**Name of journal: World Journal of Gastrointestinal Surgery**

**ESPS Manuscript NO: 17707**

**Manuscript Type: Editorial**

**Detection methods and clinical significance of free peritoneal tumor cells found during colorectal cancer surgery**

Sibio S *et al.* Free peritoneal cells in colorectal surgery

**Simone Sibio, Cristina Fiorani, Carmine Stolfi, Andrea Divizia, Roberto Pezzuto, Fabrizio Montagnese, Giulia Bagaglini, Paolo Sammartino, Giuseppe S Sica**

**Simone Sibio, Cristina Fiorani, Carmine Stolfi, Andrea Divizia, Roberto Pezzuto, Fabrizio Montagnese, Giulia Bagaglini, Paolo Sammartino, Giuseppe S Sica,** Department of Surgical Science, Tor Vergata University of Rome, 00133 Rome, Italy

**Author contributions:** Sica GS and Sibio S wrote the paper; Fiorani C, Stolfi C, Divizia A, Pezzuto R, Bagaglini G and Sammartino P performed the literature review; Montagnese F and Sica GS critically reviewed the paper.

**Conflict-of-interest statement:** All the authors of this manuscript have no conflict of interest.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Correspondence to: Giuseppe S Sica, MD, PhD,** Department of Surgical Science, Tor Vergata University of Rome, viale Oxford 81, 00133 Rome, Italy. sigisica@gmail.com

**Telephone:** +39-06-20902927

**Fax:** +39-06-20902925

**Received:** March 19, 2015

**Peer-review started:** March 21, 2015

**First decision:** April 10, 2015

**Revised:** June 10, 2015

**Accepted:** July 11, 2015

**Article in press:**

**Published online:**

**Abstract**

Peritoneal washing is now part of the standard clinical practice in several abdominal and pelvic neoplasias. However, in colorectal cancer surgery, intra-peritoneal free cancer cells (IFCC) presence is not routinely investigated and their prognostic meaning is still unclear. When peritoneal washing results are positive for the presence of IFCC a worse outcome is usually expected in these colorectal cancer operated patients, but it what is not clear is whether it is associated with an increased risk of local recurrence. It is authors’belief that one of the main reasons why IFCC are not researched as integral part of the routine staging system for colon cancer is that there still isn’t a diagnostic or detection method with enough sensibility and specificity. However, the potential clinical implications of a routine research for the presence IFCC in colon neoplasias are enormous: not only to obtain a more accurate clinical staging but also to offer different therapy protocols, based on the presence of IFCC. Based on this, adjuvant chemotherapy could be offered to those patients found to be positive for IFCC; also, protocols of proactive intraperitoneal chemotherapy could be applied). Although presence of IFCC appears to have a valid prognostic significance, further studies are needed to standardize detection and examination procedures, to determine if there are and which are the stages more likely to benefit from routine search for IFCC.

**Key words:** Colorectal cancer; Free cancer cells; Peritoneal recurrence; Cytology; Proactive treatment

**© The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** This invited editorial looks for the role of intra-peritoneal free cancer cells (IFCC) in the surgical practice for colorectal cancer. Prognostic significance of IFCC in colorectal cancer patients is still not clear. Several studies have been published but detection systems are still highly heterogeneous and results remain misleading. Peritoneal cytology could be useful in early-stage cancers to identify subsets of patients with potential worse prognosis, who may be good candidates for adjuvant treatment or even prophylactic intraperitoneal chemotherapy. Current available data need stronger validation to include IFCC in the routine staging protocols of colorectal cancer patients. However, it is the authors’ belief that cancer cells found free in the peritoneum of patients with colon cancer, must have a biological and a clinical role. The means of detection based on real time polymerase chain reaction, will surely add power to conventional cytology and with the improvement in sensibility of the methods the clinical role of IFCC could eventually become clear. New therapy protocols might be applied.

Sibio S, Fiorani C, Stolfi C, Divizia A, Pezzuto R, Montagnese F, Bagaglini G, Sammartino P, Sica GS. Detection methods and clinical significance of free peritoneal tumor cells found during colorectal cancer surgery. *World J Gastrointest Surg* 2015; In press

**INTRODUCTION**

Worldwide, colorectal cancer is the third most prevalent malignancy accounting for over 1 million new cases per year with more than 500000 deaths[1]. Incidence in Europe exceeds 400000 per year [2].

More than 25%-35% of patients with either early or advanced colorectal cancer will develop peritoneal recurrence alone after a first line treatment; peritoneal carcinomatosis is present in up to 44% of patients with recurrent colorectal cancer; the presence of synchronous or metachronous peritoneal metastasis is associated with poor prognosis[3,4] accounting for more than one third of all deaths.

Despite recent advances in gaining a thorough knowledge of clinical, biological and pathological behavior of colorectal cancer, the most commonly used staging systems for colorectal cancer are the Tumor-Node-Metastasis (TNM) and the modified Duke’s staging systems[5-7]. Expected prognosis, treatment choice and adjuvant chemotherapy are based on results provided by these staging systems which consider the local extension of the disease, the lymph nodes involvement and the eventual presence of distant metastases[8].

Both these staging systems are highly heterogeneous within each stage, sometimes resulting in very different prognosis for patients that share the same stage. This is particularly true in patients in stage II and III[9-11] and it may explain why also patients with early stage cancers could develop local or distant recurrence. Furthermore, according to this staging procedure, early stage patients are not appropriate candidates for adjuvant chemotherapy even though the probability of peritoneal recurrence in these patients seems to be equal to those with more advanced cancers.

Recent studies identified only a limited subset of patients suitable for the extensive surgical treatment and the intraperitoneal heated chemotherapy (HIPEC) with promising results in terms of both recovery from disease and overall survival[12-16]. Nevertheless, ten years survival in this limited cohort of patients appears to be a disappointing accomplishment in most qualified studies, in 10% of patients[17]. More recently extensive surgery and HIPEC has been proposed to prevent peritoneal recurrence in selected cases found to have advanced mucinous cancers with positive peritoneal lavage: although the first results seem promising, such an approach is still merely investigational[18-20].

Peritoneal cytology from peritoneal lavage is routinely performed in in staging esophageal - gastric and pancreatic malignancies and it has a definitive prognostic role in ovarian cancer[21-26]. Free intra-peritoneal cancer cells (IFCC) dissemination can occur either spontaneously or because of surgical manipulation, and follows a complex mechanism of circulation, adhesion and invasion of peritoneal surfaces.

**MECHANISM OF PERITONEAL SHEDDING, CIRCULATION AND SEEDING OF CANCER CELLS**

IFCC are found in peritoneal washing of as much as 25% of colorectal cancer patients[27]. Mechanisms of seeding and the cascade of events, which may lead to their adhesion to peritoneal surface and subsequent peritoneal metastasis development, consist in several well-defined steps. Detachment of cells from primary tumor is the first and it can occur spontaneously. Down regulation of cell adhesion molecules CAMs, such as E-cadherin, associated with high interstitial pressure due to the lack of a well organized lymphatic drainage inside the tumor explains this mechanism, which is effective just when the tumor involves the colon serosal surface (T3) or when spontaneous bowel perforation occurs[28,29].

Surgery itself represents a highly effective mechanism to (that favors) peritoneal cancer spread. Theoretically, even when tumor’s manipulation is limited, tumor spill is possible from blood or lymphatic vessels section[30]. This cells show proliferation and invasive potentials and are capable of developing metastasis. Once detached, cells follow well known peritoneal routes which are the same of peritoneal fluid drainage and reabsorption, driven by gravity force, diaphragmatic excursion and mesenteric reflections, towards and from the pelvis, along the right para-colic gutter and the sub-diaphragmatic space. Moreover, tumor cells showed inherent motility[31]. Another possible iatrogenic mechanism of free cancer cells spilling and diffusion could be associated with the laparoscopic technique[32]. However, large clinical trials found no differences in peritoneal recurrence risk between open and laparoscopic surgery for colorectal cancer, possibly because the carbon dioxide potential effect is minimized by the reduced peritoneal trauma of the laparoscopic access[33]. A preferred location for free tumor cells seeding is represented by the omentum because of its discontinuous mesothelial lining and the presence of milky spots.

Surgery contributes to tumor cells spilling and adhesion even by other post surgical physiological effects: post-operative tissue inflammation and wound healing is mediated by macrophages which produce an array of mediators able to enhance tumor growth, while pro-inflammatory cytokines enhance mesothelial adhesion molecules expression. Even fibrin can entrap tumor cells during the wound healing process. After mesothelial adhesion, tumor cells become able to pass through the peritoneal discontinuity areas or even to promote mesothelial cells apoptosis as well[34]. Matrix metallo-proteinases inhibition seems also to be associated with extracellular matrix degradation, thus allowing tumor cells invasion of layers.

**IFCC DETECTION METHODS**

Several studies tried to detect IFCC immediately before and after curative surgery for colorectal cancer[35-37], using different methods and arrays with different sensitivity. A recent large review on this issue by Mohan revised 18 studies out of 3805 found, on 3197 colorectal cancer patients; large heterogenicity was found in peritoneal washing methods in terms of volume and solutions, timing of washing, and laboratory techniques[38]. Most used techniques include conventional cytology, immunological or radio-immunoassays methods, molecular techniques as real time or endpoint polymerase chain reaction (PCR). Heterogeneity of peritoneal washing techniques, timing and samples analysis accounts for the main issue in clarifying the impact of intraperitoneal free cancer cells on prognosis and risk to develop peritoneal recurrence in colorectal cancer patients[39].

Disseminated intraperitoneal cancer cells in colorectal cancer patients may be detected using a range of techniques including examination of peritoneal fluid using conventional cytology, cytology following immune-marker staining, PCR or immunocytochemistry.

The timing of the detection may vary and can occur either pre or post-tumor resection[40].

**CONVENTIONAL CYTOLOGY AND CYTOLOGY FOLLOWING IMMUNE-MARKERS STAINING**

Peritoneal cytology can be performed without lavage when free peritoneal fluid is present. In the absence of peritoneal fluid, a lavage with saline serum (NaCl 0.9%) is needed. The volume of fluid used is extremely variable, ranging from 50 to 1000 mL, but most authors use a small amount of liquid (100-200 mL) delivered around the tumor, where most cells are usually found. Wet fixed direct smears are prepared from the aspirated material after centrifugation and discarding the supernatant. Two or three slides are fixed immediately to prevent cell degeneration. Papanicolaou is a highly suitable staining method also to sediment preparations from fluids. It offers a great advantage with regard to comparative cell studies in histological sections. However Hematoxylin-Eosin, May-Grünwald-Giemsa, Diff-Quik, and other staining methods are also used.

Cyto-centrifuge preparation is recommended for small amounts of fluid with sparse cellular content. Thin layer preparation (cytospin, ThinPrep, and others) is becoming more and more popular. The method may be preferred for adjacent analyses. The remaining cellular material should be retained and stored at 4 °C, mixed up with a certain amount of the supernatant.

Adenocarcinoma of the large bowel may be suggested by those cases that display small and large compact irregular papillary clusters. The epithelial glandular cells are large and cylindrical, and show a palisade arrangement.

Immuno-stains for CK7 and CK20 expression yield a negative and a positive staining result, respectively[41,42].

**IMMUNOCYTOCHEMISTRY**

Immuno-cytochemical methods are based on the staining of tumor cells using specific antibodies against tissue (tumor) specific antigens. Target antigens include EpCAM members of the cytokeratin family or other antigens, which are overexpressed on tumor cells (HER2-neu or MUC-1) but not on other normal cells. These antibodies are either directly labeled with horseradish peroxidase, alkaline phosphatase or fluorescent proteins, or otherwise the antigen-antibody complex is visualized by a labeled second antibody, as in the alkaline phosphatase anti-alkaline phosphatase (APAAP) method. To discriminate between malignant cells and non-specifically stained non-malignant cells, an additional evaluation step that includes morphological criteria and/or an additional counterstaining is necessary. This step employs a second tumor cell-specific antigen or an antibody against a CD marker as CD45. During the last years several computer-aided search systems have been developed which are used for scanning of microscope slide analysis. Enriched fractions are commonly centrifuged onto microscope slides (cytospins) for immune-cytochemical detections.

**PCR-BASED METHODS**

PCR-based methods are widely used for the detection of IFCC, targeting both DNA and RNA markers. DNA is generally stable and independent of the transcription mechanism of the cell. DNA markers are used based on specific genetic abnormalities that occur in certain types of cancers, although it has been reported that, at least in some cases, disseminated tumor cells are not necessarily clonal to the primary tumor. In general, few chromosomal alterations specifically characterize certain types of cancer, or even are frequent enough to serve as molecular markers. The most frequently encountered genomic alterations in colorectal cancer, commonly used for the detection of free peritoneal cancer cells, include mutations in k-Ras and p53 genes, sometimes investigated together with mRNA markers. More specifically, the detection of occult tumor cells engages targeting of tumor specific mRNA, meaning mRNA that encodes for antigens that are specific either for the malignant phenotype or for the normal tissue. The use of mRNA markers is based on the notion that tumor cells continue to display the same pattern of antigen expression as their normal tissue of origin. Once released from malignant cells, mRNA is relatively unstable; therefore, once detected, mRNA markers are indicative of the presence of viable tumor cells in the examined sample[43].

In a recent meta-analysis, positive peritoneal washing was seen as an independent prognostic factor for poor survival and was associated with a greater risk of both local and systemic recurrence in colorectal cancer patients[44]. Yield rates of intraperitoneal neoplastic cells ranged from 5% to 40% depending on the methods and on the time of detection. In general, immunocytochemistry appears to result in a far greater yield of intraperitoneal neoplastic cells than either PCR or cytopathology. Furthermore it must be considered that immunocytochemistry (along with other histological staining techniques) is subjective and depends on the strength of cellular staining, while PCR-based methods have inherent problems as they detect DNA, not viable cells, and cannot delineate cancerous cells from non-malignant cells or cellular debris.

However, several cancer cell proteins may be identified by mean of PCR based methods, such as the matrix metallo-proteinase (MMP) class and specifically the MMP-7 (Figure 1) which has been recently proved a highly sensible predictive factor involved in colorectal cancer recurrence after curative treatment. In a recent article by Sica *et al*[45] expression of MMP-7 on IFCC correlated with higher recurrence rate after curative surgery for colorectal cancer and worse prognosis[45]. Patterns of expression of MMP-7 RNA transcripts in a sample of 47 patients who underwent surgery for colorectal cancer are shown in Figure 1.

**CLINICAL AND PROGNOSTIC SIGNIFICANCE**

In the last ten years, several studies attempted to state the prognostic and clinical meaning of free peritoneal cancer cells found during colorectal cancer surgery, investigating either their presence and prognostic impact[26,38,39,40,42,43,46].

If their clinical importance in gastric cancer has been clearly identified[47-51], results from this large series of studies on colorectal cancer are misleading. The first concern has to be moved to the large heterogeneity of detection techniques used: if conventional cytology appears to be very sensitive, easily applicable and low costing, its specificity is low, yielding positive results in 4% to 35.5% of series, also providing for a 2% of inconclusive examinations[42]. Immunoassays and PCR seem to be more specific as well as more expensive and subject to laboratory availability[37].

This variability partially explains the differences in results from the studies. A recently closed large trial by French authors, based on 1364 patients, found no relationship between positive cytology and incidence of recurrence and no predictive value regarding the development of peritoneal carcinomatosis. In this study positive cytology correlated with depth of invasion of colorectal wall, synchronous presence of minimal peritoneal carcinomatosis, lymph nodes metastasis, presence of ascites or not radical surgery; this reflected on survival analysis which led to worse survival in patients with positive cytology (*P* < 0.001) in univariate analysis although it didn’t reach statistical significance as independent prognostic factor[40]. Otherwise, other studies found higher risk of overall and loco-regional recurrence when peritoneal free cancer cells are found[39] as well as predictive of poorer outcome[38].

Two studies showed that poorer outcomes are associated with positive post resection washing compared to positive pre-resection one, in terms of recurrence[52] and survival[36].

In most studies, increasing disease staging correspond to higher rates of positive cytology[53] although they can be detected also in early stage patients[54], mostly in stage 2, where Lloyd found worse survival among stage 2 patients with positive cytology rather than negative ones[36].

Some authors found correlation between positive cytology and poorly differentiated cancers[55] while the correlation with mucinous or signet ring cells histology remains unclear, even because these histological types are mostly found in advanced stage diseases.

When positive cytology is found, a prophylactic intraperitoneal chemotherapy may be considered in selected cases within clinical trials.

This proactive treatment is proposed in order to prevent peritoneal diffusion in colorectal cancer patients at high risk of peritoneal metastasis. The risk factors which were identified are as follow: mucinous or signet ring cell hystologies, T3/T4 or perforated tumors and positive peritoneal cytology.

Two recent comparative studies by Sammartino *et al*[18,19] showed that on a sample of 25 patients affected by colorectal cancer at high risk of peritoneal metastasis, a more aggressive surgical treatment including omentectomy, appendectomy, hepatic round ligament resection and oophorectomy in non-fertile women, associated with prophylactic intraperitoneal hyperthermic chemotherapy led to better disease free survival and lower peritoneal recurrence rates[18-20]. It is clear, at this point, that peritoneal washing should become a standardized procedure and that the clinical implications of IFCC are potentially enormous. Effort should be spent on obtaining reliable results in terms of sensibility and specificity of the methods of analysis.

**CONCLUSION**

Positive peritoneal washing for IFCC is associated with worse outcome in colorectal cancer patients, however it is not clear whether it is associated with an increased risk of local recurrence or not. IFCC can be found in advanced stage or in the acute setting (occlusion or perforation). However, positive cytology can occur also in colorectal cancer at an earlier stage (especially TNM stage II patients) and it could affect the strategic plan of treatment. Nevertheless, available data still do not allow to include peritoneal washing and cytology as routine procedures procedures in staging colorectal cancer.

Potentially, peritoneal washing could improve the outcome of those selected patients with apparent early stage colorectal cancer, to receive adjuvant chemotherapy.

Patients with positive cytology may also become candidates to receive proactive intraperitoneal chemotherapy.

Peritoneal washing examination techniques must be improved in order to achieve a better sensitivity. It is the authors’ belief that until a proper reliable tumor marker for RT-PCR will be identified, probably the most suitable procedure remains conventional cytology. However, giving the recent studies in this direction it is desirable that highly sensible proteins such as the metallo-proteinase (MMP) class and specifically the MMP-7 are employed to increase the specificity of conventional cytology[45].

Further studies are needed to standardize detection and examination procedures, to determine if there are and which are the stages more likely to benefit from routine search for IFCC in the view of offering a proactive management, keeping in mind what Benjamin Franklin once stated: “an ounce of prevention is worth a pound of cure”.

**REFERENCES**

1 **Parkin DM**, Bray F, Ferlay J, Pisani P. Global cancer statistics, 2002. *CA Cancer J Clin* 2005; **55**: 74-108 [PMID: 15761078]

2 **Ferlay J**, Autier P, Boniol M, Heanue M, Colombet M, Boyle P. Estimates of the cancer incidence and mortality in Europe in 2006. *Ann Oncol* 2007; **18**: 581-592 [PMID: 17287242]

3 **Gellad ZF**, Provenzale D. Colorectal cancer: national and international perspective on the burden of disease and public health impact. *Gastroenterology* 2010; **138**: 2177-2190 [PMID: 20420954]

4 **Brodsky JT**, Cohen AM. Peritoneal seeding following potentially curative resection of colonic carcinoma: implications for adjuvant therapy. *Dis Colon Rectum* 1991; **34**: 723-727 [PMID: 1855433]

5 **Dukes CE**. The classification of cancer of the rectum. *J Pathol Bacteriol* 1932; **35**: 323-332

6 **Astler VB**, Coller FA. The prognostic significance of direct extension of carcinoma of the colon and rectum. *Ann Surg* 1954; **139**: 846-852 [PMID: 13159135]

7 **Hutter RV**. At last--worldwide agreement on the staging of cancer. *Arch Surg* 1987; **122**: 1235-1239 [PMID: 3675187]

8 **Sobin LH**, Hermanek P. eds. UICC International Union against Cancer TNM classification of malignant tumors 5th ed. New York: Wiley-Liss, 1997: 66-69

9 **Merkel S**, Mansmann U, Papadopoulos T, Wittekind C, Hohenberger W, Hermanek P. The prognostic inhomogeneity of colorectal carcinomas Stage III: a proposal for subdivision of Stage III. *Cancer* 2001; **92**: 2754-2759 [PMID: 11753948]

10 **Greene FL**, Stewart AK, Norton HJ. A new TNM staging strategy for node-positive (stage III) colon cancer: an analysis of 50,042 patients. *Ann Surg* 2002; **236**: 416-421; discussion 421 [PMID: 12368669]

11 **Liefers GJ**, Cleton-Jansen AM, van de Velde CJ, Hermans J, van Krieken JH, Cornelisse CJ, Tollenaar RA. Micrometastases and survival in stage II colorectal cancer. *N Engl J Med* 1998; **339**: 223-228 [PMID: 9673300]

12 **Esquivel J**, Sticca R, Sugarbaker P, Levine E, Yan TD, Alexander R, Baratti D, Bartlett D, Barone R, Barrios P, Bieligk S, Bretcha-Boix P, Chang CK, Chu F, Chu Q, Daniel S, de Bree E, Deraco M, Dominguez-Parra L, Elias D, Flynn R, Foster J, Garofalo A, Gilly FN, Glehen O, Gomez-Portilla A, Gonzalez-Bayon L, Gonzalez-Moreno S, Goodman M, Gushchin V, Hanna N, Hartmann J, Harrison L, Hoefer R, Kane J, Kecmanovic D, Kelley S, Kuhn J, Lamont J, Lange J, Li B, Loggie B, Mahteme H, Mann G, Martin R, Misih RA, Moran B, Morris D, Onate-Ocana L, Petrelli N, Philippe G, Pingpank J, Pitroff A, Piso P, Quinones M, Riley L, Rutstein L, Saha S, Alrawi S, Sardi A, Schneebaum S, Shen P, Shibata D, Spellman J, Stojadinovic A, Stewart J, Torres-Melero J, Tuttle T, Verwaal V, Villar J, Wilkinson N, Younan R, Zeh H, Zoetmulder F, Sebbag G. Cytoreductive surgery and hyperthermic intraperitoneal chemotherapy in the management of peritoneal surface malignancies of colonic origin: a consensus statement. Society of Surgical Oncology. *Ann Surg Oncol* 2007; **14**: 128-133 [PMID: 17072675]

13 **Sugarbaker PH**. Managing the peritoneal surface component of gastrointestinal cancer. Part 2. Perioperative intraperitoneal chemotherapy. *Oncology* (Williston Park) 2004; **18**: 207-19; discussion 220-2, 227-8, 230 [PMID: 15008058]

14 **Sugarbaker PH**. Managing the peritoneal surface component of gastrointestinal cancer. Part 1. Patterns of dissemination and treatment options. *Oncology* (Williston Park) 2004; **18**: 51-59 [PMID: 14768406]

15 **Glehen O**, Mithieux F, Osinsky D, Beaujard AC, Freyer G, Guertsch P, Francois Y, Peyrat P, Panteix G, Vignal J, Gilly FN. Surgery combined with peritonectomy procedures and intraperitoneal chemohyperthermia in abdominal cancers with peritoneal carcinomatosis: a phase II study. *J Clin Oncol* 2003; **21**: 799-806 [PMID: 12610177]

16 **Sugarbaker PH**, Gianola FJ, Speyer JL, Wesley R, Barofsky I, Myers CE. Prospective randomized trial of intravenous v intraperitoneal 5-FU in patients with advanced primary colon or rectal cancer. *Semin Oncol* 1985; **12**: 101-111 [PMID: 3901269]

17 **Goéré D**, Malka D, Tzanis D, Gava V, Boige V, Eveno C, Maggiori L, Dumont F, Ducreux M, Elias D. Is there a possibility of a cure in patients with colorectal peritoneal carcinomatosis amenable to complete cytoreductive surgery and intraperitoneal chemotherapy? *Ann Surg* 2013; **257**: 1065-1071 [PMID: 23299520 DOI: 10.1097/SLA.0b013e31827e9289]

18 **Sammartino P**, Sibio S, Biacchi D, Cardi M, Mingazzini P, Rosati MS, Cornali T, Sollazzo B, Atta JM, Di Giorgio A. Long-term results after proactive management for locoregional control in patients with colonic cancer at high risk of peritoneal metastases. *Int J Colorectal Dis* 2014; **29**: 1081-1089 [PMID: 24980687 DOI: 10.1007/s00384-014-1929-4]

19 **Sammartino P**, Sibio S, Biacchi D, Cardi M, Accarpio F, Mingazzini P, Rosati MS, Cornali T, Di Giorgio A. Prevention of Peritoneal Metastases from Colon Cancer in High-Risk Patients: Preliminary Results of Surgery plus Prophylactic HIPEC. *Gastroenterol Res Pract* 2012; **2012**: 141585 [PMID: 22645605 DOI: 10.1155/2012/141585]

20 **Sugarbaker PH**. Update on the prevention of local recurrence and peritoneal metastases in patients with colorectal cancer. *World J Gastroenterol* 2014; **20**: 9286-9291 [PMID: 25071322 DOI: 10.3748/wjg.v20.i28.9286]

21 **Lowe E**, McKenna H. Peritoneal washing cytology: a retrospective analysis of 175 gynaecological patients. *Aust N Z J Obstet Gynaecol* 1989; **29**: 55-61 [PMID: 2487930]

22 **Ziselman EM**, Harkavy SE, Hogan M, West W, Atkinson B. Peritoneal washing cytology. Uses and diagnostic criteria in gynecologic neoplasms. *Acta Cytol* 1984; **28**: 105-110 [PMID: 6583966]

23 **Colgan TJ**, Boerner SL, Murphy J, Cole DE, Narod S, Rosen B. Peritoneal lavage cytology: an assessment of its value during prophylactic oophorectomy. *Gynecol Oncol* 2002; **85**: 397-403 [PMID: 12051865]

24 **Nath J**, Moorthy K, Taniere P, Hallissey M, Alderson D. Peritoneal lavage cytology in patients with oesophagogastric adenocarcinoma. *Br J Surg* 2008; **95**: 721-726 [PMID: 18412292 DOI: 10.1002/bjs.6107]

25 **Bentrem D**, Wilton A, Mazumdar M, Brennan M, Coit D. The value of peritoneal cytology as a preoperative predictor in patients with gastric carcinoma undergoing a curative resection. *Ann Surg Oncol* 2005; **12**: 347-353 [PMID: 15915368]

26 **Vogel I**, Kalthoff H. Disseminated tumour cells. Their detection and significance for prognosis of gastrointestinal and pancreatic carcinomas. *Virchows Arch* 2001; **439**: 109-117 [PMID: 11561750]

27 **Ceelen WP**, Bracke ME. Peritoneal minimal residual disease in colorectal cancer: mechanisms, prevention, and treatment. *Lancet Oncol* 2009; **10**: 72-79 [PMID: 19111247 DOI: 10.1016/S1470-2045(08)70335-8]

28 **Kokenyesi R**, Murray KP, Benshushan A, Huntley ED, Kao MS. Invasion of interstitial matrix by a novel cell line from primary peritoneal carcinosarcoma, and by established ovarian carcinoma cell lines: role of cell-matrix adhesion molecules, proteinases, and E-cadherin expression. *Gynecol Oncol* 2003; **89**: 60-72 [PMID: 12694655]

29 **McArdle CS**, McMillan DC, Hole DJ. The impact of blood loss, obstruction and perforation on survival in patients undergoing curative resection for colon cancer. *Br J Surg* 2006; **93**: 483-488 [PMID: 16555262]

30 **Hansen E**, Wolff N, Knuechel R, Ruschoff J, Hofstaedter F, Taeger K. Tumor cells in blood shed from the surgical field. *Arch Surg* 1995; **130**: 387-393 [PMID: 7710337]

31 **Lindberg U**, Karlsson R, Lassing I, Schutt CE, Höglund AS. The microfilament system and malignancy. *Semin Cancer Biol* 2008; **18**: 2-11 [PMID: 18024149]

32 **Volz J**, Köster S, Spacek Z, Paweletz N. Characteristic alterations of the peritoneum after carbon dioxide pneumoperitoneum. *Surg Endosc* 1999; **13**: 611-614 [PMID: 10347302]

33 **Jayne DG**, Guillou PJ, Thorpe H, Quirke P, Copeland J, Smith AM, Heath RM, Brown JM. Randomized trial of laparoscopic-assisted resection of colorectal carcinoma: 3-year results of the UK MRC CLASICC Trial Group. *J Clin Oncol* 2007; **25**: 3061-3068 [PMID: 17634484]

34 **Heath RM**, Jayne DG, O'Leary R, Morrison EE, Guillou PJ. Tumour-induced apoptosis in human mesothelial cells: a mechanism of peritoneal invasion by Fas Ligand/Fas interaction. *Br J Cancer* 2004; **90**: 1437-1442 [PMID: 15054468]

35 **Guller U**, Zajac P, Schnider A, Bösch B, Vorburger S, Zuber M, Spagnoli GC, Oertli D, Maurer R, Metzger U, Harder F, Heberer M, Marti WR. Disseminated single tumor cells as detected by real-time quantitative polymerase chain reaction represent a prognostic factor in patients undergoing surgery for colorectal cancer. *Ann Surg* 2002; **236**: 768-775; discussion 775-776 [PMID: 12454515]

36 **Lloyd JM**, McIver CM, Stephenson SA, Hewett PJ, Rieger N, Hardingham JE. Identification of early-stage colorectal cancer patients at risk of relapse post-resection by immunobead reverse transcription-PCR analysis of peritoneal lavage fluid for malignant cells. *Clin Cancer Res* 2006; **12**: 417-423 [PMID: 16428481]

37 **Hara M**, Nakanishi H, Jun Q, Kanemitsu Y, Ito S, Mochizuki Y, Yamamura Y, Kodera Y, Tatematsu M, Hirai T, Kato T. Comparative analysis of intraperitoneal minimal free cancer cells between colorectal and gastric cancer patients using quantitative RT-PCR: possible reason for rare peritoneal recurrence in colorectal cancer. *Clin Exp Metastasis* 2007; **24**: 179-189 [PMID: 17487561]

38 **Mohan HM**, O'Connor DB, O'Riordan JM, Winter DC. Prognostic significance of detection of microscopic peritoneal disease in colorectal cancer: a systematic review. *Surg Oncol* 2013; **22**: e1-e6 [PMID: 23481599 DOI: 10.1016/j.suronc.2013.01.001]

39 **Rekhraj S**, Aziz O, Prabhudesai S, Zacharakis E, Mohr F, Athanasiou T, Darzi A, Ziprin P. Can intra-operative intraperitoneal free cancer cell detection techniques identify patients at higher recurrence risk following curative colorectal cancer resection: a meta-analysis. *Ann Surg Oncol* 2008; **15**: 60-68 [PMID: 17909914]

40 **Cotte E**, Peyrat P, Piaton E, Chapuis F, Rivoire M, Glehen O, Arvieux C, Mabrut JY, Chipponi J, Gilly FN. Lack of prognostic significance of conventional peritoneal cytology in colorectal and gastric cancers: results of EVOCAPE 2 multicentre prospective study. *Eur J Surg Oncol* 2013; **39**: 707-714 [PMID: 23601984 DOI: 10.1016/j.ejso.2013.03.021]

41 **Spieler P**, Rossle M. Non ginecological cytopathology: a practical guide. Ed. Springer, 2012: 300-313

42 **Passot G**, Mohkam K, Cotte E, Glehen O. Intra-operative peritoneal lavage for colorectal cancer. *World J Gastroenterol* 2014; **20**: 1935-1939 [PMID: 24616569 DOI: 10.3748/wjg.v20.i8.1935]

43 **Tsouma A**, Aggeli C, Pissimissis N, Lembessis P, Zografos GN, Koutsilieris M. Circulating tumor cells in colorectal cancer: detection methods and clinical significance. *Anticancer Res* 2008; **28**: 3945-3960 [PMID: 19192655]

44 **Bosanquet DC**, Harris DA, Evans MD, Beynon J. Systematic review and meta-analysis of intraoperative peritoneal lavage for colorectal cancer staging. *Br J Surg* 2013; **100**: 853-862 [PMID: 23536330 DOI: 10.1002/bjs.9118]

45 **Sica GS**, Fiorani C, Stolfi C, Monteleone G, Candi E, Amelio I, Catani V, Sibio S, Divizia A, Tema G, Iaculli E, Gaspari AL. Peritoneal expression of Matrilysin helps identify early post-operative recurrence of colorectal cancer. *Oncotarget* 2015; **6**: 13402-13415 [PMID: 25596746]

46 **Bosch B**, Guller U, Schnider A, Maurer R, Harder F, Metzger U, Marti WR. Perioperative detection of disseminated tumour cells is an independent prognostic factor in patients with colorectal cancer. *Br J Surg* 2003; **90**: 882-888 [PMID: 12854118]

47 **La Torre M**, Ferri M, Giovagnoli MR, Sforza N, Cosenza G, Giarnieri E, Ziparo V. Peritoneal wash cytology in gastric carcinoma. Prognostic significance and therapeutic consequences. *Eur J Surg Oncol* 2010; **36**: 982-986 [PMID: 20591604]

48 **Lorenzen S**, Panzram B, Rosenberg R, Nekarda H, Becker K, Schenk U, Höfler H, Siewert JR, Jäger D, Ott K. Prognostic significance of free peritoneal tumor cells in the peritoneal cavity before and after neoadjuvant chemotherapy in patients with gastric carcinoma undergoing potentially curative resection. *Ann Surg Oncol* 2010; **17**: 2733-2739 [PMID: 20490698]

49 **Katsuragi K**, Yashiro M, Sawada T, Osaka H, Ohira M, Hirakawa K. Prognostic impact of PCR-based identification of isolated tumour cells in the peritoneal lavage fluid of gastric cancer patients who underwent a curative R0 resection. *Br J Cancer* 2007; **97**: 550-556 [PMID: 17667927]

50 **Euanorasetr C**, Lertsithichai P. Prognostic significance of peritoneal washing cytology in Thai patients with gastric adenocarcinoma undergoing curative D2 gastrectomy. *Gastric Cancer* 2007; **10**: 18-23 [PMID: 17334713]

51 **Homma Y**, Hamano T, Akazawa Y, Otsuki Y, Shimizu S, Kobayashi H, Kameoka S, Kobayashi Y. Positive peritoneal washing cytology is a potential risk factor for the recurrence of curatively resected colorectal cancer. *Surg Today* 2014; **44**: 1084-1089 [PMID: 23942820 DOI: 10.1007/s00595-013-0689-z]

52 **Hase K**, Ueno H, Kuranaga N, Utsunomiya K, Kanabe S, Mochizuki H. Intraperitoneal exfoliated cancer cells in patients with colorectal cancer. *Dis Colon Rectum* 1998; **41**: 1134-1140 [PMID: 9749497]

53 **Kanellos I**, Zacharakis E, Kanellos D, Pramateftakis MG, Betsis D. Prognostic significance of CEA levels and positive cytology in peritoneal washings in patients with colorectal cancer. *Colorectal Dis* 2006; **8**: 436-440 [PMID: 16684089]

54 **Schott A**, Vogel I, Krueger U, Kalthoff H, Schreiber HW, Schmiegel W, Henne-Bruns D, Kremer B, Juhl H. Isolated tumor cells are frequently detectable in the peritoneal cavity of gastric and colorectal cancer patients and serve as a new prognostic marker. *Ann Surg* 1998; **227**: 372-379 [PMID: 9527060]

55 **Gozalan U**, Yasti AC, Yuksek YN, Reis E, Kama NA. Peritoneal cytology in colorectal cancer: incidence and prognostic value. *Am J Surg* 2007; **193**: 672-675 [PMID: 17512274]

**P-Reviewer:** He S, Hironaka S, Noguera J, Wang YH **S-Editor:** Ji FF **L-Editor: E-Editor:**



**Figure 1 Patterns of expression of matrix metallo-proteinase-7 RNA transcripts in 47 peritoneal washing samples taken from 47 patients who had undergone surgery for colorectal cancer[45].**