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***Clinical Trials Study***

**Genome-wide methylation profiling of early colorectal cancer using an Illumina Infinium Methylation EPIC BeadChip**

Wu YL *et al*. Genome-wide methylation profiling of CRC

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**Author contributions:** Wu YL, Jiang T, Zhang PJ and Tian YP designed the study; Wu YL performed the research, wrote the paper; Wu YL and Jiang T analyzed the data, and revised the manuscript for final submission; Huang W, Wu XY participated in the processing of bioinformatic analysis; Wu YL and Jiang T contributed equally to this study; Zhang PJ and Tian YP are the co-corresponding authors; and all authors have read and approve the final manuscript.

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**Abstract**

BACKGROUND

DNA methylation is a part of epigenetic modification, that is closely related to the growth and development of colorectal cancer (CRC). Specific methylated genes and methylated diagnostic models of tumors have become current research focuses. The methylation status of circulating DNA in plasma might serve as a potential biomarker for CRC.

AIM

To investigate genome-wide methylation pattern in early CRC using the Illumina Infinium Human Methylation 850K BeadChip.

METHODS

The 850K Methylation BeadChip was used to analyze the genome-wide methylation status of early CRC patients (*n* = 5) and colorectal adenoma patients (*n* = 5). Gene ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways enrichment analyses were performed on the selected differentially methylated sites to further discover candidate methylation biomarkers in plasma.

RESULTS

A total of 1865 methylated CpG sites with significant differences were detected, including 676 hypermethylated sites and 1189 hypomethylated sites. The distribution of these sites covered from the 1st to 22nd chromosomes and are mainly distributed on the gene body and gene promoter region. GO and KEGG enrichment analysis showed that the functions of these genes were related to biological regulation, molecular binding, transcription factor activity and signal transduction pathway.

CONCLUSION

The study demonstrated that the Illumina Infinium Human Methylation 850K BeadChip can be used to investigate genome-wide methylation status of plasma DNA in early CRC and colorectal adenoma patients.

**Key Words:** Colorectal cancer; DNA methylation; 850K Methylation BeadChip; Plasma; colorectal adenoma

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**Core Tip:** DNA methylation is associated with the growth and development of colorectal cancer (CRC). CRC usually develops from its precancerous lesion, colorectal adenoma. The study demonstrated that the Illumina Infinium Human Methylation 850K BeadChip can be used to investigate genome-wide methylation status of plasma DNA in early CRC and colorectal adenoma patients.

**INTRODUCTION**

Colorectal cancer (CRC) is a common malignancy with high morbidity and mortality. Tumors are highly associated with aberrant DNA methylation alterations, which are involved in the regulation of genomic function[1]. DNA methylation in promoter regions leads to gene silencing, which plays an important role in the development and progression of CRC. The incidence of CRC is insidious, and its early manifestation is not obvious, thus, early effective screening is of great significance to reduce morbidity and mortality[2]. CRC usually develops from its precancerous lesion, colorectal adenoma. The evolution from colorectal adenoma to CRC is associated with hypermethylation of tumor-related genes and genome-wide hypomethylation of CpG sites[3,4]. Various histological subtypes of colorectal adenoma also have different methylation patterns[5]. Genome-wide methylation profiling of early CRC and colorectal adenoma will contribute to the discovery of tumor biomarkers, thus improving the early screening rate of CRC.

Circulating tumor DNA (ctDNA), a free DNA released by tumor cells, was found in plasma. The genetic alterations between ctDNA and tumor cell DNA were consistent. A variety of tumor-specific alterations could be detected through ctDNA, including abnormal methylation[6]. Therefore, the detection of ctDNA methylation in plasma can facilitate the cancer diagnosis. Some research had studied genomic methylation of CRC in plasma or serum[7-10], but few focused on the ctDNA methylation differences between early CRC and colorectal adenoma. The Illumina Infinium Human Methylation 850K BeadChip has technical superiority in high-throughput detection for genomic methylation status. The 850K chip contains 91% of the 450K chip and 413745 additional sites (a total of 866895 CpG sites), covering gene promoter regions, gene coding regions, CpG islands and enhancers in the ENCODE and FANTOM5 programs[11-13]. The research efficiency has been greatly improved.

In the present study, an Illumina Infinium Human Methylation 850K BeadChip was used to detect methylation differences of plasma DNA between CRC patients and colorectal adenoma patients, gene ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analyses was also performed to investigate the biological functions of differentially methylated sites (DMSs). This study aimed to investigate the different methylation pattern in plasma DNA between early CRC and colorectal adenoma using the Human Methylation 850K BeadChip.

**MATERIALS AND METHODS**

***Patients***

In this study, the Human Methylation 850K BeadChip assay included early CRC patients (*n* = 5) and colorectal adenoma patients (*n* = 5) hospitalized in the Chinese PLA General Hospital from January 2018 to January 2020 (Table 1). Their ages ranged from 52 to 79 years old, with an average age of 63 years old. Inclusion criteria: None of the patients had received any treatment when their peripheral blood samples were taken, including surgical resection, radiotherapy, chemotherapy, and targeted therapy; all patients who underwent colonoscopy and endoscopic biopsy, which were confirmed as colorectal adenoma (without cancerization) by pathology, were included in the adenoma group, which were confirmed as CRC by pathology and at stage I according to the American Joint Committee on Cancer tumor-node-metastasis staging system for CRC, were included in the tumor group; all patients had no other malignant tumors or serious diseases. Later clinical follow-up was required for all patients. If the postoperative pathological results of the tumor tissue were inconsistent with those of endoscopic biopsy, postoperative histopathology prevailed, and cases that did not meet the inclusion criteria were excluded. All participants signed written informed consent for the collection of samples and subsequent analyses before inclusion.

***DNA isolation and bisulfite conversion***

Peripheral blood was collected from the veins of patients and centrifuged within 30 min (4℃, 3000 rpm, 10 min). The plasma was collected and stored at 4℃ for DNA extraction within 3 days, according to the instructions of the QIAamp MinElute ccfDNA Mini Kit (QIAGEN, Germany, 55204). The DNA was treated with bisulfite to convert cytosine to uracil according to the instructions of the EZ DNA Methylation-Gold Kit (Zymo Research, USA, D5001/D5003). Subsequently Qubit ssDNA HS Assay Kit (Invitrogen, USA, Q32854) as well as Qubit 3.0 fluorescence quantizer (Thermo Fisher, USA) were used for DNA quantitative analysis after bisulfite conversion. The bisulfite-converted DNA was stored at -20℃ until use.

***850K methylation assay and bioinformatic analysis***

Genome-wide methylation profiling was perfrormed by Illumina Infinium Human Methylation 850K BeadChip (Sinotech Genomics, China), in accordance with Illumina's standard protocol. Bisulfite-converted DNA was used for alkali denaturation and genomic amplification. Then, the amplified DNA was used for hybridization, washing, fluorescent staining, single-base extension and scanning on an 850K BeadChip. An IScan software analysis system (Illumina, USA) was used for signal acquisition and analysis. The data (IDAT files) were mainly analyzed using the ChAMP package in R. We used β values (range, 0-1) to represent the DNA methylation level, wAhich were calculated from the intensity ratio of the methylated signals to the total (methylated and unmethylated) signals for each site. Then we compared the β values of each probe site in the samples between the CRC group and adenoma group, calculating the average β value of each site and the difference in β values between groups (Δβ). For plasma samples, a CpG site with |Δβ| > 0.1 and a *P* value < 0.01 was considered as a DMS (Δβ > 0.1 was considered a hypermethylated site, and Δβ < -0.1 was considered a hypomethylated site). The Benjamini and Hochberg method was applied to reduce the false-positive rate[14].

We also conducted GO and KEGG enrichment analyses for DMSs using the R package. In the functional enrichment analysis, the *P* values represented the likelihood that the gene associated with the GO term would be enriched in the input gene list, and a *P* value less than 0.05 was considered significant.

**RESULTS**

***Genome-wide detection of methylation in CRC plasma by the 850K methylation BeadChip***

The 850K methylation BeadChip included a total of 866895 CpG sites, screening out 1865 differentially methylated CpG sites between CRC and adenoma plasma samples (|Δβ| > 0.1, *P* < 0.01), including 676 hypermethylation sites and 1189 hypomethylation sites (Figure 1). These DMSs were widely distributed on all chromosomes, among which, chromosomes 1 and 2 had the most sites and chromosomes 21 and 22 had the least sites (Table 2). Figure 2A and B shows the distribution of DMSs on the chromosomes of the genome. There were also differences in the distribution of the gene structure elements, mainly in the gene body (64.1%) and gene promoter region (32.7%, including TSS1500, TSS200, 5'UTR, and 1stExon). The distribution of DMSs in and around CpG islands was also different, mainly in the CpG Shore (53.7%) and CpG Island regions (20.3%) (Figure 3A and B).

***GO enrichment analysis***

GO enrichment analysis including the biological process, cellular component, and molecular function categories, was performed. A *P* value of < 0.05 was considered statistically significant. After GO enrichment analysis of the selected DMSs, 783 GO functional annotations were obtained, including 571 terms of biological processes, 107 terms of cell components and 105 terms of molecular functions. These GO functional terms indicated that DMSs in CRC cover a variety of functional communities. In terms of biological process, differentially expressed genes were mainly enriched in biological regulation, cellular process, metabolic process, regulation of biological process, and single organism process. The cell component analysis showed that molecules distributed in the extracellular region, cell membrane, organelle and synapse were significantly enriched. At the molecular level, functional enrichment annotation showed that the highly enriched genes were associated with molecular binding, catalytic activity and signaling pathways (Figure 4A). The bubble diagram in Figure 4B shows the top 30 GO terms with the most significant enrichment effect. Significantly enriched biological processes included gamma−aminobutyric acid depletion and transport and cerebellar Purkinje cell layer development. Significantly enriched cellular components included the filopodium tip and apical cortex. Significantly enriched molecular functions included microfilament motor activity, ephrin receptor activity and phosphatidylinositol phospholipase C and sphingolipid binding (Figure 4B).

***KEGG pathway enrichment analysis***

KEGG pathways were divided into cellular processes, environmental information processing, genetic information processing, human disease metabolism, metabolism, and organismal systems. KEGG annotation results of DMSs were classified according to the above six pathways (Figure 5A). A total of 26 KEGG signaling pathways were significantly enriched, which are shown in the bubble diagram (Figure 5B). KEGG analysis showed that DMSs were involved in various cellular pathways in CRC. Significantly enriched pathways included long-term depression, calcium signaling pathway, synapse related pathway, Rap1 signaling pathway, PI3K-Akt signaling pathway and MAPK signaling pathway.

**DISCUSSION**

Cancer is associated with a series of abnormal alterations in genetics and epigenetics. Mutations or epigenetic alterations in key pathways involved in cell growth and division may induce the occurrence of tumors[15]. Aberrant methylation modifications in the colon can be observed in early premalignant lesions such as adenoma and normal mucosa of para-carcinoma tissue[16]. And DNA methylation is an important epigenetic alteration that leads to silencing of the expression of tumor-related genes, and many abnormal methylations have been found in the genomic DNA of tumor cells, which are also reflected in plasma ctDNA. Therefore, the detection of ctDNA in plasma is becoming a new method of cancer diagnosis and monitoring[9]. Current ctDNA detection techniques based on gene mutations mainly rely on the number of tumor-derived DNA molecules in the sample, the number of mutated cancer cells, the concentration fraction of ctDNA and the sensitivity of analysis[17]. Using methylation patterns to detect ctDNA in plasma has advantages over mutation-based tests. Epigenetic modifications are more consistent in tumor cells than genetic mutations[18]. Therefore, early CRC can be diagnosed by detecting the methylation pattern of ctDNA.

The detecting DNA for 850K Methylation BeadChip is first treated with bisulfite to convert methylation differences into differences in bases. The Illumina Infinium Human Methylation 850K BeadChip has two detection methods (Infinium I and Infinium II). Infinium I has two specific probes for the bisulfite-converted DNA sequence. The M-type magnetic beads match the methylated sites, and the U-type magnetic beads match the unmethylated sites. Infinium II had only one probe, and only one base is added for single nucleotide extension after pairing. Different bases are added for methylated and unmethylated sequences, producing different fluorescence signals, and the level of methylation is calculated according to the signal ratio[11,19].

This study used an Illumina Infinium Human Methylation 850K BeadChip to detect and analyze the genome-wide methylation status of 5 early CRC patients and 5 cases of adenoma. According to the research findings, there were 1865 DMSs, including 676 hypermethylated sites and 1189 hypomethylated sites. The distribution of these methylated sites in the chromosomes, gene structural elements, CpG islands and their surrounding regions were all different. Then GO and KEGG enrichment analyses were performed on the differentially methylated genes. GO and KEGG functional annotation indicated that differentially methylated genes in CRC cover a variety of different functional aspects and participate in multiple biological pathways. Examples are Calcium signaling pathway, Rap1 signaling pathway, PI3K-Akt signaling pathway, MAPK signaling pathway, cell adhesion pathway and metabolic pathway, are all important biological pathways in tumor research[20-23]. These results indicated that multiple signaling pathways might be simultaneously regulated by DNA methylation in the progression of colorectal adenoma to early CRC. The limitation of this study is the small sample size due to the high cost. Due to the low content of plasma DNA, there was basically no residual after the chip detection, so we had not conducted other studies for verification. This project used an 850K Methylation chip to detect more than 850000 sites with low initial detecting DNA content, suitable for the methylation screening of trace DNA. Compared with other methylation chips, its detection flux had a qualitative improvement. And compared with the next generation whole-genome methylation sequencing technology, it solved the contradiction that the initial DNA content of sequencing technology is high (3-5 μg) while the plasma DNA content is very low[24]. Previously, the 850K methylation chip detection for CRC mostly used tissue or mucosal samples[25], so the sampling method is invasive and might cause tumor heterogeneity due to sampling deviation. In this study, plasma samples from patients with CRC and adenoma were used for genome-wide methylation analysis to screen for differentially methylated sites. Non-invasive samples were obtained from peripheral blood of patients. Therefore, this novel technology can be used as an effective tool for screening plasma DNA methylation sites.

**CONCLUSION**

This study showed that the Illumina Infinium Human Methylation 850K BeadChip can be used to investigate genome-wide methylation pattern of plasma DNA in tumors for the discovery of biomarkers. The limitations of this study are the sample size and the deficiency of verification. Further researches are needed on how to reduce the amount of initial DNA for the chip, and how to apply the selected biomarkers to cancer diagnosis.

**ARTICLE HIGHLIGHTS**

***Research background***

DNA methylation is closely related to the growth and development of colorectal cancer (CRC). Circulating tumor DNA (ctDNA) in plasma also has tumor-specific methylation patterns, the methylation status of ctDNA might serve as a potential biomarker.

***Research motivation***

In this study, we investigated the different methylation pattern between early CRC and colorectal adenoma.

***Research objectives***

This study aimed to analyze the genomic methylation status of CRC and discover potential methylated biomarkers for CRC diagnosis by Illumina Infinium Human Methylation 850K BeadChip.

***Research methods***

We used 850K Methylation BeadChip and enrichment analysis to select the differentially methylated sites of early CRC patients (*n* = 5) and colorectal adenoma patients (*n* = 5). Then, the gene ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis was performed.

***Research results***

We found 1865 methylated sites with significant differences (676 hypermethylated sites and 1189 hypomethylated sites). The distribution of these sites covered from the 1st to 22nd chromosomes and are mainly distributed on the gene body and gene promoter region. GO and KEGG enrichment analysis identified that the functions of these genes were related to biological regulation, molecular binding, transcription factor activity and signal transduction pathway.

***Research conclusions***

The Illumina Infinium Human Methylation 850K BeadChip can be used to investigate genome-wide methylation status of plasma DNA to select potential methylated biomarkers for CRC diagnosis.

***Research perspectives***

Genome-wide methylation analysis of early CRC and screening of biomarkers by 850K Methylation BeadChip provided a new possibility for early diagnosis of CRC.

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**Footnotes**

**Institutional review board statement:** The study was reviewed and approved by the Medical Ethics Committee of PLA General Hospital.

**Clinical trial registration statement:** We declared that this study has been reviewed and approved by the Medical Ethics Committee of PLA General Hospital. The number was S2022-003-01.

**Informed consent statement:** All study participants, or their legal guardian, provided informed written consent prior to study enrollment.

**Conflict-of-interest statement:** We declare that we have no financial or personal relationships with other individuals or organizations that can inappropriately influence our work and that there is no professional or other personal interest of any nature in any product, service and/or company that could be construed as influencing the position presented in or the review of the manuscript.

**Data sharing statement:** No additional data are available.

**CONSORT 2010 statement:** The authors have read the CONSORT 2010 statement, and the manuscript was prepared and revised according to the CONSORT 2010 statement.

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**Peer-review report’s scientific quality classification**

Grade A (Excellent): A

Grade B (Very good): 0

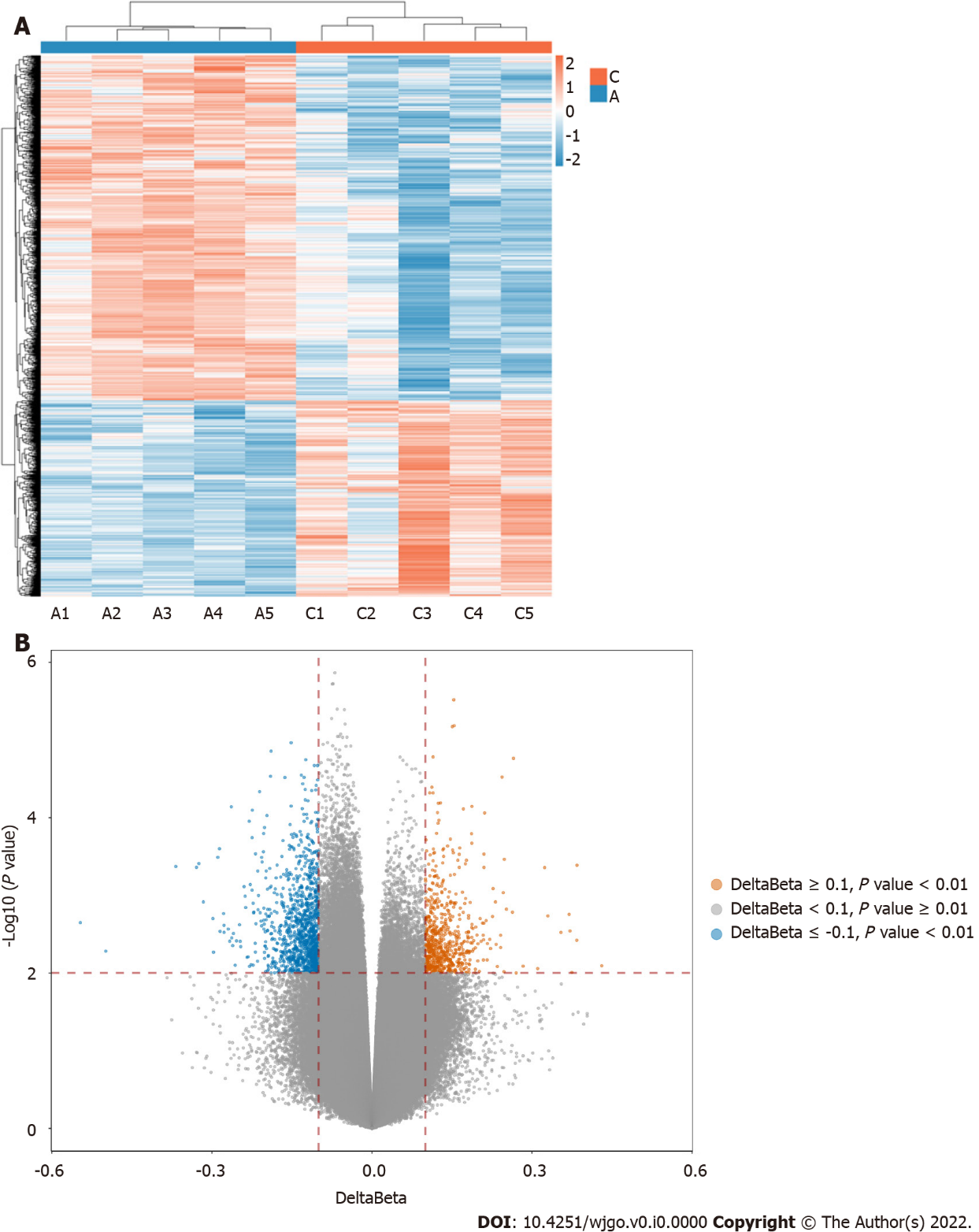
Grade C (Good): C

Grade D (Fair): 0

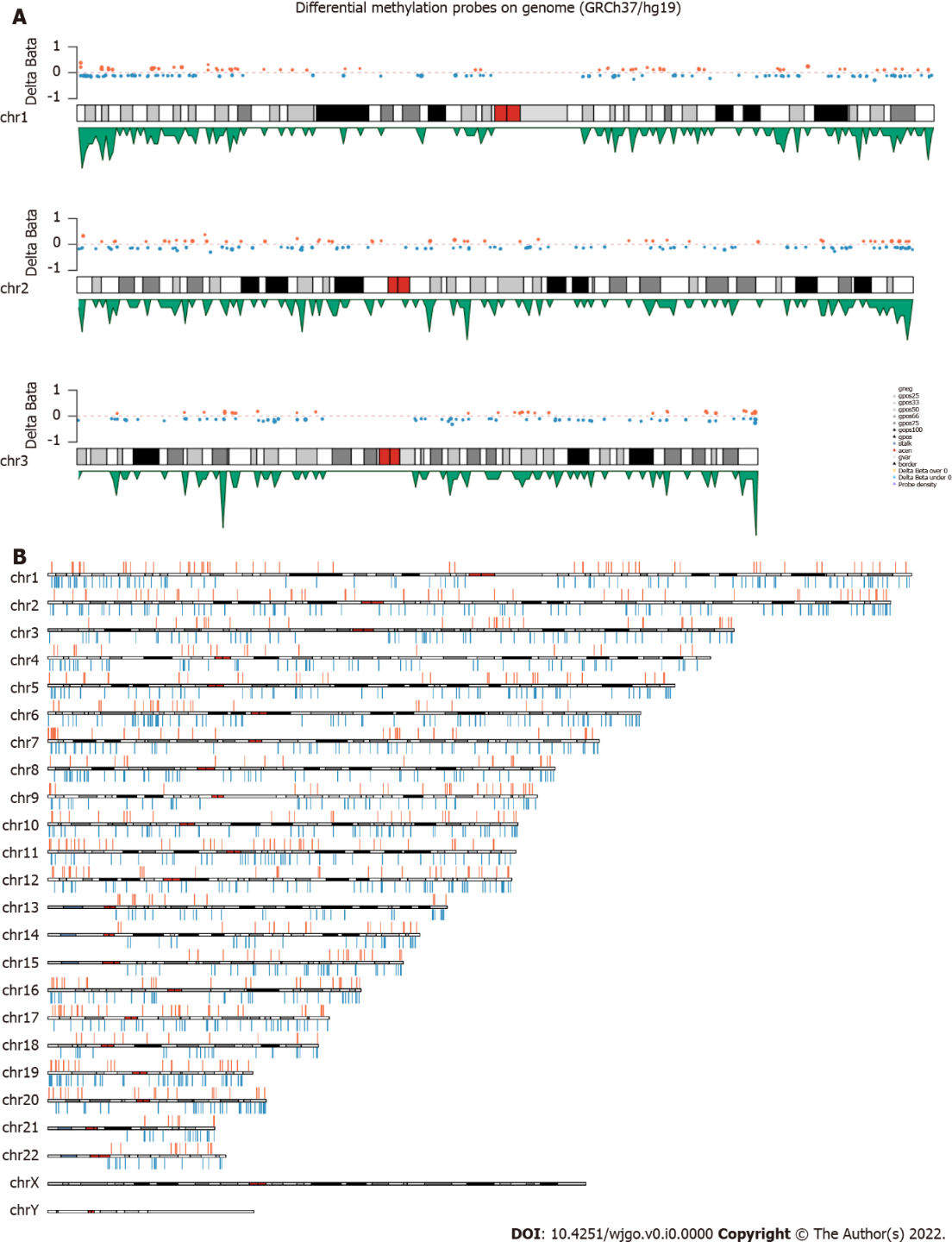
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**P-Reviewer:** Kołat D, Poland; Osera S, Japan **S-Editor:** Wang JL **L-Editor:** A **P-Editor:** Wang JL

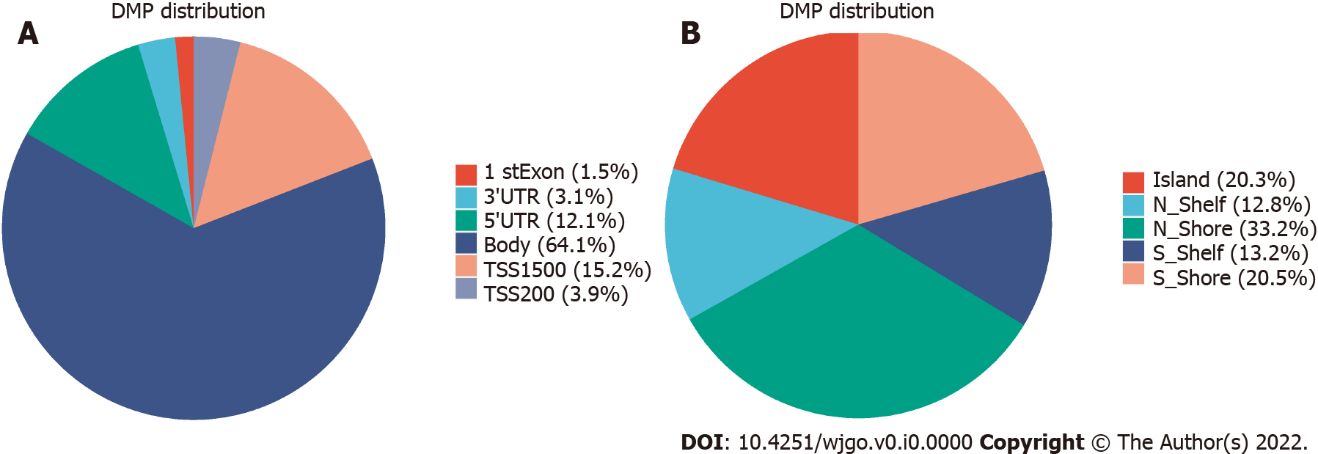
**Figure Legends**



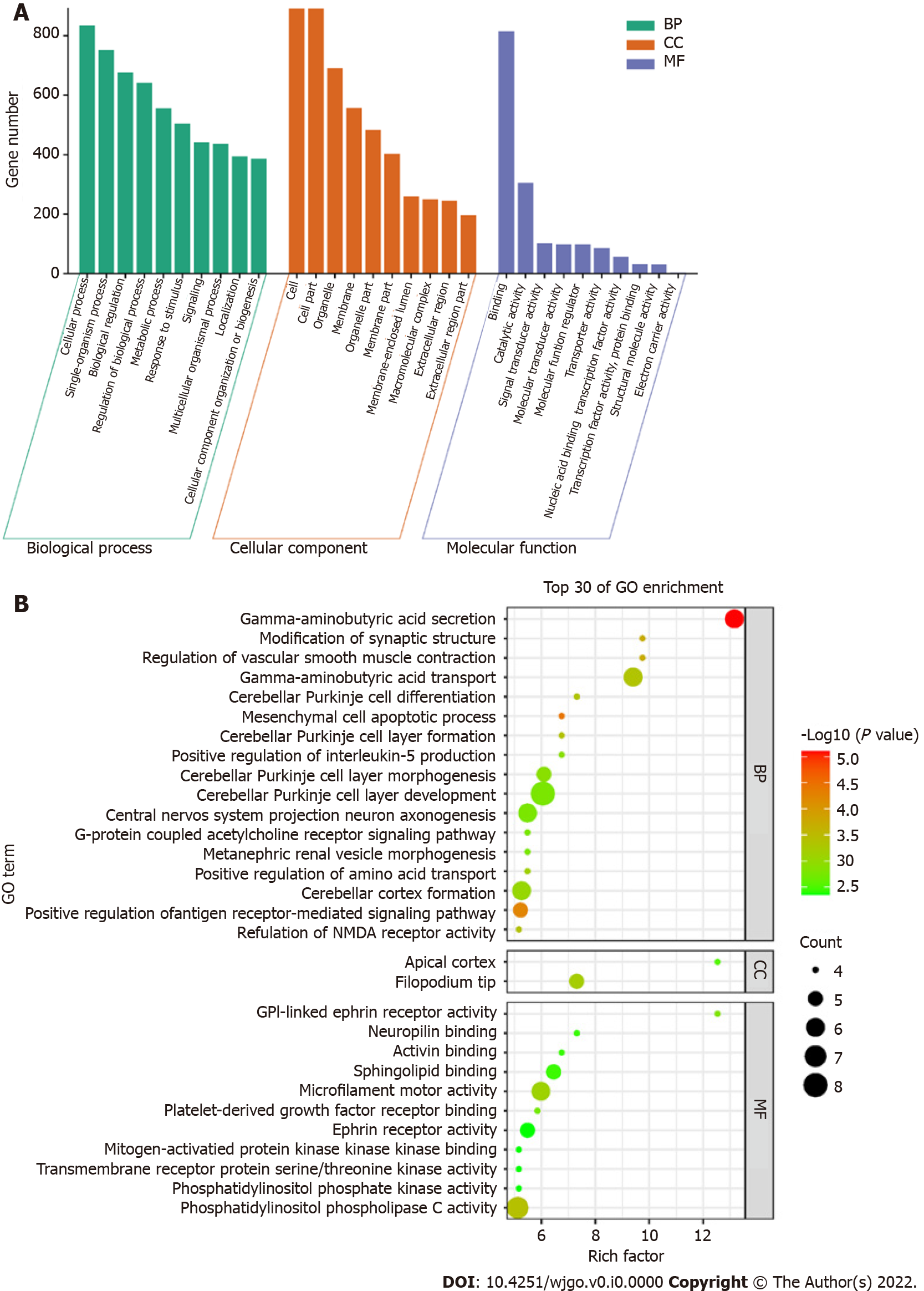
**Figure 1 The heatmap and volcano of the differentially methylated sites.** A: Adenoma group; B: Delta β data. Ca: Cancer group.



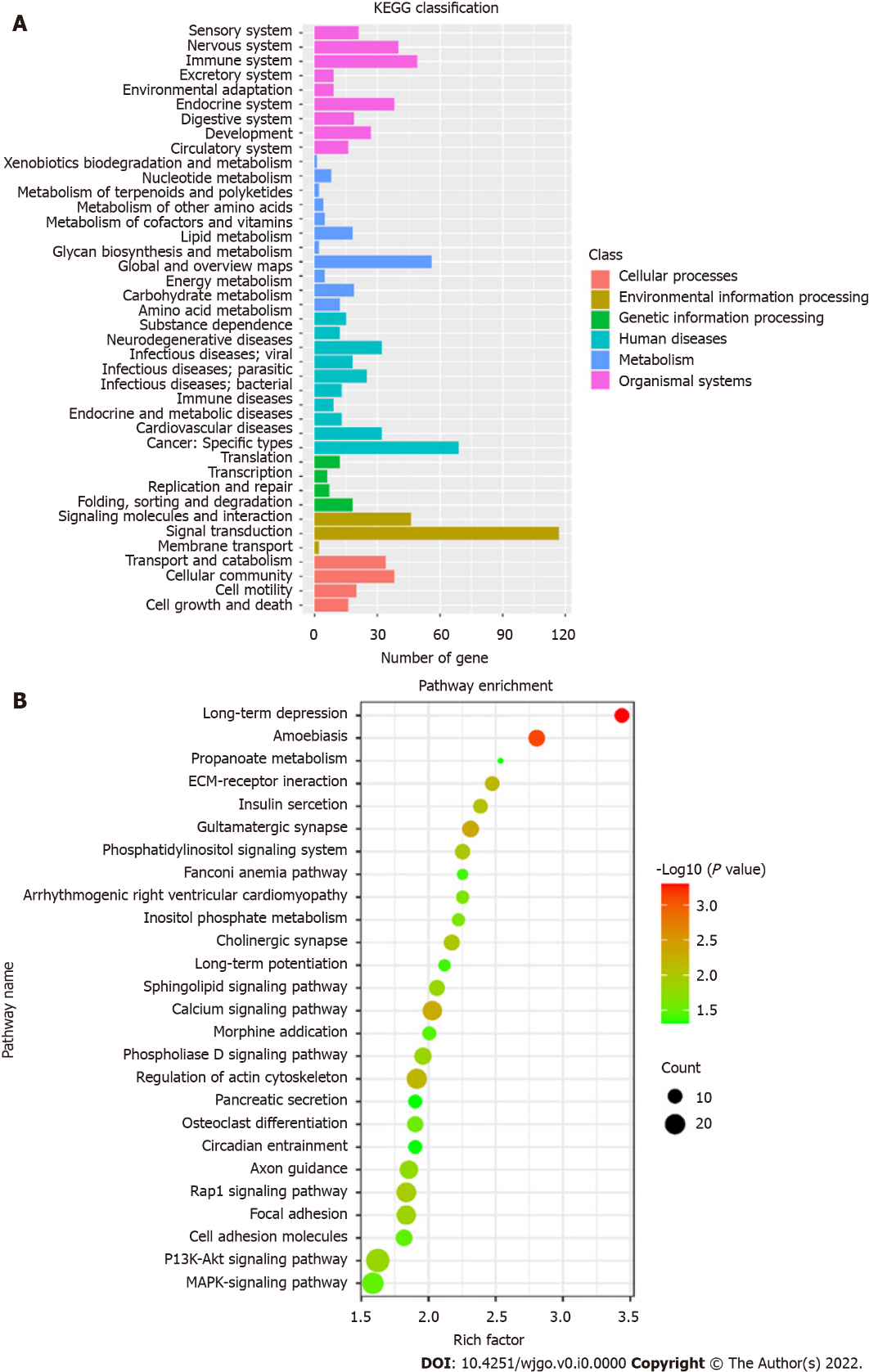
**Figure 2 The distribution of differentially methylated sites.** A: Examples of the distribution of differentially methylated sites (DMSs) on chromosome 1, 2, and 3. The orange and blue dots represent the CpGs which are Δβ > 0 and Δβ < 0 respectively, the green peak represents the density of probes; B: Schematic map of the genomic DMSs, where red and blue represent the CpGs which are Δβ > 0 and Δβ < 0 respectively.



**Figure 3 Distribution of differentially methylated sites.** A: Distribution of differentially methylated sites (DMSs) in gene structural elements; B: Distribution of DMSs in and around CpG island.



**Figure 4 Gene ontology analysis.** A: Gene ontology (GO) function distribution of differentially methylated sites; B: Bubble diagram of GO enrichment results. GO: Gene ontology; BP: Biological process; CC: Cellular component; MF: Molecular function.



**Figure 5 Kyoto Encyclopedia of Genes and Genomes analysis.** A: Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway classification map; B: Bubble diagram of KEGG enrichment pathways. KEGG: Kyoto Encyclopedia of Genes and Genomes.

**Table 1 Patient characteristics**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
|  | **Gender** | **Age** | **Group** | **TNM stage** |
| C1 | Male | 52 | CRC (I) | I |
| C2 | Male | 62 | CRC (I) | I |
| C3 | Female | 62 | CRC (I) | I |
| C4 | Male | 64 | CRC (I) | I |
| C5 | Female | 79 | CRC (I) | I |
| A1 | Female | 69 | Adenoma |  |
| A2 | Male | 69 | Adenoma |  |
| A3 | Male | 69 | Adenoma |  |
| A4 | Male | 49 | Adenoma |  |
| A5 | Male | 74 | Adenoma |  |

CRC: Colorectal cancer; TNM: Tumor-node-metastasis.

**Table 2 The distribution of differentially methylated sites, (*n*)**

|  |  |  |  |
| --- | --- | --- | --- |
| **Chromosome** | **DMSs** | **Hypermethylated** | **Hypomethylated** |
| chr1 | 162 | 62 | 100 |
| chr2 | 158 | 53 | 105 |
| chr3 | 104 | 35 | 69 |
| chr4 | 83 | 22 | 61 |
| chr5 | 112 | 35 | 77 |
| chr6 | 112 | 25 | 87 |
| chr7 | 115 | 55 | 60 |
| chr8 | 89 | 27 | 62 |
| chr9 | 79 | 33 | 46 |
| chr10 | 91 | 29 | 62 |
| chr11 | 104 | 42 | 62 |
| chr12 | 105 | 41 | 64 |
| chr13 | 53 | 18 | 35 |
| chr14 | 50 | 22 | 28 |
| chr15 | 58 | 22 | 36 |
| chr16 | 72 | 34 | 38 |
| chr17 | 76 | 35 | 41 |
| chr18 | 38 | 13 | 25 |
| chr19 | 88 | 33 | 55 |
| chr20 | 60 | 22 | 38 |
| chr21 | 25 | 7 | 18 |
| chr22 | 31 | 11 | 20 |

DMSs: Differentially methylated sites.