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***Observational Study***

**Causal roles of gut microbiota in cholangiocarcinoma etiology suggested by genetic study**

Chen ZT *et al*. Causality between gut microbiota and CCA

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**Abstract**

BACKGROUND

Cholangiocarcinoma (CCA) is a highly malignant biliary tract cancer with poor prognosis. Previous studies have implicated the gut microbiota in CCA, but evidence for causal mechanisms is lacking.

AIM

To investigate the causal relationship between gut microbiota and CCA risk.

METHODS

We performed a two-sample mendelian randomization study to evaluate potential causal associations between gut microbiota and CCA risk using genome-wide association study summary statistics for 196 gut microbial taxa and CCA. Genetic variants were used as instrumental variables. Multiple sensitivity analyses assessed result robustness.

RESULTS

Fifteen gut microbial taxa showed significant causal associations with CCA risk. Higher genetically predicted abundance of *Eubacteriumnodatum group*, *Ruminococcustorques group*, *Coprococcus*, *Dorea*, and *Actinobacteria* were associated with reduced risk of gallbladder cancer and extrahepatic CCA. Increased intrahepatic CCA risk was associated with higher abundance of *Veillonellaceae*, *Alistipes*, *Enterobacteriales*, and *Firmicutes.* Protective effects against CCA were suggested for *Collinsella*, *Eisenbergiella*, *Anaerostipes*, *Paraprevotella*, *Parasutterella*,and *Verrucomicrobia.* Sensitivity analyses indicated these findings were reliable without pleiotropy.

CONCLUSION

This pioneering study provides novel evidence that specific gut microbiota may play causal roles in CCA risk. Further experimental validation of these candidate microbes is warranted to consolidate causality and mechanisms.

**Key Words:** Cholangiocarcinoma; Mendelian randomization; Gut microbiota; Instrumental variables; Sensitivity analyses

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**Core Tip:** Cholangiocarcinoma (CCA) is a highly malignant biliary tract cancer with poor prognosis. Emerging evidence suggests the gut microbiota may play a causal role in CCA pathogenesis, but robust genetic evidence is still lacking. Using genome-wide association study summary statistics, our study provides novel evidence that 15 gut microbial taxa may confer either protective or detrimental causal effects on CCA risk.

**INTRODUCTION**

Cholangiocarcinoma (CCA) originates from the biliary epithelium and is among the most prevalent malignancies due to its significant malignancy potential[1,2]. Based on anatomical site of origin, CCA manifests as three distinct subtypes: Intrahepatic CCA (iCCA), extrahepatic CCA (eCCA), and gallbladder cancer (GC)[1]. Established risk factors for CCA include fluke infections, inflammatory bowel disease, intrahepatic bile duct stones, choledochal cysts, and primary sclerosing cholangitis (PSC)[3,4]. Despite recent progress in diagnosis and therapy, CCA prognosis remains poor with 5-year survival below 5% for advanced disease[5-7]. Further elucidation of CCA pathogenesis at the molecular, epigenetic and genomic levels is therefore critical to enable novel treatment approaches.

In recent years, the gut microbiota has emerged as a key factor governing health[8,9]. Microbiota dysbiosis can impact immune function, metabolism and physiology, contributing to diseases like obesity, diabetes, non-alcoholic fatty liver disease and cancer[10-12]. Anatomically and physiologically, the hepatobiliary duct and gastrointestinal tract comprise a “gut-liver axis” that regulates liver pathology and intrahepatic/systemic immunity[13]. The microbiota likely contributes to diverse hepatobiliary conditions including cancer, PSC, choledocholithiasis and cholelithiasis[14-17]. Previous research has revealed that the gut microbiota plays a pivotal role in the diagnosis and treatment of CCA[18,19]. In-depth investigations into the role of the gut microbiota in CCA have significantly improved the prognostic outlook for individuals affected by this disease. However, the causal relationship between the gut microbiota and CCA remains unclear. Elucidating such mechanisms would enable microbiome modulation as an early preventative approach aligning with precancer interception paradigms.

Establishing causality is challenged by limited clinical trial follow-up and potential confounding in observational studies. Mendelian randomization (MR) helps address this by using genetic variants as instrumental variables (IVs)[20]. The present study represents a pioneering effort in employing a two-sample MR approach to discern a potential causal link between particular gut microbiota taxa and CCA, thereby offering valuable insights for subsequent mechanistic inquiries.

**MATERIALS AND METHODS**

***Ethical statement and study design***

This research adheres to the STROBE-MR Guidelines, and all data employed in this study are openly available and appropriately cited[21]. Consequently, our study did not require additional ethics committee approval.

Figure 1 presents the directed acyclic graph guiding the design of the current MR study. In this framework, the gut microbiota represents the exposure variables, while CCA constitutes the outcome variable. Genetic variants associated with gut microbiota taxa were leveraged as IVs to evaluate potential causal associations of gut microbiota composition with CCA risk, thereby minimizing issues of confounding.

***Date sources***

A large-scale genome-wide association study (GWAS) encompassed 18340 participants drawn from 24 diverse cohorts spanning multiple countries, examining 122110 Loci of genetic variation. This study provided summary statistics for gut microbiota based on 16S rRNA gene sequencing data obtained from the MiBioGen (<https://mibiogen.gcc.rug.nl/>) database[22]. Among the participants, a significant majority, 13266 individuals or 72.3%, were of European ancestry. The study encompassed a broad spectrum of 211 traits, which included members from 131 genera, 35 families, 20 orders, 16 classes, and 9 phyla. In the current MR study, 15 unidentified taxa were notably excluded, resulting in the analysis incorporating 196 taxonomic units, spanning 9 phyla, 16 classes, 20 orders, 32 families, and 119 genera. Jiang *et al*[23] conducted analyses of summary statistics for malignant neoplasms of the gallbladder and extrahepatic bile ducts (195 European ancestry cases, 456153 European ancestry controls) as well as intrahepatic CCA (104 European ancestry cases, 456244 European ancestry controls), which were provided by the GWAS Catalog (https://www.ebi.ac.uk/gwas/).

***The choice of IVs***

MR analyses utilized IVs, primarily single nucleotide polymorphisms (SNPs), as mediators to explore causality between exposures and outcomes. The foundational assumption in MR necessitates that all SNPs robustly and independently predict the exposure variable at the genome-wide significance level. In present research, we utilized robust SNPs associated with gut microbiota as IVs for the exposure variable. However, applying a stringent threshold of 5 × 10-8 would have excluded the majority of these SNPs. Consequently, we opted for a relatively lenient yet still statistically significant threshold of 1 × 10-05, as supported by prior studies[24,25]. This threshold was set to encompass most gut microbiota-associated SNPs, ensuring that those with an *R2* < 0.001 and a physical distance (kb) of 10000 were included, thus mitigating linkage disequilibrium (LD). The *F* statistic was employed to assess the strength of the correlation between IVs and exposures, with an *F* statistic exceeding 10 typically indicating a substantial correlation. These screening criteria serve to establish the reliability of the findings in present MR study.

***Statistical analysis***

The analyses were conducted in RStudio (Version: 2023.06.1 + 524) using the TwoSampleMR package (version 0.5.7) and MRPRESSO package. In the context of a global-level test, a two-sided *P* value of 0.05 was considered statistically significant. In present study, we utilized a comprehensive approach, incorporating MR-Egger, weighted median, inverse variance weighted (IVW), simple mode, and weighted mode methodologies for MR analysis, enabling a thorough assessment of the causal relationship between gut microbiota and CCA. In cases where pleiotropy among IVs is absent, IVW is selected as the primary analytical method due to its superior statistical power[26]. To evaluate the reliability of our findings, we conducted a set of sensitivity analyses, including Cochran’s *Q* test, MR-Egger intercept test, and MR-PRESSO global test. Both the Cochrane’s *Q* test and MR-Egger intercept test were employed to assess the presence of SNP-associated heterogeneity and horizontal pleiotropy for each gut microbiota trait. The outcomes revealed *P* values exceeding 0.05, indicating the absence of heterogeneity and horizontal pleiotropy. Outliers were identified through the application of MR-PRESSO analysis. Additionally, we also employed funnel plots and conducted leave-one-out sensitivity tests to assess heterogeneity. The leave-one-out analysis was utilized to identify potential pleiotropic effects originating from individual SNPs. Scatter plots, forest plots, funnel plots, and leave-one-out sensitivity tests serve as valuable tools for visualizing MR results in a comprehensive manner.

**RESULTS**

***Selection of IVs***

Initially, we identified 122100 SNPs associated with gut microbiota traits through the MiBioGen Consortium dataset. Following a rigorous sequence of quality control procedures based on locus-wide statistical significance (*P* < 1 × 10-5) and the LD threshold (*R2* < 0.001, with a clumping distance of 10000 kb), 2236 SNPs associated with 196 gut microbiota trials were selected as IVs. Notably, all IVs exhibited F-statistics exceeding 10, thereby indicating the absence of evidence for weak instrument bias (Supplementary Table 1). Based on these SNPs, we have extracted corresponding pieces of information from the outcome variable dataset (Supplementary Tables 2 and 3).

***Causal inference of the relationship of the gut microbiota with GC and eCCA risk***

According to the results of the IVW method, the higher genetically predicted abundance of *genus Eubacteriumnodatum group* [odds ratio (OR) = 0.489, standard error (SE) = 0.267, 95% confidence interval (CI): 0.290-0.826, *P* = 0.007], *genus Ruminococcustorques group* (OR = 0.291, SE = 0.535, 95%CI: 0.102-0.830, *P* = 0.021), *genus Coprococcus* (OR = 0.296, SE = 0.593, 95%CI: 0.093-0.946, *P* = 0.040), *genus Dorea* (OR = 0.250, SE = 0.622, 95%CI: 0.074-0.847, *P* = 0.026), *phylum actinobacteria* (OR = 0.348, SE = 0.483, 95%CI: 0.135-0.898, *P* = 0.029) were associated with a reduced risk of GC and eCCA (Figure 2A). In contrast, genetically predicted abundance of *genus* *Collinsella* (OR = 6.977, SE = 0.621, 95%CI: 2.006-23.560, *P* = 0.002), *genus* *Eisenbergiella* (OR = 2.018, SE = 0.342, 95%CI: 1.033-3.941, *P* = 0.040)was positively related to GC and eCCA risk (Figure 2B). The weighted median, simple mode, and weighted mode exhibited the same directional impact as IVW, although the *P* values were not consistently statistically significant (Supplementary Table 4, Figure 3).

***Causal inference of the relationship of the gut microbiota with iCCA risk***

Employing the IVW method, our study found suggestive evidence of a potential causal link between genetically predicted increases in the *family Veillonellaceae* (*P* = 0.014, 95%CI: 1.292-9.929, OR = 3.582, SE = 0.520), *order Enterobacteriales*/*family Enterobacteriaceae* (*P* = 0.032, 95%CI: 1.156-27.429, OR = 5.632, SE = 0.808), *genus Alistipes* (*P* = 0.020, 95%CI: 1.316-24.245, OR = 5.648, SE = 0.743), and *phylum Firmicutes* (*P* = 0.046, 95%CI: 1.025-12.258, OR = 3.545, SE = 0.633) with an increased risk of iCCA (Figure 2B). From the earlier mentioned traits, it was noted that both the *order Enterobacteriales* and the *family Enterobacteriaceae* fall under the same bacterial category and share identical IVs. Furthermore, our findings suggest that genetically predicted increases in the *genus Anaerostipes* (*P* = 0.006, 95%CI: 0.033-0.564, OR = 0.135, SE = 0.728), the *genus Parasutterella* (*P* = 0.032, 95%CI: 0.115-0.907, OR = 0.323, SE = 0.527), the *genus Paraprevotella* (*P* = 0.005, 95%CI: 0.107-0.672, OR = 0.268, SE = 0.470), and the *phylum Verrucomicrobia* (*P* = 0.005, 95%CI: 0.048-0.588, OR = 0.168, SE = 0.640) are associated with protective effects against iCCA (Figure 3A). Additionally, the causal effect estimates derived from the weighted median, simple mode, and weighted mode methods showed similar magnitudes and directions as those obtained with the previously mentioned IVW method (Supplementary Table 4, Figure 4).

***Sensitivity analysis***

Subsequently, a comprehensive sensitivity analysis was carried out to assess the stability and reliability of the inferred causal relationship between gut microbiota and CCA. The detailed description is summarized in Table 1. Cochran’s *Q* statistics revealed the absence of significant heterogeneity among the selected IVs, with *P* values exceeding 0.05 in both IVW and MR-Egger methods. Furthermore, our analysis, employing the MR-Egger intercept method, did not reveal any indications of a horizontal pleiotropic effect (*P* > 0.05). The outcomes from the MR-PRESSO trial indicated the absence of any horizontal pleiotropic outliers. The leave-one-out analysis results demonstrated that none of the SNPs were influential outliers (Figures 5 and 6). Additionally, the use of funnel and forest plots to depict a symmetrical pattern serves to visually affirm the reliability of the study’s results (Supplementary Figures 1-4).

**DISCUSSION**

CCA is a highly diverse form of cancer, with its global incidence steadily on the rise[27]. With surgical resection being the exclusive curative treatment modality, the prognosis for individuals afflicted with CCA remains bleak[28]. In recent years, the prevention of tumor initiation and the inhibition of tumor progression have emerged as pivotal milestones in cancer management within the field of oncology. Changes in the composition of gut microbiota are closely associated with the initiation and progression of cancer[8,29]. This study represents the inaugural attempt to evaluate the causal link between gut microbiota and CCA while also endeavoring to identify particular causative microbial taxa through two-sample MR analyses based on GWAS summary statistics.

The gut microbiota constitutes a complex and dynamically evolving assembly of ecological microbial communities that reside within the human gastrointestinal tract, often referred to as a “neglected organ”[30-32]. These microorganisms assume a pivotal role in maintaining the homeostasis of the digestive system, exerting multifaceted metabolic, immunological, and protective functions that contribute to the overall health of the host[32]. Although gut microbiota plays a crucial role in facilitating various essential and advantageous physiological processes, such as the digestion of macronutrients and the synthesis of certain vitamins, a wealth of empirical data underscores their potential involvement in the emergence of detrimental phenotypes[33,34]. Notably, discernible alterations in both the structure and function of the microbial community have been linked to numerous disease states, including cancer[31]. Due to the bidirectional communication between the gastrointestinal tract and the biliary system, the liver excretes bile acids and other biologically active components *via* the bile duct to interface with the intestine. Simultaneously, the gut microbiota and its metabolites are transported to the liver through the bile duct. Therefore, the gut microbiota plays a pivotal role in the pathogenesis and progression CCA[18,35]. Hence, there is a compelling need for additional investigations to elucidate the causal connection between gut microbiota and CCA, thereby establishing a novel theoretical foundation for the prevention and treatment of CCA.

The impact of the gut microbiota in the field of oncology is a double-edged sword, and our research has equally substantiated this perspective from a genetic standpoint. The gut microbiota actively fosters the development of extraintestinal cancers by facilitating bacterial translocation and the generation of bioactive molecules within the biliary tract. Numerous research investigations have demonstrated notable distinctions in gut microbiota composition between individuals with extraintestinal cancers and those without the disease[32,35]. *Bacteroides* and *Ruminococcaceae* have been shown to potentially contribute to the pathogenesis of hepatocellular carcinoma by exacerbating hepatic inflammation, accumulating toxic compounds, and inducing liver steatosis[36]. Zhang *et al*[35] observed a depletion of *Saccharomyces cerevisiae* (*S. cerevisiae*) in iCCA. Significantly, past studies have shown that *S. cerevisiae* has the capacity to impede the growth of colorectal tumors. It achieves this by inducing apoptosis in epithelial cells, modulating intestinal immunity, and altering the composition of the gut microbiota[37]. The microbiota can indirectly influence tumor progression by generating and metabolizing bioactive molecules, which, when carried through systemic circulation, such as bacterial LPS entering the bloodstream, can impact tumor formation in distant tissues from the gastrointestinal tract[38]. *Lactic acid bacteria* and *Bifidobacterium* play a role in the regulation of pH and bile acid processes[39]. Furthermore, their enzymatic capacity to degrade potential carcinogens and their metabolites, including heterocyclic amines, nitrosamines, and aflatoxins, contributes to the inhibition of the development of various cancers, such as gastric and liver cancers[40].

In this MR study, we determined that 15 microbial taxa are causally associated with CCA. Elevated genetic predisposition towards higher abundance of the *genus Eubacteriumnodatum group*, *genus Ruminococcustorques group*, *genus Coprococcus*, *genus Dorea*, *phylum* *Actinobacteria*, *family Veillonellaceae*, *genus Alistipes*, *order Enterobacteriales*/*family Enterobacteriaceae*, and *phylum Firmicutes* were found to be associated with a decreased risk of CCA. Conversely, a genetically predicted increase in the abundance of the *genus* *Collinsella*, *genus* *Eisenbergiella*, *genus Anaerostipes*, *genus Paraprevotella*, *genus Parasutterella*, and *phylum Verrucomicrobia* exhibited a positive correlation with the risk of CCA. The abundance of the *genus Eubacteriumnodatum group* was found significantly reduced in colorectal cancer patients, and its functionality appeared to be associated with processes related to protein digestion and absorption, as well as the renin-angiotensin system pathway[41]. The *genus Ruminococcustorques group* and *phylum* *Actinobacteria* also demonstrated an association with an elevated risk of bladder cancer[42]. *Genus Coprococcus* is a bacterium known for producing butyrate, and its presence may be associated with a reduction in the effectiveness of neoadjuvant chemoradiation therapy for rectal cancer[43]. Elevated abundance of the gut *genus Dorea* has the potential to serve as a predictive factor for farnesoid X receptor deactivation, which is recognized as a risk factor for metabolic dysfunction-associated steatotic liver disease[44]. The aforementioned findings indicate that certain protective or risky gut microbiota identified in this MR analysis are consistent with previous research and are likely to play significant roles as reference points in future clinical studies. Furthermore, the causal relationship between gut microbiota and CCA warrant further investigation through clinical and *in vivo* experiments to enhance our understanding of the “gut-liver axis” theories.

One major strength of this study is the utilization of the MR method, which assists in mitigating the impact of confounding variables, thereby enhancing the persuasiveness of the findings compared to observational research. Nevertheless, our analysis has several limitations that warrant consideration. Firstly, it is important to note that MR analyses were conducted at the bacterial genus level, as opposed to a more specific species level, due to the limited resolution provided by 16S rRNA sequencing. Secondly, the significance threshold for exposure IVs was set at 1 × 10-05 due to the inadequacy of IVs reaching genome-wide significance. However, IVs with F-statistics below 10 were excluded to mitigate the potential bias associated with weak instruments.

**CONCLUSION**

In summary, this two-sample MR study offers new insights, suggesting a potential causal link between certain gut microbiota taxa and CCA. By utilizing genetic variants as IVs, we identified 15 microbial taxa that may confer either protective or detrimental effects on CCA risk. This study sheds new light on the intricate gut-liver axis interactions and microbiota-mediated mechanisms underlying CCA. Further experimental validations are warranted to consolidate the causality, delineate the molecular events, and exploit the clinical values of these candidate microbes.

**ARTICLE HIGHLIGHTS**

***Research background***

Cholangiocarcinoma (CCA) is a highly malignant biliary tract cancer with poor prognosis. Previous studies have implicated the gut microbiota in CCA, but evidence for causal mechanisms is lacking.

***Research motivation***

To investigate the causal relationship between gut microbiota and CCA risk.

***Research objectives***

To investigate the causal relationship between gut microbiota and CCA risk.

***Research methods***

We performed a two-sample mendelian randomization study to evaluate potential causal associations between gut microbiota and CCA risk using genome-wide association study summary statistics for 196 gut microbial taxa and CCA. Genetic variants were used as instrumental variables. Multiple sensitivity analyses assessed result robustness.

***Research results***

Fifteen gut microbial taxa showed significant causal associations with CCA risk. Higher genetically predicted abundance of *Eubacteriumnodatum group*, *Ruminococcustorques group*, *Coprococcus*, *Dorea*, and *Actinobacteria* were associated with reduced risk of gallbladder cancer and extrahepatic CCA. Increased intrahepatic CCA risk was associated with higher abundance of *Veillonellaceae*, *Alistipes*, *Enterobacteriales*, and *Firmicutes.* Protective effects against CCA were suggested for *Collinsella*, *Eisenbergiella*, *Anaerostipes*, *Paraprevotella*, *Parasutterella*,and *Verrucomicrobia.* Sensitivity analyses indicated these findings were reliable without pleiotropy.

***Research conclusions***

This pioneering study provides novel evidence that specific gut microbiota may play causal roles in CCA risk. Further experimental validation of these candidate microbes is warranted to consolidate causality and mechanisms.

***Research perspectives***

Experimental validation of the candidate microbes identified to be causally associated with CCA risk. Further *in vitro* and *in vivo* studies could be conducted to consolidate the causal effects and explore the underlying molecular mechanisms. Analysis of species-level resolution of gut microbiota through metagenomic shotgun sequencing or other techniques. The current study was limited to genus-level associations due to 16S rRNA gene sequencing. A more detailed characterization at the species level could provide further insights.

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**Footnotes**

**Institutional review board statement:** The research does not involve any animal experiments and clinical data or human subjects, which does not require any special ethical clearance as per the guidelines of our institution.

**Informed consent statement:** Informed consent statement is not required since our manuscript solely utilizes publicly available data for analysis.

**Conflict-of-interest statement:** The authors declare that they have no competing interests.

**STROBE statement:** The authors have read the STROBE Statement—checklist of items, and the manuscript was prepared and revised according to the STROBE Statement—checklist of items.

**Data sharing statement:** The datasets used and analyzed in the present study are available from the corresponding authors on reasonable request. The datasets generated and/or analyzed during the current study are available in GWAS Catalog (https://www.ebi.ac.uk/gwas/) and MiBioGen (https://mibiogen.gcc.rug.nl) database.

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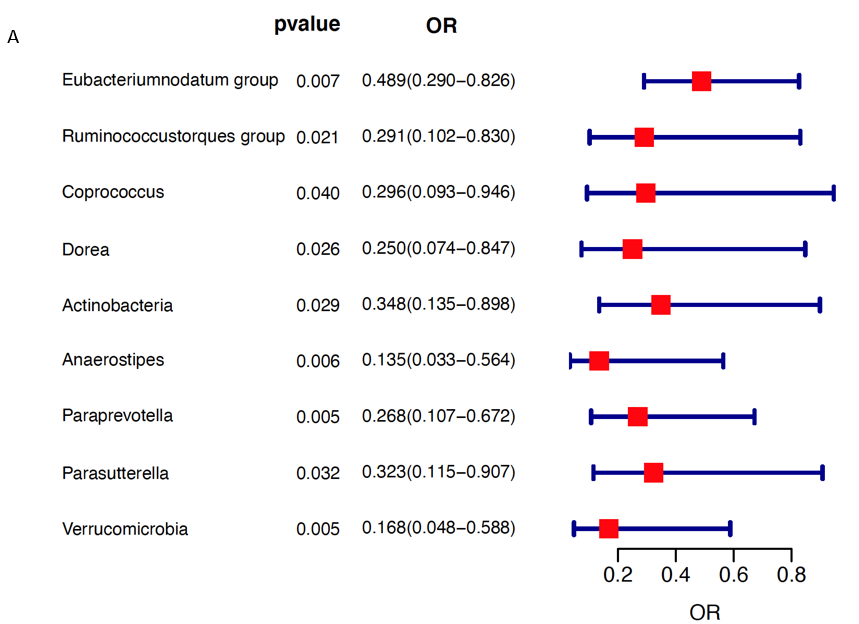
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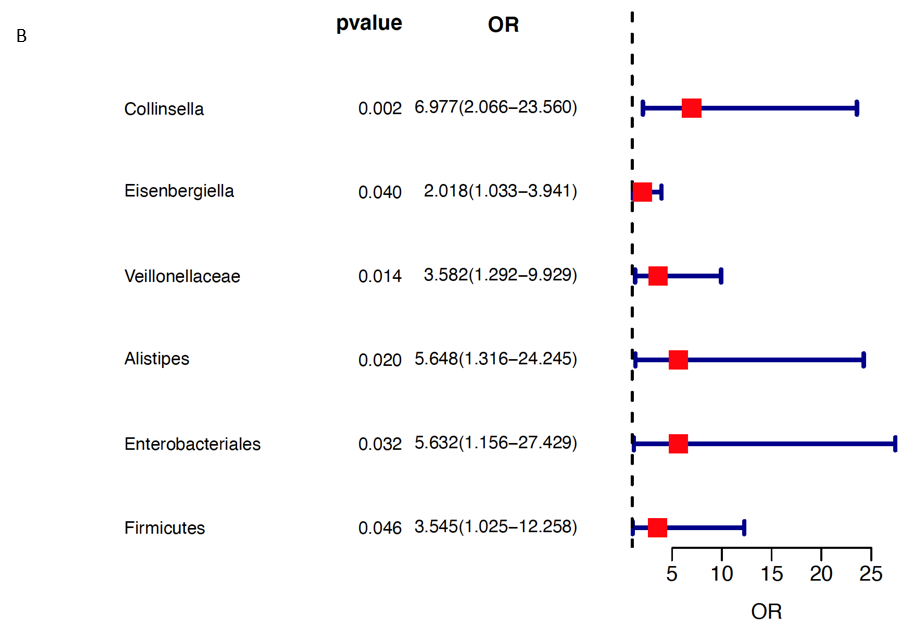
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**Figure Legends**

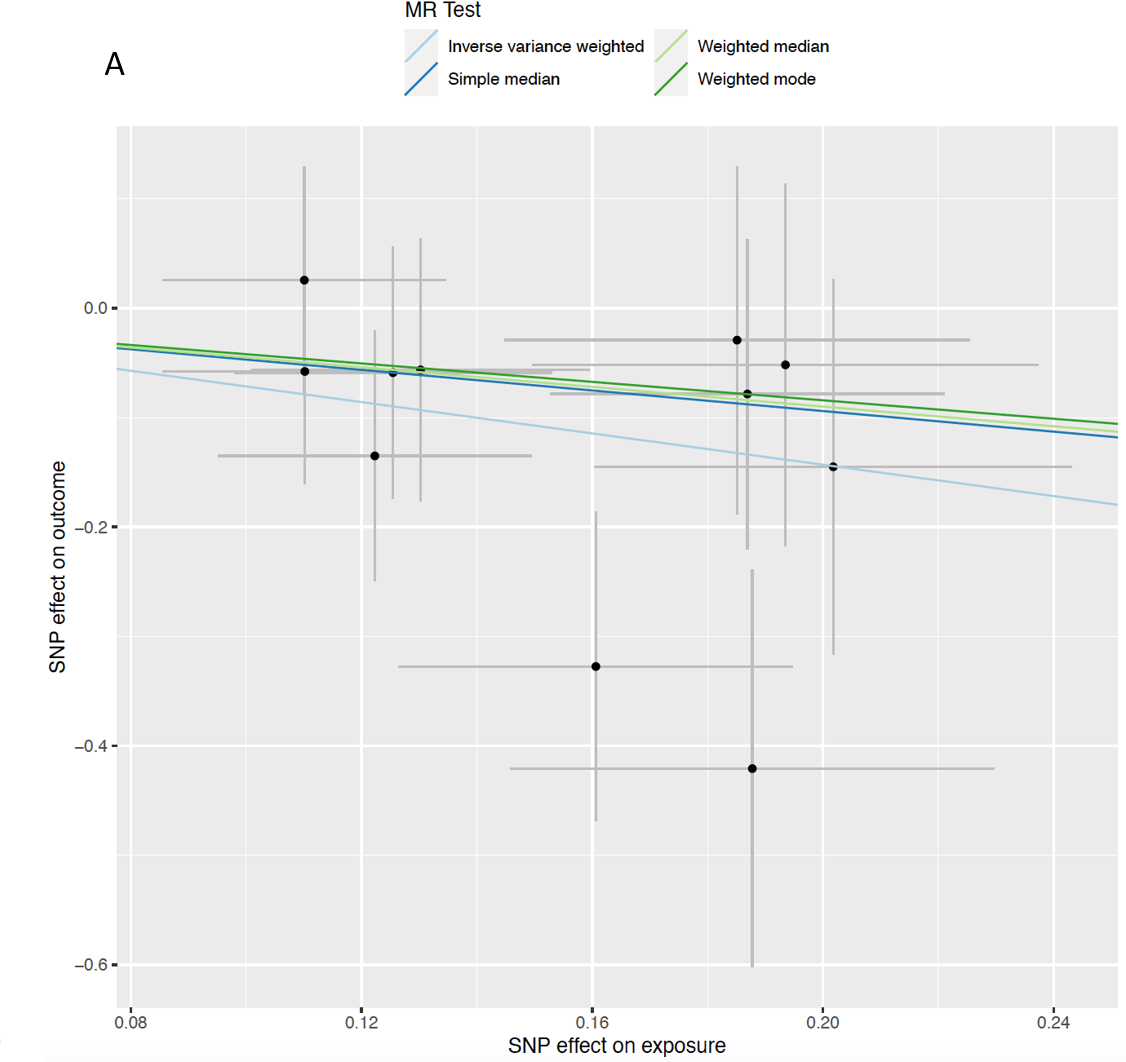
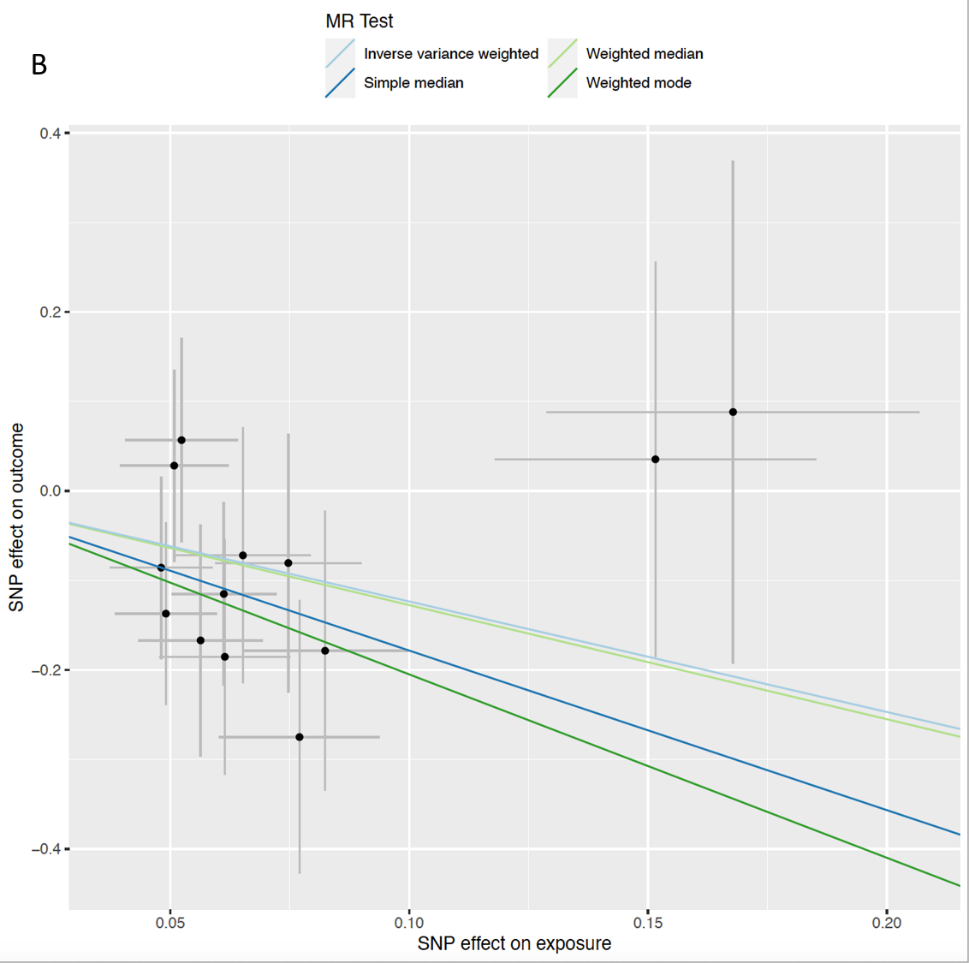
Figure 1

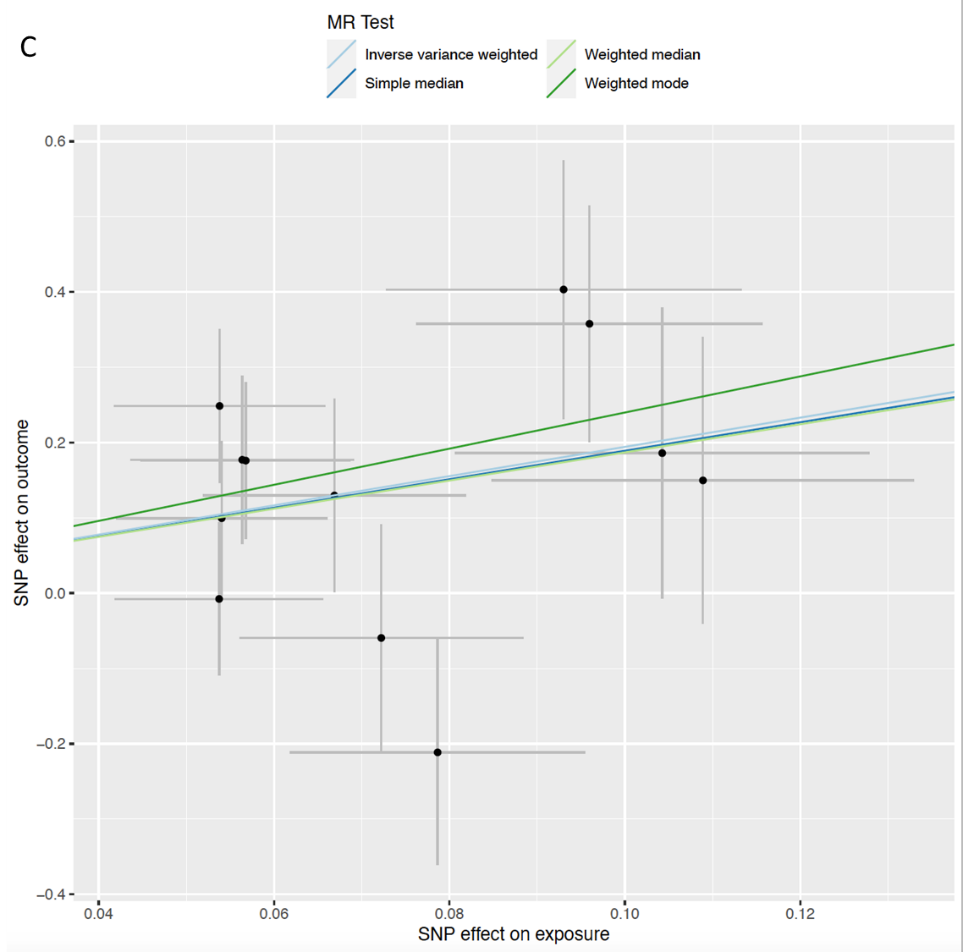
**Figure 1 The process of present mendelian randomization analyses is shown in flow chart.** A: Principle diagram of mendelian randomization study; B: Diagrammatic illustration of the complete mendelian randomization analysis process. Other risk factors influencing the occurrence of cholangiocarcinoma include exposure to fluke infections, inflammatory bowel disease, intrahepatic bile duct stones, choledochal cysts, and primary sclerosing cholangitis. Assumption 1: The instrumental variables (IVs) selected for this study should demonstrate a significant association with gut microbiota; Assumption 2: The IVs chosen for present study are required to have no significant associations with other potential confounding factors; Assumption 3: The IVs utilized in present study do not have any independent causal pathways leading to the outcome (CCA) other than through gut microbiota. MR: Mendelian randomization; IV: Instrumental variable; CCA: Cholangiocarcinoma; GC: Gallbladder cancer; eCCA: Extrahepatic cholangiocarcinoma; iCCA: Intrahepatic cholangiocarcinoma; LD: Linkage disequilibrium; SNPs: Single nucleotide polymorphisms.

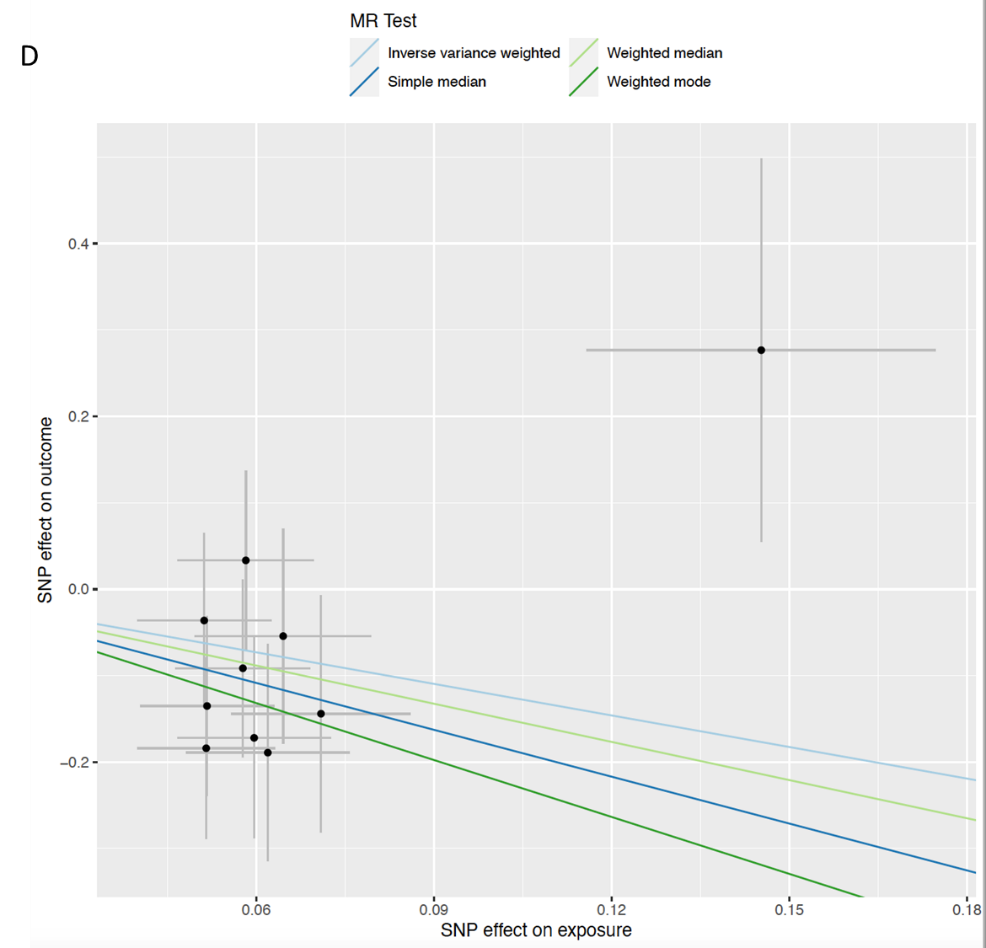


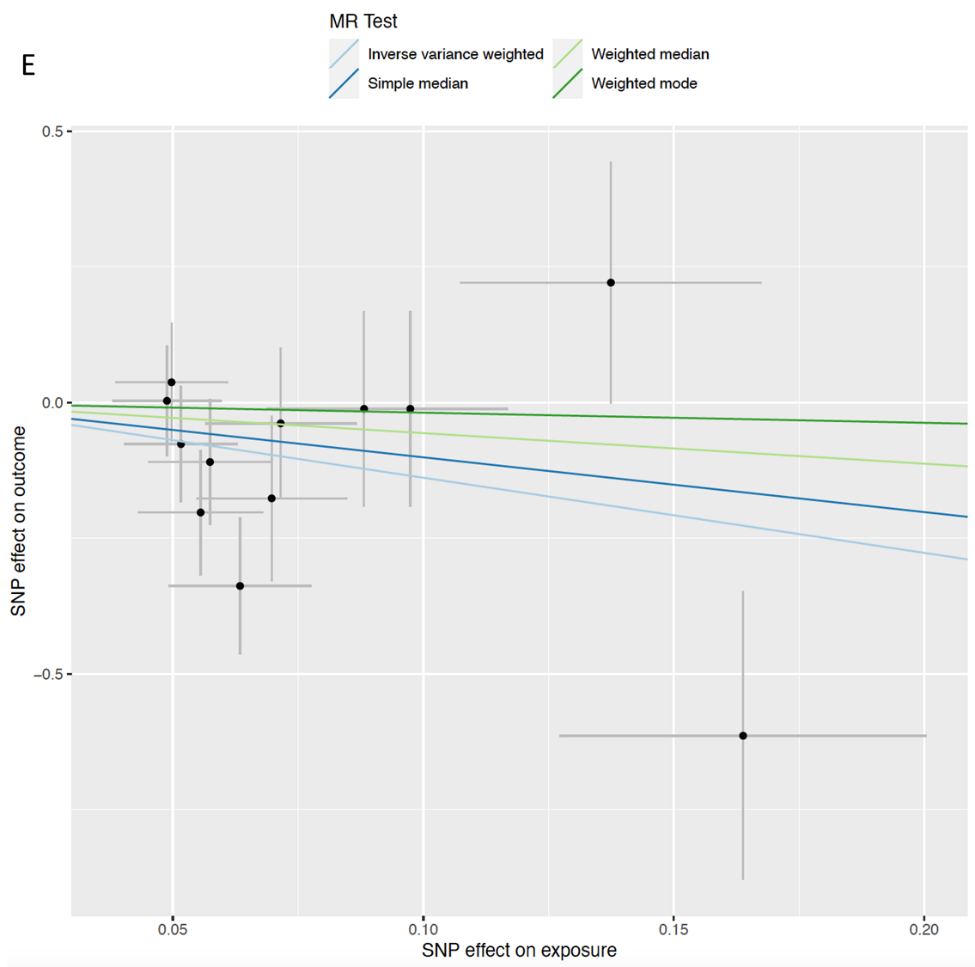


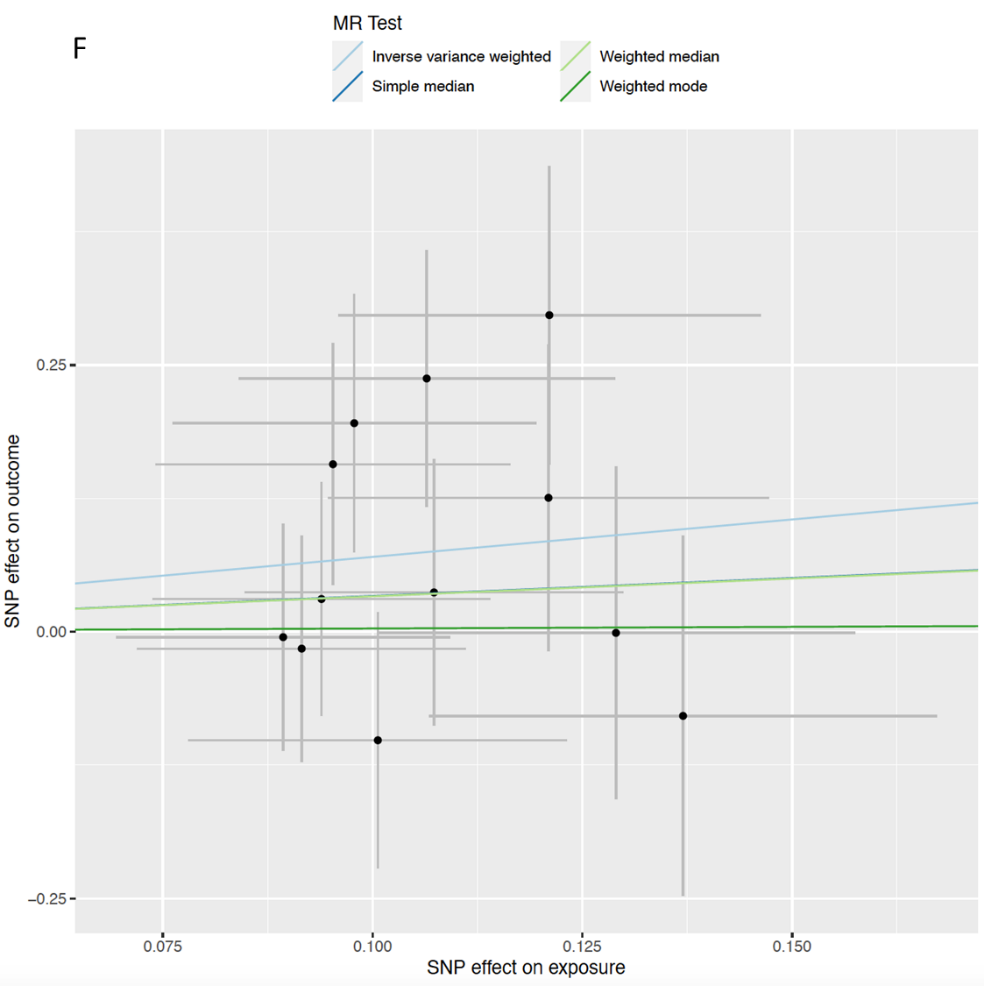
**Figure 2 The forest plot illustrates the relationships between gut microbiota and cholangiocarcinoma.** A: An increased abundance of gut microbial taxa was observed to be linked with a reduced risk of cholangiocarcinoma (CCA); B: An increased abundance of gut microbial taxa was observed to be linked with an elevated risk of CCA. OR: Odds ratio.

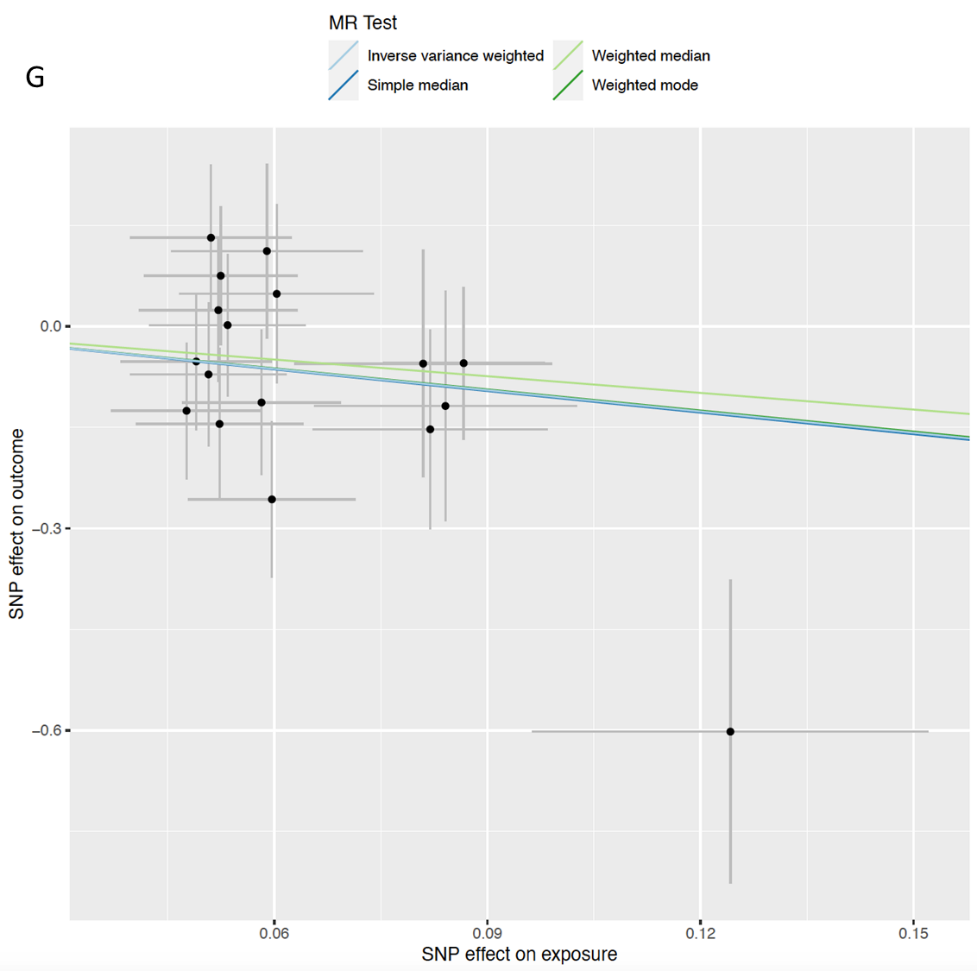
 



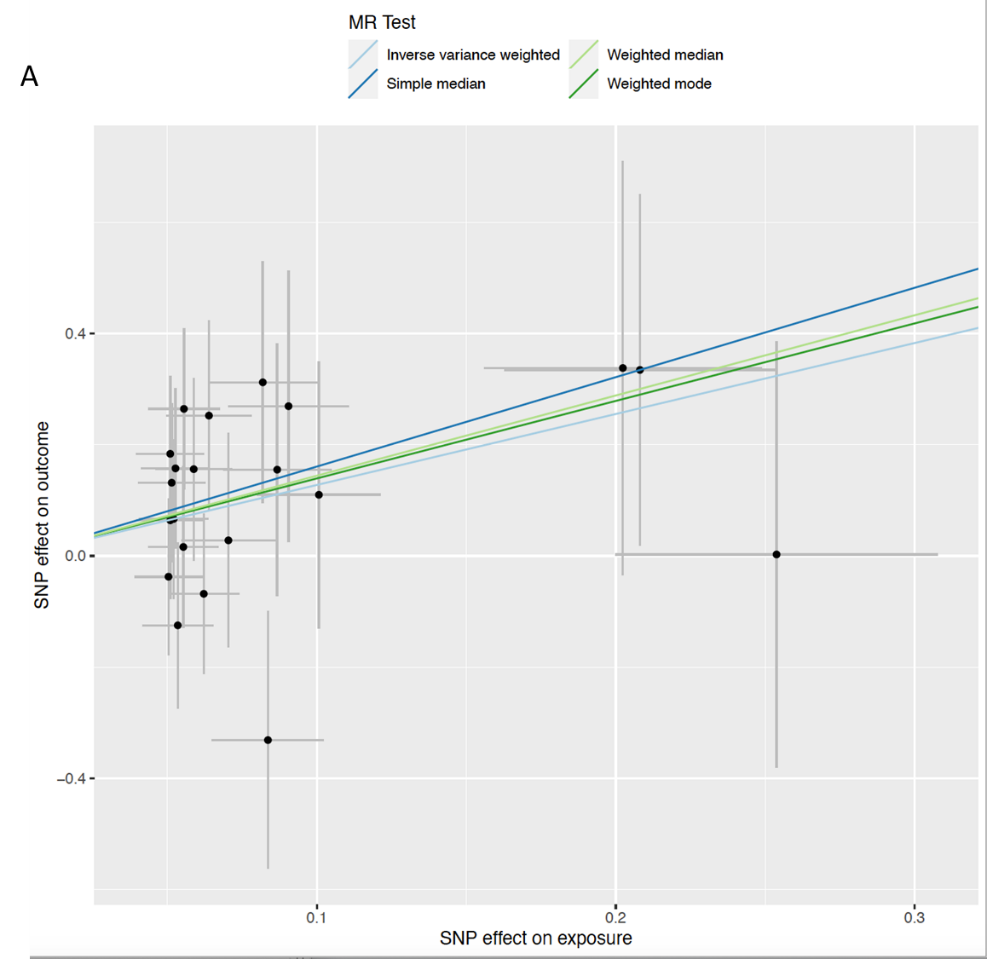


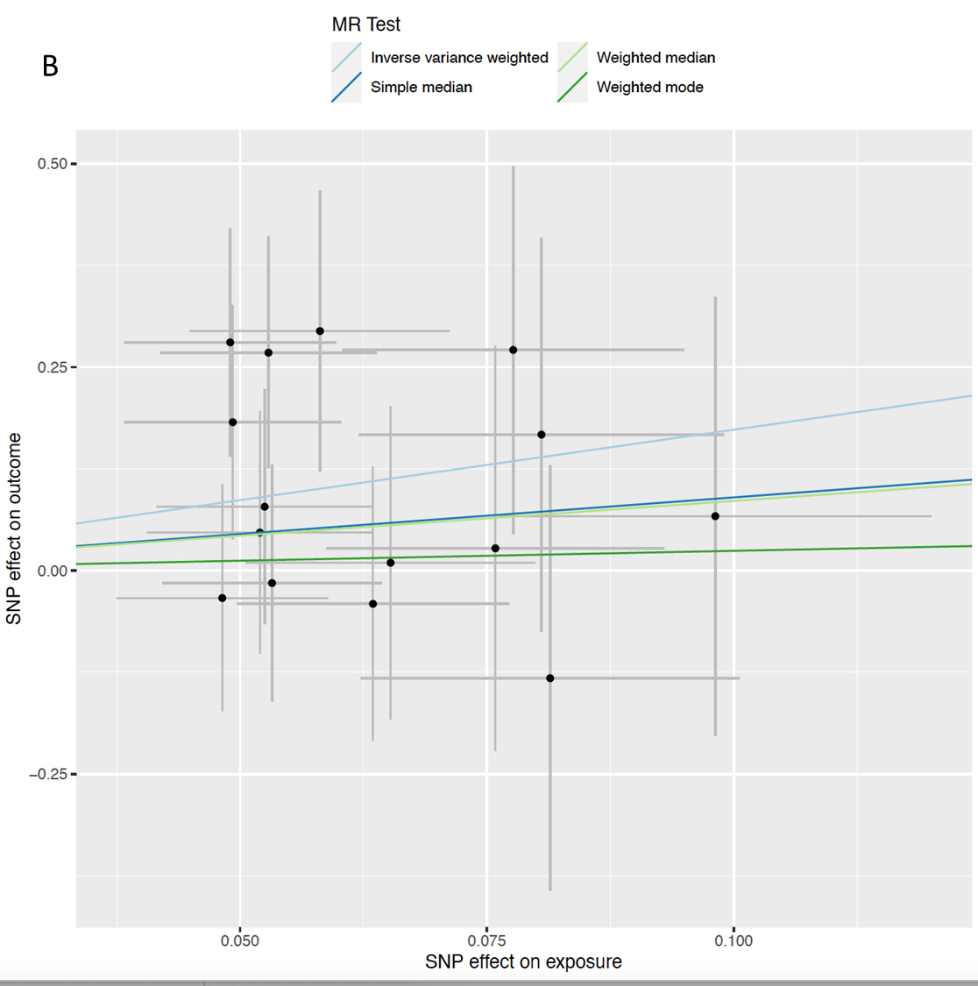
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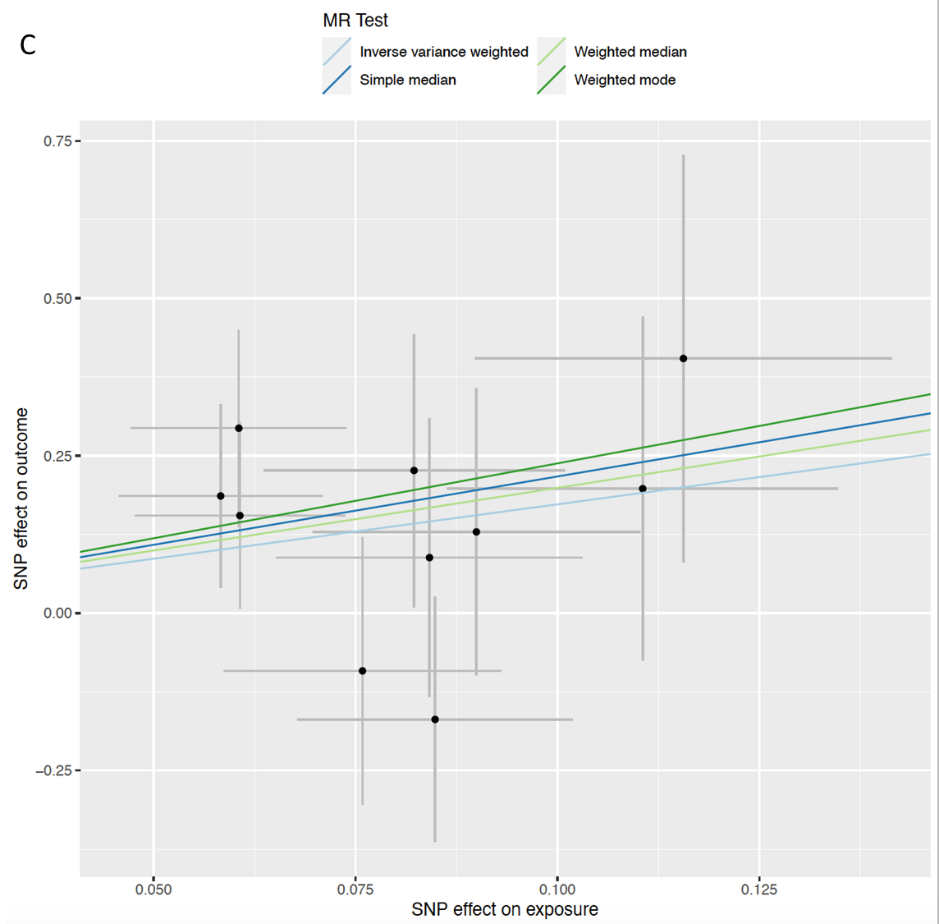
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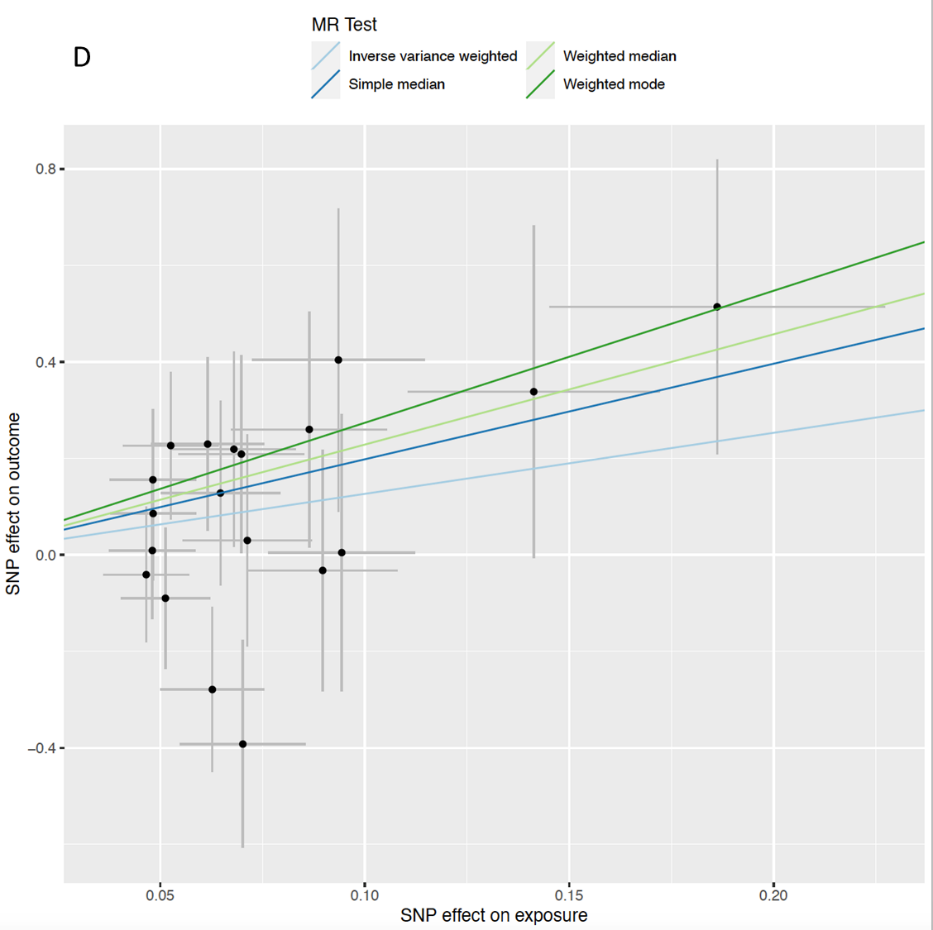
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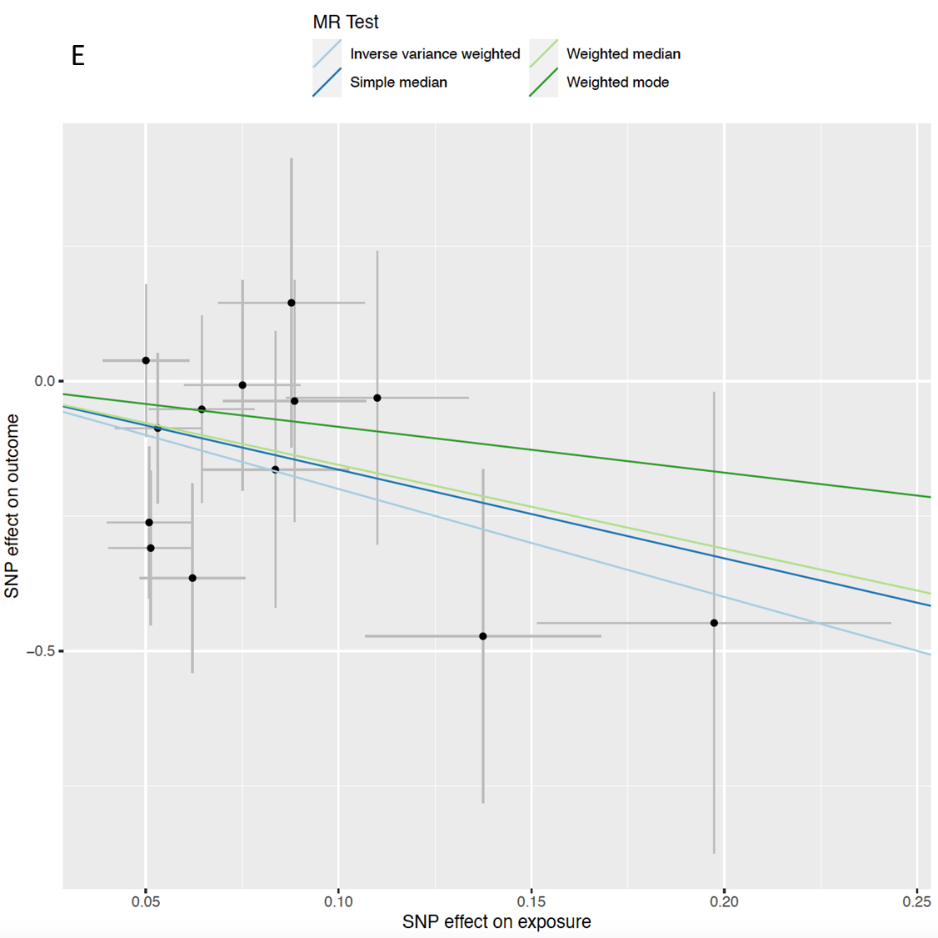
**Figure 3 Scatter plots of causal estimates of specific gut microbiota taxa on gallbladder cancer and extrahepatic cholangiocarcinoma.** The slope of each line corresponding to the estimated mendelian randomization effect in different models, including the conventional inverse variance weighted, weighted median, simple mode, and weighted mode.A: *Genus Eubacteriumnodatum group*; B: *Genus* *Ruminococcustorques* *group*; C: *Genus* *Collinsella*; D: *Genus* *Coprococcus*; E: *Genus* *Dorea*; F: *Genus* *Eisenbergiella*; G: *Phylum* *Actinobacteria.* MR: Mendelian randomization; SNP: Single nucleotide polymorphisms.

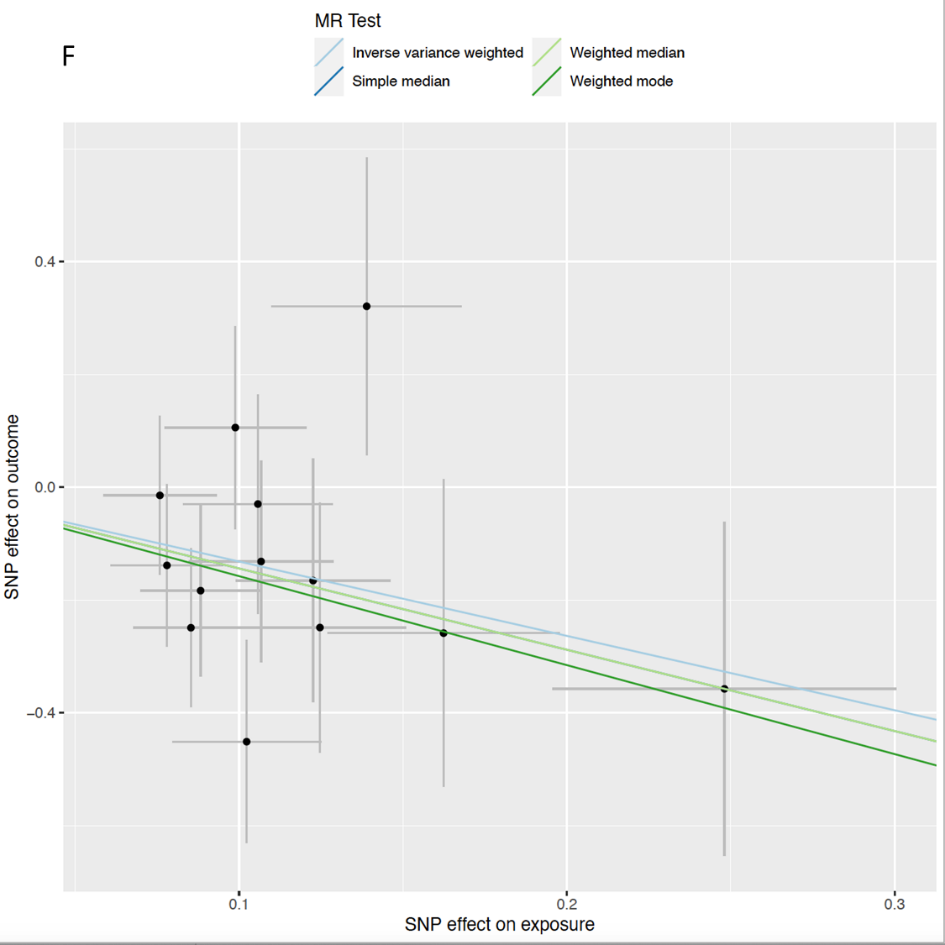


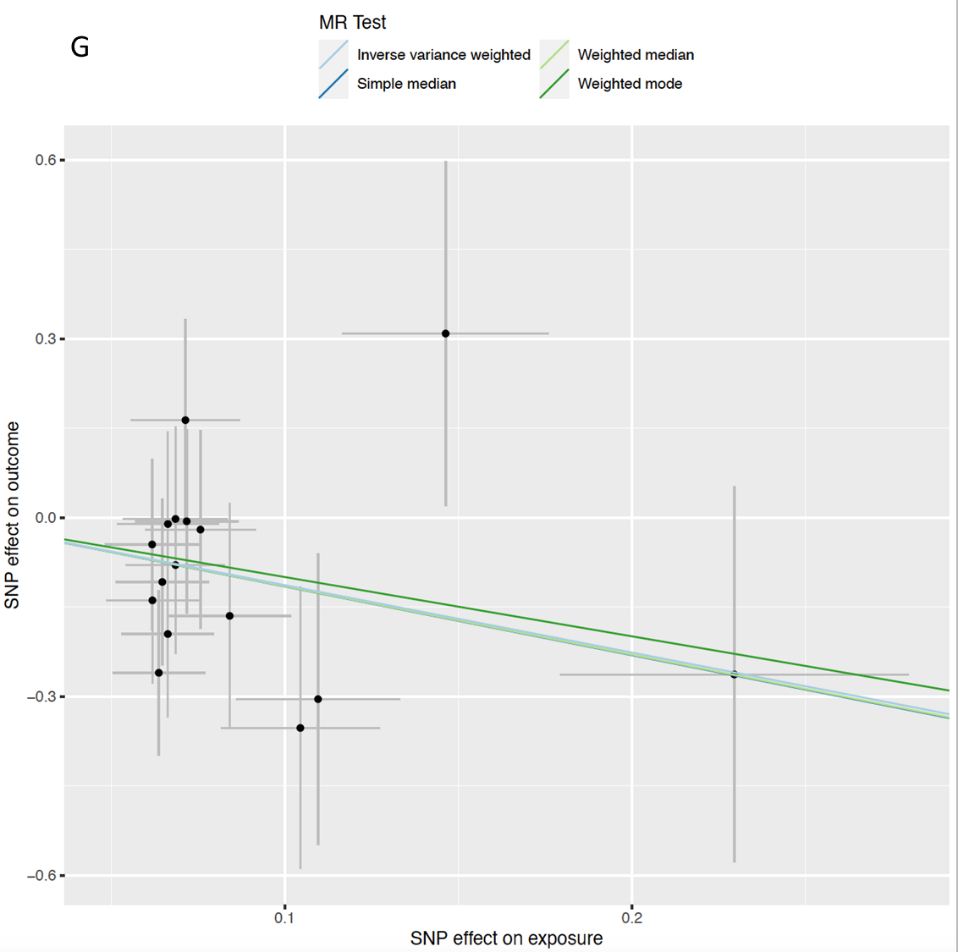


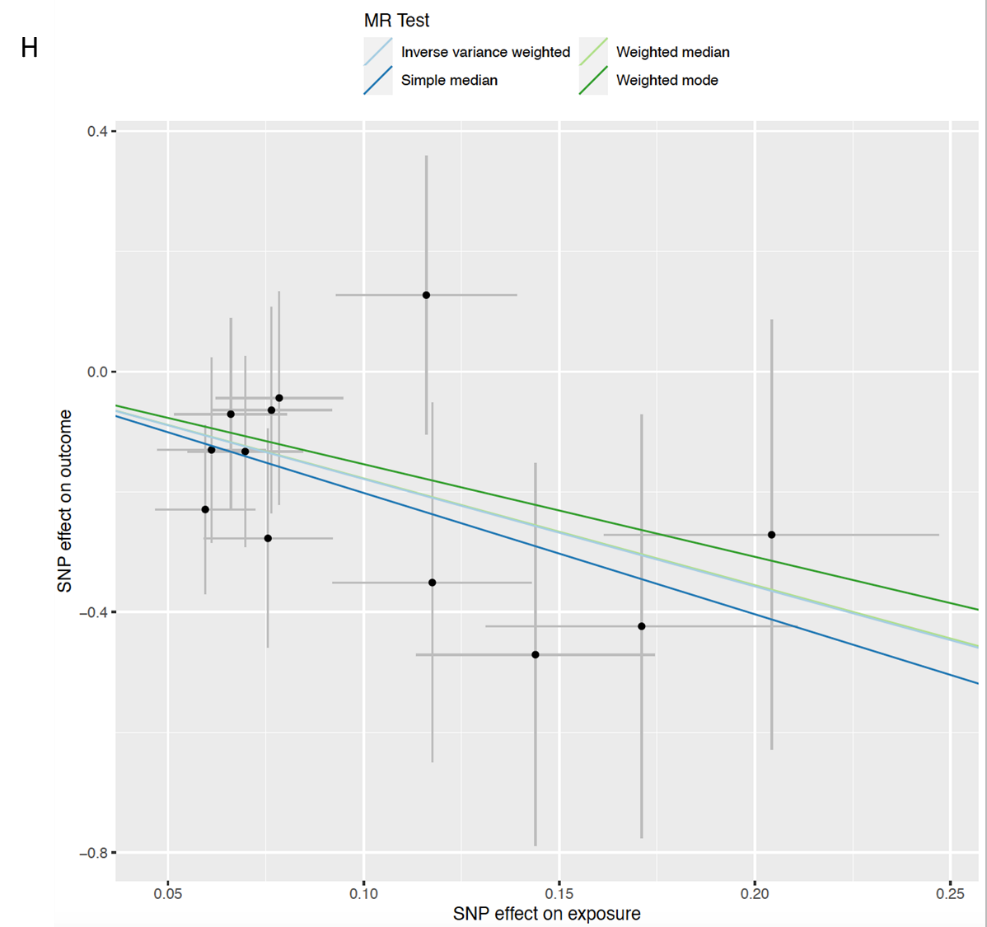
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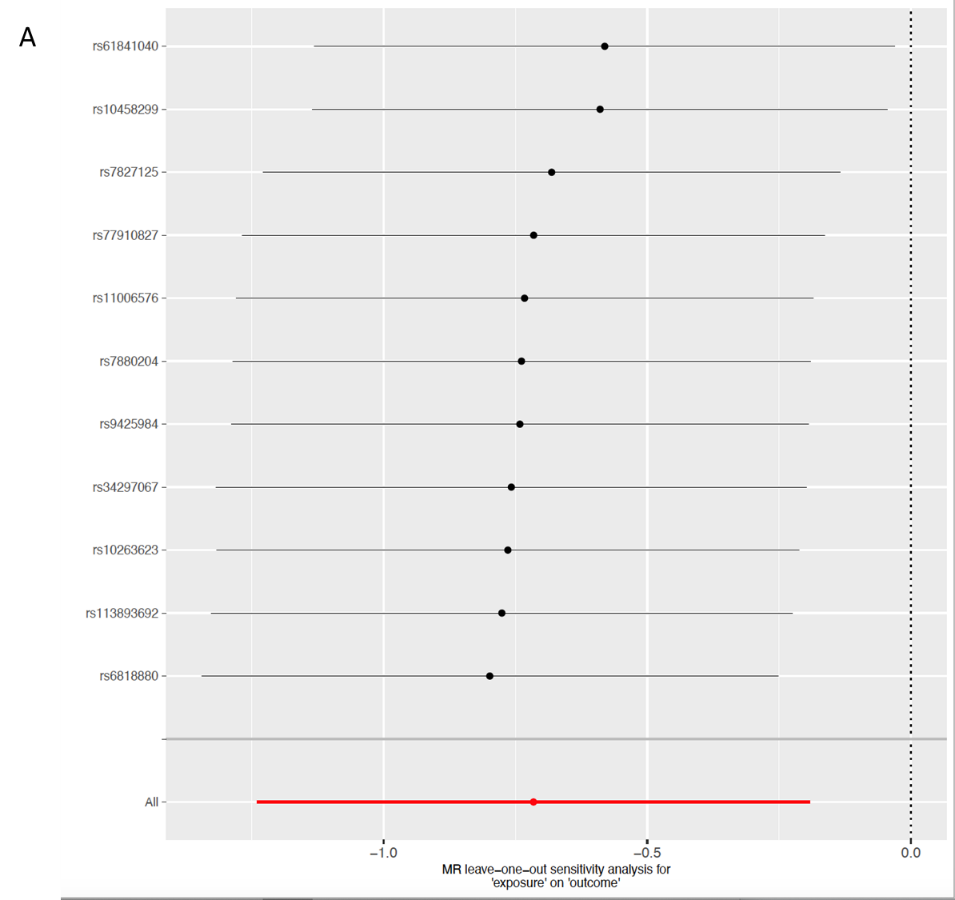
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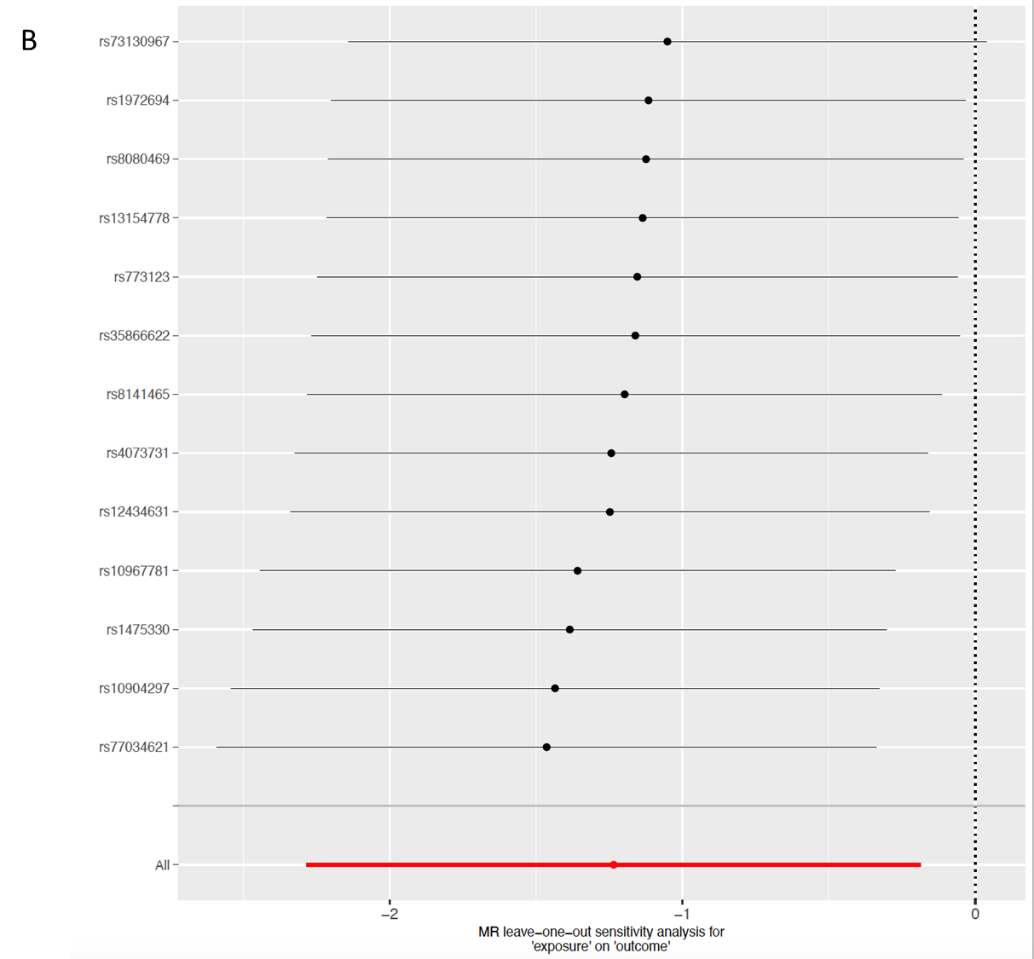
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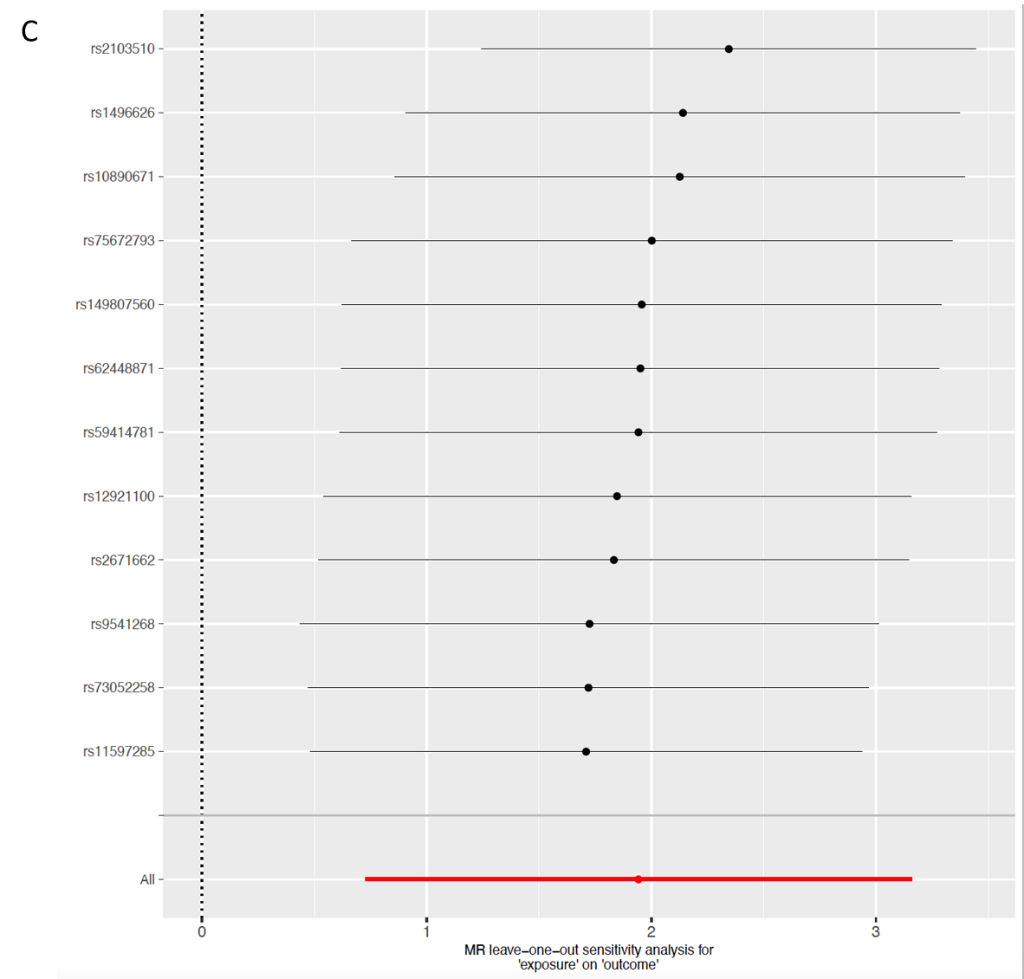
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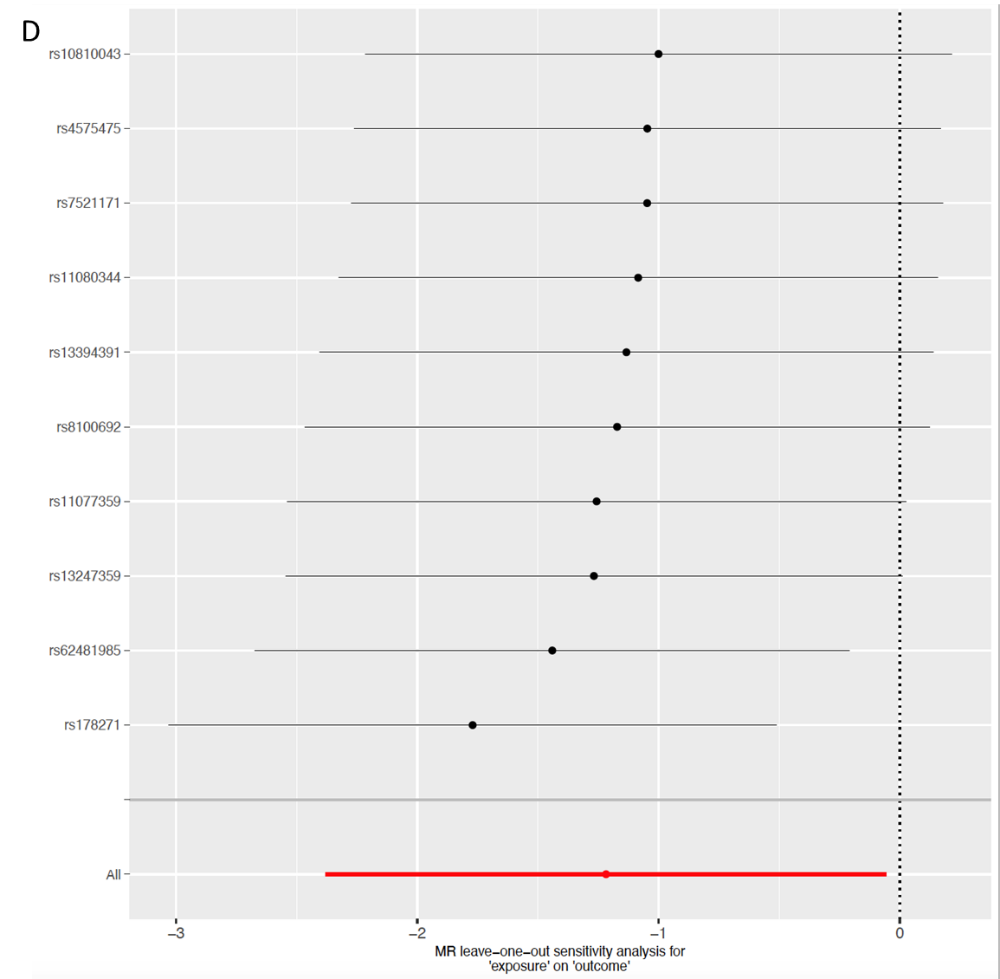
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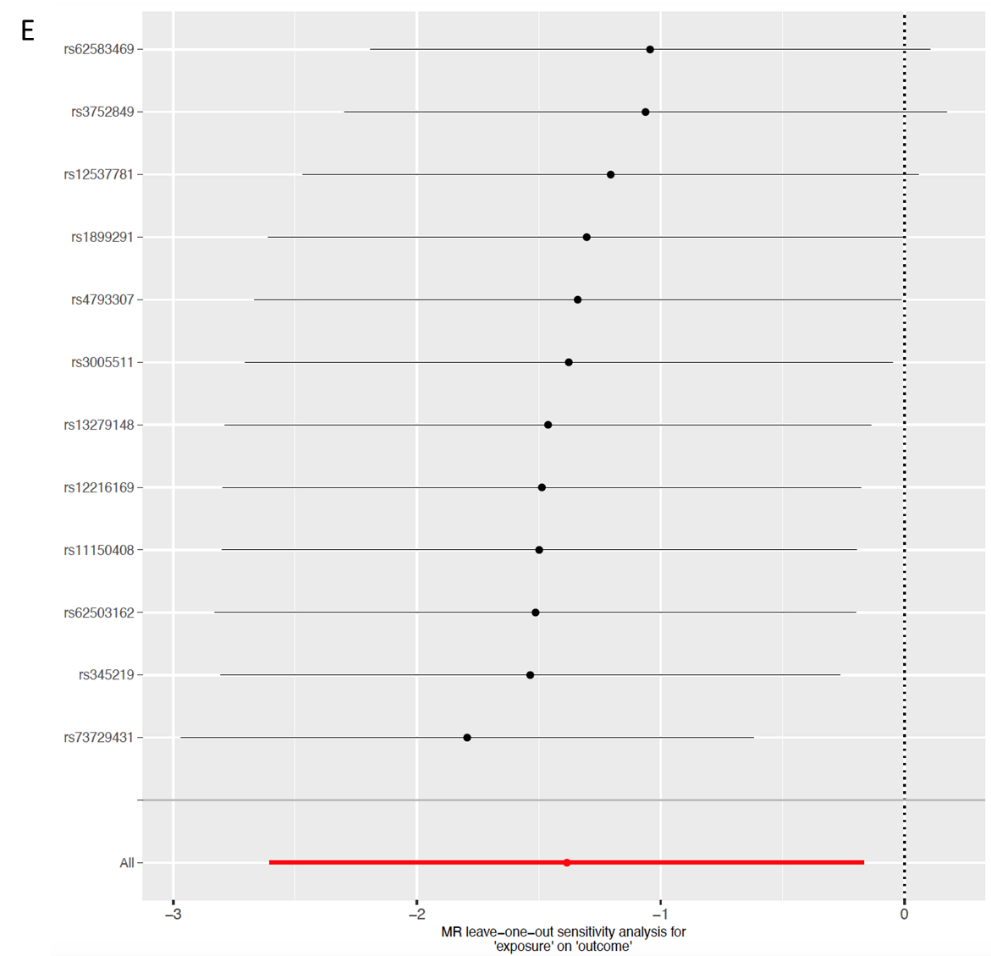
**Figure 4 Scatter plots of causal estimates of specific gut microbiota taxa on intrahepatic cholangiocarcinoma.** The slope of each line corresponding to the estimated Mendelian randomization effect in different models, including the conventional inverse variance weighted, weighted median, simple mode, and weighted mode. A: *Family* *Veillonellaceae*; B: *Genus* *Alistipes*; C: *Order* *Enterobacteriales*; D: *Phylum* *Firmicutes*; E: *Genus* *Anaerostipes*; F: *Genus* *Paraprevotella*; G: *Genus* *Parasutterella*; H: *Phylum* *Verrucomicrobia*. MR: Mendelian randomization; SNP: Single nucleotide polymorphisms.

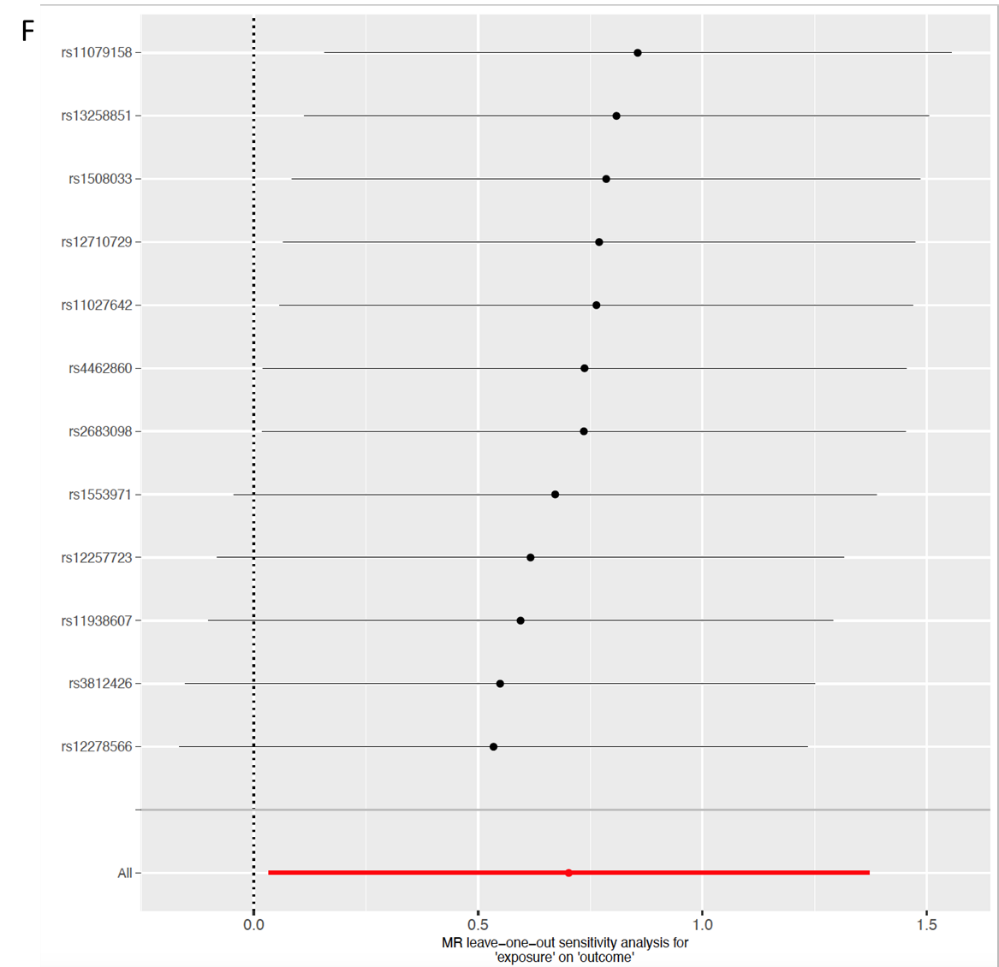


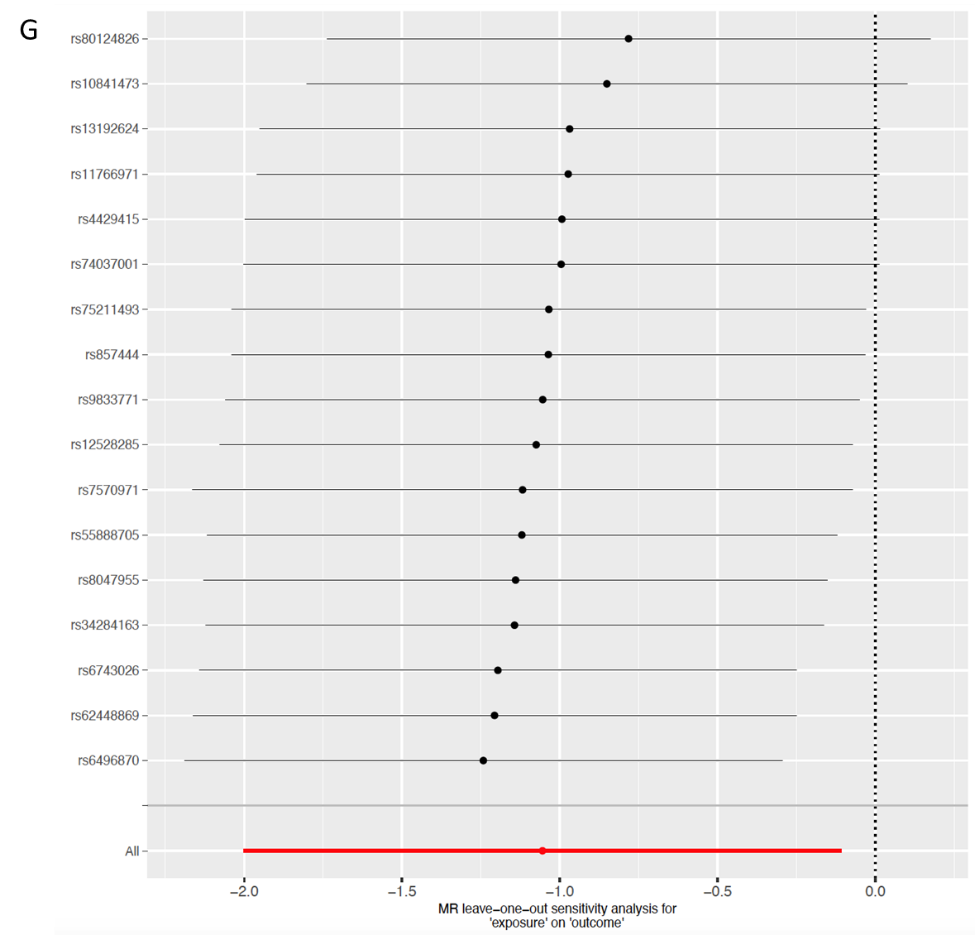




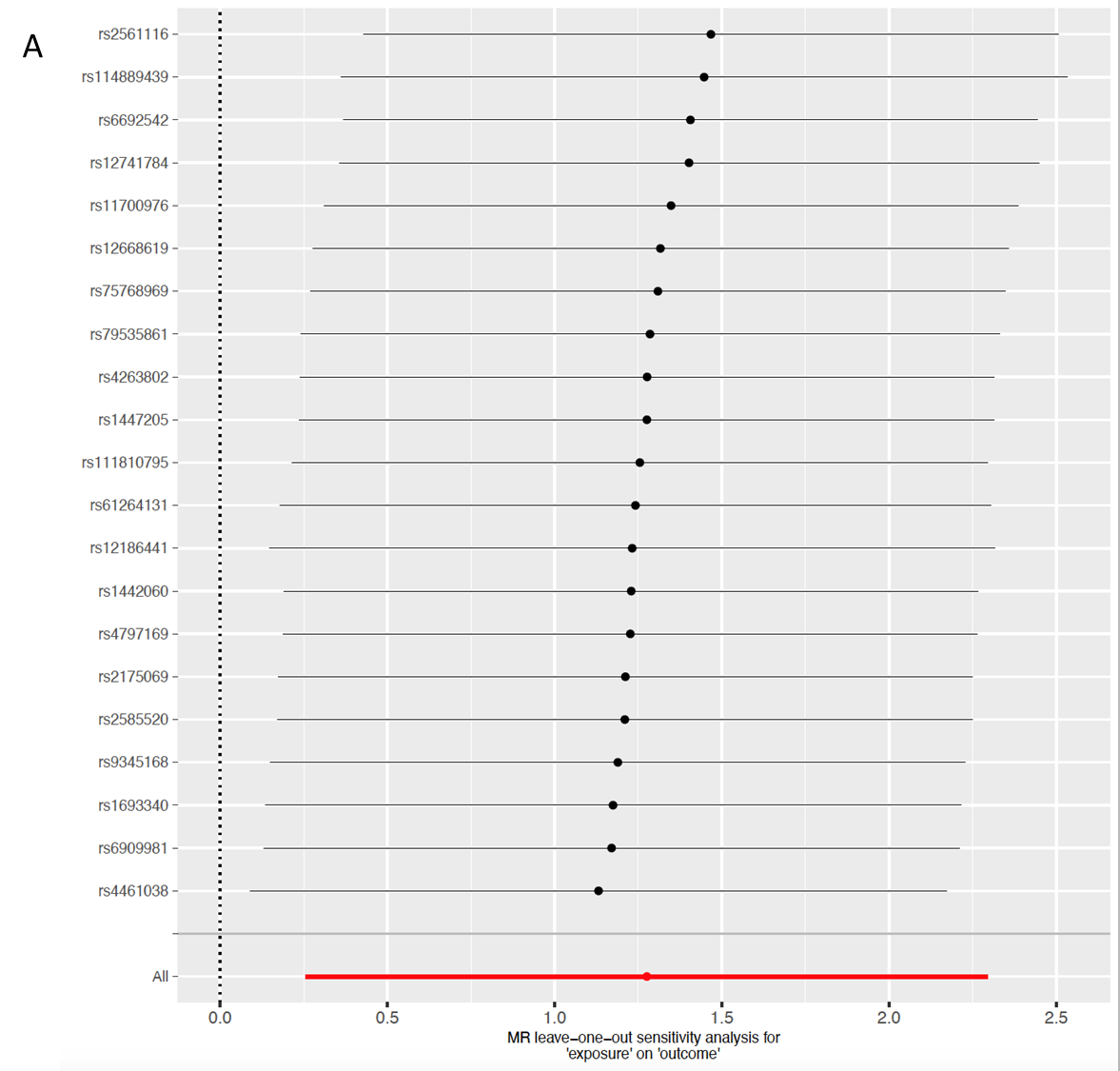


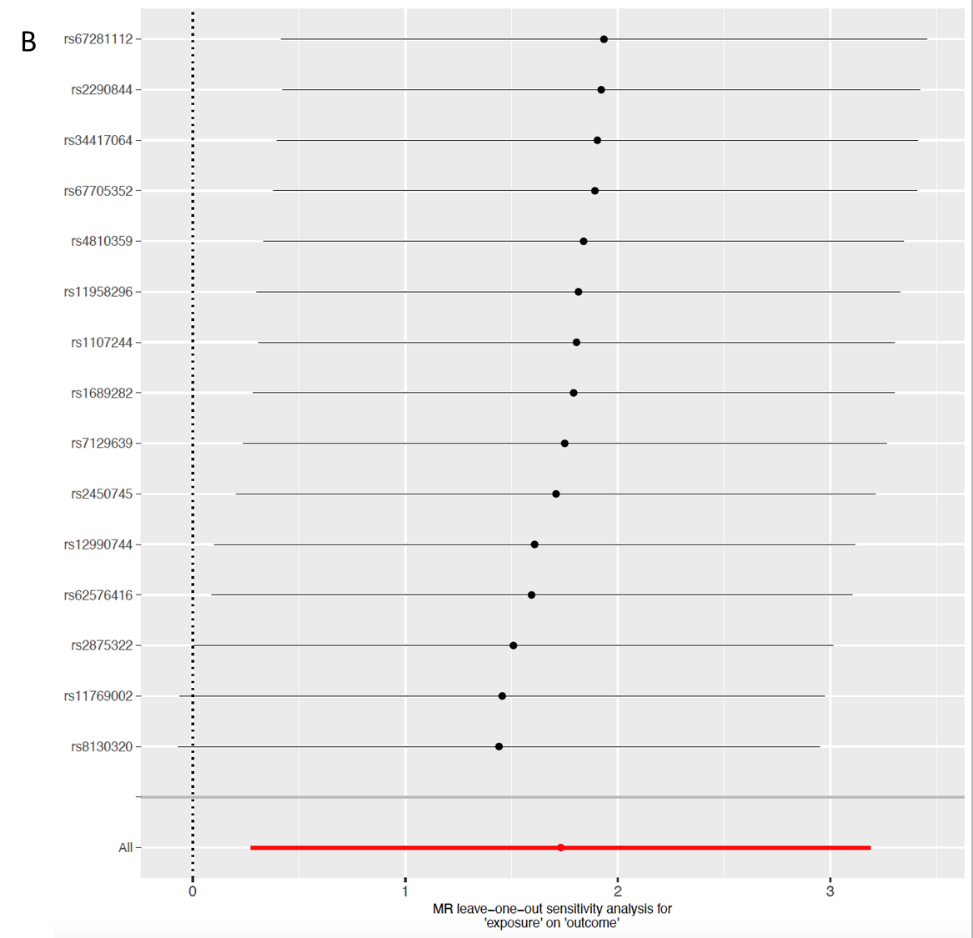


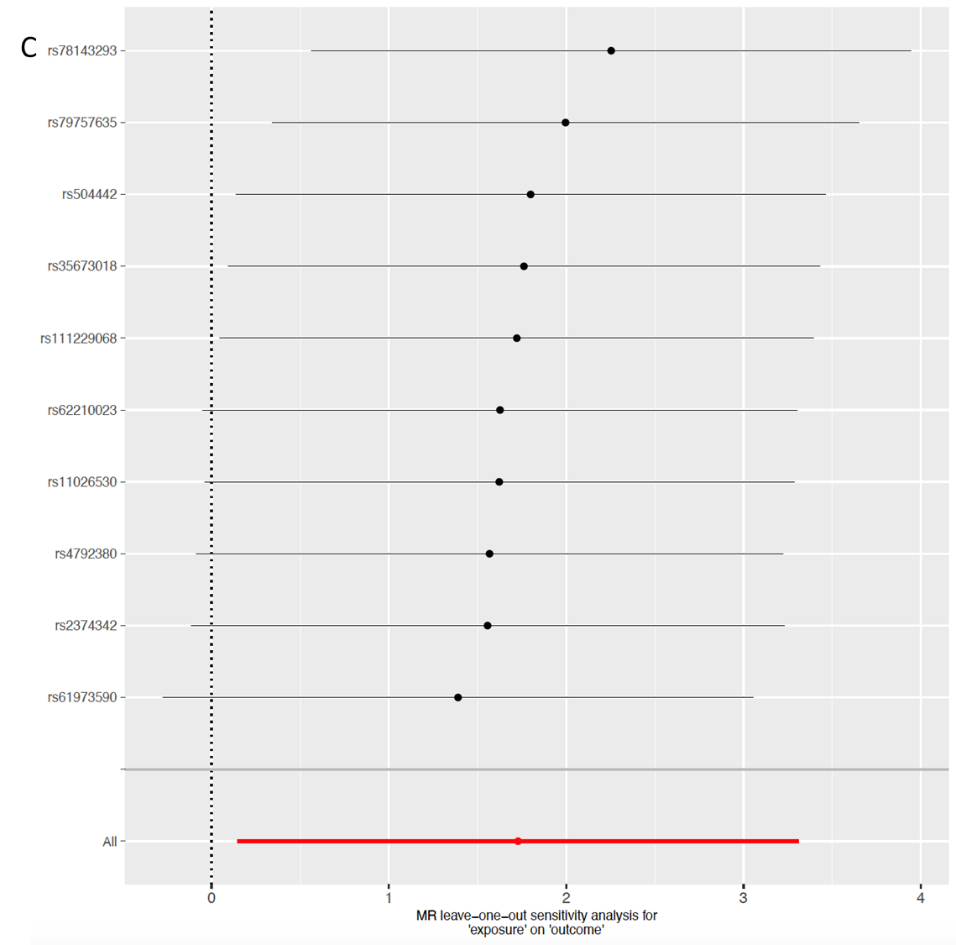
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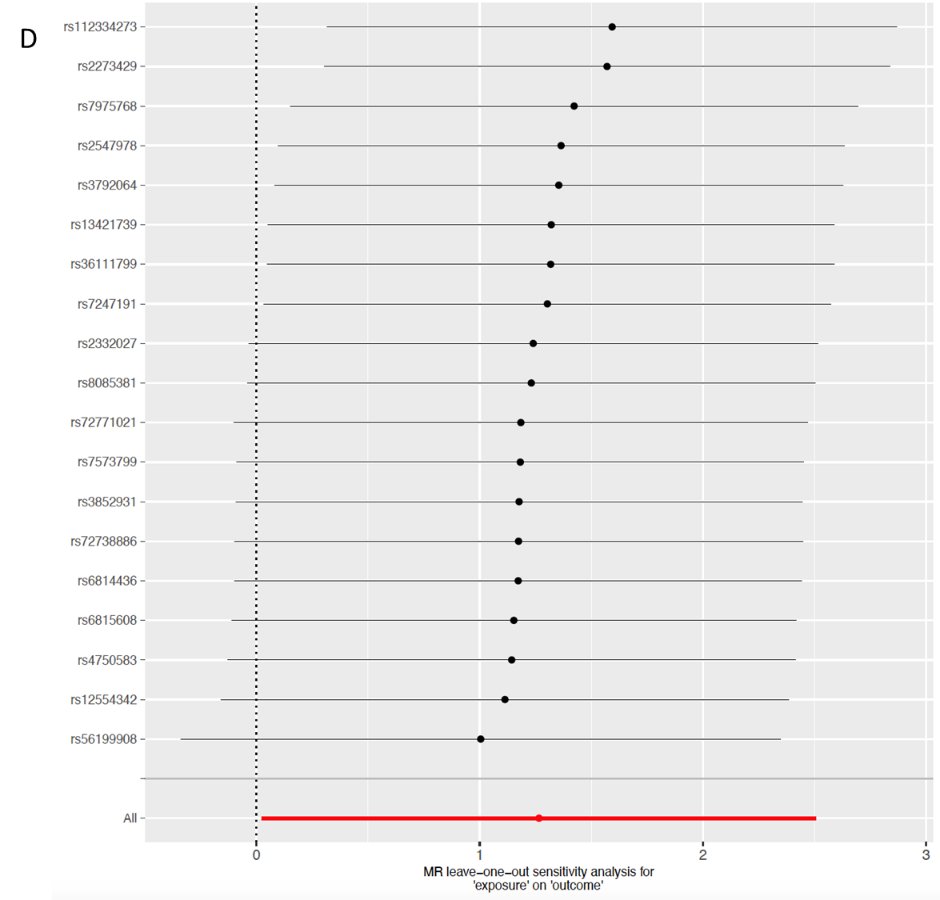
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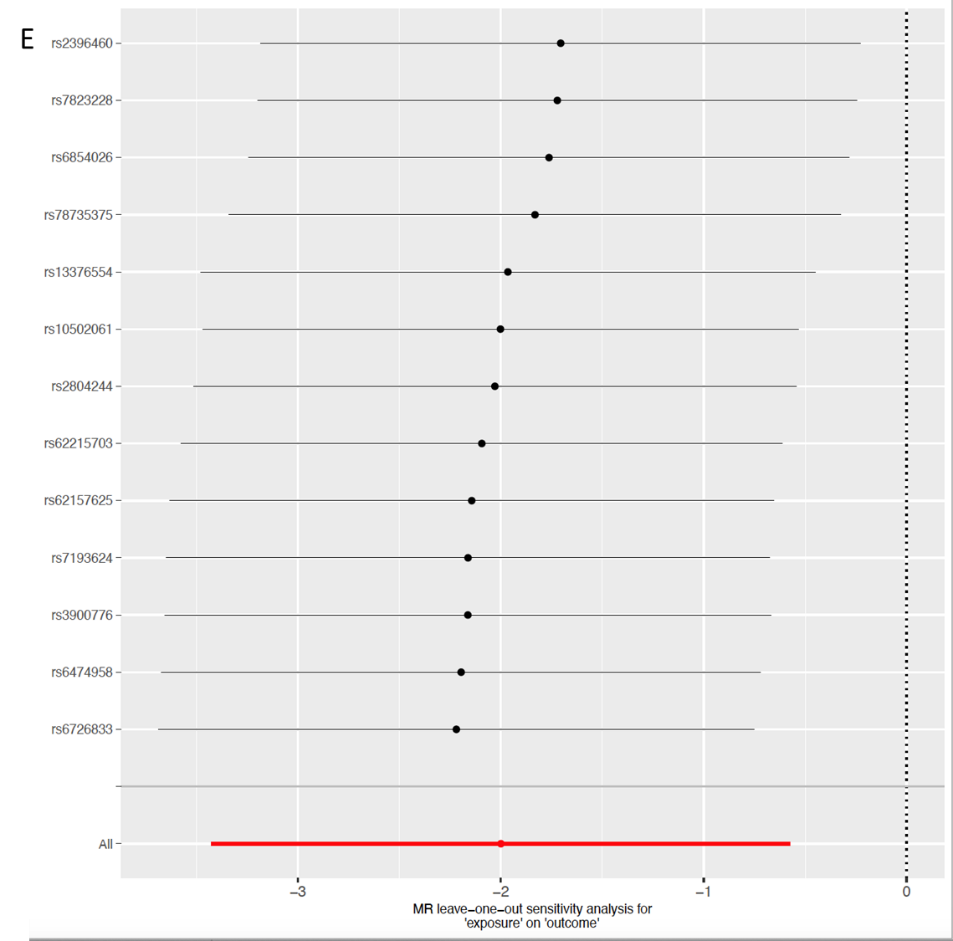
**Figure 5 Leave-one-out stability tests causal estimates of specific gut microbiota taxa on gallbladder cancer and extrahepatic cholangiocarcinoma.** Calculate the Mendelian randomization results of the remaining single nucleotide polymorphisms (SNPs) after removing the SNP one by one. A: *Genus Eubacteriumnodatum group*; B: *Genus* *Ruminococcustorques* *group*; C: *Genus* *Collinsella*; D: *Genus* *Coprococcus*; E: *Genus* *Dorea*; F: *Genus* *Eisenbergiella*; G: *Phylum* *Actinobacteria.* MR: Mendelian randomization; SNP: Single nucleotide polymorphisms.

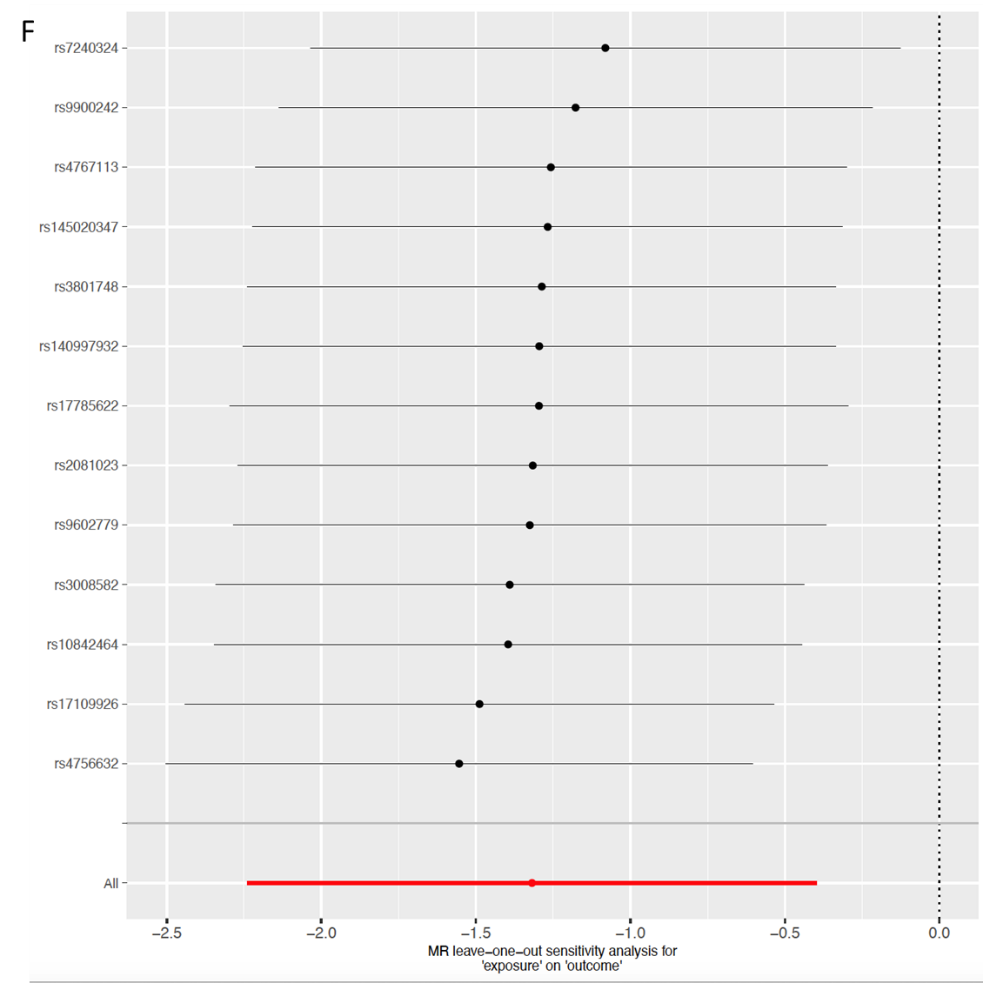


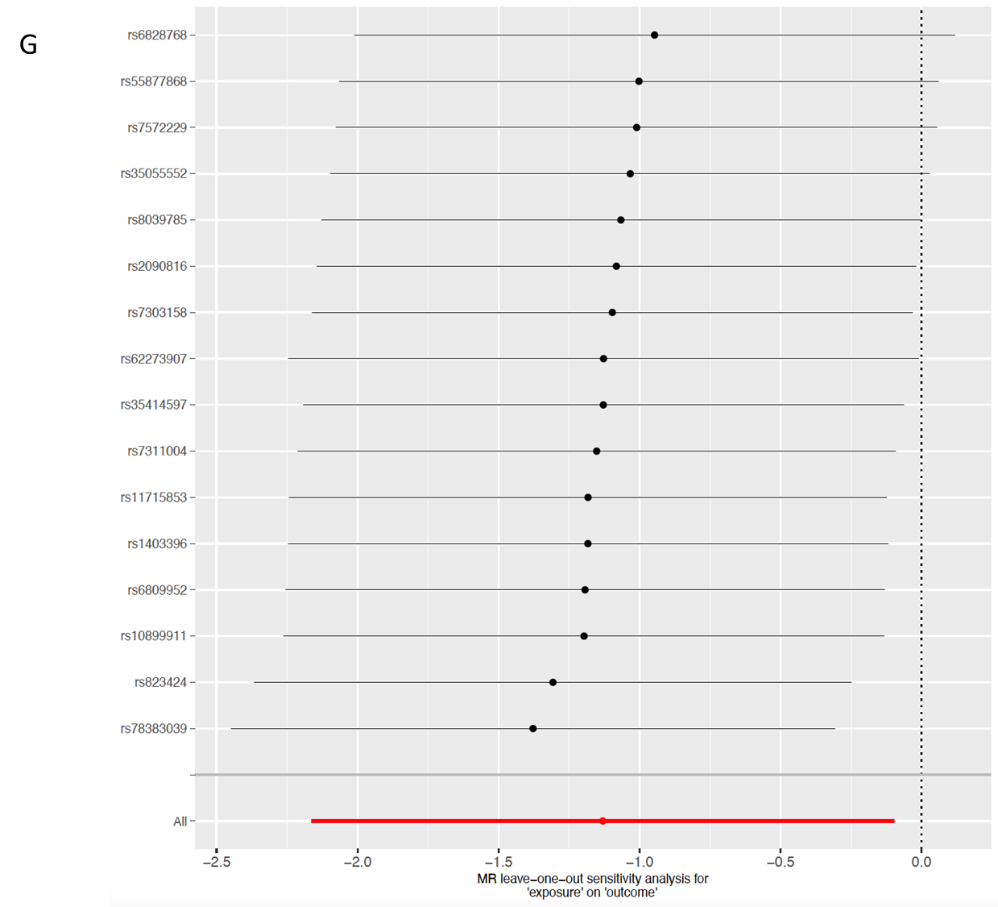


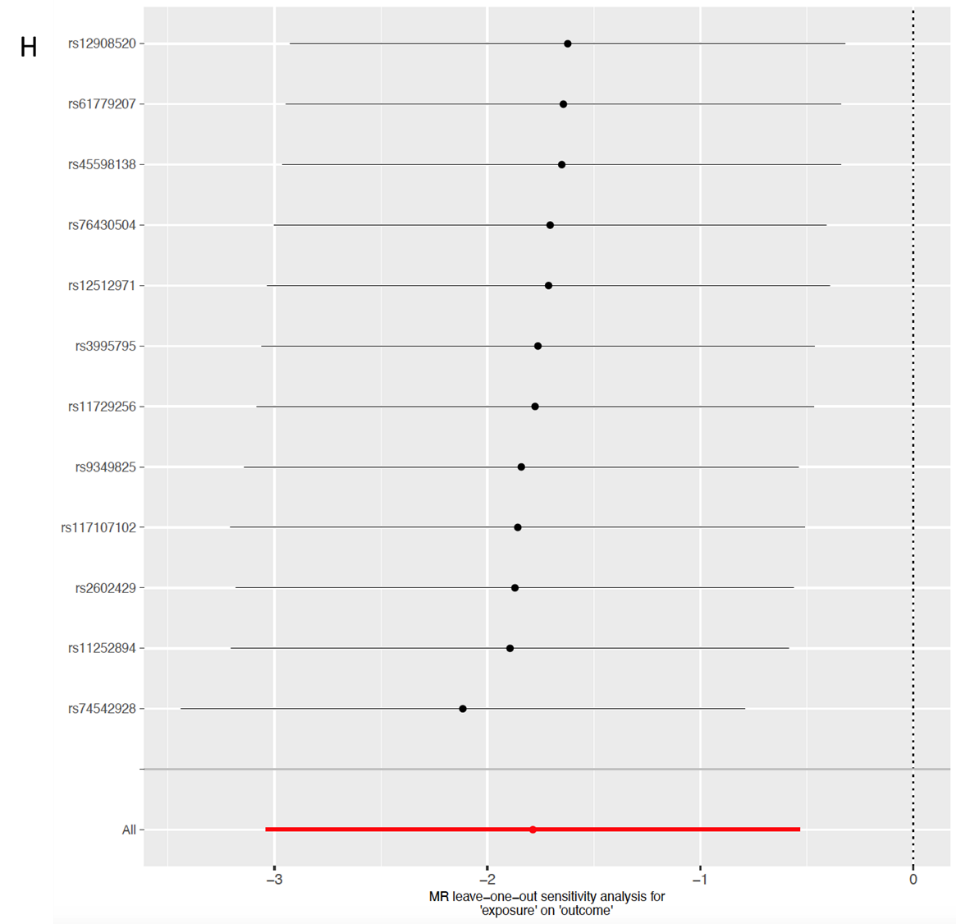


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**Figure 6 Leave-one-out stability tests causal estimates of specific gut microbiota taxa on intrahepatic cholangiocarcinoma.** Calculate the mendelian randomization results of the remaining single nucleotide polymorphisms (SNPs) after removing the SNP one by one.A: *Family* *Veillonellaceae*; B: *Genus* *Alistipes*; C: *Order* *Enterobacteriales*; D: *Phylum* *Firmicutes*; E: *Genus* *Anaerostipes*; F: *Genus* *Paraprevotella*; G: *Genus* *Parasutterella*; H: *Phylum* *Verrucomicrobia*. MR: Mendelian randomization; SNP: Single nucleotide polymorphisms.

**Table 1 Heterogeneity and pleiotropy analysis of the mendelian randomization study on gut microbiota and cholangiocarcinoma**

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| **Exposure** | **Outcome** | **Method** | **Heterogeneity** | | **Horizontal pleiotropy** | | **MR-PRESSO** |
| **Q** | **Q, *P* value** | **Egger intercept** | ***P* value** | ***P* value** |
| *Verrucomicrobia* | iCCA | MRE | 4.795 | 0.904 | -0.048 | 0.762 | 0.951 |
| IVW | 4.892 | 0.936 |  |  |  |
| *Firmicutes* | iCCA | MRE | 16.151 | 0.513 | -0.117 | 0.342 | 0.527 |
| IVW | 17.108 | 0.516 |  |  |  |
| *Enterobacteriales*/*Enterobacteriaceae* | iCCA | MRE | 5.600 | 0.692 | 0.231 | 0.433 | 0.725 |
| IVW | 6.282 | 0.711 |  |  |  |
| *Parasutterella* | iCCA | MRE | 9.901 | 0.769 | -0.062 | 0.638 | 0.805 |
| IVW | 10.133 | 0.811 |  |  |  |
| *Paraprevotella* | iCCA | MRE | 10.402 | 0.495 | -0.057 | 0.740 | 0.619 |
| IVW | 10.518 | 0.571 |  |  |  |
| *Anaerostipes* | iCCA | MRE | 9.312 | 0.593 | -0.095 | 0.526 | 0.679 |
| IVW | 9.740 | 0.639 |  |  |  |
| *Alistipes* | iCCA | MRE | 8.587 | 0.803 | 0.203 | 0.373 | 0.808 |
| IVW | 9.437 | 0.802 |  |  |  |
| *Veillonellaceae* | iCCA | MRE | 13.399 | 0.818 | 0.037 | 0.672 | 0.850 |
| IVW | 13.584 | 0.851 |  |  |  |
| *Eubacteriumnodatum group* | GC and eCCA | MRE | 6.622 | 0.676 | 0.106 | 0.555 | 0.755 |
| IVW | 6.997 | 0.726 |  |  |  |
| *Ruminococcustorques group* | GC and eCCA | MRE | 6.527 | 0.836 | -0.113 | 0.309 | 0.812 |
| IVW | 7.665 | 0.811 |  |  |  |
| *Collinsella* | GC and eCCA | MRE | 14.609 | 0.147 | 0.024 | 0.894 | 0.229 |
| IVW | 14.636 | 0.200 |  |  |  |
| *Coprococcus* | GC and eCCA | MRE | 4.421 | 0.817 | -0.311 | 0.068 | 0.442 |
| IVW | 8.877 | 0.449 |  |  |  |
| *Dorea* | GC and eCCA | MRE | 13.383 | 0.203 | -0.031 | 0.820 | 0.304 |
| IVW | 13.456 | 0.265 |  |  |  |
| *Eisenbergiella* | GC and eCCA | MRE | 10.563 | 0.393 | -0.052 | 0.857 | 0.493 |
| IVW | 10.599 | 0.477 |  |  |  |
| *Actinobacteria* | GC and eCCA | MRE | 14.593 | 0.481 | 0.182 | 0.153 | 0.406 |
| IVW | 16.855 | 0.395 |  |  |  |

MR: Mendelian randomization; CCA: Cholangiocarcinoma;iCCA:Intrahepatic cholangiocarcinoma; GC:Gallbladder cancer; eCCA: Extrahepatic cholangiocarcinoma; *Q*: Cochran’s *Q* test; MRE: MR egger; IVW: Inverse variance weighted.