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**Purinergic receptors and nucleotide processing ectoenzymes: Their roles in regulating mesenchymal stem cell functions**

Scarfì S. Purinergic receptors and nucleotide ectoenzymes in MSCs

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**Abstract**

Human mesenchymal stem cells (MSCs) are a rare population of non-hematopoietic stem cells with multilineage potential, originally identified in the bone marrow. Due to the lack of a single specific marker, MSCs can be recognized and isolated by a series of features such as plastic adherence, a panel of surface markers, the clonogenic and the differentiation abilities. The recognized role of MSCs in the regulation of hemopoiesis, in cell-degeneration protection and in the homeostasis of mesodermal tissues through their differentiation properties, justifies the current interest in identifying the biochemical signals produced by MSCs and their active crosstalk in tissue environments. Only recently extracellular nucleotides (eNTPs) and their metabolites have been included among the molecular signals produced by MSCs. These molecules are active on both ionotropic and metabotropic receptors present in most cell types. MSCs possess on their plasma membrane a significant display of these receptors and of nucleotide processing ectoenzymes. Thus, from their niche MSCs give a significant contribution to the complex signaling network of eNTPs and its derivatives. Recent studies have demonstrated the multifaceted aspects of eNTP metabolism and their signal transduction in MSCs and revealed important roles in specifying differentiation lineages and modulating MSC physiology and communication with other cells. This review discusses the roles of eNTPs, their receptors and ectoenzymes and the relevance of the signaling network and MSC functions, and also focuses on the importance of this emerging area of interest for future MSC-based cell therapies.

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**Key words:** Mesenchymal stem cell; Purinergic receptors; Ectoenzymes; ATP; -NAD; Adenosine; cADPR

**Core tip:** The multifaceted aspects of extracellular nucleotide metabolism (mainly ATP and -NAD) on mesenchymal stem cell (MSC) surface has been addressed by basic researchers only recently, sometimes revealing unexpected pivotal roles for these molecules in specifying differentiation lineages and modulating MSC physiology and communication with other cells. This review discusses the roles of extracellular nucleotides, their receptors and ectoenzymes and the relevance of their signaling network and MSC functions, and also focuses on the importance of this emerging area of interest for future MSC-based cell therapies.

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**INTRODUCTION**

Human mesenchymal stem cells (MSCs, also known as marrow stromal cells) are a rare population of non-hematopoietic stem cells with multilineage potential originally identified in the bone marrow (BM)[1,2]. BM-derived MSCs (BM-MSCs) are still considered as the gold standard for MSC applications, nevertheless the BM has several limitations as a source of MSCs, such as low frequency in this compartment, a painful isolation procedure and the loss of differentiation potential with donor's increasing age. Thus, there is growing interest in identifying alternative sources for MSCs. To this end MSCs obtained from the adipose tissue[3], dental pulp[4], placenta and Wharton's jelly[5] have gained much attention in recent times, since they can be easily isolated without any ethical concern from tissues which would be otherwise discarded.

Due to the lack of a single specific marker, MSCs can be recognized and isolated by a series of features such as plastic adherence, a panel of surface markers, the clonogenic and differentiation abilities[2,6,7]. They can be expanded in vitro for several passages without losing their lineage properties and are commonly considered the precursors of mesodermal cell types such as osteocytes, adipocytes and chondrocytes. It is still under debate, if MSCs can differentiate to non-mesodermal cell types such as hepatocytes or neurons[8-10].

In the BM MSCs play a key role in providing hemopoietic progenitors (HPs) with soluble factors essential to their proliferation and differentiation[11]. Furthermore, MSCs possess immunoregulatory functions[12]. Actually, a number of clinical trials are currently exploring the use of MSCs in cell-based therapies of various pathological conditions, such as graft *vs* host disease, renal, neurological and cardiovascular diseases[13,14]. The clinical benefit of MSC-based cell therapy seems mostly related to MSC-derived soluble factors possessing immunomodulating, growth-supporting and/or antiapoptotic activities, as demonstrated on animal models[12]. Furthermore, their differentiation and tissue regeneration potential have already been used in therapeutic clinical approaches involving tissue engineering and gene therapy[15,16].

*In vitro* differentiation of MSCs requires the activation of specific transcription factors, regulatory genes and signal cascades[17,18]. Adipogenesis induction gives rise to preadipocytes with cytoplasmic accumulation of lipid droplets and release of adipokines and extracellular matrix-associated proteins[19]. On the other hand, osteogenesis-induced osteoblasts secrete mineralized extracellular matrix, with high levels of calcium phosphate forming hydroxyapatite crystals[20]. Since both osteoblasts and adipocytes originate from a common MSC precursor, it seems obvious that osteoblast and adipocyte differentiation pathways are regulated jointly[21].

Although a plethora of studies[22-24] have shown that many substances, as well as mechanical agents, are causally related to these differentiation processes, the mechanisms involved are not yet completely defined. However, a large body of evidence supports the idea that there is an inverse relationship between the differentiation of MSCs to osteoblasts or to adipocytes, *i.e.*, conditions favoring the differentiation towards one lineage impair the differentiation to the other lineage. This seems to occur for instance during attainment of peak bone mass[25-26], when adipogenesis in the BM is inhibited favoring osteogenesis, or in aging population[27], when in the BM adipocytes are predominant respect to other cells of mesodermal origin.

MSCs regulate their fate through the complex integration of autocrine and paracrine extracellular signals (*i.e.*, hormones, cytokines, nucleotides, xenobiotics) enabling the cells to sense the external milieu and to establish a fine communication with the surrounding cell population. Hence, they calibrate their response (differentiation, immunomodulation, proliferation, migration) on the basis of the necessities of the tissue in which they reside or on the organism physio-pathological conditions.

From an evolutionary point of view, nucleotides are considered among the most ancient molecules with biological activity, and in fact they are used by living organisms for many different purposes: energy metabolism, storage of genetic information, signal transduction and extracellular communication. Nucleotides can be released or leaked into the extracellular milieu by virtually every cell in the body. Extracellular nucleotides (eNTPs) comprise both extracellular purines (ATP, ADP, -NAD, ADPR and cADPR) and extracellular pyrimidines (UTP and UDP). Once outside the cell, they either serve as signaling molecules by binding specific P2 purinergic receptors (P2X or P2Y) or are converted into other active nucleotides[28] and finally degraded to the related nucleosides. Nucleosides, mainly adenosine, can then bind different types of P1 purinergic receptors[29]. Nucleotide extracellular metabolism is mediated by special proteins located on the outer surface of the plasma membrane that possess an enzymatic domain in the extracellular region and called ectoenzymes[30]. Currently, there is an accumulating body of evidence indicating that the various ectoenzymes work in concert to dismantle eNTPs. Thus, in whatsoever milieu, the balance between nucleotides and nucleosides relies on the direct outflow of such molecules from transporters and channels in the plasma membrane[31-33], as well as on the activity of the specific ectoenzymes present on the cell surface.

It is now well established that eNTPs mediate intercellular communication virtually in all tissues. They are one of the most important indicators of cell stress in the pericellular environment[34] and the network of extracellular nucleotides/nucleosides serves multiple functions in a balanced and finely tuned fashion[35-37].

MSCs possess on their plasma membrane a significant display of purinergic receptors and ectoenzymes[38-40] and these cells have been reported to actively release nucleotides such as ATP and -NAD upon certain stimuli[39-42] (Figure 1). Thus, from their niche these cell types give a significant contribution to the complex network of signaling involving eNTPs and its derivatives, and accumulating literature indicates that MSC functions are also autocrinally influenced by eNTPs affecting their differentiation properties as well as their immunomodulatory activity.

Here is discussed the role of eNTPs, its receptors and converting ectoenzymes and the relevance of this signaling network in MSC functions, also focusing on the importance of this emerging area of interest for future MSC-based cell therapies.

**P1 RECEPTORS IN MSC**

Purinergic receptors (Ps) are plasma membrane receptors specific for adenosine, purine and pyrimidine nucleotides, which are expressed throughout the mammalian organism in all cell types. Upon their physiological agonist, Ps can be classified into P1 receptors, which natural ligand is adenosine, and P2 receptors, which recognized natural ligands are nucleotides (mainly ATP and UTP, see Figure 1)[29]. The adenosine receptors are G protein-coupled seven-transmembrane proteins further classified into the A1R, A2AR, A2BR, and A3R subtypes[29]. In particular the P1 signaling pathway involves cyclic adenosine monophosphate (cAMP) synthesis upon A2AR and A2BR activation, or cAMP inhibition upon A1R and A3R activation[29].

Adenosine can be directly released by cells[31,32] or generated by the dephosphorylation of adenine nucleotides, which in many tissues are dephosphorylated to AMP by the ecto-nucleoside triphosphate phosphohydrolase (CD39), AMP is then further dephosphorylated to adenosine by ecto-5’-nucleotidase (CD73)[30]. The resulting adenosine has an essential role in the attenuation of inflammation and in damaged tissue healing. Furthermore, it mediates diverse cardioprotective, neuroprotective, vasodilatatory and angiogenic responses[43-46], in many cases counteracting the ATP inflammatory/stress signal triggered by P2 purinergic receptor activation.

Several studies in the last decade have established the presence of both P1 and P2 receptor family members on MSC surface (Figure 1) trying to elucidate their role in the homeostasis and differentiation properties of this cell type both *in vitro* and *in vivo*.

Adenosine receptor presence and function on MSC surface was first evidenced by Evans and coworkers[47] demonstrating for the first time formation of extracellular adenosine by an osteoprogenitor cell line and by MSCs. In that occasion the presence of all four adenosine receptor subtypes, and especially A2bR, was ascertained demonstrating a causal role of their activation in active secretion of the inflammatory cytokine IL-6 and of the osteoclastogenesis inhibitory factor osteoprotegerin. These data indicate that adenosine production, as well as its activity through adenosine receptors, could be a potential target for pharmacological interventions in the bone for many diseases, including osteoporosis[48].

A further study[49] demonstrated that adenosine signaling affects proliferation and development of BM-MSCs. Perhaps the most significant finding of this work is the demonstration that adenosine A2AR deletion or blockade diminishes the number of colony forming unit-fibroblasts (CFU-F) in cultured BM-MSCs. Thus the authors speculated that adenosine, targeting the A2AR, could increase the proliferation of MSCs, as also reported for other cell types[50,51]. Alternatively they suggest that since A2AR stimulation has been shown to diminish apoptosis in other cell types[52,53], an increased survival of MSCs could enhance CFU-F yield from freshly isolated adult stem cells. Interestingly, they confirmed that A2AR and CD73 are coordinately regulated in MSCs as in other cell types[54], strengthening the idea of an active crosstalk in adenosine signaling between the adenosine receptor and the ectoenzymes able to generate the nucleoside in the pericellular space.

More recently both *in vitro*[55] and *in vivo*[56,57] studies have evaluated the contribution of adenosine signaling in MSC differentiation. Gharibi *et al*[55] in particular investigated the *in vitro* expression of adenosine receptor subtypes and the adenosine metabolism as they differentiated MSCs into osteoblasts or adipocytes. They found differential expression of the adenosine receptor subtypes during differentiation as well as in mature cells. Differential expression was related both to the progression of lineage specificity (A2BR dominant in osteoblast differentiation; A1R and A2AR in adipogenic differentiation) and to the maintenance of specialized features in the two lineages (A2AR essential to ALP expression in osteoblasts; A1R involved in lipogenic activity in adipocytes).

These data suggest that in case of diseases associated with an imbalance in the differentiation and function of these two lineages, useful strategies could include the targeting of the adenosine signaling pathway. These researches will be useful in preventing or treating conditions with insufficient bone or excessive adipocyte formation[25-27].

Finally, in recent reports[56,57] an essential role of adenosine signaling through A2BR in *in vivo* osteoblast differentiation and bone formation seems definitely confirmed. Both studies suggest that the pharmacological stimulation of this signaling pathway may enhance bone density and bone fracture healing in variously compromised situations such as non-healing fractures in osteoporosis[56] and osteolytic bone lesions in multiple myeloma[57]. In general, all the above-mentioned studies confirm an essential, functional role of extracellular adenosine and its signaling pathway in MSC physiology, homeostasis and intervention in bone and adipose tissue reconstitution allowing the identification of new pharmacological targets.

**P2 RECEPTORS IN MSC**

Extracellular nucleotides have been definitely recognized as autocrine/paracrine signaling molecules[58] released from cells in response to physiological and pathological stimulation, such as mechanical stress, hypoxia, inflammation, and other agonists. The mechanisms of nucleotide release comprise exocytosis, ATP-binding cassette transporters, connexin hemichannels, and voltage-dependent anion channels[33]. Many signaling roles for nucleotides have been demonstrated in several tissues, including neurotransmission[33], rhythm regulation in the myocardium[59]; gastrointestinal and liver function[60], regulation of epithelial cell responses[61]; blood flow distribution, oxygen delivery and endothelial barrier integrity[62,63]; immune responses[43,64] and activation of platelets at sites of vascular injury[65]. Besides acute signaling events, there is increasing evidence that purines and pyrimidines also have potent long-term roles in cell proliferation and growth[34], induction of apoptosis and anticancer activity [43] and atherosclerotic plaque formation[66]. These effects are mediated by extracellular stimulation of P2 purinergic receptors of which two major subfamilies, P2X and P2Y, have been described. The ionotropic P2X receptors are ligand-gated channels that gate extracellular cations in response to ATP and comprise seven receptor subtypes (P2X1-P2X7)[29]. Conversely, the metabotropic P2Y receptors are G-protein-coupled proteins that alternatively couple to Gq (P2Y1-2, P2Y4, P2Y6, and P2Y11) and therefore activate phospholipase C-β, or to Gi (P2Y12-14) that inhibit adenylyl cyclase and regulate ion channels[29]. Notably, P2Y11 receptor is dually coupled to phospholipase C and adenylyl cyclase stimulation.

P2Y receptors can be divided into: (1) adenine nucleotide-preferring receptors, mainly responding to ATP and ADP (P2Y1, P2Y11-13), (2) uracil nucleotide-preferring receptors (P2Y4 and P2Y6) responding to both UTP and UDP, (3) receptors of mixed selectivity (P2Y2), and (4) nucleotide sugar-preferring P2Y14 receptor responding to UDP-glucose and UDP-galactose[29]. Finally, the P2Y1 and P2Y11 receptors have been also described as -NAD receptors with diverse functional activities[64,67-68], in particular P2Y1 is also a receptor for ADPR, a -NAD metabolite generated by the cycling/hydrolysing activity of CD38 and BST1/CD157 ectoenzymes[36,68].

Only recently P2 receptors and the related activating nucleotides have been object of investigation in relation to MSC functions (Figure 1). In earlier reports[41,42] the spontaneous release of ATP from MSCs *via* gap junction hemichannels was assessed, in one occasion demonstrating a direct stimulation of P2Y1 receptor triggering intracellular Ca2+ oscillations[41] while in another showing the concurrent activation of P2X and P2Y receptors by ATP, resulting in a modulation of the proliferation rate at early passages of MSC cultivation[42].

Presence of the G-protein coupled P2Y2 receptor has also been recently demonstrated on rat MSCs as well as its activation by the preferred agonist UTP inducing intracellular Ca2+ oscillations or elevating Ca2+ levels depending on cell density, and suggesting that these different Ca2+ responses in MSCs may be correlated with cell cycle progression[69].

More recently, different investigations have been directed to the pleiotropic effects of P2 receptor activation by ATP focusing on MSC functionality in the hematopoietic niche and on the differentiation properties of these cells[70-73]. In a recent paper analyzing the effects of ATP on MSC functions Ferrari and collaborators[70] observed a downregulation of genes related to cell proliferation and anti-inflammatory cytokines and concurrently an upregulation of pro-inflammatory cytokines and cell migration related genes. These data confirm the *in vitro* inhibitory activity of ATP on MSC proliferation as already observed in a previous work[42] and demonstrate an *in vivo* potentiated homing capacity to the BM of ATP-pretreated MSCs that could be useful in supporting therapies for BM engraftment.

The role of ATP during MSC differentiation has also been addressed in the last years[38,71-73]. The related studies indicate that (1) a variety of metabolically active P2X (P2X3-7) and P2Y (all subtypes) receptors are detectable in MSCs (Figure ) and are up- or downregulated during adipogenic and osteogenic differentiation; in particular, P2Y4 and P2Y14 seem to be important for the onset of MSC commitment (regulated both in adipogenic and in osteogenic differentiation), P2Y1 and P2Y2 are downregulated in osteogenic differentiation while P2Y11 is significantly upregulated in adipogenic commitment[38]; (2) significant ATP release by MSCs, especially observed during shockwave treatment, is able to promote osteogenic differentiation through P2X7 receptor activation with a significant positive impact in bone healing[71]; and (3) ATP treatment modulates the expression of several genes governing adipogenic and osteogenic differentiation of MSCs which can be tuned from one lineage to the other by specific culture conditions in the presence of this nucleotide[72]. In addition, evidences from Ciciarello and coworkers[72] seem to indicate that ATP is able to promote adipogenesis through its triphosphate form while osteogenic differentiation seems to be induced by its nucleoside adenosine, as also proposed by others[55-57], resulting from ATP degradation by the CD39/CD73 system or directly released by cells. Thus, based on these findings, it is proposed that adipogenic differentiation is mainly mediated by activation of P2Y1 and P2Y4, receptors while stimulation of the adenosine receptor subtype A2BR is involved in osteogenic differentiation. In another recent investigation also P2Y13 receptor has been implicated in *in vivo* osteogenic differentiation through the study of impaired bone turnover in a P2Y13-KO mouse model[73]. In this study P2Y13 activation and consequent osteogenic induction, at the expenses of adipocyte differentiation, seems to be orchestrated by ADP stimulation and not ATP, thus complicating the picture of nucleotide involvement in the MSC differentiation process.

In their whole all these data provide new insights into the molecular regulation of MSC differentiation and demonstrate the necessity to further deepen this topic of investigation in order to better understand the pleiotropic effects of ATP and its derivatives on MSC differentiating abilities and to finally merge current, sometimes contrasting, observations.

Besides ATP and its derivatives, also the dinucleotide -NAD has been shown to activate P2 receptors (P2Y1 and P2Y11), its effects mainly investigated in cell types of the immune system and in the neuromuscular transmission[64,67-68]. Interestingly, it has been recently demonstrated that this nucleotide has a significant impact also on MSC functions[39]. In particular, -NAD can be released in the extracellular milieu upon stimuli able to open CX43 hemichannels in MSCs (*i.e.*, low extracellular calcium, shear stress, inflammatory stimuli) and this release is functional to increase MSC proliferation, migration and production of immunomodulatory cytokines without compromising the differentiation abilities of these cells. Such effects are observable in MSCs in the presence of -NAD both extracellularly added or autocrinally released and are dependent from P2Y11 activation (Figure 1). Thus, as for adenosine and its preferential receptors, also -NAD through its specific P2Y11 target, can exert a beneficial role in modulating cell protective functions relevant to MSC-based cell therapies.

**NUCLEOTIDE-DEGRADING ECTOENZYMES IN MSC**

Ectoenzymes are a family of cell surface molecules whose catalytic domain lies in the extracellular region. A subset of this family, the nucleotide-metabolizing ectoenzymes, are key components in the regulation of the extracellular balance between nucleotides and nucleosides together with equilibrative transporters and channels enabling direct outflow of these molecules[31-33].

Following the signal transduction, eNTPs need to be rapidly inactivated, mainly to adenosine which in turn has other pharmacological/counteracting properties. Nucleotide hydrolysing enzymes include the nucleoside triphosphate diphosphohydrolase (NTPDase) family[74], the nucleotide pyrophosphatase/ phosphodiesterase (NPP) family[75,76], and the ecto-5'-nucleotidase[77].

NTPDases are capable of hydrolysing a broad range of nucleoside tri and diphosphates, but not monophosphates. Namely, half of the eight different NTPDase genes (NTPDase1, 2, 3, and 8) are expressed as cell surface-located enzymes. The prototypic member of the NTPDase family is the cell activation antigen CD39 (NTPDase1)[78] which expression has been demonstrated on a variety of cells: vascular endothelial and smooth muscle cells[79], exocrine pancreas[80], dendritic cells[81], lymphocytes[82] and recently also MSCs[40] (Figure 1). On the other hand, The NPP family consists of seven related ectoenzymes possessing surprisingly broad substrate specificity capable of hydrolysing pyrophosphate and phosphodiester bonds generating for instance, AMP from ATP, or AMP and NMN (nicotinamide monophosphate) from -NAD[83]. The first three members of this family, NPP1-3 hydrolyze various nucleotides therefore being relevant in the purinergic signaling cascade[75]. In particular, human NPP1 is highly expressed in bone and cartilage, and less in other organs and tissues[75]. In bone tissue, NPP1 acts as a PPi-generating ectoenzyme ensuring normal bone matrix mineralization and soft tissue calcification[84]. In MSCs the presence and enzymatic activity of NPP1 and NPP3 has been recently demonstrated[39] (Figure 1), once more attesting the existence of an active and complex extracellular nucleotide metabolism in these cells.

Extracellular AMP, generated either from ATP or from -NAD degradation, can be further metabolized by the ecto-5'-nucleotidase CD73 releasing adenosine[77]. CD73 is expressed to a variable extent in different tissues, with abundant expression in the colon, kidney, brain, liver, heart, lung and large vessel endothelium[77,85,86]. Notably, CD73 is coexpressed with CD39 on the surface of CD4+ Treg cells being an important constituent of the suppressive machinery that converts ATP to the anti-inflammatory mediator adenosine with subsequent inhibition of T cell proliferation and cytokine secretion[82]. Interestingly, this situation closely resembles that of MSCs which immunomodulatory activity also has been recently related to the CD39/CD73 enzymatic axis actively producing extracellular adenosine with paracrine/immunosuppressive effects also in these cells[40] (Figure 1). These data may indicate a key role of adenosine in switching the stem cell properties of MSCs towards an immunomodulatory/pro-healing phenotype which in so many occasions has demonstrated its utility[14], suggesting a possible pharmacological use of adenosine in potentiating these features in cell-based therapies.

Although CD73 is one major cell surface marker defining MSCs according to the International Society for Cellular Therapy (ISCT), it is surprising how little is known about the enzymatic function of CD73 in these cells[87]. Notably, CD73 expression is regulated by Wnt-β-catenin signaling, one of the major pathways in stem cell and bone homeostasis[88]. Recently CD73 has been reported to be involved in osteogenic differentiation where loss of this ectoenzyme causes a lower bone mineral content in mouse trabecular bone with decreased osteocalcin serum levels and reduced expression of osteogenic mRNA markers[89]. Little is known about the role of CD73 in chondrogenesis, except that CD73 is downregulated during differentiation[90,91]. In a recent investigation further insights into CD73 relation to osteogenic/chondrogenic differentiation have been added to the literature[92] using an *in vitro* model of MSCs differentiated after cyclic-compressive loading. In these conditions Ode *et al*[92] observed increased chondrogenic differentiation accompanied by a decreased CD73 expression; besides, inhibition of CD73 enzymatic activity further increased chondrogenic matrix deposition. In contrast, in the same experimental setting, but in conditions of osteogenic induction and in the presence of a CD73 inhibitor, MSCs showed a reduction of osteogenic marker expression and of mineral matrix deposition suggesting that CD73 and its metabolite adenosine, as well as P1 receptors, belong to alternative differentiation pathways in MSCs whose expression enhance (osteogenic) or inhibit (chondrogenic) specific cell lineages. So far, and to our knowledge, no investigations have been undertaken to test the role of CD73 as an ectoenzyme during adipocyte differentiation in MSCs. Since it is known that this protein is expressed on mature adipocytes and that CD73-derived adenosine is functionally involved in body fat homeostasis, mainly inhibiting lipolysis[93], it is highly probable that this topic will be eventually addressed in the near future, hopefully adding new bricks to the comprehension of adipose tissue formation mechanism and complex homeostasis.

Another well-known class of ectoenzymes are -NAD-consuming surface proteins, primarily represented by the CD38-BST1 system[36]. The CD38 gene codes for a type II transmembrane protein distributed in a broad range of cell types[36]. The other member of the family is BST1/CD157, which differs in structure and tissue distribution[36]. The dual cycling/hydrolysing metabolism of -NAD by CD38 leads to the generation of potent intracellular Ca2+ mobilizing compounds, including cADPR (from cycling activity) and ADPR (from both cycling and hydrolysing activities)[94].

It has been recently demonstrated that MSCs show both a significant -NAD release from CX43 hemichannels and an active extracellular metabolism of this dinucleotide due not only to NPP1/3 and CD73 degradation to adenosine but also to CD38-BST1 secondary metabolite production[39] (Figure 1). The release of -NAD in the BM milieu from MSCs is essential not only for autocrine physiological, as well as immunomodulatory functions[39], but also for HP proliferation and stem cell niche maintenance[95-97]. Thus, the bilateral nucleotide network generated upon -NAD release from MSCs in the BM comprises the following enzymatic steps and functional effects: (1) -NAD released in the BM milieu directly stimulates MSC and HP functions through the purinergic receptor P2Y11[39,98], (2) extracellular -NAD can be substrate of various ectoenzymes present either on MSCs, possessing both NPP-CD73 and CD38-BST1 ectoenzymes, or on HP displaying the CD38 activity[39,99-101], (3) these enzymatic activities are able to release secondary metabolites in the BM milieu, namely adenosine, ADPR and cADPR, which again can exert autocrine and paracrine regulatory effects on MSCs and HPs[28,39,99-102]. Indeed, nanomolar/low micromolar concentrations of cADPR, such as those produced by variously stimulated CD38-BST1 positive BM cells[99,100], significantly increase the *in vitro*[99-102] and *in vivo*[96,103] proliferation and engraftment of human HPs and MSCs indicating a relevant role for this network of nucleotide-responding and nucleotide-metabolizing proteins in the BM.

## CONCLUSION

The increasingly recognized role of MSCs in the homeostasis of mesodermal tissues through their proliferation/differentiation properties, and in the regulation of hemopoiesis and cell-degeneration protection through the production of paracrine signals, justifies the current interest in identifying the biochemical signals produced by MSCs and their active crosstalk in tissue environments. Only recently such signals have been shown to belong also to the network of eNTPs and their metabolites produced by specialized ectoenzymes[39,40,87,89-92,99] and active on both ionotropic[41,71] and metabotropic receptors [38,39,42,69,70,72,73] in MSCs (Figure 1). Researchers have just began to uncover the multifaceted aspects of eNTP network on MSCs, sometimes revealing unexpected pivotal roles for these molecules and their derivatives in specifying differentiation lineages and in modulating MSC physiology and signaling towards other cells.

Thus, while extracellular -NAD and cADPR signalings seem more related to MSC homeostasis/proliferation and to the maintenance of an optimal stem cell niche for the harmonious growth of HPs and MSCs in the BM[39,95-97,99-103], ATP and adenosine demonstrate more pleiotropic roles affecting both the immunomodulatory properties of these cells as well as their lineage commitment. In particular, the nucleotide has been more frequently associated to inhibition of proliferation[42,69], pro-inflammatory and cell migration properties[70], as well as to an enhancement of both adipogenic and osteogenic differentiation[38,71-73] in MSCs. Conversely, adenosine has been associated to an autocrine protective[49], as well as a paracrine immunosuppressive[40] activity counteracting ATP stimulation. Furthermore, in MSCs, adenosine seems to have a significant role in alternative lineage specification by concomitant promotion of bone formation[55-57,72,90-92] and inhibition of cartilage production[92]. In agreement with this, it has been suggested that the positive effect of ATP on osteocyte differentiation could be just a consequence of adenosine production on MSCs through surface activity of degrading ectoenzymes[72].

The prosecution of these studies, which on the basis of what discovered until now and summarized in this review, seems to be essential for a thorough comprehension of MSC physiology, in the future will enable researchers to define precisely the involvement of these cells in tissue repair and to finally address the current clinical issues related to their use in cell-based therapies.

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**Figure 1 Surface network of purinergic receptors and nucleotide ectoenzymes on mesenchymal stem cells.** On the basis of the recent findings the Figure shows all the purinergic receptors and ectoenzymes which presence has been ascertained on mesenchymal stem cells through qPCR analyses and/or demonstration of a clear physiological function (see text for references). Furthermore, both ATP and -NAD stimulation mechanisms and metabolisms are summarized as an example of the finely tuned extracellular balance between nucleotides and nucleosides and their pleiotropic effects. CX43 HC： CX43 hemichannels; Ade: Adenosine; NMP: Nicotinamide monophosphate; ER: Endoplasmic reticulum; RyR1,3: ryanodine receptors 1 and 3.

