**Name of Journal:** *World Journal of Hepatology*

**Manuscript NO:** 68457

**Manuscript Type:** MINIREVIEWS

**Immunomodulation by probiotics and prebiotics in hepatocellular carcinoma**

Russo E *et al*. Immunomodulation in hepatocellular carcinoma

Edda Russo, Camila Fiorindi, Francesco Giudici, Amedeo Amedei

**Edda Russo, Francesco Giudici, Amedeo Amedei,** Department of Experimental and Clinical Medicine, University of Florence, Florence 50134, Tuscany, Italy

**Camila Fiorindi,** Department of Health Professions, Dietary Production Line and Nutrition, University Hospital of Careggi, Florence 50134, Italy

**Author contributions:** Russo E, Giudici F, Fiorindi C and Amedei A conceptualized the review; Russo E and Fiorindi C wrote the paper, collected the data and prepared the figures; Giudici F and Amedei A corrected the final version; Russo E, Giudici F, Amedei A were responsible for funding acquisition; and all authors critically revised the manuscript.

**Supported by** a grant from ECCO 2021 to Francesco Giudici.

**Corresponding author: Amedeo Amedei, PhD, Professor,** Department of Experimental and Clinical Medicine, University of Florence, Complesso Cubo, Viale Pieraccini 6, Florence 50134, Tuscany, Italy. amedeo.amedei@unifi.it

**Received:** May 27, 2021

**Revised:** July 21, 2021

**Accepted: January 25, 2022**

**Published online:**

**Abstract**

Hepatocellular carcinoma (HCC) is the most prevalent primary malignancy in patients suffering from chronic liver diseases and cirrhosis. Recent attention has been paid to the involvement of the gut-liver axis (GLA) in HCC pathogenesis. This axis results from a bidirectional, anatomical and functional relationship between the gastrointestinal system and the liver. Moreover, the complex network of interactions between the intestinal microbiome and the liver plays a crucial role in modulation of the HCC-tumor microenvironment, contributing to the pathogenesis of HCC by exposing the liver to pathogen-associated molecular patterns, such as bacterial lipopolysaccharides, DNA, peptidoglycans and flagellin. Indeed, the alteration of gut microflora may disturb the intestinal barrier, bringing several toll-like receptor ligands to the liver thus activating the inflammatory response. This review explores the new therapeutic opportunities that may arise from novel insights into the mechanisms by which microbiota immunomodulation, represented by probiotics, and prebiotics, affects HCC through the GLA.

**Key Words:** Hepatocellular carcinoma; Gut microbiota; Probiotics; Prebiotics; Gut-liver axis; Immunomodulation

Russo E, Fiorindi C, Giudici F, Amedei A. Immunomodulation by probiotics and prebiotics in hepatocellular carcinoma. *World J Hepatol* 2022; In press

**Core Tip:** In patients with chronic liver disease and cirrhosis, hepatocellular carcinoma (HCC) is the most common primary malignancy. Recent attention has been paid to the involvement of the gut-liver axis (GLA) in HCC pathogenesis. This review explores the potential for new treatment options as a result of novel insights into the processes by which microbiota immunomodulation, represented by probiotics and prebiotics, affects HCC through the GLA.

**INTRODUCTION**

Hepatocellular carcinoma (HCC) is the leading cause of cancer-related death worldwide and it is the most common primary tumor in people with cirrhosis and chronic liver disease[1]. Males are diagnosed with HCC at a greater rate than females (2.4:1) in Eastern and Southern Asia, Middle and Western Africa, Micronesia/Polynesia and Melanesia[2]. Today, infection with hepatitis B virus (HBV) and hepatitis C virus (HCV), as well as dietary aflatoxin and alcohol abuse, are all significant risk factors for HCC occurrence. Despite the fact that HBV and HCV account for 80%-90% of total HCC cases, the obesity epidemic, the development of effective direct acting antivirals for HCV, and the availability of a universal HBV vaccination may alter HCC epidemiology in the future[3].

Recently, an increase in the incidence of non-alcoholic steatohepatitis (NASH) and non-alcoholic fatty liver disease (NAFLD) has been accompanied by an increase in the incidence of NASH-related HCC[4]. Aristolochic acid and tobacco have also been identified as probable pathogenetic cofactors in HCC, according to several findings on mutational signatures[5].

Specific advances in our understanding of the processes related to NASH-associated HCC have offered new insights into the tumor microenvironment contributions, generated by a mutual interplay between the immune system and gut microbiota (GM), defined as the assemblage of microorganisms such as bacteria, eukaryotes, archaea, and viruses inhabiting the intestine[6-9]. Indeed, important factors linked to the immune-microbiome interplay such as leaky gut, endotoxemia, toll-like receptors (TLR), dysbiosis, and immunomodulation have been associated with HCC development[10]. Although the liver does not come into direct contact with bacteria, it is anatomically connected to the gut[11]. Of note, the gastrointestinal tract influences homeostasis, preserving an intact barrier against bacterial lipopolysaccharides (LPS) and intestinal bacteria. The most often utilized marker for the translocation of inflammatory bacterial microbiota-associated molecular patterns (MAMPs) is LPS.

LPS is an element found in gram-negative bacteria cell walls that triggers inflammation *via* TLR4. Low-grade exposure to GM-derived metabolites and MAMPs occurs during the physiologic transit of nutrient-rich blood from the colon to the liver, despite the extremely effective multi-level intestinal barrier[11]. Bacterial translocation and LPS build up cause intestinal bacterial overgrowth and changes in GM composition when intestinal permeability is enhanced. Furthermore, a leaky gut allows dysbiotic microbiota-associated bacterial metabolites and MAMPs to more easily translocate and reach the liver. Degradation, detoxification, and clearance of LPS and other microbial products are all hampered in patients with chronic liver diseases or cirrhosis, since the damaged liver is exposed to a wider spectrum of TLR ligands, as well as other bacterial products and metabolites[12]. Indeed, it has been noted that altered microbiota is generally present in HCC patients[13]. In addition, unlike bacterial species, it was discovered that the iron transport, microbial metabolism, and energy-producing system of HCC patients and healthy controls varied considerably[13].

TLR4 produced by activated stellate cells reacts to low LPS concentrations, causing fibrosis and cirrhosis development. An animal model of hepatocarcinogenesis showed that the GM and TLR4 activation have been shown to enhance HCC development by increasing cell proliferation and suppressing apoptosis[14].

As the gut-liver axis (GLA) is involved in HCC pathogenesis, the study of the mutual interplay between the microbiota and immune response and their cross-talk with the tumor microenvironment are an important focus of current clinical research. Our review investigates and analyzes the potential therapeutic benefits of emerging insights into the mechanisms by which microbiota immunomodulation, as represented by probiotics and prebiotics, impacts HCC *via* the GLA.

***GLA***

The GM as a “virtual metabolic organ” establishes an axis with several extraintestinal organs, such as the brain, kidneys, bone and cardiovascular system. However, in recent years, the GLA has received considerable interest[15]. The GLA is the result of a bidirectional, anatomical and functional relationship between the gastrointestinal system and the liver, largely *via* the portal circulation (Figure 1). A complex network of interactions between the enteric microbiome and the liver regulates and stabilizes their symbiotic connection, which includes metabolic and immunological crosstalk[16,17]. Antigens (from harmful microorganisms or food) enter through these linkages and are identified by dendritic cells, which then activate the adaptive immune system by regulating T cell responses. Pathogen-associated molecular patterns (PAMPs) (*e.g.,* LPS, DNA and flagellin activate nuclear factor kappa B and peptidoglycans) *via* nod-like receptors and TLRs, as a result, inflammatory cytokines and chemokines are secreted and reach the portal circulation. PAMPs can activate stellate cells implicated in fibrosis development and progression, in addition to hepatocyte injury, and Kupffer cells are much more susceptible to LPS than hepatocytes[18].

***Involvement of the microbiome in HCC***

As previously reported, the GM contribution to HCC etiopathogenesis is complex and elaborate. A disturbed intestinal barrier brings a series of TLR ligands to the liver and activates the inflammatory response in different ways: (1) *Via* upregulation of hepatic stellate cell proliferation and downregulation of hepatocyte apoptosis, the TLR signaling pathways induce liver tumorigenesis[14]; and (2) Lastly, in HCC, inadequate immunosurveillance is linked to an aberrant intestinal microbiota. Furthermore, *via* increasing oxidative stress, inflammatory response and steatosis, the microbiota dysbiosis might be linked to HCC development[19].

In general, changes in the makeup of microbial profiles are thought to have a role in tumorigenesis[9]. In fact, recent research has revealed a link between certain bacterial profiles and HCC patients[20], showing high amounts of *Escherichia coli* and other gram-negative bacteria in the intestinal bacterial flora, which are linked to elevated LPS levels in serum[21]. Moreover, *Fusobacterium* and *Oribacterium* are the bacteria most often identified from a tongue swab of HCC affected subjects. On the other hand, the intestinal HCC microbiome showed reduced amounts of *Lactobacillus spp.*, *Bifidobacterium spp.*, and *Enterococcus spp.*[22]. A more recent report examined bacterial diversity in cirrhosis and HCC patients[20]. A decrease in the fecal microbial diversity from healthy controls to cirrhosis and an increase from cirrhosis to early HCC with cirrhosis (both induced by chronic HBV infection) was observed. In addition, different microbiota markers were detected[20]. Moreover, the authors observed a decrease in *Verrucomicrobia* with a simultaneous increase in *Actinobacteria*.

Furthermore, augmented levels of *Bacteroides* and *Ruminococcaceae* and increased levels of *Akkermansia* and *Bifidobacterium* were detected in patients with NASH-induced cirrhosis and HCC, in comparison to NASH-induced cirrhosis without HCC[23]. In the same study, the authors discovered a link between enteric microbiota patterns and calprotectin levels as well as systemic inflammation. The GM of patients with HBV-associated HCC and non-HBV non-HCV (NBNC) associated HCC was compared in further research. In comparison to healthy controls or to patients with NBNC-related HCC, those with HBV-related HCC had a substantially richer fecal microbiota. Patients with NBNC-related HCC had higher amounts of pro-inflammatory bacteria (*Escherichia coli*, *Enterococcus*) and lower amounts of anti-inflammatory bacteria (*Faecalibacterium*, *Ruminococcus*, *Ruminoclostridium*), leading to lower quantities of anti-inflammatory short-chain fatty acids (SCFA) in their feces[24]. The GM of HCC patients receiving liver transplantation was also compared to the intestinal microbiota of individuals who did not have HCC but had a similar etiology of cirrhosis. An augmented abundance of fecal *Escherichia coli* was linked with HCC[25]. Additionally, *Helicobacter spp.*, was found in liver HCC suggesting that intestine translocation might be a possible cause of carcinogenesis[26]. To this end, the enteric microflora profile might potentially indicate reaction rates in HCC patients undergoing treatment with immune checkpoint inhibitors[27], suggesting that the microbiome could be used for liver cancer immunotherapy[28].

Overall, alterations in microbial profiles in HCC did not appear to be consistent among investigations, presumably due to differences in etiologies, geographical locations, and dietary intakes. Differences between patients with cirrhosis and HCC, as well as cirrhosis alone, appear to be lower than those between healthy individuals and those with cirrhosis. As a result, microbiome-based diagnostic tools are anticipated to be more potent for cirrhosis identification than for HCC diagnosis. However, rather than particular HCC-associated abnormalities, the microbiome's functional effect on HCC development is more likely to be linked to cirrhosis-related changes, which may increase HCC advancement.

***Therapeutic impact of probiotics in HCC***

A recent study has shown that antibiotics can help with small-intestinal bacterial overgrowth-related liver damage, demonstrating the connection between intestinal microbiota and liver disorders[29]. Probiotics are microorganisms that have a beneficial effect on humans and are presently being studied as a potential therapy for chronic liver disease[16,29]. Indeed, they can support the growth of enteric microbes producing anti-inflammatory metabolites, which exert an immune suppressive effect.

Probiotics can help microbiota generate anti-inflammatory compounds with tumor-suppressing properties. Probiotics have a significant influence on the GLA, with anti-inflammatory and immunomodulatory effects on GM and gut barrier function, as well as a metabolic influence on organs beyond the gastrointestinal tract. They can affect the generation of immunomodulating GM compounds that show anti-tumor properties (Figure 2). Furthermore, supplementation with probiotics increased the expression of some anti-inflammatory cytokines, such as interleukin (IL)-13, IL-10, and IL-27.

Downregulation of angiogenic factors and receptors, VEGFA, Fms related receptor tyrosine kinase 1, ANGPT2, and kinase insert domain receptor, were seen in mice fed a particular probiotic combination[30]. However, not all probiotic species have the same immunomodulatory effect on the gut microflora. Different strains of *Lactobacillus spp.*, for example, are linked to both pro-obesity and anti-obesity effects[31]. Research on mice with diet-induced obesity found that distinct targeted enteric bacteria modification using vancomycin *vs* probiotic strain *Lactobacillus salivarius* resulted in divergent metabolic effects, although microbiota modifications were identical. By contrast, a recent human investigation in obese adolescents focussing on the impact of *Lactobacillus salivarius Ls*-33 on a number of inflammatory biomarkers, found no evidence of an effect on the metabolic syndrome[32,33]. These findings indicate that the role of the same probiotics could be different due to various conditions; therefore, attention should be paid to evaluating the different impacting factors.

The use of antimicrobials and probiotics in chronic liver illnesses should be based on the GLA pathophysiology. Despite a consistent number of findings from animal and human research, further case-control prospective studies with a large number of patients are needed to fully understand this issue. Obesity-induced intestinal microbial dysbiosis can lead to HCC, according to a mouse model[34]. Aflatoxin-induced HCC is the subject of research in relation to therapeutic HCC prevention with probiotics. In fact, aflatoxin contamination of foods is an etiological risk factor for HCC in underdeveloped nations. Probiotics may help to suppress aflatoxin B-induced hepatocarcinogenesis, correct gut dysbiosis, lower LPS levels, and shrink tumors, according to findings from human and animal model studies[14,35]. Moreover, in a mouse model, treatment with *Lactobacillus plantarum* (*L* *plantarum*) C88 (isolated from Chinese traditional fermented meals) boosted fecal aflatoxin B1 excretion and controlled the defence system's antioxidant deficiency[35]. Additionally, a probiotic yogurt containing *Lactobacillus rhamnosus*, *Streptococcus thermophilus*, and *Wiessella cibaria* also reduced aflatoxin metabolites in the urine of children who ate aflatoxin B1-infected maize.

The binding of aflatoxin metabolites to probiotic bacteria may have resulted in lower intestinal absorption, inducing a reduction in aflatoxin metabolites in urine[36].

In addition, the ability of probiotic bacteria to facilitate epigenetic modification of host gene expression is advantageous in reducing HCC development[37]. The crosstalk between host and enteric microflora, where gene expression is controlled by several methods such as DNA methylation and histone modification, demonstrates bacterial control of host gene expression[38]. In mice treated with a colon carcinogen, it has been observed that *Lactobacillus acidophilus* (*L.* *acidophilus*) and *Bifidobacterium bifidum* can decrease the expression of Kristen rat sarcoma viral oncogene homolog, oncomirs (microRNA-221and microRNA 155) and the oncogenes BCL2-like 2 (Bcl-w) in the liver. Moreover, mice treated with these probiotic bacteria had higher levels of the tumor suppressor microRNA-122 and the tumor suppressor gene transcription factor PU.1.[37]. Probiotic supplementation may minimize the incidence of HCC by safeguarding the hepatocyte genome, which is important in the pathophysiology of HCC. Probiotic fermented milk and chlorophyllin were shown to reduce the expression of rasp-21, c-myc, cyclin D1, and Bcl-2 in an HCC rat model, slowing tumor development and volume by 40%. Mice treated with probiotics have significant amounts of fecal *Oscillibacter* and *Prevotella*[39].

Microbial PAMPs can induce liver cancer progression *via* TLR-mediated inflammatory responses, as previously described. The treatment with bacteria having probiotic properties has been demonstrated to decrease the development of HCC in the liver by reducing the expression of TLR-induced inflammation. When rats with induced liver cirrhosis were treated with *L* *plantarum* probiotic bacteria, they showed lower TLR4 expression and less liver damage[40]. In addition, gut sterilization and TLR4 inactivation reduced HCC by 80% to 90%, indicating that they might be used as HCC preventive methods[14].

Furthermore, the antiviral action of probiotic bacteria may slow HCC progression by avoiding persistent HBV infection. The release of HBV surface antigen (HBsAg) was diminished by a cell extract of *Bifidobacterium adolescentis* by reducing the transcription of HBsAg gene, which contrasted the infection. Despite the fact that the amount of HBV DNA in the cells did not change considerably, probiotic therapy drastically reduced the amount of extracellular HBV DNA available[41]. Furthermore, treatment with *Lactobacillus bulgaricus* induced lowered viral load and cellular deterioration[42]. On the other hand, probiotic bacteria supplements can also help liver function during HCV infection. In addition, HCV-positive patients, treated with *Enterococcus faecalis* lowered blood levels of the aspartate aminotransferase (AST) and alanine aminotransferase (ALT) (liver damage indicators)[43].

Furthermore, NAFLD is also a key etiological risk factor in HCC development. Supplementation with the probiotic bacteria *L. acidophilus* and *Bifidobacterium lactis* can help NAFLD patients with liver damage, as seen by lower ALT, AST, and total cholesterol blood levels[44]. Probiotic therapy lowered body weight and total body fat content in obese NAFLD patients. By suppressing the pro-inflammatory cytokine tumor necrosis factor-α, probiotics also reduced hepatic inflammation in obese NAFLD patients[45]. Additionally, GM changes seem to be the cause of the hepatoprotective and anti-inflammatory actions of probiotic bacteria in NAFLD patients. In rats fed with a high-sucrose and a high-fat diet, probiotic meals improved diet-induced loss of intestinal microbiota diversity, intestinal epithelial barrier function and colonization resistance. Restoration of enteric microbiota and gut epithelial barrier function reduced NAFLD progression by lowering serum LPS levels and reducing TLR4-mediated hepatic inflammation[36].

Finally, probiotic cell components and metabolites increase the production of tight junction proteins, which helps to maintain gut epithelial integrity. Tight junction integrity is critical for preventing translocation of pathogens or exogenous substances across intestinal epithelia and consequently intestinal inflammation. Constituents of tight junctions comprise integral membrane proteins, such as [occludin](https://www.sciencedirect.com/topics/medicine-and-dentistry/occludin), [claudin](https://www.sciencedirect.com/topics/medicine-and-dentistry/claudin) (CLDN) family members, JAMs 1-3, [cingulin](https://www.sciencedirect.com/topics/biochemistry-genetics-and-molecular-biology/cingulin), and cytoplasmic plaque proteins, such as zonular occludins-1 (ZO-1) and ZO-2[46,47]. Different components of the CLDN family have been reported to be affected during hepatocarcinogenesis[48-51]. However, in HCC patients undergoing hepatectomy, downregulation of ZO-1 is linked to a poor prognosis[52]. Probiotic supplementation can elicit considerable upregulation and relocalization of interepithelial tight junction proteins by activating numerous TLR produced by the intestinal epithelium. In terms of the effect of probiotic therapy on tight junctions, it was discovered that a VSL#3 combination maintained the epithelial barrier in acute colitis by avoiding reduced tight junction protein expression and an increased apoptotic ratio[53]. Moreover, in a mouse model of high-fat diet or obesity-induced liver steatosis, supplementation with a multispecies probiotic (including *Bifidobacteria*, *Lactobacilli* and *Streptococcus*) formulation helped to maintain tight-junction proteins ZO-1 and ZO-2, and reduced hepatic triglyceride concentration compared with a high-fat diet alone[54]. In older rats, a probiotic cocktail including *Lactobacillus* and *Enterococcus* strains reduced microbiota dysbiosis, leaky gut, inflammation, metabolic dysfunctions, and physical function loss caused by a high-fat diet. The GM regulated by probiotics decreased leaky gut by strengthening tight junctions, which lowered inflammation[55].

***Prebiotics as a novel therapeutic approach***

Prebiotics are substrates that are used selectively by host bacteria to provide a health benefit[56]. Health effects are mostly related to the production of SCFA through prebiotics fermentation by a limited number of genera/species of the intestinal microbiota such as *Lactobacilli* and *Bifidobacteria*[57]. *Bifidobacteria* are the most common bacterial species in the human GM, and they are thought to benefit human health by preserving the resident microbiota's balance. SCFAs such as acetate, propionate, and butyrate exert several biologic activities that positively influence the structure and function of the microbial com­munity. Some of these beneficial effects include controlling colonocyte function, promoting water and electrolyte absorption, decreasing the intraluminal pH, inhibiting pathogen proliferation, modifying the immune homeostasis of the gut and modulating the inflammatory processes[56].

The improvement of mucosal barrier functions exerted by the modulation of GM composition improves conditions such as cirrhosis and consequently may prevent HCC. Prebiotics such as butyrate are not metabolized in the mitochondria of tumor cells but enter the cell nucleus to regulate gene expression. In fact, the functions of butyrate including inhibition of histone deacetylase, decrease tumorigenesis (Figure 3)[58].

The most studied prebiotic classes are galactans (galacto-oligosaccharides) and inulin-type fructans [*e.g.,* fructo-oligosaccharides (FOS), inulin, oligofructose]. There are other oligosaccharides such as isomalto-oligosaccharides and soybean oligosaccharides that have recently gained interest. Dietary sources of prebiotic oligosaccharides are various types of vegetables, fruits, grains and legumes. Depending on their water solubility, dietary fibers are classed as insoluble (*e.g.,* hemicellulose, cellulose, and lignin) and soluble (*e.g.,* fructans, gums, and pectins). Only soluble fibers can be metabolized by the GM and have “prebiotic” functions which can affect host health[59].

In particular, the two prebiotics most researched are FOS and inulin, which have been shown to stimulate the growth of beneficial *Bifidobacteria* that are implicated in cancer prevention (Figure 3)[60].

Inulin is a non-digestible functional polysaccharide, present in plants such as onions, chicory, Jerusalem artichoke, leeks, garlic, bananas, rye, barley and wheat[61]. A recent study showed that inulin modulates GM and hepatic fatty acid composition limiting dysbiosis and its negative effect on host health[62].

FOS are mostly present in plants such as garlic, onions, leeks, chicory root, Jerusalem artichokes, asparagus, and bananas. New findings suggest that FOS can help to achieve colonization resistance against *Clostridium difficile*. Several studies have reported the association between liver cancer and intestinal microflora and indicate that *Clostridium* cluster XIVa, *Clostridium* cluster XI, or other strains catalyzing the transformation from primary to secondary bile acids could be triggers of hepatic cancerogenesis[34,63,64]. Moreover, research has shown that *Clostridia* species are enriched in the guts of mice with liver cancer, documenting the association between cancer and dysbiosis[65].

A study evaluating the relationship between food groups and liver cancer risk reported that specific subgroups of vegetables, rich in inulin and FOS, such as celery, mushrooms, Chinese chives, onions, garlic, garlic shoots, asparagus, lettuce, garland chrysanthemum, legumes, squash and carrots, had a strong inverse relationship with liver cancer, indicating their protective effects against HCC[66] (Figure 4). A cohort study involving 125455 individuals from the Nurses' Health Study and the Health Professionals Follow-up Study found that wholegrain, bran, and cereal fiber consumption may help reduce HCC[67].

Moreover, eating whole grains and dietary fiber, particularly cereal fiber, has been linked to a decreased risk of obesity and NAFLD, both of which are known predisposing factors for HCC, as previously documented[68]. In fact, long-term consumption of whole grains has been suggested to reduce the risk of HCC by improving gut integrity and changing GM composition[69]. Moreover, a meta-analysis of 19 studies involving 1290045 participants (3912 with HCC) showed that every 100 g/d increase in vegetable intake decreases the risk of HCC by 8%[70].

A potential role of diet in HCC development has recently been clarified by a systematic review (30 studies involving 5222534 participants) that investigated the association between diet and HCC. In particular, certain dietary patterns, such as the Mediterranean diet, the Urban Prudent Dietary Pattern, the Alternative Healthy Eating Index-2010, and the Traditional Cantonese Dietary Pattern, were found to lower the incidence of HCC[71].

Recently, other substances besides non-digestible oligosaccharides, such as plant polyphenols are considered to have prebiotic effects. These compounds are not absorbed in the small intestine and, therefore, reach the colon where they are processed by the colonic microbiota conferring benefit to the host health[71]. They comprise flavonoids, phenolic acids and lignins found in nuts, tea, wine, vegetables and fruits. Polyphenols, due their immunomodulation activity, can have chemopreventive effects in HCC progression[72]. In particular, polyphenols contained in tea have been reported to have positive effects on GM homeostasis by decreasing the growth of pathogenic bacteria[73]. One of them is the antioxidant ellagic acid, present especially in nuts and berries, which is metabolized by microbiota in the colon into urolithins that have cancer preventive properties[74].

A prospective cohort study found that a higher consumption of tree nuts (hazelnuts, almonds, macadamias, pecans, cashews, and pistachios) (mean 1.25 serving per week) was associated with a reduced risk of HCC[75,76].

Coffee contains phenolic agents which have anticarcinogenic properties and its consumption has been demonstrated by various studies to reduce HCC risk[77].

Moreover, the cancer preventive effect of the association of oligosaccharides with polyphenols was also recently investigated. A study examined the effect of the combination of FOS and pectins with raspberry polyphenols on cecal microbial fermentation and regulation of liver lipid metabolism and inflammation, and concluded that FOS and pectins enhanced the effects of the raspberry polyphenolic extract against disorders related to NAFLD[78].

Furthermore, prebiotics use has been explored as an approach to modulate intestinal barrier function through maintaining tight junction (TJ) integrity. Prebiotics have demonstrated effects on GM composition that could lead to beneficial changes in TJ protein expression and distribution. However, the effect of prebiotics on TJ by microbiota composition modulation is less recognized than those produced by probiotic supplementation.

The prebiotics which showed beneficial effects on intestinal TJs in various studies are FOS, inulin, and galacto-oligosaccharide. A recent study investigated the effect of FOS on AMP-activated protein kinase (AMPK) activity and TJ assembly under non-inflammatory and inflammatory conditions using T84 cells as an intestinal epithelial cell model, and showed that FOS promoted intestinal epithelial TJ assembly through AMPK activation[79]. In addition, other studies demonstrated that prebiotic supplementation results in robust activation of AMPK[80,81].

A study showed that inulin fermentation increased occludin, CLDN-3 and ZO-1 RNA expression that could reinforce the barrier function[82]. Even galacto-oligosaccharide supplementation increased ZO-1, occludin and CLDN-1 protein expression[83].

**CONCLUSION**

HCC development is a complicated process influenced by a number of etiological risk factors. However, several studies have shown that probiotics and prebiotics have a beneficial effect on GM regulation and decrease procarcinogenic factors in the liver. It is noteworthy that the most remarkable aspect is the importance of a balanced diet with proper nutrition (which includes higher intake of vegetables, fish, white meat, and coffee, and reduced intake of fat, red meat, and alcohol), especially for those patients suffering liver, and especially chronic, diseases[84]. A Mediterranean diet has recently been shown to reduce the incidence of HCC, providing a new paradigm for future research[85]. Moreover, scientific investigations have revealed the possibility of developing cancer-preventive symbiotic (combination of probiotic and prebiotic) functional foods[86]. Of course, further research is needed to fully identify the bioactive probiotic metabolites (post-biotics) of certain dietary phytochemicals and to understand the possible mechanism(s) by which these post-biotics interact with the host[87]. In the future, the development of dietary methods and adjunct treatments to prevent the development of HCC may be supported by the creation of combinations of symbiotics or post-biotics with improved anticancer potential.

**ACKNOWLEDGEMENTS**

The authors thank Dr. Elisangela Miceli, PhD for English revisions.

**REFERENCES**

1 **Kulik L**, El-Serag HB. Epidemiology and Management of Hepatocellular Carcinoma. *Gastroenterology* 2019; **156**: 477-491.e1 [PMID: 30367835 DOI: 10.1053/j.gastro.2018.08.065]

2 **Ferlay J**, Shin HR, Bray F, Forman D, Mathers C, Parkin DM. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. *Int J Cancer* 2010; **127**: 2893-2917 [PMID: 21351269 DOI: 10.1002/ijc.25516]

3 **Nordenstedt H**, White DL, El-Serag HB. The changing pattern of epidemiology in hepatocellular carcinoma. *Dig Liver Dis* 2010; **42** Suppl 3: S206-S214 [PMID: 20547305 DOI: 10.1016/S1590-8658(10)60507-5]

4 **Cholankeril G**, Patel R, Khurana S, Satapathy SK. Hepatocellular carcinoma in non-alcoholic steatohepatitis: Current knowledge and implications for management. *World J Hepatol* 2017; **9**: 533-543 [PMID: 28469809 DOI: 10.4254/wjh.v9.i11.533]

5 **Schulze K**, Imbeaud S, Letouzé E, Alexandrov LB, Calderaro J, Rebouissou S, Couchy G, Meiller C, Shinde J, Soysouvanh F, Calatayud AL, Pinyol R, Pelletier L, Balabaud C, Laurent A, Blanc JF, Mazzaferro V, Calvo F, Villanueva A, Nault JC, Bioulac-Sage P, Stratton MR, Llovet JM, Zucman-Rossi J. Exome sequencing of hepatocellular carcinomas identifies new mutational signatures and potential therapeutic targets. *Nat Genet* 2015; **47**: 505-511 [PMID: 25822088 DOI: 10.1038/ng.3252]

6 **Anstee QM**, Reeves HL, Kotsiliti E, Govaere O, Heikenwalder M. From NASH to HCC: current concepts and future challenges. *Nat Rev Gastroenterol Hepatol* 2019; **16**: 411-428 [PMID: 31028350 DOI: 10.1038/s41575-019-0145-7]

7 **Friedman SL**, Neuschwander-Tetri BA, Rinella M, Sanyal AJ. Mechanisms of NAFLD development and therapeutic strategies. *Nat Med* 2018; **24**: 908-922 [PMID: 29967350 DOI: 10.1038/s41591-018-0104-9]

8 **Russo E**, Taddei A, Ringressi MN, Ricci F, Amedei A. The interplay between the microbiome and the adaptive immune response in cancer development. *Therap Adv Gastroenterol* 2016; **9**: 594-605 [PMID: 27366226 DOI: 10.1177/1756283X16635082]

9 **Russo E**,Amedei A. The Role of the Microbiota in the Genesis of Gastrointestinal Cancers. *Front Anti-Infective Drug Discov* 2018; **7**: 1-44 [DOI: 10.2174/9781681085623118070003]

10 **Wan MLY**, El-Nezami H. Targeting gut microbiota in hepatocellular carcinoma: probiotics as a novel therapy. *Hepatobiliary Surg Nutr* 2018; **7**: 11-20 [PMID: 29531939 DOI: 10.21037/hbsn.2017.12.07]

11 **Pradere JP**, Troeger JS, Dapito DH, Mencin AA, Schwabe RF. Toll-like receptor 4 and hepatic fibrogenesis. *Semin Liver Dis* 2010; **30**: 232-244 [PMID: 20665376 DOI: 10.1055/s-0030-1255353]

12 **Tao X**, Wang N, Qin W. Gut Microbiota and Hepatocellular Carcinoma. *Gastrointest Tumors* 2015; **2**: 33-40 [PMID: 26673641 DOI: 10.1159/000380895]

13 **Lu H**, Ren Z, Li A, Zhang H, Jiang J, Xu S, Luo Q, Zhou K, Sun X, Zheng S, Li L. Deep sequencing reveals microbiota dysbiosis of tongue coat in patients with liver carcinoma. *Sci Rep* 2016; **6**: 33142 [PMID: 27605161 DOI: 10.1038/srep33142]

14 **Dapito DH**, Mencin A, Gwak GY, Pradere JP, Jang MK, Mederacke I, Caviglia JM, Khiabanian H, Adeyemi A, Bataller R, Lefkowitch JH, Bower M, Friedman R, Sartor RB, Rabadan R, Schwabe RF. Promotion of hepatocellular carcinoma by the intestinal microbiota and TLR4. *Cancer Cell* 2012; **21**: 504-516 [PMID: 22516259 DOI: 10.1016/j.ccr.2012.02.007]

15 **Konturek PC**, Harsch IA, Konturek K, Schink M, Konturek T, Neurath MF, Zopf Y. Gut⁻Liver Axis: How Do Gut Bacteria Influence the Liver? *Med Sci (Basel)* 2018; **6** [PMID: 30227645 DOI: 10.3390/medsci6030079]

16 **Vajro P**, Paolella G, Fasano A. Microbiota and gut-liver axis: their influences on obesity and obesity-related liver disease. *J Pediatr Gastroenterol Nutr* 2013; **56**: 461-468 [PMID: 23287807 DOI: 10.1097/MPG.0b013e318284abb5]

17 **Golonka RM**, Vijay-Kumar M. Atypical immunometabolism and metabolic reprogramming in liver cancer: Deciphering the role of gut microbiome. *Adv Cancer Res* 2021; **149**: 171-255 [PMID: 33579424 DOI: 10.1016/bs.acr.2020.10.004]

18 **Yiu JH**, Dorweiler B, Woo CW. Interaction between gut microbiota and toll-like receptor: from immunity to metabolism. *J Mol Med (Berl)* 2017; **95**: 13-20 [PMID: 27639584 DOI: 10.1007/s00109-016-1474-4]

19 **Tripathi A**, Debelius J, Brenner DA, Karin M, Loomba R, Schnabl B, Knight R. The gut-liver axis and the intersection with the microbiome. *Nat Rev Gastroenterol Hepatol* 2018; **15**: 397-411 [PMID: 29748586 DOI: 10.1038/s41575-018-0011-z]

20 **Ren Z**, Li A, Jiang J, Zhou L, Yu Z, Lu H, Xie H, Chen X, Shao L, Zhang R, Xu S, Zhang H, Cui G, Chen X, Sun R, Wen H, Lerut JP, Kan Q, Li L, Zheng S. Gut microbiome analysis as a tool towards targeted non-invasive biomarkers for early hepatocellular carcinoma. *Gut* 2019; **68**: 1014-1023 [PMID: 30045880 DOI: 10.1136/gutjnl-2017-315084]

21 **Grąt M**, Wronka KM, Krasnodębski M, Masior Ł, Lewandowski Z, Kosińska I, Grąt K, Stypułkowski J, Rejowski S, Wasilewicz M, Gałęcka M, Szachta P, Krawczyk M. Profile of Gut Microbiota Associated With the Presence of Hepatocellular Cancer in Patients With Liver Cirrhosis. *Transplant Proc* 2016; **48**: 1687-1691 [PMID: 27496472 DOI: 10.1016/j.transproceed.2016.01.077]

22 **Zhang HL**, Yu LX, Yang W, Tang L, Lin Y, Wu H, Zhai B, Tan YX, Shan L, Liu Q, Chen HY, Dai RY, Qiu BJ, He YQ, Wang C, Zheng LY, Li YQ, Wu FQ, Li Z, Yan HX, Wang HY. Profound impact of gut homeostasis on chemically-induced pro-tumorigenic inflammation and hepatocarcinogenesis in rats. *J Hepatol* 2012; **57**: 803-812 [PMID: 22727732 DOI: 10.1016/j.jhep.2012.06.011]

23 **Ponziani FR**, Bhoori S, Castelli C, Putignani L, Rivoltini L, Del Chierico F, Sanguinetti M, Morelli D, Paroni Sterbini F, Petito V, Reddel S, Calvani R, Camisaschi C, Picca A, Tuccitto A, Gasbarrini A, Pompili M, Mazzaferro V. Hepatocellular Carcinoma Is Associated With Gut Microbiota Profile and Inflammation in Nonalcoholic Fatty Liver Disease. *Hepatology* 2019; **69**: 107-120 [PMID: 29665135 DOI: 10.1002/hep.30036]

24 **Liu Q**, Li F, Zhuang Y, Xu J, Wang J, Mao X, Zhang Y, Liu X. Alteration in gut microbiota associated with hepatitis B and non-hepatitis virus related hepatocellular carcinoma. *Gut Pathog* 2019; **11**: 1 [PMID: 30675188 DOI: 10.1186/s13099-018-0281-6]

25 **Sheng L**, Jena PK, Hu Y, Liu HX, Nagar N, Kalanetra KM, French SW, French SW, Mills DA, Wan YY. Hepatic inflammation caused by dysregulated bile acid synthesis is reversible by butyrate supplementation. *J Pathol* 2017; **243**: 431-441 [PMID: 28892150 DOI: 10.1002/path.4983]

26 **Krüttgen A**, Horz HP, Weber-Heynemann J, Vucur M, Trautwein C, Haase G, Luedde T, Roderburg C. Study on the association of Helicobacter species with viral hepatitis-induced hepatocellular carcinoma. *Gut Microbes* 2012; **3**: 228-233 [PMID: 22572832 DOI: 10.4161/gmic.19922]

27 **Routy B**, Le Chatelier E, Derosa L, Duong CPM, Alou MT, Daillère R, Fluckiger A, Messaoudene M, Rauber C, Roberti MP, Fidelle M, Flament C, Poirier-Colame V, Opolon P, Klein C, Iribarren K, Mondragón L, Jacquelot N, Qu B, Ferrere G, Clémenson C, Mezquita L, Masip JR, Naltet C, Brosseau S, Kaderbhai C, Richard C, Rizvi H, Levenez F, Galleron N, Quinquis B, Pons N, Ryffel B, Minard-Colin V, Gonin P, Soria JC, Deutsch E, Loriot Y, Ghiringhelli F, Zalcman G, Goldwasser F, Escudier B, Hellmann MD, Eggermont A, Raoult D, Albiges L, Kroemer G, Zitvogel L. Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. *Science* 2018; **359**: 91-97 [PMID: 29097494 DOI: 10.1126/science.aan3706]

28 **Russo E**, Nannini G, Dinu M, Pagliai G, Sofi F, Amedei A. Exploring the food-gut axis in immunotherapy response of cancer patients. *World J Gastroenterol* 2020; **26**: 4919-4932 [PMID: 32952339 DOI: 10.3748/wjg.v26.i33.4919]

29 **Sajjad A**, Mottershead M, Syn WK, Jones R, Smith S, Nwokolo CU. Ciprofloxacin suppresses bacterial overgrowth, increases fasting insulin but does not correct low acylated ghrelin concentration in non-alcoholic steatohepatitis. *Aliment Pharmacol Ther* 2005; **22**: 291-299 [PMID: 16097995 DOI: 10.1111/j.1365-2036.2005.02562.x]

30 **Li J**, Sung CY, Lee N, Ni Y, Pihlajamäki J, Panagiotou G, El-Nezami H. Probiotics modulated gut microbiota suppresses hepatocellular carcinoma growth in mice. *Proc Natl Acad Sci U S A* 2016; **113**: E1306-E1315 [PMID: 26884164 DOI: 10.1073/pnas.1518189113]

31 **Million M**, Angelakis E, Paul M, Armougom F, Leibovici L, Raoult D. Comparative meta-analysis of the effect of Lactobacillus species on weight gain in humans and animals. *Microb Pathog* 2012; **53**: 100-108 [PMID: 22634320 DOI: 10.1016/j.micpath.2012.05.007]

32 **Gøbel RJ**, Larsen N, Jakobsen M, Mølgaard C, Michaelsen KF. Probiotics to adolescents with obesity: effects on inflammation and metabolic syndrome. *J Pediatr Gastroenterol Nutr* 2012; **55**: 673-678 [PMID: 22695039 DOI: 10.1097/MPG.0b013e318263066c]

33 **Larsen N**, Vogensen FK, Gøbel RJ, Michaelsen KF, Forssten SD, Lahtinen SJ, Jakobsen M. Effect of Lactobacillus salivarius Ls-33 on fecal microbiota in obese adolescents. *Clin Nutr* 2013; **32**: 935-940 [PMID: 23510724 DOI: 10.1016/j.clnu.2013.02.007]

34 **Yoshimoto S**, Loo TM, Atarashi K, Kanda H, Sato S, Oyadomari S, Iwakura Y, Oshima K, Morita H, Hattori M, Honda K, Ishikawa Y, Hara E, Ohtani N. Obesity-induced gut microbial metabolite promotes liver cancer through senescence secretome. *Nature* 2013; **499**: 97-101 [PMID: 23803760 DOI: 10.1038/nature12347]

35 **Huang L**, Duan C, Zhao Y, Gao L, Niu C, Xu J, Li S. Reduction of Aflatoxin B1 Toxicity by Lactobacillus plantarum C88: A Potential Probiotic Strain Isolated from Chinese Traditional Fermented Food "Tofu". *PLoS One* 2017; **12**: e0170109 [PMID: 28129335 DOI: 10.1371/journal.pone.0170109]

36 **Nduti NN**, McMillan AM, Seney S, Sumarah MW, Njeru PN, Mwaniki MW, Reid G. Investigating probiotic yoghurt to reduce an aflatoxin B1 biomarker among school children in Eastern Kenya: Preliminary study. *Int Dairy J* 2016; **63**: 124–129 [DOI: 10.1016/j.idairyj.2016.07.014]

37 **Heydari Z**, Rahaie M, Alizadeh AM. Different anti-inflammatory effects of Lactobacillus acidophilus and Bifidobactrum bifidioum in hepatocellular carcinoma cancer mouse through impact on microRNAs and their target genes. *J Nutr Intermed Metab* 2019; **16**: 100096 [DOI: 10.1016/j.jnim.2019.100096]

38 **Miro-Blanch J**, Yanes O. Epigenetic Regulation at the Interplay Between Gut Microbiota and Host Metabolism. *Front Genet* 2019; **10**: 638 [PMID: 31338107 DOI: 10.3389/fgene.2019.00638]

39 **Kumar M**, Verma V, Nagpal R, Kumar A, Gautam SK, Behare PV, Grover CR, Aggarwal PK. Effect of probiotic fermented milk and chlorophyllin on gene expressions and genotoxicity during AFB₁-induced hepatocellular carcinoma. *Gene* 2011; **490**: 54-59 [PMID: 21963996 DOI: 10.1016/j.gene.2011.09.003]

40 **Elshaer AM**, El-Kharashi OA, Hamam GG, Nabih ES, Magdy YM, Abd El Samad AA. Involvement of TLR4/ CXCL9/ PREX-2 pathway in the development of hepatocellular carcinoma (HCC) and the promising role of early administration of lactobacillus plantarum in Wistar rats. *Tissue Cell* 2019; **60**: 38-47 [PMID: 31582017 DOI: 10.1016/j.tice.2019.07.010]

41 **Lee DK**, Kang JY, Shin HS, Park IH, Ha NJ. Antiviral activity of Bifidobacterium adolescentis SPM0212 against Hepatitis B virus. *Arch Pharm Res* 2013; **36**: 1525-1532 [PMID: 23657805 DOI: 10.1007/s12272-013-0141-3]

42 **El-Adawi HN,** I.; Fattouh, F.; El-Deeb, N. . Investigation of the antiviral bioactivity of Lactobacillus bulgaricus 761N extracellular extract against hepatitis C virus (HCV). *Int J Pharmacol* 2015; **11**: 19–26 [DOI: 10.3923/ijp.2015.19.26]

43 **Oo KM**, Lwin AA, Kyaw YY, Tun WM, Fukada K, Goshima A, Shimada T, Okada S. Safety and long-term effect of the probiotic FK-23 in patients with hepatitis C virus infection. *Biosci Microbiota Food Health* 2016; **35**: 123-128 [PMID: 27508113 DOI: 10.12938/bmfh.2015-024]

44 **Nabavi S**, Rafraf M, Somi MH, Homayouni-Rad A, Asghari-Jafarabadi M. Effects of probiotic yogurt consumption on metabolic factors in individuals with nonalcoholic fatty liver disease. *J Dairy Sci* 2014; **97**: 7386-7393 [PMID: 25306266 DOI: 10.3168/jds.2014-8500]

45 **Ahn SB**, Jun DW, Kang BK, Lim JH, Lim S, Chung MJ. Randomized, Double-blind, Placebo-controlled Study of a Multispecies Probiotic Mixture in Nonalcoholic Fatty Liver Disease. *Sci Rep* 2019; **9**: 5688 [PMID: 30952918 DOI: 10.1038/s41598-019-42059-3]

46 **König J**, Wells J, Cani PD, García-Ródenas CL, MacDonald T, Mercenier A, Whyte J, Troost F, Brummer RJ. Human Intestinal Barrier Function in Health and Disease. *Clin Transl Gastroenterol* 2016; **7**: e196 [PMID: 27763627 DOI: 10.1038/ctg.2016.54]

47 **Song CH**, Kim N, Sohn SH, Lee SM, Nam RH, Na HY, Lee DH, Surh YJ. Effects of 17β-Estradiol on Colonic Permeability and Inflammation in an Azoxymethane/Dextran Sulfate Sodium-Induced Colitis Mouse Model. *Gut Liver* 2018; **12**: 682-693 [PMID: 30400733 DOI: 10.5009/gnl18221]

48 **Bouchagier KA**, Assimakopoulos SF, Karavias DD, Maroulis I, Tzelepi V, Kalofonos H, Karavias DD, Kardamakis D, Scopa CD, Tsamandas AC. Expression of claudins-1, -4, -5, -7 and occludin in hepatocellular carcinoma and their relation with classic clinicopathological features and patients' survival. *In Vivo* 2014; **28**: 315-326 [PMID: 24815833]

49 **Holczbauer Á**, Gyöngyösi B, Lotz G, Törzsök P, Kaposi-Novák P, Szijártó A, Tátrai P, Kupcsulik P, Schaff Z, Kiss A. Increased expression of claudin-1 and claudin-7 in liver cirrhosis and hepatocellular carcinoma. *Pathol Oncol Res* 2014; **20**: 493-502 [PMID: 24696415 DOI: 10.1007/s12253-013-9683-4]

50 **Huang GW**, Ding X, Chen SL, Zeng L. Expression of claudin 10 protein in hepatocellular carcinoma: impact on survival. *J Cancer Res Clin Oncol* 2011; **137**: 1213-1218 [PMID: 21647678 DOI: 10.1007/s00432-011-0987-z]

51 **Cheung ST**, Leung KL, Ip YC, Chen X, Fong DY, Ng IO, Fan ST, So S. Claudin-10 expression level is associated with recurrence of primary hepatocellular carcinoma. *Clin Cancer Res* 2005; **11**: 551-556 [PMID: 15701840]

52 **Nagai T**, Arao T, Nishio K, Matsumoto K, Hagiwara S, Sakurai T, Minami Y, Ida H, Ueshima K, Nishida N, Sakai K, Saijo N, Kudo K, Kaneda H, Tamura D, Aomatsu K, Kimura H, Fujita Y, Haji S, Kudo M. Impact of Tight Junction Protein ZO-1 and TWIST Expression on Postoperative Survival of Patients with Hepatocellular Carcinoma. *Dig Dis* 2016; **34**: 702-707 [PMID: 27750241 DOI: 10.1159/000448860]

53 **Mennigen R**, Nolte K, Rijcken E, Utech M, Loeffler B, Senninger N, Bruewer M. Probiotic mixture VSL#3 protects the epithelial barrier by maintaining tight junction protein expression and preventing apoptosis in a murine model of colitis. *Am J Physiol Gastrointest Liver Physiol* 2009; **296**: G1140-G1149 [PMID: 19221015 DOI: 10.1152/ajpgi.90534.2008]

54 **Briskey D**, Heritage M, Jaskowski LA, Peake J, Gobe G, Subramaniam VN, Crawford D, Campbell C, Vitetta L. Probiotics modify tight-junction proteins in an animal model of nonalcoholic fatty liver disease. *Therap Adv Gastroenterol* 2016; **9**: 463-472 [PMID: 27366215 DOI: 10.1177/1756283X16645055]

55 **Ahmadi S**, Wang S, Nagpal R, Wang B, Jain S, Razazan A, Mishra SP, Zhu X, Wang Z, Kavanagh K, Yadav H. A human-origin probiotic cocktail ameliorates aging-related leaky gut and inflammation via modulating the microbiota/taurine/tight junction axis. *JCI Insight* 2020; **5** [PMID: 32302292 DOI: 10.1172/jci.insight.132055]

56 **Gibson GR**, Hutkins R, Sanders ME, Prescott SL, Reimer RA, Salminen SJ, Scott K, Stanton C, Swanson KS, Cani PD, Verbeke K, Reid G. Expert consensus document: The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. *Nat Rev Gastroenterol Hepatol* 2017; **14**: 491-502 [PMID: 28611480 DOI: 10.1038/nrgastro.2017.75]

57 **Roberfroid M**, Gibson GR, Hoyles L, McCartney AL, Rastall R, Rowland I, Wolvers D, Watzl B, Szajewska H, Stahl B, Guarner F, Respondek F, Whelan K, Coxam V, Davicco MJ, Léotoing L, Wittrant Y, Delzenne NM, Cani PD, Neyrinck AM, Meheust A. Prebiotic effects: metabolic and health benefits. *Br J Nutr* 2010; **104** Suppl 2: S1-63 [PMID: 20920376 DOI: 10.1017/S0007114510003363]

58 **Bultman SJ**. The microbiome and its potential as a cancer preventive intervention. *Semin Oncol* 2016; **43**: 97-106 [PMID: 26970128 DOI: 10.1053/j.seminoncol.2015.09.001]

59 **Makki K**, Deehan EC, Walter J, Bäckhed F. The Impact of Dietary Fiber on Gut Microbiota in Host Health and Disease. *Cell Host Microbe* 2018; **23**: 705-715 [PMID: 29902436 DOI: 10.1016/j.chom.2018.05.012]

60 **Davis CD**, Milner JA. Gastrointestinal microflora, food components and colon cancer prevention. *J Nutr Biochem* 2009; **20**: 743-752 [PMID: 19716282 DOI: 10.1016/j.jnutbio.2009.06.001]

61 **Sousa V**, Santos E, Sgarbieri V. The Importance of Prebiotics in Functional Foods and Clinical Practice. *Food Sci Nutr* 2011; **2** [DOI: 10.4236/fns.2011.22019]

62 **Albouery M**, Bretin A, Buteau B, Grégoire S, Martine L, Gambert S, Bron AM, Acar N, Chassaing B, Bringer MA. Soluble Fiber Inulin Consumption Limits Alterations of the Gut Microbiota and Hepatic Fatty Acid Metabolism Caused by High-Fat Diet. *Nutrients* 2021; **13** [PMID: 33806985 DOI: 10.3390/nu13031037]

63 **Loo TM**, Kamachi F, Watanabe Y, Yoshimoto S, Kanda H, Arai Y, Nakajima-Takagi Y, Iwama A, Koga T, Sugimoto Y, Ozawa T, Nakamura M, Kumagai M, Watashi K, Taketo MM, Aoki T, Narumiya S, Oshima M, Arita M, Hara E, Ohtani N. Gut Microbiota Promotes Obesity-Associated Liver Cancer through PGE2-Mediated Suppression of Antitumor Immunity. *Cancer Discov* 2017; **7**: 522-538 [PMID: 28202625 DOI: 10.1158/2159-8290.CD-16-0932]

64 **Ma C**, Han M, Heinrich B, Fu Q, Zhang Q, Sandhu M, Agdashian D, Terabe M, Berzofsky JA, Fako V, Ritz T, Longerich T, Theriot CM, McCulloch JA, Roy S, Yuan W, Thovarai V, Sen SK, Ruchirawat M, Korangy F, Wang XW, Trinchieri G, Greten TF. Gut microbiome-mediated bile acid metabolism regulates liver cancer via NKT cells. *Science* 2018; **360** [PMID: 29798856 DOI: 10.1126/science.aan5931]

65 **Ridlon JM**, Kang DJ, Hylemon PB, Bajaj JS. Bile acids and the gut microbiome. *Curr Opin Gastroenterol* 2014; **30**: 332-338 [PMID: 24625896 DOI: 10.1097/MOG.0000000000000057]

66 **Zhang W**, Xiang YB, Li HL, Yang G, Cai H, Ji BT, Gao YT, Zheng W, Shu XO. Vegetable-based dietary pattern and liver cancer risk: results from the Shanghai women's and men's health studies. *Cancer Sci* 2013; **104**: 1353-1361 [PMID: 23841909 DOI: 10.1111/cas.12231]

67 **Yang W**, Ma Y, Liu Y, Smith-Warner SA, Simon TG, Chong DQ, Qi Q, Meyerhardt JA, Giovannucci EL, Chan AT, Zhang X. Association of Intake of Whole Grains and Dietary Fiber With Risk of Hepatocellular Carcinoma in US Adults. *JAMA Oncol* 2019; **5**: 879-886 [PMID: 30789662 DOI: 10.1001/jamaoncol.2018.7159]

68 **Makarova-Rusher OV**, Altekruse SF, McNeel TS, Ulahannan S, Duffy AG, Graubard BI, Greten TF, McGlynn KA. Population attributable fractions of risk factors for hepatocellular carcinoma in the United States. *Cancer* 2016; **122**: 1757-1765 [PMID: 26998818 DOI: 10.1002/cncr.29971]

69 **Costabile A**, Klinder A, Fava F, Napolitano A, Fogliano V, Leonard C, Gibson GR, Tuohy KM. Whole-grain wheat breakfast cereal has a prebiotic effect on the human gut microbiota: a double-blind, placebo-controlled, crossover study. *Br J Nutr* 2008; **99**: 110-120 [PMID: 17761020 DOI: 10.1017/S0007114507793923]

70 **Yang Y**, Zhang D, Feng N, Chen G, Liu J, Chen G, Zhu Y. Increased intake of vegetables, but not fruit, reduces risk for hepatocellular carcinoma: a meta-analysis. *Gastroenterology* 2014; **147**: 1031-1042 [PMID: 25127680 DOI: 10.1053/j.gastro.2014.08.005]

71 **George ES**, Sood S, Broughton A, Cogan G, Hickey M, Chan WS, Sudan S, Nicoll AJ. The Association between Diet and Hepatocellular Carcinoma: A Systematic Review. *Nutrients* 2021; **13** [PMID: 33430001 DOI: 10.3390/nu13010172]

72 **Ferguson LR**, Philpott M. Cancer prevention by dietary bioactive components that target the immune response. *Curr Cancer Drug Targets* 2007; **7**: 459-464 [PMID: 17691905 DOI: 10.2174/156800907781386605]

73 **Darvesh AS**, Bishayee A. Chemopreventive and therapeutic potential of tea polyphenols in hepatocellular cancer. *Nutr Cancer* 2013; **65**: 329-344 [PMID: 23530632 DOI: 10.1080/01635581.2013.767367]

74 **Larrosa M**, González-Sarrías A, García-Conesa MT, Tomás-Barberán FA, Espín JC. Urolithins, ellagic acid-derived metabolites produced by human colonic microflora, exhibit estrogenic and antiestrogenic activities. *J Agric Food Chem* 2006; **54**: 1611-1620 [PMID: 16506809 DOI: 10.1021/jf0527403]

75 **Acharya S**, Adamová D, Adhya SP, Adler A, Adolfsson J, Aggarwal MM, Aglieri Rinella G, Agnello M, Agrawal N, Ahammed Z, Ahmad S, Ahn SU, Aiola S, Akindinov A, Al-Turany M, Alam SN, Albuquerque DSD, Aleksandrov D, Alessandro B, Alfanda HM, Alfaro Molina R, Ali B, Ali Y, Alici A, Alkin A, Alme J, Alt T, Altenkamper L, Altsybeev I, Anaam MN, Andrei C, Andreou D, Andrews HA, Andronic A, Angeletti M, Anguelov V, Anson C, Antičić T, Antinori F, Antonioli P, Anwar R, Apadula N, Aphecetche L, Appelshäuser H, Arcelli S, Arnaldi R, Arratia M, Arsene IC, Arslandok M, Augustinus A, Averbeck R, Aziz S, Azmi MD, Badalà A, Baek YW, Bagnasco S, Bailhache R, Bala R, Baldisseri A, Ball M, Baral RC, Barbera R, Barioglio L, Barnaföldi GG, Barnby LS, Barret V, Bartalini P, Barth K, Bartsch E, Bastid N, Basu S, Batigne G, Batyunya B, Batzing PC, Bauri D, Bazo Alba JL, Bearden IG, Bedda C, Behera NK, Belikov I, Bellini F, Bellwied R, Beltran LGE, Belyaev V, Bencedi G, Beole S, Bercuci A, Berdnikov Y, Berenyi D, Bertens RA, Berzano D, Betev L, Bhasin A, Bhat IR, Bhatt H, Bhattacharjee B, Bianchi A, Bianchi L, Bianchi N, Bielčík J, Bielčíková J, Bilandzic A, Biro G, Biswas R, Biswas S, Blair JT, Blau D, Blume C, Boca G, Bock F, Bogdanov A, Boldizsár L, Bolozdynya A, Bombara M, Bonomi G, Bonora M, Borel H, Borissov A, Borri M, Botta E, Bourjau C, Bratrud L, Braun-Munzinger P, Bregant M, Broker TA, Broz M, Brucken EJ, Bruna E, Bruno GE, Buckland MD, Budnikov D, Buesching H, Bufalino S, Buhler P, Buncic P, Busch O, Buthelezi Z, Butt JB, Buxton JT, Caffarri D, Caines H, Caliva A, Calvo Villar E, Camacho RS, Camerini P, Capon AA, Carnesecchi F, Castillo Castellanos J, Castro AJ, Casula EAR, Catalano F, Ceballos Sanchez C, Chakraborty P, Chandra S, Chang B, Chang W, Chapeland S, Chartier M, Chattopadhyay S, Chattopadhyay S, Chauvin A, Cheshkov C, Cheynis B, Chibante Barroso V, Chinellato DD, Cho S, Chochula P, Chowdhury T, Christakoglou P, Christensen CH, Christiansen P, Chujo T, Cicalo C, Cifarelli L, Cindolo F, Cleymans J, Colamaria F, Colella D, Collu A, Colocci M, Concas M, Conesa Balbastre G, Conesa Del Valle Z, Contin G, Contreras JG, Cormier TM, Corrales Morales Y, Cortese P, Cosentino MR, Costa F, Costanza S, Crkovská J, Crochet P, Cuautle E, Cunqueiro L, Dabrowski D, Dahms T, Dainese A, Damas FPA, Dani S, Danisch MC, Danu A, Das D, Das I, Das S, Dash A, Dash S, Dashi A, De S, De Caro A, de Cataldo G, de Conti C, de Cuveland J, De Falco A, De Gruttola D, De Marco N, De Pasquale S, De Souza RD, Deb S, Degenhardt HF, Deisting A, Deja KR, Deloff A, Delsanto S, Dhankher P, Di Bari D, Di Mauro A, Diaz RA, Dietel T, Dillenseger P, Ding Y, Divià R, Djuvsland Ø, Dmitrieva U, Dobrin A, Domenicis Gimenez D, Dönigus B, Dordic O, Dubey AK, Dubla A, Dudi S, Duggal AK, Dukhishyam M, Dupieux P, Ehlers RJ, Elia D, Engel H, Epple E, Erazmus B, Erhardt F, Erokhin A, Ersdal MR, Espagnon B, Eulisse G, Eum J, Evans D, Evdokimov S, Fabbietti L, Faggin M, Faivre J, Fantoni A, Fasel M, Fecchio P, Feldkamp L, Feliciello A, Feofilov G, Fernández Téllez A, Ferrero A, Ferretti A, Festanti A, Feuillard VJG, Figiel J, Filchagin S, Finogeev D, Fionda FM, Fiorenza G, Flor F, Foertsch S, Foka P, Fokin S, Fragiacomo E, Francisco A, Frankenfeld U, Fronze GG, Fuchs U, Furget C, Furs A, Fusco Girard M, Gaardhøje JJ, Gagliardi M, Gago AM, Gal A, Galvan CD, Ganoti P, Garabatos C, Garcia-Solis E, Garg K, Gargiulo C, Garner K, Gasik P, Gauger EF, Gay Ducati MB, Germain M, Ghosh J, Ghosh P, Ghosh SK, Gianotti P, Giubellino P, Giubilato P, Glässel P, Goméz Coral DM, Gomez Ramirez A, Gonzalez V, González-Zamora P, Gorbunov S, Görlich L, Gotovac S, Grabski V, Graczykowski LK, Graham KL, Greiner L, Grelli A, Grigoras C, Grigoriev V, Grigoryan A, Grigoryan S, Groettvik OS, Gronefeld JM, Grosa F, Grosse-Oetringhaus JF, Grosso R, Guernane R, Guerzoni B, Guittiere M, Gulbrandsen K, Gunji T, Gupta A, Gupta R, Guzman IB, Haake R, Habib MK, Hadjidakis C, Hamagaki H, Hamar G, Hamid M, Hamon JC, Hannigan R, Haque MR, Harlenderova A, Harris JW, Harton A, Hassan H, Hatzifotiadou D, Hauer P, Hayashi S, Heckel ST, Hellbär E, Helstrup H, Herghelegiu A, Hernandez EG, Herrera Corral G, Herrmann F, Hetland KF, Hilden TE, Hillemanns H, Hills C, Hippolyte B, Hohlweger B, Horak D, Hornung S, Hosokawa R, Hristov P, Huang C, Hughes C, Huhn P, Humanic TJ, Hushnud H, Husova LA, Hussain N, Hussain SA, Hussain T, Hutter D, Hwang DS, Iddon JP, Ilkaev R, Inaba M, Ippolitov M, Islam MS, Ivanov M, Ivanov V, Izucheev V, Jacak B, Jacazio N, Jacobs PM, Jadhav MB, Jadlovska S, Jadlovsky J, Jaelani S, Jahnke C, Jakubowska MJ, Janik MA, Jercic M, Jevons O, Jimenez Bustamante RT, Jin M, Jonas F, Jones PG, Jusko A, Kalinak P, Kalweit A, Kang JH, Kaplin V, Kar S, Karasu Uysal A, Karavichev O, Karavicheva T, Karczmarczyk P, Karpechev E, Kebschull U, Keidel R, Keil M, Ketzer B, Khabanova Z, Khan AM, Khan S, Khan SA, Khanzadeev A, Kharlov Y, Khatun A, Khuntia A, Kileng B, Kim B, Kim B, Kim D, Kim DJ, Kim EJ, Kim H, Kim JS, Kim J, Kim J, Kim J, Kim M, Kim S, Kim T, Kim T, Kindra K, Kirsch S, Kisel I, Kiselev S, Kisiel A, Klay JL, Klein C, Klein J, Klein S, Klein-Bösing C, Klewin S, Kluge A, Knichel ML, Knospe AG, Kobdaj C, Kofarago M, Köhler MK, Kollegger T, Kondratyev A, Kondratyeva N, Kondratyuk E, Konopka PJ, Konyushikhin M, Koska L, Kovalenko O, Kovalenko V, Kowalski M, Králik I, Kravčáková A, Kreis L, Krivda M, Krizek F, Krizkova Gajdosova K, Krüger M, Kryshen E, Krzewicki M, Kubera AM, Kučera V, Kuhn C, Kuijer PG, Kumar L, Kumar S, Kundu S, Kurashvili P, Kurepin A, Kurepin AB, Kushpil S, Kvapil J, Kweon MJ, Kwon Y, La Pointe SL, La Rocca P, Lai YS, Langoy R, Lapidus K, Lardeux A, Larionov P, Laudi E, Lavicka R, Lazareva T, Lea R, Leardini L, Lee S, Lehas F, Lehner S, Lehrbach J, Lemmon RC, León Monzón I, Lettrich M, Lévai P, Li X, Li XL, Lien J, Lietava R, Lim B, Lindal S, Lindenstruth V, Lindsay SW, Lippmann C, Lisa MA, Litichevskyi V, Liu A, Liu S, Ljunggren HM, Llope WJ, Lodato DF, Loginov V, Loizides C, Loncar P, Lopez X, López Torres E, Luettig P, Luhder JR, Lunardon M, Luparello G, Lupi M, Maevskaya A, Mager M, Mahmood SM, Mahmoud T, Maire A, Majka RD, Malaev M, Malik QW, Malinina L, Mal'Kevich D, Malzacher P, Mamonov A, Manko V, Manso F, Manzari V, Mao Y, Marchisone M, Mareš J, Margagliotti GV, Margotti A, Margutti J, Marín A, Markert C, Marquard M, Martin NA, Martinengo P, Martinez JL, Martínez MI, Martínez García G, Martinez Pedreira M, Masciocchi S, Masera M, Masoni A, Massacrier L, Masson E, Mastroserio A, Mathis AM, Matuoka PFT, Matyja A, Mayer C, Mazzilli M, Mazzoni MA, Mechler AF, Meddi F, Melikyan Y, Menchaca-Rocha A, Meninno E, Meres M, Mhlanga S, Miake Y, Micheletti L, Mieskolainen MM, Mihaylov DL, Mikhaylov K, Mischke A, Mishra AN, Miśkowiec D, Mitu CM, Mohammadi N, Mohanty AP, Mohanty B, Khan MM, Mondal MM, Mordasini C, Moreira De Godoy DA, Moreno LAP, Moretto S, Morreale A, Morsch A, Mrnjavac T, Muccifora V, Mudnic E, Mühlheim D, Muhuri S, Mulligan JD, Munhoz MG, Münning K, Munzer RH, Murakami H, Murray S, Musa L, Musinsky J, Myers CJ, Myrcha JW, Naik B, Nair R, Nandi BK, Nania R, Nappi E, Naru MU, Nassirpour AF, Natal da Luz H, Nattrass C, Nayak K, Nayak R, Nayak TK, Nazarenko S, Negrao De Oliveira RA, Nellen L, Nesbo SV, Neskovic G, Ng F, Nielsen BS, Nikolaev S, Nikulin S, Nikulin V, Noferini F, Nomokonov P, Nooren G, Noris JCC, Norman J, Nowakowski P, Nyanin A, Nystrand J, Ogino M, Ohlson A, Oleniacz J, Oliveira Da Silva AC, Oliver MH, Onderwaater J, Oppedisano C, Orava R, Ortiz Velasquez A, Oskarsson A, Otwinowski J, Oyama K, Pachmayer Y, Pacik V, Pagano D, Paić G, Palni P, Pan J, Pandey AK, Panebianco S, Papikyan V, Pareek P, Park J, Parkkila JE, Parmar S, Passfeld A, Pathak SP, Patra RN, Paul B, Pei H, Peitzmann T, Peng X, Pereira LG, Pereira Da Costa H, Peresunko D, Perez GM, Perez Lezama E, Peskov V, Pestov Y, Petráček V, Petrovici M, Pezzi RP, Piano S, Pikna M, Pillot P, Pimentel LODL, Pinazza O, Pinsky L, Pisano S, Piyarathna DB, Płoskoń M, Planinic M, Pliquett F, Pluta J, Pochybova S, Poghosyan MG, Polichtchouk B, Poljak N, Poonsawat W, Pop A, Poppenborg H, Porteboeuf-Houssais S, Pozdniakov V, Prasad SK, Preghenella R, Prino F, Pruneau CA, Pshenichnov I, Puccio M, Punin V, Puranapanda K, Putschke J, Quishpe RE, Ragoni S, Raha S, Rajput S, Rak J, Rakotozafindrabe A, Ramello L, Rami F, Raniwala R, Raniwala S, Räsänen SS, Rascanu BT, Rath R, Ratza V, Ravasenga I, Read KF, Redlich K, Rehman A, Reichelt P, Reidt F, Ren X, Renfordt R, Reshetin A, Revol JP, Reygers K, Riabov V, Richert T, Richter M, Riedler P, Riegler W, Riggi F, Ristea C, Rode SP, Rodríguez Cahuantzi M, Røed K, Rogalev R, Rogochaya E, Rohr D, Röhrich D, Rokita PS, Ronchetti F, Rosas ED, Roslon K, Rosnet P, Rossi A, Rotondi A, Roukoutakis F, Roy A, Roy P, Rueda OV, Rui R, Rumyantsev B, Rustamov A, Ryabinkin E, Ryabov Y, Rybicki A, Rytkonen H, Saarinen S, Sadhu S, Sadovsky S, Šafařík K, Saha SK, Sahoo B, Sahoo P, Sahoo R, Sahoo S, Sahu PK, Saini J, Sakai S, Sambyal S, Samsonov V, Sandoval A, Sarkar A, Sarkar D, Sarkar N, Sarma P, Sarti VM, Sas MHP, Scapparone E, Schaefer B, Schambach J, Scheid HS, Schiaua C, Schicker R, Schmah A, Schmidt C, Schmidt HR, Schmidt MO, Schmidt M, Schmidt NV, Schmier AR, Schukraft J, Schutz Y, Schwarz K, Schweda K, Scioli G, Scomparin E, Šefčík M, Seger JE, Sekiguchi Y, Sekihata D, Selyuzhenkov I, Senyukov S, Serradilla E, Sett P, Sevcenco A, Shabanov A, Shabetai A, Shahoyan R, Shaikh W, Shangaraev A, Sharma A, Sharma A, Sharma M, Sharma N, Sheikh AI, Shigaki K, Shimomura M, Shirinkin S, Shou Q, Sibiriak Y, Siddhanta S, Siemiarczuk T, Silvermyr D, Simatovic G, Simonetti G, Singh R, Singh R, Singh VK, Singhal V, Sinha T, Sitar B, Sitta M, Skaali TB, Slupecki M, Smirnov N, Snellings RJM, Snellman TW, Sochan J, Soncco C, Song J, Songmoolnak A, Soramel F, Sorensen S, Sputowska I, Stachel J, Stan I, Stankus P, Steffanic PJ, Stenlund E, Stocco D, Storetvedt MM, Strmen P, Suaide AAP, Sugitate T, Suire C, Suleymanov M, Suljic M, Sultanov R, Šumbera M, Sumowidagdo S, Suzuki K, Swain S, Szabo A, Szarka I, Tabassam U, Taillepied G, Takahashi J, Tambave GJ, Tang S, Tarhini M, Tarzila MG, Tauro A, Tejeda Muñoz G, Telesca A, Terrevoli C, Thakur D, Thakur S, Thomas D, Thoresen F, Tieulent R, Tikhonov A, Timmins AR, Toia A, Topilskaya N, Toppi M, Torales-Acosta F, Torres SR, Tripathy S, Tripathy T, Trogolo S, Trombetta G, Tropp L, Trubnikov V, Trzaska WH, Trzcinski TP, Trzeciak BA, Tsuji T, Tumkin A, Turrisi R, Tveter TS, Ullaland K, Umaka EN, Uras A, Usai GL, Utrobicic A, Vala M, Valle N, van der Kolk N, van Doremalen LVR, van Leeuwen M, Vande Vyvre P, Varga D, Vargas A, Vargyas M, Varma R, Vasileiou M, Vasiliev A, Vázquez Doce O, Vechernin V, Veen AM, Vercellin E, Vergara Limón S, Vermunt L, Vernet R, Vértesi R, Vickovic L, Viinikainen J, Vilakazi Z, Villalobos Baillie O, Villatoro Tello A, Vino G, Vinogradov A, Virgili T, Vislavicius V, Vodopyanov A, Volkel B, Völkl MA, Voloshin K, Voloshin SA, Volpe G, von Haller B, Vorobyev I, Voscek D, Vrláková J, Wagner B, Wang M, Watanabe Y, Weber M, Weber SG, Wegrzynek A, Weiser DF, Wenzel SC, Wessels JP, Westerhoff U, Whitehead AM, Widmann E, Wiechula J, Wikne J, Wilk G, Wilkinson J, Willems GA, Willsher E, Windelband B, Witt WE, Wu Y, Xu R, Yalcin S, Yamakawa K, Yang S, Yano S, Yin Z, Yokoyama H, Yoo IK, Yoon JH, Yuan S, Yuncu A, Yurchenko V, Zaccolo V, Zaman A, Zampolli C, Zanoli HJC, Zardoshti N, Zarochentsev A, Závada P, Zaviyalov N, Zbroszczyk H, Zhalov M, Zhang X, Zhang Y, Zhang Z, Zhao C, Zherebchevskii V, Zhigareva N, Zhou D, Zhou Y, Zhou Z, Zhu H, Zhu J, Zhu Y, Zichichi A, Zimmermann MB, Zinovjev G, Zurlo N; A Large Ion Collider Experiment Collaboration. Investigations of Anisotropic Flow Using Multiparticle Azimuthal Correlations in pp, p-Pb, Xe-Xe, and Pb-Pb Collisions at the LHC. *Phys Rev Lett* 2019; **123**: 142301 [PMID: 31702212 DOI: 10.1103/PhysRevLett.123.142301]

76 **Lamuel-Raventos RM**, Onge MS. Prebiotic nut compounds and human microbiota. *Crit Rev Food Sci Nutr* 2017; **57**: 3154-3163 [PMID: 27224877 DOI: 10.1080/10408398.2015.1096763]

77 **Bøhn SK**, Blomhoff R, Paur I. Coffee and cancer risk, epidemiological evidence, and molecular mechanisms. *Mol Nutr Food Res* 2014; **58**: 915-930 [PMID: 24668519 DOI: 10.1002/mnfr.201300526]

78 **Fotschki B**, Juśkiewicz J, Jurgoński A, Sójka M. Fructo-Oligosaccharides and Pectins Enhance Beneficial Effects of Raspberry Polyphenols in Rats with Nonalcoholic Fatty Liver. *Nutrients* 2021; **13** [PMID: 33802455 DOI: 10.3390/nu13030833]

79 **Wongkrasant P**, Pongkorpsakol P, Ariyadamrongkwan J, Meesomboon R, Satitsri S, Pichyangkura R, Barrett KE, Muanprasat C. A prebiotic fructo-oligosaccharide promotes tight junction assembly in intestinal epithelial cells via an AMPK-dependent pathway. *Biomed Pharmacother* 2020; **129**: 110415 [PMID: 32603892 DOI: 10.1016/j.biopha.2020.110415]

80 **Wang J**, Tian S, Yu H, Wang J, Zhu W. Response of Colonic Mucosa-Associated Microbiota Composition, Mucosal Immune Homeostasis, and Barrier Function to Early Life Galactooligosaccharides Intervention in Suckling Piglets. *J Agric Food Chem* 2019; **67**: 578-588 [PMID: 30562014 DOI: 10.1021/acs.jafc.8b05679]

81 **Peng L**, Li ZR, Green RS, Holzman IR, Lin J. Butyrate enhances the intestinal barrier by facilitating tight junction assembly via activation of AMP-activated protein kinase in Caco-2 cell monolayers. *J Nutr* 2009; **139**: 1619-1625 [PMID: 19625695 DOI: 10.3945/jn.109.104638]

82 **Julie Uerlingsac MS,** Els Willemsb, Sofie Tangheb, Geert Bruggemanb, Jérôme Bindellea, Nadia Everaert. Differential effects of inulin or its fermentation metabolites on gut barrier and immune function of porcine intestinal epithelial cells. *J Funct Foods* 2020; **67**: 103855 [DOI: 10.1016/j.jff.2020.103855]

83 **Wang G**, Sun W, Pei X, Jin Y, Wang H, Tao W, Xiao Z, Liu L, Wang M. Galactooligosaccharide pretreatment alleviates damage of the intestinal barrier and inflammatory responses in LPS-challenged mice. *Food Funct* 2021; **12**: 1569-1579 [PMID: 33459741 DOI: 10.1039/d0fo03020a]

84 **Mandair DS**, Rossi RE, Pericleous M, Whyand T, Caplin M. The impact of diet and nutrition in the prevention and progression of hepatocellular carcinoma. *Expert Rev Gastroenterol Hepatol* 2014; **8**: 369-382 [PMID: 24597926 DOI: 10.1586/17474124.2014.894879]

85 **Turati F**, Trichopoulos D, Polesel J, Bravi F, Rossi M, Talamini R, Franceschi S, Montella M, Trichopoulou A, La Vecchia C, Lagiou P. Mediterranean diet and hepatocellular carcinoma. *J Hepatol* 2014; **60**: 606-611 [PMID: 24240052 DOI: 10.1016/j.jhep.2013.10.034]

86 **Thilakarathna WPDW**, Langille MG, Rupasinghe HPV. Polyphenol-based prebiotics and synbiotics: Potential for cancer chemoprevention. *Curr Opin Food Sci* 2018; **20**: 51-57 [DOI: 10.1016/j.cofs.2018.02.011]

87 **Russo E**, Giudici F, Fiorindi C, Ficari F, Scaringi S, Amedei A. Immunomodulating Activity and Therapeutic Effects of Short Chain Fatty Acids and Tryptophan Post-biotics in Inflammatory Bowel Disease. *Front Immunol* 2019; **10**: 2754 [PMID: 31824517 DOI: 10.3389/fimmu.2019.02754]

**Footnotes**

**Conflict-of-interest statement:** Authors declare no any conflict of interest.

**Open-Access:** This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: https://creativecommons.org/Licenses/by-nc/4.0/

**Provenance and peer review:** Invited article; Externally peer reviewed.

**Peer-review model:** Single blind

**Peer-review started:** May 27, 2021

**First decision:** July 6, 2021

**Article in press:**

**Specialty type:** Gastroenterology and hepatology

**Country/Territory of origin:** Italy

**Peer-review report’s scientific quality classification**

Grade A (Excellent): A

Grade B (Very good): 0

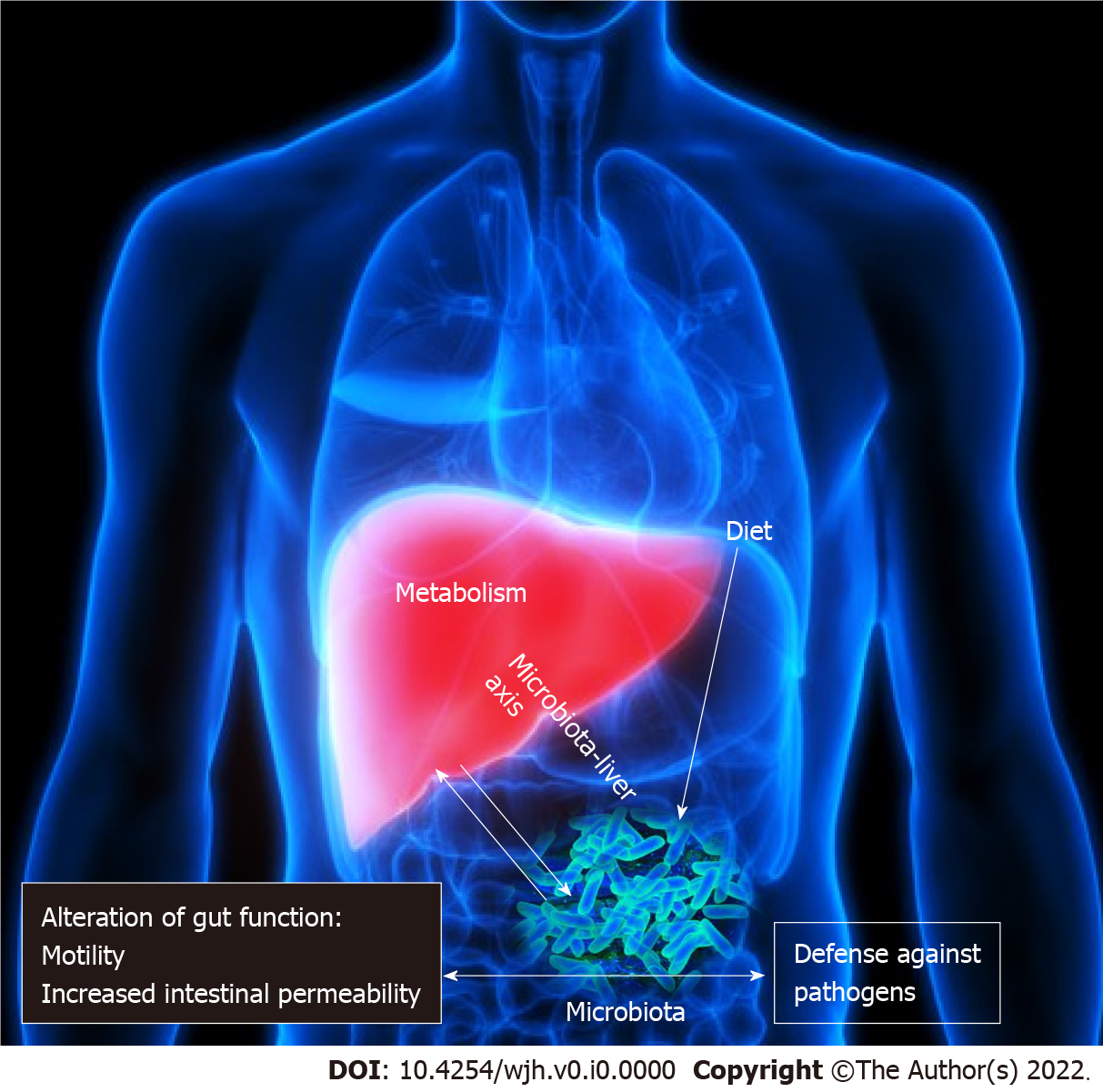
Grade C (Good): C

Grade D (Fair): 0

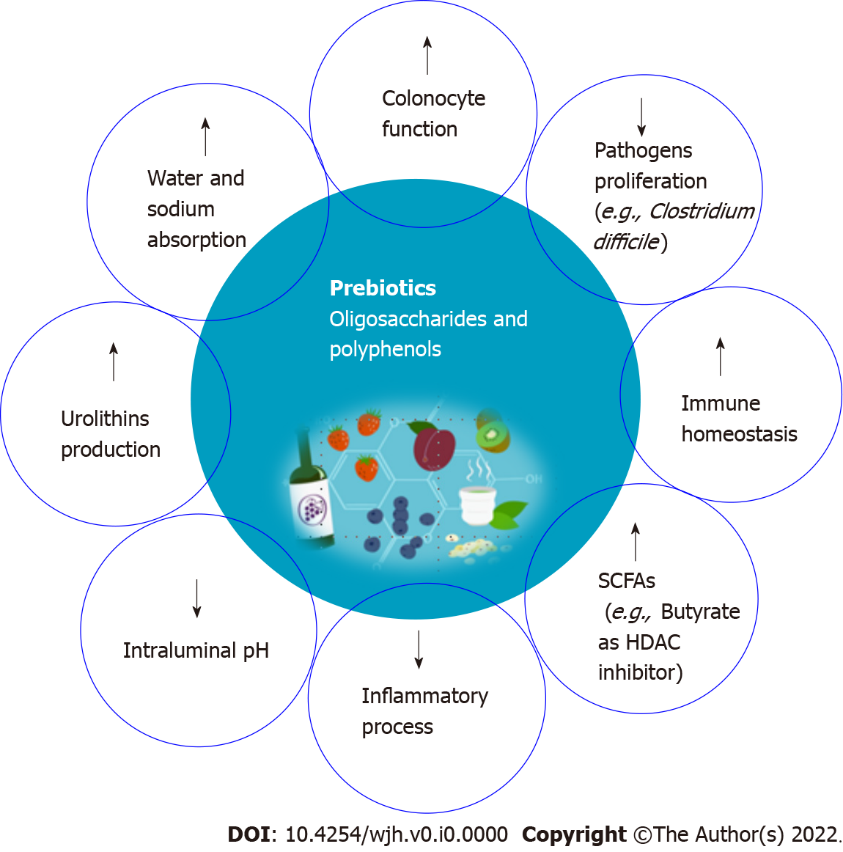
Grade E (Poor): 0

**P-Reviewer:** Vairappan B, Zhang L **S-Editor:** Wang JJ **L-Editor:** Webster JR **P-Editor:**

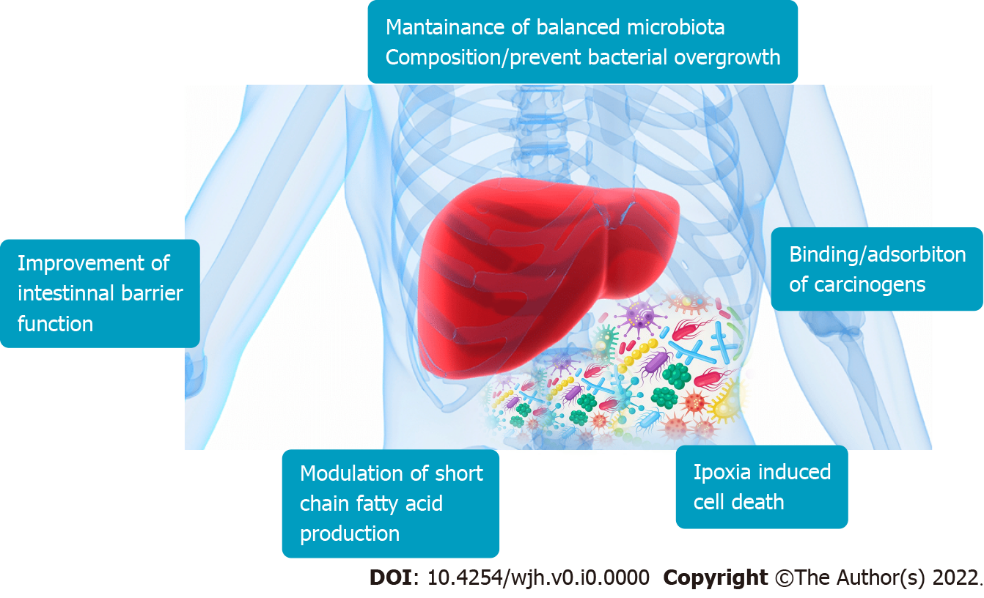
**Figure Legends**



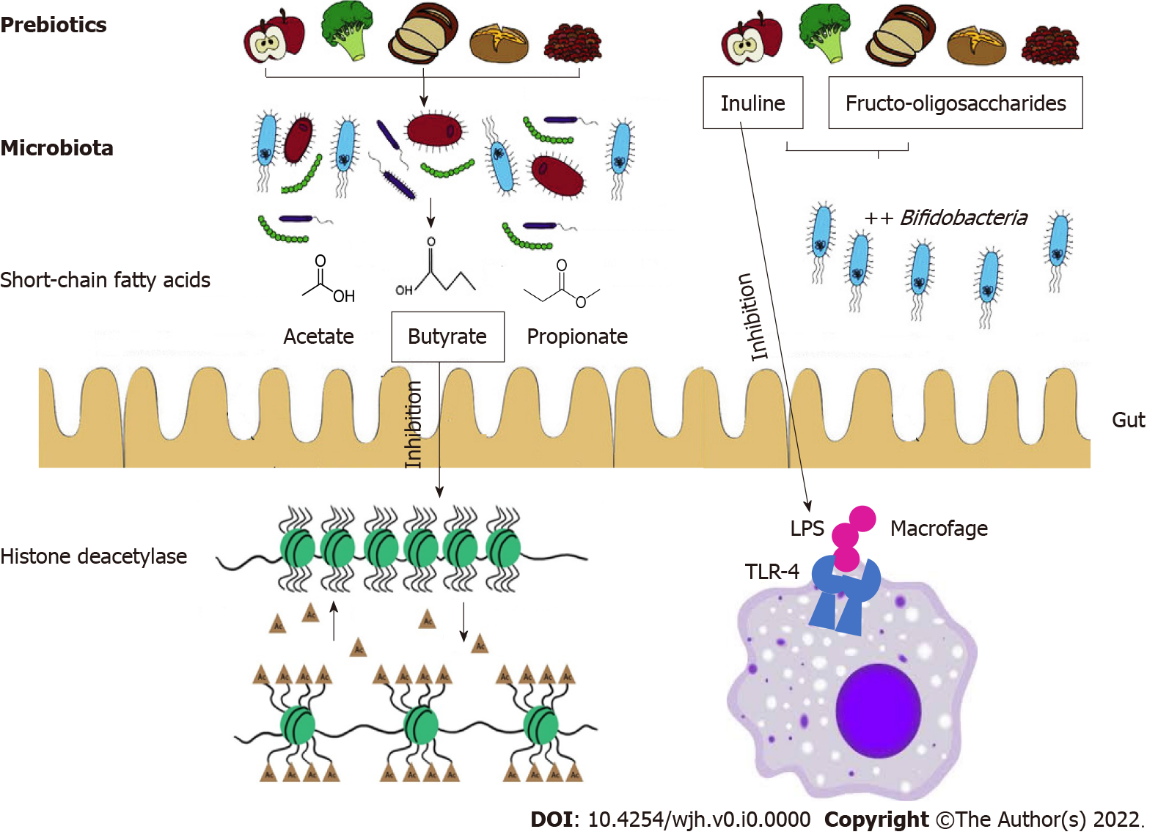
**Figure 1 The gut liver axis and microbiota.**



**Figure 2 Proposed mechanism of action of probiotics against hepatocellular carcinoma progression.** HDAC: Histone deacetylase; SCFAs: Short-chain fatty acids.



**Figure 3 Role of prebiotics on immunomodulation in hepatocellular carcinoma.** Prebiotics contribute to short-chain fatty acids such as acetate, propionate, and butyrate production, which positively influence the structure and function of the microbial community. Butyrate functions include inhibition of histone deacetylase, and decreased tumorigenesis. In addition, inulin improves gut barrier integrity, thereby reducing pro-inflammatory lipopolysaccharides-toll-like receptors 4 signaling in macrophages during alcoholic liver disease. Fructo-oligosaccharides and inulin stimulates the growth of beneficial *Bifidobacteria* that are implicated in cancer prevention.



**Figure 4 Effect of prebiotics in cancer prevention.** LPS: Lipopolysaccharides; TLR: Toll-like receptors.