# World Journal of *Stem Cells*

World J Stem Cells 2021 August 26; 13(8): 971-1159





Published by Baishideng Publishing Group Inc

World Journal of Stem Cells

#### Contents

#### Monthly Volume 13 Number 8 August 26, 2021

#### **REVIEW**

Differences and similarities between mesenchymal stem cell and endothelial progenitor cell 971 immunoregulatory properties against T cells

Razazian M, Khosravi M, Bahiraii S, Uzan G, Shamdani S, Naserian S

- 985 Inter-regulatory role of microRNAs in interaction between viruses and stem cells Afshari A, Yaghobi R, Rezaei G
- 1005 Mesenchymal stem cells for enhancing biological healing after meniscal injuries Rhim HC, Jeon OH, Han SB, Bae JH, Suh DW, Jang KM
- 1030 Modulating poststroke inflammatory mechanisms: Novel aspects of mesenchymal stem cells, extracellular vesicles and microglia

Xin WQ, Wei W, Pan YL, Cui BL, Yang XY, Bähr M, Doeppner TR

#### **MINIREVIEWS**

1049 Antler stem cells and their potential in wound healing and bone regeneration Zhang W, Ke CH, Guo HH, Xiao L

1058 Therapeutic prospects of mesenchymal stem/stromal cells in COVID-19 associated pulmonary diseases: From bench to bedside

Zhang LS, Yu Y, Yu H, Han ZC

1072 Mesenchymal stem cells as a potential therapeutic tool to cure cognitive impairment caused by neuroinflammation

Skok M

1084 Effects of radiation and chemotherapy on adipose stem cells: Implications for use in fat grafting in cancer patients

Platoff R, Villalobos MA, Hagaman AR, Liu Y, Matthews M, DiSanto ME, Carpenter JP, Zhang P

1094 Methods to produce induced pluripotent stem cell-derived mesenchymal stem cells: Mesenchymal stem cells from induced pluripotent stem cells

Dupuis V, Oltra E

1112 Central nervous system tumors and three-dimensional cell biology: Current and future perspectives in modeling

Abou-Mrad Z, Bou Gharios J, Moubarak MM, Chalhoub A, Moussalem C, Bahmad HF, Abou-Kheir W

1127 Regulators of liver cancer stem cells Liu K, Ou JHJ



#### Contents

World Journal of Stem Cells

#### Monthly Volume 13 Number 8 August 26, 2021

#### SYSTEMATIC REVIEWS

Induced pluripotent stem cells as suitable sensors for fibromyalgia and myalgic encephalomyelitis/chronic 1134 fatigue syndrome

Monzón-Nomdedeu MB, Morten KJ, Oltra E

#### **CASE REPORT**

1151 Treatment of acute ischemic stroke by minimally manipulated umbilical cord-derived mesenchymal stem cells transplantation: A case report

Ahn H, Lee SY, Jung WJ, Lee KH



### Contents

Monthly Volume 13 Number 8 August 26, 2021

#### **ABOUT COVER**

Editorial Board Member of World Journal of Stem Cells, Radwa A Mehanna, MD, PhD, Professor, Physiology Department, Vice Executive Manager, Center of Excellence For Research in Regenerative Medicine and its Applications CERRMA, Faculty of Medicine, Alexandria University, Alexandria 21561, Egypt. radwa.mehanna@alexmed.edu.eg

#### **AIMS AND SCOPE**

The primary aim of World Journal of Stem Cells (WJSC, World J Stem Cells) is to provide scholars and readers from various fields of stem cells with a platform to publish high-quality basic and clinical research articles and communicate their research findings online. WJSC publishes articles reporting research results obtained in the field of stem cell biology and regenerative medicine, related to the wide range of stem cells including embryonic stem cells, germline stem cells, tissue-specific stem cells, adult stem cells, mesenchymal stromal cells, induced pluripotent stem cells, embryonal carcinoma stem cells, hemangioblasts, lymphoid progenitor cells, etc.

#### **INDEXING/ABSTRACTING**

The WJSC is now indexed in Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports/Science Edition, Biological Abstracts, BIOSIS Previews, Scopus, PubMed, and PubMed Central. The 2021 Edition of Journal Citation Reports<sup>®</sup> cites the 2020 impact factor (IF) for WJSC as 5.326; IF without journal self cites: 5.035; 5-year IF: 4.956; Journal Citation Indicator: 0.55; Ranking: 14 among 29 journals in cell and tissue engineering; Quartile category: Q2; Ranking: 72 among 195 journals in cell biology; and Quartile category: Q2. The WJSC's CiteScore for 2020 is 3.1 and Scopus CiteScore rank 2020: Histology is 31/60; Genetics is 205/325; Genetics (clinical) is 64/87; Molecular Biology is 285/382; Cell Biology is 208/279.

#### **RESPONSIBLE EDITORS FOR THIS ISSUE**

Production Editor: Yan-Xia Xing, Production Department Director: Yu-Jie Ma; Editorial Office Director: Ze-Mao Gong.

NAME OF JOURNAL	INSTRUCTIONS TO AUTHORS		
World Journal of Stem Cells	https://www.wignet.com/bpg/gerinfo/204		
<b>ISSN</b>	GUIDELINES FOR ETHICS DOCUMENTS		
ISSN 1948-0210 (online)	https://www.wjgnet.com/bpg/GerInfo/287		
LAUNCH DATE	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH		
December 31, 2009	https://www.wjgnet.com/bpg/gerinfo/240		
FREQUENCY	PUBLICATION ETHICS		
Monthly	https://www.wjgnet.com/bpg/GerInfo/288		
EDITORS-IN-CHIEF	PUBLICATION MISCONDUCT		
Shengwen Calvin Li, Tong Cao, Carlo Ventura	https://www.wjgnet.com/bpg/gerinfo/208		
EDITORIAL BOARD MEMBERS	ARTICLE PROCESSING CHARGE		
https://www.wjgnet.com/1948-0210/editorialboard.htm	https://www.wjgnet.com/bpg/gerinfo/242		
PUBLICATION DATE	STEPS FOR SUBMITTING MANUSCRIPTS		
August 26, 2021	https://www.wjgnet.com/bpg/GerInfo/239		
COPYRIGHT	ONLINE SUBMISSION		
© 2021 Baishideng Publishing Group Inc	https://www.f6publishing.com		

© 2021 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: bpgoffice@wjgnet.com https://www.wjgnet.com



W J S C World Journal of Stem Cells

Submit a Manuscript: https://www.f6publishing.com

World J Stem Cells 2021 August 26; 13(8): 1084-1093

DOI: 10.4252/wisc.v13.i8.1084

ISSN 1948-0210 (online)

MINIREVIEWS

# Effects of radiation and chemotherapy on adipose stem cells: Implications for use in fat grafting in cancer patients

Rebecca Platoff, Miguel A Villalobos, Ashleigh Rapp Hagaman, Yuan Liu, Martha Matthews, Michael E DiSanto, Jeffrey P Carpenter, Ping Zhang

**ORCID number:** Rebecca Platoff 0000-0002-2285-7502; Miguel A Villalobos 0000-0002-9518-5647: Ashleigh Rapp Hagaman 0000-0003-0567-2984; Yuan Liu 0000-0001-6378-0436; Martha Matthews 0000-0002-9470-0002; Michael E DiSanto 0000-0003-4754-7794; Jeffrey P Carpenter 0000-0003-4569-3106; Ping Zhang 0000-0001-6581-7941.

Author contributions: Platoff R reviewed the literature and drafted the manuscript; Villalobos MA and Hagaman AR performed the majority of experiments and drafted the manuscript; Liu Y and Matthews M conception and design; DiSanto ME and Carpenter JP critical revision and editing of the manuscript; Zhang P conceived the manuscript, reviewed the literature, performed conception and design, wrote the article and approved the final version as the corresponding author.

Conflict-of-interest statement: The authors declare that there is no conflict of interest.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0)

Rebecca Platoff, Miguel A Villalobos, Ashleigh Rapp Hagaman, Yuan Liu, Martha Matthews, Jeffrey P Carpenter, Ping Zhang, Department of Surgery, Cooper University Health Care, Camden, NJ 08103. United States

Yuan Liu, Martha Matthews, Jeffrey P Carpenter, Ping Zhang, Department of Surgery, Cooper Medical School of Rowan University, Camden, NJ 08103, United States

Michael E DiSanto, Department of Biomedical Sciences, Cooper Medical School of Rowan University, Camden, NJ 08103, United States

Corresponding author: Ping Zhang, DDS, PhD, Assistant Professor, Department of Surgery, Cooper University Health Care, 401 Haddon Avenue, Camden, NJ 08103, United States. zhang-ping@cooperhealth.edu

## Abstract

Autologous fat transplantation is a versatile tool in reconstructive surgery. Adipose-derived stem cells (ASCs) increase survival of fat grafts and thus are increasingly used for breast reconstruction in breast cancer patients. However, radiation and/or chemotherapy have been proposed to inhibit soft tissue regeneration in wound healing thus suggesting alteration in stem cell pathways. Therefore, elucidating effects of radiation and chemotherapy on ASCs is critical if one desires to enhance the survival of fat grafts in patients. This review outlines our work evaluating the function and recoverability of ASCs from radiation or chemotherapy patients, focusing specifically on their availability as a source of autologous stem cells for fat grafting and breast reconstruction in cancer patients. Even though evidence suggests radiation and chemotherapy negatively influence ASCs at the cellular level, the efficiency of the isolation and differentiation capacity did not appear influenced in patients after receiving chemotherapy treatment, although fat from radiated patients exhibited significantly altered ASC differentiation into endothelial-like cells. Further, the *in vitro* growth rates of patient's ASCs do not differ significantly before or after treatment. Taken together, these studies suggest ASCs as an important new tool for grafting and reconstruction even when radiation and chemotherapy treatment are involved.

Key Words: Fat grafting; Breast reconstruction; Stem cells; Breast cancer; Radiation; Chemotherapy



license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Invited manuscript

Specialty type: Cell and tissue engineering

Country/Territory of origin: United States

#### Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B, B Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Received: February 25, 2021 Peer-review started: February 25, 2021 First decision: April 20, 2021 Revised: April 30, 2021 Accepted: July 29, 2021 Article in press: July 29, 2021 Published online: August 26, 2021

P-Reviewer: Frasca A, Portincasa A, Rojas A S-Editor: Gao CC L-Editor: A P-Editor: Ma YJ



#### ©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Breast reconstruction with fat grafting after surgery is a major therapy that is enhanced by the use of autologous adipose-derived stem cells (ASCs) following surgery for breast cancer. Emerging studies suggest that cancer treatment therapies have a cytotoxic effect that may limit stem cell cellular functions. In this review, we summarize our work on evaluating the functional recovery potential of autologous ASCs in patients post-radiation or chemotherapy. In addition, we provide evidence that ASCs may represent a novel and effective cellular mechanism for enhancing fat grafting and reconstruction outcomes especially in those patients with cancer.

Citation: Platoff R, Villalobos MA, Hagaman AR, Liu Y, Matthews M, DiSanto ME, Carpenter JP, Zhang P. Effects of radiation and chemotherapy on adipose stem cells: Implications for use in fat grafting in cancer patients. World J Stem Cells 2021; 13(8): 1084-1093 URL: https://www.wjgnet.com/1948-0210/full/v13/i8/1084.htm

DOI: https://dx.doi.org/10.4252/wjsc.v13.i8.1084

#### INTRODUCTION

Autologous fat transplantation is a versatile tool in reconstructive surgery. As a supplement to whole-fat grafts, adipose-derived stem cells (ASCs) can exhibit a positive effect on wound healing and increase survival rates as evidenced by their increasing use in plastic and reconstructive procedures[1-4]. ASCs have been used for nearly 20 years in reconstructive surgery to help with soft tissue healing by improving tissue vascularity, elasticity, and healing capacity. At the same time, ASCs help to minimize inflammation, and overall, lead to tissue self-renewal, and differentiation into specialized cell types[5-7]. In addition, ASCs, as an abundant source of adult mesenchymal stem cells, have been shown to secrete several growth factors that play a key role in promoting neovascularization which contributes to adipose tissue regeneration<sup>[7,8]</sup>. In addition, ASCs secrete multiple anti-apoptotic growth factors to attenuate adipocyte loss, which ultimately enhances the viability of fat grafts[9].

Recently, clinical studies concluded that autologous fat grafting with a stromal vascular fraction, a rich source of ASCs, improves clinical outcomes in breast augmentation and facial lipoatrophy patients [3,10-12]. This is due to ASCs' versatility to develop into a variety of mature tissues as well as their great capacity for proliferation[13]. The ability of ASCs to produce adipose tissue regeneration has been shown to enhance volume and improve cosmesis and symmetry in breast reconstruction[6]. The use of ASCs is also thought to improve the survival of fat grafting because it can boost angiogenesis via ASC's differentiation into endothelial cells (EC) and vascular endothelial growth factor (VEGF) secretion[14]. Moreover, ASCs have a longer lifespan in culture than bone marrow stromal cells before becoming senescent which allows greater flexibility in the lab environment[15,16]. The beneficial properties of ASCs described above highlight the potential importance of ASCs in the maintenance of transplanted tissue volume, an important consideration in adipose tissue engineering[8].

In recent years, the use of ASCs within the fat graft is considered particularly useful and has been increasingly employed following breast-conserving cancer surgery. Currently, approximately 93000 breast reconstructions are performed each year in the United States, supporting the clinical significance of this work[12,17,18]. Breast reconstruction with fat grafting after surgery is a major area that is enhanced by the use of autologous ASCs following surgery for breast cancer [5,19]. However, approximately two-thirds of all cancer patients will undergo radiation and/or chemotherapy as part of their treatment plan. In this context, however, one must consider that radiation and chemotherapy in cancer patients might limit stem cell cellular functions important for soft tissue wound healing. If this possibility is real, then the isolation, banking, and use of autologous ASCs may be critical in order to minimize wound adverse events. Based upon this reasoning, the characterization and usage of ASCs for clinical applications in cancer patients has become a recent focus of research.



Although ASC-assisted autologous fat transfer approaches for breast reconstruction have been shown to enhance graft survival and local angiogenesis, examination of the ability of ASCs to retain their inherent cellular functions and the efficiency of recovering these stem cells in the tumor/cancer treatment arena is lacking. To date, most studies have shown that radiation and chemotherapy have a cytotoxic effect on the stem cell's proliferative and differentiation potential [20-23]. For clinical translation to patients, a better understanding of cancer treatments on ASCs improvement of fat graft survival in actual patients is sorely needed.

This review describes our work defining two critical characteristics of stem cells isolated from human adipose tissue: (1) Availability in cancer patients after receiving radiation or chemotherapy most likely to appear to require a viable source of autologous stem cells; and (2) Ability to retain their great function and recovery capacity post-radiation/chemotherapy. Within this substructure, we summarize and highlight the practical usefulness of these cells in fat grafting and reconstructive procedures in cancer patients undergoing radiation and chemotherapy.

#### EFFECT OF RADIATION ON ADIPOSE STEM CELLS

Radiation therapy is a routine treatment for patients with cancer, either before or after surgical resection. However, in patients receiving radiation, there is a local injury to the surrounding soft tissues and when the surgical insult is added to the radiated soft tissue, the result is a high rate of wound complications[24,25]. An important complication of radiation therapy in breast cancer patients is iatrogenic damage of normal breast tissue that results in chronic, painful, and disfiguring wounds[26,27]. Additionally, when tissue damage does occur, the reconstructive surgeon is faced with the challenge of restoring normal tissue health or being left with poor survival[28]. To restore the damaged tissue a better understanding is needed concerning whether stem cells contribute to wound healing and the degree to which radiation might hinder the ability of stem cells to participate in tissue recovery. It is well known that ASCs can promote neovascularization and healing of damaged tissues in tissue engineering applications[29,30] yet applicability in the "cancer" population has been questioned as it is likely that cancer treatment and co-morbidity adversely affect many of the cell populations. Thus, it is beneficial to determine the influence of radiation therapy on ASCs isolation and cellular functions that are considered clinically important to improve the radiation-induced wound healing in favor of positive outcomes of tissue reconstruction and repair in patients. Starting from this background, we have studied and evaluated the deficiency that radiated breast tissues have in the number and functional abilities of ASCs with emphasis on endothelial differentiation in breast cancer patients.

#### Availability and growth rate of ASCs in patients treated with radiation

Many initial studies that sought to evaluate fat as a source of stem cells examined liposuction specimens obtained from young, healthy plastic surgery patients[31-34]. However, the effects of radiation on ASC cells remain largely unknown. Several studies have shown that the growth rates of ASCs are decreased and the number of apoptotic cells is remarkably increased after irradiation[19,35-37]. However, little is known about the cell viability of ASCs from patients after exposure to radiation therapy. Also, a cell type employed in tissue engineering must be readily and abundantly available in the specific target patient population (e.g., breast cancer patients) that it is intended for use in [38-40]. Therefore, we have studied the availability of ASCs in breast cancer patients undergoing elective radiation surgical procedures (full manuscript in preparation). Each patient that had previously received radiation therapy in one breast, but not in the contralateral breast for cancer treatment, donated approximately 25-50 g of fat from each breast to evaluate any deficiency that the radiated tissues had in ASCs numbers and functions.

To determine the influence of radiation on stem cells isolation, ASCs were isolated from normal and radiated breast tissue specimens from the same patient. The major finding of our study was that radiation did not appear to significantly influence the number of stem cells harvested as there was no difference in the number of ASCs obtained between the radiated and non-irradiated breasts. Based on the number of cells obtained one week after harvest, we did not observe any significant change in the in vitro growth rate between radiated and non-radiated ASCs in terms of the doubling time. This result suggests that ASCs can be isolated from the patients after radiation exposure and used for fat grafting and reconstruction purposes in patients post-



radiation treatment (Figure 1A).

#### Radiation effect on ASCs endothelial differentiation

The role of stem cells in the replacement of senescent or deteriorated cells of the human body is defined by their capacity for self-renewal and multilineage differentiation. Recently, cell-assisted lipotransfer studies concluded that ASCs stimulate angiogenesis and improve both graft revascularization and survival[41-43]. Indeed the ability to acquire characteristics of cells resident within neovascularization, as the result of differentiation into EC, represents another important aspect of the ASC's usefulness in fat grafting and healing of damaged tissues.

ASCs can differentiate into a wide range of cell types, including EC and also ASCs have been shown to express higher levels of the angiogenic factors such as VEGF, hepatocyte growth factor (HGF), and insulin-like growth factor (IGF), which enhance their involvement in angiogenesis that promotes graft retention [27,43,44]. Furthermore, a sufficient density of mesenchymal stem cells and proper differentiation is necessary to improve the viability of the fat grafts and promote wound healing. To further understand the use of autologous ASCs in reconstructive procedures following radiation, we have examined the effects of radiation on angiogenic capabilities of ASCs obtained from breast tissue specimens in patients after exposure to the radiation treatment, and especially their ability to differentiate into a functional EC-phenotype which is known to participate in the regeneration of capillary networks.

In these experiments, the functions of ASCs from the radiated breast tissue specimens with an emphasis on endothelial differentiation, including expression of endothelial-specific markers PECAM-1 (CD31), von Willebrand factor (vWF), and endothelial nitric oxide synthase (eNOS) were compared to the ASCs from the nonradiated side of the breast. After three weeks of culture in EC-differentiation media, we found there was a very large and significant difference in EC differentiation capacity in the radiated ASCs based on the decreased expression of all three above endothelial markers between radiated and non-radiated breast tissue specimens using real-time PCR analysis (Figure 1B). Taken together, our studies characterized the ASC cell viability and differentiation capacity in response to irradiation and suggest that radiation therapy has deleterious effects on ASC differentiation capacity towards EC which may represent the root cause of chronic wounding and poor fat graft survival in patients, i.e., radiation appears to damage the ability of ASCs to repopulate the microvasculature with the correct cell types. This finding requires further study with a larger number of patients examining whether ASCs have the function recovery potential of pro-angiogenic and pro-adipogenic phenotypes following radiation treatment after fat transplantation.

#### EFFECT OF CHEMOTHERAPY ON ADIPOSE STEM CELLS

To date, many of the original studies have mainly focused on cytotoxic damage to mesenchymal stem cells resulting from chemotherapy with little attention given to the recovery of the stem cells' viability and cellular function capability after exposure to chemotherapeutic drug treatment [22,23,45,46]. The *in vitro* human data from our lab and others demonstrate that direct exposure to chemotherapeutic agents decreases the proliferation rate and multi-potency differentiating abilities of ASCs[21,22,24]. However, little is known about the effects of chemotherapy on ASCs viability outcomes in patients. It is therefore important and necessary to understand the ASCs damage pattern caused by chemotherapy and to know what preserves or destroys stem cells for best clinical practice. In these studies, we have determined the function recovery potential of ASCs by examining: (1) In vitro human ASCs treated with three commonly utilized clinical chemotherapeutic agents: paclitaxel (PTX), 5-fluorouracil, and doxorubicin for 3 d followed by a washout period (no drugs) of a week; (2) In vivo rats given intravenous PTX injections for 2 wk followed by cessation of drug treatment for an additional 2 wk; and (3) Isolation and evaluation of ASCs functional capacity from patients that received neoadjuvant chemotherapy (NAC) compared with the ASCs from patients not-receiving chemotherapy treatment.

#### Recovery potential of ASCs after post-chemo-treatment

Recovering cell viability and differentiation capability after treatment with chemotherapeutic drugs represents another important aspect of determining stem cell usefulness in cancer patients for stem cell-based targeted therapy and reconstruction. A few reports have now demonstrated the resistance of ASCs to chemotherapeutic



Characteristics	Patient popul	ation ( <i>n</i> = 8)		25		
	Post radiation	No-radiation	cont	20 -		
ASC yield	3.5% of SVF	4.2% of SVF	io the	15 -		
Growth rate (dT)	25.68(h)	25.73 (h)	ed t	10-		
Adipocyte	(+)	(+)	par	10		T
Osteocyte	(+)	(+)	ШQ	5 -		
Endothelial	(-)	(+)	e	-	т	

Figure 1 Availability of adipose-derived stem cells in patients after radiation exposure. A: The number of stem cells harvested and growth rates did not appear to be affected by radiation; B: Decreased expression of endothelial markers of CD31, von Willebrand factor, and endothelial nitric oxide synthase between the adipose-derived stem cells from radiated (orange) and non-radiated (blue) breast tissue specimens. vWF: von Willebrand factor; eNOS: Endothelial nitric oxide synthase; ASC: Adipose-derived stem cell.

Fold

agents and their maintenance of phenotype including their potential of differentiating into multiple cell lineages *in vitro* after treatment[46-48]. To further understand the use of autologous ASCs in reconstructive procedures following chemotherapy, we have examined if ASCs can recover their cellular activities post chemotherapy treatment using *in vitro* and *in vivo* models[23].

CD31

R-

vWF

eNOS

After the withdrawal of the drugs, the cells were cultured for an additional 9-d. The ASCs showed slow recovery of cell growth capacity in lower doses of the drugs, but not full recovery since ASCs numbers remained significantly below that of controls[23, 49]. As this time-lapse is considerably after the withdrawal of the drugs for longer than the 9-d, ASCs likely recovered from the inhibitory effects on cell growth with the extra time. Additionally, as the current clinical practice is to provide the patient chemotherapy drugs before and after surgical excision of the tumor, evaluation of the functional recovery of ASCs in the patient should be mapped to the patient's treatment and recovery time[23].

To complement our *in vitro* findings, we investigated *in vivo* the ASCs from the animals treated for 2-wk with PTX followed by washout of drug for an additional 2-wk. We observed that ASCs displayed recovery of adipogenic, osteogenic, and endothelial differentiation potentials in the cells isolated from the 2-wk post cessation PTX injection groups when compared with the active PTX-treated groups[23]. However, full recovery of ASC viability was not achieved as the number of ASCs was still significantly lower than for controls[23]. This finding requires further confirmation with a comparison of ASCs before, post-drug administration, and after cessation of treatment in the same animal to precisely determine if the chemotherapy is responsible for the alteration in the function and recovery. Taken together, our *in vitro* cell and *in vivo* animal results support ASCs having the potential to recover differentiation capacity after exposure to chemotherapeutic agents (Figure 2)[23].

#### Availability of ASCs in patients receiving chemotherapy

Recently, ASC-based therapies were successfully used for regenerative medicine such as breast cancer and reconstructive surgery [12,44,50]. Accordingly, our group examined the availability of ASCs in breast cancer patients. ASCs were isolated from adipose tissue from the cancerous breast and the opposite side noncancerous breast from the same patient post-NAC treatment about 6-8 wk[49]. These findings were compared with the patients that did not receive NAC with regard to their ASC cell yield, proliferation rates, and particularly their potential for differentiation into an adipocyte-phenotype[49].

The numbers of ASCs obtained were not altered in patients after receiving the chemo treatment compared with the non-chemotherapy treatment patients. The cellular growth rates of ASCs isolated from the chemo-treatment patients were also not affected upon culture *in vitro*, and these cells appeared to retain the capacity to acquire adipocyte traits similar to those ASCs isolated from non-chemotherapy patients. Additionally, we found that the number of ASCs yielded and their proliferation rates from the tumor primary breast tissues were decreased compared to ASCs obtained from the normal breast tissue in both NAC and non-NAC treatment groups. However, the adipogenic differentiation capacity of ASCs from the tumor primary

A





Figure 2 In vitro and in vivo evaluation of recovery potency of adipose-derived stem cells after chemotherapy exposure. In both the in vitro and animal studies, after cessation of drugs, adipose-derived stem cells exhibited partial recovery (-/+) of cell growth and recovery (+) of multipotent differentiation capabilities

> breast tissue and the normal breast tissue were similar (Table 1). This finding indicated that in patients who received chemotherapy, the treatment does not significantly change the ASCs phenotype acquired in the tumor environment. In this study, we found that notwithstanding the in vitro evidence of negative effects on ASCs after exposure to chemotherapeutic agents, clinical relevance is questionable as examination of the patient's ASCs reveals that these cells exhibit functional recovery of adipogenic differentiation after receiving NAC treatment[49]. Moreover, the cancerous side of the breast did appear to adversely affect stem cell yield, but not to a point where stem cell harvest would be considered impractical at the post-chemo treatment stage in this patient population. Hence, more studies are needed to evaluate the influence of chemotherapy in breast cellular interactions involving ASCs/or cancer stem cells and tumor cells. Taken together, these results indicate that stem cell availability was not proven inferior for the population receiving chemotherapy or having the cancerous disease.

#### ADDITIONAL STUDIES WITH ASCS

An important aspect of designing a fat graft is to enhance the viability and survival rate of transplanted fat tissues. However, one of the main challenges in enhancing the viability of fat grafting is to provide a sufficient and functional vasculature[43]. The preliminary data above suggests that radiation therapy has deleterious effects on ASC differentiation capacity towards EC, i.e., appears to damage the ability of ASCs to repopulate the microvasculature. Therefore, successful use of these stem cells will likely require isolation and banking before radiation treatment. Fat grafting used in breast reconstruction following surgery and radiation for breast cancer is accompanied by a relatively high failure rate due to absorption of the grafted fat, and it is suspected that this absorption results from the lack of vascular support to the grafted tissue[17, 51]. Therefore, it remains a challenge for researchers to seek an effective solution to improve angiogenesis that would result in boosting the efficiency of survival in fat



Table 1 Availability of adipose-derived stem cells in patients receiving chemotherapy							
Patient population	ASC yield (million/gram of fat)	d (million/gram of fat) Growth rates (dT) (h)					
Received NAC ( $n = 11$ )							
Tumor side	$0.69 \pm 0.20$	37.44	+				
Normal side	$0.99 \pm 0.29$	36.48	+				
	P = NS vs no received NAC	P = NS					
No receiving NAC ( <i>n</i> = 10)							
Tumor side	$0.52\pm0.10$	33.84	+				
Normal side	$0.71 \pm 0.17$	31.20	+				
	P = NS vs normal side	P = NS					

The number of adipose-derived stem cells (ASCs) harvested and growth rates did not appear to be significantly affected in patients post-chemotherapy; these cells appeared to retain the capacity to acquire adipocyte traits similar to the ASCs from patients not receiving chemotherapy. ASC: Adipose-derived stem cell; NAC: neoadjuvant chemotherapy; NS: Not significant.

transplantation[42,43,52].

In our study, we focused on investigating the potential of ASCs differentiation toward an endothelial phenotype requirement into the fat transplantation, thereby improving both survival and neovascularization for autologous fat transplantation. Hereby, taking advantage of the ASCs responsiveness to endothelial differentiation, we have examined the effect of fat grafting assisted with ASCs, or ASCs differentiated to a functional EC-phenotype, on the degree of survival and neovascularization in the animal model. Some studies suggested that fat graft survival is mainly dependent on successful vascularization so that adipogenesis is associated with capillary angiogenesis allowing adipocyte differentiation within clusters of endothelial and stromal cells[53,54]. Importantly, ASCs induced to an EC-like phenotype with lipotransfer increased the graft volume retention and revascularization, which represent potential mechanisms for adipose transplantation (Figure 3)[43].

#### CONCLUSION

Adult autologous adipose derived-stem cells represent an important source of cells for fat grafting and breast reconstruction. Their usefulness is directly related to the availability of the cells in patients that undergo cancer treatment as well as their maintenance of cell yield, growth, and multipotent differentiation potential postradiation or chemo-treatment. Studies from our group and others suggest that stem cells derived from cancer patient adipose tissue appear to maintain their important characteristics; ASCs provide a potential to retain their capacity as a source of autologous stem cells for fat grafting and reconstruction in cancer patients. Going forward, it will be necessary to determine whether fat grafting assisted with ASCs differentiated to a functional EC phenotype can be achieved following post-radiation therapy. This will likely depend upon the ASCs' ability to neovascularize secondary to improved graft survival and healing of damaged tissues. To determine if ASCs altered function and recovery components may be due to the chemotherapy, it is important to perform more preclinical and clinical studies to elucidate the effects of chemotherapy before, during, and after the cessation period in the same patient. Furthermore, there has been a keen interest in optimizing the microenvironmental cues important to modulate the local environment and thus stimulate angiogenesis of the ASCs for improving grafted fat survival in a previous radiotherapy-treated field and this line of research should be aggressively pursued.



Figure 3 Adipose-derived stem cell-assisted transplanted fat lipoaspirate improved fat graft angiogenesis. Immunofluorescence micrograph (200 ×; CD31 and human nuclear stain) of human adipose-derived stem cells (ASCs) with fat lipoaspirate injected into the rat for 8 wk. Fat lipoaspirate was mixed with human endothelial differentiated ASCs and then subcutaneously injected into the adult male Sprague-Dawley rat's dorsum. Immunofluorescence staining analysis of the transplants was performed with an anti-human nuclear antibody (red) to detect if the human ASCs proliferated in transplanted tissues. The CD31 staining (green) was used to detect capillary endothelial cells. The merging of the red fluorescence of anti-human nuclear with the green fluorescence of CD31 revealed 3 yellow endothelial cells, indicating that the delivery of human ASCs promoted neovascularization.

#### REFERENCES

- Coleman SR. Structural fat grafting: more than a permanent filler. Plast Reconstr Surg 2006; 118: 1 108S-120S [PMID: 16936550 DOI: 10.1097/01.prs.0000234610.81672.e7]
- Pearl RA, Leedham SJ, Pacifico MD. The safety of autologous fat transfer in breast cancer: lessons from stem cell biology. J Plast Reconstr Aesthet Surg 2012; 65: 283-288 [PMID: 21820375 DOI: 10.1016/j.bjps.2011.07.017]
- Yoshimura K, Sato K, Aoi N, Kurita M, Inoue K, Suga H, Eto H, Kato H, Hirohi T, Harii K. Cell-3 assisted lipotransfer for facial lipoatrophy: efficacy of clinical use of adipose-derived stem cells. Dermatol Surg 2008; 34: 1178-1185 [PMID: 18513295 DOI: 10.1111/j.1524-4725.2008.34256.x]
- Hassan WU, Greiser U, Wang W. Role of adipose-derived stem cells in wound healing. Wound Repair Regen 2014; 22: 313-325 [PMID: 24844331 DOI: 10.1111/wrr.12173]
- Yoshimura K. Cell-Assisted Lipotransfer and Therapeutic Use of Adipose Stem Cells Thereafter. 5 Aesthetic Plast Surg 2020; 44: 1266-1267 [PMID: 32766904 DOI: 10.1007/s00266-020-01781-4]
- 6 Naderi N, Combellack EJ, Griffin M, Sedaghati T, Javed M, Findlay MW, Wallace CG, Mosahebi A, Butler PE, Seifalian AM, Whitaker IS. The regenerative role of adipose-derived stem cells (ADSC) in plastic and reconstructive surgery. Int Wound J 2017; 14: 112-124 [PMID: 26833722 DOI: 10.1111/iwj.12569
- Zuk PA, Zhu M, Mizuno H, Huang J, Futrell JW, Katz AJ, Benhaim P, Lorenz HP, Hedrick MH. 7 Multilineage cells from human adipose tissue: implications for cell-based therapies. Tissue Eng 2001; 7: 211-228 [PMID: 11304456 DOI: 10.1089/107632701300062859]
- 8 Wilson A, Butler PE, Seifalian AM. Adipose-derived stem cells for clinical applications: a review. Cell Prolif 2011; 44: 86-98 [PMID: 21199013 DOI: 10.1111/j.1365-2184.2010.00736.x]
- 9 Nan H, Huang J, Li H, Li Q, Liu D. Assessment of biological characteristics of adipose tissue-derived stem cells co-labeled with Molday ION Rhodamine BTM and green fluorescent protein in vitro. Mol Med Rep 2013; 8: 1446-1452 [PMID: 24065138 DOI: 10.3892/mmr.2013.1694]
- Moseley TA, Zhu M, Hedrick MH. Adipose-derived stem and progenitor cells as fillers in plastic and 10 reconstructive surgery. Plast Reconstr Surg 2006; 118: 121S-128S [PMID: 16936551 DOI: 10.1097/01.prs.0000234609.74811.2e]
- Matsumoto D, Sato K, Gonda K, Takaki Y, Shigeura T, Sato T, Aiba-Kojima E, Iizuka F, Inoue K, 11 Suga H, Yoshimura K. Cell-assisted lipotransfer: supportive use of human adipose-derived cells for soft tissue augmentation with lipoinjection. Tissue Eng 2006; 12: 3375-3382 [PMID: 17518674 DOI: 10.1089/ten.2006.12.3375]
- 12 Fu S, Luan J, Xin M, Wang Q, Xiao R, Gao Y. Fate of adipose-derived stromal vascular fraction cells after co-implantation with fat grafts: evidence of cell survival and differentiation in ischemic adipose tissue. Plast Reconstr Surg 2013; 132: 363-373 [PMID: 23897335 DOI: 10.1097/PRS.0b013e31829588b3
- 13 Tan SS, Ng ZY, Zhan W, Rozen W. Role of Adipose-derived Stem Cells in Fat Grafting and Reconstructive Surgery. J Cutan Aesthet Surg 2016; 9: 152-156 [PMID: 27761084 DOI: 10.4103/0974-2077.191672
- Kim EH, Heo CY. Current applications of adipose-derived stem cells and their future perspectives. 14 World J Stem Cells 2014; 6: 65-68 [PMID: 24567789 DOI: 10.4252/wjsc.v6.i1.65]
- Kern S, Eichler H, Stoeve J, Klüter H, Bieback K. Comparative analysis of mesenchymal stem cells 15



from bone marrow, umbilical cord blood, or adipose tissue. Stem Cells 2006; 24: 1294-1301 [PMID: 16410387 DOI: 10.1634/stemcells.2005-0342]

- Yoshimura K, Shigeura T, Matsumoto D, Sato T, Takaki Y, Aiba-Kojima E, Sato K, Inoue K, 16 Nagase T, Koshima I, Gonda K. Characterization of freshly isolated and cultured cells derived from the fatty and fluid portions of liposuction aspirates. J Cell Physiol 2006; 208: 64-76 [PMID: 16557516 DOI: 10.1002/jcp.20636]
- 17 Malhaire C, Hequet D, Falcou MC, Feron JG, Tardivon A, Leduey A, Guillot E, Mosseri V, Rouzier R, Couturaud B, Reyal F. Outcome of oncoplastic breast-conserving surgery following bracketing wire localization for large breast cancer. Breast 2015; 24: 370-375 [PMID: 25913288 DOI: 10.1016/j.breast.2015.02.037]
- 18 American Society of Plastic Surgeons. Report of the 2010 plastic surgery statistics: ASPS national clearinghouse of plastic surgery procedural statistics. [cited 15 Feb 2021]. In: American Society of Plastic Surgeons [Internet]. Available from: https://www.plasticsurgery.org
- Forcheron F, Agay D, Scherthan H, Riccobono D, Herodin F, Meineke V, Drouet M. Autologous 19 adipocyte derived stem cells favour healing in a minipig model of cutaneous radiation syndrome. PLoS One 2012; 7: e31694 [PMID: 22348120 DOI: 10.1371/journal.pone.0031694]
- Mvula B, Mathope T, Moore T, Abrahamse H. The effect of low level laser irradiation on adult 20 human adipose derived stem cells. Lasers Med Sci 2008; 23: 277-282 [PMID: 17713825 DOI: 10.1007/s10103-007-0479-1]
- Pike S, Zhang P, Wei Z, Wu N, Klinger A, Chang S, Jones R, Carpenter J, Brown SA, DiMuzio P, Tulenko T, Liu Y. In vitro effects of tamoxifen on adipose-derived stem cells. Wound Repair Regen 2015; 23: 728-736 [PMID: 26043659 DOI: 10.1111/wrr.12322]
- 22 Li J, Law HK, Lau YL, Chan GC. Differential damage and recovery of human mesenchymal stem cells after exposure to chemotherapeutic agents. Br J Haematol 2004; 127: 326-334 [PMID: 15491295 DOI: 10.1111/j.1365-2141.2004.05200.x]
- Harris WM, Zhang P, Plastini M, Ortiz T, Kappy N, Benites J, Alexeev E, Chang S, Brockunier R, 23 Carpenter JP, Brown SA. Evaluation of function and recovery of adipose-derived stem cells after exposure to paclitaxel. Cytotherapy 2017; 19: 211-221 [PMID: 27887867 DOI: 10.1016/j.jcyt.2016.10.010]
- 24 Gülçelik MA, Doğan L, Karaman N, Turan M, Kahraman YS, Akgül GG, Özaslan C. Intraoperative boost radiation effects on early wound complications in breast cancer patients undergoing breastconserving surgery Turk J Med Sci 2017; 47: 1185-1190 [PMID: 29156861 DOI: 10.3906/sag-1605-48]
- Ruano-Ravina A, Cantero-Muñoz P, Eraso Urién A. Efficacy and safety of intraoperative 25 radiotherapy in breast cancer: a systematic review. Cancer Lett 2011; 313: 15-25 [PMID: 21930343 DOI: 10.1016/j.canlet.2011.08.020]
- Yun JH, Diaz R, Orman AG. Breast Reconstruction and Radiation Therapy. Cancer Control 2018; 26 25: 1073274818795489 [PMID: 30132338 DOI: 10.1177/1073274818795489]
- Nelson JA, Disa JJ. Breast Reconstruction and Radiation Therapy: An Update. Plast Reconstr Surg 27 2017; 140: 60S-68S [PMID: 29064923 DOI: 10.1097/PRS.000000000003943]
- Rigotti G, Marchi A, Galiè M, Baroni G, Benati D, Krampera M, Pasini A, Sbarbati A. Clinical 28 treatment of radiotherapy tissue damage by lipoaspirate transplant: a healing process mediated by adipose-derived adult stem cells. Plast Reconstr Surg 2007; 119: 1409-1422 [PMID: 17415234 DOI: 10.1097/01.prs.0000256047.47909.71]
- 29 Miranville A, Heeschen C, Sengenès C, Curat CA, Busse R, Bouloumié A. Improvement of postnatal neovascularization by human adipose tissue-derived stem cells. Circulation 2004; 110: 349-355 [PMID: 15238461 DOI: 10.1161/01.CIR.0000135466.16823.D0]
- Lopatina T, Kalinina N, Karagyaur M, Stambolsky D, Rubina K, Revischin A, Pavlova G, 30 Parfyonova Y, Tkachuk V. Adipose-derived stem cells stimulate regeneration of peripheral nerves: BDNF secreted by these cells promotes nerve healing and axon growth de novo. PLoS One 2011; 6: e17899 [PMID: 21423756 DOI: 10.1371/journal.pone.0017899]
- Miana VV, González EAP. Adipose tissue stem cells in regenerative medicine. 31 Ecancermedicalscience 2018; 12: 822 [PMID: 29662535 DOI: 10.3332/ecancer.2018.822]
- Katz AJ, Tholpady A, Tholpady SS, Shang H, Ogle RC. Cell surface and transcriptional 32 characterization of human adipose-derived adherent stromal (hADAS) cells. Stem Cells 2005: 23: 412-423 [PMID: 15749936 DOI: 10.1634/stemcells.2004-0021]
- 33 Zuk PA, Zhu M, Ashjian P, De Ugarte DA, Huang JI, Mizuno H, Alfonso ZC, Fraser JK, Benhaim P, Hedrick MH. Human adipose tissue is a source of multipotent stem cells. Mol Biol Cell 2002; 13: 4279-4295 [PMID: 12475952 DOI: 10.1091/mbc.e02-02-0105]
- 34 DiMuzio P, Tulenko T. Tissue engineering applications to vascular bypass graft development: the use of adipose-derived stem cells. J Vasc Surg 2007; 45 Suppl A: A99-103 [PMID: 17544030 DOI: 10.1016/j.jvs.2007.02.046]
- Chang P, Qu Y, Liu Y, Cui S, Zhu D, Wang H, Jin X. Multi-therapeutic effects of human adipose-35 derived mesenchymal stem cells on radiation-induced intestinal injury. Cell Death Dis 2013; 4: e685 [PMID: 23788042 DOI: 10.1038/cddis.2013.178]
- Mvula B, Moore TJ, Abrahamse H. Effect of low-level laser irradiation and epidermal growth factor 36 on adult human adipose-derived stem cells. Lasers Med Sci 2010; 25: 33-39 [PMID: 19172344 DOI: 10.1007/s10103-008-0636-1
- Drake DB, Oishi SN. Wound healing considerations in chemotherapy and radiation therapy. Clin 37



Plast Surg 1995; 22: 31-37 [PMID: 7743707]

- Saha S, Bhanja P, Liu L, Alfieri AA, Yu D, Kandimalla ER, Agrawal S, Guha C. TLR9 agonist 38 protects mice from radiation-induced gastrointestinal syndrome. PLoS One 2012; 7: e29357 [PMID: 22238604 DOI: 10.1371/journal.pone.0029357]
- 39 Jeong W, Yang X, Lee J, Ryoo Y, Kim J, Oh Y, Kwon S, Liu D, Son D. Serial changes in the proliferation and differentiation of adipose-derived stem cells after ionizing radiation. Stem Cell Res Ther 2016; 7: 117 [PMID: 27530249 DOI: 10.1186/s13287-016-0378-0]
- 40 Baaße A, Machoy F, Juerß D, Baake J, Stang F, Reimer T, Krapohl BD, Hildebrandt G. Radiation Sensitivity of Adipose-Derived Stem Cells Isolated from Breast Tissue. Int J Mol Sci 2018; 19 [PMID: 29986519 DOI: 10.3390/ijms19071988]
- Toyserkani NM, Quaade ML, Sørensen JA. Cell-Assisted Lipotransfer: A Systematic Review of Its 41 Efficacy. Aesthetic Plast Surg 2016; 40: 309-318 [PMID: 26893280 DOI: 10.1007/s00266-016-0613-1
- Jiang A, Li M, Duan W, Dong Y, Wang Y. Improvement of the survival of human autologous fat 42 transplantation by adipose-derived stem-cells-assisted lipotransfer combined with bFGF. ScientificWorldJournal 2015; 2015: 968057 [PMID: 25695105 DOI: 10.1155/2015/968057]
- Harris WM, Plastini M, Kappy N, Ortiz T, Chang S, Brown S, Carpenter JP, Zhang P. Endothelial Differentiated Adipose-Derived Stem Cells Improvement of Survival and Neovascularization in Fat Transplantation. Aesthet Surg J 2019; 39: 220-232 [PMID: 29846494 DOI: 10.1093/asj/sjy130]
- 44 Zhu M, Zhou Z, Chen Y, Schreiber R, Ransom JT, Fraser JK, Hedrick MH, Pinkernell K, Kuo HC. Supplementation of fat grafts with adipose-derived regenerative cells improves long-term graft retention. Ann Plast Surg 2010; 64: 222-228 [PMID: 20098110 DOI: 10.1097/SAP.0b013e31819ae05c
- 45 Mueller LP, Luetzkendorf J, Mueller T, Reichelt K, Simon H, Schmoll HJ. Presence of mesenchymal stem cells in human bone marrow after exposure to chemotherapy: evidence of resistance to apoptosis induction. Stem Cells 2006; 24: 2753-2765 [PMID: 16931776 DOI: 10.1634/stemcells.2006-0108]
- Liang W, Xia H, Li J, Zhao RC. Human adipose tissue derived mesenchymal stem cells are resistant 46 to several chemotherapeutic agents. Cytotechnology 2011; 63: 523-530 [PMID: 21761127 DOI: 10.1007/s10616-011-9374-5]
- 47 Beane OS, Fonseca VC, Darling EM. Adipose-derived stem cells retain their regenerative potential after methotrexate treatment. Exp Cell Res 2014; 327: 222-233 [PMID: 24992046 DOI: 10.1016/j.vexcr.2014.06.015]
- Oi Z, Zhang Y, Liu L, Guo X, Qin J, Cui G. Mesenchymal stem cells derived from different origins 48 have unique sensitivities to different chemotherapeutic agents. Cell Biol Int 2012; 36: 857-862 [PMID: 22694597 DOI: 10.1042/CBI20110637]
- Hagaman AR, Zhang P, Koko KR, Nolan RS, Fromer MW, Gaughan J, Matthews M. Isolation and 49 identification of adipose-derived stromal/stem cells from breast cancer patients after exposure neoadjuvant chemotherapy. World J Exp Med 2020; 10: 26-40 [PMID: 32399395 DOI: 10.5493/wjem.v10.i3.26]
- Zhou Y, Wang J, Li H, Liang X, Bae J, Huang X, Li Q. Efficacy and Safety of Cell-Assisted 50 Lipotransfer: A Systematic Review and Meta-Analysis. Plast Reconstr Surg 2016; 137: 44e-57e [PMID: 26710060 DOI: 10.1097/PRS.0000000000001981]
- 51 Philips BJ, Marra KG, Rubin JP. Healing of grafted adipose tissue: current clinical applications of adipose-derived stem cells for breast and face reconstruction. Wound Repair Regen 2014; 22 Suppl 1: 11-13 [PMID: 24813358 DOI: 10.1111/wrr.12164]
- Zamora DO, Natesan S, Becerra S, Wrice N, Chung E, Suggs LJ, Christy RJ. Enhanced wound 52 vascularization using a dsASCs seeded FPEG scaffold. Angiogenesis 2013; 16: 745-757 [PMID: 23709171 DOI: 10.1007/s10456-013-9352-y]
- Philips BJ, Grahovac TL, Valentin JE, Chung CW, Bliley JM, Pfeifer ME, Roy SB, Dreifuss S, 53 Kelmendi-Doko A, Kling RE, Ravuri SK, Marra KG, Donnenberg VS, Donnenberg AD, Rubin JP. Prevalence of endogenous CD34+ adipose stem cells predicts human fat graft retention in a xenograft model. Plast Reconstr Surg 2013; 132: 845-858 [PMID: 23783061 DOI: 10.1097/PRS.0b013e31829fe5b11
- Paik KJ, Zielins ER, Atashroo DA, Maan ZN, Duscher D, Luan A, Walmsley GG, Momeni A, 54 Vistnes S, Gurtner GC, Longaker MT, Wan DC. Studies in Fat Grafting: Part V. Cell-Assisted Lipotransfer to Enhance Fat Graft Retention Is Dose Dependent. Plast Reconstr Surg 2015; 136: 67-75 [PMID: 25829158 DOI: 10.1097/PRS.000000000001367]





## Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

