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**Deciphering the role of transforming growth factor-beta 1 as a diagnostic-prognostic-therapeutic candidate against hepatocellular carcinoma**

Devan AR *et al*. Deciphering role of TGF-β1 against HCC

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**Abstract**

Transforming growth factor-beta (TGF-β) is a multifunctional cytokine that performs a dual role as a tumor suppressor and tumor promoter during cancer progression. Among different ligands of the TGF-β family, TGF-β1 modulates most of its biological outcomes. Despite the abundant expression of TGF-β1 in the liver, steatosis to hepatocellular carcinoma (HCC) progression triggers elevated TGF-β1 levels, contributing to a poor prognosis and survival. Additionally, elevated TGF-β1 levels in the tumor microenvironment create an immunosuppressive stage *via* various mechanisms. TGF-β1 has a prime role as a diagnostic and prognostic biomarker in HCC. Moreover, TGF-β1 is widely studied as a therapeutic target either as monotherapy or combined with immune checkpoint inhibitors. This review provides clinical relevance and up-to-date information regarding the potential of TGF-β1 in diagnosis, prognosis, and therapy against HCC.

**Key Words:** Transforming growth factor-beta 1; Inflammation; Immunosuppression; Fibrogenesis; Hepatocellular carcinoma; Biomarker; Immunotherapy

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**Core Tip:** Transforming growth factor-beta 1 (TGF-β1) exhibits a progressive elevation throughout hepatic dysfunction starting from hepatitis to hepatocellular carcinoma (HCC) as an inflammatory cytokine, pro-fibrogenic marker, immunosuppressive agent, and pro-carcinogenic growth factor. Aberrant TGF-β1 activation in HCC is associated with a poor prognosis and survival. TGF-β1 mediated immunosuppression disturbs the anticancer surveillance and the efficacy of immunotherapeutic agents. This pleiotropic effect of TGF-β1 in the context of HCC makes it ideal as a diagnostic, prognostic, and therapeutic candidate in HCC.

**INTRODUCTION**

***Hepatocellular carcinoma, an aggressive and refractory cold tumor***

Liver cancer, specifically hepatocellular carcinoma (HCC), is often recognized as an aggressive malignancy, ranking 6th in incidence and 3rd in terms of mortality among all malignancies in 2020, where the mortality rate is roughly equivalent to the incidence rate[1]. As it develops in the background of chronic inflammation starting from the fatty liver, HCC remains undiagnosed for years until it worsens. The progressive transformation from cirrhosis to HCC also creates longer delays in diagnosis[2]. HCC is commonly diagnosed by liver imaging techniques such as ultrasound, computed tomography, and magnetic resonance imaging. Blood biomarkers such as alpha fetoprotein (AFP), protein induced by vitamin K absence or antagonist II levels, and liver biopsy are also used to diagnose HCC[3,4]. Even though the recent guidelines recommend HCC surveillance biannually and antiviral vaccination, the lack of effective surveillance programs significantly contributes to HCC progression to the advanced stage, particularly in high-risk individuals. Once established, HCC cells rapidly proliferate and spread to the extrahepatic site, such as the lungs, portal vein, and lymph nodes. In such cases, in an advanced stage, curative interventions such as liver transplantation, resection, percutaneous ablation, and chemoembolization are not responsive. Systemic drug therapy remains the primary treatment modality[5,6], where immunotherapy and tyrosine kinase inhibitors are the approved treatment options[7]. However, limited response to therapy, the emergence of multidrug resistance, immunosuppressive tumor microenvironment, and lack of validated diagnostic and prognostic biomarkers pose significant obstacles in establishing an effective treatment against HCC[8,9].

Transforming growth factor-beta (TGF-β) is a critical homeostasis regulator, which is aberrantly activated during inflammation, fibrosis, and carcinogenesis[10]. Among the three isoforms, TGF-β1 is superior in TGF-β signal transduction, especially those related to chronic liver diseases. Since HCC is an inflammation-induced immunosuppressed malignancy, the role of TGF-β1 signaling in HCC has been extensively evaluated recently[11]. This review presents the potential of the TGF-β superfamily of ligands, specifically TGF-β1, to develop as a therapeutic and prognostic-diagnostic marker candidate against HCC.

***TGF-β1 signaling in HCC***

The TGF-β superfamily of ligands are dimeric peptide growth factors, with more than 30 members in humans, mainly TGF-βs, activins, inhibins, and bone morphogenetic proteins. TGF-βs are categorized into three different isoforms: TGF-β1, TGF-β2, and TGF-β3. Of the three isoforms, TGF-β1 is the most evaluated and abundant, found in epithelial, endothelial, hematopoietic, and connective tissues. TGF-β2 is expressed in epithelial and neuronal cells, while TGF-β3 is found in mesenchymal cells[12]. TGF-βs are implicated in diverse physiological processes, including cell homeostasis and embryonic development[13]. They are pleiotropic factors that regulate inflammation, fibrogenesis, cell differentiation, proliferation, epithelial-mesenchymal transition (EMT), extracellular matrix (ECM) formation, and elicits tumor-suppressive and pro-tumor effects in a cell-context dependent manner[14].

The three isoforms of TGF-β have structural similarity and functional redundancy[15]. However, TGF-β1 is often considered a potent and superior isoform with significant physiological and pathological importance[16]. Importantly, TGF-β1 exerts a cell-context dependent effect in the liver[17,18]. Normal to activated TGF-β1 signature confers a protective effect by inhibiting hepatocyte proliferation and hepatic stellate cell (HSC) activation, inducing apoptosis, preventing fibrosis, and improving liver function. While, aberrantly activated TGF-β1 signature manifests as HSC activation and worsening fibrosis to HCC, where tumor cells lose their sensitivity toward the inhibitory effect of TGF-β1[19-21]. Exposure of hepatocytes to various causative factors such as viruses, alcohol, toxicants, and other metabolic disorders leads to the release of TGF-β1. Also, other pro-inflammatory cytokines such as tumor necrosis factor-α and growth factors from non-parenchymal liver cells switch on inflammation, production of the ECM, and accumulation of fibrous material that eventually progress to cirrhosis[22,23]. A simultaneous increase in integrins, a class of vital cell adhesion molecules, is observed as the fibrogenesis continues. These integrins interact with TGF-β1 and other ECM proteins, altering signal transduction pathways[24]. Along with the accumulation of genetic mutations, HSCs induce TGF-β and β-catenin-dependent EMT, leading to tumor growth in the liver. TGF-β1 continues to increase, promoting neo-angiogenesis by interacting with other pathways and mediating stromal-tumor cell interaction, conferring aggressive phenotype and metastasis[25] (Figure 1). Wang *et al*[26] demonstrated that both TGF-β1 and TGFβR1 have a crucial role in regulating proliferation, invasion, metastasis, and immune response in HCC cells.

TGF-β1 exerts biological and pathological effects *via* Smad and non-Smad pathways. TGF-βs are synthesized in the inactivated form and exist as a latent TGF-β complex (LTC) by binding with latency-associated protein. Later, the LTC is converted to a large latent complex (LLC) by interacting with latent TGF-β binding protein in the ECM. Integrin signaling plays a significant role in the activation and subsequent release of TGF-β1 from the LLC. It is also mediated by other factors such as pH and protease enzyme[27]. In the canonical Smad pathway, activated TGF-β first binds with the extracellular domain of TGF-β receptor type II, which triggers the cross phosphorylation of the kinase domain of TGF-β receptor type I. TGF-β R1 activation leads to the phosphorylation of Smad proteins, Smad 2 and Smad 3. Later, Smad-2 and 3 complex binds with the co-Smad Smad-4 to form a ternary complex. This ternary complex is then translocated into the nucleus, binds to Smad binding elements in DNA, and activates the transcription of TGF-β-dependent genes[28]. Binding of the inhibitory Smad, Smad-7 will shut down the activated pathway[29]. In addition to the canonical Smad pathway, TGF-β can exert biological functions by activating other diverse signaling molecules such as P38, JNK, PI3K/AKT, RAS-ERK, and RHO-ROCK, which constitute the non-Smad pathway of TGF-β signal transduction[30] (Figure 2).

The Smad pathway of TGF-β signal transduction also enhances the transcription of FoxP3, predominantly present in T regulatory (Treg) cells[31,32]. A high amount of tumor-infiltrating Treg cells and FoxP3 positive Treg cells in blood is reported in HCC patients, leading to the deterioration of effector T cells such as CD4+ and CD+ cytotoxic T lymphocytes, which are pillars of anticancer immunity[33,34]. Together, TGF-β inhibits natural killer (NK) cells, blocks interferon (IFN)-γ secretion, and prevents effector immune cells recruitment to tumor tissue[35,36]. Additionally, TGF-β inhibits IFN-γ secretion by interacting with the activating transcription factor 1[37]. Likewise, TGF-β-RUNX3 transcription factor interaction and co-expression of programmed death ligand 1 (PD-L1) and interleukin-10 promote the transformation of naïve B cells to immunoglobulin A producing B cells, which are crucial in HCC development from non-alcoholic fatty liver[38]. Elevated TGF-β can directly enhance the transcription of PD-1 in HCC. The interaction of PD-1 with PD-L1 causes significant immunosuppression by T cell exhaustion, which manifests as inhibition of T cell activation, proliferation, and cytotoxic action[39,40]. In a recent study, Bao *et al*[41] reported that TGF-β1 triggers the expression of immune checkpoints such as PD-1 and CTLA4 on HCC cells and attenuates T-cell-mediated anti-tumor immune surveillance. Therefore, up-regulated TGF-β, mainly isoform TGF-β 1, directly affects immune checkpoint inhibition, and it works as an indicator of T cell exhaustion. This evidence suggests the potential of TGF-β1 targeted immunotherapies against HCC. The pivotal role of the TGF-β1 signature in hepatic dysfunction and HCC extends its potential as a biomarker molecule for diagnosis and prognostic prediction and a therapeutic target.

***Clinical utility of TGF-β1 as a diagnostic marker of HCC***

As an inflammatory-fibrogenic cytokine molecule, the involvement of TGF-β1 in all stages of liver injury, starting from fatty liver, steatosis, and fibrosis to cirrhosis and HCC, is evident. Intergromic analysis of *TGF-β* gene alterations among the 33 cancer types in the TGCA dataset revealed 39% alterations. Gastrointestinal cancers and HCC exhibited prominent mutations compared with other cancer types[42]. Later, in an HCC-specific transcriptomic analysis, 40% of HCC samples were found with mutations in genes of the TGF-β pathway[43]. Higher TGF-β1 levels in HCC correlate with a high rate of extrahepatic metastasis (EHM), poor prognosis, and low survival rate[44]. After acute/chronic liver injury, liver sinusoidal endothelial cells and HSCs secrete TGF-β1 and up-regulate TGF-β receptors[45,46]. Elevated TGF-β1 level was found in viral and alcohol-induced fibrosis[47]. Thus, TGF-β levels can be used to track the response to therapy, and the decrease in TGF-β 1 level followed by IFN treatment in hepatitis B virus (HBV) patients is associated with improved treatment outcomes. Apart from HSC-triggered TGF-β secretion, hepatitis C virus (HCV) infection can also induce TGF-β1 production in hepatocytes[48]. Likewise, proteomic and phospho-proteomic characterization of 110 tumor and non-tumor tissues of early-stage HBV-associated HCC found increased expression of *TGF-β* genes compared with the non-tumor tissue[49]. However, the dichromatic role of TGF-β1 on cancer growth, *i.e.,* tumor suppressive in early stage or oncogenic effect in late stage, is a matter of concern. Another study indicated a comparative functional genomic approach and illustrated the link between TGF-β expression signature and HCC subtypes. The study showed that TGF-β positive HCC clusters can be categorized into two. HCC with early TGF-β signature exhibit physiological responses while HCC associated with late TGF-β signature showed metastasis and poor survival[50]. N-2-fluorenylacetamide-induced rat hepatoma model was used to investigate the association of TGF-β1 expression with different stages of hepatocarcinogenesis and a progressive elevation of hepatic TGF-β1 and *TGF-β1* mRNA was found during the transformation of hepatocytes to malignant cells[51]. Another study indicated that elevated plasma TGF-β1 level was found in 89.5% of HCC patients, and interestingly, among these patients, 93.3% had an AFP level less than 400 μg/L[52]. This suggested that TGF-β1 expression can be a more accurate and sensitive biomarker for early diagnosis of HCC for monitoring the disease progression.

The diagnostic importance of TGF-β1 was established significantly earlier itself. In 1997, Tsai *et al*[53] investigated the correlation of urine TGF-β1 level with HCC. They found a significant increase in TGF-β1 level in HCC patients compared with the healthy control group and cirrhotic chronic hepatitis patients. They also reported the association of TGF-β 1 levels with a poor prognosis and shorter survival[53]. Later, the same team compared urinary TGF-β1 with another important tumor marker, AFP, and found that disease progression from cirrhosis to HCC is characterized by a typical elevation in both urinary TGF-β1 and serum AFP with a diagnostic accuracy of about 90%[54,55]. These collective data suggest the potential of TGF-β1 to be used along with AFP as a complementary tumor marker to differentiate HCC from cirrhosis correctly. Another study investigated the rationale for parallel determination of TGF-β1 and AFP to diagnose HCC. They found that the TGF-β1 level exhibited a stage-dependent increase in all liver diseases where AFP showed an HCC-specific elevation[56]. As TGF-β1 estimation tracks the disease progression pattern, TGF-β can be considered a more sensitive diagnostic marker of HCC. Its specificity is higher when it is analyzed along with AFP. To diagnose and select patients for galunisertib (TGF-β inhibitor) therapy, Cao Y *et al*[57] in 2017 performed next-generation sequencing-based analysis in HCC samples and found that the mRNA levels of *TGF-β1* along with *SKIL* and *PMEPA1* could be better diagnostic markers as well as makers to select patients who are more likely to respond to galunisertib.

Another study investigated the association of serum TGF-β1 with disease severity in HCC using 180 subjects in different stages of HCC. A group of cirrhotic patients, as well as healthy controls, were also maintained. Consistent with the previous reports, this study also found a significant increase in TGF-β1 level in HCC (1687.47 ± 1462.81 pg/mL) as compared with cirrhotic patients (487.98 ± 344.23 pg/mL) and controls (250.16 ± 284.16 pg/mL). Additionally, the serum level of TGF-β1 showed an exponential elevation as the disease progressed from early to advanced, *i.e.,* during progression from Barcelona Clinic Liver Cancer stage A to D, TGF-β 1 level increased from 652.83 to 1668.78 pg/mL[56]. The best cut-off value of TGF-β1 detection was determined as 301.9 pg/mL, comparable with the value (370 pg/mL) reported by Shehata *et al*[58] (Table 1).

Background inflammation and indolent transformation are the critical factors that create a waiting time paradox in diagnosing HCC, making the tumor more aggressive and refractory. Since TGF-β, specifically TGF-β1, plays an essential function from the initial hepatic injury to hepatocarcinogenesis, it holds immense potential to be validated as a diagnostic marker of HCC. Although the dual functioning of TGF-β1 is still debatable, the diagnostic relevance of TGF-β1 is well evident, and thus, it warrants further investigations and clinical validation.

***Clinical utility of TGF-β1 as a prognostic marker of HCC***

Poor prognostic characteristics of HCC contribute to late detection, aggressiveness, and failure of therapeutic interventions[70]. Molecular pathways of hepato-carcinogenesis are still confusing because of the involvement of diverse molecular pathways, genetic alterations, and evolution of malignant cells. Thus, this ultimately results in the worst prognosis within the early stage itself[71]. The expression of TGF-β1 is remarkably increased at the advanced stages of HCC and is involved in initiating EMT, regulating tumor proliferation, and promoting immunosuppressive tumor microenvironment during HCC progression under the challenges like liver cirrhosis and HBV and HCV infections. This warrants screening TGF-β1 levels from the early stages of HCC as a tool for evaluating the clinical outcomes. Depending upon the expression profile of TGF-β1, it is effortless to estimate the clinical impact of therapeutic strategies[72].

A research study conducted by Giannelli *et al*[73] proposed that TGF-β1 promotes EMT by stimulating homologous proteins like snail and slug. Secretion of TGF-β1 by HCC invasive cell lines, especially cell lines with α3β1-integrin expression, is significantly higher than in non-invasive and cirrhotic cell lines. The patients at the initial and advanced stages of HCC with a higher profile of TGF-β1 possess a poor prognostic ratio with lower overall survival (OS) and disease-free survival (DFS) rates[73,74]. Likewise, another notable experimental study by Lee *et al*[75] demonstrated that plasma TGF-β1 is positively correlated with critical conditions like EHM, portal vein thrombosis, and regional lymph node involvement. Statistical studies involving the detailed examination of overall and cumulative survival rates of HCC patients showed that candidates with abundant levels of plasma TGF-β1 manifested remarkably lower survival rates than the candidates with lower expression of TGF-β1. This evidence points to the usefulness of TGF-β1 as a prognostic marker in HCC.

Wang Y *et al*[76] elucidated the crucial involvement of TGF-β1 in tumor progression. A total of 180 patients with HCC were selected for the study, out of which 105 were found with solid expression of TGF-β1. This study showed a positive correlation between TGF-β1 and Treg cells. The increased secretion of TGF-β1 at the starting stage of HCC indicates that the tumor may be one of the most critical sources of TGF-β1 in HCC patients. Earlier studies also provided evidence that TGF-β1 promotes the regulatory phenotype and modulates the biological functions of Tregs. By Kaplan-Meier evaluation, it was found that HCC patients overexpressing TGF-β1 in neoplastic tissues had a considerably shorter OS and greater recurrence rate than patients with lower expression[76]. A meta-analysis study conducted by Peng *et al*[77] reported that TGF-β1 implements an unfavorable prognosis on OS rates of HCC patients with a hazard ratio of about 1.71 and 2.29 from both univariate and multivariate analyses. Additionally, the study indicated the worst prognosis of TGF-β1 upon DFS, relapse-free survival, and progression-free survival of 1422 patients *via* COX univariate analysis with a hazard ratio of about 1.60. In summary, the results from these studies draw out the negative prognostic impact of high TGF-β1 expression on the OS in HCC patients.

TGF-β1 possesses a dual functional role in malignancy; initially, it acts by blocking epidermal growth and promoting tumor suppression, but in later stages, it appears to be involved in the up-gradation of advanced tumors[78]. Embryonic liver fodrin (ELF), a novel form of β-spectrin, is involved as a Smad3/4 adaptor in the TGF-β mediated tumor suppression signaling pathway. Miscolocalization of Smad3 and Smad4 caused by the dysregulation of ELF resulted in the disruption of TGF-β signaling pathways[79]. A research study conducted by Ji *et al*[80] investigated the predictive value of both TGF-β1 and ELF in HCC patients after hepatic resection. The expression of TGF-β1 is significantly higher in HCC tissues than in normal liver tissues, while the incidence of ELF is higher in normal liver tissue in contrast with HCC samples. The reports of postoperative survival rates of HCC patients with lower expression of TGF-β1 showed that DFS and OS rates of HCC patients at 1 (79.4%), 3 (73.5%), and 5 (62.0%) years were significantly higher than those of the patients with higher expression of TGF-β1 (28.0%, 12.0%, and 12.0%, respectively). The study also showed a negative correlation between TGF-β1 and ELF levels. The study also indicated that DFS rates of HCC patients with higher expression of ELF and lower TGF-β1 levels are remarkably more elevated than the HCC patients with low expression of ELF and higher TGF-β1 levels at 1, 3, and 5 years (75.0%, 60.0%, and 57.5% *vs* 25.0%, 15.9%, and 10.2%, respectively; *P <* 0.001). Data from clinicopathological examination exhibited that TGF-β1 positively relates with hepatitis B surface antigen, tumor size, tumor number, TNM, and recurrence, while ELF is negatively correlated with all metastatic characteristics, suggesting that ELF is associated with tumor suppressing features. This research study indicated that both TGF-β1 and ELF can be included in the category of relevant biomarkers as prognostic agents for evaluating clinical results after hepatic resection[80].

An experimental approach described the correlation and the possibility of fibroblast growth factor (FGF) receptor 4 (FGFR4) and TGF-β1 as prognostic biomarkers in HCC. FGFR4 is the most predominant isoform of the FGFR family and is actively involved in various biological activities, including metastasis, differentiation, embryonic development, proliferation, apoptosis, and angiogenesis[81]. Multiple studies showed that FGFR4 plays a clear-cut role in the pathogenesis of HCC and the up-regulation of FGFR4 possesses resistance to various targeted therapies[82]. A clinicopathological examination conducted by Chen *et al*[83] showed that elevated expression of both TGF-β1 and FGFR4 enhances tumors’ invasiveness and metastatic nature. Clinicopathologic characteristics revealed that HCC patients at advanced stages with high TGF-β1 and FGFR4 expression were more likely to be at a higher TNM stage. Statistical data showed that the OS of patients at 5 years is about 8.5%, and the median survival duration is 32.3 mo in case of TGF-β1 positive cases. In contrast, in TGF-β1 negative cases, the OS is about 45.6% and the median survival rate is 50.4 mo. Candidates with high TGF-β1 expression had a lower OS rate in contrast to those with negative TGF-β1 expression profiles. Likewise, in cases of high levels of FGFR4, the OS rate of patients is very low, that is, the 5-year survival rate is only about 8.3%, and median survival rate is 29.4 mo while in the condition of impeded FGFR4 expression, the 5-year survival rate is about 70.1% and the median survival period is 51.2 mo. This study showed a positive correlation between TGF-β1 and FGFR4 as prognostic markers in HCC. Additionally, the results from univariate and multivariate analyses showed that both TGF-β1 and FGFR4 are independent and reliable prognostic factors in HCC for evaluating the therapeutic response in HCC patients, especially after postoperative procedures[83]. The strong correlation between TGF-β1 expression and survival rates of HCC patients suggests its potential as a prognostic biomarker for HCC (Table 2). In addition to the role of TGF-β1 as an effective prognostic marker, it can also be used for targeted therapeutic strategies.

***Clinical utility of TGF-β1 as a therapeutic target in HCC***

The tyrosine kinase inhibitor Sorafenib was the first approved first-line therapy for advanced HCC. Sorafenib was the first-line therapy for 10 years until another tyrosine kinase inhibitor, Levatinib, was approved in 2018. Even though tyrosine kinase inhibitors dominated HCC therapy as first-line or second-line options, the efficacy was only modest, with limited treatment outcome, the emergence of drug resistance, and also relapse[85]. Recently, treatment strategies adopted a paradigm shift to immunotherapeutic approaches because of the importance of the immune microenvironment in carcinogenesis.

The liver is the most extensive reticuloendothelial system and peripheral immunomodulatory organ in the human body. Immunotherapy is being extensively evaluated in liver cancer[86]. The liver constitutes a vast repository of immune cells including NK cells, kupffer cells, sinusoidal endothelial cells, and innate T cells[87]. Aberrant immune checkpoint activation makes HCC a cold tumor, where anti-tumor immune surveillance is completely abolished[88]. The combination of immune checkpoint inhibitors (ICIs) such as Nivolumab (PD-1 inhibitor) + Ipilimumab (CTLA4 inhibitor) and Atezolizumab (PD-L1 inhibitor) + Bevacizumab [vascular endothelial growth factor (VEGF) inhibitor] got approval as first-line therapy for various cancers such as non-small cell lung cancer (NSCLC) and HCC in 2020[89]. Alhough ICIs exerted a superior effect to tyrosine inhibitors, current immunotherapeutic drugs failed to establish an effective anticancer immunity against HCC. Although the immunotherapeutic approaches modify effector immune cell functions to elicit anti-tumor immune response, the immunosuppressive tumor microenvironment neutralizes the effects of immunotherapy[90].

Analysis of the TGF-β profile of HCC samples in the TGCA data set revealed four categories with typical TGF-β expression[91]. The cluster with a highly activated TGF-β signature, which accounts for 14.5% of samples, exhibited prominent immune exhaustion and a poor prognosis. ICIs may not work well in this cluster. Additionally, anti-inflammatory/anti-fibrotic agents targeting TGF-β can improve the immune milieu[92,93]. The majority of HCC samples belong to a cluster of activated TGF-β signature (45%) and showed a low level of an immune response. Hence, combining a TGF-β inhibitor with an immune checkpoint inhibitor can exert a synergistic effect. Thirty percent of HCC samples showed a normal TGF-β signature associated with active immune surveillance; therefore, immunotherapy will be most suitable for this cluster. The fourth cluster is a minor population (9.9%) that exhibits an inactivated TGF-β signature with poor immune cell activation and response[94]. With this evidence, it is clear that TGF-β1 signature can be used to decide the suitable therapy or predict the outcome of immunotherapy.

Several specific and non-specific inhibitors of TGF-β, mainly TGF-β1 and 2 inhibitors, are being developed and evaluated against various tumors, including HCC. Regarding non-specific inhibitors, as the primary molecular target is different, their ability to block the TGF-β pathway offers additional benefits as anticancer agents. One such example is Halofuginone, an alkaloid coccidiostat with reported preclinical activity against HCC. In addition to the prominent inhibition of collagen synthesis, Halofuginone also blocks TGF-β1, inhibits ECM formation and fibroblast proliferation, and increases IFN-γ and anti-tumor immune response[95,96]. The effect of Halofuginone against advanced progressive solid tumor has been evaluated in phase I clinical trial (NCT00027677), and in 2000, the United States Food and Drug Administration gave orphan drug approval status to Halofuginone for treatment of scleroderma[97]. Histone deacetylase exerts epigenetic regulation of TGF-β1 mediated fibrosis and carcinogenesis[98]. Studies indicate that histone deacetylase inhibitors such as Panobinostat have shown effectiveness in HCC animal models and phase I human trials when combined with sorafenib (NCT00823290)[99]. Apart from the non-specific inhibitors, recent preclinical interventions combined TGF-β1 targeting antibodies or TGF-βR1 inhibitor with PD-L1 inhibitors and obtained prominent cytotoxic effect and anticancer immune surveillance in various solid tumors[100,101]. Likewise, M7824 is a bifunctional fusion protein with dual targeting of PD-L1 and TGF-β, which has been evaluated in animal models of various cancers either alone or in combination with vaccines[102]. M7824 exerted a significant inhibitory effect on TGF-β1[103]. Among the different TGF-β inhibitors, LY2109761 is an orally bioavailable TGF-β receptor type I inhibitor, which exhibited an antitumor effect in various HCC animal models[104]. LY2109761 inhibited TGF-β1 induced migration, invasion, and anoikis in HCC cells[105,106]. Another study suggested the anti-angiogenic potential of LY2109761, which was superior to the typical VEGF inhibitor Bevacizumab, and the effect was mediated by suppression of VEGF through inhibition of Smad dependent TGF-β1 signaling[107,108]. In addition to this, anti-TGF-β agents targeting other isoforms are also developed. AP-12009 is a TGF-β2 specific antisense oligonucleotide in a clinical trial to treat glioma and anaplastic astrocytoma[109,110]. Likewise, TGF-β1 directed mRNA was developed as AP-11011, which was evaluated against NSCLC and colon cancer in preclinical models[111]. Lordelimumab is a TGF-β2 specific monoclonal antibody with an anti-fibrotic effect[112]. Many preclinical studies investigated different TGF-β inhibitors, yet, Galunisertib (LY2157299), a kinase inhibitor of TGF-β1, is only the one in current clinical trials[113]. Several clinical trials of Galunisertib are ongoing or have been completed either alone or combined with Sorafenib, ICIs, and alkylating agents (Table 3).

**CONCLUSION**

TGF-β1 exerts a unique regulatory power on inflammation, fibrogenesis, and immune response in HCC. Among other TGF-β isoforms, significant and progressive expression of TGF-β1 during the entire course of HCC pathogenesis, starting from chronic hepatitis to HCC, makes it a sensitive and accurate diagnostic marker of HCC. The specificity and sensitivity of TGF-β1 based diagnosis of HCC can be improved by parallel estimation of serum AFP. Even after establishing HCC, TGF-β1 continues to elevate as HCC progresses and is associated with a poor prognosis and shorter survival. Since TGF-β1 is the master regulator of the immunosuppressive tumor milieu in HCC, TGF-β1 inhibition could sensitize ICI, tyrosine kinase inhibitors, and other systemic or curative interventions. HCC remains the deadliest refractory tumor predominantly due to its delayed diagnosis. In that context, TGF-β1 is relevant for early diagnosis, prognosis, and therapy. Even though a plethora of supporting evidence is available, still TGF-β1 is not much studied and evaluated compared with other markers such as AFP. Notably, the dichotomic nature of TGF-β signaling in HCC needs to be defined accurately to establish the clinical utility of TGF-β1. Thus, proper and careful determination of the TGF-β1 profile of patients is necessary to choose the suitable patients for TGF-β1 targeted therapy.

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**Figure Legends**

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**Figure 1 Involvement of transforming growth factor-beta 1 in various stages of liver dysfunction.** Transforming growth factor-beta (TGF-β) regulates cell homeostasis in normal physiological conditions. As an inflammatory-fibrogenic cytokine molecule, the involvement of TGF-β1 in all phases of liver injury, from fatty liver, steatosis, and fibrosis to cirrhosis and hepatocellular carcinoma, is evident. Causative agents like viral infection, alcohol, and co-morbidities like diabetes and obesity trigger the release of a pro-inflammatory cytokine such as TGF-β1, which stimulates inflammation, extracellular matrix (ECM) production, and accumulation of fibrous material that eventually progress to cirrhosis. As the fibrosis continues, the interaction of overexpressed TGF-β1 with integrins and other ECM proteins can alter signaling, accumulate gene mutations, and induce epithelial-mesenchymal transition and hepatocarcinogenesis. TGF: Transforming growth factor; ECM: Extracellular matrix; HCC: Hepatocellular carcinoma.

**图示

描述已自动生成**

**Figure 2 Transforming growth factor-beta 1 signaling pathway.** Smad and non-Smad pathways mainly regulate transforming growth factor-beta 1 (TGF-β1) signal transduction. Integrin signaling triggers the activation and release of TGF-β1 from the LCC (large latent complex). Activated TGF-β1 binds TGF-β type II receptor, leading to phosphorylation of TGF-β type I receptor and associated Smad proteins, mainly Smad-2 and Smad-3. Phosphorylated Smad-2 and 3, complex with the co-Smad Smad-7, form a ternary complex. This ternary complex then translocates into the nucleus, binds to Smad binding elements, and regulates the transcription of TGF-β-related genes. TGF: Transforming growth factor; LAP: Latency associated protein; LTAB: Latent transforming growth factor-beta binding protein; SBE: Smad binding elements; ECM: Extracellular matrix; mTOR: Mammalian target of Rapamycin; PI3K: Phosphoinositide 3-kinase; AKT: AKT serine/threonine kinase 1; MKK: Mitogen-activated protein kinase kinase.; JNK: C-Jun N-terminal kinase.

**Table 1 Clinical utility of** **transforming growth factor-beta 1 as a diagnostic marker against** **hepatocellular carcinoma**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Ref.** | **Sample size HCC/control** | **Assay type** | **TGF-β1 level** | **Sample type** | **Outcome of the study** |
| [60,61] | 26/20 | ELISA | Control: 1.4 ± 0.8 ng/mL | Plasma | TGF-β1 level showed a progressive elevation from cirrhotic to HCC patients to normal subjects. No significant association was found between plasma TGF-β1 and serum AFP levels |
| HCC: 19.3 ± 1.95 ng/mL (*P* < 0.05) |
| [62,63] | 70 | ELISA | Control: 2.7 ± 0.7 ng/mL | Plasma | Elevated plasma TGF-β1 levels in HCC patients are associated with increased tumor size, overexpression of tissue inhibitor of metalloproteinase-1, and tumor severity |
| HCC: 7.3 ± 4.3 ng/mL (*P* < 0.05) |
| [54,55] | 94/50 | 125I-Radio Immuno Assay Kit | Control: 1.5-33.6 μg/g creatinine |  | Urinary TGF-β1 and serum AFP levels were higher in HCC than in cirrhotic patients. The study suggested that both TGF-β and AFP can be used as complementary biomarkers to distinguish between HCC and cirrhosis |
| Cirrhotic: 4.3-52.5 μg/g creatinine |
| HCC: 3.5-184 μg/g creatinine (*P* < 0.0001) |
| [64] | 54/30 | ELISA | TGF-β1 score | Serum | The study team calculated the serum concentration score based on the cut-off limit of 74 pg/mL and 637 pg/mL for TGF-β1 and sFas, respectively. TGF-β1 levels were higher than the cut off value in 23% of HCC patients with negative AFP values, suggesting its diagnostic potential in AFP negative HCC |
| Control: 0.6 ± 0.2 |
| HCC: 1.6 ± 0.5 |
| [65] | 38/23 | ELISA | Control: 300 pg/mL | Plasma | Elevated plasma TGF-β1 level can be a useful diagnostic marker in detecting small HCC, with higher sensitivity than AFP |
| HCC: 954.9 pg/mL (*P* < 0.0001) |
| [66] | 70/32 | ELISA | Control: 2 ng/mL | Plasma | Higher circulating TGF-β1 in HCC patients is associated with suppression of anti-tumor immunity and disease progression |
| HCC: 7.5 ng/mL (*P* < 0.0001) |
| [52] | 50/30 | ELISA | Control: 0.67 ± 0.1 μg/mL | Serum | Aberrant TGF-β1 expression in HCC is associated with differentiation and worsening of HBV infection |
| HCC: 2.21 ± 1.1 μg/mL (sensitivity = 89.5%, specificity = 94%) |
| RT-PCR | Overexpression of *TGF-β1* mRNA in HCC patients, *P* < 0.0001 | Circulating TGF-β1 level and *TGF-β1* mRNA expression can be used as sensitive biomarkers for diagnosing HBV induced HCC |
| [56] | 23/40 | ELISA | Control: 14.35 ± 8.76 ng/mL | Serum | TGF-β1 is a sensitive diagnostic marker for HCC than AFP. Specificity can be increased with combined evaluation of TGF-β1 and AFP levels |
| HCC: 64.35 ± 33.68 ng/mL (*P* < 0.05) |
| [67] | 54/30 | ELISA | Control: 39.5 ± 9.8 pg/mL | Serum | The study suggested elevated TGF-β1 and EGFR levels as reliable diagnostic markers for HCC induced, AFP negative HCC |
| HCC: 1194 ± 331 pg/mL (*P* < 0.0001) |
| [68] | 120/30 | ELISA | Control: 250.16 ± 284.61 pg/mL | Serum | TGF-β1 showed progressive elevation during various stages of liver dysfunction. Higher TGF-β1 level in HCC is associated with tumor grade, pathological stage, and invasiveness |
| Cirrhotic: 487.98 ± 344.23 pg/mL |
| HCC: 1687.47 ± 1642 pg/mL (*P* < 0.0001) |
| [69] | 100/36 | ELISA | Control: 57.29 ± 11.70 ng/mL | Serum | Serum levels of TGF-β were significantly higher in HCC patients than in normal controls |
| HCC: 225.82 ± 48.93 ng/mL (*P* < 0.0001) |

ELISA: Enzyme-linked immunosorbent assay; RT-PCR: Reverse transcription-polymerase chain reaction; HCC: Hepatocellular carcinoma; TGF-β: Transforming growth factor-beta; AFP: Alpha fetoprotein; HBV: Hepatitis B virus; EGFR: Epidermal growth factor receptor.

**Table 2 Clinical utility of transforming growth factor-beta 1 as a prognostic marker against hepatocellular carcinoma**

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| **Ref.** | **Sample size HCC/control** | **Sample and assay type** | **TGF-β1 level** | | **Survival rate (%), (patients with higher TGF-β1 *vs* patients with lower TGF-β1)** | | **Outcome of the study** |
| [75] | 571/551 | Plasma | Control: 3.58 ± 0.17 log10 pg/mL | | 1 yr survival (47 *vs* 60) | 3 yr survival (28 *vs* 36) *P* < 0.05 | Plasma TGF-β1 levels showed a positive correlation with tumor size, invasion, and extrahepatic metastasis and inversely correlated with survival rates in HCC patients |
| ELISA | Cirrhotic: 3.20 ± 0.37 log10 pg/mL | |
| HCC: 3.83 ± 0.31 log10 pg/mL | |
| [83] | 126 | Tumor tissue | 84% of samples (106/126) showed high intra-tumoral TGF-β1 expression | | 5 yr survival (8.5 *vs* 45.6) | | TGF-β1 and FGFR4 were positively correlated in HCC tumor tissues and showed a significant association with lower survival rates in patients |
| Immunohistochemistry | 64.3% samples (81/126) showed high peri-tumoral TGF-β1 expression | |
| [80] | 84/20 | Tumor tissue | TGF-β1 overexpression found in 59.5% of samples (50/84) than that of normal liver tissue | | 1 yr survival (28 *vs* 79.4) | 5 yr survival (12 *vs* 62.6) | TGF-β1 expression was dominant, whereas ELF expression was suppressed in HCC tissues |
| Immunohistochemistry | Patients with high TGF-β1 and lower ELF expression are associated with a poor overall survival and postoperative disease free survival compared with low TGF-β1 and high ELF group |
| [76] | 184/30 | Plasma and tumor tissue | Elevated plasma TGF-β1 level | | 2 yr survival ( 51 *vs* 77) | 3 yr survival (4 *vs* 68), *P* < 0.05 | Higher TGF-β1 expression in tumor tissues triggers Treg cell mediated immunosuppression in tumor microenvironment and contributes to a poor prognosis in HCC |
| ELISA and immunohistochemistry | TGF-β1 was strongly stained in tumor tissue | |
| [84] | 40 | Serum | Before RFA: 63.22 ± 23.61 ng/mL | After RFA: 56.33 ± 24.24 ng/mL | NA | | Radiofrequency ablation lowered TGF-β1 and AFP-L3% expression in HCC patients |
| ELISA | Low TGF-β1 and AFP-L3% levels were observed in the no recurrence group, suggesting its potential as prognostic markers for HCC |

ELISA: Enzyme-linked immunosorbent assay; HCC: Hepatocellular carcinoma; TGF-β: Transforming growth factor-beta; AFP: Alpha fetoprotein; ELF: Embryonic liver fodrin; FGFR: Fibroblast growth factor receptor 4.

**Table 3 Clinical trials of transforming growth factor-beta 1 blockade with Galunisertib in hepatocellular carcinoma and other cancers**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Drug** | **Title of the study** | **Treatment** | **Phase** | **Status** | **Trial ID** |
| Galunisertib | A study of Galunisertib on the immune system in participants with cancer | Monotherapy | Phase I | Completed | NCT02304419 |
| Galunisertib | Galunisertib (LY2157299) and stereotactic body radiotherapy in advanced hepatocellular carcinoma | Combination with radiotherapy | Phase I | Completed | NCT02906397 |
| Galunisertib | A study of LY2157299 in participants with unresectable hepatocellular cancer | Combination with Nivolumab | Phase II | Completed | NCT02423343 |
| Galunisertib | A study of LY2157299 in participants with unresectable hepatocellular cancer | Combination with Sorafenib | Phase I | Completed | NCT02240433 |
| Galunisertib | A study of LY2157299 in participants with advanced hepatocellular carcinoma | Combination with Sorafenib | Phase II | Completed | NCT02178358 |
| Galunisertib | A study of LY2157299 in participants with hepatocellular carcinoma | Combination with Sorafenib/Ramucirumumab | Phase II | Completed | NCT01246986 |
| Galunisertib | Galunisertib and Capecitabine in advanced resistant TGF-beta activated colorectal cancer (EORTC1615) | Combination with Capecitabine | Phase II | Withdrawn | NCT03470350 |
| Galunisertib | A study of LY2157299 in participants with pancreatic cancer (advanced or has spread to another part of the body) | Combination with Gemcitabine | Phase I | Completed | NCT02154646 |
| Galunisertib | A study of Galunisertib (LY2157299) and Durvalumab (MEDI4736) in participants with metastatic pancreatic cancer | Combination with Durvalumab | Phase I | Completed | NCT02734160 |

TGF: Transforming growth factor.



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