

WJG 20<sup>th</sup> Anniversary Special Issues (5): Colorectal cancer

## Strategies to overcome resistance to epidermal growth factor receptor monoclonal antibody therapy in metastatic colorectal cancer

Woo-Jeong Jeong, Pu-Hyeon Cha, Kang-Yell Choi

Woo-Jeong Jeong, Pu-Hyeon Cha, Kang-Yell Choi, Translational Research Center for Protein Function Control, Department of Biotechnology, College of Life Science and Biotechnology, Yonsei University, Seoul 120-749, South Korea

**Author contributions:** Jeong WJ and Cha PH wrote the manuscript draft and edited the final version of the article; Choi KY revised it critically for important intellectual content and approved the final version.

**Correspondence to:** Kang-Yell Choi, Professor, Translational Research Center for Protein Function Control, Department of Biotechnology, College of Life Science and Biotechnology, Yonsei University, 134 Shinchon-Dong, Seodemun-Gu, Seoul 120-749, South Korea. [kychoi@yonsei.ac.kr](mailto:kychoi@yonsei.ac.kr)

Telephone: +82-2-21236592 Fax: +82-2-3627265

Received: November 1, 2013 Revised: February 14, 2014

Accepted: April 15, 2014

Published online: August 7, 2014

### Abstract

Administration of monoclonal antibodies (mAbs) against epidermal growth factor receptor (EGFR) such as cetuximab and panitumumab in combination with conventional chemotherapy substantially prolongs survival of patients with metastatic colorectal cancer (mCRC). However, the efficacy of these mAbs is limited due to genetic variation among patients, in particular *K-ras* mutations. The discovery of *K-ras* mutation as a predictor of non-responsiveness to EGFR mAb therapy has caused a major change in the treatment of mCRC. Drugs that inhibit transformation caused by oncogenic alterations of Ras and its downstream components such as BRAF, MEK and AKT seem to be promising cancer therapeutics as single agents or when given with EGFR inhibitors. Although multiple therapeutic strategies to overcome EGFR mAb-resistance are under investigation, our understanding of their mode of action is limited. Rational drug development based on stringent preclinical data, biomarker validation, and

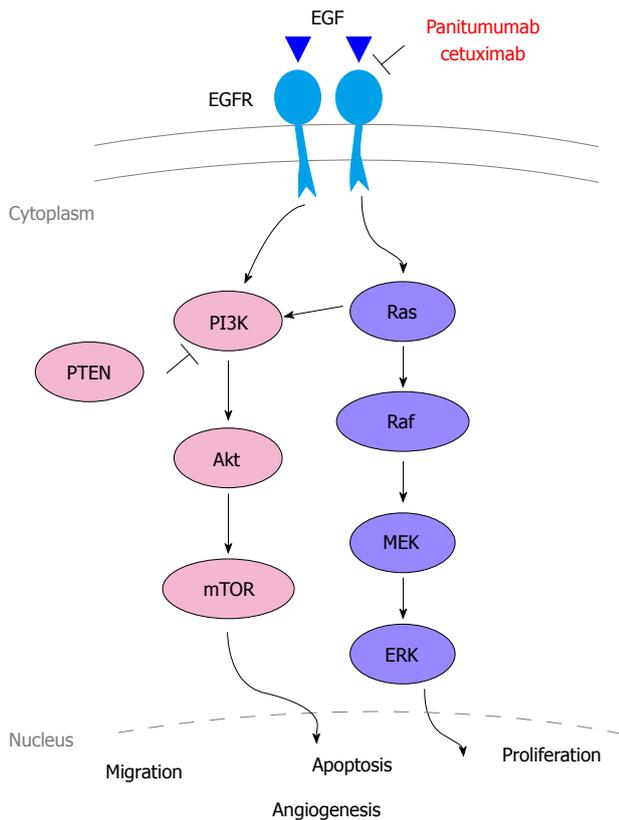
proper selection of patients is of paramount importance in the treatment of mCRC. In this review, we will discuss diverse approaches to overcome the problem of resistance to existing anti-EGFR therapies and potential future directions for cancer therapies related to the mutational status of genes associated with EGFR-Ras-ERK and PI3K signalings.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

**Key words:** Colorectal cancer; Epidermal growth factor receptor; Resistance; *K-ras* mutation; Combinational therapy

**Core tip:** Personalized treatment of patients with metastatic colorectal cancer (mCRC) based on genetic profiling of individual tumors is considered the future direction of cancer therapy. The important discovery that mutation of the *K-ras* gene is a predictor of resistance to epidermal growth factor receptor (EGFR) monoclonal antibodies is only the first of a series of genetic predictors and an increasing number of molecular alterations have since been hypothesized to play a role in resistance to anti-EGFR drugs in CRC, including activating mutations in B-Raf and PIK3CA, and loss of expression of PTEN. A comprehensive molecular characterization of mCRC and a better understanding of the functional interactions within the RTK-activated intracellular pathway will be necessary in order to select the most appropriate therapy for each individual patient.

Jeong WJ, Cha PH, Choi KY. Strategies to overcome resistance to epidermal growth factor receptor monoclonal antibody therapy in metastatic colorectal cancer. *World J Gastroenterol* 2014; 20(29): 9862-9871 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v20/i29/9862.htm> DOI: <http://dx.doi.org/10.3748/wjg.v20.i29.9862>



**Figure 1 Epidermal growth factor receptor and its downstream signaling in colorectal cancer.** Binding of ligands such as epidermal growth factor (EGF) to EGF receptor (EGFR) activates downstream Ras/ERK and PI3K/Akt pathways and regulates various physiological processes. EGFR administration of monoclonal antibodies (mAbs) (cetuximab and panitumumab) block activation of these pathways. Mutations of downstream molecules such as Ras, PI3K or Raf are associated with resistance to EGFR mAbs in patients with metastatic colorectal cancer (mCRC).

## INTRODUCTION

Colorectal cancer (CRC) is the third most frequently diagnosed type of cancer and the leading cause of cancer-related deaths worldwide<sup>[1,2]</sup>. CRC is highly treatable when diagnosed and surgically removed at an early stage; however, 5-year survival is less than 10% in patients with unresectable metastasis<sup>[3,4]</sup>. Approximately 40%-50% of CRC patients develop metastatic cancer and 80%-90% of these have unresectable metastases<sup>[5]</sup>. Chemotherapy is usually suggested for the treatment of metastatic CRC (mCRC), because surgery is limited to patients who have no metastasis outside of the liver or those who would have an appropriate amount of liver left after the surgery<sup>[4]</sup>. Conventional chemotherapy such as 5-fluorouracil (5-FU)/leucovorin (LV), irinotecan, or oxaliplatin is still mainly used as treatment for patients with mCRC<sup>[6]</sup>. Moreover, combinational therapy of oxaliplatin or irinotecan with 5-FU/LV has considerably improved the therapeutic outcome of this group of patients<sup>[7-10]</sup>. However, these chemotherapeutic agents have various adverse effects such as hair loss, nausea and vomiting<sup>[11]</sup> because they interfere with the division or reproduction of rapidly growing normal cells such as bone marrow cells in addi-

tion to their desired effect on cancer cells.

The recent development of targeted or biological therapeutics represents a substantial advance in treatment for mCRC. Although the efficacy of these targeted therapeutics is restricted to certain individuals because the drugs work on specific target proteins, these approaches have critically improved the survival of patients with metastases. When used appropriately to treat patients according to their molecular profiles, targeted therapeutics significantly prolongs overall survival and disease-free survival. Moreover, these treatments showed fewer adverse effects such as hair loss and nausea than conventional chemotherapy. Most of the targeted therapeutic agents currently in development or in clinical usage are molecules with high affinity for growth factor receptors, such as epidermal growth factor receptor (EGFR)<sup>[4]</sup>.

The recent introduction of monoclonal antibody (mAb) drugs targeting EGFR such as cetuximab (Erbix; ImClone, Branchburg, United States) and panitumumab (ABX-EGF; Amgen, Thousand Oaks, United States), into combination chemotherapy regimens with currently used drugs for the treatment of mCRC patients has been shown to be effective and has widened treatment options. However, the efficacy of these two mAbs is limited by the unresponsiveness of patients harboring a *K-ras* mutation<sup>[12]</sup>. Here, we review the mechanisms underlying resistance to EGFR mAb therapies due to *K-ras* mutations and discuss the current status of drug development strategies to overcome the problem of resistance in the treatment of patients with mCRC.

## MONOCLONAL ANTIBODIES TARGETING EGFR FOR THE TREATMENT OF CRC

### EGFR

The EGFR is a receptor tyrosine kinase (RTK) belonging to the ErbB family of cell membrane receptors. Binding of ligands, such as EGF or transforming growth factor alpha (TGF $\alpha$ ) to EGFR induces dimerization and activation of the receptors. This RTK is auto-phosphorylated and induces activation of multiple downstream signaling pathways including extracellular-signal-regulated kinase (ERK) and phosphatidylinositol 3-kinase (PI3K)/protein kinase B (Akt) pathways (Figure 1). These two pathways are involved in the regulation of various cell physiological cellular processes such as proliferation, migration, apoptosis, and angiogenesis<sup>[13]</sup> (Figure 1). Therefore, dysregulation of EGFR signaling can induce malignant transformation and tumor progression through activation of downstream signaling.

EGFR is frequently overexpressed or mutated, and such changes are associated with tumor progression and poor prognosis in many types of cancers including head and neck cancers<sup>[14]</sup>. EGFR has also been shown to be highly overexpressed in 25%-82% of cases of CRC<sup>[15]</sup>. Although the clinical significance of EGFR overexpression or gene copy number in CRC is controversial, recent studies indicate that these genetic characteristics are as-

sociated with prognosis or survival of CRC patients<sup>[16-18]</sup>. Consequently, EGFR has attracted great attention in the field of anti-cancer drug development because of its presumed role in tumor growth and progression. Cetuximab and panitumumab, two monoclonal antibodies against EGFR, have recently been used in the treatment of mCRC patients and have shown effective clinical benefits in 10%-20% of patients<sup>[19-22]</sup>.

## CETUXIMAB AND PANITUMUMAB

Cetuximab and panitumumab inhibit EGFR downstream signaling pathways, such as Ras/ERK and PI3K/Akt pathways. The safety and efficacy of these two mAbs has been studied in patients with EGFR-overexpressing CRC, and both agents have shown reliable efficacy in these patients<sup>[19-22]</sup>. In a randomized controlled trial including 329 patients who received either cetuximab (400 mg/m<sup>2</sup> initial dose followed by 250 mg/m<sup>2</sup>) and/or the topoisomerase I inhibitor irinotecan<sup>[23]</sup>, patients that receive both drugs had an objective response rate of 22.9% compared with 10.8% for those receiving only cetuximab. Cetuximab is used in combination with irinotecan to treat mCRC patients who are refractory to irinotecan-based chemotherapy<sup>[24]</sup>. It is also used as a single agent for mCRC patients with intolerance to the irinotecan-based chemotherapy<sup>[24]</sup>. The efficacy of panitumumab was studied in 463 patients who received panitumumab (6 mg/kg) with the best supportive care (BSC) or only BSC<sup>[25]</sup>. Panitumumab significantly prolonged progression-free survival (PFS) of patients treated with panitumumab and BSC compared with patients who received only BSC<sup>[25]</sup>; the mean PFS in patients receiving both panitumumab and BSC was 96 d whereas that of patients receiving BSC alone was 60 d<sup>[25]</sup>. Panitumumab is approved by the United States Food and Drug Administration (FDA) for the treatment of patients with EGFR expressing mCRC after following fluoropyrimidine-, oxaliplatin- and irinotecan-containing chemotherapy<sup>[26]</sup>. This agent is also approved by the FDA as a single agent for the treatment of mCRCs<sup>[26]</sup>.

Although cetuximab and panitumumab have been shown efficacy in patients with EGFR-expressing mCRC, their benefit is restricted to only a small proportion (8%-23%) of patients because mCRC harboring a *K-ras* mutation is resistant to these mAbs. Therefore, the FDA suggested that the *K-ras* gene mutational status of mCRC patients should be evaluated prior to administration of EGFR mAbs to avoid wasting time and money<sup>[27]</sup>.

## MECHANISMS OF RESISTANCE TO EGFR-TARGETED mABS

Despite evidence of the efficacy of cetuximab in the treatment of mCRC patients, the low response rate was the proof of concept for resistance to treatment with anti-EGFR mAbs. There is a large body of evidence supporting the existence of negative predictors that identify patients who should not be treated with anti-EGFR

mAbs. The identification of additional genetic determinants of primary resistance to EGFR-targeted therapies in CRCs is important to allow prospective identification of patients who should not be treated with cetuximab, thus avoiding their exposure to ineffective and expensive therapy. Recent work has therefore focused on the analysis of oncogenic mutations in genes encoding key downstream effectors of the EGFR signaling pathway<sup>[28,29]</sup>.

### K-Ras

*K-ras*, a member of the rat sarcoma virus (*ras*) gene family of oncogenes (which includes *K-ras*, *H-ras* and *N-ras*), encodes the guanosine diphosphate (GDP)- and guanosine triphosphate (GTP)-binding protein Ras that acts as a self-inactivating intracellular signal transducer<sup>[30]</sup>. K-Ras act as an important, but not exclusive effector of EGFR<sup>[12,31]</sup>, signaling mainly but not exclusively through BRAF and the ERK axis. K-Ras can also activate PI3K through direct interaction with its catalytic subunit<sup>[32]</sup>. When K-Ras is mutated, PI3K results in constitutive activation of its downstream signaling pathway that causes the cells to become independent of EGFR signaling activation. Somatic mutations of *K-ras* occur in 30%-40% of CRCs and predominantly in codon 12 (approximately 70%-80%) and codon 13 (approximately 15%-20%) of exon 2 (Table 1). *K-ras* mutations have emerged as the major negative predictor of efficacy in patients receiving cetuximab. Strong evidence that *K-ras* mutations are associated with the lack of response to cetuximab in chemorefractory mCRC patients led the FDA and the European Medicines Agency (EMA) to restrict the use of cetuximab as monotherapy or in combination with chemotherapy, to patients with *K-ras* wild type (WT) tumors<sup>[33]</sup>. Clearly, the *K-ras* biomarker identifies CRC patients likely to benefit from anti-EGFR therapy. However, because only 20%-40% of patients with *K-ras* WT will respond to cetuximab, either in monotherapy or in combination therapy, *K-ras* status alone does not accurately predict the subset of CRC patients that will respond to EGFR mAbs.

### N-Ras

Due to the low frequency of *N-ras* mutations in mCRC (approximately 3%-5%; Table 1), mutational status of *N-ras* have not been considered as predictive biomarkers in the treatment of mCRC to be applied for the anti-EGFR mAb therapy.

Recently, in a randomized phase 3 study of panitumumab monotherapy<sup>[34]</sup> and other preliminary findings<sup>[35-40]</sup> suggest that *N-ras* mutations are associated with the resistance to cetuximab and panitumumab. In the *K-ras* WT patients, carriers of *B-Raf* and *N-ras* mutations had a significantly lower response rate than those harboring WT *B-Raf* and *N-ras*<sup>[39]</sup>. Therefore, checking the mutational status of all *ras* isotype can provide additional predictive information for the prescription of EGFR mAb therapy in mCRC<sup>[40]</sup>.

### B-Raf

BRAF is a cytoplasmic serine/threonine kinase that di-

**Table 1** Components of the epidermal growth factor receptor signaling pathway and their abnormalities in colorectal cancer

Component (gene/protein)	Defect in CRC	Frequency in CRC
EGFR/EGFR	Protein expression Mutation	25%-90% Rare
<i>K-ras</i> / <i>K-Ras</i>	Increased copy number Activating mutation (exon 2, codon 12, 13, exon 3/4, codon 61, 117, 146)	0%-50% 30%-40%
<i>N-ras</i> / <i>N-Ras</i>	Activating mutation (exon 1, codon 12, 13, exon 2, codon 61)	3%-5%
<i>B-Raf</i> / <i>BRAF</i>	Activating mutation (V600E)	10%-15%
<i>PIK3CA</i> / <i>PI3KCA</i>	Activating mutation (exons 9 and 20)	15%-18%
<i>PTEN</i> / <i>PTEN</i>	Loss of protein expression Mutation Loss of heterozygosity	13%-19%

CRC: Colorectal cancer; EGFR: Epidermal growth factor receptor.

rectly interacts with Ras<sup>[41]</sup>. Known *B-Raf* mutations are mainly located in the kinase domain, with a single substitution of glutamic acid for valine at codon 600 (V600E) accounting for 80% of all mutations although other, less frequent, activating mutations affect the same residue, including V600A, V600D, V600G, V600K, V600M and V600R<sup>[42]</sup>. The V600E amino acid substitution is thought to be responsible for the oncogenic properties of BRAF through insertion of a negatively charged amino acid in the activation segment, thus mimicking phosphorylation of the kinase and causing it to be constitutively active<sup>[42]</sup>. BRAF V600E is the most common point mutation in mCRC, and is present in approximately 10%-15% of cases (Table 1)<sup>[43]</sup>. *K-Ras* and BRAF function in the same pathway downstream of EGFR and mutations in the genes encoding these proteins are mutually exclusive, therefore one could speculate that the presence of an active mutation in one of these two molecules is sufficient to drive constitutive activation of the pathway<sup>[43]</sup>. Because RAF is an important downstream effector of Ras, targeting RAF could be an effective strategy for the treatment of *K-ras* or *B-Raf* mutated tumors. Surprisingly, vemurafenib (PLX4032), a selective BRAF inhibitor that showed pronounced efficacy in *B-Raf*-mutant melanoma patients, had only modest clinical activity in a study evaluating 19 mCRC patients with the BRAF V600E mutation, suggesting that the biology of BRAF activation in patients with mCRC is more heterogeneous than that in melanoma<sup>[44]</sup>. In other studies, no response to cetuximab or panitumumab was observed in patients with *B-Raf*-mutant mCRC<sup>[39,45]</sup>.

### PI3K/PTEN

One of the major pathways activated by EGFR is the PI3K/Akt signaling pathway. This pathway can be de-regulated by either inactivation of the phosphatase and tensin homologue (PTEN) or by activating mutations of the PI3K p110 catalytic subunit (PIK3CA). Activa-

tion of PIK3CA results in increased AKT/mTOR pathway signaling and increased cellular proliferation<sup>[46]</sup>. Class IA PI3Ks are heterodimers composed of regulatory (p85) and catalytic (p110) subunits. Mutations in the *PIK3CA* gene occur in approximately 15%-18% (Table 1) of CRCs<sup>[47,48]</sup>. More than 80% of *PIK3CA* mutations in CRCs occur in exon 9 (60%-65%) or exon 20 (20%-25%)<sup>[39]</sup>. The gain of function induced by mutation in exon-9 (helical-domain) is independent of binding to the p85 regulatory subunit but requires interaction with Ras-GTP. In contrast, mutations in exon-20 (kinase-domain) are active in the absence of Ras-GTP binding but are highly dependent on the interaction with p85<sup>[49]</sup>. *PIK3CA* mutations can be found together with *K-ras* or *B-Raf* mutations in the same tumor, and this makes it difficult to evaluate their individual role in defining sensitivity to anti-EGFR mAbs<sup>[39,50]</sup>. Patients with mutation in *K-ras* or *B-Raf* and WT *PIK3CA* do not respond to cetuximab whereas patients with WT *K-ras* and *B-Raf* but mutations in *PIK3CA* may have different sensitivity depending on the characteristics of the mutations they harbor: *PIK3CA* exon 20 mutations are associated with resistance to cetuximab whereas *PIK3CA* exon 9 variants have no significant effect on response<sup>[39]</sup>. PI3K-initiated signaling is inhibited by PTEN. Recent reports suggested that inactivation of PTEN is associated with resistance to EGFR targeting agents<sup>[51,52]</sup>. However, the role of PTEN loss in CRC is unclear and the lack of a standardized method for PTEN detection limits the possibility of using this marker in the clinical setting.

## COMBINATIONAL THERAPIES

Because of the crosstalk between many of the RTK signaling pathways, we do not expect a single gene dependency for cancer phenotypes. Moreover, cancer cells that are treated with drugs that block a single molecular target are often able to activate alternative pathways as escape mechanisms to overcome the blockade. In addition, the effectiveness of drugs varies depending on the mutational status of the relevant gene. Therefore, appropriate selection of patients for treatment with anti-EGFR drugs is a major challenge in the management of mCRC. Collectively, the data available from clinical studies suggest that these drugs are active only in a subset of patients. The most promising approaches are rational combinations of targeted treatments that include inhibitors of downstream effectors of the EGFR pathway. At the present time, several drugs that inhibit activated BRAF, MEK, PI3K, Akt and mTOR are available and evaluations of these drugs in clinical trials are actively ongoing (Table 2)<sup>[53-77]</sup>. However, a comprehensive understanding of the precise role of these potential drug targets in CRC and the oncogenic dependence of tumors on these components is still lacking<sup>[78]</sup>.

### RAS inhibition

The Ras pathway is central to many nodes of RTK signaling and because it is constitutively activated in many

**Table 2** RAF, MEK and PI3K/mTOR inhibitors presently in the status of clinical trial in colorectal cancer

Drug	Target	Manufacturer	Phase	Indication
Sorafenib	BRAF	Bayer	II	Colon cancer (combination with cetuximab)
PLX4032	BRAFV600E	Plexxikon	I	Melanoma, colon cancer
XL281	BRAF	Exelixis	I	Solid tumors
GSK1120212	MEK	GlaxoSmithKline	I	Solid tumors, Lymphoma
AZD6244	MEK	AstraZeneca	II	Colon cancer (combination with capecitabine)
AS703026	MEK	EMD Serono	I	Solid tumors
GDC-0973	MEK	Genentech	I	Solid tumors
RO5126766	MEK	Hoffman-La Roche	I	Solid tumors
TAK-733	MEK	Millenium	I	Solid tumors
RDEA119	MEK	Ardea Biosciences	I	Solid tumors
BGT226	PI3K	Novartis	I / II	Solid tumors, Her2 positive Breast Cancer
XL147	PI3K	Exelixis	I	Solid tumors, Lymphoma
XL765	PI3K	Exelixis	I	Solid tumors
BEZ235	PI3K	Novartis	I	Solid tumors
GDC-0941	PI3K	Genentech	I	Solid tumors
PX-866	PI3K	ProIX	I	Solid tumors
SF1126	PI3K	Semafore	I	Solid tumors
Everolimus	mTOR	Novartis	II	Colon cancer (combination with cetuximab)

human cancers including CRC, it can bypass the EGFR-driven signaling cascade and reduce the clinical efficacy of EGFR inhibitors. Therefore, development of new therapeutic strategies for CRC with mutant *K-ras*, is critically needed. Several different strategies have been applied to target oncogenic Ras. One of the initial strategies used was inhibition of Ras farnesylation, a post-translational modification that is required for localization of Ras to the plasma membrane. Preclinical studies of farnesyltransferase inhibitors (FTIs) in transgenic mouse models that overexpress oncogenic Ras showed potent antitumor activity<sup>[79]</sup>. However, FTIs have shown little, if any clinical activity in patients with solid tumors, probably because of alternative modification of Ras such as geranylgeranylation<sup>[80,81]</sup>. As another approach to targeting oncogenic Ras, synthetic lethal screening has been used to identify novel anticancer agents capable of selectively killing tumor cells harboring a specific mutation<sup>[82-84]</sup>. The idea of reducing Ras expression by antisense or RNA interference is promising, but successful application of this technology is currently limited by lack of efficient delivery, uptake, and gene silencing. Using high-throughput screening approaches with loss-of-function RNAi, several groups have identified proteins that, when lost, elicit a synthetic lethal response when combined with mutant Ras oncogenes but have no effect on cells with WT Ras<sup>[85-89]</sup>. Despite all these efforts, there are still no effective

therapeutic agents or regimens available in the clinic for patients with tumors associated with *K-ras* mutation.

### RAF inhibitors

RAF is an important effector that functions downstream of Ras in the ERK signaling pathway and therefore represents a potential target for the treatment of tumors with mutant *K-ras*. Although the BRAF V600E inhibitor vemurafenib (PLX4032) shows pronounced activity in patients with *B-Raf* mutated melanoma<sup>[44]</sup>, the clinical activity of vemurafenib in previously treated patients with *B-Raf*-mutated mCRC was more modest, with a response rate of only 5% (one partial response, no complete responses) among 20 patients<sup>[44]</sup>. Interestingly, resistance to therapy in *B-Raf*-mutated CRC appears to be caused by persistent activation of the EGFR signaling pathway. Recently, two research groups independently reported that blockade of BRAF causes rapid feedback activation of EGFR<sup>[90,91]</sup>, which upon phosphorylation triggers sustained activation of ERK signaling and cell proliferation through activation of Ras and CRAF. *In vitro*, inhibition of EGFR activity by cetuximab restores sensitivity to vemurafenib. Clinical trials of a combination of vemurafenib and cetuximab in metastatic *B-Raf*-mutated CRC are currently underway (Table 2)<sup>[92]</sup>. Additionally, resistance to BRAF inhibition may also develop through activation of other signaling pathways. CRC demonstrates a higher level of PI3K/Akt signaling than melanoma, and *B-Raf* mutated colorectal cells display lower sensitivity *in vitro* to vemurafenib in the presence of concomitant PTEN or PI3K mutations<sup>[93]</sup>. Although these findings confirm mutant *B-Raf* as a therapeutic target in this disease, they also show that the biology of BRAF activation is clearly more heterogeneous in CRC than in other tumor types.

### MEK inhibitors

Selective inhibitors of MEK kinases seem an attractive target for tumors that preferentially signal through the Ras-RAF-MEK-ERK pathway. Proof of concept was provided in a preclinical study with the MEK inhibitors AS703026 or AZD6244 (Table 2), which inhibited the growth of xenograft tumors formed by CRC cells with mutant *K-ras*<sup>[94]</sup>. We further investigated the effect of MEK inhibitors on cells with cetuximab resistance attributed to *K-ras* mutation using isogenic DLD-1 CRC cell lines (D-WT and D-MUT) that harbor WT or mutant *K-ras* alleles respectively, and found that the MEK inhibitors suppressed cetuximab-resistance of CRC cells that was attributed to *K-ras* mutation both *in vitro* and *in vivo*<sup>[94]</sup>. Recent studies showed that a compensatory or activating feedback loop between RAF-MEK-ERK and PI3K pathways counteracts the effect of MEK inhibition<sup>[95]</sup>. Moreover, dual inhibition with MEK and PI3K inhibitors resulted in marked inhibition of tumor cell growth. *In vitro* studies in *K-ras*-mutant CRC cell lines showed that the presence of activating mutations in PIK3CA or loss-of-function mutations in PTEN resulted in insensitivity

to MEK inhibitor<sup>[96]</sup>. These studies also showed that mutational activation of PIK3CA is not functionally equivalent to PTEN loss. Therefore, the authors concluded that PI3K pathway activation is a major resistance mechanism that impairs the efficacy of MEK inhibitors in *K-ras* mutated cancers, and it is therefore important to test whether pan-PI3K inhibitors will act synergistically with MEK inhibitors in cancers with coexisting *PTEN* and *K-ras* mutations. Together with the previous study, this provides a strong rationale for combination treatment with PI3K and MEK inhibitors. In conclusion, although MEK inhibition is theoretically an interesting approach to targeting *K-ras* activated tumors, it is very likely that MEK inhibitors will only be efficient in a subgroup of *K-ras* mutant CRCs. As we discuss in this manuscript, combination with other targeted agents is probably a more efficient approach.

### PI3K/Akt/mTOR pathway

Inhibition of PI3K could be a feasible approach to the treatment of CRC that is resistant to EGFR monoclonal antibodies because of abnormal PTEN/PI3K status. Moreover, because the oncogene *K-ras* can activate the PI3K-Akt-mTOR pathway and such activation has been suggested as a possible mechanism of resistance to MEK inhibitors, researchers are also interested in determining whether inhibitors of the PI3K-Akt-mTOR pathway are effective in the treatment of *K-ras*-mutant CRC. Zhang *et al.*<sup>[97]</sup> examined the effects of co-treatment with the mTOR inhibitor rapamycin and the MEK inhibitor PD89059 in *K-ras* mutant CRC cell lines. This combination inhibited cell proliferation with cell cycle arrest and induced apoptosis. Combinatory treatment with PI3K/Akt inhibitor and BRAF inhibitor showed synergistic growth inhibition in *B-Raf* mutated CRC cell lines that were resistant to a BRAF inhibitor<sup>[98,99]</sup>. Given the selectivity of BRAF inhibitors for the mutant form of *B-Raf*, the combination of BRAF inhibitors with PI3K/Akt pathway inhibitors is worthy of further investigation. EGFR or PI3K/Akt inhibitors combined with BRAF inhibition could be considered for individual cases of *B-Raf*-mutant CRCs with specific mechanisms of PI3K pathway activation, such as PTEN loss.

### CONCLUSION

Personalized treatment of patients with mCRC based on genetic profiling of individual tumors is considered the future direction of cancer therapy. The important discovery that mutation of the *K-ras* gene is a predictor of resistance to EGFR monoclonal antibodies has brought this approach into clinical practice as an important innovation for the treatment of mCRC. However, this is only the first of a series of genetic predictors and an increasing number of molecular alterations have since been hypothesized to play a role in resistance to anti-EGFR drugs in CRC, including activating mutations in *N-Ras*, *B-Raf* and *PIK3CA*, and loss of expression of *PTEN*. These find-

ings suggest that resistance to anti-EGFR agents involves a complex network of molecular alterations. Assessment of the effects of these alterations on the efficacy of new drugs that selectively target proteins introduces a new paradigm to clinical oncology. Because of the complexity and heterogeneity of molecular alterations in patients, the aim for the near future is the development of personalized anti-cancer drugs for the treatment of mCRC through definition of the mutation profile of key signaling genes in individual tumors. A comprehensive molecular characterization of mCRC and a better understanding of the functional interactions within the RTK-activated intracellular pathway will be necessary in order to select the most appropriate therapy for each individual patient.

### REFERENCES

- 1 **Wong A**, Ma BB. Personalizing therapy for colorectal cancer. *Clin Gastroenterol Hepatol* 2014; **12**: 139-144 [PMID: 24025538 DOI: 10.1016/j.cgh.2013.08.040]
- 2 **Hawk ET**, Levin B. Colorectal cancer prevention. *J Clin Oncol* 2005; **23**: 378-391 [PMID: 15637400 DOI: 10.1200/JCO.2005.08.097]
- 3 **Sanoff HK**, Sargent SD, Campbell ME. Survival update and prognostic factor analysis of oxaliplatin (Ox) and irinotecan (Iri) combinations for metastatic colorectal cancer (MCRC). *J Clin Oncol* 2007; **25**: 4067
- 4 **Silvestri A**, Pin E, Huijbers A, Pellicani R, Parasido EM, Pierobon M, Petricoin E, Liotta L, Belluco C. Individualized therapy for metastatic colorectal cancer. *J Intern Med* 2013; **274**: 1-24 [PMID: 23527888 DOI: 10.1111/joim.12070]
- 5 **Field K**, Lipton L. Metastatic colorectal cancer-past, progress and future. *World J Gastroenterol* 2007; **13**: 3806-3815 [PMID: 17657834]
- 6 Modulation of fluorouracil by leucovorin in patients with advanced colorectal cancer: evidence in terms of response rate. Advanced Colorectal Cancer Meta-Analysis Project. *J Clin Oncol* 1992; **10**: 896-903 [PMID: 1534121]
- 7 **de Gramont A**, Figer A, Seymour M, Homerin M, Hmissi A, Cassidy J, Boni C, Cortes-Funes H, Cervantes A, Freyer G, Papamichael D, Le Bail N, Louvet C, Hendler D, de Braud F, Wilson C, Morvan F, Bonetti A. Leucovorin and fluorouracil with or without oxaliplatin as first-line treatment in advanced colorectal cancer. *J Clin Oncol* 2000; **18**: 2938-2947 [PMID: 10944126]
- 8 **Douillard JY**, Cunningham D, Roth AD, Navarro M, James RD, Karasek P, Jandik P, Iveson T, Carmichael J, Alakl M, Gruia G, Awad L, Rougier P. Irinotecan combined with fluorouracil compared with fluorouracil alone as first-line treatment for metastatic colorectal cancer: a multicentre randomised trial. *Lancet* 2000; **355**: 1041-1047 [PMID: 10744089]
- 9 **Goldberg RM**, Sargent DJ, Morton RF, Fuchs CS, Ramathan RK, Williamson SK, Findlay BP, Pitot HC, Alberts SR. A randomized controlled trial of fluorouracil plus leucovorin, irinotecan, and oxaliplatin combinations in patients with previously untreated metastatic colorectal cancer. *J Clin Oncol* 2004; **22**: 23-30 [PMID: 14665611 DOI: 10.1200/JCO.2004.09.046]
- 10 **Grothey A**, Sargent D, Goldberg RM, Schmoll HJ. Survival of patients with advanced colorectal cancer improves with the availability of fluorouracil-leucovorin, irinotecan, and oxaliplatin in the course of treatment. *J Clin Oncol* 2004; **22**: 1209-1214 [PMID: 15051767 DOI: 10.1200/JCO.2004.11.037]
- 11 **Love RR**, Leventhal H, Easterling DV, Nerenz DR. Side effects and emotional distress during cancer chemotherapy. *Cancer* 1989; **63**: 604-612 [PMID: 2912536]

- 12 **Lièvre A**, Bachelot JB, Le Corre D, Boige V, Landi B, Emile JF, Côté JF, Tomasic G, Penna C, Ducreux M, Rougier P, Penault-Llorca F, Laurent-Puig P. KRAS mutation status is predictive of response to cetuximab therapy in colorectal cancer. *Cancer Res* 2006; **66**: 3992-3995 [PMID: 16618717 DOI: 10.1158/0008-5472.CAN-06-0191]
- 13 **Citri A**, Yarden Y. EGF-ERBB signalling: towards the systems level. *Nat Rev Mol Cell Biol* 2006; **7**: 505-516 [PMID: 16829981 DOI: 10.1038/nrm1962]
- 14 **Chang SS**, Califano J. Current status of biomarkers in head and neck cancer. *J Surg Oncol* 2008; **97**: 640-643 [PMID: 18493942 DOI: 10.1002/jso.21023]
- 15 **Spano JP**, Fagard R, Soria JC, Rixe O, Khayat D, Milano G. Epidermal growth factor receptor signaling in colorectal cancer: preclinical data and therapeutic perspectives. *Ann Oncol* 2005; **16**: 189-194 [PMID: 15668269 DOI: 10.1093/annonc/mdi057]
- 16 **Jiang Z**, Li C, Li F, Wang X. EGFR gene copy number as a prognostic marker in colorectal cancer patients treated with cetuximab or panitumumab: a systematic review and meta analysis. *PLoS One* 2013; **8**: e56205 [PMID: 23441167 DOI: 10.1371/journal.pone.0056205]
- 17 **Galizia G**, Lieto E, Ferraraccio F, De Vita F, Castellano P, Orditura M, Imperatore V, La Mura A, La Manna G, Pinto M, Catalano G, Pignatelli C, Ciardiello F. Prognostic significance of epidermal growth factor receptor expression in colon cancer patients undergoing curative surgery. *Ann Surg Oncol* 2006; **13**: 823-835 [PMID: 16614884 DOI: 10.1245/ASO.2006.05.052]
- 18 **Nicholson RI**, Gee JM, Harper ME. EGFR and cancer prognosis. *Eur J Cancer* 2001; **37** Suppl 4: S9-15 [PMID: 11597399]
- 19 **Saltz LB**, Meropol NJ, Loehrer PJ, Needle MN, Kopit J, Mayer RJ. Phase II trial of cetuximab in patients with refractory colorectal cancer that expresses the epidermal growth factor receptor. *J Clin Oncol* 2004; **22**: 1201-1208 [PMID: 14993230 DOI: 10.1200/JCO.2004.10.182]
- 20 **Cunningham D**, Humblet Y, Siena S, Khayat D, Bleiberg H, Santoro A, Bets D, Mueser M, Harstrick A, Verslype C, Chau I, Van Cutsem E. Cetuximab monotherapy and cetuximab plus irinotecan in irinotecan-refractory metastatic colorectal cancer. *N Engl J Med* 2004; **351**: 337-345 [PMID: 15269313 DOI: 10.1056/NEJMoa033025]
- 21 **Meyerhardt JA**, Mayer RJ. Systemic therapy for colorectal cancer. *N Engl J Med* 2005; **352**: 476-487 [PMID: 15689586 DOI: 10.1056/NEJMra040958]
- 22 **Van Cutsem E**, Peeters M, Siena S, Humblet Y, Hendlisz A, Neyns B, Canon JL, Van Laethem JL, Maurel J, Richardson G, Wolf M, Amado RG. Open-label phase III trial of panitumumab plus best supportive care compared with best supportive care alone in patients with chemotherapy-refractory metastatic colorectal cancer. *J Clin Oncol* 2007; **25**: 1658-1664 [PMID: 17470858 DOI: 10.1200/JCO.2006.08.1620]
- 23 **Information FI**. FDA labelling information. Available from: URL: <http://www.fda.gov/cder/foi/label/2004/125084lbl.pdf>
- 24 **Graham J**, Muhsin M, Kirkpatrick P. Cetuximab. *Nat Rev Drug Discov* 2004; **3**: 549-550 [PMID: 15272498 DOI: 10.1038/nrd1445]
- 25 FDA labelling information. Available from: URL: <http://www.fda.gov/cder/foi/label/2006/125147s0000lbl.pdf>
- 26 **Saltz L**, Easley C, Kirkpatrick P. Panitumumab. *Nat Rev Drug Discov* 2006; **5**: 987-988 [PMID: 17201026 DOI: 10.1038/nrd2204]
- 27 **August J**. Market watch: emerging companion diagnostics for cancer drugs. *Nat Rev Drug Discov* 2010; **9**: 351 [PMID: 20431558 DOI: 10.1038/nrd3173]
- 28 **Morgillo F**, Cantile F, Fasano M, Troiani T, Martinelli E, Ciardiello F. Resistance mechanisms of tumour cells to EGFR inhibitors. *Clin Transl Oncol* 2009; **11**: 270-275 [PMID: 19451059]
- 29 **Bardelli A**, Siena S. Molecular mechanisms of resistance to cetuximab and panitumumab in colorectal cancer. *J Clin Oncol* 2010; **28**: 1254-1261 [PMID: 20100961 DOI: 10.1200/JCO.2009.24.6116]
- 30 **Bos JL**. ras oncogenes in human cancer: a review. *Cancer Res* 1989; **49**: 4682-4689 [PMID: 2547513]
- 31 **Benvenuti S**, Sartore-Bianchi A, Di Nicolantonio F, Zanon C, Moroni M, Veronese S, Siena S, Bardelli A. Oncogenic activation of the RAS/RAF signaling pathway impairs the response of metastatic colorectal cancers to anti-epidermal growth factor receptor antibody therapies. *Cancer Res* 2007; **67**: 2643-2648 [PMID: 17363584 DOI: 10.1158/0008-5472.CAN-06-4158]
- 32 **Rodriguez-Viciana P**, Warne PH, Dhand R, Vanhaesebroeck B, Gout I, Fry MJ, Waterfield MD, Downward J. Phosphatidylinositol-3-OH kinase as a direct target of Ras. *Nature* 1994; **370**: 527-532 [PMID: 8052307 DOI: 10.1038/370527a0]
- 33 **Siena S**, Sartore-Bianchi A, Di Nicolantonio F, Balfour J, Bardelli A. Biomarkers predicting clinical outcome of epidermal growth factor receptor-targeted therapy in metastatic colorectal cancer. *J Natl Cancer Inst* 2009; **101**: 1308-1324 [PMID: 19738166 DOI: 10.1093/jnci/djp280]
- 34 **Peeters M**, Oliner KS, Parker A, Siena S, Van Cutsem E, Huang J, Humblet Y, Van Laethem JL, André T, Wiezorek J, Reese D, Patterson SD. Massively parallel tumor multi-gene sequencing to evaluate response to panitumumab in a randomized phase III study of metastatic colorectal cancer. *Clin Cancer Res* 2013; **19**: 1902-1912 [PMID: 23325582 DOI: 10.1158/1078-0432.CCR-12-1913]
- 35 **De Roock W**, Jonker DJ, Di Nicolantonio F, Sartore-Bianchi A, Tu D, Siena S, Lamba S, Arena S, Frattini M, Piessevaux H, Van Cutsem E, O'Callaghan CJ, Khambata-Ford S, Zalcborg JR, Simes J, Karapetis CS, Bardelli A, Tejpar S. Association of KRAS p.G13D mutation with outcome in patients with chemotherapy-refractory metastatic colorectal cancer treated with cetuximab. *JAMA* 2010; **304**: 1812-1820 [PMID: 20978259 DOI: 10.1001/jama.2010.1535]
- 36 **Peeters M**, Douillard JY, Van Cutsem E, Siena S, Zhang K, Williams R, Wiezorek J. Mutant KRAS codon 12 and 13 alleles in patients with metastatic colorectal cancer: assessment as prognostic and predictive biomarkers of response to panitumumab. *J Clin Oncol* 2013; **31**: 759-765 [PMID: 23182985 DOI: 10.1200/JCO.2012.45.1492]
- 37 **André T**, Blons H, Mabro M, Chibaudel B, Bachelot JB, Tournigand C, Bennamoun M, Artru P, Nguyen S, Ebenezer C, Aissat N, Cayre A, Penault-Llorca F, Laurent-Puig P, de Gramont A. Panitumumab combined with irinotecan for patients with KRAS wild-type metastatic colorectal cancer refractory to standard chemotherapy: a GERCOR efficacy, tolerance, and translational molecular study. *Ann Oncol* 2013; **24**: 412-419 [PMID: 23041588 DOI: 10.1093/annonc/mds465]
- 38 **Loupakis F**, Ruzzo A, Cremolini C, Vincenzi B, Salvatore L, Santini D, Masi G, Stasi I, Canestrari E, Rulli E, Floriani I, Bencardino K, Galluccio N, Catalano V, Tonini G, Magnani M, Fontanini G, Basolo F, Falcone A, Graziano F. KRAS codon 61, 146 and BRAF mutations predict resistance to cetuximab plus irinotecan in KRAS codon 12 and 13 wild-type metastatic colorectal cancer. *Br J Cancer* 2009; **101**: 715-721 [PMID: 19603018 DOI: 10.1038/sj.bjc.6605177]
- 39 **De Roock W**, Claes B, Bernasconi D, De Schutter J, Biesmans B, Fountzilas G, Kalogeris KT, Kotoula V, Papamichael D, Laurent-Puig P, Penault-Llorca F, Rougier P, Vincenzi B, Santini D, Tonini G, Cappuzzo F, Frattini M, Molinari F, Saletti P, De Dosso S, Martini M, Bardelli A, Siena S, Sartore-Bianchi A, Tabernero J, Macarulla T, Di Fiore F, Gangloff AO, Ciardiello F, Pfeiffer P, Qvortrup C, Hansen TP, Van Cutsem E, Piessevaux H, Lambrechts D, Delorenzi M, Tejpar S. Effects of KRAS, BRAF, NRAS, and PIK3CA mutations on the efficacy of cetuximab plus chemotherapy in chemotherapy-refractory metastatic colorectal cancer: a retrospective

- consortium analysis. *Lancet Oncol* 2010; **11**: 753-762 [PMID: 20619739 DOI: 10.1016/S1470-2045(10)70130-3]
- 40 **Douillard JY**, Oliner KS, Siena S, Tabernero J, Burkes R, Barugel M, Humblet Y, Bodoky G, Cunningham D, Jassem J, Rivera F, Kocákova I, Ruff P, Błasińska-Morawiec M, Šmakal M, Canon JL, Rother M, Williams R, Rong A, Wizezorek J, Sidhu R, Patterson SD. Panitumumab-FOLFOX4 treatment and RAS mutations in colorectal cancer. *N Engl J Med* 2013; **369**: 1023-1034 [PMID: 24024839 DOI: 10.1056/NEJMoa1305275]
- 41 **Sridhar SS**, Hedley D, Siu LL. Raf kinase as a target for anticancer therapeutics. *Mol Cancer Ther* 2005; **4**: 677-685 [PMID: 15827342 DOI: 10.1158/1535-7163.MCT-04-0297]
- 42 **Davies H**, Bignell GR, Cox C, Stephens P, Edkins S, Clegg S, Teague J, Woffendin H, Garnett MJ, Bottomley W, Davis N, Dicks E, Ewing R, Floyd Y, Gray K, Hall S, Hawes R, Hughes J, Kosmidou V, Menzies A, Mould C, Parker A, Stevens C, Watt S, Hooper S, Wilson R, Jayatilake H, Gusterson BA, Cooper C, Shipley J, Hargrave D, Pritchard-Jones K, Maitland N, Chenevix-Trench G, Riggins GJ, Bigner DD, Palmieri G, Cossu A, Flanagan A, Nicholson A, Ho JW, Leung SY, Yuen ST, Weber BL, Seigler HF, Darrow TL, Patterson H, Marais R, Marshall CJ, Wooster R, Stratton MR, Futreal PA. Mutations of the BRAF gene in human cancer. *Nature* 2002; **417**: 949-954 [PMID: 12068308 DOI: 10.1038/nature00766]
- 43 **Rajagopalan H**, Bardelli A, Lengauer C, Kinzler KW, Vogelstein B, Velculescu VE. Tumorigenesis: RAF/RAS oncogenes and mismatch-repair status. *Nature* 2002; **418**: 934 [PMID: 12198537 DOI: 10.1038/418934a]
- 44 **Kopetz JD**, Chan E, Hecht JR, O'Dwyer PJ, Lee RJ, Nolop KB, Saltz L. PLX4032 in metastatic colorectal cancer patients with mutant BRAF tumors. *J Clin Oncol* 2010; **28**: abstr 3534
- 45 **Laurent-Puig P**, Cayre A, Manceau G, Buc E, Bachet JB, Lecomte T, Rougier P, Lievre A, Landi B, Boige V, Ducreux M, Ychou M, Bibeau F, Bouché O, Reid J, Stone S, Penault-Llorca F. Analysis of PTEN, BRAF, and EGFR status in determining benefit from cetuximab therapy in wild-type KRAS metastatic colon cancer. *J Clin Oncol* 2009; **27**: 5924-5930 [PMID: 19884556 DOI: 10.1200/JCO.2008.21.6796]
- 46 **Vivanco I**, Sawyers CL. The phosphatidylinositol 3-Kinase AKT pathway in human cancer. *Nat Rev Cancer* 2002; **2**: 489-501 [PMID: 12094235 DOI: 10.1038/nrc839]
- 47 **Barault L**, Veyrie N, Jooste V, Lecorre D, Chapusot C, Ferraz JM, Lièvre A, Cortet M, Bouvier AM, Rat P, Roignot P, Faivre J, Laurent-Puig P, Piard F. Mutations in the RAS-MAPK, PI(3)K (phosphatidylinositol-3-OH kinase) signaling network correlate with poor survival in a population-based series of colon cancers. *Int J Cancer* 2008; **122**: 2255-2259 [PMID: 18224685 DOI: 10.1002/ijc.23388]
- 48 **Nosho K**, Kawasaki T, Ohnishi M, Suemoto Y, Kirkner GJ, Zepf D, Yan L, Longtine JA, Fuchs CS, Ogino S. PIK3CA mutation in colorectal cancer: relationship with genetic and epigenetic alterations. *Neoplasia* 2008; **10**: 534-541 [PMID: 18516290]
- 49 **Zhao L**, Vogt PK. Helical domain and kinase domain mutations in p110alpha of phosphatidylinositol 3-kinase induce gain of function by different mechanisms. *Proc Natl Acad Sci USA* 2008; **105**: 2652-2657 [PMID: 18268322 DOI: 10.1073/pnas.0712169105]
- 50 **Sartore-Bianchi A**, Di Nicolantonio F, Nichelatti M, Molinari F, De Dosso S, Saletti P, Martini M, Cipani T, Marrapese G, Mazzucchelli L, Lamba S, Veronese S, Frattini M, Bardelli A, Siena S. Multi-determinants analysis of molecular alterations for predicting clinical benefit to EGFR-targeted monoclonal antibodies in colorectal cancer. *PLoS One* 2009; **4**: e7287 [PMID: 19806185 DOI: 10.1371/journal.pone.0007287]
- 51 **Frattini M**, Saletti P, Romagnani E, Martin V, Molinari F, Ghisletta M, Camponovo A, Etienne LL, Cavalli F, Mazzucchelli L. PTEN loss of expression predicts cetuximab efficacy in metastatic colorectal cancer patients. *Br J Cancer* 2007; **97**: 1139-1145 [PMID: 17940504 DOI: 10.1038/sj.bjc.6604009]
- 52 **Perrone F**, Lampis A, Orsenigo M, Di Bartolomeo M, Gevorgyan A, Losa M, Frattini M, Riva C, Andreola S, Bajetta E, Bertario L, Leo E, Pierotti MA, Pilotti S. PI3KCA/PTEN deregulation contributes to impaired responses to cetuximab in metastatic colorectal cancer patients. *Ann Oncol* 2009; **20**: 84-90 [PMID: 18669866 DOI: 10.1093/annonc/mdn541]
- 53 **Hainsworth JD**, Waterhouse DM, Penley WC, Shipley DL, Thompson DS, Webb CD, Anthony Greco F. Sorafenib and everolimus in advanced clear cell renal carcinoma: a phase I/II trial of the SCRI Oncology Research Consortium. *Cancer Invest* 2013; **31**: 323-329 [PMID: 23614653 DOI: 10.3109/07357907.2013.789900]
- 54 **Ng K**, Tabernero J, Hwang J, Bajetta E, Sharma S, Del Prete SA, Arrowsmith ER, Ryan DP, Sedova M, Jin J, Malek K, Fuchs CS. Phase II study of everolimus in patients with metastatic colorectal adenocarcinoma previously treated with bevacizumab-, fluoropyrimidine-, oxaliplatin-, and irinotecan-based regimens. *Clin Cancer Res* 2013; **19**: 3987-3995 [PMID: 23743569 DOI: 10.1158/1078-0432.CCR-13-0027]
- 55 **Hjelmeland AB**, Lattimore KP, Fee BE, Shi Q, Wickman S, Keir ST, Hjelmeland MD, Batt D, Bigner DD, Friedman HS, Rich JN. The combination of novel low molecular weight inhibitors of RAF (LBT613) and target of rapamycin (RAD001) decreases glioma proliferation and invasion. *Mol Cancer Ther* 2007; **6**: 2449-2457 [PMID: 17766837 DOI: 10.1158/1535-7163.MCT-07-0155]
- 56 **Yamaguchi T**, Kakefuda R, Tajima N, Sowa Y, Sakai T. Antitumor activities of JTP-74057 (GSK1120212), a novel MEK1/2 inhibitor, on colorectal cancer cell lines in vitro and in vivo. *Int J Oncol* 2011; **39**: 23-31 [PMID: 21523318 DOI: 10.3892/ijo.2011.1015]
- 57 **Honda K**, Yamamoto N, Nokihara H, Tamura Y, Asahina H, Yamada Y, Suzuki S, Yamazaki N, Ogita Y, Tamura T. Phase I and pharmacokinetic/pharmacodynamic study of RO5126766, a first-in-class dual Raf/MEK inhibitor, in Japanese patients with advanced solid tumors. *Cancer Chemother Pharmacol* 2013; **72**: 577-584 [PMID: 23860959 DOI: 10.1007/s00280-013-2228-4]
- 58 **Iverson C**, Larson G, Lai C, Yeh LT, Dadson C, Weingarten P, Appleby T, Vo T, Maderna A, Vernier JM, Hamatake R, Miner JN, Quart B. RDEA119/BAY 869766: a potent, selective, allosteric inhibitor of MEK1/2 for the treatment of cancer. *Cancer Res* 2009; **69**: 6839-6847 [PMID: 19706763 DOI: 10.1158/0008-5472.CAN-09-0679]
- 59 **Wong H**, Vernillet L, Peterson A, Ware JA, Lee L, Martini JF, Yu P, Li C, Del Rosario G, Choo EF, Hoeflich KP, Shi Y, Aftab BT, Aoyama R, Lam ST, Belvin M, Prescott J. Bridging the gap between preclinical and clinical studies using pharmacokinetic-pharmacodynamic modeling: an analysis of GDC-0973, a MEK inhibitor. *Clin Cancer Res* 2012; **18**: 3090-3099 [PMID: 22496205 DOI: 10.1158/1078-0432.CCR-12-0445]
- 60 **von Euw E**, Atefi M, Attar N, Chu C, Zachariah S, Burgess BL, Mok S, Ng C, Wong DJ, Chmielowski B, Lichter DI, Koya RC, McCannel TA, Izmailova E, Ribas A. Antitumor effects of the investigational selective MEK inhibitor TAK733 against cutaneous and uveal melanoma cell lines. *Mol Cancer* 2012; **11**: 22 [PMID: 22515704 DOI: 10.1186/1476-4598-11-22]
- 61 **Markman B**, Tabernero J, Krop I, Shapiro GI, Siu L, Chen LC, Mita M, Melendez Cuero M, Stutvoet S, Birl D, Anak O, Hackl W, Baselga J. Phase I safety, pharmacokinetic, and pharmacodynamic study of the oral phosphatidylinositol-3-kinase and mTOR inhibitor BGT226 in patients with advanced solid tumors. *Ann Oncol* 2012; **23**: 2399-2408 [PMID: 22357447 DOI: 10.1093/annonc/mds011]
- 62 **Mahadevan D**, Chiorean EG, Harris WB, Von Hoff DD, Stejskal-Barnett A, Qi W, Anthony SP, Younger AE, Rens-

- vold DM, Cordova F, Shelton CF, Becker MD, Garlich JR, Durden DL, Ramanathan RK. Phase I pharmacokinetic and pharmacodynamic study of the pan-PI3K/mTORC vascular targeted pro-drug SF1126 in patients with advanced solid tumours and B-cell malignancies. *Eur J Cancer* 2012; **48**: 3319-3327 [PMID: 22921184 DOI: 10.1016/j.ejca.2012.06.027]
- 63 **Chiarini F**, Grimaldi C, Ricci F, Tazzari PL, Evangelisti C, Ognibene A, Battistelli M, Falcieri E, Melchionda F, Pession A, Pagliaro P, McCubrey JA, Martelli AM. Activity of the novel dual phosphatidylinositol 3-kinase/mammalian target of rapamycin inhibitor NVP-BEZ235 against T-cell acute lymphoblastic leukemia. *Cancer Res* 2010; **70**: 8097-8107 [PMID: 20876803 DOI: 10.1158/0008-5472.CAN-10-1814]
- 64 **Chung EJ**, Brown AP, Asano H, Mandler M, Burgan WE, Carter D, Camphausen K, Citrin D. In vitro and in vivo radiosensitization with AZD6244 (ARRY-142886), an inhibitor of mitogen-activated protein kinase/extracellular signal-regulated kinase 1/2 kinase. *Clin Cancer Res* 2009; **15**: 3050-3057 [PMID: 19366835 DOI: 10.1158/1078-0432.CCR-08-2954]
- 65 **Bennouna J**, Lang I, Valladares-Ayerbes M, Boer K, Adenis A, Escudero P, Kim TY, Pover GM, Morris CD, Douillard JY. A Phase II, open-label, randomised study to assess the efficacy and safety of the MEK1/2 inhibitor AZD6244 (ARRY-142886) versus capecitabine monotherapy in patients with colorectal cancer who have failed one or two prior chemotherapeutic regimens. *Invest New Drugs* 2011; **29**: 1021-1028 [PMID: 20127139 DOI: 10.1007/s10637-010-9392-8]
- 66 **Haass NK**, Sproesser K, Nguyen TK, Contractor R, Medina CA, Nathanson KL, Herlyn M, Smalley KS. The mitogen-activated protein/extracellular signal-regulated kinase kinase inhibitor AZD6244 (ARRY-142886) induces growth arrest in melanoma cells and tumor regression when combined with docetaxel. *Clin Cancer Res* 2008; **14**: 230-239 [PMID: 18172275 DOI: 10.1158/1078-0432.CCR-07-1440]
- 67 **Yao E**, Zhou W, Lee-Hoeflich ST, Truong T, Haverty PM, Eastham-Anderson J, Lewin-Koh N, Gunter B, Belvin M, Murray LJ, Friedman LS, Sliwkowski MX, Hoeflich KP. Suppression of HER2/HER3-mediated growth of breast cancer cells with combinations of GDC-0941 PI3K inhibitor, trastuzumab, and pertuzumab. *Clin Cancer Res* 2009; **15**: 4147-4156 [PMID: 19509167 DOI: 10.1158/1078-0432.CCR-08-2814]
- 68 **Raynaud FI**, Eccles SA, Patel S, Alix S, Box G, Chuckowree I, Folkes A, Gowan S, De Haven Brandon A, Di Stefano F, Hayes A, Henley AT, Lensun L, Pergl-Wilson G, Robson A, Saghir N, Zhyvoloup A, McDonald E, Sheldrake P, Shuttleworth S, Valenti M, Wan NC, Clarke PA, Workman P. Biological properties of potent inhibitors of class I phosphatidylinositol 3-kinases: from PI-103 through PI-540, PI-620 to the oral agent GDC-0941. *Mol Cancer Ther* 2009; **8**: 1725-1738 [PMID: 19584227 DOI: 10.1158/1535-7163.MCT-08-1200]
- 69 **Folkes AJ**, Ahmadi K, Alderton WK, Alix S, Baker SJ, Box G, Chuckowree IS, Clarke PA, Depledge P, Eccles SA, Friedman LS, Hayes A, Hancox TC, Kugendradas A, Lensun L, Moore P, Olivero AG, Pang J, Patel S, Pergl-Wilson GH, Raynaud FI, Robson A, Saghir N, Salphati L, Sohal S, Ultsch MH, Valenti M, Wallweber HJ, Wan NC, Wiesmann C, Workman P, Zhyvoloup A, Zvelebil MJ, Shuttleworth SJ. The identification of 2-(1H-indazol-4-yl)-6-(4-methanesulfonyl-piperazin-1-ylmethyl)-4-morpholin-4-yl-thieno[3,2-d]pyrimidine (GDC-0941) as a potent, selective, orally bioavailable inhibitor of class I PI3 kinase for the treatment of cancer. *J Med Chem* 2008; **51**: 5522-5532 [PMID: 18754654 DOI: 10.1021/jm800295d]
- 70 **Liu TJ**, Koul D, LaFortune T, Tiao N, Shen RJ, Maira SM, Garcia-Echeverria C, Yung WK. NVP-BEZ235, a novel dual phosphatidylinositol 3-kinase/mammalian target of rapamycin inhibitor, elicits multifaceted antitumor activities in human gliomas. *Mol Cancer Ther* 2009; **8**: 2204-2210 [PMID: 19671762 DOI: 10.1158/1535-7163.MCT-09-0160]
- 71 **Serra V**, Markman B, Scaltriti M, Eichhorn PJ, Valero V, Guzman M, Botero ML, Llonch E, Atzori F, Di Cosimo S, Maira M, Garcia-Echeverria C, Parra JL, Arribas J, Baselga J. NVP-BEZ235, a dual PI3K/mTOR inhibitor, prevents PI3K signaling and inhibits the growth of cancer cells with activating PI3K mutations. *Cancer Res* 2008; **68**: 8022-8030 [PMID: 18829560 DOI: 10.1158/0008-5472.CAN-08-1385]
- 72 **Howes AL**, Chiang GG, Lang ES, Ho CB, Powis G, Vuori K, Abraham RT. The phosphatidylinositol 3-kinase inhibitor, PX-866, is a potent inhibitor of cancer cell motility and growth in three-dimensional cultures. *Mol Cancer Ther* 2007; **6**: 2505-2514 [PMID: 17766839 DOI: 10.1158/1535-7163.MCT-06-0698]
- 73 **Ihle NT**, Williams R, Chow S, Chew W, Berggren ML, Paine-Murrieta G, Minion DJ, Halter RJ, Wipf P, Abraham R, Kirkpatrick L, Powis G. Molecular pharmacology and antitumor activity of PX-866, a novel inhibitor of phosphoinositide-3-kinase signaling. *Mol Cancer Ther* 2004; **3**: 763-772 [PMID: 15252137]
- 74 **Molckovsky A**, Siu LL. First-in-class, first-in-human phase I results of targeted agents: highlights of the 2008 American society of clinical oncology meeting. *J Hematol Oncol* 2008; **1**: 20 [PMID: 18959794 DOI: 10.1186/1756-8722-1-20]
- 75 **Sala E**, Mologni L, Truffa S, Gaetano C, Bollag GE, Gambacorti-Passerini C. BRAF silencing by short hairpin RNA or chemical blockade by PLX4032 leads to different responses in melanoma and thyroid carcinoma cells. *Mol Cancer Res* 2008; **6**: 751-759 [PMID: 18458053 DOI: 10.1158/1541-7786.MCR-07-2001]
- 76 **McCubrey JA**, Milella M, Tafuri A, Martelli AM, Lunghi P, Bonati A, Cervello M, Lee JT, Steelman LS. Targeting the Raf/MEK/ERK pathway with small-molecule inhibitors. *Curr Opin Investig Drugs* 2008; **9**: 614-630 [PMID: 18516761]
- 77 **McCubrey JA**, Steelman LS, Abrams SL, Bertrand FE, Ludwig DE, Bäsecke J, Libra M, Stivala F, Milella M, Tafuri A, Lunghi P, Bonati A, Martelli AM. Targeting survival cascades induced by activation of Ras/Raf/MEK/ERK, PI3K/PTEN/Akt/mTOR and Jak/STAT pathways for effective leukemia therapy. *Leukemia* 2008; **22**: 708-722 [PMID: 18337766 DOI: 10.1038/leu.2008.27]
- 78 **De Roock W**, De Vriendt V, Normanno N, Ciardiello F, Tejpar S. KRAS, BRAF, PIK3CA, and PTEN mutations: implications for targeted therapies in metastatic colorectal cancer. *Lancet Oncol* 2011; **12**: 594-603 [PMID: 21163703 DOI: 10.1016/S1470-2045(10)70209-6]
- 79 **Kohl NE**, Omer CA, Conner MW, Anthony NJ, Davide JP, deSolms SJ, Giuliani EA, Gomez RP, Graham SL, Hamilton K. Inhibition of farnesyltransferase induces regression of mammary and salivary carcinomas in ras transgenic mice. *Nat Med* 1995; **1**: 792-797 [PMID: 7585182]
- 80 **Adjei AA**, Mauer A, Bruzek L, Marks RS, Hillman S, Geyer S, Hanson LJ, Wright JJ, Erlichman C, Kaufmann SH, Vokes EE. Phase II study of the farnesyl transferase inhibitor R115777 in patients with advanced non-small-cell lung cancer. *J Clin Oncol* 2003; **21**: 1760-1766 [PMID: 12721252 DOI: 10.1200/JCO.2003.09.075]
- 81 **Macdonald JS**, McCoy S, Whitehead RP, Iqbal S, Wade JL, Giguere JK, Abbruzzese JL. A phase II study of farnesyl transferase inhibitor R115777 in pancreatic cancer: a Southwest oncology group (SWOG 9924) study. *Invest New Drugs* 2005; **23**: 485-487 [PMID: 16133800 DOI: 10.1007/s10637-005-2908-y]
- 82 **Dolma S**, Lessnick SL, Hahn WC, Stockwell BR. Identification of genotype-selective antitumor agents using synthetic lethal chemical screening in engineered human tumor cells. *Cancer Cell* 2003; **3**: 285-296 [PMID: 12676586]
- 83 **Guo W**, Wu S, Liu J, Fang B. Identification of a small molecule with synthetic lethality for K-ras and protein kinase C iota. *Cancer Res* 2008; **68**: 7403-7408 [PMID: 18794128 DOI: 10.1158/0008-5472.CAN-08-1449]

- 84 **Torrance CJ**, Agrawal V, Vogelstein B, Kinzler KW. Use of isogenic human cancer cells for high-throughput screening and drug discovery. *Nat Biotechnol* 2001; **19**: 940-945 [PMID: 11581659 DOI: 10.1038/nbt1001-940]
- 85 **Barbie DA**, Tamayo P, Boehm JS, Kim SY, Moody SE, Dunn IF, Schinzel AC, Sandy P, Meylan E, Scholl C, Fröhling S, Chan EM, Sos ML, Michel K, Mermel C, Silver SJ, Weir BA, Reiling JH, Sheng Q, Gupta PB, Wadlow RC, Le H, Hoersch S, Wittner BS, Ramaswamy S, Livingston DM, Sabatini DM, Meyerson M, Thomas RK, Lander ES, Mesirov JP, Root DE, Gilliland DG, Jacks T, Hahn WC. Systematic RNA interference reveals that oncogenic KRAS-driven cancers require TBK1. *Nature* 2009; **462**: 108-112 [PMID: 19847166 DOI: 10.1038/nature08460]
- 86 **Scholl C**, Fröhling S, Dunn IF, Schinzel AC, Barbie DA, Kim SY, Silver SJ, Tamayo P, Wadlow RC, Ramaswamy S, Döhner K, Bullinger L, Sandy P, Boehm JS, Root DE, Jacks T, Hahn WC, Gilliland DG. Synthetic lethal interaction between oncogenic KRAS dependency and STK33 suppression in human cancer cells. *Cell* 2009; **137**: 821-834 [PMID: 19490892 DOI: 10.1016/j.cell.2009.03.017]
- 87 **Vicent S**, Chen R, Sayles LC, Lin C, Walker RG, Gillespie AK, Subramanian A, Hinkle G, Yang X, Saif S, Root DE, Huff V, Hahn WC, Sweet-Cordero EA. Wilms tumor 1 (WT1) regulates KRAS-driven oncogenesis and senescence in mouse and human models. *J Clin Invest* 2010; **120**: 3940-3952 [PMID: 20972333 DOI: 10.1172/JCI44165]
- 88 **Wang Y**, Ngo VN, Marani M, Yang Y, Wright G, Staudt LM, Downward J. Critical role for transcriptional repressor Snail2 in transformation by oncogenic RAS in colorectal carcinoma cells. *Oncogene* 2010; **29**: 4658-4670 [PMID: 20562906 DOI: 10.1038/onc.2010.218]
- 89 **Luo J**, Emanuele MJ, Li D, Creighton CJ, Schlabach MR, Westbrook TF, Wong KK, Elledge SJ. A genome-wide RNAi screen identifies multiple synthetic lethal interactions with the Ras oncogene. *Cell* 2009; **137**: 835-848 [PMID: 19490893 DOI: 10.1016/j.cell.2009.05.006]
- 90 **Corcoran RB**, Ebi H, Turke AB, Coffee EM, Nishino M, Cogdill AP, Brown RD, Della Pelle P, Dias-Santagata D, Hung KE, Flaherty KT, Piris A, Wargo JA, Settleman J, Mino-Kenudson M, Engelman JA. EGFR-mediated reactivation of MAPK signaling contributes to insensitivity of BRAF mutant colorectal cancers to RAF inhibition with vemurafenib. *Cancer Discov* 2012; **2**: 227-235 [PMID: 22448344 DOI: 10.1158/2159-8290.CD-11-0341]
- 91 **Prahallad A**, Sun C, Huang S, Di Nicolantonio F, Salazar R, Zecchin D, Beijersbergen RL, Bardelli A, Bernards R. Unresponsiveness of colon cancer to BRAF(V600E) inhibition through feedback activation of EGFR. *Nature* 2012; **483**: 100-103 [PMID: 22281684 DOI: 10.1038/nature10868]
- 92 **Morris V**, Kopetz S. BRAF inhibitors in clinical oncology. *F1000Prime Rep* 2013; **5**: 11 [PMID: 23585929 DOI: 10.12703/P5-11]
- 93 **Mao M**, Tian F, Mariadason JM, Tsao CC, Lemos R, Dayyani F, Gopal YN, Jiang ZQ, Wistuba II, Tang XM, Bornman WG, Bollag G, Mills GB, Powis G, Desai J, Gallick GE, Davies MA, Kopetz S. Resistance to BRAF inhibition in BRAF-mutant colon cancer can be overcome with PI3K inhibition or demethylating agents. *Clin Cancer Res* 2013; **19**: 657-667 [PMID: 23251002 DOI: 10.1158/1078-0432.CCR-11-1446]
- 94 **Yoon J**, Koo KH, Choi KY. MEK1/2 inhibitors AS703026 and AZD6244 may be potential therapies for KRAS mutated colorectal cancer that is resistant to EGFR monoclonal antibody therapy. *Cancer Res* 2011; **71**: 445-453 [PMID: 21118963 DOI: 10.1158/0008-5472.CAN-10-3058]
- 95 **Mirzoeva OK**, Das D, Heiser LM, Bhattacharya S, Siwak D, Gendelman R, Bayani N, Wang NJ, Neve RM, Guan Y, Hu Z, Knight Z, Feiler HS, Gascard P, Parvin B, Spellman PT, Shokat KM, Wyrobek AJ, Bissell MJ, McCormick F, Kuo WL, Mills GB, Gray JW, Korn WM. Basal subtype and MAPK/ERK kinase (MEK)-phosphoinositide 3-kinase feedback signaling determine susceptibility of breast cancer cells to MEK inhibition. *Cancer Res* 2009; **69**: 565-572 [PMID: 19147570 DOI: 10.1158/0008-5472.CAN-08-3389]
- 96 **Wee S**, Jagani Z, Xiang KX, Loo A, Dorsch M, Yao YM, Sellers WR, Lengauer C, Stegmeier F. PI3K pathway activation mediates resistance to MEK inhibitors in KRAS mutant cancers. *Cancer Res* 2009; **69**: 4286-4293 [PMID: 19401449 DOI: 10.1158/0008-5472.CAN-08-4765]
- 97 **Zhang YJ**, Tian XQ, Sun DF, Zhao SL, Xiong H, Fang JY. Combined inhibition of MEK and mTOR signaling inhibits initiation and progression of colorectal cancer. *Cancer Invest* 2009; **27**: 273-285 [PMID: 19194827 DOI: 10.1080/07357900802314893]
- 98 **Yang H**, Higgins B, Kolinsky K, Packman K, Bradley WD, Lee RJ, Schostack K, Simcox ME, Kopetz S, Heimbros D, Lestini B, Bollag G, Su F. Antitumor activity of BRAF inhibitor vemurafenib in preclinical models of BRAF-mutant colorectal cancer. *Cancer Res* 2012; **72**: 779-789 [PMID: 22180495 DOI: 10.1158/0008-5472.CAN-11-2941]
- 99 **Oikonomou E**, Koc M, Sourkova V, Andera L, Pintzas A. Selective BRAFV600E inhibitor PLX4720, requires TRAIL assistance to overcome oncogenic PIK3CA resistance. *PLoS One* 2011; **6**: e21632 [PMID: 21738740 DOI: 10.1371/journal.pone.0021632]

P- Reviewer: Nishida T, Yu B S- Editor: Zhai HH

L- Editor: A E- Editor: Ma S





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>



ISSN 1007-9327

