**Name of journal:** **World Journal of Gastroenterology**

**ESPS Manuscript No: 6637**

**Columns: TOPIC HIGHLIGHTS**

WJG 20th Anniversary Special Issues (6): *Helicobacter pylori*

**Potential mechanism of corpus-predominant gastritis after PPI therapy in *Helicobacter pylori-*positive patients with GERD**

Mukaisho K *et al*. Bile acids, pH, and *H. pylori*

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**Received:** October 25, 2013 **Revised:** March 7, 2014

**Accepted:** April 30, 2014

**Published online:**

**Abstract**

The long-term use of proton pump inhibitors (PPIs) exacerbates corpus atrophic gastritis in patients with *Helicobacter pylori* (*H. pylori*) infection. To identify a potential mechanism for this change, we discuss interactions between pH, bile acids, and *H. pylori*. Duodenogastric reflux, which includes bile, occurs in healthy individuals, and bile reflux is increased in patients with gastroesophageal reflux disease (GERD). Diluted human plasma and bile acids have been found to be significant chemoattractants and chemorepellents, respectively, for the bacillus *H. pylori*. Although only taurine conjugates, with a *pKa* of 1.8–1.9, are soluble in an acidic environment, glycine conjugates, with a *pKa* of 4.3–5.2, as well as taurine-conjugated bile acids are soluble in the presence of PPI therapy. Thus, the soluble bile acid concentrations in the gastric contents of patients with GERD after continuous PPI therapy are considerably higher than that in those with intact acid production. In the distal stomach, the high concentration of soluble bile acids is likely to act as a bactericide or chemorepellent for *H. pylori*. In contrast, the mucous layer in the proximal stomach has an optimal bile concentration that forms chemotactic gradients with plasma components required to direct *H. pylori* to the epithelial surface. *H. pylori* may then colonize in the stomach body rather than in the pyloric antrum, which may explain the occurrence of corpus-predominant gastritis after PPI therapy in *H. pylori*-positive patients with GERD.

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**Key words:** *Helicobacter pylori*;Proton pump inhibitor; Corpus-predominant gastritis; Bile acids; Gastroesophageal reflux disease; Chemotactic gradient

**Core tip:** It has been widely accepted that the long-term use of proton pump inhibitors (PPIs) exacerbates corpus atrophic gastritis in patients with *Helicobacter pylori* (*H. pylori*) infection. Recently, we successfully demonstrated that long-term PPI administration promotes corpus atrophic gastritis in Mongolian gerbils, which are excellent models of *H. pylori*-related gastritis and adenocarcinoma. Here, we suggest a potential mechanism for corpus-predominant gastritis after PPI therapy in *H. pylori*-positive patients with gastroesophageal reflux disease that includes interactions between bile acids, pH, and *H. pylori*.

Mukaisho K, Hagiwara T, Nakayama T, Hattori T, Sugihara H. Potential mechanism of corpus-predominant gastritis after PPI therapy in *Helicobacter pylori-*positive patients with GERD. *World J Gastroenterol* 2014; In press

**Introduction**

It has been widely accepted that the long-term use of proton pump inhibitors (PPIs) exacerbates corpus atrophic gastritis in patients with *Helicobacter pylori (H. pylori)* infection[1-3]. Furthermore, we successfully demonstrated that long-term PPI administration promotes corpus atrophic gastritis in Mongolian gerbils, which are excellent models of *H. pylori*-related gastritis and adenocarcinoma[4-6]. Kuipers hypothesized that corpus atrophic gastritis is caused by a change from antral-predominant to corpus-predominant gastritis related to PPI therapy in *H. pylori*-positive patients[7]. However, the mechanism for this change has not been determined. In this article, we suggest a potential mechanism for corpus-predominant gastritis related to PPI therapy in *H. pylori-*positive patients with gastroesophageal reflux disease (GERD) that includes interactions between pH, bile acids, and *H. pylori*.

**Bile reflux and GERD**

The use of PPIs for GERD treatment has increased substantially over the past 2 decades[8]. Duodenogastric reflux, which includes bile, occurs in healthy individuals[9,10], but increases in patients with GERD[11], and bile acids have been detected in esophageal aspiration samples in up to 86% of patients with GERD[12]. These findings suggest that patients with GERD are likely to have increased concentration of bile acids in their gastric contents.

**Bile acids and *pKa***

The concentration of bile acids in the duodenum ranges from 2–3 mmol/L at fasting to 6–10 mmol/L after a meal[13,14]. The chemical characteristics of bile in humans are governed by the *pKa* of the individual bile acids. Free bile acids have a *pKa* of approximately 7 and comprise approximately 2% of bile. Glycine-conjugated bile acids have a *pKa* of 4.3–5.2 and comprise more than 60% of bile, and taurine-conjugated bile acids have a *pKa* of 1.8–1.9 and comprise 20% of bile, resulting in a ratio of glycine to taurine conjugates of about 3:1[15,16]. Although both glycine and taurine conjugates are soluble in neutral solutions, only taurine conjugates are soluble in an acidic environment. However, both glycine- and taurine-conjugated bile acids are soluble in the presence of PPI therapy. Thus, the concentrations of soluble bile acids markedly increase (approximately 4 times) with PPI administration.

**Bile acids and *H. pylori***

Experimentally, highly diluted human plasma and gallbladder bile (2% diluted in buffer) were found to be significant chemoattractants and chemorepellents, respectively, for the bacillus *H. pylori*[17]. Of the bile acids, which were all chemorepellents, the greatest effects were demonstrated by taurocholic and taurodeoxycholic acids[17]. In contrast, another study reported that glycine-conjugated bile acids markedly inhibit *H. pylori* growth and more so than the taurine conjugates[18]. It is strongly suspected that important chemotactic gradients are formed from chemoattractant plasma components that transude from capillaries into the mucosa and chemorepellent duodenal bile contents that reflux into the stomach[17].

It has also been reported that bile is bactericidal for *H. pylori* at higher concentrations; *H. pylori* is reduced by 0.5–1% bile and inhibited by 5% bile[19]. Several studies have reported an inverse relationship between bile reflux and the presence of *H. pylori*[20-22], which may account for the absence of *H. pylori* in the stomach with persistent biliary reflux and in the lower gastrointestinal tract[19]. These results imply that an optimal bile concentration that forms chemotactic gradients with plasma is required to direct *H. pylori* to the epithelial surface from the gastric lumen and this concentration is likely to be < 5%.

**Evidence of duodenal ulcer treatment by antisecretory therapy**

Since the 1940s, it has been recognized that the duodenal bulb pH is lower in patients with ulcer disease than in those without[23] and that antacids and antisecretory therapy, which reduce the duodenal acid load, accelerate ulcer healing. Graham et al. reported that the combination of a high duodenal acid load and *H. pylori* infection is possibly a critical event in the pathogenesis of *H. pylori*-related duodenal ulcer disease[18]. They suggested that, because glycine-conjugated bile acids are precipitated at an acidic pH, *H. pylori* can survive in gastric metaplasia, and any event that leads to an increase in the duodenal acid load predisposes patients with *H. pylori* infection to duodenal ulcer diseases[24,25]. Furthermore, any condition that reduces the duodenal acid load (*e.g.*, antisecretory therapy) allows the bile acids to remain in the solution, inhibits the growth of *H. pylori*, and promotes ulcer healing[18]. Therefore, inhibition of acid by antisecretory therapy increases the duodenal bulb pH, allowing soluble glycine-conjugated bile acids to inhibit *H. pylori* growth[24].

**Potential mechanism of corpus-predominant pangastritis after PPI therapy**

In patients with intact gastric acid production, the pyloric antrum mucous layer may have the optimal bile concentration to direct *H. pylori* to the epithelial surface and thus enable *H. pylori* colonization, while the stomach body may be less optimal for *H. pylori* colonization[17]. The relative bile concentrations, particularly those of both soluble glycine- and taurine-conjugated bile acids, in the gastric contents in patients with GERD, in whom the gastric acid production is inhibited by maintenance PPI therapy, are considerably higher than in those with intact acid production. Therefore, in the distal stomach, the high concentration of soluble bile acids likely acts as a bactericide or chemorepellent for *H. pylori*, and chemotactic gradients with plasma components are not formed to direct the *H. pylori* toward the epithelial surface. In contrast, the mucous layer in the proximal stomach has the optimal bile acid concentration to form chemotactic gradients with plasma components for *H. pylori* colonization in the presence of PPI therapy. *H. pylori* may then colonize in the stomach body and not in the pyloric antrum (Figure 1). This may explain the mechanism for corpus-predominant gastritis after PPI therapy in *H. pylori-*positive patients with GERD.

In conclusion, interactions between bile acids, pH, and *H. pylori* seem to be associated with the occurrence of corpus-predominant gastritis after PPI therapy in *H. pylori*-positive patients with GERD. It has been reported that the corpus-predominant gastritis might promote adenocarcinoma development[7]. In the clinical setting, it is recommended that patients being considered for long-term PPI therapy should be tested for *H. pylori* infection, and if present, this pathogen should be eradicated[6].

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**P-Reviewers: Kim BW, Reyes ve S-Editor:** Ma YJ **L-Editor:** **E-Editor:**

**Figure 1 Potential mechanism for thechanges in *Helicobactor pylori* colonization with proton pump inhibitor therapy.** A: Normal subject with intact acid production; B: patient with gastroesophageal reflux disease (GERD) and proton pump inhibitor (PPI) therapy. These figures show the intragastric conditions. The yellow area represents the reflux of soluble bile acids, and the blue area represents gastric juice. In normal subjects, the concentration of soluble bile acids may be < 5% in the gastric contents of the pyloric antrum (A). In contrast, increased soluble bile acid reflux with decreased gastric acid secretion acts as a bactericide for *Helicobacter pylori* (*H. pylori*) in the distal stomach; the concentration of soluble bile acids in patients with GERD under PPI maintenance therapy is considerably higher than that in normal subjects with intact acid production, especially in the distal stomach (B). The colonization of *H. pylori* therefore changes the pattern from antral-predominant to corpus-predominant. E-C junction: esophago-gastric junction.