8.1-fold and 4.6-fold higher in patients, so that the ratio of primary/secondary BA was 3.6-fold higher. Therefore, % primary BA was 1.4-fold higher, while % secondary BA was 0.80-fold lower in patients *vs* controls.

BA profile in low vs medium-MELD patients

Table 5 compares representative urinary BA concentrations and indices between lowand medium-MELD patients. Total BA concentrations was twice and individual BA concentrations were (1.15-fold to 3.9-fold) higher in medium *vs* low-MELD patients (Table 5).

Unamidated BA concentration was lower, while G-amidated and T-amidated BA were higher in the medium-MELD patients. Therefore, % T-amidation was 1.5-fold higher, while there was minimal difference in the % amidation and % G-amidation between medium and low-MELD patients. Similarly, the concentrations of both sulfated and unsulfated were 1.3- and 2-fold higher in medium *vs* low-MELD. On the other hand, the % sulfation of BA was only 1.07-fold higher, but it was statistically significant.

The absolute concentrations of mono-, di-, and tri-OH BA were also (1.8-2-fold) higher in medium-MELD patients, but the % mono-OH decreased (0.86-fold); while % di- and % tri-OH remained unchanged.

Total 12 α -OH and non-12 α -OH BA were both higher in medium *vs* low-MELD patients, but to different extents so that % non-12 α -OH BA remained unchanged, while % 12 α -OH decreased and the ratio of 12 α -OH/ non-12 α -OH was approximately 0.7-fold lower.

Total primary BA were 3.4-fold higher, while total secondary BA were slightly (0.9-fold) lower in medium-MELD patients, so that the ratio of primary/secondary BA was 2.3-fold higher. Similarly, % primary BA was 1.4-fold higher, while % secondary BA was 0.6-fold lower in medium- MELD patients.

BA profile in compensated vs decompensated patients

Table 6 compares representative urinary BA concentrations and indices between decompensated and compensated patients. In general, the same trend in the higher *vs* lower MELD patients comparison was observed in the decompensated *vs* compensated patients. Total BA was 1.3-fold higher, all individual BA were higher, but to variable extents. The percentage of CDCA, HDCA, CA, and HCA were higher (1.3-2.1-fold), while the percentage of LCA, UDCA, DCA, and MDCA were lower (0.3-0.7-fold) in decompensated *vs* compensated patients.

The % T-amidation was 1.3-fold higher in decompensated vs. compensated patients, while there was no difference in the % amidation, % G-amidation, or % sulfation. The % mono-OH decreased (0.73-fold), % di-OH remained unchanged, and % tri-OH slightly increased (1.13-fold) due to increasing % CA and % HCA. The ratio of 12 α -OH/ non-12 α -OH lower, the % 12 α -OH, and CA/CDCA ratio decreased (0.7-0.8-fold), while % non-12 α -OH BA remained unchanged.

Total primary BA were two-fold higher, while total secondary BA were 0.8-fold lower, so that the ratio of primary/secondary BA was 2.6-fold higher in decompensated patients. Therefore, % primary BA was 1.5-fold higher, while % secondary BA was 0.56-fold lower in decompensated patients.

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Table 4 Representative bile acids concentrations and indices in controls vs patients							
	Controls		Patients	Patients		Patients vs controls	
BA (µmoi/L) or BA indices	mean	SE	mean	SE	Ratio	<i>P</i> value	
Total BA	9.30	0.33	54.6	5.20	5.87	0.000	
Total LCA	1.01	0.05	2.92	0.24	2.88	0.000	
Total UDCA	1.56	0.07	18.6	3.23	11.9	0.001	
Total CDCA	2.52	0.13	21.0	2.09	8.35	0.000	
Total DCA	3.16	0.16	4.92	0.58	1.56	0.072	
Total HDCA	0.01	0.00	0.01	0.00	1.57	0.051	
Total MDCA	0.06	0.01	0.05	0.01	0.90	0.992	
Total CA	0.44	0.03	3.13	0.44	7.09	0.003	
Total MCA	0.36	0.02	2.79	0.34	7.83	0.000	
Total HCA	0.03	0.00	0.27	0.04	10.6	0.001	
Other BA ¹	0.16	0.01	0.86	0.13	5.54	NA	
% LCA	11.5	0.38	9.20	0.39	0.79	0.002	
% UDCA	17.7	0.49	21.3	0.88	1.21	0.138	
% CDCA	27.1	0.65	36.3	0.94	1.34	0.000	
% DCA	31.1	0.68	14.6	0.53	0.47	0.000	
% HDCA	0.07	0.01	0.04	0.00	0.54	0.052	
% MDCA	0.64	0.04	0.36	0.05	0.56	0.135	
% CA	5.25	0.27	6.27	0.25	1.19	0.064	
% MCA	4.03	0.16	6.39	0.34	1.58	0.003	
% HCA	0.30	0.02	0.45	0.04	1.52	0.018	
Total Unamidated	1.00	0.05	3.33	0.33	3.34	0.000	
Total G-amidated	7.59	0.29	44.6	4.46	5.88	0.000	
Total T-amidated	0.71	0.03	6.61	0.85	9.37	0.001	
% Amidation	87.7	0.47	86.9	0.65	0.99	0.053	
% G-amidation	79.7	0.49	76.0	0.71	0.95	0.000	
% T-amidation	7.98	0.26	10.8	0.46	1.35	0.005	
Total Unsulfated	0.94	0.05	5.91	0.57	6.26	0.000	
Total Sulfated	8.35	0.31	48.7	4.77	5.83	0.000	
% Sulfation	88.5	0.46	82.9	0.60	0.94	0.000	
Total Mono-OH	1.01	0.05	2.92	0.24	2.88	0.000	
Total Di-OH	7.30	0.29	44.6	4.58	6.11	0.000	
Total Tri-OH	0.82	0.04	6.19	0.65	7.52	0.000	
% Mono-OH	11.5	0.38	9.16	0.39	0.79	0.002	
% Di-OH	76.6	0.50	72.7	0.65	0.95	0.001	
% Tri-OH	9.58	0.33	13.1	0.43	1.37	0.000	
Total 12α-OH	3.62	0.17	8.35	0.83	2.30	0.001	
Total non-12α-OH	5.67	0.20	46.2	4.68	8.15	0.000	
12a-OH/non12a-OH	0.65	0.02	0.33	0.01	0.51	0.000	
CA/CDCA	0.24	0.01	0.24	0.02	1.00	0.625	
% 12α-ΟΗ	36.7	0.62	22.1	0.54	0.60	0.000	

% non-12a-OH	63.3	0.62	77.9	0.54	1.23	0.000
Total Primary	3.34	0.15	27.2	2.59	8.15	0.000
Total Secondary	5.95	0.23	27.4	3.52	4.59	0.000
Primary/ Secondary	0.69	0.03	2.52	0.22	3.63	0.000
% Primary	36.7	0.70	49.4	1.06	1.35	0.000
% Secondary	63.3	0.70	50.6	1.06	0.80	0.000

¹Other bile acids: Nor-deoxycholic acid, 12-oxo-chenodeoxycholic acid, 3-dehydrocholic acid, 6-oxo-lithocholic acid, 7-oxo-lithocholic acid, 12-oxolithocholic acid, isolithocholic acid, and isodeoxycholic acid.

NA: Not available; BA: Bile acids; G: Glycine; T: Taurine; CDCA: Chenodeoxycholic acid; CA: Cholic acid; LCA: Lithocholic acid; UDCA: Ursodeoxycholic acid; DCA: Deoxycholic acid; HDCA: Hyodeoxycholic acid; MDCA: Murideoxycholic acid; MCA: Muricholic acid; HCA: Hyocholic acid.

ROC curve analysis

Supplementary Table 3 lists the AUC for BA concentrations and indices. Supplementary Table 4 shows the full list of BA concentrations and indices. Total BA, CDCA, CA, % DCA, % HDCA, % MDCA, total G-Amidated, total unsulfated, total sulfated, total di-OH, total tri-OH, total non-12a-OH, 12a-OH/non12a-OH, % 12a-OH, % non-12α-OH, total primary, primary/secondary, % primary, and % secondary produced AUC > 0.7. Figure 1 shows ROC curves of BA indices with AUC > 0.7. Potential cut-off values selected based on the optimum specificity and sensitivity for BA indices with AUC > 0.7 are listed in Supplementary Table 5.

Risk analysis: Logistic regression analysis

Table 7 shows the results of logistic regression analyzes for BA indices with ROC (AUC) > 0.7. Logistic regression analysis detects whether there is a risk of liver disease associated with changes in BA indices. The risk of liver disease increased with changing levels of all BA indices (P < 0.05) except (% HDCA and % MDCA). Additionally, the OR from the logistic regression analysis quantifies the magnitude of the risk of developing liver diseases per unit (10% and 20% of the normal value) changes in BA indices. For example, for every 20% increase in the % non-12a-OH BA, the likelihood of having a liver disease increases 2.72-folds (OR: 2.72; P < 0.05). In contrast for every 20% increase in the % 12a-OH BA, the likelihood of having a liver disease decreases 0.56-folds (OR: 0.56; *P* < 0.05).

BA profile in different liver disease subtypes

Table 8 compare BA indices with ROC-AUC > 0.7 between controls vs patients with specific liver disease subtype. Mixed effects models were used to compare disease subtypes individually vs controls. The goal was to identify BA indices that can serve as diagnostic biomarkers for specific liver disease subtypes.

We have found that most BA indices were significantly different between controls vs all individual liver disease subtypes. Total BA, total CDCA, total CA, total Gamidated, total unsulfated, total sulfated, total di-OH, total tri-OH, Total non-12a-OH, % non-12α-OH and total primary were higher (1.1- to 39.5-fold) in every liver disease group compared with controls. % Primary and primary/secondary were higher (1.1fold to 9.27-fold) in all liver disease group compared with controls except in PBC. % DCA, % HDCA, % 12a-OH, and 12a-OH/non-12a-OH were lower (0.07-fold to 0.85fold) in every liver disease group compared with controls. % MDCA and % secondary was lower in all liver disease group compared with controls except in elevated LFT and PBC, respectively.

Non-BA parameters

In addition to BA indices, we have also examined other biomarkers currently used in the clinic to evaluate liver functions. These non-BA parameters include AST, ALT, AST/ALT, bilirubin, albumin, INR, protime, creatinine, APRI, and MELD. Table 9 compares the non-BA parameters in controls and patients using mixed effects models. All the non-BA parameters were higher in patients compared to controls except albumin and protime, which were lower in patients. Within the patient population, all non-BA parameters were higher in medium compared to low- MELD patients except albumin, and ALT. The same results also applied to decompensated vs compensated patients.

The AUC for non-BA parameters was > 0.7 for all of them except creatinine,



Table 5 Representative bile acids concentrations and indices in medium- vs low- model for end-stage liver disease patients						
	Low-MELD		Medium-MELD		Medium- vs low-MELD	
BA (µmol/L) or BA indices	mean	SE	mean	SE	Ratio	P value
Total BA	59.2	7.94	116	24.8	1.96	1.000
Total LCA	3.40	0.35	6.01	1.72	1.77	0.175
Total UDCA	24.4	5.34	18.6	6.30	0.76	0.172
Total CDCA	18.3	2.31	71.4	16.3	3.90	0.000
Total DCA	5.30	0.96	6.08	1.47	1.15	1.000
Total HDCA	0.01	0.00	0.01	0.00	0.61	1.000
Total MDCA	0.05	0.01	0.06	0.01	1.28	1.000
Total CA	2.80	0.48	10.6	4.45	3.79	0.000
Total MCA	3.58	0.57	2.15	0.46	0.60	0.210
Total HCA	0.25	0.04	0.86	0.36	3.48	0.002
% LCA	9.31	0.53	7.97	1.47	0.86	1.000
% UDCA	23.1	1.29	14.3	2.52	0.62	1.000
% CDCA	34.7	1.21	55.6	3.17	1.60	0.000
% DCA	13.8	0.65	7.18	1.33	0.52	0.005
% HDCA	0.04	0.01	0.01	0.00	0.32	0.661
% MDCA	0.29	0.04	0.13	0.03	0.43	1.000
% CA	5.75	0.30	8.70	1.25	1.51	0.145
% MCA	7.15	0.48	3.70	0.91	0.52	0.000
% HCA	0.46	0.07	0.75	0.15	1.61	0.148
Total Unamidated	4.24	0.55	2.87	0.72	0.68	0.062
Total G-amidated	48.4	6.89	92.8	19.7	1.92	1.000
Total T-amidated	6.58	1.04	20.7	7.30	3.15	0.040
% Amidation	86.7	0.87	94.4	1.28	1.09	0.005
% G-amidation	75.5	0.96	77.2	2.73	1.02	1.000
% T-amidation	11.2	0.64	17.1	2.15	1.53	0.002
Total unsulfated	6.99	0.93	9.04	2.42	1.29	1.000
Total sulfated	52.3	7.21	107	23.2	2.05	1.000
% Sulfation	82.4	0.81	88.3	1.34	1.07	0.009
Total mono-OH	3.40	0.35	6.01	1.72	1.77	0.175
Total di-OH	48.1	7.01	96.2	20.9	2.00	1.000
Total tri-OH	6.63	0.90	13.6	4.90	2.06	0.301
% Mono-OH	9.31	0.53	7.97	1.47	0.86	1.000
% Di-OH	72.0	0.90	77.2	2.14	1.07	0.058
% Tri-OH	13.4	0.59	13.1	1.40	0.98	0.274
Total 12α-OH	8.55	1.23	16.8	4.86	1.96	0.053
Total non-12α-OH	50.7	7.21	99.6	21.5	1.96	1.000
12a-OH/non12a-OH	0.30	0.01	0.20	0.02	0.68	0.135
CA/CDCA	0.21	0.01	0.17	0.03	0.81	1.000
% 12a-OH	21.0	0.69	16.1	1.44	0.77	0.008
% non-12a-OH	79.0	0.69	83.9	1.44	1.06	0.008

Total primary	25.0	3.08	85.1	19.5	3.41	0.000
Total secondary	34.3	5.78	31.3	8.05	0.91	0.316
Primary/secondary	2.19	0.24	5.02	1.16	2.29	1.000
% Primary	48.1	1.40	68.7	3.10	1.43	0.014
% Secondary	51.9	1.40	31.3	3.10	0.60	0.014

BA: Bile acids; MELD: Model for end-stage liver disease; G: Glycine; T: Taurine; CDCA: Chenodeoxycholic acid; CA: Cholic acid; LCA: Lithocholic acid; UDCA: Ursodeoxycholic acid; DCA: Deoxycholic acid; HDCA: Hyodeoxycholic acid; MDCA: Murideoxycholic acid; MCA: Muricholic acid; HCA: Hyocholic acid.

> protime, and AST/ALT ratio. Also, per logistic regression analysis, the risk of being diagnosed with a liver disease increased to various extents with changing levels of all non-BA parameters (P < 0.05) except creatinine and AST/ALT. For example, for every 20% increase in the albumin and protime, the likelihood of having a liver disease decreases 0.28-fold and 0.85-fold, respectively. In contrast for every 20% increase in the other non-BA parameters, the likelihood of having a liver disease increases 1.13-fold to 3-fold (Supplementary Table 6).

> In addition, we have found that most non-BA parameters were significantly different between controls vs. all individual liver disease subtypes (Supplementary Table 7). Creatinine, INR, AST, ALT, bilirubin, AST/ALT, and MELD were higher in most liver disease group compared with controls. In contrast, albumin and protime were lower in most liver disease group compared with controls.

Association between non-BA parameters and BA indices

Supplementary Table 8 shows the association between non-BA parameters and BA indices using mixed effects models. We have found that all non-BA parameters were significantly associated with most BA concentrations/indices, except creatinine (P >0.05).

DISCUSSION

To ensure that the difference in the BA profiles between patients and controls are not due to the differences in the demographics we showed that: (1) Most of BA were not associated with demographic covariates, and (2) The ones that were associated had the same extent of association in the patient and control groups (Supplementary Table 1).

Patients were categorized based on the severity of the liver disease using MELD^[33-37] and the compensation status^[28]. Accordingly, we have compared the BA profiles between entire patient vs control populations as well as among the patients with different levels of disease severity. Most BA (except MDCA) were higher, but to different extents, in patients vs controls (Table 4) and in the more-severe patient groups, *i.e.*, medium vs low-MELD (Table 5) as well as decompensated vs compensated (Table 6). In particular, the percentages of the primary BA (CDCA, CA, and HCA) were higher, while the percentage of the secondary BA (DCA) was lower. The % primary BA was 1.4-fold higher, while % secondary BA was 0.8-fold lower and the ratio of primary/secondary BA was 3.6-fold higher in patients vs controls (Table 4). The same trend was also observed in the patients with more severe form of the disease, where the % of primary BA also increased with the severity of the liver disease (medium-MELD > low-MELD > controls) and (decompensated > compensated > controls), whereas % secondary BA decreased with the severity of the disease. (Tables 5 and 6).

Cholestatic diseases are associated with impaired bile flow to the intestine, which translates into reduced transformation of primary into secondary BA by intestinal bacteria^[9,25,38-40]. Therefore, while all BA concentrations were higher in patients due to the impairment of bile flow, the proportion of secondary BA (formed in the intestine) decreased with the severity of the cholestatic disease, which may reflect the extent of bile flow impairment.

The conjugation of BA with G and T decreases their pKa, increases their ionization and solubility, enhances their urinary elimination, and decreases their toxicity^[30,41-44]. However, T-amidated BA are generally less cytotoxic and more ionized than Gamidated BA^[43,45,46]. Even though unamidated as well as T-and G-amidated BAs were



Table 6 Representative bile acids concentrations and indices in compensated vs decompensated patients						
BA (umol/L) or BA indicos	Compensated		Decompensated		Decompensated vs compensated	
BA (pinoi/L) of BA indices	mean	SE	mean	SE	Ratio	P value
Total BA	66.6	10.8	86.9	14.9	1.31	0.160
Total LCA	3.73	0.54	4.26	0.70	1.14	0.547
Total UDCA	27.0	6.49	21.0	9.82	0.78	0.687
Total CDCA	20.4	3.42	45.0	6.28	2.20	0.001
Total DCA	6.85	1.76	4.93	0.73	0.72	0.394
Total HDCA	0.01	0.00	0.02	0.01	1.61	0.430
Total MDCA	0.06	0.01	0.05	0.01	0.86	0.619
Total CA	2.62	0.40	6.28	1.51	2.39	0.024
Total MCA	4.48	0.93	4.07	0.83	0.91	0.864
Total HCA	0.20	0.04	0.64	0.14	3.23	0.002
% LCA	9.00	0.64	6.61	0.64	0.73	0.020
% UDCA	24.9	1.97	12.0	1.32	0.48	0.007
% CDCA	33.2	1.62	54.74	2.05	1.65	0.000
% DCA	14.3	0.98	9.17	1.00	0.64	0.000
% HDCA	0.02	0.01	0.03	0.01	1.42	0.532
% MDCA	0.34	0.14	0.11	0.01	0.33	0.264
% CA	6.07	0.54	7.58	0.48	1.25	0.262
% MCA	7.26	0.72	7.21	0.82	0.99	0.542
% HCA	0.35	0.05	0.74	0.08	2.09	0.005
Total unamidated	4.35	0.69	3.88	1.04	0.89	0.876
Total G-amidated	56.2	9.81	70.5	12.5	1.25	0.240
Total T-amidated	5.97	0.79	12.6	2.58	2.11	0.010
% Amidation	87.9	1.15	93.6	0.75	1.06	0.003
% G-amidation	76.5	1.30	78.8	1.23	1.03	0.161
% T-amidation	11.5	0.93	14.8	1.02	1.29	0.161
Total unsulfated	7.84	1.19	9.53	1.85	1.22	0.310
Total sulfated	58.7	9.97	77.4	13.4	1.32	0.156
% Sulfation	82.7	1.12	85.2	0.99	1.03	0.054
Total mono-OH	3.73	0.54	4.26	0.70	1.14	0.547
Total di-OH	54.4	9.55	70.9	13.1	1.31	0.174
Total tri-OH	7.30	1.25	11.0	1.96	1.51	0.085
% Mono-OH	9.00	0.64	6.61	0.64	0.73	0.020
% Di-OH	72.7	1.14	76.0	1.31	1.05	0.016
% Tri-OH	13.7	0.92	15.5	0.95	1.13	0.674
Total 12a-OH	10.1	2.08	11.44	1.75	1.14	0.554
Total non-12a-OH	56.5	9.36	75.51	14.0	1.34	0.137
12α-OH/non12α-OH	0.33	0.02	0.24	0.02	0.71	0.002
CA/CDCA	0.21	0.02	0.17	0.01	0.79	0.043
% 12α-ΟΗ	22.0	1.07	17.3	0.99	0.79	0.001
% non-12α-OH	78.0	1.07	82.7	0.99	1.06	0.001

Total primary	27.7	4.46	56.0	7.59	2.02	0.001
Total secondary	38.8	7.43	31.0	10.3	0.80	0.874
Primary/secondary	2.27	0.44	5.98	0.69	2.64	0.001
% Primary	46.9	2.05	70.3	1.88	1.50	0.000
% Secondary	53.1	2.05	29.7	1.88	0.56	0.000

BA: Bile acids; G: Glycine; T: Taurine; CDCA: Chenodeoxycholic acid; CA: Cholic acid; LCA: Lithocholic acid; UDCA: Ursodeoxycholic acid; DCA: Deoxycholic acid; HDCA: Hyodeoxycholic acid; MDCA: Murideoxycholic acid; MCA: Muricholic acid; HCA: Hyocholic acid.

Table 7 Univariate logistic regression analysis of bile acids concentrations and indices ¹								
DA (umal/I) as DA indiana	Duchus (regression coefficient)	Dualua	Exp(B)-odds ratio					
BA (μmol/L) or BA indices	B value (regression coemcient)	P value	1-unit change	10% change	20% change			
Total BA	0.080	0.000	1.08	1.08	1.16			
Total CDCA	0.226	0.000	1.25	1.06	1.12			
Total CA	1.181	0.000	3.26	1.05	1.11			
% DCA	-0.080	0.000	0.92	0.78	0.61			
% HDCA	-1.898	0.069	0.15	0.99	0.97			
% MDCA	-0.174	0.162	0.84	0.99	0.98			
Total G-amidated	0.084	0.000	1.09	1.07	1.14			
Total unsulfated	0.784	0.000	2.19	1.08	1.16			
Total sulfated	0.080	0.000	1.08	1.07	1.14			
Total di-OH	0.094	0.000	1.10	1.07	1.15			
Total tri-OH	0.731	0.000	2.08	1.06	1.13			
Total non-12a-OH	0.146	0.000	1.16	1.09	1.18			
12a-OH/non12a-OH	-2.349	0.000	0.10	0.86	0.74			
% 12α-ΟΗ	-0.079	0.000	0.92	0.75	0.56			
% non-12a-OH	0.079	0.000	1.08	1.65	2.72			
Total primary	0.190	0.000	1.21	1.07	1.14			
Primary/secondary	0.834	0.000	2.30	1.06	1.12			
% Primary	0.033	0.000	1.03	1.13	1.27			
% Secondary	-0.033	0.000	0.97	0.81	0.66			

¹Bile acids with receiver operating characteristic (ROC)-areas under the ROC curve > 0.7 were included in this table.

BA: Bile acids; G: Glycine; CDCA: Chenodeoxycholic acid; CA: Cholic acid; DCA: Deoxycholic acid; HDCA: Hyodeoxycholic acid; MDCA: Murideoxycholic acid.

> higher in patients, the increase in T-amidated BA was the most profound. Therefore, % T-amidation increased, while % G-amidation decreased in patients vs controls (Table 4) as well as in medium-MELD vs low-MELD (Table 5) and decompensated vs compensated patients (Table 6). The preferential accumulation of T-amidated BA can be interpreted as an adaptive compensating response to protect the liver from BA toxicity by increasing elimination of the more toxic G-amidated and unamidated compared to the less toxic T-amidated BA^[9,26,47]. In addition, T-amidated BA has the highest affinity as substrates for the canalicular transporter, Bile Salt Export Pump (BSEP) (T-amidated > G-amidated > unamidated BA)^[48-50]. Therefore, an impairment of the BA transport by BSEP, as documented in some cholestatic diseases^[51-53], is expected to preferential accumulation T-amidated BA.

> Both sulfated and unsulfated BA were higher in patients (Table 4), but % sulfation was slightly higher in medium- compared with low-MELD and in decompensated



Table 8 Bile acids concentrations and indices in controls and patients with specific liver disease subtype ¹																								
BA (μmol/L) or BA indices	Controls		Hepatitis s C		Hepatitis B		Laennec cirrhosis		Primary biliary cholangitis		Primary sclerosing cholangitis		Autoimmune Hepatitis		α-1 antitrypsin deficiency		NASH		Carcinoma		Cryptogenic cirrhosis		Elevated LFT	
	<i>n</i> = 103		<i>n</i> = 71		<i>n</i> = 15		<i>n</i> = 117		<i>n</i> = 12		<i>n</i> = 17		n = 27		<i>n</i> = 6		<i>n</i> = 56		<i>n</i> = 26		<i>n</i> = 11		n = 22	
	mean	SE	mean	SE	mean	SE	mean	SE	mean	SE	mean	SE	mean	SE	mean	SE	mean	SE	mean	SE	mean	SE	mean	SE
Total BA	9.30	0.33	53.3 ^a	9.96	13.7 ^a	5.23	62.0 ^a	9.44	237 ^a	69.8	124 ^a	27.4	71.9 ^a	15.2	25.4 ^a	6.79	29.8 ^a	4.31	90.9 ^a	26.7	56.0 ^a	16.3	106 ^a	69.3
Total CDCA	2.52	0.13	27.0 ^a	4.89	6.76 ^a	4.16	30.0 ^a	4.25	28.6 ^a	9.99	39.4 ^a	10.7	29.2 ^a	8.46	9.14 ^a	3.12	13.6 ^a	2.33	31.9 ^a	8.76	27.7 ^a	7.39	31.3 ^a	18.8
Total CA	0.44	0.03	3.05 ^a	0.54	1.16 ^a	0.70	4.00 ^a	1.02	5.07 ^a	2.27	6.47 ^a	2.25	1.96 ^a	0.39	2.44 ^a	0.85	1.65 ^a	0.23	3.54 ^a	1.11	2.70 ^a	0.59	5.55 ^a	3.09
% DCA	31.1	0.68	16.2 ^a	1.27	19.9 ^a	3.26	13.1 ^a	0.95	7.99 ^a	2.22	9.01 ^a	1.86	15.6 ^a	1.43	18.9	4.23	15.7 ^a	1.22	14.9 ^a	1.43	7.93 ^a	2.68	17.4 ^a	3.07
% HDCA	0.07	0.01	0.02 ^a	0.00	0.06	0.02	0.02 ^a	0.01	0.03	0.02	0.01	0.00	0.02	0.00	0.03	1.21	0.05	0.01	0.06	0.02	0.04	0.03	0.05	0.04
% MDCA	0.64	0.04	0.19 ^a	0.04	0.38	0.08	0.16 ^a	0.03	0.15 ^a	0.07	0.18 ^a	0.06	0.22 ^a	0.05	0.38	0.16	0.49	0.21	0.16 ^a	0.05	0.07 ^a	0.01	1.34	0.98
Total G-amidated	7.59	0.29	44.8 ^a	9.11	11.6 ^a	4.52	50.8 ^a	8.26	210 ^a	60.4	106 ^a	24.0	61.8 ^a	13.5	16.3 ^a	4.58	26.0 ^a	4.01	78.1 ^a	24.5	49.2 ^a	14.9	86.8 ^a	58.2
Total unsulfated	0.94	0.05	7.69 ^a	1.43	1.62 ^a	0.38	7.94 ^a	1.21	17.6 ^a	8.66	6.21 ^a	1.13	6.06 ^a	1.44	3.82 ^a	1.21	4.64 ^a	0.65	13.0 ^a	3.16	3.58 ^a	0.58	13.0 ^a	9.22
Total sulfated	8.35	0.31	45.6 ^a	8.73	12.1 ^a	4.96	54.1 ^a	8.59	219 ^a	62.0	117 ^a	26.5	65.9 ^a	14.7	21.6 ^a	6.11	25.2 ^a	4.04	77.9 ^a	24.2	52.4 ^a	16.0	92.9 ^a	60.2
Total di-OH	7.30	0.29	41.0 ^a	7.98	10.6 ^a	4.41	49.05 ^a	7.97	214 ^a	62.50	111 ^a	25.9	60.9 ^a	13.7	15.9 ^a	4.72	23.2 ^a	3.87	71.9 ^a	23.0	50.3 ^a	16.0	91.0 ^a	61.0
Total tri-OH	0.82	0.04	8.61 ^a	1.63	1.82 ^a	0.80	8.59 ^a	1.54	12.48 ^a	6.02	9.28 ^a	2.44	4.83 ^a	0.76	5.24 ^a	1.83	4.27 ^a	0.66	13.8 ^a	3.62	3.92 ^a	0.65	10.8 ^a	6.50
Total non-12a-OH	5.67	0.20	41.8 ^a	7.46	10.3 ^a	4.52	50.8 ^a	7.85	224 ^a	66.9	113 ^a	26.5	62.3 ^a	14.2	19.4 ^a	5.59	23.8 ^a	3.74	75.7 ^a	22.1	51.1 ^a	16.1	94.7 ^a	66.1
12a-OH/non12a-OH	0.65	0.02	0.37 ^a	0.05	0.51	0.08	0.31 ^a	0.02	0.15 ^a	0.05	0.29 ^a	0.05	0.33 ^a	0.04	0.40	0.07	0.34 ^a	0.02	0.28 ^a	0.03	0.22 ^a	0.05	0.37 ^a	0.06
% 12α-ΟΗ	36.7	0.62	22.8 ^a	1.28	31.1	2.9	21.3 ^a	1.0	10.8 ^a	2.69%	18.8 ^a	2.6	22 ^a	1.69	27.1	3.18	23.5 ^a	1.14	20.6 ^a	1.55	16.2 ^a	2.88	24.8 ^a	2.8
% non-12a-OH	63.3	0.62	77.2 ^a	1.28	68.9	2.9	78.7 ^a	1.0	89.2 ^a	2.69	81.2 ^a	2.6	78.0 ^a	1.69	72.9	3.18	76.5 ^a	1.14	79.4 ^a	155	83.8 ^a	2.88	75.2 ^a	2.8
Total primary	3.34	0.15	35.6 ^a	6.23	8.58 ^a	4.93	38.6 ^a	5.47	41.1 ^a	15.6	48.6 ^a	12.6	34.1 ^a	8.91	14.4 ^a	4.33	17.9 ^a	2.70	45.7 ^a	12.1	31.6 ^a	7.89	42.1 ^a	23.9
Primary/secondary	0.69	0.03	3.70 ^a	0.55	1.52 ^a	0.59	4.33 ^a	0.60	0.30	0.10	2.88 ^a	0.68	2.09 ^a	0.60	1.26	0.26	2.28 ^a	0.30	2.32 ^a	0.40	6.43 ^a	2.09	1.70 ^a	0.43
% Primary	36.7	0.70	60.2 ^a	1.99	47.0 ^a	3.93	60.9 ^a	1.7	18.0 ^a	2.97	51.6 ^a	5.09	50.1 ^a	2.77	50.3	5.43	52.8 ^a	2.21	56.1 ^a	3.25	68.2 ^a	5.80	49.3 ^a	4.6
% Secondary	63.3	0.70	39.8 ^a	1.99	53.0 ^a	3.93	39.1 ^a	1.7	82.0 ^a	2.97	48.4 ^a	5.09	49.9 ^a	2.77	49.7	5.43	47.2 ^a	2.21	43.9 ^a	3.25	31.8 ^a	5.80	50.7 ^a	4.6

 1 Bile acids with receiver operating characteristic (ROC)-areas under the ROC curve > 0.7 were included in this table.

^aSignificant difference between each specific liver disease subtype vs controls (P < 0.05).

BA: Bile acids; G: Glycine; CDCA: Chenodeoxycholic acid; CA: Cholic acid; DCA: Deoxycholic acid; HDCA: Hyodeoxycholic acid; MDCA: Murideoxycholic acid.

compared with compensated patients (Tables 5 and 6). The upregulation of sulfation of BA by SULT2A1 in patients with liver diseases is thought as a compensatory response to eliminate and detoxify the accumulated toxic BA^[8-13,54,55]. However, it is also possible that sulfation activity in these patients may eventually decrease due to exhaustion or defects of these recovery mechanisms. Therefore, while liver insults can be remediated by upregulating BA sulfation under normal conditions and in milder forms of liver diseases, but subjects who fail to upregulate this defensive mechanism or exhaust it under more severe forms of the diseases are at higher risk of developing the disease and/or may have worse prognosis^[26]. Another explanation for the preferential accumulation of BA-sulfates could be related to the inhibition of their canalicular transport into bile by efflux transporters, mainly the multidrug resistance-associated proteins 2-4 (MRP 2-4). These transporters preferentially transport divalent amidated and conjugated (sulfated and glucuronidated) BA^[56-59]. MRPs including MRP2 activity is known to be compromised in various cholestatic liver diseases due downregulation of their expression and/or membrane localization^[60-62], which may lead to the preferential retention of their substrates including BA-sulfates in the liver and systemic circulation.

CYP8B1 catalyzes 12a-hydroxylation of the di-OH CDCA to the tri-OH CA. The CA/CDCA or the 12a-OH/non-12a ratios are used as probes to measure CYP8B1 activity [$^{63-65}$]. The 12a-OH/non-12a-OH ratio was 50% lower in patients compared with controls (Table 4). Also, both ratios were lower in medium-MELD vs low-MELD as well as decompensated vs compensated patients (Tables 5 and 6). This indicates that CYP8B1 activity, which exclusively takes place in the liver^[66,67], may be compromised during liver diseases in general and is further compromised with disease severity. Also, CDCA has a much higher affinity to BSEP than CA and other 12a-OH BA^[49,68]. Therefore, when BSEP activity is compromised in the more severe liver diseases, it is expected to lead to the preferential accumulation of its high-affinity substrates including CDCA, which will also decrease the CA/CDCA and 12a-OH/non-12a ratios.

Many BA concentrations and indices demonstrated AUC > 0.7 supporting their potential as biomarkers for the diagnosis of liver diseases (Supplementary Table 3). We identified three potential cut-off values, which achieve a good balance between specificity and sensitivity (Supplementary Table 5). BA indices have higher AUC values than the absolute BA concentrations, which indicates that BA indices are more accurate in distinguishing between controls and patients.

We found correlation between the risk of developing a liver disease and many BA indices using logistic regression analysis (P < 0.05). The univariate logistic regression associated with a 20% change from the mean value for the absolute BA concentrations ranged from 1.11 to 1.18, whereas it was as high as 2.72 for BA indices (Table 7). This suggests that BA indices are more sensitive than absolute BA concentrations in terms of predicting larger magnitudes of the risk of developing a liver disease.

All the above analyses demonstrate that BA indices can serve as a global marker to differentiate the pooled cholestatic liver disease population from controls in this study. In addition, we have divided the patients into different individual disease groups and performed similar analyses in these groups vs. controls, for the individual diseases. Most BA indices with ROC-AUC > 0.7 were significantly different between controls vs most of the individual liver disease subtypes (Table 8). In particular, hepatitis C and cirrhosis were the largest subpopulations in our study, and all global diagnostic BA indices from the pooled patients vs. control analyses (P < 0.05 and ROC-AUC > 0.7) were also specific diagnostic markers for these two particular liver diseases vs. controls (P < 0.05).

We have found a significant correlation between BA indices and non-BA parameters, except creatinine (P > 0.05) (Supplementary Table 8). However, BA indices, in general, outperformed non-BA parameters as biomarkers for liver diseases on many levels. Non-BA parameters were 0.76-fold to 2.5-fold higher (Table 9), whereas BA indices were as high as approximately 12-fold higher (total UDCA) in patients compared to controls (Table 4). Similarly, the magnitude of change within the MELD groups, compensation status, and among individual diseases were all much higher in BA vs non-BA.

This study has the following limitations: (1) Severity of the liver diseases were assessed using MELD score, compensation status, and a panel of liver enzymes. However, liver histological evaluation was not included because it is not a routine practice to perform liver histology on all patients, but rather for specific patients as required by the hepatologists. And (2) we have enough subjects in this study to perform solid statistics, but smaller number of subjects in many individual disease subgroups. Also, distribution of subjects between disease groups was unbalanced.



Table 9 Summary of non-bile acids parameters

Non-BA parameters	Oracharda		Patients											
	Controls		Pooled		Low-MELD		Medium-MELD		Compensated		Decompensate	AUC		
	mean	SE	mean	SE	mean	SE	mean	SE	mean	SE	mean	SE	- AUC	
Creatinine (mg/dL)	0.87	0.01	0.99	0.05	0.93	0.07	1.33 ^b	0.16	1.05	0.15	1.05	0.06	0.539	
Albumin (g/dL)	3.96	0.02	3.61 ^a	0.03	3.61	0.03	2.82 ^b	0.10	3.69	0.04	3.03 ^c	0.06	0.713	
INR	0.99	0.01	1.18 ^a	0.02	1.11	0.01	1.63 ^b	0.10	1.15	0.03	1.36 ^c	0.03	0.758	
Protime (s)	13.4	0.10	10.2 ^a	0.33	13.6	0.13	19.4 ^b	0.98	11.2	0.52	13.7 ^c	0.64	0.591	
AST (U/L)	22.8	0.34	53.2 ^a	2.31	52.1	2.59	79.2 ^b	10.4	52.6	3.97	61.7	4.85	0.876	
ALT (U/L)	21.0	0.46	51.0 ^a	2.60	51.0	3.24	46.0	5.54	49.0	4.09	40.6	3.55	0.825	
Bilirubin (mg/dL)	0.62	0.03	1.58 ^a	0.09	1.31	0.05	5.02 ^b	0.68	1.42	0.12	3.04 ^c	0.29	0.804	
AST/ALT	1.15	0.01	1.22	0.02	1.21	0.03	1.79 ^b	0.09	1.21	0.04	1.61 ^c	0.05	0.500	
MELD	7.13	0.10	10.3 ^a	0.24	9.07	0.16	18.9	0.42	9.54	0.37	14.0 ^c	0.46	0.747	
APRI	NA	NA	0.93	0.06	1.05	0.07	2.44 ^b	0.42	0.94	0.08	1.63 ^c	0.18	NA	

¹Areas under the receiver operating characteristic curve from receiver operating characteristic analysis of pooled patients vs controls.

^aSignificant difference between patients vs controls (P < 0.05).

^bSignificant difference between medium-model for end-stage liver disease vs low-model for end-stage liver disease groups (P < 0.05).

^cSignificant difference between decompensated vs compensated patients (P < 0.05).

NA: Not available; BA: Bile acids; MELD: Model for end-stage liver disease; INR: International normalized ratio; ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; APRI: Aspartate aminotransferase/platelet ratio index; ROC: Receiver operating characteristic curve; AUC: Areas under the receiver operating characteristic curve.

CONCLUSION

In summary, the results of this study demonstrated that total and all individual BA increased in patients with 11 different cholestatic diseases. However, the high interindividual variability of BA absolute concentrations makes most of them statistically insignificant and prevent their utilization as diagnostic markers. In contrast, BA indices had much lower inter- and intra-individual variability, which allowed their use as diagnostic and prognostic markers for liver diseases. Furthermore, we have shown that several BA indices outperformed non-BA markers, currently used in the clinic, as diagnostic markers to differentiate our patient pool as well as individual cholestatic diseases against healthy controls.

The increase in the total BA concentration in patients can be attributed to specific changes in the BA pool composition. This increase primarily resulted from primary BA



1-specificity

Figure 1 Receiver operating characteristics curves of bile acids concentrations and indices with area under the receiver operating characteristics curve > 0.7. The area under the receiver operating characteristics curve (AUC) for differentiating patients from healthy controls. The scale of both the Y-axis (sensitivity) and the X-axis (1-specificity) is 0-1. Bile acids (BA) indices are higher in patients vs. controls, and the positive actual state was patients except the ones annotated with "*", where BA indices were lower in patients compared to controls. For these BA indices, "1-AUC" instead of "AUC" was calculated. AUC: Area under the receiver operating characteristics curve; BA: Bile acids; CDCA: Chenodeoxycholic acid; CA: Cholic acid; DCA: Deoxycholic acid; HDCA: Hyodeoxycholic acid; MDCA: Murideoxycholic acid; G: Glycine.

> (CDCA, CA, and HCA), while the % of the secondary BA (LCA and DCA) were lower. This lead to about 4-fold increase in the primary/secondary BA ratio. Consequently, the BA pool has drastically shifted in patients from being 37% primary to approximately 50% primary BA. The increase in T-amidated BA was more profound than that of G-amidated BA, which lead to a marked increase in the % T-amidation. Furthermore, this trend of elevated primary and amidated BA was exacerbated with disease severity. This pattern can be a sign of less transformation of primary into secondary and less deconjugation of amidated BA by intestinal bacteria associated with more impairment of bile flow associated with more severe cholestatic diseases. % Sulfation was higher in patients with more severe forms of liver diseases indicating the upregulation of sulfation in these patients as a compensatory response to detoxify BA accumulation. Finally, the increase in non-12α-OH was more profound than that of 12α-OH BA, which indicates that hepatic CYP8B1 activity is compromised in liver diseases in general and is further compromised with disease severity.

In the 2nd paper of this series, we have utilized BA indexes to build a survival model called "The Bile Acid Score", which we showed was able to predict the prognosis into adverse events including death and liver transplant in liver patients.

ARTICLE HIGHLIGHTS

Research background

Bile acids (BA) have been extensively investigated for decades as biomarkers for numerous hepatobiliary diseases. However, these efforts never translated into a widespread in the clinic, due to the extreme inter-and intra-individual variability of total and individual BA concentrations and the marked differences in the physiological and pathological properties of the different individual BA. To this end, we have developed the concept of "BA indices", which demonstrated their use as diagnostic biomarkers for cholestatic liver diseases.

Research motivation

Biomarkers currently used in the clinic are not specific to the liver or bile duct injurie. BA were extensively investigated for decades as biomarkers for numerous hepatobiliary diseases. This could be attributed to the marked differences in the physiological and pathological properties of the different individual BA. BA indices have much lower variability than the absolute BA concentrations used to calculate them. Indeed, we have demonstrated that BA indices offered numerous advantages over absolute total and individual BA concentrations including low inter- and intraindividual variability and were resistant to covariate influences such as age, gender, body mass index, food consumption, and moderate alcohol consumption.

Research objectives

The objective of this project was to discover and validate diagnostic biomarkers of cholestatic liver diseases based on the urinary BA profile. We have developed the concept of "BA indices", which are ratios calculated from the absolute concentrations of individual BA and their metabolites. BA indices have much lower variability than the absolute BA concentrations used to calculate them, which enabled their use as diagnostic biomarkers for cholestatic liver diseases.

Research methods

We analyzed urine samples by liquid chromatography-tandem mass spectrometry and compared the urinary BA profile between patients with hepatobiliary diseases vs healthy controls by statistical analysis (independent sample-t-test, Mann-Whitney test, Mixed effects models, by pairwise comparisons using Bonferroni's adjustment, receiver operating characteristic curve analyses, Univariate and multivariate logistic regression analysis).

Research results

The results of this study demonstrated that total and all individual BA increased in patients with 11 different cholestatic diseases. However, the high inter-individual variability of BA absolute concentrations makes most of them statistically insignificant and prevent their utilization as diagnostic markers. In contrast, BA indices had much lower inter- and intra-individual variability, which allowed their use as diagnostic and prognostic markers for liver diseases. Furthermore, we have shown that several BA indices outperformed non-BA markers, currently used in the clinic, as diagnostic markers to differentiate our patient pool as well as individual cholestatic diseases against healthy controls.

Research conclusions

BA indices demonstrated high area under the receiver operating characteristic curves, and changes of BA indices were associated with the risk of having a liver disease as determined by the logistic regression analysis, which demonstrated their use as diagnostic biomarkers for cholestatic liver diseases.

Research perspectives

We have developed survival models based on BA indices to predict the prognosis of hepatobiliary diseases which is illustrated in the second paper of this series.



ACKNOWLEDGEMENTS

The authors wish to thank the nurses of the CRC (Mary Ann Martin, Cindy Cowarden, Caroline Peterson, Claire Haier, and Mary Phillips) and the staff for their valuable contributions to managing the health control arm of the study, recruiting subjects, and collecting samples.

REFERENCES

- Hofmann AF. The continuing importance of bile acids in liver and intestinal disease. Arch Intern 1 Med 1999; 159: 2647-2658 [PMID: 10597755 DOI: 10.1001/archinte.159.22.2647]
- 2 Monte MJ, Marin JJ, Antelo A, Vazquez-Tato J. Bile acids: chemistry, physiology, and pathophysiology. World J Gastroenterol 2009; 15: 804-816 [PMID: 19230041 DOI: 10.3748/wjg.15.804]
- 3 Khurana S, Raufman JP, Pallone TL. Bile acids regulate cardiovascular function. Clin Transl Sci 2011; 4: 210-218 [PMID: 21707953 DOI: 10.1111/j.1752-8062.2011.00272.x]
- Thomas C, Pellicciari R, Pruzanski M, Auwerx J, Schoonjans K. Targeting bile-acid signalling for metabolic diseases. Nat Rev Drug Discov 2008; 7: 678-693 [PMID: 18670431 DOI: 10.1038/nrd2619]
- Maillette de Buy Wenniger L, Beuers U. Bile salts and cholestasis. Dig Liver Dis 2010; 42: 409-418 [PMID: 20434968 DOI: 10.1016/j.dld.2010.03.015]
- Pauli-Magnus C, Meier PJ. Hepatocellular transporters and cholestasis. J Clin Gastroenterol 2005; 6 39: S103-S110 [PMID: 15758645 DOI: 10.1097/01.mcg.0000155550.29643.7b]
- 7 Palmer RH. Bile acids, liver injury, and liver disease. Arch Intern Med 1972; 130: 606-617 [PMID: 4627840
- 8 Alnouti Y. Bile Acid sulfation: a pathway of bile acid elimination and detoxification. Toxicol Sci 2009; 108: 225-246 [PMID: 19131563 DOI: 10.1093/toxsci/kfn268]
- Bathena SP, Thakare R, Gautam N, Mukherjee S, Olivera M, Meza J, Alnouti Y. Urinary bile acids 9 as biomarkers for liver diseases II. Signature profiles in patients. Toxicol Sci 2015; 143: 308-318 [PMID: 25344563 DOI: 10.1093/toxsci/kfu228]
- 10 Makino I, Hashimoto H, Shinozaki K, Yoshino K, Nakagawa S. Sulfated and nonsulfated bile acids in urine, serum, and bile of patients with hepatobiliary diseases. Gastroenterology 1975; 68: 545-553 [PMID: 1112456]
- Summerfield JA, Cullen J, Barnes S, Billing BH. Evidence for renal control of urinary excretion of 11 bile acids and bile acid sulphates in the cholestatic syndrome. Clin Sci Mol Med 1977; 52: 51-65 [PMID: 606464 DOI: 10.1042/cs0520051]
- Takikawa H, Beppu T, Seyama Y. Urinary concentrations of bile acid glucuronides and sulfates in 12 hepatobiliary diseases. Gastroenterol Jpn 1984; 19: 104-109 [PMID: 6735068 DOI: 10.1007/bf02806931]
- van Berge Henegouwen GP, Brandt KH, Eyssen H, Parmentier G. Sulphated and unsulphated bile 13 acids in serum, bile, and urine of patients with cholestasis. Gut 1976; 17: 861-869 [PMID: 1001976 DOI: 10.1136/gut.17.11.861]
- LaRusso NF, Shneider BL, Black D, Gores GJ, James SP, Doo E, Hoofnagle JH. Primary sclerosing 14 cholangitis: summary of a workshop. Hepatology 2006; 44: 746-764 [PMID: 16941705 DOI: 10.1002/hep.21337
- 15 Dueland S, Reichen J, Everson GT, Davis RA. Regulation of cholesterol and bile acid homoeostasis in bile-obstructed rats. Biochem J 1991; 280 (Pt 2): 373-377 [PMID: 1747109 DOI: 10.1042/bj2800373
- 16 Kawai H, Kudo N, Kawashima Y, Mitsumoto A. Efficacy of urine bile acid as a non-invasive indicator of liver damage in rats. J Toxicol Sci 2009; 34: 27-38 [PMID: 19182433 DOI: 10.2131/jts.34.27
- Geuken E, Visser D, Kuipers F, Blokzijl H, Leuvenink HG, de Jong KP, Peeters PM, Jansen PL, 17 Slooff MJ, Gouw AS, Porte RJ. Rapid increase of bile salt secretion is associated with bile duct injury after human liver transplantation. J Hepatol 2004; 41: 1017-1025 [PMID: 15582136 DOI: 10.1016/j.jhep.2004.08.023]
- Buis CI, Geuken E, Visser DS, Kuipers F, Haagsma EB, Verkade HJ, Porte RJ. Altered bile 18 composition after liver transplantation is associated with the development of nonanastomotic biliary strictures. J Hepatol 2009; 50: 69-79 [PMID: 19012987 DOI: 10.1016/j.jhep.2008.07.032]
- Ramaiah SK. A toxicologist guide to the diagnostic interpretation of hepatic biochemical parameters. 19 Food Chem Toxicol 2007; 45: 1551-1557 [PMID: 17658209 DOI: 10.1016/j.fct.2007.06.007]
- 20 Ozer J, Ratner M, Shaw M, Bailey W, Schomaker S. The current state of serum biomarkers of hepatotoxicity. Toxicology 2008; 245: 194-205 [PMID: 18291570 DOI: 10.1016/j.tox.2007.11.021]
- 21 Huang WM, Seubert DE, Donnelly JG, Liu M, Javitt NB. Intrahepatic cholestasis of pregnancy: detection with urinary bile acid assays. J Perinat Med 2007; 35: 486-491 [PMID: 18052835 DOI: 10.1515/JPM.2007.128]
- Muraji T, Harada T, Miki K, Moriuchi T, Obatake M, Tsugawa C. Urinary sulfated bile acid 22 concentrations in infants with biliary atresia and breast-feeding jaundice. Pediatr Int 2003; 45: 281-



283 [PMID: 12828581 DOI: 10.1046/j.1442-200x.2003.01710.x]

- Sinakos E, Lindor KD. Bile acid profiles in intrahepatic cholestasis of pregnancy: is this the solution 23 to the enigma of intrahepatic cholestasis of pregnancy? Am J Gastroenterol 2010; 105: 596-598 [PMID: 20203641 DOI: 10.1038/ajg.2009.639]
- 24 Meng LJ, Reyes H, Palma J, Hernandez I, Ribalta J, Sjövall J. Profiles of bile acids and progesterone metabolites in the urine and serum of women with intrahepatic cholestasis of pregnancy. J Hepatol 1997; 27: 346-357 [PMID: 9288610 DOI: 10.1016/s0168-8278(97)80181-x]
- 25 Trottier J, Białek A, Caron P, Straka RJ, Milkiewicz P, Barbier O. Profiling circulating and urinary bile acids in patients with biliary obstruction before and after biliary stenting. PLoS One 2011; 6: e22094 [PMID: 21760958 DOI: 10.1371/journal.pone.0022094]
- Bathena SP, Thakare R, Gautam N, Mukherjee S, Olivera M, Meza J, Alnouti Y. Urinary bile acids 26 as biomarkers for liver diseases I. Stability of the baseline profile in healthy subjects. Toxicol Sci 2015; 143: 296-307 [PMID: 25344562 DOI: 10.1093/toxsci/kfu227]
- 27 U. S. Department of Health and Human Services and U.S. Department of Agriculture. 2015-2020 Dietary Guidelines for Americans. [cited September 15, 2019]. In: USDA and HHS [Internet]. Available from: https://health.gov/sites/default/files/2019-09/2015-2020 Dietary Guidelines.pdf
- Zipprich A, Garcia-Tsao G, Rogowski S, Fleig WE, Seufferlein T, Dollinger MM. Prognostic 28 indicators of survival in patients with compensated and decompensated cirrhosis. Liver Int 2012; 32: 1407-1414 [PMID: 22679906 DOI: 10.1111/j.1478-3231.2012.02830.x]
- 29 Huang J, Bathena SP, Csanaky IL, Alnouti Y. Simultaneous characterization of bile acids and their sulfate metabolites in mouse liver, plasma, bile, and urine using LC-MS/MS. J Pharm Biomed Anal 2011; 55: 1111-1119 [PMID: 21530128 DOI: 10.1016/j.jpba.2011.03.035]
- 30 Bathena SP, Mukherjee S, Olivera M, Alnouti Y. The profile of bile acids and their sulfate metabolites in human urine and serum. J Chromatogr B Analyt Technol Biomed Life Sci 2013; 942-943: 53-62 [PMID: 24212143 DOI: 10.1016/j.jchromb.2013.10.019]
- Thakare R, Alamoudi JA, Gautam N, Rodrigues AD, Alnouti Y. Species differences in bile acids II. 31 Bile acid metabolism. J Appl Toxicol 2018; 38: 1336-1352 [PMID: 29845631 DOI: 10.1002/jat.3645]
- 32 Pagano M, Gauvreau K. Principles of Biostatistics. 2nd ed. Brooks/Cole: Duxbury, 2000: 259-331
- 33 Malinchoc M, Kamath PS, Gordon FD, Peine CJ, Rank J, ter Borg PC. A model to predict poor survival in patients undergoing transjugular intrahepatic portosystemic shunts. Hepatology 2000; 31: 864-871 [PMID: 10733541 DOI: 10.1053/he.2000.5852]
- 34 Kamath PS, Wiesner RH, Malinchoc M, Kremers W, Therneau TM, Kosberg CL, D'Amico G, Dickson ER, Kim WR. A model to predict survival in patients with end-stage liver disease. Hepatology 2001; 33: 464-470 [PMID: 11172350 DOI: 10.1053/jhep.2001.22172]
- 35 Murray KF, Carithers RL Jr; AASLD. AASLD practice guidelines: Evaluation of the patient for liver transplantation. *Hepatology* 2005; **41**: 1407-1432 [PMID: 15880505 DOI: 10.1002/hep.20704]
- Zipprich A, Kuss O, Rogowski S, Kleber G, Lotterer E, Seufferlein T, Fleig WE, Dollinger MM. 36 Incorporating indocyanin green clearance into the Model for End Stage Liver Disease (MELD-ICG) improves prognostic accuracy in intermediate to advanced cirrhosis. Gut 2010; 59: 963-968 [PMID: 20581243 DOI: 10.1136/gut.2010.208595]
- 37 Farnsworth N, Fagan SP, Berger DH, Awad SS. Child-Turcotte-Pugh vs MELD score as a predictor of outcome after elective and emergent surgery in cirrhotic patients. Am J Surg 2004; 188: 580-583 [PMID: 15546574 DOI: 10.1016/j.amjsurg.2004.07.034]
- Trottier J, Białek A, Caron P, Straka RJ, Heathcote J, Milkiewicz P, Barbier O. Metabolomic 38 profiling of 17 bile acids in serum from patients with primary biliary cirrhosis and primary sclerosing cholangitis: a pilot study. Dig Liver Dis 2012; 44: 303-310 [PMID: 22169272 DOI: 10.1016/j.dld.2011.10.025
- Modica S, Petruzzelli M, Bellafante E, Murzilli S, Salvatore L, Celli N, Di Tullio G, Palasciano G, 39 Moustafa T, Halilbasic E, Trauner M, Moschetta A. Selective activation of nuclear bile acid receptor FXR in the intestine protects mice against cholestasis. Gastroenterology 2012; 142: 355-65. e1-4 [PMID: 22057115 DOI: 10.1053/j.gastro.2011.10.028]
- Luo L, Aubrecht J, Li D, Warner RL, Johnson KJ, Kenny J, Colangelo JL. Assessment of serum bile 40 acid profiles as biomarkers of liver injury and liver disease in humans. PLoS One 2018; 13: e0193824 [PMID: 29513725 DOI: 10.1371/journal.pone.0193824]
- 41 Heuman DM, Pandak WM, Hylemon PB, Vlahcevic ZR. Conjugates of ursodeoxycholate protect against cytotoxicity of more hydrophobic bile salts: in vitro studies in rat hepatocytes and human erythrocytes. Hepatology 1991; 14: 920-926 [PMID: 1937396 DOI: 10.1002/hep.1840140527]
- Rolo AP, Oliveira PJ, Moreno AJ, Palmeira CM. Bile acids affect liver mitochondrial bioenergetics: 42 possible relevance for cholestasis therapy. Toxicol Sci 2000; 57: 177-185 [PMID: 10966524 DOI: 10.1093/toxsci/57.1.177
- Li Y, Zhang X, Chen J, Feng C, He Y, Shao Y, Ding M. Targeted metabolomics of sulfated bile acids 43 in urine for the diagnosis and grading of intrahepatic cholestasis of pregnancy. Genes Dis 2018; 5: 358-366 [PMID: 30591938 DOI: 10.1016/j.gendis.2018.01.005]
- Hofmann AF, Hagey LR. Bile acids: chemistry, pathochemistry, biology, pathobiology, and 44 therapeutics. Cell Mol Life Sci 2008; 65: 2461-2483 [PMID: 18488143 DOI: 10.1007/s00018-008-7568-6
- Rust C, Karnitz LM, Paya CV, Moscat J, Simari RD, Gores GJ. The bile acid 45 taurochenodeoxycholate activates a phosphatidylinositol 3-kinase-dependent survival signaling cascade. J Biol Chem 2000; 275: 20210-20216 [PMID: 10770953 DOI: 10.1074/jbc.M909992199]



- 46 Rust C, Bauchmuller K, Fickert P, Fuchsbichler A, Beuers U. Phosphatidylinositol 3-kinasedependent signaling modulates taurochenodeoxycholic acid-induced liver injury and cholestasis in perfused rat livers. Am J Physiol Gastrointest Liver Physiol 2005; 289: G88-G94 [PMID: 15746212 DOI: 10.1152/ajpgi.00450.2004]
- Perreault M, Białek A, Trottier J, Verreault M, Caron P, Milkiewicz P, Barbier O. Role of 47 glucuronidation for hepatic detoxification and urinary elimination of toxic bile acids during biliary obstruction. PLoS One 2013; 8: e80994 [PMID: 24244729 DOI: 10.1371/journal.pone.0080994]
- Gerloff T, Stieger B, Hagenbuch B, Madon J, Landmann L, Roth J, Hofmann AF, Meier PJ. The 48 sister of P-glycoprotein represents the canalicular bile salt export pump of mammalian liver. J Biol Chem 1998; 273: 10046-10050 [PMID: 9545351 DOI: 10.1074/jbc.273.16.10046]
- 49 Hayashi H, Takada T, Suzuki H, Onuki R, Hofmann AF, Sugiyama Y. Transport by vesicles of glycine- and taurine-conjugated bile salts and taurolithocholate 3-sulfate: a comparison of human BSEP with rat Bsep. Biochim Biophys Acta 2005; 1738: 54-62 [PMID: 16332456 DOI: 10.1016/j.bbalip.2005.10.006]
- 50 Noé J, Stieger B, Meier PJ. Functional expression of the canalicular bile salt export pump of human liver. Gastroenterology 2002; 123: 1659-1666 [PMID: 12404240 DOI: 10.1053/gast.2002.36587]
- Jansen PL, Strautnieks SS, Jacquemin E, Hadchouel M, Sokal EM, Hooiveld GJ, Koning JH, De 51 Jager-Krikken A, Kuipers F, Stellaard F, Bijleveld CM, Gouw A, Van Goor H, Thompson RJ, Müller M. Hepatocanalicular bile salt export pump deficiency in patients with progressive familial intrahepatic cholestasis. Gastroenterology 1999; 117: 1370-1379 [PMID: 10579978 DOI: 10.1016/s0016-5085(99)70287-8]
- van der Woerd WL, Houwen RH, van de Graaf SF. Current and future therapies for inherited 52 cholestatic liver diseases. World J Gastroenterol 2017; 23: 763-775 [PMID: 28223721 DOI: 10.3748/wjg.v23.i5.763
- 53 Stieger B. Role of the bile salt export pump, BSEP, in acquired forms of cholestasis. Drug Metab Rev 2010; **42**: 437-445 [PMID: 20028269 DOI: 10.3109/03602530903492004]
- Stiehl A, Raedsch R, Rudolph G, Gundert-Remy U, Senn M. Biliary and urinary excretion of 54 sulfated, glucuronidated and tetrahydroxylated bile acids in cirrhotic patients. *Hepatology* 1985; 5: 492-495 [PMID: 3997078 DOI: 10.1002/hep.1840050325]
- 55 Barrett KG, Fang H, Cukovic D, Dombkowski AA, Kocarek TA, Runge-Morris M. Upregulation of UGT2B4 Expression by 3'-Phosphoadenosine-5'-Phosphosulfate Synthase Knockdown: Implications for Coordinated Control of Bile Acid Conjugation. Drug Metab Dispos 2015; 43: 1061-1070 [PMID: 25948711 DOI: 10.1124/dmd.114.061440]
- Meier PJ, Stieger B. Bile salt transporters. Annu Rev Physiol 2002; 64: 635-661 [PMID: 11826283 56 DOI: 10.1146/annurev.physiol.64.082201.100300]
- 57 Hirohashi T, Suzuki H, Takikawa H, Sugiyama Y. ATP-dependent transport of bile salts by rat multidrug resistance-associated protein 3 (Mrp3). J Biol Chem 2000; 275: 2905-2910 [PMID: 10644759 DOI: 10.1074/jbc.275.4.2905]
- 58 Zelcer N, Reid G, Wielinga P, Kuil A, van der Heijden I, Schuetz JD, Borst P. Steroid and bile acid conjugates are substrates of human multidrug-resistance protein (MRP) 4 (ATP-binding cassette C4). Biochem J 2003; 371: 361-367 [PMID: 12523936 DOI: 10.1042/BJ20021886]
- 59 Stieger B, Fattinger K, Madon J, Kullak-Ublick GA, Meier PJ. Drug- and estrogen-induced cholestasis through inhibition of the hepatocellular bile salt export pump (Bsep) of rat liver. Gastroenterology 2000; 118: 422-430 [PMID: 10648470 DOI: 10.1016/s0016-5085(00)70224-1]
- 60 Kubitz R, Wettstein M, Warskulat U, Häussinger D. Regulation of the multidrug resistance protein 2 in the rat liver by lipopolysaccharide and dexamethasone. Gastroenterology 1999; 116: 401-410 [PMID: 9922322 DOI: 10.1016/s0016-5085(99)70138-1]
- 61 Mottino AD, Cao J, Veggi LM, Crocenzi F, Roma MG, Vore M. Altered localization and activity of canalicular Mrp2 in estradiol-17beta-D-glucuronide-induced cholestasis. Hepatology 2002; 35: 1409-1419 [PMID: 12029626 DOI: 10.1053/jhep.2002.33327]
- Kojima H, Nies AT, König J, Hagmann W, Spring H, Uemura M, Fukui H, Keppler D. Changes in 62 the expression and localization of hepatocellular transporters and radixin in primary biliary cirrhosis. J Hepatol 2003; 39: 693-702 [PMID: 14568249 DOI: 10.1016/s0168-8278(03)00410-0]
- 63 Pandak WM, Bohdan P, Franklund C, Mallonee DH, Eggertsen G, Björkhem I, Gil G, Vlahcevic ZR, Hylemon PB. Expression of sterol 12alpha-hydroxylase alters bile acid pool composition in primary rat hepatocytes and in vivo. Gastroenterology 2001; 120: 1801-1809 [PMID: 11375960 DOI: 10.1053/gast.2001.24833]
- Zhang M, Chiang JY. Transcriptional regulation of the human sterol 12alpha-hydroxylase gene 64 (CYP8B1): roles of heaptocyte nuclear factor 4alpha in mediating bile acid repression. J Biol Chem 2001; 276: 41690-41699 [PMID: 11535594 DOI: 10.1074/jbc.M105117200]
- 65 Mörk LM, Strom SC, Mode A, Ellis EC. Addition of Dexamethasone Alters the Bile Acid Composition by Inducing CYP8B1 in Primary Cultures of Human Hepatocytes. J Clin Exp Hepatol 2016; 6: 87-93 [PMID: 27493455 DOI: 10.1016/j.jceh.2016.01.007]
- 66 Li T, Apte U. Bile Acid Metabolism and Signaling in Cholestasis, Inflammation, and Cancer. Adv Pharmacol 2015; 74: 263-302 [PMID: 26233910 DOI: 10.1016/bs.apha.2015.04.003]
- 67 Chiang JY. Negative feedback regulation of bile acid metabolism: impact on liver metabolism and diseases. Hepatology 2015; 62: 1315-1317 [PMID: 26122550 DOI: 10.1002/hep.27964]
- 68 Mita S, Suzuki H, Akita H, Hayashi H, Onuki R, Hofmann AF, Sugiyama Y. Inhibition of bile acid transport across Na+/taurocholate cotransporting polypeptide (SLC10A1) and bile salt export pump



(ABCB 11)-coexpressing LLC-PK1 cells by cholestasis-inducing drugs. *Drug Metab Dispos* 2006; **34**: 1575-1581 [PMID: 16760228 DOI: 10.1124/dmd.105.008748]





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