

# World Journal of *Clinical Cases*

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## Retrospective Study

# Evaluation of high-risk factors and the diagnostic value of alpha-fetoprotein in the stratification of primary liver cancer

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## Abstract

### BACKGROUND

Alpha-fetoprotein (AFP) is one of the diagnostic standards for primary liver cancer (PLC); however, AFP exhibits insufficient sensitivity and specificity for diagnosing PLC.

### AIM

To evaluate the effects of high-risk factors and the diagnostic value of AFP in stratified PLC.

### METHODS

In total, 289 PLC cases from 2013 to 2019 were selected for analysis. First, the contributions of high-risk factors in stratifying PLC were compared according to the following criteria: Child-Pugh score, clinical stage of liver cirrhosis, tumor size, and Barcelona Clinic Liver Cancer (BCLC) stage. Then, the diagnostic value of AFP was evaluated in different stratifications of PLC by receiver operating characteristic curves. For PLC cases in which AFP played little role, the diagnostic values of carcinoembryonic antigen (CEA), carbohydrate antigen 19-9 (CA 19-9), gamma-glutamyl transferase (GGT), and AFP were analyzed.

### RESULTS

The roles of high-risk factors differed in stratified PLC. The incidence of smoking and drinking history was higher in PLC with Child-Pugh scores of C ( $P < 0.0167$ ).

The hepatitis B virus (HBV) infection rate in PLC with cirrhosis was more than in PLC without cirrhosis ( $P < 0.0167$ ). Small tumors were more prone to cirrhosis than large tumors ( $P < 0.005$ ). BCLC stage D PLC was more likely to be associated with HBV infection and cirrhosis ( $P < 0.0083$ ). AFP levels were higher in PLC with cirrhosis, diffuse tumors, and BCLC stage D disease. In diagnosing PLC defined as Child–Pugh A, B, and C, massive hepatoma, diffuse hepatoma, BCLC stage B, C, and D, and AFP showed significant diagnostic value [all area under the curve (AUC)  $> 0.700$ ]. However, these measures were meaningless (AUC  $< 0.600$ ) in small hepatomas and BCLC A stage PLC, but could be replaced by the combined detection of CEA, CA 19-9, GGT, and AFP (AUC = 0.810 and 0.846, respectively).

### CONCLUSION

Stratification of PLC was essential for precise diagnoses and benefited from evaluating AFP levels.

**Key Words:** Primary liver cancer; Stratification; Risk factors; Alpha-fetoprotein; Receiver operating characteristic curve; Diagnostics

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**Core Tip:** To evaluate the effects of high-risk factors and the diagnostic value of alpha-fetoprotein (AFP) in stratified primary liver cancer (PLC), 289 cases were selected for analysis. First, the contributions of high-risk factors in stratifying PLC were compared. Then, the diagnostic value of AFP was evaluated in different stratifications of PLC by receiver operating characteristic curves. For PLC cases in which AFP played little role, the diagnostic values of carcinoembryonic antigen (CEA), carbohydrate antigen 19-9, gamma-glutamyl transferase, and AFP were analyzed. It was concluded that stratification of PLC was essential for precise diagnoses and it benefited from diagnostic values of AFP.

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## INTRODUCTION

Primary liver cancer (PLC) was the second leading cause of cancer-related mortality in 2014, with its death toll accounting for 51% of total global deaths[1]. Among all liver cancer patients, the 5-year overall survival rate is only 10.1%, and cirrhosis is the primary cause of death for PLC patients[2,3]. PLC incidence rates vary across clinical etiologies and conditions such as liver disease severity; even within the same clinical entity, individual PLC risk is heterogeneous across patients for unknown reasons[4]. Hence, clinically meaningful utility must be demonstrated under specific clinical scenarios for a diagnostic modality to be adopted into regular use. This was the initial purpose of stratifying the PLC cases in this study. Most previous studies have included hepatitis B virus (HBV) and/or hepatitis C virus (HCV) infection, alcohol use, non-alcoholic fatty liver, cirrhosis, gender, and age as the high-risk factors related to PLC[5]. Of these, infection by hepatoma viruses was the primary contributing factor to liver cancer in developing countries such as Asia and Africa[6,7]. HBV promotes malignant changes in liver cells by infecting the host and integrating the genome[8,9]. In contrast, non-alcoholic fatty liver disease and other metabolic diseases are the main susceptibility factors for PLC in developed countries [10,11].

Early diagnosis is vital for expanding treatment choices and improving the prognosis and quality of life of PLC patients. Alpha-fetoprotein (AFP) is a widely used serological marker that shows increased levels 8 to 11 mo before symptoms occur; thus, it is one of the diagnostic standards in the guidelines for PLC in China and Japan[12]. However, related reports have demonstrated that AFP shows deficient sensitivity and specificity for diagnosing PLC, causing missed diagnoses and misdiagnoses[13]. Therefore, this retrospective study aimed to evaluate the role of high-risk factors in diagnosing stratified PLC cases, especially the diagnostic value of AFP.

## MATERIALS AND METHODS

### Subjects

In total, 289 untreated PLC patients who were initially diagnosed at the North China University of Science and Technology Affiliated Hospital according to the guidelines for the diagnosis and treatment of PLC in China (2017 Edition)[14] were selected as the observation group. The control group consisted of 217 untreated cases with chronic hepatitis B and 279 cases with cirrhosis. There were no differences between the two groups in terms of age ( $\chi^2 = 0.536$ ,  $P = 0.765$ ) or gender ( $F = 2.869$ ,  $P = 0.057$ ).

Clinical criteria of the observation group were determined as follows: (1) PLC tumor size classifications were based on the Expert consensus on pathological diagnosis of PLC[15]; and (2) Child-Pugh liver function scores (similar to assessments in the Japan Society of Hepatology guidelines) and Barcelona Clinic Liver Cancer (BCLC) staging standards were derived from guidelines for the diagnosis and treatment of PLC in China (2017 Edition).

The exclusion criteria for this study were: (1) Metastatic cancer; (2) prior treatment for PLC; (3) the size and number of PLC and/or metastatic lesions were unclear from imaging examinations such as magnetic resonance imaging (MRI) or computed tomography (CT); and (4) the serological tumor markers AFP, carcinoembryonic antigen (CEA), carbohydrate antigen 19-9 (CA 19-9), and gamma-glutamyl transferase (GGT) were not detected.

Clinical criteria in the control group were determined as follows: (1) Chronic hepatitis B cases were defined as chronic inflammatory diseases of the liver caused by HBV that had lasted for > 6 mo and were diagnosed according to the Guidelines for the Prevention and Treatment of Chronic Hepatitis B (2019 Version)[16]; and (2) cirrhosis was judged by the Chinese Liver Disease Diagnosis and Treatment Management Standards[17] and confirmed by imaging and/or pathological examinations.

### Data collection

**General data collection:** Relevant patient information (gender, age, smoking and drinking history, and other basic information) were collected from the electronic medical records system. Patients with a history of smoking were defined as those who had smoked more than 1 cigarette per day for more than one year in a row. Patients with a history of drinking alcohol were defined as those who consumed more than 100 mL per day for more than 1 year in a row.

The main symptoms and signs of PLC included systemic symptoms (fatigue, loss of appetite, edema, and liver disease face), digestive symptoms (bloating, diarrhea, and abdominal pain), and bleeding symptoms ranging from a bleeding tendency to anemia. For patients with PLC with cirrhosis, neuropsychiatric performance metrics were also collected including personality, communication, behavior, calculation ability, intelligence, consciousness, orientation, and whether there were flapping tremors, increased muscle tone, tendon hyperreflexia, ankle clonus or Babinski signs, and other abnormal nervous system parameters, in addition to whether ascites were present.

**Imaging data collection:** Imaging data were collected from the examination results of PLC patients' abdominal ultrasound, abdominal MRI enhancement, or abdominal CT enhancement scans, including the size and number of PLC and metastases, and the involvement of blood vessels such as the portal vein.

**Laboratory-related data collection:** Venous blood samples were collected after fasting (> 12 h) to detect serum markers including HBV antigen (HBsAg), HCV-Ag, albumin, total bilirubin, direct bilirubin, prothrombin time (PT), AFP, CEA, CA 19-9, and GGT. Among them, the liver function items were detected using a Beckman Coulter AU5800 or AU5821 automatic biochemical analyzer (Beckman Coulter, Brea, CA, United States); serum tumor markers were detected using a Roche 602 electrochemiluminescence immunoassay (Roche, Basel, Switzerland); PT was detected using a STAGO STAR Max instrument (Stago, Inc., Parsippany, NJ, United States); HBsAg and HCV-Ag were detected using a Mindray CL6000i (Mindray, Shenzhen, China). The cut-off level for AFP was 400 ng/mL in accordance with guidelines for the diagnosis and treatment of PLC in China (2017 Edition).

### Statistical analysis

All data were analyzed using SPSS 19.0 software (IBM, Armonk, NY, United States). Counting data are presented as rates, and differences among groups were compared with the  $\chi^2$  test. For pairwise comparisons between multiple sets of rates, the test standard adjustment method was used. Measurement data were first tested with the Kolmogorov-Smirnov test to check whether the measurement data of each group were normally distributed. Normally distributed data are presented as mean  $\pm$  standard deviation, and differences between two groups were analyzed with the T test and differences among multiple groups were analyzed by ANOVA. Non-normally distributed measurement data are presented as median (interquartile range), and differences between two groups were analyzed with the Mann-Whitney U test, differences among independent samples were analyzed with the Kruskal-Wallis H test, and differences between two sets of measurement data were analyzed with the Bonferroni test. The method of adjusting test standards took  $\alpha'' = \alpha/N$  as the test standard. In the formula  $N = k(k-1) \div 2$ ,  $k$  is the number of sample rates. The other methods used  $\alpha = 0.05$  as the test

standard.

**Statement:** Statistical review of the study was performed by a biomedical statistician.

## RESULTS

### ***Influence of high-risk factors on stratifying PLC cases***

PLC cases were stratified according to liver function Child–Pugh scores, clinical stage of liver cirrhosis, classification of liver cancer size, and BCLC stage. Subsequently, the influences of high-risk factors (age, gender, HBV and/or HCV infection, smoking, alcohol use, non-alcoholic fatty liver, and cirrhosis) on the stratification of PLC were compared to provide more reference. Tables 1-4 show the general data for each item with cases and rates. The main results are presented below.

When PLC cases were stratified by Child–Pugh scores, the rate of smoking and drinking history played the greatest roles in distinguishing class C ( $P = 0.013$  and  $P = 0.007$ , respectively, both  $< 0.0167$ ). When PLC cases were stratified by the clinical stage of cirrhosis, HBsAg showed the biggest prognostic difference. To be precise, compared with PLC cases without liver cirrhosis, PLC with cirrhosis had higher rates of HBV infection, which was not associated with the severity of cirrhosis ( $P < 0.001$ ). Accordingly, when PLC cases were stratified by tumor size, the data demonstrated that liver cirrhosis occurred more frequently in patients with smaller lesions compared with patients with massive liver tumors ( $P = 0.002$ ,  $< 0.005$ ). Finally, when the PLC cases were stratified by BCLC stage, both HBsAg and liver cirrhosis changed at different stages. In particular, the rate of HBV infection in BCLC stage D cases was substantially increased compared with stages A, B, and C. There was likely marked cirrhosis in stage D PLC cases. In summary, the different high-risk factors weighted distinctively when PLC was stratified by each criterion, meaning it is necessary to evaluate them in certain stratifications of PLC. Otherwise the mixed factors may confuse our judgement as to the condition of individual PLC patients.

### ***Analysis of serum AFP in PLC cases***

**The diagnostic value of serum AFP levels in PLC, chronic hepatitis, and cirrhosis:** Table 5 shows that AFP levels in the PLC group were significantly increased compared with those in the chronic hepatitis B group ( $P < 0.001$ ) and the cirrhosis group ( $P < 0.001$ ), demonstrating its diagnostic value ( $P < 0.001$ ). As a confirmed diagnostic biomarker, AFP levels in PLC were higher than those in the other two groups. Noticeably, the median AFP level in PLC was 63.69 ng/mL, much lower than the 400 ng/mL listed in the guidelines, which was in accordance with clinical situations. Actually, AFP-negative PLC is not uncommon in clinical practice, which diminishes the diagnostic value of AFP. Although, the rate of AFP-positive PLC cases is not 100%, theoretically owing to various reasons including testing methods and the period of PLC, these data still surprised us. Our data showed that AFP levels were commonly below conceivable diagnostic expectations (only 35.6% of these cases were  $> 400$  ng/mL), which is in accordance with other studies[18].

**AFP levels and diagnostic value in PLC stratified by different criteria:** Tables 6 and 7 show AFP levels and its related diagnostic efficiency in each PLC stratification. AFP levels varied in different stratifications (Table 6). Furthermore, serum AFP levels in PLC with either decompensated or compensated liver cirrhosis were dramatically increased compared with PLC without cirrhosis ( $P = 0.004$  and  $P = 0.005$ , respectively). Additionally, AFP levels in PLC with diffuse liver cancer were strikingly increased compared with cases of small liver cancers ( $P = 0.007$ ,  $< 0.05$ ). BCLC stage D PLC patients had significantly increased AFP levels compared with BCLC stage A PLC patients ( $P = 0.009$ ,  $< 0.05$ ).

Next, the diagnostic values of AFP for stratified PLC cases were analyzed by receiver operating characteristic curves (Table 7 and Figure 1). When ranked in terms of diagnostic value, AFP had the greatest impact on PLC featuring Child–Pugh grade B, then compensated cirrhosis, Child–Pugh grade C, large tumors, and decompensated cirrhosis; moreover, area under the curve (AUC) values for AFP in these criteria were all  $> 0.800$  (0.847, 0.846, 0.821, 0.805, and 0.800, respectively), suggesting relatively better reliability. In contrast, AUC values for AFP were decreased in PLC featuring BCLC stage D, diffuse tumors, Child–Pugh grade A, and BCLC stages B and C (0.799, 0.785, 0.759, 0.741, and 0.731, respectively, all  $< 0.800$ ). Although the  $P$  values were all  $< 0.05$ , the diagnostic efficiency of serum AFP for small liver cancers and BCLC stage A liver cancer was low and meaningless (AUC = 0.595 and 0.592, respectively).

**The diagnostic value of combining CEA, CA 19-9, GGT, and AFP in PLC with small tumors and BCLC stage A cases:** The above findings demonstrated that AFP had little diagnostic value in PLC with small tumors and BCLC stage A cases. Thus, we next examined whether combined detection of CEA, CA 19-9, GGT, and AFP would have better results in this population. Considering that all four biomarkers are not only markers of gastrointestinal tumors, but also have intensive connections with liver diseases, the study was performed to supplement the disadvantages of AFP alone.

**Table 1 Influence of high-risk factors on primary liver cancer stratified by Child-Pugh score**

Factors	Criterion: Child-Pugh score			Statistics	P value
	A	B	C		
Male (%)	40 (76.9)	61 (77.2)	16 (76.9)	0.06	0.814
Age (yr)	59.40 ± 10.18	62.83 ± 9.85	58.50 ± 7.20	2.78	0.065
History of smoking (%)	16 (30.2)	22 (27.8) <sup>a</sup>	12 (60.0)	7.74	0.021
History of drinking (%)	13 (24.5) <sup>a</sup>	26 (32.9)	11 (55.0)	6.11	0.047
HBsAg (+) (%)	38 (71.7)	56 (70.9)	14 (70.0)	0.02	0.989
HCV-Ag (+) (%)	3 (5.7)	5 (6.3)	0 (0.0)	1.31	0.520
Non-alcoholic fatty liver (%)	9 (17.0)	18 (22.8)	1 (5.0)	3.47	0.176

<sup>a</sup>Compared with “C”, *P* < 0.0167. HBsAg: Hepatitis B virus antigen; HCV: Hepatitis C virus.

**Table 2 Influence of high-risk factors on primary liver cancer stratified by clinical stage of cirrhosis**

Factors	Criterion: Clinical stage of cirrhosis			Statistics	P value
	No cirrhosis	Compensated	Decompensated		
Male (%)	99 (72.3)	33 (78.6)	84 (77.1)	1.08	0.582
Age (yr)	62.93 ± 11.85	61.21 ± 1.58	60.93 ± 9.04	1.14	0.322
History of smoking (%)	47 (34.3)	13 (30.2)	37 (33.9)	0.26	0.880
History of drinking (%)	50 (36.5)	11 (26.2)	39 (35.8)	1.59	0.451
HBsAg (+) (%)	46 (33.6)	32 (76.2) <sup>a</sup>	76 (69.1) <sup>a</sup>	41.27	< 0.001
HCV-Ag (+) (%)	9 (6.6)	2 (4.8)	6 (5.5)	0.25	0.883
Non-alcoholic fatty liver (%)	39 (28.5)	9 (21.4)	19 (17.3)	4.38	0.112

<sup>a</sup>Compared with “No cirrhosis”, *P* < 0.0167. HBsAg: Hepatitis B virus antigen; HCV: Hepatitis C virus.

**Table 3 Influence of high-risk factors on primary liver cancer stratified by pathological classification of tumor size**

Factors	Criterion: Pathological classification of tumor size					Statistics	P value
	Small	Medium	Large	Massive	Diffuse		
Male (%)	30 (68.2)	17 (70.8)	30 (66.7)	22 (75.9)	117 (80.1)	5.05	0.283
Age (yr)	62.86 ± 9.03	59.45 ± 10.43	61.86 ± 8.86	57.79 ± 11.69	63.03 ± 11.75	1.75	0.139
History of smoking (%)	9 (20.5)	9 (37.5)	12 (26.7)	12 (41.4)	55 (37.4)	6.29	0.179
History of drinking (%)	10 (22.7)	8 (33.3)	14 (31.1)	11 (37.9)	57 (38.8)	4.28	0.370
HBsAg (+) (%)	20 (45.5)	14 (58.3)	23 (51.1)	14 (48.3)	83 (56.5)	2.30	0.680
HCV-Ag (+) (%)	3 (6.8)	1 (4.2)	4 (8.9)	2 (6.9)	7 (4.8)	1.32	0.858
Cirrhosis (%)	31 (70.5)	17 (70.0)	20 (44.4)	10 (34.5) <sup>a</sup>	74 (50.3)	14.15	0.007
Non-alcoholic fatty liver (%)	10 (22.7)	7 (29.2)	7 (15.6)	7 (24.1)	36 (24.5)	2.11	0.715

<sup>a</sup>Compared with “Small”, *P* < 0.005. HBsAg: Hepatitis B virus antigen; HCV: Hepatitis C virus.

Combining CEA, CA 19-9, and GGT improved the diagnostic performance of AFP (all AUC values > 0.600 and greater than AFP alone) (Tables 8 and 9, Figure 2). In particular, the diagnostic value of the combination had significantly increased values for small tumors and BCLC stage A (AUC = 0.810 and 0.846, respectively, *P* < 0.0001). Therefore, the combination of CEA, CA 19-9, GGT and AFP is worth using to diagnose PLC with small tumors or at BCLC stage A.

**Table 4 Influence of high-risk factors on primary liver cancer stratified by Barcelona stage**

Factors	Criterion: Barcelona stage				Statistics	P value
	A stage	B stage	C stage	D stage		
Male (%)	36 (67.9)	42 (76.4)	51 (75.0)	87 (77.7)	1.90	0.594
Age (yr)	60.25 ± 11.57	65.15 ± 12.40	62.50 ± 11.28	60.80 ± 9.05	2.53	0.057
History of smoking (%)	14 (26.4)	20 (36.4)	23 (33.8)	40 (35.4)	1.58	0.664
History of drinking (%)	15 (28.3)	16 (29.1)	28 (41.2)	41 (36.3)	3.11	0.375
HBsAg (+) (%)	21 (39.6) <sup>a</sup>	24 (43.6) <sup>a</sup>	31 (45.6) <sup>a</sup>	78 (69.0)	18.90	< 0.001
HCV-Ag (+) (%)	4 (7.5)	4 (7.3)	3 (4.4)	6 (5.3)	0.79	0.852
Cirrhosis (%)	11 (20.8) <sup>a</sup>	13 (23.6) <sup>a</sup>	15 (22.1) <sup>a</sup>	113 (100.0)	167.33	< 0.001
Non-alcoholic fatty liver (%)	14 (26.4)	18 (32.7)	15 (22.1)	20 (17.7)	5.08	0.166

<sup>a</sup>Compared with “D”, *P* < 0.0083. HBsAg: Hepatitis B virus antigen; HCV: Hepatitis C virus.

**Table 5 Analysis of alpha-fetoprotein level and diagnosis positive rate in patients with primary liver cancer, chronic hepatitis and cirrhosis**

Group	<i>n</i>	Men:Women	Age (mean ± S)	AFP (ng/mL)	Diagnosis positive rate, %
Chronic hepatitis B	217	157:60	59.85 ± 7.25	2.77 (1.67, 5.15) <sup>a</sup>	1.4 <sup>b</sup>
Cirrhosis	279	204:75	60.99 ± 9.91	3.26 (1.74, 9.80) <sup>a</sup>	4.7 <sup>b</sup>
PLC	289	216:73	61.92 ± 10.84	63.69 (3.28, 1210.00)	35.6
Statistics		0.536	2.869	121.974	147.565
<i>P</i> value		0.765	0.057	< 0.001	< 0.001

<sup>a</sup>Compared with “PLC group”, *P* < 0.05.

<sup>b</sup>Compared with “PLC group”, *P* < 0.0167. AFP: Alpha-fetoprotein; PLC: Primary liver cancer.

## DISCUSSION

Early diagnosis is crucial to decreasing the recurrence rate of PLC after surgery. Considering that current treatments tend to be more accurate and individualized, stratifying cases might play a role in making more precise PLC diagnoses. Previous studies have pointed out that chronic HBV exposure, fatty liver, cigarette and alcohol use, and liver cirrhosis could help tumor cells escape immune surveillance and promote tumor proliferation and metastasis[19,20,11]. Additionally, the prognosis of PLC is impacted by tumor size, number, the presence of vascular invasion, and lymph node metastasis [21]. In clinical practice, these high-risk factors are mixed in with the different criteria we analyzed; however, there is no good model featuring the different phases of PLC, meaning it is essential to treat the factors distinctively.

Our results demonstrate that certain factors have different diagnostic roles when PLC cases are stratified. Currently, there are various PLC scoring systems that use a combination of routine clinical features[22]. We first found when PLC was stratified by Child-Pugh score, the rates of smoking and drinking history played the biggest diagnostic roles. Generally, smoking and drinking induce direct or indirect toxic effects that increase the risk of developing PLC among chronic liver disease patients[23]. Second, when we stratified cases by clinical stage of cirrhosis, HBsAg was the most discriminatory factor. The literature suggests that HBV proteins are involved in hepatocarcinogenesis[24]. On the basis of our results, patients with HBV infection are advised to be on alert for liver cirrhosis to reduce the incidence of PLC. Subsequently, PLC was stratified by tumor size, and the data demonstrated that liver cirrhosis occurred more frequently in patients with small liver tumors than in those with massive liver tumors. It has been reported that the presence of cirrhosis or advanced liver fibrosis is a distinct predisposing factor for liver cancer, predominantly hepatocellular carcinoma[25]. Briefly, caution should be taken when treating patients with small tumors that are inclined to cirrhosis, surgery should be advised for these patients as it could improve their prognosis. Finally, when our data were stratified by BCLC stage, both HBsAg and liver cirrhosis changed at different stages. The BCLC staging system classifies cases based on the patient's life expectancy, meaning more emphasis is placed on both HBV infection

**Table 6 Alpha-fetoprotein expression levels in primary liver cancer by different standards**

Layering standard	Group	AFP (ng / mL)	F	P value
Cirrhosis clinical stage	No cirrhosis	7.32 (2.28, 7.32)	10.03	0.007
	Compensated cirrhosis	267.40 (12.56, 1210.00) <sup>a</sup>		
	Decompensated cirrhosis	130.60 (6.69, 1210.00) <sup>a</sup>		
Child-Pugh classification	A grade	461.16 ± 546.16	0.26	0.774
	B grade	516.74 ± 534.18		
	C grade	549.64 ± 561.80		
Classification of tumor size	Small liver cancer	4.54 (1.78, 99.19)	21.70	< 0.001
	Medium liver cancer	12.56 (2.66, 521.65)		
	Large liver cancer	17.58 (2.10, 782.25)		
	Massive liver cancer	249.90 (4.14, 1210.00)		
	Diffuse liver cancer	267.40 (3.69, 1210.00) <sup>b</sup>		
Barcelona stage	Phase A	3.76 (1.78, 478.95)	8.56	0.036
	Phase B	103.90 (3.17, 1149.50)		
	Phase C	111.90 (3.18, 1210.00)		
	Phase D	116.25 (6.59, 1210.00) <sup>c</sup>		

<sup>a</sup>Compared with “no cirrhosis”, *P* < 0.05.

<sup>b</sup>Compared with “small liver cancer”, *P* < 0.05.

<sup>c</sup>Compared with “Phase A”, *P* < 0.05. AFP: Alpha-fetoprotein.

and cirrhosis when evaluating prognosis.

As mentioned above, stratification might make a difference when assessing PLC. Thus, we next examined the trend of biomarkers in each phase. Toader *et al*[26] found that AFP was still a reliable diagnostic and prognostic tool for PLC patients, especially in developing countries. In our study, the diagnostic value of AFP alone for PLC was not as good as expected, and there was even a small proportion of AFP-negative PLC cases[27]. A meta-analysis of studies showed that concomitant use of ultrasound and AFP improved early PLC detection compared with ultrasound alone[28]. Moreover, AFP is still regarded as a biomarker in some PLC guidelines. Thus, we thought it would be valuable to further study AFP according to patient stratification. Our analysis showed that AFP had quite different functions in different stratifications of PLC, having greater utility in patients with poor liver function, advanced, massive, and multiple tumors.

We found that AFP was not suitable for diagnosing small and early-stage liver cancers. However, there has been increasing recognition that a single biomarker may not be sufficient and that a combination of biomarkers may be needed to optimize sensitivity for small and early PLC. Currently, new serum tumor markers such as Golgi protein 73[29], glypican-3[30], and liver cancer-related miRNA [31] have been shown to be promising biomarkers that could improve the diagnostic efficiency of PLC, but most of these indicators are still not suitable for clinical use. Thus, we analyzed classic liver markers including CEA, CA 19-9[32], and GGT[33]. CEA and CA 19-9 are serum tumor markers that are primarily used for screening and diagnosis of gastrointestinal and other digestive system tumors. Both show a certain degree of expression in PLC. Our results confirmed that the combined detection of these four biomarkers contributed to increasing diagnostic performance for small and early-stage liver cancers.

Due to my limited energy, no further in-depth study was conducted. In future studies, we should study AFP in patients with liver cancer and patient prognosis, 5-year survival rate, *etc.*

**Table 7 Analysis of diagnostic value of alpha-fetoprotein for primary liver cancer with different criteria**

Layering standard	Group	AUC	Std.Error	P value	95%CI	Sensitivity, %	Specificity, %
Child-Pugh classification	A grade	0.759	0.0424	< 0.0001	(0.721, 0.794)	62.26	84.68
	B grade	0.847	0.0279	< 0.0001	(0.815, 0.875)	77.22	81.45
	C grade	0.821	0.0575	< 0.0001	(0.785, 0.853)	75.00	81.25
Cirrhosis clinical stage	No	0.647	0.0303	< 0.0001	(0.608, 0.684)	38.69	94.56
	Compensated cirrhosis	0.846	0.0369	< 0.0001	(0.813, 0.875)	78.57	81.05
	Decompensated cirrhosis	0.800	0.0276	< 0.0001	(0.766, 0.8310)	69.09	83.47
Classification of tumor size	Small	0.595	0.0514	0.0642	(0.552, 0.637)	38.64	84.68
	Medium	0.684	0.0636	0.0038	(0.642, 0.724)	66.67	73.39
	Large	0.675	0.0520	0.0008	(0.634, 0.714)	66.67	71.98
	Massive	0.805	0.0411	< 0.0001	(0.769, 0.838)	51.72	96.17
	Diffuse	0.785	0.0257	< 0.0001	(0.751, 0.816)	62.59	90.93
Barcelona stage	Phase A	0.592	0.0481	0.0547	(0.550, 0.634)	30.19	96.17
	Phase B	0.741	0.0415	< 0.0001	(0.703, 0.778)	52.73	90.93
	Phase C	0.731	0.0403	< 0.0001	(0.692, 0.767)	52.94	93.35
	Phase D	0.799	0.0271	< 0.0001	(0.765, 0.830)	68.14	83.47

AUC: Area under the curve.

**Table 8 Evaluation of four indicators for diagnosing small liver cancer alone and in combination**

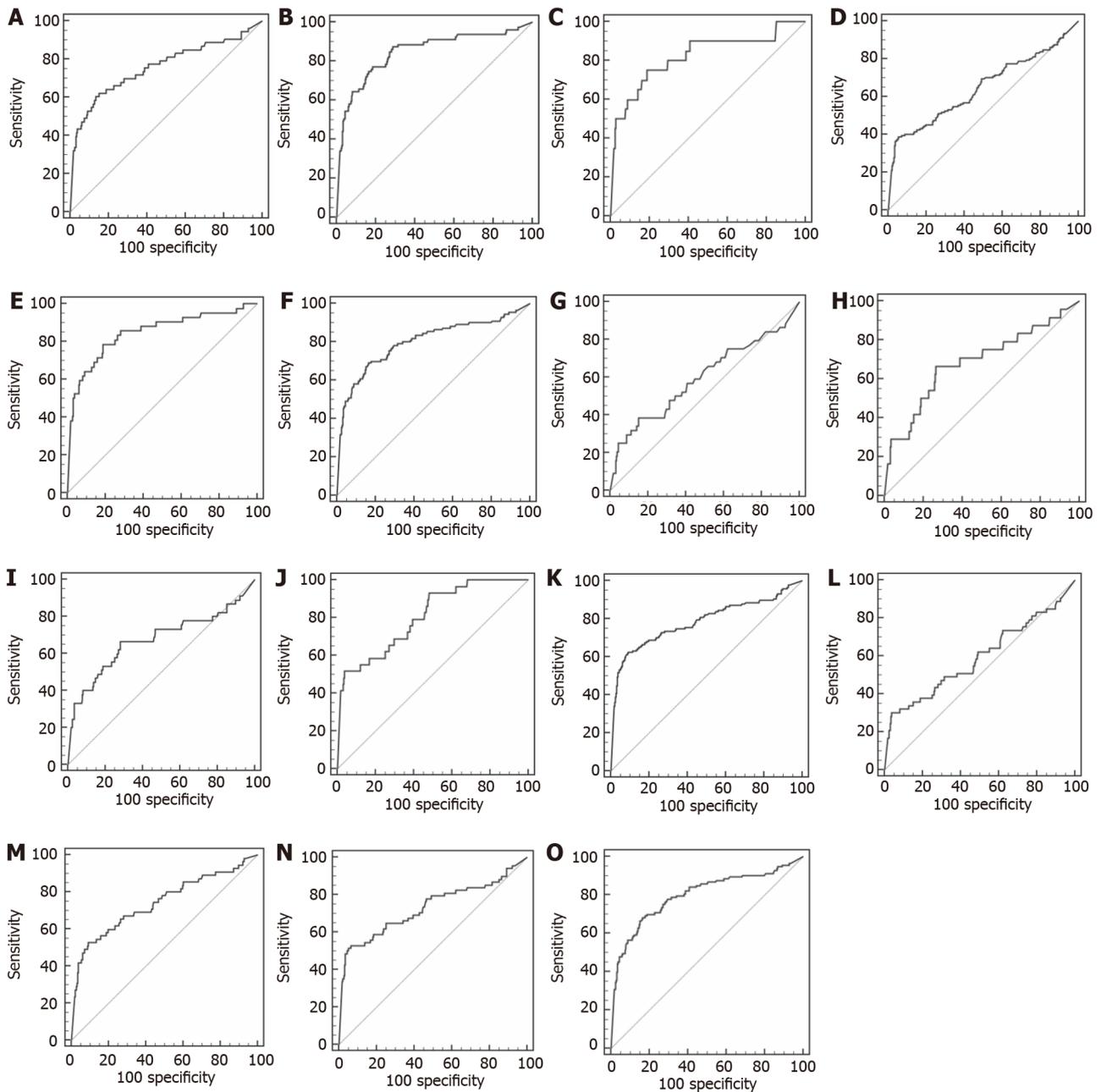
Diagnostic indicators	AUC	Sensitivity, %	Specificity, %	Std.Error	95%CI	Z	P value
AFP	0.595	38.64	84.68	0.0519	(0.552, 0.632)	1.93	0.067
CEA	0.649	52.27	75.22	0.0461	(0.589, 0.706)	3.23	0.0013
CA-199	0.719	81.82	66.18	0.0467	(0.671, 0.763)	4.69	< 0.0001
GGT	0.728	77.27	56.89	0.0390	(0.671, 0.780)	5.84	< 0.0001
Combined diagnosis	0.810	68.18	79.56	0.0349	(0.758, 0.855)	8.90	< 0.0001

AUC: Area under the curve; AFP: Alpha-fetoprotein; CEA: Carcinoembryonic antigen; CA 19-9: Carbohydrate antigen 19-9; GGT: Gamma-glutamyl transferase.

**Table 9 Evaluation of the four indicators for the diagnosis of Barcelona Clinic Liver Cancer stage A liver cancer alone and in combination**

Diagnostic indicators	AUC	Sensitivity, %	Specificity, %	Std.Error	95%CI	Z	P value
AFP	0.593	30.19	96.17	0.0481	(0.550, 0.634)	1.93	0.0538
CEA	0.611	32.08	96.00	0.0462	(0.551, 0.669)	1.197	0.2313
CA-199	0.608	75.47	42.86	0.0429	(0.548, 0.666)	2.511	0.0121
GGT	0.743	60.38	78.22	0.0380	(0.688, 0.794)	6.409	< 0.0001
Combined diagnosis	0.843	66.04	91.96	0.0319	(0.794, 0.883)	10.738	< 0.0001

AUC: Area under the curve; AFP: Alpha-fetoprotein; CEA: Carcinoembryonic antigen; CA 19-9: Carbohydrate antigen 19-9; GGT: Gamma-glutamyl transferase.

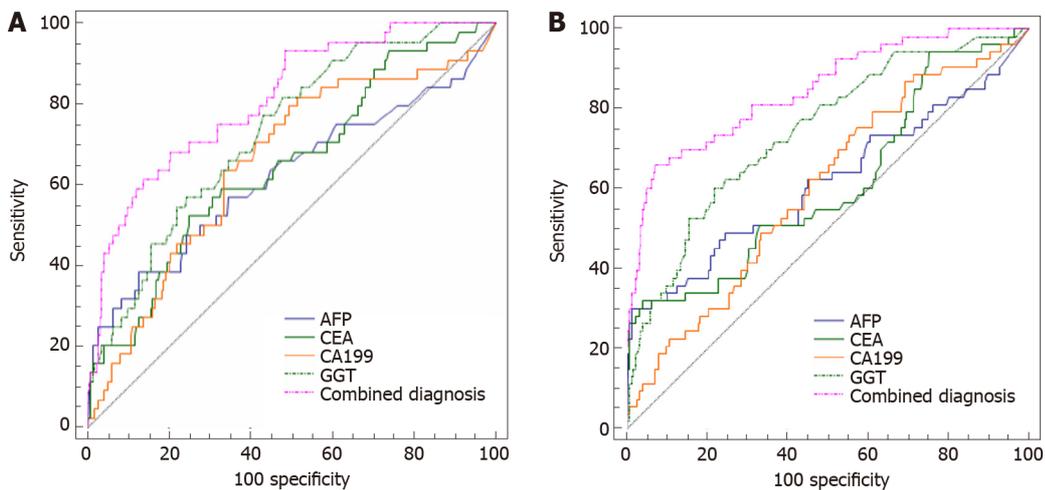


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**Figure 1 Receiver operating characteristic curves in different stratifications of primary liver cancer.** A-O: Represent primary liver cancer (PLC) of Child-Pugh A, PLC of Child-Pugh B, PLC of Child-Pugh C, PLC without cirrhosis, PLC with the compensated phase of cirrhosis, PLC with the decompensated phase of cirrhosis, small tumors, medium tumors, large tumors, massive tumors, diffuse tumors, PLC at Barcelona Clinic Liver Cancer (BCLC) stage A, PLC at BCLC stage B, PLC at BCLC stage C, and PLC at BCLC stage D, respectively. The contributions of related high-risk factors to stratified PLC were compared according to Child-Pugh score, clinical stage of concurrent liver cirrhosis, tumor size and BCLC stage, respectively.

## CONCLUSION

In summary, stratified diagnosis of PLC was essential, and the high-risk factors had distinct roles in PLC classifications. AFP level functioned as a diagnostic biomarker in the stratified PLC population that included the following: poor function, advanced hepatoma, and massive and diffuse tumors. For small hepatomas and BCLC A stage PLC, combined detection of CEA, CA 19-9, GGT, and AFP is a more promising approach to diagnosing PLC compared with testing AFP alone.



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**Figure 2 Area under the curve.** A-B: Represent small tumors and primary liver cancer (PLC) at Barcelona Clinic Liver Cancer (BCLC) stage A, respectively. Data above demonstrated that alpha-fetoprotein (AFP) had little diagnostic value in PLC patients with small tumors and at BCLC stage A. As shown, carcinoembryonic antigen, carbohydrate antigen 19-9, and gamma-glutamyl transferase improved the diagnostic value of AFP. AFP: Alpha-fetoprotein; BCLC: Barcelona Clinic Liver Cancer; CEA: Carcinoembryonic antigen; CA 19-9: Carbohydrate antigen 19-9; GGT: Gamma-glutamyl transferase.

## ARTICLE HIGHLIGHTS

### Research background

Stratified diagnosis of primary liver cancer (PLC) was essential, and the high-risk factors had distinct roles in PLC classifications. Alpha-fetoprotein (AFP) level functioned as a diagnostic biomarker in the stratified PLC population that included the following: Poor function, advanced hepatoma, and massive and diffuse tumors. For small hepatomas and Barcelona Clinic Liver Cancer (BCLC) A stage PLC, combined detection of carcinoembryonic antigen (CEA), carbohydrate antigen 19-9 (CA 19-9), gamma-glutamyl transferase (GGT), and AFP is a more promising approach to diagnosing PLC compared with testing AFP alone.

### Research motivation

Stratification of PLC was essential for precise diagnoses and benefited from evaluating AFP levels.

### Research objectives

This study aimed to evaluate the role of high-risk factors in diagnosing stratified PLC cases, especially the diagnostic value of AFP.

### Research methods

First, the contributions of high-risk factors in stratifying PLC were compared. Then, the diagnostic value of AFP was evaluated in different stratifications of PLC by receiver operating characteristic curves. For PLC cases in which AFP played little role, the diagnostic values of CEA, CA 19-9, GGT, and AFP were analyzed.

### Research results

The roles of high-risk factors differed in stratified PLC. AFP levels were higher in PLC with cirrhosis, diffuse tumors, and BCLC stage D disease. However, these measures were meaningless [area under the curve (AUC) < 0.600] in small hepatomas and BCLC A stage PLC, but could be replaced by the combined detection of CEA, CA 19-9, GGT, and AFP (AUC = 0.810 and 0.846, respectively).

### Research conclusions

PLC incidence rates vary across clinical etiologies and conditions such as liver disease severity; even within the same clinical entity, individual PLC risk is heterogeneous across patients for unknown reasons. Hence, clinically meaningful utility must be demonstrated under specific clinical scenarios for a diagnostic modality to be adopted into regular use.

### Research perspectives

PLC was the second leading cause of cancer-related mortality in 2014. PLC incidence rates vary across clinical etiologies and conditions.

## FOOTNOTES

**Author contributions:** Wu JH contributed to conception and design; Jiao HB contributed to collection and assembly of data; Jiao HB and Wang W contributed to manuscript writing, data analysis and interpretation; all authors contributed to final approval of the manuscript; Jiao HB and Wang W contributed equally to this work.

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**Data sharing statement:** Dataset available from the corresponding author at [tswujinghua@126.com](mailto:tswujinghua@126.com).

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