

Contribution of miRNAs to ion-channel remodelling in atrial fibrillation

José Manuel Vilches, Diego Franco, Amelia E Aránega

José Manuel Vilches, Diego Franco, Amelia E Aránega, Cardiac and Skeletal Myogenesis Group, Department of Experimental Biology, University of Jaén, 23071 Jaén, Spain
Author contributions: All the authors equally contributed to this work.

Supported by Grants BFU2012-38111 (Ministerio de Economía y Competitividad, Gobierno de España), Nos. CTS-1614, P08-CTS-03878, BIO-302 (Junta de Andalucía) and AFM2012-16074 (AFM).

Conflict-of-interest: The authors declare that they have no competing interests.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Amelia E Aránega, PhD, Cardiac and Skeletal Myogenesis Group, Department of Experimental Biology, University of Jaén, CU Las Lagunillas B3-362, 23071 Jaén, Spain. aaaranega@ujaen.es

Telephone: +34-953-212763

Fax: +34-953-211875

Received: October 7, 2014

Peer-review started: October 8, 2014

First decision: November 1, 2014

Revised: November 12, 2014

Accepted: December 10, 2014

Article in press: December 10, 2014

Published online: February 23, 2015

Abstract

Atrial fibrillation (AF) is the most commonly encountered clinical arrhythmia associated with pronounced mortality and morbidity, which are related to palpitations, fainting, congestive heart failure, and stroke. Prolonged episodes of AF promote AF persistence mainly due to electrical remodelling that alters ion-channel expression and/or function. MicroRNAs (miRNAs), a new class of non-

coding mRNAs of around 22 nucleotides in length, have recently emerged as one of the key players in the gene-expression regulatory networks. The potential roles of miRNAs in controlling AF have recently been investigated. Several recent studies have provided promising results for a better understanding of the molecular mechanisms of AF. In this review, we summarize the mechanism of miRNAs as regulators of ion-channel gene expression and their role in causing AF through electrical remodelling.

Key words: Atrial fibrillation; MicroRNA; Ion channels; Electrical remodelling

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Atrial fibrillation (AF) is the most common sustained arrhythmia in clinical practice associated with pronounced mortality and morbidity. MicroRNAs (miRNAs), which are approximately 22 nucleotide long RNA regulators of gene expression, have become a major focus of research in molecular biology miRNAs have emerged as key post-transcriptional regulators of gene expression therefore, it is important to comment on the roles of these small non-protein-coding mRNAs in the cardiovascular system and their involvement in AF. In this review, we summarize the mechanism of miRNAs as regulators of ion-channels genes and their role in electrical remodelling caused by AF.

Vilches JM, Franco D, Aránega AE. Contribution of miRNAs to ion-channel remodelling in atrial fibrillation. *World J Hypertens* 2015; 5(1): 6-13 Available from: URL: <http://www.wjgnet.com/2220-3168/full/v5/i1/6.htm> DOI: <http://dx.doi.org/10.5494/wjh.v5.i1.6>

INTRODUCTION

Atrial fibrillation (AF) is the most common sustained

arrhythmia in clinical practice, with high prevalence in the geriatric population^[1]. AF is a condition in which heart-rhythm control is displaced from the normal sinus node pacemaker by rapid activity in different areas of the atria, resulting in rapid and irregular atrial activity. This pathology is often underestimated because it is usually asymptomatic and rarely life-threatening. However, it can induce palpitations, fainting, congestive heart failure, and stroke due to electrical disturbance, as well as thromboembolism, all of which are associated with high morbidity and mortality^[2]. Therefore, a better understanding of the mechanisms of and molecular basis for AF may uncover safer, more effective therapeutic targets and new treatment methods^[3].

AF progresses as three different clinical forms, which have been previously well described. Initially, AF occurs in a paroxysmal form characterized by episodes of the arrhythmia that terminate spontaneously. Persistent AF is defined by episodes that are sustained longer than 7 d and require termination by pharmacological or direct-current electric cardioversion. In permanent AF the normal heart rhythm cannot be converted to sinus rhythm. It is believed that AF development from paroxysmal to persistent and permanent forms occurs through the influence of atrial remodelling caused by the arrhythmia itself and/or the progression of underlying heart disease^[3]. Therefore, the molecular and electrical remodelling of ion channels determining the potential duration has been proposed as a major mechanism in chronic atrial fibrillation. Prolonged episodes of AF alter atrial properties (“atrial remodelling”), promoting AF persistence. Electrical remodelling alters ion-channel expression and/or function, increasing the likelihood of ectopic firing or re-entry, thereby promoting AF initiation and/or recurrence. Ion-channel remodelling shortens the atrial effective refractory period (ERP), favouring re-entrant arrhythmias, mainly due to the shortening of atrial action potential duration (APD)^[4-6]. ERP reduction promotes the conduction block and wave break underlying fibrillatory conduction. APD shortening may lead to AF-related atrial contractile dysfunction^[7-11]. The balance between inward currents that depolarize and outward currents that repolarize cardiac myocytes determines APD, and thus ERP. Thus, deregulation of ion expression as a molecular mechanism for AF has been considered quite interesting by many researchers. In addition to electrical remodelling it has been shown that structural alterations and fibrosis are implicated in AF generation and perpetuation^[12,13].

Recently microRNA (miRNA) research has evolved rapidly, therefore it is important to comment on the roles of these non-coding short RNA molecules in the cardiovascular system and their involvement in AF. Primarily, miRNAs suppress protein synthesis by inhibiting the translation of protein from mRNA or by promoting the degradation of mRNA, thereby silencing gene expression on the post-transcriptional level. Notably, many recent reports have demonstrated that miRNAs regulate several properties of cardiac physiology and excitability,

including automaticity, Ca²⁺ handling, conduction, and repolarization^[14]. Several miRNAs have been shown to have a role in the triggering or prolonging AF, while other miRNAs target several genes considered to have the potential to regulate AF. Available results show the potential role of miRNAs involved in AF.

In this review, we summarize the mechanism of miRNAs as regulators of ion-channels genes and their role in electrical remodelling caused by AF.

ROLE OF MIRNAS IN CARDIAC DISEASES, REMODELLING, AND HOMEOSTASIS

MiRNAs are endogenous, conserved, single-stranded, small (c. 22 nucleotides in length), noncoding RNAs that repress gene expression at the post-transcriptional level by targeting mRNAs^[15,16]. A single miRNA can regulate multiple targets; therefore, modulating the expression of one miRNA often results in the modulation of an entire gene network. Genes coding for miRNAs, such as the protein-coding genes, are located in chromosomes as an integral part of the genome; however, these genes may appear more genomically diverse and dynamic. Based on the genomic organization of miRNA genes, miRNAs can be grouped into two classes: (1) intergenic miRNAs, when the miRNA-coding genes are located in genomic regions distinct from known transcription units and are found between protein-coding genes; and/or (2) intragenic miRNAs, if the miRNA-coding genes are located within annotated genes either of protein-coding genes or of long noncoding RNAs as their host genes and they are transcribed by using the host regulatory elements. Curiously, some miRNAs are transcribed from multiple copies of their genes whereas many other miRNAs appear in clusters on a single polycistronic transcript.

Two converging pathways have been demonstrated for the biogenesis of miRNAs: the canonical pathway and the noncanonical pathway^[17]. The canonical pathway is Drosha-dependent and is usually used to generate intergenic miRNAs and intronic miRNAs. The first step of this pathway, transcription of miRNAs genes yields transcripts, termed primary miRNA transcripts (pri-miRNAs) with a characteristic hairpin morphology, comprising a loop and an imperfectly paired stem incorporating the mature miRNA sequence on one of the strands, operated mainly by RNA polymerase II. Subsequently, these pri-miRNAs are processed by the Class 2 Rnase enzyme Drosha together with Pasha protein to become precursor miRNAs (pre-miRNAs), which are then transported to the cytoplasm through the Ran-GTP-dependent nuclear pore exportin-5, where they are processed by Dicer (another RNase III enzyme) to generate the final approximately 22 nucleotide product of mature miRNAs. The mature miRNAs are then loaded into the miRNA-associated multi-protein RNA-induced silencing complex (RISC) that includes the Argonaute

Proteins (mature miRNAs). A perfect base-pairing of the miRNA (within miRISC complex) motif(s), mainly in the 3'-untranslated region termed the "seed sequence", a critical site for miRNA actions^[18,19], is required for effective repression of translation of the protein. Also, some miRNAs are involved in gene silencing through mRNA degradation. Alternatively, Drosha-independent and either Dicer-dependent or Dicer-independent noncanonical pathways are used to generate Mirtrons, small nucleolar miRNAs and short hairpin miRNAs transfer miRNAs. It is important to highlight that some properties of the miRNAs, such as their ability to influence whole cellular processes instead of single genes, the easy manipulation of miRNA activity *in vivo* using miRNA inhibitors, and the high intracellular stability of miRNAs, make them a promising therapeutic target^[20].

To date, over a thousand miRNAs have been identified and they have been reported to participate in many fundamental biological processes, including cell proliferation, growth, differentiation, apoptosis, and tissue remodelling. The essential role of miRNA in early heart development was first revealed by a Dicer knockout. The early lethality of Dicer mutants, along with the complete depletion of mature miRNAs, indicated the crucial role of miRNAs in early heart development^[21]. Additionally, Dicer activity is also required for normal functioning of the mature heart, as adult mice lacking Dicer in the myocardium suffer from high incidence of sudden death, cardiac hypertrophy, and reactivation of the foetal cardiac-gene programme^[22]. These data demonstrate that miRNAs play an important role during normal heart development, cardiac remodelling, and homeostasis. Dysregulated miRNAs have been shown to be related to the pathology of many cardiovascular diseases, including arrhythmia, hypertrophy, and heart failure, gaining a critical position as potential therapeutical targets for cardiac diseases. For instance, multiple miRNAs, such as *miR-1*, *miR-133*, *miR-208*, *miR-195*, *miR-21*, and *miR-18b*, have been identified to participate in cardiac hypertrophy and can independently determine this pathological process. Importantly, *miR-1*, *miR-133*, *miR-21*, *miR-29*, *miR-199a*, and *miR-320* have also been reported to be involved in myocardial infarction^[23]. Among them, *miR-1* and *miR-133* are considered muscle-specific miRNAs and they represent the most abundant miRNAs expressed in the heart^[14].

Interestingly, recent studies have highlighted circulating micRNAs as biomarkers for the AF^[24]. Moreover, several miRNAs such as *miR-24*, *miR-29b* and *miR133* have been shown to be involved in myocardial fibrosis^[25,26] and open new perspectives to analyse the role of miRNAs in the structural alterations and fibrosis involved in AF generation and perpetuation.

Finally, a few miRNAs have been reported to regulate cardiac excitability through targeting genes encoding ion channels^[27-30]. Here we focus on miRNAs which target different ion channels that play key roles in the electrical remodelling of AF.

ION-CHANNEL DYSFUNCTION IN ATRIAL FIBRILLATION

It has been previously shown that the factors responsible for onset of AF include triggers that induce arrhythmia. Triggers include not only sympathetic or parasympathetic stimulation, bradycardia, atrial premature beats or tachycardia, accessory AV pathways, and acute atrial stretch, but also ectopic foci occurring in "sleeves" of atrial tissue within the pulmonary veins or vena caval junctions as well^[31].

After the onset of AF, prolonged episodes alter atrial properties ("atrial remodelling") promoting AF persistence. AF-associated remodelling events within the atria include electrical remodelling and, secondarily, structural remodelling and autonomic-nervous-system changes. AF-related electrical remodelling can result from altered expression and/or function of cardiac ion channels, leading to the development of functional re-entry substrates and finally contributing to persistent AF. Therefore, electrical remodelling shortens the atrial ERP and APD, and reduces its physiological rate of adaptation as a result of ion-channel remodelling and abnormal Ca^{2+} handling (Figure 1).

The inward sodium current (I_{Na}), which is encoded by the voltage-gated cardiac sodium channel *SCN5A*, initiates the cardiac action potential and this I_{Na} is involved in repolarization and refractoriness. It bears highlight that rapid cardiac rates can significantly reduce (I_{Na}) density and conduction velocity, triggering AF (Figure 1)^[32]. In addition, mutations in *SCN5A* are related to diseases caused by abnormalities in cardiac conduction, including AF. In this context, Darbar *et al*^[11] tested the hypothesis that vulnerability to AF is associated with variation in *SCN5A*, the gene encoding the cardiac sodium channel. They found novel as well as rare variants in nearly 6% of the population, including alleles that segregate with AF in other family members, supporting the hypothesis that *SCN5A* is an important AF-susceptibility gene.

It has been well documented that increased K^+ currents or decreased Ca^{2+} currents shorten APD, promoting re-entrant AF. Because Ca^{2+} enters atrial cells during AP depolarization, the high atrial rate in AF increase Ca^{2+} loading, which leads to activation of auto-protective mechanisms to reduce chronic Ca^{2+} overload. Therefore, in patients with chronic AF, mRNA transcription of the auxiliary L-type Ca^{2+} -channel subunits are significantly diminished reducing the inward Ca^{2+} currents, maintaining the AP plateau, shortening the AP duration, and finally contributing to re-entry^[33]. Moreover, alterations in Ca^{2+} handling promote diastolic Ca^{2+} release by increasing open probability of the sarcoplasmic reticulum Ca^{2+} release channels (RyR2), this in turn promoting delayed after-depolarizations (DADs) and ectopic activity (Figure 1)^[5,6,34].

Additionally, AF-related shortening of APD can also result from an increase in K^+ channel gene products and/or activity. The inward rectified K^+ current (I_{K1}) and I_{K-ACh} are also altered in AF and both AF-related I_{K1} up-

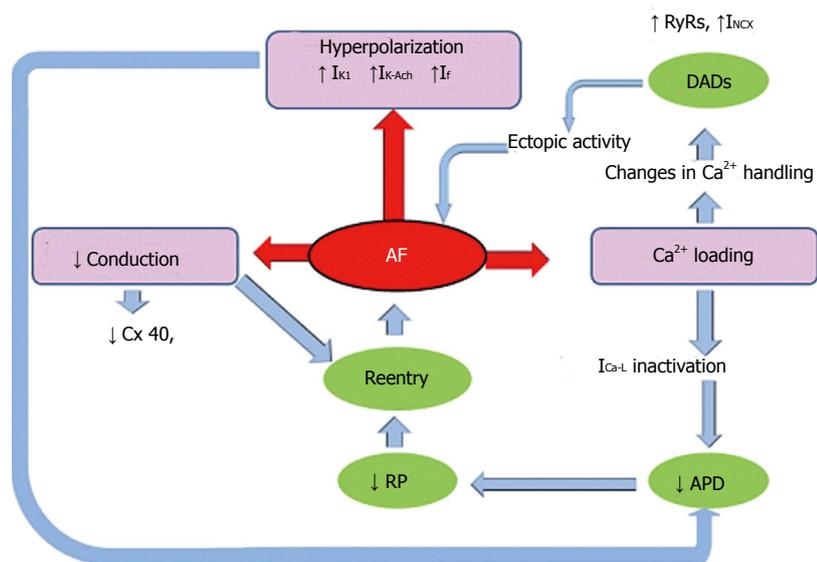


Figure 1 Mechanisms of atrial fibrillation. Cytotoxic overloading of Ca^{2+} occurs by a rapid atrial rate. As a result, this acts as a mechanism of cell protection via the inactivation of L-type Ca^{2+} current (I_{Ca-L}), which in turn reduces the action potential duration (APD) and this consequently shortens the refractory period (RP). This overloading of Ca^{2+} also contributes to an abnormal function of ryanodine receptor channels (RyRs) and intensifies the inward Na^+Ca^{2+} -exchange current (I_{Ncx}), leading to delayed afterpolarizations (DADs) and ectopic activity. In addition, AF produces hyperpolarization due to increased I_{Ca-L} , which increases the inward rectifier current (I_{K1}) and acetylcholine-regulated K^+ current (I_{KAch}). This also reduces APD and promotes conduction by decreasing of connexin-40 (Cx40).

regulation and enhancement of the constitutive form of I_{KAch} promote the persistence of AF by reducing APD (Figure 1)^[35]. *KCNN3*, which encodes the small conductance calcium-activated potassium channel 3 (SK3), is a new locus strongly associated with AF^[36]. The role of SK channels remains unclear, but it is believed that they contribute to the repolarization of cardiac action^[37,38]. A recent study showed SK3 to be strongly down-regulated in permanent AF, which contributed to AF persistence and susceptibility by abbreviating APD^[39].

Similarly, it has been well established that chronic AF in humans reduces the transient outward potassium currents (I_{to}) by transcriptional down-regulation of the Kv4.3 potassium channel^[7,8]. The ultrarapid delayed rectifier K^+ channel (I_{kur}) also plays a major part since it has been shown that the amplitude of I_{kur} is significantly reduced and the current density is lower in patients with AF^[10]. More recently it has been demonstrated in human that chronic AF modifications in transcriptional and posttranscriptional mechanisms of HCN channels occur and are associated with a slight yet significant gain-of-function of the hyperpolarization activated “funny” current (I_f), which may contribute to enhanced atrial ectopy^[40].

The slow delayed rectifier potassium channel (I_{Ks}), made up of a pore-forming subunit encoded by *KCNQ1*, has also been implicated in AF. Thus, several mutations in *KCNQ1* have been described in AF patients^[41,42], some of which are associated with I_{Ks} block and AP^[43]. The potassium channel voltage-gated subfamily member 2-(*KCNH2*) is an ion-channel gene that encodes an inwardly rectifying potassium channel (I_{Kr}), and a genetic variation in the *KCNH2* gene reportedly predisposes Chinese Han individuals to the risk of acquired AF^[44]. In addition, lower expression levels of the ATP-sensitive potassium channel (*Kir6.2*); the acetylcholine-dependent potassium channel (*Kir3.4*) and the ultra-rapid delayed rectifier potassium channel (*Kv1.5*) in patients with AF have also been described^[10,45]. However, the underlying mechanisms leading to these changes remain unclear.

Finally, although relatively little is known about gap-junction remodelling in the atria, changes in ion channels involved in cardiac conduction have also been reported in relation to AF. Atrial gap junctions are composed of two different subunit proteins, *i.e.*, Connexin40 (Cx40) and Connexin43 (Cx43). In the goat, Cx40 expression decreases and the protein becomes more heterogeneously distributed after long periods of fast pacing, contributing to the onset and self-perpetuation of re-entry pathways and AF^[46]. Moreover, pacing can induce atrial tachyarrhythmias or fibrillation in Cx40-null mice^[47]. These results are consistent with the extensive clinical evidence pointing to disturbances in connexin-40 as a basis for genetic AF predisposition. Importantly, a very recent report has shown a clear reduction in Cx40 but not Cx43 expression (both mRNA and protein) in atrial appendage tissues from patients with lone AF with no Cx40 coding-region mutations identified^[47].

In summary, despite that many studies have pointed out the relevance of the AF-related electrical remodelling, our understanding of the underlying molecular mechanisms leading to and perpetuating ion-channel remodelling during AF is very limited. Better knowledge and deeper insights into the molecular mechanisms underlying AF may help to identify new and selective drug targets for improved AF treatment.

MICRORNAS AND ELECTRICAL REMODELLING IN AF

As mentioned above, electrical remodelling is a maladaptive process that can trigger and prolong AF. Recently, multiple research groups have uncovered an important role of several miRNAs in regulating cardiac excitability and arrhythmogenesis by targeting specific ion channels, which might have an important impact on AF onset and/or persistence^[48,49].

Microarray analyses in a canine model of AF showed that *miR-223*, *miR-328*, and *miR-664* were significantly up-

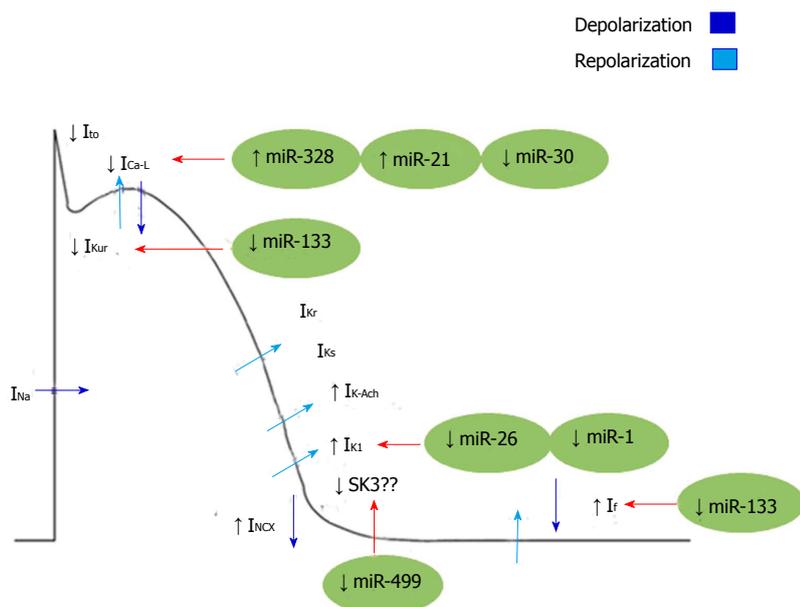


Figure 2 Mechanisms of electrical remodelling in atrial fibrillation regulated by microRNAs. *I_{Na}*: Inward sodium current; *I_{K1}*: Inward rectified K⁺ current; *I_{to}*: Transient outward potassium currents; *I_{Ca-L}*: Inactivation of L-type Ca²⁺ current; *I_{Kur}*: Ultrarapid delayed rectifier K⁺ channel; *I_{Ks}*: Slow delayed rectifier potassium channel; *I_{K-Ach}*: Acetylcholine-activated inward-rectifying potassium channel; *I_{NCX}*: Inward Na⁺-Ca²⁺-exchange current; SK3: Small conductance calcium-activated potassium channel 3; *I_f*: Funny current.

regulated, whereas *miR-101*, *miR-320*, and *miR-499* were down-regulated^[14,50]. Among these, *miR-328* was the most significantly up-regulated and forced expression of *miR-328* through adenovirus infection in canine atrium as well as a transgenic approach in mice recapitulated the phenotypes of AF. Enhanced AF vulnerability by diminished L-type Ca²⁺ current, led to a shortened atrial action potential. Normalization of *miR-328* levels with antagomiR reversed the conditions, and genetic knockdown of endogenous *miR-328* dampened AF vulnerability, highlighting the importance of *miR-328* as a potential therapeutic target for AF^[49].

Other miRNAs such as *miR-21* can also act as regulators of L-type Ca²⁺ channel. Barana *et al*^[51] showed that *miR-21* was expressed in human myocytes with chronic AF. They demonstrated that this disease increases the expression of *miR-21*, which decreases *I_{Ca-L}*. Down-regulation of *I_{Ca-L}* by *miR-21*, suggests that this miR could have an important function in AF through electrical remodelling (Figure 2).

On the other hand, *miR-26* has been demonstrated to target *KCNJ2*, the gene encoding the main component of the inward rectifier potassium-channel current *I_{K1}* (KIR 2.1). As stated above, the *I_{K1}* is an important regulator of reentrant-spiral dynamics and a major component of AF-related electrical remodelling. Recently, Luo *et al*^[52] has found that *miR-26* was down-regulated in AF patients accompanied by *I_{K1}*/KIR2.1 protein up-regulation. Moreover, by gain and loss-of-function experiments in mice, these authors demonstrated that *miR-26* controls the expression of *KCNJ2* and thereby promotes AF.

In addition, it has also been shown that expression levels of *miR-1* (a muscle specific miRNA) are greatly reduced in the atrial tissue of AF patients. Yang *et al*^[53] showed that the elimination of *miR-1* by an antisense inhibitor in infarcted rat hearts relieved arrhythmogenesis, whereas *miR-1* overexpression slowed conduction and depolarized the cytoplasmic membrane by post-transcriptionally repressing *KCNJ2* and *GJA1* (which encodes connexin 43). More

recently it has been documented that increased *I_{K1}* currents in left atrial cardiomyocytes of AF patients are associated with greater *Kir2.1* expression and lower levels of *miR-1*, reinforcing the notion of a role for *miR-1* in regulating atrial remodelling in AF^[48]. However, no significant changes in *Cx43* mRNA and protein expression nor in *Cx43* localization were found in those cardiomyocytes, suggesting that *miR-1* may differentially regulate subunits such as *Kir2.1* and *Cx43* in different cell subtypes^[48]. Moreover, a very recent miR array analysis in humans has revealed that *miR-1* is greater in the left atrium than in the right one, underscoring the notion that AF pathophysiology may affect the two atria differently^[54] (Figure 2).

The possibility that another muscle-specific miRNA such as *miR-133* plays a role in AF has also been proposed, since down-regulation of *miR-133* favours atrial remodelling by derepression of *KCNH2/I_{Kr}* and *KCNH1/I_{Ks}*. However, there is no direct evidence on how *miR-133* affects *I_{Ks}* or *I_{Kr}* current^[55,56]. In this way, Luo *et al*^[52] found that *miR-1* and *miR-133* act to restrict overexpression of *HCN2* and *HCN4* (encoding for important components of *I_f* currents), and down-regulation of *miR-1/miR-133* acts partially causing the anomalous re-expression of *HCN2* and *HCN4* in cardiac hypertrophy. Yet it remains unclear whether *miR-1* and *miR-133* affect *HCN* channels in the context of AF^[57]. Notwithstanding, *miR-133* together with *miR-30* were found to be down-regulated in a canine model of AF, reinforcing the potential role of *miR-133* in AF^[58] (Figure 2).

Together with *miR-1* and *miR-133*, *miR-208a* is also a muscle-specific miRNA, which is encoded within an intron of the α -cardiac muscle myosin heavy-chain gene (*Myh6*). Although no direct evidence for the involvement of *miR-208a* in atrial electrical remodelling have been reported, *miR-208a* transgenic gain of function in mice lead to arrhythmias. In addition, *Cx40* expression was increased in those transgenic mice, indicating that *miR-208a* could modulate electrical conduction in the heart^[59] (Figure 2).

Finally, in a recently reported finding, a potential role

of *miR-499* in AF-related electrical remodelling has been proposed. Atrial *miRNA-499* is significantly upregulated in AF, (Figure 2), leading to down-regulation of the small-conductance SK3, possibly contributing to the electrical remodelling in AF^[39].

MiRNA research has highlighted the roles of miRNAs in the cardiovascular system and their involvement in AF. Further studies on miRNA and AF will provide new insights into the mechanisms underlying that disorder and may offer new strategies for the treatment of AF.

CONCLUDING REMARKS AND CLINICAL PERSPECTIVES

Electrical remodelling secondary to the onset of atrial fibrillation dramatically contributes to the recurrence of AF, eventually leading to persistent AF. Many recent studies demonstrate that miRNAs are becoming one of the most fascinating regulatory molecules, due to their critical roles in fine-tuning of physiological processes and their deregulation in several disorders, including AF. The functional significance of miRs as direct or indirect post-transcriptional regulators of ion channels involved in electrical remodelling strongly suggests that these miRNAs may serve as potential biomarkers or promising drug targets in the treatment, and management of AF, though considerable work remains to be done. Further studies making use of existing AF models, miRNA expression signatures and miRNA target gene prediction should provide information finally applicable to clinical practice.

REFERENCES

- 1 **Kannel WB**, Wolf PA, Benjamin EJ, Levy D. Prevalence, incidence, prognosis, and predisposing conditions for atrial fibrillation: population-based estimates. *Am J Cardiol* 1998; **82**: 2N-9N [PMID: 9809895 DOI: 10.1016/S0002-9149(98)00583-9]
- 2 **Nattel S**, Harada M. Atrial remodeling and atrial fibrillation: recent advances and translational perspectives. *J Am Coll Cardiol* 2014; **63**: 2335-2345 [PMID: 24613319 DOI: 10.1016/j.jacc.2014.02.555]
- 3 **Fujiki A**. Progression of atrial fibrillation from paroxysmal to persistent. *Circ J* 2014; **78**: 1058-1060 [PMID: 24621659 DOI: 10.1253/circj.CJ-14-0264]
- 4 **Oh S**, Kim KB, Ahn H, Cho HJ, Choi YS. Remodeling of ion channel expression in patients with chronic atrial fibrillation and mitral valvular heart disease. *Korean J Intern Med* 2010; **25**: 377-385 [PMID: 21179275 DOI: 10.3904/kjim.2010.25.4.377]
- 5 **Kaumann AJ**, Lynham JA, Brown AM. Comparison of the densities of 5-HT4 receptors, beta 1- and beta 2-adrenoceptors in human atrium: functional implications. *Naunyn Schmiedebergs Arch Pharmacol* 1996; **353**: 592-595 [PMID: 8740155 DOI: 10.1007/BF00169181]
- 6 **Bosch RF**, Zeng X, Grammer JB, Popovic K, Mewis C, Kühlkamp V. Ionic mechanisms of electrical remodeling in human atrial fibrillation. *Cardiovasc Res* 1999; **44**: 121-131 [PMID: 10615396 DOI: 10.1016/S0008-6363(99)00178-9]
- 7 **Grammer JB**, Bosch RF, Kühlkamp V, Seipel L. Molecular remodeling of Kv4.3 potassium channels in human atrial fibrillation. *J Cardiovasc Electrophysiol* 2000; **11**: 626-633 [PMID: 10868735 DOI: 10.1111/j.1540-8167.2000.tb00024.x]
- 8 **Atienza F**, Almendral J, Moreno J, Vaidyanathan R, Talkachou A, Kalifa J, Arenal A, Villacastín JP, Torrecilla EG, Sánchez A,

- Ploutz-Snyder R, Jalife J, Berenfeld O. Activation of inward rectifier potassium channels accelerates atrial fibrillation in humans: evidence for a reentrant mechanism. *Circulation* 2006; **114**: 2434-2442 [PMID: 17101853 DOI: 10.1161/CIRCULATIONAHA.106.633735]
- 9 **Dobrev D**, Friedrich A, Voigt N, Jost N, Wettwer E, Christ T, Knaut M, Ravens U. The G protein-gated potassium current I(K,ACh) is constitutively active in patients with chronic atrial fibrillation. *Circulation* 2005; **112**: 3697-3706 [PMID: 16330682 DOI: 10.1161/CIRCULATIONAHA.105.575332]
- 10 **Brandt MC**, Priebe L, Böhle T, Südkamp M, Beuckelmann DJ. The ultrarapid and the transient outward K(+) current in human atrial fibrillation. Their possible role in postoperative atrial fibrillation. *J Mol Cell Cardiol* 2000; **32**: 1885-1896 [PMID: 11013132 DOI: 10.1006/jmcc.2000.1221]
- 11 **Darbar D**, Kannankeril PJ, Donahue BS, Kucera G, Stubblefield T, Haines JL, George AL, Roden DM. Cardiac sodium channel (SCN5A) variants associated with atrial fibrillation. *Circulation* 2008; **117**: 1927-1935 [PMID: 18378609 DOI: 10.1161/CIRCULATIONAHA.107.757955]
- 12 **Kallergis EM**, Manios EG, Kanoupakis EM, Mavrakis HE, Arfanakis DA, Maliaraki NE, Lathourakis CE, Chlouverakis GI, Vardas PE. Extracellular matrix alterations in patients with paroxysmal and persistent atrial fibrillation: biochemical assessment of collagen type-I turnover. *J Am Coll Cardiol* 2008; **52**: 211-215 [PMID: 18617070 DOI: 10.1016/j.jacc.2008.03.045]
- 13 **Fujita M**, Cheng XW, Inden Y, Shimano M, Yoshida N, Inoue A, Yamamoto T, Takeshita K, Kyo S, Taguchi N, Shi GP, Kuzuya M, Okumura K, Murohara T. Mechanisms with clinical implications for atrial fibrillation-associated remodeling: cathepsin K expression, regulation, and therapeutic target and biomarker. *J Am Heart Assoc* 2013; **2**: e000503 [PMID: 24342995 DOI: 10.1161/JAHA.113.000503]
- 14 **Luo X**, Zhang H, Xiao J, Wang Z. Regulation of human cardiac ion channel genes by microRNAs: theoretical perspective and pathophysiological implications. *Cell Physiol Biochem* 2010; **25**: 571-586 [PMID: 20511702 DOI: 10.1159/000315076]
- 15 **Xu J**, Zhao J, Evan G, Xiao C, Cheng Y, Xiao J. Circulating microRNAs: novel biomarkers for cardiovascular diseases. *J Mol Med (Berl)* 2012; **90**: 865-875 [PMID: 22159451 DOI: 10.1007/s00109-011-0840-5]
- 16 **Kumarswamy R**, Thum T. Non-coding RNAs in cardiac remodeling and heart failure. *Circ Res* 2013; **113**: 676-689 [PMID: 23989712 DOI: 10.1161/CIRCRESAHA.113.300226]
- 17 **Havens MA**, Reich AA, Duelli DM, Hastings ML. Biogenesis of mammalian microRNAs by a non-canonical processing pathway. *Nucleic Acids Res* 2012; **40**: 4626-4640 [PMID: 22270084 DOI: 10.1093/nar/gks026]
- 18 **Han J**, Lee Y, Yeom KH, Kim YK, Jin H, Kim VN. The Drosha-DGCR8 complex in primary microRNA processing. *Genes Dev* 2004; **18**: 3016-3027 [PMID: 15574589 DOI: 10.1101/gad.1262504]
- 19 **Han J**, Lee Y, Yeom KH, Nam JW, Heo I, Rhee JK, Sohn SY, Cho Y, Zhang BT, Kim VN. Molecular basis for the recognition of primary microRNAs by the Drosha-DGCR8 complex. *Cell* 2006; **125**: 887-901 [PMID: 16751099 DOI: 10.1016/j.cell.2006.03.043]
- 20 **Kwekkeboom RF**, Lei Z, Doevendans PA, Musters RJ, Sluijter JP. Targeted delivery of miRNA therapeutics for cardiovascular diseases: opportunities and challenges. *Clin Sci (Lond)* 2014; **127**: 351-365 [PMID: 24895056 DOI: 10.1042/CS20140005]
- 21 **Wilkins BJ**, Molkenin JD. Calcium-calcineurin signaling in the regulation of cardiac hypertrophy. *Biochem Biophys Res Commun* 2004; **322**: 1178-1191 [PMID: 15336966 DOI: 10.1016/j.bbrc.2004.07.121]
- 22 **da Costa Martins PA**, Bourajaj M, Gladka M, Kortland M, van Oort RJ, Pinto YM, Molkenin JD, De Windt LJ. Conditional dicer gene deletion in the postnatal myocardium provokes spontaneous cardiac remodeling. *Circulation* 2008; **118**: 1567-1576 [PMID: 18809798 DOI: 10.1161/circulationaha.108.769984]
- 23 **Wang Z**. *MicroRNAs and Cardiovascular Disease*. Bentham

Science Publishers, 2010

- 24 **Gomes da Silva AM**, Silbiger VN. miRNAs as biomarkers of atrial fibrillation. *Biomarkers* 2014; **19**: 631-636 [PMID: 25171770 DOI: 10.3109/1354750X.2014.954001]
- 25 **Wang J**, Huang W, Xu R, Nie Y, Cao X, Meng J, Xu X, Hu S, Zheng Z. MicroRNA-24 regulates cardiac fibrosis after myocardial infarction. *J Cell Mol Med* 2012; **16**: 2150-2160 [PMID: 22260784 DOI: 10.1111/j.1582-4934.2012.01523.x]
- 26 **Piccinini AM**, Midwood KS. Illustrating the interplay between the extracellular matrix and microRNAs. *Int J Exp Pathol* 2014; **95**: 158-180 [PMID: 24761792 DOI: 10.1111/iep.12079]
- 27 **Wang Z**, Yang B. MicroRNA Expression Detection Methods. Berlin, Heidelberg, Germany: Springer, 2010 [DOI: 10.1007/978-3-642-04928-6]
- 28 **Latronico MV**, Condorelli G. MicroRNAs and cardiac pathology. *Nat Rev Cardiol* 2009; **6**: 419-429 [DOI: 10.1038/nrcardio.2009.56]
- 29 **Latronico MV**, Condorelli G. RNA silencing: small RNA-mediated posttranscriptional regulation of mRNA and the implications for heart electrophysiology. *J Cardiovasc Electrophysiol* 2009; **20**: 230-237 [PMID: 19017333 DOI: 10.1111/j.1540-8167.2008.01357.x]
- 30 **Wang Z**. miRNA in the regulation of ion channel/transporter expression. *Compr Physiol* 2013; **3**: 599-653 [PMID: 23720324 DOI: 10.1002/cphy.c110002]
- 31 **Maass AH**, Van Gelder IC. Atrial resynchronization therapy: a new concept for treatment of heart failure with preserved ejection fraction and prevention of atrial fibrillation? *Eur J Heart Fail* 2012; **14**: 227-229 [PMID: 22357574 DOI: 10.1093/eurjhf/hfs014]
- 32 **Gaspo R**, Bosch RF, Bou-Abboud E, Nattel S. Tachycardia-induced changes in Na⁺ current in a chronic dog model of atrial fibrillation. *Circ Res* 1997; **81**: 1045-1052 [PMID: 9400386 DOI: 10.1161/01.RES.81.6.1045]
- 33 **Van Wagoner DR**, Pond AL, Lamorgese M, Rossie SS, McCarthy PM, Nerbonne JM. Atrial L-type Ca²⁺ currents and human atrial fibrillation. *Circ Res* 1999; **85**: 428-436 [PMID: 10473672 DOI: 10.1161/01.RES.85.5.428]
- 34 **Greiser M**, Halaszovich CR, Frechen D, Boknik P, Ravens U, Dobrev D, Lückhoff A, Schotten U. Pharmacological evidence for altered src kinase regulation of I (Ca,L) in patients with chronic atrial fibrillation. *Naunyn Schmiedebergs Arch Pharmacol* 2007; **375**: 383-392 [PMID: 17593353 DOI: 10.1007/s00210-007-0174-6]
- 35 **Voigt N**, Dobrev D. Ion Channel Remodelling in Atrial Fibrillation. *European Cardiology* 2011; **7**: 97-103 [DOI: 10.15420/ecr.2011.7.2.97]
- 36 **Ellinor PT**, Lunetta KL, Glazer NL, Pfeufer A, Alonso A, Chung MK, Sinner MF, de Bakker PI, Mueller M, Lubitz SA, Fox E, Darbar D, Smith NL, Smith JD, Schnabel RB, Soliman EZ, Rice KM, Van Wagoner DR, Beckmann BM, van Noord C, Wang K, Ehret GB, Rotter JI, Hazen SL, Steinbeck G, Smith AV, Launer LJ, Harris TB, Makino S, Nelis M, Milan DJ, Perz S, Esko T, Köttgen A, Moebus S, Newton-Cheh C, Li M, Möhlenkamp S, Wang TJ, Kao WH, Vasani RS, Nöthen MM, MacRae CA, Stricker BH, Hofman A, Uitterlinden AG, Levy D, Boerwinkle E, Metspalu A, Topol EJ, Chakravarti A, Gudnason V, Psaty BM, Roden DM, Meitinger T, Wichmann HE, Witteman JC, Barnard J, Arking DE, Benjamin EJ, Heckbert SR, Kääb S. Common variants in KCNN3 are associated with lone atrial fibrillation. *Nat Genet* 2010; **42**: 240-244 [PMID: 20173747 DOI: 10.1038/ng.537]
- 37 **Tuteja D**, Xu D, Timofeyev V, Lu L, Sharma D, Zhang Z, Xu Y, Nie L, Vázquez AE, Young JN, Glatzer KA, Chiamvimonvat N. Differential expression of small-conductance Ca²⁺-activated K⁺ channels SK1, SK2, and SK3 in mouse atrial and ventricular myocytes. *Am J Physiol Heart Circ Physiol* 2005; **289**: H2714-H2723 [PMID: 16055520 DOI: 10.1152/ajpheart.00534.2005]
- 38 **Li N**, Timofeyev V, Tuteja D, Xu D, Lu L, Zhang Q, Zhang Z, Singapur A, Albert TR, Rajagopal AV, Bond CT, Periasamy M, Adelman J, Chiamvimonvat N. Ablation of a Ca²⁺-activated K⁺ channel (SK2 channel) results in action potential prolongation in atrial myocytes and atrial fibrillation. *J Physiol* 2009; **587**: 1087-1100 [PMID: 19139040 DOI: 10.1113/jphysiol.2008.167718]
- 39 **Ling TY**, Wang XL, Chai Q, Lau TW, Koestler CM, Park SJ, Daly RC, Greason KL, Jen J, Wu LQ, Shen WF, Shen WK, Cha YM, Lee HC. Regulation of the SK3 channel by microRNA-499—potential role in atrial fibrillation. *Heart Rhythm* 2013; **10**: 1001-1009 [PMID: 23499625 DOI: 10.1016/j.hrthm.2013.03.005]
- 40 **Stilitano F**, Lonardo G, Giunti G, Del Lungo M, Coppini R, Spinelli V, Sartiani L, Poggesi C, Mugelli A, Cerbai E. Chronic atrial fibrillation alters the functional properties of If in the human atrium. *J Cardiovasc Electrophysiol* 2013; **24**: 1391-1400 [PMID: 23869794 DOI: 10.1111/jce.12212]
- 41 **Chu HM**, Feng MJ, Li YG, Zhang YX, Ma JF, He B, Yu YB, Liu J, Chen XM. Polymorphisms but not mutations of the KCNQ1 gene are associated with lone atrial fibrillation in the Chinese Han population. *ScientificWorldJournal* 2013; **2013**: 373454 [PMID: 23710137 DOI: 10.1155/2013/373454]
- 42 **Ki CS**, Jung CL, Kim HJ, Baek KH, Park SJ, On YK, Kim KS, Noh SJ, Youm JB, Kim JS, Cho H. A KCNQ1 mutation causes age-dependant bradycardia and persistent atrial fibrillation. *Pflugers Arch* 2014; **466**: 529-540 [PMID: 23989646 DOI: 10.1007/s00424-013-1337-6]
- 43 **Hancox JC**, Kharche S, El Harchi A, Stott J, Law P, Zhang H. In silico investigation of a KCNQ1 mutation associated with familial atrial fibrillation. *J Electrocardiol* 2014; **47**: 158-165 [PMID: 24411289 DOI: 10.1016/j.jelectrocard.2013.12.004]
- 44 **Wang QS**, Wang XF, Chen XD, Yu JF, Wang J, Sun J, Lu SB, Shen MY, Lu M, Li YG, Jin L. Genetic polymorphism of KCNH2 confers predisposition of acquired atrial fibrillation in Chinese. *J Cardiovasc Electrophysiol* 2009; **20**: 1158-1162 [PMID: 19490382 DOI: 10.1111/j.15408167.2009.01494.x]
- 45 **Kovoor P**, Wickman K, Maguire CT, Pu W, Gehrman J, Berul CI, Clapham DE. Evaluation of the role of I(KACh) in atrial fibrillation using a mouse knockout model. *J Am Coll Cardiol* 2001; **37**: 2136-2143 [PMID: 11419900 DOI: 10.1016/S0735-1097(01)01304-3]
- 46 **van der Velden HM**, Ausma J, Rook MB, Hellemons AJ, van Veen TA, Allessie MA, Jongasma HJ. Gap junctional remodeling in relation to stabilization of atrial fibrillation in the goat. *Cardiovasc Res* 2000; **46**: 476-486 [PMID: 10912458 DOI: 10.1016/S0008-6363(00)00026-2]
- 47 **Bagwe S**, Berenfeld O, Vaidya D, Morley GE, Jalife J. Altered right atrial excitation and propagation in connexin40 knockout mice. *Circulation* 2005; **112**: 2245-2253 [PMID: 16203917 DOI: 10.1161/CIRCULATIONAHA.104.527325]
- 48 **Girmatsion Z**, Biliczki P, Bonauer A, Wimmer-Greinecker G, Scherer M, Moritz A, Bukowska A, Goette A, Nattel S, Hohnloser SH, Ehrlich JR. Changes in microRNA-1 expression and IK1 up-regulation in human atrial fibrillation. *Heart Rhythm* 2009; **6**: 1802-1809 [PMID: 19959133 DOI: 10.1016/j.hrthm.2009.08.035]
- 49 **Lu Y**, Zhang Y, Wang N, Pan Z, Gao X, Zhang F, Zhang Y, Shan H, Luo X, Bai Y, Sun L, Song W, Xu C, Wang Z, Yang B. MicroRNA-328 contributes to adverse electrical remodeling in atrial fibrillation. *Circulation* 2010; **122**: 2378-2387 [PMID: 21098446 DOI: 10.1161/circulationaha.110.958967]
- 50 **Olesen MS**, Refsgaard L, Holst AG, Larsen AP, Grubb S, Haunsø S, Svendsen JH, Olesen SP, Schmitt N, Calloe K. A novel KCND3 gain-of-function mutation associated with early-onset of persistent lone atrial fibrillation. *Cardiovasc Res* 2013; **98**: 488-495 [PMID: 23400760 DOI: 10.1093/cvr/cvt028]
- 51 **Barana A**, Matamoros M, Dolz-Gaitón P, Pérez-Hernández M, Amorós I, Núñez M, Sacristán S, Pedraz Á, Pinto Á, Fernández-Avilés F, Tamargo J, Delpón E, Caballero R. Chronic atrial fibrillation increases microRNA-21 in human atrial myocytes decreasing L-type calcium current. *Circ Arrhythm Electrophysiol* 2014; **7**: 861-868 [PMID: 25107449 DOI: 10.1161/CIRCEP.114.001709]

- 52 **Luo X**, Pan Z, Shan H, Xiao J, Sun X, Wang N, Lin H, Xiao L, Maguy A, Qi XY, Li Y, Gao X, Dong D, Zhang Y, Bai Y, Ai J, Sun L, Lu H, Luo XY, Wang Z, Lu Y, Yang B, Nattel S. MicroRNA-26 governs profibrillatory inward-rectifier potassium current changes in atrial fibrillation. *J Clin Invest* 2013; **123**: 1939-1951 [PMID: 23543060 DOI: 10.1172/JCI62185]
- 53 **Yang B**, Lin H, Xiao J, Lu Y, Luo X, Li B, Zhang Y, Xu C, Bai Y, Wang H, Chen G, Wang Z. The muscle-specific microRNA miR-1 regulates cardiac arrhythmogenic potential by targeting GJA1 and KCNJ2. *Nat Med* 2007; **13**: 486-491 [PMID: 17401374 DOI: 10.1038/nm1569]
- 54 **Slagsvold KH**, Johnsen AB, Rognmo O, Høydal M, Wisløff U, Wahba A. Comparison of left versus right atrial myocardium in patients with sinus rhythm or atrial fibrillation - an assessment of mitochondrial function and microRNA expression. *Physiol Rep* 2014; pii: e12124 [PMID: 25168873 DOI: 10.14814/phy2.12124]
- 55 **Wang Z**, Fermini B, Nattel S. Rapid and slow components of delayed rectifier outward current in human atrial myocytes. *Cardiovasc Res* 1994; **28**: 1540-1515 [DOI: 10.1093/cvr/28.10.1540]
- 56 **Luo X**, Xiao J, Lin H, Li B, Lu Y, Yang B, Wang Z. Transcriptional activation by stimulating protein 1 and post-transcriptional repression by muscle-specific microRNAs of IKs-encoding genes and potential implications in regional heterogeneity of their expressions. *J Cell Physiol* 2007; **212**: 358-367 [PMID: 17443681 DOI: 10.1002/jcp.21030]
- 57 **Luo X**, Lin H, Pan Z, Xiao J, Zhang Y, Lu Y, Yang B, Wang Z. Down-regulation of miR-1/miR-133 contributes to re-expression of pacemaker channel genes HCN2 and HCN4 in hypertrophic heart. *J Biol Chem* 2008; **283**: 20045-20052 [PMID: 18458081 DOI: 10.1074/jbc.M801035200]
- 58 **Li H**, Li S, Yu B, Liu S. Expression of miR-133 and miR-30 in chronic atrial fibrillation in canines. *Mol Med Rep* 2012; **5**: 1457-1460 [PMID: 22407060 DOI: 10.3892/mmr.2012.831]
- 59 **Callis TE**, Pandya K, Seok HY, Tang RH, Tatsuguchi M, Huang ZP, Chen JF, Deng Z, Gunn B, Shumate J, Willis MS, Selzman CH, Wang DZ. MicroRNA-208a is a regulator of cardiac hypertrophy and conduction in mice. *J Clin Invest* 2009; **119**: 2772-2786 [PMID: 19726871 DOI: 10.1172/JCI36154]

P- Reviewer: Cheng XW, Liu C, Salles GF **S- Editor:** Tian YL
L- Editor: A **E- Editor:** Wu HL





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>

