**Name of Journal:** *World Journal of Hepatology*

**Manuscript NO:** 67651

**Manuscript Type:** ORIGINAL ARTICLE

***Retrospective Study***

**Acute liver failure with hemolytic anemia in children with Wilson’s disease: Genotype-phenotype correlations?**

Pop TL *et al*. Genotype-phenotype correlation in WD

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**Received:** April 30, 2021

**Revised:** June 23, 2021

**Accepted:** September 3, 2021

**Published online:** October 27, 2021

**Abstract**

BACKGROUND

Wilson’s disease (WD) is a rare autosomal recessive inherited disorder of copper metabolism. Acute liver failure (ALF) and hemolytic anemia represent the most severe presentation of WD in children. No clear genotype-phenotype correlations exist in WD. Protein-truncating nonsense, frame-shift, or splice-site variants may be associated with more severe disease. In contrast, missense variants may be associated with late-onset, less severe disease, and more neurological manifestations. Recently, a gene variant (HSD17B13:TA, rs72613567) with a possible hepatic protective role against toxins was associated with a less severe hepatic phenotype in WD.

AIM

To analyze the possible genotype-phenotype correlations in children with WD presented with ALF and non-immune hemolytic anemia.

METHODS

The medical records of children with WD diagnosed and treated in our hospital from January 2006 to December 2020 were retrospectively analyzed. The clinical manifestations (ALF with non-immune hemolytic anemia or other less severe forms), laboratory parameters, copper metabolism, *ATP7B* variants, and the HSD17B13:TA (rs72613567) variant were reviewed to analyze the possible genotype-phenotype correlations.

RESULTS

We analyzed the data of 51 patients with WD, 26 females (50.98%), with the mean age at the diagnosis of 12.36 ± 3.74 years. ALF and Coombs-negative hemolytic anemia was present in 8 children (15.67%), all adolescent girls. The Kayser-Fleisher ring was present in 9 children (17.65%). The most frequent variants of the *ATP7B* gene were p.His1069Gln (c.3207A>G) in 38.24% of all alleles, p.Gly1341Asp (c.4021G>A) in 26.47%, p.Trp939Cys (c.2817G>T) in 9.80%, and p.Lys844Ter (c.2530A>T) in 4.90%. In ALF with hemolytic anemia, p.Trp939Cys (c.2817G>T) and p.Lys844Ter (c.2530A>T) variants were more frequent than in other less severe forms, in which p.His1069Gln (c.3207A>G) was more frequent. p.Gly1341Asp (c.4021G>A) has a similar frequency in all hepatic forms. For 33 of the patients, the HSD17B13 genotype was evaluated. The overall HSD17B13:TA allele frequency was 24.24%. Its frequency was higher in patients with less severe liver disease (26.92%) than those with ALF and hemolytic anemia (14.28%).

CONCLUSION

It remains challenging to prove a genotype-phenotype correlation in WD patients. In children with ALF and hemolytic anemia, the missense variant~~s~~ other than p.His1069Gln (c.3207A>G) and frame-shift variants were the most frequently present in homozygous status or compound heterozygous status with site splice variants. As genetic analysis is usually time-consuming and the results are late, the importance at the onset of the ALF is questionable. If variants proved to be associated with severe forms are found in the pre-symptomatic phase of the disease, this could be essential to predict a possible severe evolution.

**Key Words:** Wilson’s disease; Children; Acute liver failure; Hemolytic anemia; *ATP7B* variant; Genotype-phenotype correlation

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**Citation:** Pop TL, Grama A, Stefanescu AC, Willheim C, Ferenci P. Acute liver failure with hemolytic anemia in children with Wilson’s disease: Genotype-phenotype correlations? *World J Hepatol* 2021; 13(10): 1428-1438

**URL:** https://www.wjgnet.com/1948-5182/full/v13/i10/1428.htm

**DOI:** https://dx.doi.org/10.4254/wjh.v13.i10.1428

**Core Tip:** Acute liver failure (ALF) and hemolytic anemia represent the most severe presentation of Wilson’s disease (WD) in children, with a possible fatal evolution. There is no definite genotype-phenotype correlation in WD, but many studies try to solve this puzzle. Our research reports a higher presence of a missense [p.Trp939Cys (c.2817G>T)] and frame-shift variant [p.Lys844Ter (c.2530A>T)] in children with ALF and hemolytic anemia, while in less severe form, p.His1069Gln (c.3207A>G) was more frequent. HSD17B13:TA variant may be associated with less severe liver disease, as it was proved to have a protective role against liver toxins.

**INTRODUCTION**

Wilson’s disease (WD) is a rare autosomal recessive inherited disorder of copper metabolism caused by homozygous or compound heterozygous variants of the *ATP7B* gene. The prevalence of WD is estimated as 1/30000[1]. The *ATP7B* gene encodes transmembrane copper-transporting ATPase (*ATP7B*) and is located on chromosome 13q14.3, containing 20 introns and 21 exons[2]. According to the Human Gene Mutation Database, more than 800 variants of the *ATP7B* gene have been described. More than half of these variants are single nucleotide missense and nonsense, and the others are insertions/deletions and splice-site variants[3,4].

The clinical forms of WD are very variable due to the copper accumulation in different organs. The age of onset has been reported to be between 2 and 70 years[5,6]. Liver disease is the first clinical manifestation in 40%-60% of WD patients, more often in the first decade[1,5,7,8]. In children, WD patients present with an incidental finding of high levels of transaminases in an asymptomatic child, acute or chronic hepatitis, or decompensation of cirrhosis (in older children and adolescents)[5,9]. As an initial form of presentation, neurological disease is described in 18%-68% of patients, mainly in young adults (20-30 years). WD’s most common neurological features are tremor, dystonia, parkinsonism, associated with dysarthria, gait and posture disturbances, drooling, and dysphagia[1]. Also, psychiatric disease (mainly mood disturbances, depression, or bipolar disease) may be present, mainly in adulthood. A decline in school performances, impulsiveness, and inappropriate behavior was reported in adolescents[1]. Hematologic disease, renal disease, skeletal and cardiac disease may be described in WD patients[1,10,11].

Acute liver failure (ALF) may be the initial presentation or a complication during WD evolution in children and young adults[12]. Approximately 2%-6% of ALF cases may be caused by WD[13,14]. A rare presentation of WD (5%[15]), ALF accompanied by a hemolytic crisis may have a severe evolution, with coagulopathy, encephalopathy, and progressive renal failure, resulting in death without an emergency liver transplantation[1,10,11,16]. This clinical form of WD occurred in 30% of children with ALF requesting liver transplantation and 60% of those with fatal evolution before transplantation[5]. Therefore, early diagnosis and referral to specialized centers are determinants for the prognosis in these patients[12].

There is a continuous interest in genotype-phenotype correlations in WD. Based on the phenotypic classification, studies tried to find a link between the genetic variants and clinical forms or severity of WD disease, important for the prognosis of the disease[2,17]. The *ATP7B* gene variants may have different effects on the presence and function of the ATPase encoded with various consequences on the clinical presentation. Many studies have tried to analyze these correlations regarding the age of the onset, neurological or hepatic form, ceruloplasmin activity, hepatic copper level, or the presence of Kayser-Fleischer (KF) ring[18-20]. Still, there is no definite genotype-phenotype correlation so far, which may be due to the disease’s high genetic heterogeneity and rareness[2,21]. Some authors suggest that the severe hepatic phenotype and earlier onset are more likely associated with the nonsense or frame-shift variants. A less severe hepatic or a neurologic phenotype is linked to missense variants [2,22-26]. The clinical presentation in WD may also be influenced by environmental and epigenetic factors or modifier genes[4]. Recently, a gene variant (HSD17B13:TA, rs72613567) with a possible hepatic protective role against toxins was associated with a less severe hepatic phenotype in WD[27].

Our study aimed to analyze the possible genotype-phenotype correlations in children with WD presented with ALF and non-immune hemolytic anemia and to investigate the most common *ATP7B* variants in our patients with this severe form of the disease.

**MATERIALS AND METHODS**

The medical records of children with WD diagnosed and treated in our hospital from January 2006 to December 2020 were retrospectively analyzed. The clinical manifestations (acute or chronic liver disease, neurologic disease, ALF with non-immune hemolytic anemia), laboratory parameters, copper metabolism, and *ATP7B* variants were reviewed.

Diagnostic of WD was based on positive family history, clinical symptoms (including the presence of KF ring), and laboratory tests (low serum ceruloplasmin, < 20 mg/dL, elevated 24-h urinary copper excretion, baseline or stimulated by penicillamine) following the current diagnostic and management guidelines[28,29]. ALF with hemolytic anemia was diagnosed on the coagulopathy (prolonged prothrombin time, increased international normalized ratio (INR) > 2 without hepatic encephalopathy or > 1.5 in the presence of encephalopathy), low hemoglobin level, and negative Coombs test. Laboratory tests were performed using standard methods. None of our patients had a liver biopsy to assess the histology, as we could not measure the copper content in our service. The severity of the fibrosis was evaluated at diagnosis and during the follow-up using a non-invasive assessment of liver stiffness by transient elastography (FibroScan, Echosense, France)[9].

The molecular analysis of the *ATP7B* gene was performed using a semi-nested polymerase chain reaction-based restriction fragment length polymorphism assay for p.His1069Gln (c.3207A>G) variant detection as previously described. If negative or heterozygous for p.His1069Gln (c.3207A>G) variant, samples were Sanger sequenced by the ABI Prism 310 Genetic Analyzer (Perkin Elmer; Norwalk, CT, United States) until 2012, followed by the 3500 Genetic Analyzer (Applied Biosystems; Foster City, CA, United States) using published primers.

The HSD17B13:TA (rs72613567) variant was determined using allelic discrimination real-time polymerase chain reaction and validated by Sanger sequencing in normal controls having different HSD17B13 genotypes. Unfortunately, this evaluation was technically possible only for the second half of the study, and only 33 patients were assessed. We included only the children with WD confirmed by molecular analysis, and we excluded all suspected WD patients without genetic confirmation or with incomplete data.

We analyzed the clinical and laboratory features, including the most frequent variants in children with ALF and hemolytic anemia compared to those with other clinical forms.

Statistical analysis of the data collected was performed using Statistica 13.5 (Tibco Software; Palo Alto, CA, United States). The variables with normal distribution were presented as mean and standard. Comparison of continuous variables was performed using the Student t-test. Categorical variables were presented as numbers and percentages; they were compared using the Chi-square test. Two-sided *P*-values were analyzed, and the *P*-value < 0.05 was considered statistically significant.

**RESULTS**

During the last 15 years, 67 patients with WD were diagnosed and treated in our clinic. After reviewing genetic data, we included 51 patients, 26 females (50.98%), with the mean age at WD diagnosis of 12.36 ± 3.74 years (between 5 and 23 years).

Almost all patients included in our study presented liver diseases; only one was with a neurological form, and one was diagnosed following the screening due to WD in the family. Our clinic is the main pediatric hepatology service and center for expertise in pediatric liver rare disorders in Transylvania, Romania. Therefore, the selection of the patients referred to our center would be biased regarding the clinical presentation in our WD patients. ALF and Coombs-negative hemolytic anemia was present in 8 children (15.67%), all adolescent girls. The KF ring was present in 9 children (17.65%). The clinical and laboratory characteristics of the WD patients included in this study, based on the clinical onset, are presented in Table 1.

In two girls, ALF with hemolytic anemia was not the initial presentation that led to the WD diagnostic. Initially, they had only increased transaminases but progressed shortly to this severe clinical evolution.

In our patients, the most frequent variant of the *ATP7B* gene was p.His1069Gln (c.3207A>G), present in 12 children in homozygous status and 17 children in compound heterozygous status (38.24% of all alleles). p.Gly1341Asp (c.4021G>A) variant was present in 10 children in homozygous status and 7 patients in compound heterozygous status (26.47%). The other two frequent variants (mainly in patients with ALF and hemolytic anemia) were p.Trp939Cys (c.2817G>T), in 5 children in homozygous status (9.80%) and p.Lys844Ter (c.2530A>T), in one child as homozygous status and three children as a part of compound heterozygous status (4.90%). In Table 2, we present the most frequent variants grouped by the clinical form of presentation. In ALF with hemolytic anemia, p.Trp939Cys (c.2817G>T) and p.Lys844Ter (c.2530A>T) were more frequent than in other less severe forms, in which p.His1069Gln (c.3207A>G) was more frequent. p.Gly1341Asp (c.4021G>A) variant has a similar frequency in all hepatic forms.

For 33 of the patients included in our study, the HSD17B13 genotype was evaluated. The overall HSD17B13:TA allele frequency in our study was 24.24%. HSD17B13:TA allele frequency was higher in patients with less severe liver disease (26.92%) than ALF and hemolytic anemia (14.28%). Table 3 presents the demographic, clinical, and genotype association in patients investigated for the HSD17B13:TA variant.

Two patients with ALF were transplanted, five survived with the native liver following supportive intensive care, and one girl had a fatal evolution on the second day after admission. Also, another child with cirrhosis died due to severe complications before liver transplantation was possible.

**DISCUSSION**

Our study aimed to assess the possible genotype-phenotype correlations regarding WD’s most severe clinical form in children and adolescents. This endeavor in patients with WD is challenging, as was proved by many studies already published. So far, the research failed to conclude this issue due to the high heterogeneity of *ATP7B* variants (more than 1300 described) and the rareness of the disease (small series of patients). Furthermore, the increased number of compound heterozygotes involving different kinds of variants makes this analysis more difficult[18,19,24,30,31]. As phenotypic differences were reported in siblings with the same genotype or monozygotic twins, the involvement of other factors is possible[2,30,32-35]. Environmental factors (nutritional copper intake, infections, drugs, or other toxins), modifier genes, and epigenetic factors’ interaction with the genetic variants may explain the different clinical presentations in WD[4,8,18,22].

The introduction of a phenotype classification tried to ease analyzing the clinical forms in WD[17]. Our patients mainly have the hepatic form (the most frequent one in children and adolescents). Some of our older patients also had neurological and psychiatric manifestations. Not all our patients suspected of WD had a genetic confirmation of the *ATP7B* variants. Therefore, only patients with two WD variants in cis were included in our study to analyze the possible genotype-phenotype correlations.

ALF with hemolytic anemia was present in 8 children, all girls. The age of onset was higher than other hepatic presentations (acute or chronic hepatitis, autoimmune features, or cirrhosis, data not shown). The increased frequency of ALF described in females is not fully understood, but it may be explained by hormonal differences or the intervention of epigenetic factors (methylome and transcriptome differences)[4,13,18,19].

The KF ring was described in 9 children with liver disease (17.65%), three of them with ALF, and 6 in the other forms. Four of those six children with KF ring in other forms of liver disease presented neurological manifestations in their evolution. Other studies proved that ocular involvement is less frequent in hepatic than in neurological involvement[31]. The presence of the KF ring was reported lower in children. In Greek children with WD, the KF ring was present in 48.7% of those with liver disease and 16% of those diagnosed through family screening[7], while in the Italian children, only in 8.6% of those with liver disease[8]. The KF ring was present in more than half of the ALF patients, compared to 37.5% in our small series[10].

Regarding the laboratory results, the differences in children with ALF and hemolytic anemia and the other forms are expected for the bilirubin, hemoglobin, and INR. The number of white blood cells (WBC) is higher in children with ALF, and platelets are lower. There are no significant differences in the serum level of transaminases (even lower in ALF patients) and gamma-glutamyl transferase. The number of WBCs is an important risk factor as it was included in the prognostic score to predict mortality and evaluate the need for liver transplantation[5]. Also, the low level of transaminases in children with the ALF form of WD is a well-known feature and would help the diagnostic. High aspartate-aminotransferase (AST) to ALT ratio and low alkaline phosphatase to serum bilirubin ratio may be used to differentiate the WD patients in ALF[13,15]. In our cohort, two children with ALF had an AST/ALT ratio higher than 4 and only one alkaline phosphatase to bilirubin level ratio lower than 2. In children, the ratio between serum alkaline phosphatase and total bilirubin level may not always be helpful due to bone-derived alkaline phosphatase[5]. The transaminase level was lower than in children with acute hepatitis. As the age is higher in patients presenting with ALF, the evolution of the disease without any clinical sign for years explains the severe fibrosis or cirrhosis in these patients. In a large study that aims to analyze the genotype-phenotype correlation, fulminant liver failure or hemolysis were associated with liver cirrhosis in 93.4% and 66.7% of patients, higher than in the other milder presentations of WD[19].

In our children with ALF and hemolytic anemia, the serum ceruloplasmin level was lower than in the other patients but not significantly. In the meantime, the urinary copper excretion was higher, as can be expected, due to the severe necrosis associated with ALF.

No clear genotype-phenotype correlations exist in WD. Protein-truncating nonsense, frame-shift, or splice-site variants have a significant functional and structural impact on the ATP7B protein and may be associated with more severe disease (early-onset, low ceruloplasmin level, high copper content in liver). In contrast, missense variants are associated with late-onset, less severe disease and more neurological manifestations[18,25,26,36]. There are also reports of some missense variants associated with the early onset of disease with various severity in the same family[30]. Previous reports proposed the association of exon 18-20 variants with hepatic and hematological onset but not with neurological disease[37].

The most frequent variants in Central Europe, p.His1069Gln (c.3207A>G), was also the most frequent one in our cohort. It was found in homozygous or heterozygous status in 38.24% of all alleles in our study, compared with 72% in Poland, 35% in Greece, and 38% in a previous study from Romania[17,21,23,26,38]. This variant is more frequent in older patients with the neurological form of WD[3,7,23]. In our cohort, there was no child with ALF and hemolytic anemia with p.His1069Gln (c.3207A>G) variant. This is a missense variant and is probably associated with protein misfolding, abnormal phosphorylation of the P-domain, and altered ATP binding orientation and affinity[13]. R969Q, another missense variant present in our children, is almost exclusively associated with late-onset liver disease[3,23].

Another missense variant, p.Gly1341Asp (c.4021G>A), was the second most frequent one in our children. p.Gly1341Asp (c.4021G>A) is a variant of the transmembrane domain of the *ATP7B* gene and, in homozygous status, was proved to be associated with more severe and early onset of WD[39]. This variant was associated in homozygous status with ALF and/or hemolytic anemia in two children. In one girl, hemolytic anemia developed after treatment with zinc for a chronic increase of transaminases with questionable compliance. The second girl with this genotype-phenotype association has a younger sister with the same genetic status presenting only an increase of transaminases. The most frequent variant in our patients with ALF and/or hemolytic anemia was p.Trp939Cys (c.2817G>T), described previously in early-onset hepatic disease and with a high risk for liver failure in homozygotes[24]. Three adolescents (girls) with ALF presented this variant in homozygous status; the other two children (males) had the same status but did not have a severe form. The p.Lys844Ter (c.2530A>T) variant is the fourth most frequent in our cohort; it was present more in children with a severe form of WD. One girl was homozygote, and in another two girls, the variant was associated with splice-site variants in a compound heterozygous status. The p.Lys844Ter (c.2530A>T) variant is a frame-shift variant presumed to be associated with severe clinical evolution, as are also splice-site variants. It was previously described in WD patients of Hungarian origin[40] and few patients with late-onset of WD[41].

The early diagnosis of WD in children would probably prevent the evolution and sometimes the onset of the disease with a severe form. As mentioned in other studies, gender would modify the disease presentation due to different hormone balance[18,19]. If we analyze the possible influence of the sex of the patients, the severe form of the disease was present in two of the four girls and none of the boys with p.Gly1341Asp (c.4021G>A) homozygous status. All children with p.Gly1341Asp (c.4021G>A) variant in compound heterozygous status associated with p.His1069Gln (c.3207A>G) variant experienced a less severe form of WD.

HSD17B13 encodes a protein involved in regulating the biosynthesis of lipids, and by its enzymatic roles, is implicated in lipid-mediated inflammation. Recently, a protein-truncation variant (HSD17B13:TA, rs72613567) was shown to have a protective role against liver toxins, including copper toxicity in WD[27]. In our cohort, the allele frequency of HSD17B13:TA was similar to other results for the Caucasian population, higher in patients with less severe liver disease than those presented with ALF and hemolytic anemia. The age of diagnosis was higher in patients homozygous for this variant than in heterozygous status or without this variant. Even without statistical significance, these results suggest the possible role of the HSD17B13:TA variant in the modulation of the WD severity together with factors, including sex, age, *ATP7B* variant, and other gene variants.

ALF was fatal only in one of our cases included in this study. Two girls underwent emergency liver transplantation on the fourth day after their presentation in our service. The liver transplantation was performed at the Fundeni Institute in Bucharest, Romania. This clinical presentation should be regarded as an emergency[5,42]. The patients should be referred as soon as possible to a center that could provide intensive care, including extrahepatic liver support, until liver transplantation would be possible for severe cases. Unfortunately, one girl died the second day after her admission to our center.

Strengths and limitations. This study presents the largest cohort of children with genetically confirmed WD from our country and the neighboring region. It represents the first description of the possible correlation of ALF and hemolytic anemia with p.Trp939Cys (c.2817G>T) and p.Lys844Ter (c.2530A>T) variants in Eastern European children with WD. However, there are some limitations of our study. Firstly, the small number of children with this severe form made the statistical analysis of our findings difficult. Another issue is represented by the selection of patients, as our pediatric hepatology service admits mainly children and adolescents with hepatic disease. A significant limitation was the difficulty of considering and analyzing other possible factors that would lead to an acute, severe clinical form compared to children with the same genotype [p.Gly1341Asp (c.4021G>A)].

In the future, with the onset of a National Registry for patients with WD, including the genetic analyzes, more data on WD patients from Romania would be available. In the severe clinical form of WD, the genetic background would be less critical from the point of view of immediate medical care. The result of the genetic analysis would arrive with the clinician late, after the evolution of the patient would be clear. With the recent progress in screening for WD[43], the genetic analysis in children with an early suspected disease would help predict future evolution. When nonsense, frame-shift, or splicing-site variants are identified in a pre-symptomatic period, the importance of this genotype-phenotype correlation for the prognostic is evident.

**CONCLUSION**

It remains challenging to prove a genotype-phenotype correlation in WD patients due to the small number of patients in the reported series and the increased genetic heterogeneity. In children with ALF and non-immune hemolytic anemia, the nonsense variants other than p.His1069Gln [as p.Trp939Cys (c.2817G>T)] and frame-shift variants [p.Lys844Ter (c.2530A>T)] were the most frequently present in homozygous status or compound heterozygous status with site splice variants. As genetic analysis is usually time-consuming and the results are late (except in the screening of the relative of an index patient), the importance for the prognosis at the onset of the ALF is questionable. However, if variants proved to be associated with severe forms are found early in the evolution of the disease, this could be essential to predict a possible severe evolution if the patients would not follow treatment.

**ARTICLE HIGHLIGHTS**

***Research background***

There is a continuous interest in genotype-phenotype correlations in Wilson’s disease (WD).

***Research motivation***

The aim is to study the possible genotype-phenotype correlations in children with acute liver failure (ALF) and hemolytic anemia in WD.

***Research objectives***

The objectives include the analysis of *ATP7B* variants in children with ALF and hemolytic anemia in WD compared to the other clinical presentations and the possible role of the HSD17B13:TA variant in the modulation of the WD severity.

***Research methods***

The retrospective study included 63 children with WD diagnosed and follow-up during 2006-2020. The clinical manifestations (acute or chronic liver disease, neurologic disease, ALF with non-immune hemolytic anemia), laboratory parameters, copper metabolism, *ATP7B* variants, and the HSD17B13:TA (rs72613567) variant were reviewed.

***Research results***

In our cohort, in children with ALF and non-immune hemolytic anemia, the nonsense variants other than p.His1069Gln (c.3206A>G), as p.Trp939Cys (c.2817G>T), and frame-shift variants, as p.Lys844Ter (c.2530A>T), were the most frequently present. The allele frequency of HSD17B13:TA was similar to other results for the Caucasian population, higher in patients with the less severe liver disease than those presented with ALF and hemolytic anemia.

***Research conclusions***

It remains challenging to prove a genotype-phenotype correlation in WD patients due to the small number of patients in the reported series and the increased genetic heterogeneity. When nonsense, frame-shift, or splicing-site variants are identified in a pre-symptomatic period, the importance of this genotype-phenotype correlation for the prognostic is evident.

***Research perspectives***

A more extensive study involving children and adolescents with ALF and hemolytic anemia form of WD should be provided to confirm the findings. New studies are needed to evaluate the role of protective variant, HSD17B13:TA (rs72613567), in association with other factors, in less severe forms of WD in children.

**REFERENCES**

1 **Członkowska A**, Litwin T, Dusek P, Ferenci P, Lutsenko S, Medici V, Rybakowski JK, Weiss KH, Schilsky ML. Wilson disease. *Nat Rev Dis Primers* 2018; **4**: 21 [PMID: 30190489 DOI: 10.1038/s41572-018-0018-3]

2 **Chang IJ**, Hahn SH. The genetics of Wilson disease. *Handb Clin Neurol* 2017; **142**: 19-34 [PMID: 28433102 DOI: 10.1016/B978-0-444-63625-6.00003-3]

3 **Cooper DN,** Ball EV, Stenson PD, Phillips AD, Evans K, Heywood S, Hayden MJ, Chapman MM, Mort ME, Azevedo L, Millar DS. The Human Gene Mutation Database. Qiagen. [cited 22 April 2021]. Available from: http://www.hgmd.cf.ac.uk/ac/index.php

4 **Medici V**, LaSalle JM. Genetics and epigenetic factors of Wilson disease. *Ann Transl Med* 2019; **7**: S58 [PMID: 31179295 DOI: 10.21037/atm.2019.01.67]

5 **Dhawan A**, Taylor RM, Cheeseman P, De Silva P, Katsiyiannakis L, Mieli-Vergani G. Wilson's disease in children: 37-year experience and revised King's score for liver transplantation. *Liver Transpl* 2005; **11**: 441-448 [PMID: 15776453 DOI: 10.1002/lt.20352]

6 **Schmidt HH**. Role of genotyping in Wilson's disease. *J Hepatol* 2009; **50**: 449-452 [PMID: 19118916 DOI: 10.1016/j.jhep.2008.11.008]

7 **Manolaki N**, Nikolopoulou G, Daikos GL, Panagiotakaki E, Tzetis M, Roma E, Kanavakis E, Syriopoulou VP. Wilson disease in children: analysis of 57 cases. *J Pediatr Gastroenterol Nutr* 2009; **48**: 72-77 [PMID: 19172127 DOI: 10.1097/MPG.0b013e31817d80b8]

8 **Nicastro E**, Loudianos G, Zancan L, D'Antiga L, Maggiore G, Marcellini M, Barbera C, Marazzi MG, Francavilla R, Pastore M, Vajro P, D'Ambrosi M, Vegnente A, Ranucci G, Iorio R. Genotype-phenotype correlation in Italian children with Wilson's disease. *J Hepatol* 2009; **50**: 555-561 [PMID: 19118915 DOI: 10.1016/j.jhep.2008.09.020]

9 **Stefanescu AC**, Pop TL, Stefanescu H, Miu N. Transient elastography of the liver in children with Wilson's disease: Preliminary results. *J Clin Ultrasound* 2016; **44**: 65-71 [PMID: 26177868 DOI: 10.1002/jcu.22281]

10 **Shribman S**, Warner TT, Dooley JS. Clinical presentations of Wilson disease. *Ann Transl Med* 2019; **7**: S60 [PMID: 31179297 DOI: 10.21037/atm.2019.04.27]

11 **Walshe JM**. The acute haemolytic syndrome in Wilson's disease--a review of 22 patients. *QJM* 2013; **106**: 1003-1008 [PMID: 23842488 DOI: 10.1093/qjmed/hct137]

12 **Mainardi V**, Rando K, Valverde M, Olivari D, Castelli J, Rey G, Gerona S. Acute Liver Failure due to Wilson Disease: Eight Years of the National Liver Transplant Program in Uruguay. *Ann Hepatol* 2019; **18**: 187-192 [PMID: 31113589 DOI: 10.5604/01.3001.0012.7911]

13 **Camarata MA**, Gottfried M, Rule JA, Ala A, Lee WM, Todd Stravitz R, Schilsky ML. Outcomes of Acute Liver Injury in Adults Due to Wilson's Disease: Is Survival Without Transplant Possible? *Liver Transpl* 2020; **26**: 330-336 [PMID: 31838773 DOI: 10.1002/lt.25703]

14 **Grama A**, Aldea CO, Burac L, Delean D, Bulata B, Sirbe C, Duca E, Boghitoiu D, Coroleuca A, Pop TL. Etiology and Outcome of Acute Liver Failure in Children-The Experience of a Single Tertiary Care Hospital from Romania. *Children (Basel)* 2020; **7** [PMID: 33317098 DOI: 10.3390/children7120282]

15 **Eisenbach C**, Sieg O, Stremmel W, Encke J, Merle U. Diagnostic criteria for acute liver failure due to Wilson disease. *World J Gastroenterol* 2007; **13**: 1711-1714 [PMID: 17461475 DOI: 10.3748/wjg.v13.i11.1711]

16 **Tian Y**, Gong GZ, Yang X, Peng F. Diagnosis and management of fulminant Wilson's disease: a single center's experience. *World J Pediatr* 2016; **12**: 209-214 [PMID: 26041495 DOI: 10.1007/s12519-015-0026-2]

17 **Ferenci P**, Caca K, Loudianos G, Mieli-Vergani G, Tanner S, Sternlieb I, Schilsky M, Cox D, Berr F. Diagnosis and phenotypic classification of Wilson disease. *Liver Int* 2003; **23**: 139-142 [PMID: 12955875 DOI: 10.1034/j.1600-0676.2003.00824.x]

18 **Ferenci P**. Phenotype-genotype correlations in patients with Wilson's disease. *Ann N Y Acad Sci* 2014; **1315**: 1-5 [PMID: 24517292 DOI: 10.1111/nyas.12340]

19 **Ferenci P**, Stremmel W, Członkowska A, Szalay F, Viveiros A, Stättermayer AF, Bruha R, Houwen R, Pop TL, Stauber R, Gschwantler M, Pfeiffenberger J, Yurdaydin C, Aigner E, Steindl-Munda P, Dienes HP, Zoller H, Weiss KH. Age and Sex but Not ATP7B Genotype Effectively Influence the Clinical Phenotype of Wilson Disease. *Hepatology* 2019; **69**: 1464-1476 [PMID: 30232804 DOI: 10.1002/hep.30280]

20 **Shah AB**, Chernov I, Zhang HT, Ross BM, Das K, Lutsenko S, Parano E, Pavone L, Evgrafov O, Ivanova-Smolenskaya IA, Annerén G, Westermark K, Urrutia FH, Penchaszadeh GK, Sternlieb I, Scheinberg IH, Gilliam TC, Petrukhin K. Identification and analysis of mutations in the Wilson disease gene (ATP7B): population frequencies, genotype-phenotype correlation, and functional analyses. *Am J Hum Genet* 1997; **61**: 317-328 [PMID: 9311736 DOI: 10.1086/514864]

21 **Cocoş R**, Şendroiu A, Schipor S, Bohîlţea LC, Şendroiu I, Raicu F. Genotype-phenotype correlations in a mountain population community with high prevalence of Wilson's disease: genetic and clinical homogeneity. *PLoS One* 2014; **9**: e98520 [PMID: 24897373 DOI: 10.1371/journal.pone.0098520]

22 **Horslen S**, Hahn SH. Genotype-phenotype correlation in Wilson disease. *J Clin Gastroenterol* 2010; **44**: 387-388 [PMID: 20421802 DOI: 10.1097/MCG.0b013e3181d96ac4]

23 **Panagiotakaki E**, Tzetis M, Manolaki N, Loudianos G, Papatheodorou A, Manesis E, Nousia-Arvanitakis S, Syriopoulou V, Kanavakis E. Genotype-phenotype correlations for a wide spectrum of mutations in the Wilson disease gene (ATP7B). *Am J Med Genet A* 2004; **131**: 168-173 [PMID: 15523622 DOI: 10.1002/ajmg.a.30345]

24 **Usta J**, Abu Daya H, Halawi H, Al-Shareef I, El-Rifai O, Malli AH, Sharara AI, Habib RH, Barada K. Homozygosity for Non-H1069Q Missense Mutations in ATP7B Gene and Early Severe Liver Disease: Report of Two Families and a Meta-analysis. *JIMD Rep* 2012; **4**: 129-137 [PMID: 23430908 DOI: 10.1007/8904\_2011\_91]

25 **Merle U**, Weiss KH, Eisenbach C, Tuma S, Ferenci P, Stremmel W. Truncating mutations in the Wilson disease gene ATP7B are associated with very low serum ceruloplasmin oxidase activity and an early onset of Wilson disease. *BMC Gastroenterol* 2010; **10**: 8 [PMID: 20082719 DOI: 10.1186/1471-230X-10-8]

26 **Gromadzka G**, Schmidt HH, Genschel J, Bochow B, Rodo M, Tarnacka B, Litwin T, Chabik G, Członkowska A. Frameshift and nonsense mutations in the gene for ATPase7B are associated with severe impairment of copper metabolism and with an early clinical manifestation of Wilson's disease. *Clin Genet* 2005; **68**: 524-532 [PMID: 16283883 DOI: 10.1111/j.1399-0004.2005.00528.x]

27 **Ferenci P**, Pfeiffenberger J, Stättermayer AF, Stauber RE, Willheim C, Weiss KH, Munda-Steindl P, Trauner M, Schilsky M, Zoller H. HSD17B13 truncated variant is associated with a mild hepatic phenotype in Wilson's Disease. *JHEP Rep* 2019; **1**: 2-8 [PMID: 32039348 DOI: 10.1016/j.jhepr.2019.02.007]

28 **European Association for Study of Liver.**. EASL Clinical Practice Guidelines: Wilson's disease. *J Hepatol* 2012; **56**: 671-685 [PMID: 22340672 DOI: 10.1016/j.jhep.2011.11.007]

29 **Socha P**, Janczyk W, Dhawan A, Baumann U, D'Antiga L, Tanner S, Iorio R, Vajro P, Houwen R, Fischler B, Dezsofi A, Hadzic N, Hierro L, Jahnel J, McLin V, Nobili V, Smets F, Verkade HJ, Debray D. Wilson's Disease in Children: A Position Paper by the Hepatology Committee of the European Society for Paediatric Gastroenterology, Hepatology and Nutrition. *J Pediatr Gastroenterol Nutr* 2018; **66**: 334-344 [PMID: 29341979 DOI: 10.1097/MPG.0000000000001787]

30 **Barada K**, Nemer G, ElHajj II, Touma J, Cortas N, Boustany RM, Usta J. Early and severe liver disease associated with homozygosity for an exon 7 mutation, G691R, in Wilson's disease. *Clin Genet* 2007; **72**: 264-267 [PMID: 17718866 DOI: 10.1111/j.1399-0004.2007.00853.x]

31 **Merle U**, Schaefer M, Ferenci P, Stremmel W. Clinical presentation, diagnosis and long-term outcome of Wilson's disease: a cohort study. *Gut* 2007; **56**: 115-120 [PMID: 16709660 DOI: 10.1136/gut.2005.087262]

32 **Sapuppo A**, Pavone P, Praticò AD, Ruggieri M, Bertino G, Fiumara A. Genotype-phenotype variable correlation in Wilson disease: clinical history of two sisters with the similar genotype. *BMC Med Genet* 2020; **21**: 128 [PMID: 32532207 DOI: 10.1186/s12881-020-01062-6]

33 **Riordan SM**, Williams R. The Wilson's disease gene and phenotypic diversity. *J Hepatol* 2001; **34**: 165-171 [PMID: 11211896 DOI: 10.1016/s0168-8278(00)00028-3]

34 **Takeshita Y**, Shimizu N, Yamaguchi Y, Nakazono H, Saitou M, Fujikawa Y, Aoki T. Two families with Wilson disease in which siblings showed different phenotypes. *J Hum Genet* 2002; **47**: 543-547 [PMID: 12376745 DOI: 10.1007/s100380200082]

35 **Kegley KM**, Sellers MA, Ferber MJ, Johnson MW, Joelson DW, Shrestha R. Fulminant Wilson's disease requiring liver transplantation in one monozygotic twin despite identical genetic mutation. *Am J Transplant* 2010; **10**: 1325-1329 [PMID: 20346064 DOI: 10.1111/j.1600-6143.2010.03071.x]

36 **Lee BH**, Kim JH, Lee SY, Jin HY, Kim KJ, Lee JJ, Park JY, Kim GH, Choi JH, Kim KM, Yoo HW. Distinct clinical courses according to presenting phenotypes and their correlations to ATP7B mutations in a large Wilson's disease cohort. *Liver Int* 2011; **31**: 831-839 [PMID: 21645214 DOI: 10.1111/j.1478-3231.2011.02503.x]

37 **Leggio L**, Addolorato G, Loudianos G, Abenavoli L, Gasbarrini G. Genotype-phenotype correlation of the Wilson disease ATP7B gene. *Am J Med Genet A* 2006; **140**: 933 [PMID: 16532467 DOI: 10.1002/ajmg.a.31191]

38 **Iacob R**, Iacob S, Nastase A, Vagu C, Ene AM, Constantinescu A, Anghel D, Banica C, Paslaru L, Coriu D, Dima S, Gheorghe C, Ionica E, Gheorghe L. The His1069Gln mutation in the ATP7B gene in Romanian patients with Wilson's disease referred to a tertiary gastroenterology center. *J Gastrointestin Liver Dis* 2012; **21**: 181-185 [PMID: 22720308]

39 **Scortova E.** Evaluarea expresiei genei ATP7B in liniile celulare CHO transgenice la nivelul proteinei. [Assesing the ATP7B gene expression in CHO transgenic cell lines at protein levels]. Romanian. Sanatate Publica. *Eco Si Manag Med* 2014; **1**: 54-57

40 **Folhoffer A**, Horváth A, Hegedüs D, Firneisz G, Dunkel K, Willheim C, Ferenci P, Szönyi L, Abonyi M, Lakatos PL, Szalay F. [ATP7B gene mutations in Hungarian patients with Wilson disease--case reports to illustrate the diverse clinical presentations]. *Orv Hetil* 2003; **144**: 2509-2515 [PMID: 14974157]

41 **Ferenci P**, Członkowska A, Merle U, Ferenc S, Gromadzka G, Yurdaydin C, Vogel W, Bruha R, Schmidt HT, Stremmel W. Late-onset Wilson's disease. *Gastroenterology* 2007; **132**: 1294-1298 [PMID: 17433323 DOI: 10.1053/j.gastro.2007.02.057]

42 **Nazer H**, Ede RJ, Mowat AP, Williams R. Wilson's disease: clinical presentation and use of prognostic index. *Gut* 1986; **27**: 1377-1381 [PMID: 3792921 DOI: 10.1136/gut.27.11.1377]

43 **Collins CJ**, Yi F, Dayuha R, Duong P, Horslen S, Camarata M, Coskun AK, Houwen RHJ, Pop TL, Zoller H, Yoo HW, Jung SW, Weiss KH, Schilsky ML, Ferenci P, Hahn SH. Direct Measurement of ATP7B Peptides Is Highly Effective in the Diagnosis of Wilson Disease. *Gastroenterology* 2021; **160**: 2367-2382.e1 [PMID: 33640437 DOI: 10.1053/j.gastro.2021.02.052]

**Footnotes**

**Institutional review board statement:** The data analyzed in this manuscript came from research approved by the Emergency Clinical Hospital for Children Cluj-Napoca Institutional Review Board.

**Informed consent statement:** All involved subjects (or parents or legal representatives) signed a written informed consent that clinical and laboratory data might be used in further research. All details that might disclose the identity of the subjects under study were omitted or anonymized.

**Conflict-of-interest statement:** Pop TL, Grama A, Stefanescu AC, Willheim C have no conflicting interests related to the present work. Ferenci P reports personal fees from Alexion, personal fees from Univar, personal fees from Vivet Therapeutics, grants from Gilead, during the conduct of the study.

**Data sharing statement:** No additional data are available.

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**Manuscript source:** Invited manuscript

**Peer-review started:** April 30, 2021

**First decision:** June 17, 2021

**Article in press:** September 3, 2021

**Specialty type:** Gastroenterology and hepatology

**Country/Territory of origin:** Romania

**Peer-review report’s scientific quality classification**

Grade A (Excellent): A

Grade B (Very good): 0

Grade C (Good): C

Grade D (Fair): 0

Grade E (Poor): 0

**P-Reviewer:** Rodrigues AT, Sessa A **S-Editor:** Fan JR **L-Editor:** A **P-Editor:** Guo X

**Table 1 Clinical and laboratory characteristics in Wilson’s disease children with acute liver failure and hemolytic anemia**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Parameter** | | **All (*n* = 51)** | **ALF and hemolytic anemia (8 pts)** | **Other clinical forms (43 pts)** | ***P* value** |
| *Females, n (%)* | | 26 (50.98) | 8 (100) | 18 (41.86) | 0.00252 |
| *Mean age (yr)* | | 12.36 ± 3.74 | 14.59 ± 2.21 | 11.59 ± 3.79 | 0.03598 |
| *ALT (IU/L)* | | 131.54 ± 119.69 | 87.38 ±154.07 | 142.24 ± 110. 04 | 0.24969 |
| *AST (IU/L)* | | 125.05 ± 88.73 | 78.62 ± 47.44 | 136.30 ± 93.16 | 0.09950 |
| *TB (mg/dL)* | | 5.41 ± 12.93 | 22.55 ± 21.95 | 0.99 ± 0.95 | 0.000002 |
| *DB (mg/dL)* | | 4.20 ± 11.56 | 18.84 ± 20.40 | 0.42 ± 0.51 | 0.000007 |
| *GGT (IU/L)* | | 83.54 ± 43.44 | 97.71 ± 57.14 | 80.44 ± 40.33 | 0.34731 |
| *WBC (mm3)* | | 8568 ± 7360 | 14838 ± 14185 | 6666 ± 1799 | 0.00415 |
| *Hb (g/dL)* | | 12.15 ±2.96 | 7.4 ± 2.39 | 13.51 ± 1.22 | 0.000000 |
| *PLT (mm3)* | | 253306 ±125421 | 164875 ± 74043 | 278571 ± 126455 | 0.02142 |
| *INR* | | 2.94 ± 6.37 | 7.53 ± 11.34 | 1.18 ± 0.27 | 0.01359 |
| *KF ring, n (%)* | | 9 (17.65) | 3 (37.50) | 6 (13.95) | 0.10868 |
| *Ceruloplasmin (mg/dL)* | | 9.81 ± 6.118 | 7.98 ± 5.77 | 10.11 ± 6.19 | 0.39976 |
| *Urinary copper (μg/24 h)* | | 648.94 ± 1093.90 | 2 236.33 ± 2 174.46 | 384.38 ± 455.04 | 0.000006 |
| *Outcome, n (%)* | |  |  |  | 0.00121 |
|  | Survivors | 47 (92.16) | 5 (62.50) | 42 (97.67) |  |
|  | Transplanted | 2 (3.92) | 2 (25.00) | 0 |  |
|  | Deceased | 2 (3.92) | 1 (12.50) | 1 (2.33) |  |

ALF: Acute liver failure; ALT: Alanine-aminotransferase; AST: Aspartate-aminotransferase; TB: Total bilirubin; DB: Direct bilirubin; GGT: Gamma-glutamyl transferase; WBC: White blood cells; Hb: Hemoglobin; PLT: Platelets; INR: International normalized ratio; KF ring: Kayser-Fleisher ring.

**Table 2 Variants of *ATP7B* gene in children with hemolytic anemia and acute liver failure**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Variants** |  | **Hemolytic Anemia + ALF (8 patients)** | **Other clinical forms(44 patients)** | ***P* value** |
| *p.Trp939Cys (c.2817G>T)* | 5 homozygotes | 6 (42.86%) | 4 (4.55%) | 0.0000 |
| *p.Lys844Ter (c.2530A>T)* | 1 homozygotes | 4 (28.57%) | 1 (1.14%) | 0.0000 |
| 3 heterozygotes |
| *p.Gly1341Asp (c.4021G>A)* | 10 homozygotes | 4 (28.57%) | 23 (26.14%) | 0.8482 |
| 7 heterozygotes |
| *p.His1069Gln (c.3207A>G)* | 12 homozygotes | 0 | 39 (44.32%) | 0.0015 |
| 15 heterozygotes |
| *Other variants* | 1 homozygotes | 2 (14.29%) | 19 (21.59%) | 0.5304 |
| 19 heterozygotes |
| *Total alleles* |  | 14 (100%) | 88 (100%) |  |

ALF: Acute liver failure.

**Table 3 Data regarding the patients evaluated for the HSD17B13:TA variant**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
|  | **All** | **HSD17B13 genotype** | | |
| **T/T** | **TA/T** | **TA/TA** |
| *n* (%) | 33 | 19 (57.58) | 12 (36.36) | 2 (6.06) |
| Females, *n* (%) | 16 | 11 | 5 | 0 |
| Mean age (yr) | 11.67 ± 3.12 | 11.09 ± 3.45 | 12.14 ± 2.25 | 14.70 ± 3.24 |
| **Clinical presentation, *n* (%)** | | | | |
| ALF and/or hemolytic anemia | 7 (21.21) | 5 (71.43) | 2 (28.57) | 0 |
| Less severe hepatic forms | 26 (78.79) | 14 (53.85) | 10 (38.46) | 2 (7.69) |
| **ATP7B variant** | | | | |
| p.His1069Gln (c.3206A>G)/p.His1069Gln (c.3206A>G) | 5 | 2 | 3 | 0 |
| p.His1069Gln (c.3206A>G)/p.Gly1341Asp (c.4021G>A) | 6 | 3 | 3 | 0 |
| p.His1069Gln (c.3206A>G)/other | 7 | 4 | 3 | 0 |
| p.Gly1341Asp (c.4021G>A)/p.Gly1341Asp (c.4021G>A) | 7 | 3 | 2 | 2 |
| p.Trp939Cys (c.2817G>T)/p.Trp939Cys (c.2817G>T) | 4 | 4 | 0 | 0 |
| p.Trp939Cys (c.2817G>T)/other | 1 | 1 | 0 | 0 |
| p.Lys844Ter (c.2530A>T)/p.Lys844Ter (c.2530A>T) | 1 | 1 | 0 | 0 |
| p.Lys844Ter (c.2530A>T)/ other | 2 | 1 | 1 | 0 |

ALF: Acute liver failure.



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