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**Acute-on-chronic liver failure induced by antiviral therapy for chronic hepatitis C: A case report**

Zhong JL *et al*. Liver failure induced by antiviral therapy

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**Abstract**

BACKGROUND

There have been no reports of acute-on-chronic liver failure (ACLF) during treatment of chronic hepatitis C (CHC) with direct-acting antivirals (DAAs).

CASE SUMMARY

We report a 50-year-old male patient with CHC. The patient sought medical attention from the Department of Infectious Diseases at our hospital due to severe yellowing of the skin and sclera, which developed 3 mo previously and attended two consecutive hospitals without finding the cause of liver damage. It was not until 1 mo ago that he was diagnosed with CHC at our hospital. After discharge, he was treated with DAAs. During treatment, ACLF occurred, and timely measures such as liver protection, enzyme lowering, anti-infective treatment, and suppression of inflammatory storms were implemented to control the condition.

CONCLUSION

DAA drugs significantly improve the cure rate of CHC. However, when patients have factors such as autoimmune attack, coinfection, or unclear hepatitis C virus genotype, close monitoring is required during DAA treatment.

**Key Words:** Chronic hepatitis C; Acute-on-chronic liver failure; Direct acting antivirals; Sofosbuvir-velpatasvir; Case report

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**Core Tip:** The advent of direct-acting antivirals (DAAs) for chronic hepatitis C (CHC) has improved the rate of sustained virology response, resulting in clinical cure of CHC. We report a rare case of CHC where the patient developed acute-on-chronic liver failure during DAA therapy. Based on comprehensive analysis, the genotype of hepatitis C virus in this patient was unclear, and he was in an autoimmune hyperimmune state at the time and was coinfected with bacteria. When CHC is combined with the above conditions, close monitoring should be carried out during treatment to avoid a poor prognosis.

**INTRODUCTION**

Hepatitis C is an infectious disease caused by hepatitis C virus (HCV). HCV exposure can cause acute hepatitis C, which is defined as the 6-mo period after HCV exposure. Patients who fail to spontaneously clear the virus during acute infection develop persistent infection, which can cause liver inflammation and other serious liver damage. Chronic hepatitis C (CHC) occurs in 50%-80% of patients[1], and 5%-30% of CHC patients develop liver cirrhosis, liver failure and even hepatocellular carcinoma within 20-30 years[2]. Therefore, early diagnosis and treatment of CHC are very important. Over the past 10 years, direct-acting antivirals (DAAs) have revolutionized HCV treatment, increasing cure rates from < 50% to > 95%. However, we report a CHC patient in our hospital who developed acute-on-chronic liver failure (ACLF) during DAA treatment.

**CASE PRESENTATION**

***Chief complaints***

A 50-year-old male patient presented with yellow staining of the skin and sclera, poor appetite and fatigue for 1 wk.

***History of present illness***

Symptoms started 1 wk before presentation with yellow staining of the skin and sclera, poor appetite and fatigue.

***History of past illness***

Three months ago, due to yellow staining of the skin and sclera, the patient went to two tertiary hospitals for consecutive visits. Examination showed severe liver damage but the following causes were excluded: Viral hepatitis (negative for hepatitis A, B, C and E); autoimmune liver diseases (negative for autoimmune hepatitis antibody 1, autoimmune hepatitis antibody 2, immunoglobulin quantification and IgG4); [negative for rubella virus, Epstein–Barr virus (EBV), cytomegalovirus (CMV), and herpesvirus]; *Toxoplasma gondii* and hepatolenticular degeneration (negative for ceruloplasmin). Endoscopy showed chronic nonatrophic gastritis. Abdominal ultrasound showed rough echo in liver parenchyma. Upper abdominal magnetic resonance imaging (MRI) showed hepatitis or liver injury, reactive cholecystitis, and splenomegaly. Liver pathological biopsy showed that liver cells were edematous, focal necrosis, scattered lymphocytes, neutrophil infiltration, and chronic inflammatory cell infiltration in the portal area with fibrosis, in line with chronic hepatitis grade 2 and stage 1. Bilirubin level gradually increased after conservative treatment with drugs, and the patient was admitted to our hospital. The admission examination showed that the total bilirubin was increased to 389.5 mmol/L, hepatitis C antibody was weakly positive, and hepatitis C RNA load was 2.281 × 103 IU/mL. The HCV genotype could not be typed due to low viral load. The patient’s other examinations revealed no abnormalities. He was diagnosed with severe CHC and received medication (glycyrrhetinic acid monoamine S 160 mg ivgtt qd, Shuganning injection 10 mL, ursodeoxycholic acid capsule 250 mg po tid) and artificial liver treatment (plasma exchange + double plasma molecular adsorption system) on March 31 and April 2, 2022, respectively. The patient was discharged on April 18, 2022 with improved liver function. After discharge, he was treated with sofosbuvir–velpatasvir 400:100 mg one tablet/d.

***Personal and family history***

The patient had smoked 10 cigarettes/d for > 20 years, and had no history of drinking, drug use, blood transfusion, or family history of CHC.

***Physical examination***

On physical examination, the vital signs were as follows: body temperature, 36.2°C; blood pressure, 127/78 mmHg; heart rate, 88 bpm; respiratory rate, 20 breaths/min. He also had severe yellowing of the whole body skin and sclera. Moist rales were heard in both lungs on auscultation.

***Laboratory examinations***

HCV antibody was still positive, HCV RNA was < 50IU/mL, and the genotype still could not be typed. Liver function parameters were: Alanine aminotransferase (ALT) 499 U/L, aspartate aminotransferase (AST) 935 U/L, AST/ALT 2.01, alkaline phosphatase 316 U/L, γ-glutamyl transferase 203 U/L, total bilirubin 337 μmol/L, direct bilirubin 176.6 μmol/L, total protein 62.7 g/L, albumin 42 g/L, globulin 21 g/L, and prealbumin 67 mg/L. Coagulation function was: International normalized ratio (INR) 1.12 and prothrombin time activity (PTA) 82%. Routine blood analysis showed the following: White blood cells (WBCs) 10.5 × 109/L, neutrophils 5.36 × 109/L, lymphocytes 3.46 × 109/L, hemoglobin 151 g/L, and platelets 504 × 109/L.

***Imaging examinations***

Upper abdominal MRI (plain scan + enhancement + hepatobiliary pancreatic MRI water imaging) showed liver cirrhosis, splenomegaly, portal hypertension (maximum diameter of main portal vein approximately 15 mm) and suspected cholecystitis. Multiple lymph nodes in the abdominal cavity and retroperitoneum were observed. Chest computed tomography showed bilateral lower lobe pneumonia.

***Further diagnostic work-up***

Hepatitis B surface antigen, hepatitis virus A and E antibodies, ceruloplasmin, transferrin saturation, EBV DNA, CMV DNA, a-fetoprotein and thyroid function were all negative. Autoimmune hepatitis antibodies show positivity for anti-nuclear and anti-mitochondrial antibody M2, immunoglobulin quantitative: negative. Samples were sent to Jinyu Medical Test Center to examine the eight items of autoimmune hepatitis antibody, among which anti-mitochondrial subtype-2 antibody was positive. Dynamic monitoring of liver function, coagulation function, and routine blood changes during hospitalization are shown in Table 1.

**FINAL DIAGNOSIS**

Combined with the patient’s medical history, the final diagnosis was: ACLF, CHC and pulmonary infection.

**TREATMENT**

The patient was admitted to the hospital on April 27, 2022. In order to rule out drug factors, sofosbuvir and velpatasvir were discontinued. The patient received a hepatoprotective treatment (magnesium isopyrrhizinate injection 150 mg ivgtt qd, Shuganning injection 10 mL ivgtt qd, ursodeoxycholic acid capsule 250 mg po tid) and anti-infective treatment (ceftazidime 2 g ivgtt q8h). However, on April 30, the patient’s bilirubin continued to rise to 415.7 mmol/L, PTA continued to decrease to 33.1%, and blood cell counts were WBCs 13.91 × 109/L, atypical lymphocytes 6%, neutrophils 9.88 × 109/L, and lymphocytes 2.64 ×109/L. The patient had depression, the gastrointestinal symptoms worsened, and hiccups occurred. ACLF was considered. Therefore, the antibiotics were adjusted to piperacillin–tazobactam sodium 3.75 g ivgtt q8h to continue antibacterial treatment. Aciclovir 0.25 g ivgtt q8h antiviral treatment, and hormones (methylprednisolone 40 mg qd) were given to suppress immunity, over a course of 5 d. Simultaneously, artificial liver (plasma exchange + double plasma molecular adsorption system) adjuvant therapy was administered. On May 5, routine blood examination showed normal results, and EBV and CMV DNA were negative. As the patient's atypical lymphocytes only appeared once, it was considered secondary to immune disorders. Therefore, ganciclovir and piperacillin–tazobactam sodium were discontinued, but hepatoprotective treatment was continued. Re-examination on May 17 showed that liver function parameters were ALT 27 U/L, AST 36 U/L, and total bilirubin 77.4 μmol/L; therefore, the patient was discharged from hospital on May 19, 2022. He continued to take sofosbuvir–velpatasvir 400: 100 mg 1 tablet/d for antiviral treatment.

**OUTCOME AND FOLLOW-UP**

The outpatient department checked that the patient’s liver function was normal on June 23, 2022, and he received antiviral treatment until August 20. Follow-up to March 1, 2023 showed that HCV RNA was consistently below the detection limit, liver function, routine blood examination and a-fetoprotein were normal. Abdominal ultrasound showed that the light spots on the liver had thickened, instantaneous elastic imaging of the liver showed a hardness of 15.7 kPa and fat attenuation of 247 dB/m. Unexpectedly, autoimmune hepatitis and mitochondrial antibodies were negative.

**DISCUSSION**

Sofosbuvir–velpatasvir is a combined oral DAA. Sofosbuvir is a nucleotide analog NS5B polymerase inhibitor that inhibits viral replication by targeting key targets of RNA replication, while velpatasvir is a second-generation NS5A replication complex inhibitor with high antiviral activity against all HCV genotypes[3]. In noncirrhotic patients, the sustained virological response rate (SVR) can reach 95%[4]. Even in patients with HCV-related decompensated cirrhosis, the SVR rate is > 80%[5]. The patient had no previous underlying diseases, and only took sofosbuvir–velpatasvir following the diagnosis of hepatitis C, without drug interaction. There is no pharmacokinetic basis for liver damage due to sofosbuvir–velpatasvir. The occurrence of ACLF during DAA treatment may be related to the following factors.

***Autoimmunity***

The emergence of autoimmune diseases may be related to viral infection, especially chronic viral infection. HCV infection has long been suspected to be associated with the development of autoimmune diseases, as demonstrated by cryoglobulinemia[6], and antineutrophil and smooth muscle actin are the most frequently detected autoantibodies[7]. The mechanisms by which these antibodies are produced are not fully understood, but HCV can trigger a B-lymphocyte-mediated immune response shortly after immune system activation. B-lymphocyte-driven humoral immunity produces specific antibodies that are unable to inactivate virus production and replication. Therefore, the continuous replication of HCV results in constant stimulation of B cells, which may lead to B-cell dysfunction and abnormal antibody production[8]. Alternatively, the presence of autoantibodies in HCV patients may be caused by chronic apoptotic hepatocytes. Viruses, unlike bacteria and fungi, cannot reproduce on their own and must use the host-cell processes to replicate as they cannot synthesize their own proteins[9,10]. However, the pathogenic mechanism of the virus and whether antibody production truly represents an independent autoimmune disease have not been fully elucidated. During the progression of CHC in the present case, autoimmune hepatitis antibodies showed positivity for anti-nuclear and anti-mitochondrial antibody M2, and atypical lymphocytes briefly appeared, which is rare in viral hepatitis[11]. Timely use of artificial liver replacement therapy and hormonal suppression of immunity can control disease development, indicating that autoimmunity plays an important role in the progression of hepatitis C to liver failure. When hepatitis C was cured, the above antibody test results were negative. It can be seen that these antibodies became negative after HCV clearance and were not an independent factor in liver function damage.

***Bacterial infection***

Bacterial infection may be another important reason for the rapid progression to ACLF in this case. The Asia Pacific Liver Research Association defines ACLF as an acute liver injury characterized by jaundice [serum bilirubin ≥ 5 mg/dL (85 μmol/L) and coagulation disorders (INR ≥ 1.5 or PTA < 40%), accompanied by clinical ascites and/or hepatic encephalopathy within 4 wk, with or without prior diagnosis of chronic liver disease/cirrhosis, and associated with a high 28-d mortality rate[12]. It is well known that bacterial infection is the most common precipitating factor of ACLF. One study demonstrated that the overall rate of ACLF related to bacterial infection was 48%, but the rate varied between geographical regions (38% in southern Europe, and 75% in the Indian subcontinent[13]. In particular, extensively drug resistant bacteria caused by spontaneous bacterial peritonitis, pneumonia, or infection are more frequently associated with ACLF. Timely empirical antibiotic treatment can change the balance between bacteria and the host, which is beneficial for bacterial clearance. Bacterial infection may be another important reason for rapid progression to ACLF. Our patient had a pulmonary infection during his second admission, and the condition was still progressing after ceftazidime treatment for the infection. The infection was gradually controlled by changing to piperacillin–tazobactam sodium, and the bacteria that may have caused the patient's pulmonary infection were sensitive to piperacillin-tazobactam sodium.

***Refractory genotypes and resistance-associated variants***

The primary goal of DAA therapy is SVR, which is defined as undetectable HCV RNA 12 wk after the end of antiviral therapy[14,15]. Viral resistance is a major cause of virological failure in patients receiving DAAs for CHC. Selection of the DAA regimen needs to take account of the drug resistance of the virus and HCV genotype. The proportion of patients with HCV genotype 3 was higher in the population who experienced DAA failure[16]. The current first-line drugs for hepatitis C, sofosbuvir and velpatasvir, have high antiviral activity and a high resistance barrier, and resistance-associated variants may exist in patients with HCV genotype 3 and other rare HCV genotypes. This patient was diagnosed with a low viral load and genotype was not detected during two consecutive hospitalizations and ACLF occurred during treatment. After active treatment, the patient improved and then continued to take antivirals. Follow-up showed that the treatment was effective, and no drug resistance had developed, but the genotype was still unknown. The global distribution of HCV genotypes is regional, with 1b being the main genotype in China. At the same time, there are significant population differences in the distribution of HCV genotypes, and transmission methods may also vary. Our patient has no history of blood transfusion or drug use. During antiviral treatment, liver failure occurred, and the genotype was not detected during two consecutive hospitalizations. We therefore speculate that this patient is rare in terms of the HCV genotypes, or there might be a seventh genotype that we do not know.

**CONCLUSION**

DAAs have significantly improved the cure rate of CHC. However, this case also suggests that there is still a risk of liver failure during CHC treatment with DAAs if there are factors such as autoimmunity, combined bacterial infection, or unclear HCV genotype, and timely therapy requires close monitoring.

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**Table 1 Laboratory results during hospitalization**

|  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  | **April 27** | **April 29** | **April 30** | **May 1** | **May 3** | **May 5** | **May 7** | **May 9** | **May 11** | **May 15** | **May 17** |
| ALT (U/L) | 495 | 917 | 859 | 372 | 176 | 110 | 61 | 51 | 47 | 34 | 27 |
| AST (U/L) | 935 | 1841 | 1176 | 323 | 90 | 45 | 28 | 34 | 49 | 38 | 36 |
| ALP (U/L) | 316 | 251 | 278 | 168 | 148 | 199 | 198 | 162 | 165 | 175 | 169 |
| GGT (U/L) | 203 | 110 | 121 | 58 | 52 | 66 | 64 | 83 | 83 | 85 | 78 |
| TBIL (μmol/L) | 337 | 335.1 | 415.7 | 255.1 | 155.1 | 132.9 | 116.8 | 158 | 150.9 | 101.8 | 77.4 |
| ALB (g/L) | 42 | 29.4 | 35 | 30.1 | 30.6 | 31.2 | 32.3 | 37.1 | 36.8 | 40.1 | 37.8 |
| PTA (%) | 82 | 40.8 | 33.1 | 26 | 67.5 | 110.3 | 135.1 | 150 | 139.3 | 110 | - |
| PT-INR | 1.12 | 1.82 | 2.20 | 2.78 | 1.24 | 0.94 | 0.85 | 0.78 | 0.84 | 0.95 | - |
| WBC (109/L) | 10.5 | 10.16 | 13.91 | 16.05 | 9.71 | 7.91 | - | 7.20 | - | 5.07 | - |
| N (109/L) | 5.36 | 5.28 | 9.88 | 11.88 | 7.19 | 5.06 | - | 4.61 | - | 3.19 | - |
| L (109/L) | 3.46 | 3.35 | 2.64 | 3.05 | 1.84 | 2.21 | - | 2.09 | - | 1.42 | - |
| PLT (1012/L) | 504 | 455 | 639 | 529 | 482 | 418 | - | 311 | - | 330 | - |

ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; ALP: Alkaline phosphatase; GGT: γ-Glutamyl transferase; TBIL: Total bilirubin; ALB: Albumin; PTA: Prothrombin time activity; PT-INR: International normalized ratio; WBC: White blood cells; N: Neutrophil absolute value; L: Lymphocyte absolute value; PLT: Platelets.