



Circulating predictive and diagnostic biomarkers for hepatitis B virus-associated hepatocellular carcinoma

Stijn Van Hees, Peter Michielsens, Thomas Vanwolleghem

Stijn Van Hees, Peter Michielsens, Thomas Vanwolleghem, Department of Gastroenterology and Hepatology, Antwerp University Hospital, 2650 Edegem, Belgium

Stijn Van Hees, Peter Michielsens, Thomas Vanwolleghem, Laboratory of Experimental Medicine and Pediatrics, University of Antwerp, 2610 Antwerp, Belgium

Author contributions: Van Hees S and Vanwolleghem T conceptualized the manuscript; Van Hees S wrote the manuscript; all authors contributed to the critical revision and editing of the paper; Michielsens P and Vanwolleghem T approved the final version.

Supported by Foundation Against Cancer Belgium, No. 2014-087.

Conflict-of-interest statement: The authors declare no potential conflict of interest.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: Thomas Vanwolleghem, MD, PhD, Department of Gastroenterology and Hepatology, Antwerp University Hospital, Wilrijkstraat 10, 2650 Edegem, Belgium. thomas.vanwolleghem@uza.be
Telephone: +32-3-8213853
Fax: +32-3-8214478

Received: April 26, 2016
Peer-review started: April 27, 2016
First decision: June 20, 2016
Revised: July 18, 2016
Accepted: August 5, 2016

Article in press: August 5, 2016
Published online: October 7, 2016

Abstract

Chronic hepatitis B virus (HBV) infected patients have an almost 100-fold increased risk to develop hepatocellular carcinoma (HCC). HCC is the fifth most common and third most deadly cancer worldwide. Up to 50% of newly diagnosed HCC cases are attributed to HBV infection. Early detection improves survival and can be achieved through regular screening. Six-monthly abdominal ultrasound, either alone or in combination with alpha-fetoprotein serum levels, has been widely endorsed for this purpose. Both techniques however yield limited diagnostic accuracy, which is not improved when they are combined. Alternative circulating or histological markers to predict or diagnose HCC are therefore urgently needed. Recent advances in systems biology technologies have enabled the identification of several new putative circulating biomarkers. Although results from studies assessing combinations of these biomarkers are promising, evidence for their clinical utility remains low. In addition, most of the studies conducted so far show limitations in design. Attention must be paid for instance to different ethnicities and different etiologies when studying biomarkers for hepatocellular carcinoma. This review provides an overview on the current understandings and recent progress in the field of diagnostic and predictive circulating biomarkers for hepatocellular carcinoma in chronically infected HBV patients and discusses the future prospects.

Key words: Hepatocellular carcinoma; Hepatitis B virus infection; Biomarkers; Predictive; Diagnostic; Alpha-fetoprotein; Validation; Limitations; MicroRNA

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Regular screening for hepatocellular carcinoma (HCC) in patients at risk improves their survival rates. Currently available screening methods include abdominal ultrasound and alpha-fetoprotein serum levels, but both methods lack diagnostic accuracy. Recent technological advances have enabled the identification of new predictive and diagnostic hepatitis B virus (HBV)-associated HCC biomarkers. Nevertheless, most of the studies conducted so far show design limitations. This review provides an overview on the current understanding and future prospects of circulating predictive and diagnostic biomarkers for HBV-associated HCC.

Van Hees S, Michielsen P, Vanwolleghem T. Circulating predictive and diagnostic biomarkers for hepatitis B virus-associated hepatocellular carcinoma. *World J Gastroenterol* 2016; 22(37): 8271-8282 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i37/8271.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i37.8271>

INTRODUCTION

Hepatocellular carcinoma (HCC) is the fifth most common cancer worldwide and ranks third as cancer-related death cause due to a 5-year survival of only 15%^[1]. Moreover, at a time of decreasing overall cancer-related deaths due to an immense progress in cancer diagnostics and treatment options, mortality from hepatocellular carcinoma is increasing^[1,2].

Chronic hepatitis B virus (HBV) infection is a major risk factor for HCC development. Prospective cohort studies have revealed an up to 100-fold increased risk for HCC in chronically infected HBV patients^[3]. Up to 50% of newly diagnosed HCC cases are attributed to HBV infection, due to both direct and indirect oncogenic effects of the virus^[2,4-7]. Integration of HBV DNA in the human genome may result in genomic instability, while inflammation-related oxidative stress, caused by immunological responses, may contribute indirectly to HCC development^[6-9].

Four clinical phases can be distinguished during the natural course of a HBV infection: an immune-tolerance phase, an immune active phase, an inactive carrier phase and a hepatitis B e antigen (HBeAg) negative phase^[2,10]. Patients in the immune active phase and the HBeAg negative phase are at increased risk for progression towards fibrosis and ultimately the development of cirrhosis, which is a major risk factor for HCC^[11,12].

Several years to decades are needed for HCC to develop in a HBV infected liver^[13]. Early diagnosis of HCC in HBV patients is challenging but is proven to result in an improved long-term survival due to an increased chance to detect tumors at a resectable stage^[14-19].

Importantly, also a significant number of HBV patients develop HCC in a non-cirrhotic liver^[20]. Current guidelines therefore advise 6-monthly abdominal ultrasound (US) surveillance for HCC in advanced fibrosis or cirrhotic HBV patients and in non-cirrhotic patients depending on ethnic background and age^[21-25]. The technique, however, faces a disappointing 63% sensitivity to detect HCC and is hampered by inter- and intra-observer variability^[20]. Finding biomarkers to better predict or diagnose HCC therefore remains an important clinical and research priority.

Serum alpha-fetoprotein (AFP) levels are widely used for HCC screening and diagnostics, but the clinical utility to rule out or detect HCC is still a matter of debate. The protein lacks sensitivity and specificity to detect HCC. The recent improvement of systems biology techniques, such as proteomics and genomics, has enabled the identification of several new putative biomarkers^[26,27]. This review provides an overview of diagnostic and predictive serum biomarkers for HBV-associated hepatocellular carcinoma and discusses future prospects.

DIAGNOSTIC BIOMARKERS FOR HCC: AN OVERVIEW

An overview of the discussed diagnostic circulating biomarkers with their respective sensitivities and specificities to detect HCC is displayed in Table 1. An overview of their cellular origin is displayed in Figure 1.

Alpha-fetoprotein goes off stage, its glycoforms come on stage

Alpha-fetoprotein is an oncofetal protein produced by the fetal yolk sac and liver^[28]. The protein, like albumin, binds exogenous as well as endogenous substances in blood^[29]. Physiologically elevated AFP levels are found in pregnant women and newborns, but decrease quickly after birth. Upregulation of AFP later on in life has been associated with various pathological conditions such as acute hepatitis, endodermal sinus tumors and HCC^[30-32].

Alpha-fetoprotein was discovered in the late 1950s and has been of interest for the monitoring of HCC development in viral hepatitis patients since the early 1970s^[33-35]. For a long time, AFP has been widely used together with abdominal US in routine HCC screening. Nevertheless, the most recent European and American guidelines do not endorse this practice anymore since its diagnostic accuracy is low^[21-23]. The protein, most often detected by enzyme-like immunosorbent assays (ELISA), indeed faces a lack of sensitivity and specificity to detect early stage HCC in HBV patients^[36]. Only 70% of all HCC's are characterized by markedly elevated AFP levels at the time of diagnosis^[37-41]. Large HBV cohort studies showed a maximal sensitivity for AFP of about 75% to detect HCC at optimal cut-off levels^[32,41-45].

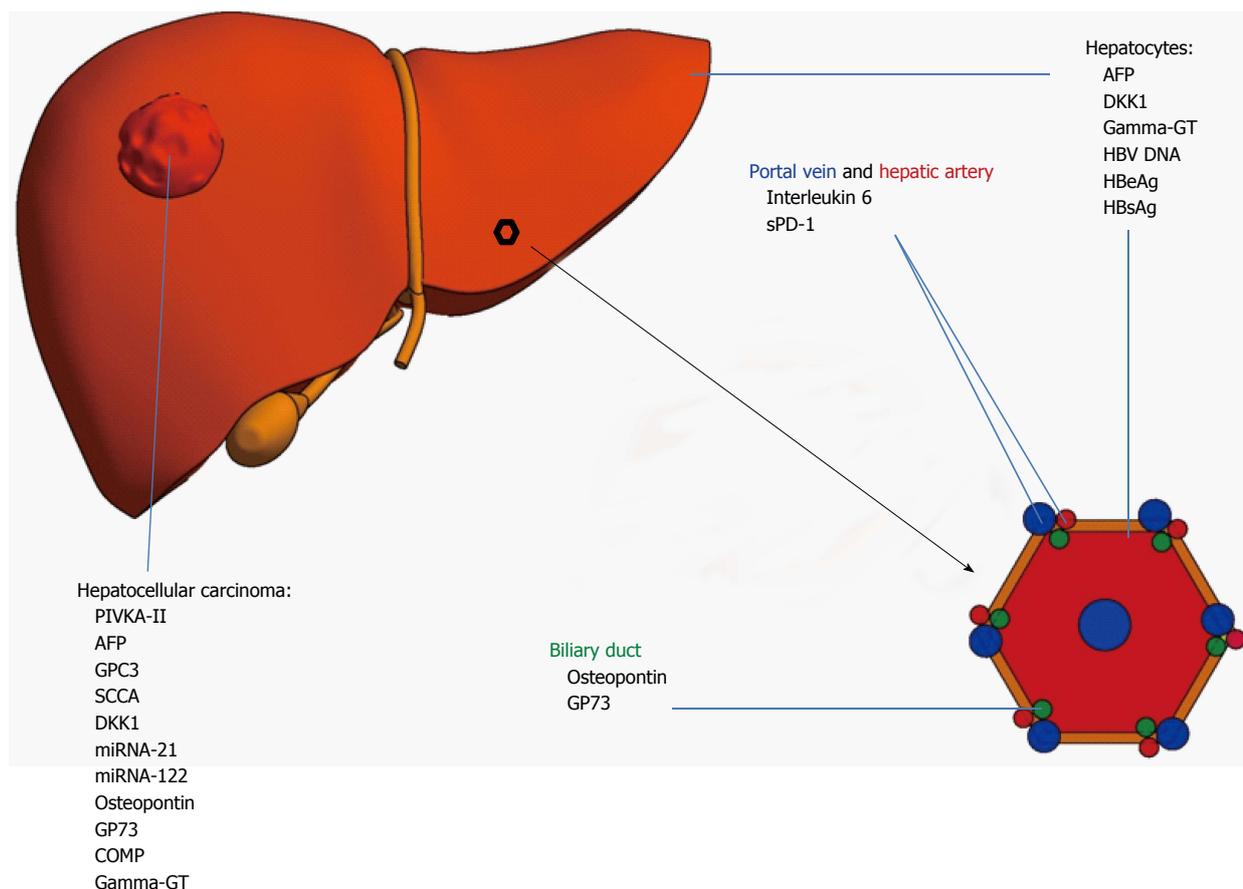


Figure 1 Cellular origin of the discussed predictive and diagnostic biomarkers in a physiological and oncological setting. Predictive biomarkers are displayed in italics. PIVKA-II: Protein induced by vitamin K absence; AFP: Alpha-fetoprotein; GPC3: Glypican-3; SCCA: Squamous cell carcinoma antigen; DKK1: Dickkopf-1 protein; miRNA: MicroRNA; COMP: Cartilage oligomeric matrix protein; GP73: Glycoprotein-73; sPD-1: Soluble programmed death-1; Gamma-GT: Gamma-glutamyltransferase.

Table 1 Diagnostic serum biomarkers for hepatitis B virus-associated hepatocellular carcinoma					
Marker	Cut-off ¹	Sensitivity	Specificity	Detection method (most reliable)	Ref.
AFP	7.7-112.0 ng/mL	25%-90%	87%-97%	ELISA	[36,73]
AFP-I3	3%-20%	36%-96%	89%-94%	Liquid-Phase Binding Assay	[46,47,52]
DCP	40-150 mAU/mL	44%-91%	68%-99%	Electrochemiluminescence immunoassay	[36,53]
Osteopontin	9.3-642.5 ng/mL	73%-97%	55%-100%	ELISA	[65]
GP73	78-150 ng/mL	68%-95%	9%-97%	Immunoblotting, Western Blotting or ELISA ²	[36,71,72]
GPC-3	2-300 ng/mL	36%-100%	40%-100%	ELISA	[36,78]
SCCA	0.12-3.80 ng/mL	42%-80%	50%-88%	ELISA	[36,81]
DKK1	1.01-2.15 ng/mL	69%-91%	62%-91%	ELISA	[86-88]
miRNA-21	NA	84%-90%	71%-92%	qRT-PCR	[83]
miRNA-122	NA	70%-82%	69%-84%	qRT-PCR	[83]

¹Range of cut-off values used in different studies included in the discussed meta-analyses and systematic reviews; ²Equally reliable. Relative levels were used with different internal standards to measure miRNA-concentrations. The choice of the most reliable detection method was based on recent studies involving at least 100 patients. Only biomarkers of which at least 4 different studies discussing their diagnostic potential exist, were included. AFP: Alpha-fetoprotein; DCP: Des-gamma carboxy prothrombin; GP73: Golgi-protein 73; GPC-3: Glypican-3; SCCA: Squamous cell carcinoma antigen; DKK1: Dickkopf-1; miRNA: MicroRNA; ELISA: Enzyme-linked immunosorbent assay; qRT-PCR: Quantitative reverse transcriptase polymerase chain reaction; NA: Not applicable.

AFP-I3: The rising star under the AFP glycoforms
 AFP is a glycoprotein of which three glycoforms exist: AFP-I1, AFP-I2 and AFP-I3. They are all characterized by an increased binding affinity for *Lens culinaris* agglutinin. AFP-I3, which shows the highest binding affinity is of particular interest as a biomarker for

hepatocellular carcinoma. This glycoform is secreted by malignant HCC cells even at early tumor stages and in the absence of elevated AFP levels and can be detected using liquid-phase binding assays^[46,47]. In addition, the fraction of AFP-I3 to total AFP in the serum correlates with the degree of malignancy^[48].

Over 15 studies have addressed the clinical potential of AFP-I3 so far with sensitivity and specificity ranging from 21% to 84% and from 89% to 94% respectively^[48-52]. However these studies assessing the clinical potential of AFP-I3 use different cut-off levels, test methods and patient numbers, resulting in a wide range of detected sensitivity. A study from 2009 measuring the fraction of AFP-I3 to total AFP using an automated immunologic analyzer and a cutoff of 10% AFP-I3 in 419 HCC patients and 417 cirrhotic controls, found a sensitivity of 42% to detect HCC^[53]. AFP-I3 fractions were measured using Western blotting in another study, involving 388 HCC patients and 212 controls with a cutoff of 15% AFP-I3 to total AFP, resulting in a sensitivity of 21%^[54]. In order to unequivocally demonstrate the superiority of AFP-I3 to AFP, large cohort studies using the same cutoff and detection method are needed. Recently AFP-I3 was suggested to be especially useful in the diagnosis of HCC in absence of elevated AFP levels, but further validation is needed^[55].

Des-gamma carboxy prothrombin

Des-gamma carboxy prothrombin (DCP) is a non-carboxylated form of prothrombin, also known as protein induced by vitamin K absence (PIVKA-II). Carboxylation takes place in the hepatocytes before the protein is released into the circulation. Release of the non-carboxylated form has been associated with vitamin K deficiency and presence of HCC^[56]. Elevated DCP levels, preferably measured using electrochemoluminescence assays, were found in sera of HCC patients, suggesting proper DCP synthesis in hepatoma cells^[36,57,58]. DCP has been investigated as a potential HCC diagnostic biomarker in several studies, showing a comparable to slightly higher diagnostic performance compared to AFP^[59-63].

Osteopontin

Osteopontin (OPN) is a glycoprotein that constitutes a major part of the extracellular matrix of bones and teeth. In addition, low levels of the protein are being secreted by biliary epithelial cells. OPN is involved in developmental as well as immunological, tumorigenic and bone homeostatic processes^[64]. Overexpression of the protein, detected using ELISA assays, was found in a wide range of tumor types including pancreas cancer, multiple myeloma and HCC^[64-66]. Seven retrospective cohort studies have investigated the diagnostic potential of OPN for HCC. So far, OPN does not outperform AFP as a diagnostic marker^[65].

Golgi protein-73

Golgi protein-73 (GP73) is a transmembrane protein physiologically located on the Golgi membrane of epithelial cells in different tissues, including the biliary tract^[67]. Its function remains largely unknown. Liver damage, caused by viral as well as non-viral agents

leads to GP73 upregulation^[68,69]. Increasing GP73 serum levels are associated with advanced fibrosis stages in HBV patients^[70,71]. A recent meta-analysis showed that GP73's diagnostic accuracy for HCC outperforms that of AFP^[72,73]. The protein can be detected using either ELISA assays, immunoblotting or Western blot. Previous studies have shown a comparable efficacy for all three methods^[36,71].

Glypican-3

Glypican-3 (GPC3) is a member of the heparan sulfate proteoglycans. It is an oncofetal antigen involved in embryonal morphogenesis^[74]. Significant expression in human adults can occur in different tissues including breast and liver and indicates ongoing pathological, mostly carcinogenic processes^[75,76]. GPC3 has been proposed as a novel serum marker for HCC^[75]. The protein promotes HCC tumor growth through stimulation of the Wnt signaling pathway^[77]. A recent meta-analysis showed an acceptable accuracy of the protein to detect HCC with a mean pooled sensitivity and specificity of 56% and 89% respectively^[78]. The protein is preferably detected using ELISA assays^[36].

Squamous cell carcinoma antigen

Squamous cell carcinoma antigen (SCCA) is a serine protease inhibitor, physiologically located in squamous epithelial cells. It is also expressed by neoplastic epithelial cells, *e.g.*, neoplastic liver cells in which it promotes tumor growth through inhibition of apoptosis^[79,80]. Increased serum levels have been detected using ELISA assays in HCC patients^[36]. The protein's diagnostic accuracy for HCC has been investigated in over 12 studies and turned out to be moderate with a pooled sensitivity and specificity of 59.0% and 76.0% respectively. Nevertheless some design limitations of these studies such as a small sample size need to be taken into account^[81].

Dickkopf-1 protein

Dickkopf-1 protein (DKK1) is a glycoprotein secreted by human hepatocytes. Upregulation of DKK1 expression takes place in a wide variety of cancers including prostate cancer, multiple myeloma and hepatocellular carcinoma^[82-84]. Overexpression of the protein is detected in tissue as well as serum from hepatocellular carcinoma patients. Although the protein is suggested to be an inhibitor of the Wnt/ β -catenin signaling pathway, its exact functions have not been fully elucidated^[82,85]. A meta-analysis showed an acceptable diagnostic accuracy of DKK1, comparable to AFP, to detect HCC with a pooled sensitivity and specificity of 65% and 94% respectively^[86-88]. Detection of the protein in serum is performed using ELISA assays^[87].

miRNA's: Promising biomarkers for HCC detection

MicroRNA's (miRNA) are small non-coding RNA's regulating gene expression by binding to messenger-

RNA (mRNA)^[89]. During recent years, circulating miRNAs have gained increasing attention for the early diagnosis and screening of hepatocellular carcinoma^[90]. So far, two miRNAs, miRNA-21 and miRNA-122 show particularly high potential in HCC diagnostics^[91]. miRNA-122 is a liver specific miRNA, whereas miRNA-21 is produced by different tissues including the colon, liver and heart in which it is involved in respectively tumor growth and cardiac disease development^[92-94]. miRNA-21 inhibits tumor suppression by inhibiting tumor suppressor pathway activating phosphatases (*e.g.*, ATK and MAPK), whereas miRNA-122 inhibits tumor growth by acting as a tumor suppressor gene^[93,95-97]. A direct correlation was observed between increasing miRNA-21 levels and increased cell proliferation^[98]. In addition, high circulating miRNA-21 levels were found to be correlated with more differentiated and progressive hepatocellular carcinoma thus indicating a bad prognosis^[94]. Serum miRNA-122 levels correlate inversely with the severity of liver fibrosis^[99]. The antitumor properties of miRNA-122 have been successfully applied in a preclinical model to prevent HCC development^[96]. The diagnostic accuracy of miRNA-21 slightly outperforms that of miRNA-122 with a pooled sensitivity and specificity of 87% and 80% respectively for miRNA-21 vs 68% and 73% for miRNA-122^[91].

Other diagnostic biomarkers

Based on systems biology approaches, more markers with diagnostic potential in HCC screening settings have recently been identified, including fucosylated fetuin A, inter-alpha-trypsin inhibitor H4, clusterin, endoglin, soluble Axl, latent TGF- β binding-protein 2 as well as peroxiredoxin 1, 2 and 3. The evidence for clinical utility of these markers remains low due to a lack of sufficiently large cohort studies^[100-108].

In addition, several studies have been published on circulating tumor cells for HCC. However, most of published studies focus on prognosis after HCC diagnosis and prediction of disease progression rather than on the diagnosis of HCC^[109-111].

PREDICTION OF THE RISK TO DEVELOP HCC

Three strategies can be applied when assessing the long-term risk for HCC. Firstly, clinical risk scores, *e.g.*, REACH-B and PAGE-B, can be calculated based on viral and host-related (*e.g.*, age and gender) clinical parameters. Most of these models have however been developed in Asian populations and lack validation in non-Asian populations^[112,113]. HBeAg positivity and HBV DNA levels above 1 million copies/mL are associated with a 4- and 11-fold increased HCC-risk during 8 and 11 years of follow-up respectively^[114-116]. Hepatitis B surface antigen (HBsAg) levels above 1000 IU/mL are accompanied with an up to 6.5-fold increased HCC risk

in men and an up to 11-fold increased risk in women within 15 years^[117]. HBsAg levels are suggested to be especially useful in case of low HBV DNA levels^[118,119].

Secondly, genome-wide association studies have enabled the linkage of genetic variants to specific disease outcomes. Single nucleotide polymorphisms (SNPs) in a wide range of genes, including the Interleukin-21 and the CRP-gene, have been associated with an increased susceptibility for HCC over a variable time course^[120-124]. Increasing evidence indicates that SNP's in the STAT4, MDM2 and HFE gene, determined on whole blood, are germline risk factors for HCC^[125,126]. On the other hand, also somatically acquired mutations, *e.g.*, in the TP53 gene, have been associated with an increased risk for HCC^[127]. All together these findings are strongly suggestive for interindividual differences in the genetic predisposition for HCC development, a predisposition that can be boosted by additional somatic mutations.

Thirdly, circulating biomolecules would be ideal as a non-invasive, predictive biomarker for HCC. An overview on the discussed predictive biomarkers including their respective increase in HCC risk is displayed in Table 2. The clinical utility of Gamma-Glutamyl Transferase (Gamma-GT) Iso-enzyme II was first evaluated as a predictive HCC marker in 1992. Patients showing persistently elevated levels of Gamma-GT Iso-enzyme II at presentation had a 86.7% risk to develop HCC within 10 years' time^[128]. Gamma-GT levels above 41 U/L and AFP-levels > 5 ng/mL have later been associated with an 8-fold increased risk for HCC in a large HBV cohort followed up for 6 years^[129]. The usefulness of AFP-levels for HCC prediction has, however, been assessed in several other studies with contradictory results^[128-130].

Cartilage oligomeric matrix protein (COMP) is an extracellular matrix protein involved in tissue genesis and remodeling^[131]. The protein is released into the circulation upon cartilage damage^[132]. Overexpression of the protein in serum from HCC patients suggested that serum COMP levels reflected an individual's fibrosis stage and subsequent risk for HCC^[133]. A recent study in Greece supports this hypothesis: COMP positivity (> 15 U/L) was associated with a 3-fold increased HCC risk during a median follow-up of 8 years^[132,134].

In a study of 27 serum cytokines and growth factors, interleukin-6 (IL-6) levels were found to predict cancer development within a timeframe of 8 to 11 years with moderate accuracy^[135]. Levels above 7 pg/mL were associated with a 3-fold increased HCC risk. As IL-6 induces C-reactive protein (CRP), the potential value of CRP in HCC risk prediction was assessed, but turned out to be disappointing^[136,137]. Recently, soluble programmed death-1 (sPD-1), a soluble form of the membrane-bound programmed death 1 on T cells with a largely unknown function was put forward as a marker^[138,139]. sPD-1 levels above 637.6 pg/mL at baseline reflected a 2-fold increased risk to develop HCC during a median follow-up time of 20 years, when

Table 2 Predictive serum biomarkers for hepatitis B virus-associated hepatocellular carcinoma

	Marker	Cut-off	Increased risk for HCC	Control group ¹	Follow-up time	Ref.
Viral	HBeAg	Positive	4-fold	HBeAg negative HBV patients	8 yr	[114]
	HBV DNA	> 1 million copies/mL	11-fold	HBV patients with HBV DNA < 300 copies/mL	11 yr	[115]
Host	HBsAg	> 1000 IU/mL	3-fold	HBV patients with HBsAg 5-9 IU/mL	14.7 yr	[117]
	Gamma-GT Iso-enzyme II	Positive	86-fold	GGT Iso-enzyme II negative HBV patients	10 yr	[128]
	Gamma-GT	> 41 U/L	8-fold	HBV patients with Gamma-GT ≥ 41 U/L	5.9 yr	[129]
	AFP	> 5 ng/mL	8-fold	HBV patients with AFP ≤ 5 ng/mL	5.9 yr	[129]
	COMP	Positive	3-fold	COMP negative HBV and HCV patients	8 yr	[134]
	IL-6	> 7 pg/mL	3-fold	HBV patients with IL-6 < 7 pg/mL	7.25 yr	[135]
	sPD-1	> 637.6 pg/mL	2-fold	HBV patients with sPD-1 < 117.3 pg/mL	20 yr	[138]

¹Control group: group included in the study, to which the increased HCC risk was calculated. HBeAg: Hepatitis B e antigen; HBV: Hepatitis B virus; HBsAg: Hepatitis B surface antigen; Gamma-GT: Gamma glutamyltransferase; AFP: Alpha-fetoprotein; COMP: Cartilage oligomeric matrix protein; IL-6: Interleukin 6; sPD-1: Soluble programmed death 1.

compared to sPD-1 levels below 117.3 pg/mL^[138].

DISCUSSION

Despite its disappointing sensitivity and specificity, AFP still remains the most widely used serum HCC biomarker. Some newly discovered circulating biomarkers, *e.g.*, AFP-I3, DCP and microRNA's show promising potential for implementation in clinical practice. However, only GP73 strongly outperforms AFP in terms of diagnostic accuracy. Large variations in sensitivity and specificity are noticed between different studies assessing the same biomarker (Table 1).

Due to the heterogeneity of HCC, one single biomarker with 100% sensitivity and specificity in all HCC cases will be hard to find. A more rational approach to increase the diagnostic accuracy might be the combination of different biomarkers^[41,52,55,62,65,72,140-142]. The most recent APASL guidelines indeed recommend the combined use of AFP, AFP-I3 and DCP in HCC screening^[22,142]. In favor of this approach, a meta-analysis showed that combined testing of GP73 and AFP increased the pooled sensitivity without decreasing the specificity to detect hepatocellular carcinoma. The pooled sensitivity and specificity were 87% and 85% respectively when biomarkers were combined, compared to 77% and 91% for GP73 and 62% and 84% for AFP when used alone^[72].

All currently identified circulating biomarkers and their combinations definitely need more validation studies. Most of the biomarker discovery studies have been performed in cohorts of a few 100 patients. The majority of identified markers has so far not been subject of large, external validation studies^[100-103,108]. Five subsequent steps are to be followed in cancer biomarker discovery. The first step is the implementation of preclinical exploratory studies. Step 2 is the development of a clinical assay. Step 3 involves retrospective studies, step 4 prospective studies and step 5 large, randomized controlled trials. Biomarkers identified in step 1 must pass all other steps before

they can be termed validated biomarkers^[143]. So far, only AFP has reached step 5^[15].

In addition, the studies that have been conducted over the last decades are hampered by limitations in their study design. As an example the patient cohorts for HCC biomarker discovery studies are often heterogeneous regarding liver disease etiology and ethnicity. In their paper, da Costa *et al.*^[144] proved the need to validate biomarkers in different ethnic populations. They investigated the potential of osteopontin and latent Transforming Growth Factor beta binding-protein in HCC diagnosis in separate cohorts in Gambia, Korea, Thailand and France. The sensitivity and specificity of both markers differed (> 10%) among ethnicities. The onset of HCC occurs at a median age of 45 in sub-Saharan African people, whereas a mean age of 52 to 65 has been observed in the rest of the world^[145,146]. In addition, HCC incidence varies among HBV and hepatitis C virus (HCV) patients, highlighting the importance of homogeneous patient cohorts. Future biomarker discovery and validation studies should therefore distinguish between different ethnicities and etiologies as this most probably explains the variation in sensitivities and specificities noticed between studies assessing the same biomarker (Table 1).

The inclusion of clinical parameters into biomarker scores could increase their performance. One study demonstrated that incorporation of age into combined models of biomarker testing significantly improved the diagnostic performance for HCC^[140].

From a clinical point of view, however, predictive serum biomarkers would be preferred over diagnostic biomarkers to tailor HCC surveillance according to the individual needs. Proteomic approaches are encouraging, but also need a validation in larger cohorts^[147,148]. Expression of Heat-shock Protein 27 was *e.g.*, detected in 90% of sera from HCC patients and in 0% of sera from non-HCC patients, which seems promising^[148]. Other groups have focused on genomics and have identified a gene signature in

liver tissue of HCV infected patients predictive of HCC development^[149,150]. It could be of interest to identify corresponding secretory biomarkers in blood.

CONCLUSION

Monitoring HCC development in chronic hepatitis B patients based on serum biomarkers remains challenging. During recent years, new predictive and diagnostic circulating biomarkers have been proposed. Combinations of these biomarkers show a higher potential for implementation in clinical practice, but large validation studies in homogeneous ethnic and etiological populations are urgently needed to unequivocally demonstrate their clinical utility.

ACKNOWLEDGMENTS

The authors thank professor Benedicte Y De Winter for the critical revision and language editing of the manuscript.

REFERENCES

- 1 Siegel RL, Miller KD, Jemal A. Cancer statistics, 2016. *CA Cancer J Clin* 2016; **66**: 7-30 [PMID: 26742998 DOI: 10.3322/caac.21332]
- 2 Dienstag JL. Hepatitis B virus infection. *N Engl J Med* 2008; **359**: 1486-1500 [PMID: 18832247 DOI: 10.1056/NEJMra0801644]
- 3 El-Serag HB. Epidemiology of viral hepatitis and hepatocellular carcinoma. *Gastroenterology* 2012; **142**: 1264-1273.e1 [PMID: 22537432 DOI: 10.1053/j.gastro.2011.12.061]
- 4 Kew MC. Epidemiology of chronic hepatitis B virus infection, hepatocellular carcinoma, and hepatitis B virus-induced hepatocellular carcinoma. *Pathol Biol (Paris)* 2010; **58**: 273-277 [PMID: 20378277 DOI: 10.1016/j.patbio.2010.01.005]
- 5 El-Serag HB. Hepatocellular carcinoma. *N Engl J Med* 2011; **365**: 1118-1127 [PMID: 21992124 DOI: 10.1056/NEJMr1001683]
- 6 Higgs MR, Chouteau P, Lerat H. 'Liver let die': oxidative DNA damage and hepatotropic viruses. *J Gen Virol* 2014; **95**: 991-1004 [PMID: 24496828 DOI: 10.1099/vir.0.059485-0]
- 7 Sukowati CH, El-Khobar KE, Ie SI, Anfuso B, Muljono DH, Tiribelli C. Significance of hepatitis virus infection in the oncogenic initiation of hepatocellular carcinoma. *World J Gastroenterol* 2016; **22**: 1497-1512 [PMID: 26819517 DOI: 10.3748/wjg.v22.i4.1497]
- 8 Bonilla Guerrero R, Roberts LR. The role of hepatitis B virus integrations in the pathogenesis of human hepatocellular carcinoma. *J Hepatol* 2005; **42**: 760-777 [PMID: 15826727 DOI: 10.1016/j.jhep.2005.02.005]
- 9 Michielsen P, Ho E. Viral hepatitis B and hepatocellular carcinoma. *Acta Gastroenterol Belg* 2011; **74**: 4-8 [PMID: 21563647]
- 10 Vanwolleghem T, Hou J, van Oord G, Andeweg AC, Osterhaus AD, Pas SD, Janssen HL, Boonstra A. Re-evaluation of hepatitis B virus clinical phases by systems biology identifies unappreciated roles for the innate immune response and B cells. *Hepatology* 2015; **62**: 87-100 [PMID: 25808668 DOI: 10.1002/hep.27805]
- 11 Lok AS. Hepatitis B: liver fibrosis and hepatocellular carcinoma. *Gastroenterol Clin Biol* 2009; **33**: 911-915 [PMID: 19577871 DOI: 10.1016/j.gcb.2009.06.001]
- 12 Vlachogiannakos J, Papatheodoridis GV. HBV: Do I treat my immunotolerant patients? *Liver Int* 2016; **36** Suppl 1: 93-99 [PMID: 26725904 DOI: 10.1111/liv.12996]
- 13 Di Bisceglie AM. Hepatitis B and hepatocellular carcinoma. *Hepatology* 2009; **49**: S56-S60 [PMID: 19399807 DOI: 10.1002/hep.22962]
- 14 McMahon BJ, Bulkow L, Harpster A, Snowball M, Lanier A, Sacco F, Dunaway E, Williams J. Screening for hepatocellular carcinoma in Alaska natives infected with chronic hepatitis B: a 16-year population-based study. *Hepatology* 2000; **32**: 842-846 [PMID: 11003632 DOI: 10.1053/jhep.2000.17914]
- 15 Zhang BH, Yang BH, Tang ZY. Randomized controlled trial of screening for hepatocellular carcinoma. *J Cancer Res Clin Oncol* 2004; **130**: 417-422 [PMID: 15042359 DOI: 10.1007/s00432-004-0552-0]
- 16 Miller ZA, Lee KS. Screening for hepatocellular carcinoma in high-risk populations. *Clin Imaging* 2015; **40**: 311-314 [PMID: 26898986 DOI: 10.1016/j.clinimag.2015.11.010]
- 17 Yuen MF, Cheng CC, Laufer JJ, Lam SK, Ooi CG, Lai CL. Early detection of hepatocellular carcinoma increases the chance of treatment: Hong Kong experience. *Hepatology* 2000; **31**: 330-335 [PMID: 10655254 DOI: 10.1002/hep.510310211]
- 18 van Meer S, de Man RA, Coenraad MJ, Sprengers D, van Nieuwkerk KM, Klumpen HJ, Jansen PL, IJzermans JN, van Oijen MG, Siersema PD, van Erpecum KJ. Surveillance for hepatocellular carcinoma is associated with increased survival: Results from a large cohort in the Netherlands. *J Hepatol* 2015; **63**: 1156-1163 [PMID: 26100498 DOI: 10.1016/j.jhep.2015.06.012]
- 19 Stravitz RT, Heuman DM, Chand N, Sterling RK, Shiffman ML, Luketic VA, Sanyal AJ, Habib A, Mihas AA, Giles HC, Maluf DG, Cotterell AH, Posner MP, Fisher RA. Surveillance for hepatocellular carcinoma in patients with cirrhosis improves outcome. *Am J Med* 2008; **121**: 119-126 [PMID: 18261500 DOI: 10.1016/j.amjmed.2007.09.020]
- 20 Velázquez RF, Rodríguez M, Navascués CA, Linares A, Pérez R, Sotorrios NG, Martínez I, Rodrigo L. Prospective analysis of risk factors for hepatocellular carcinoma in patients with liver cirrhosis. *Hepatology* 2003; **37**: 520-527 [PMID: 12601348 DOI: 10.1053/jhep.2003.50093]
- 21 European Association For The Study Of The Liver; European Organisation For Research And Treatment Of Cancer. EASL-EORTC clinical practice guidelines: management of hepatocellular carcinoma. *J Hepatol* 2012; **56**: 908-943 [PMID: 22424438 DOI: 10.1016/j.jhep.2011.12.001]
- 22 Omata M, Lesmana LA, Tateishi R, Chen PJ, Lin SM, Yoshida H, Kudo M, Lee JM, Choi BI, Poon RT, Shiina S, Cheng AL, Jia JD, Obi S, Han KH, Jafri W, Chow P, Lim SG, Chawla YK, Budihusodo U, Gani RA, Lesmana CR, Putranto TA, Liaw YF, Sarin SK. Asian Pacific Association for the Study of the Liver consensus recommendations on hepatocellular carcinoma. *Hepatol Int* 2010; **4**: 439-474 [PMID: 20827404 DOI: 10.1007/s12072-010-9165-7]
- 23 Bruix J, Sherman M. Management of hepatocellular carcinoma: an update. *Hepatology* 2011; **53**: 1020-1022 [PMID: 21374666 DOI: 10.1002/hep.24199]
- 24 Kokudo N, Hasegawa K, Akahane M, Igaki H, Izumi N, Ichida T, Uemoto S, Kaneko S, Kawasaki S, Ku Y, Kudo M, Kubo S, Takayama T, Tateishi R, Fukuda T, Matsui O, Matsuyama Y, Murakami T, Arii S, Okazaki M, Makuuchi M. Evidence-based Clinical Practice Guidelines for Hepatocellular Carcinoma: The Japan Society of Hepatology 2013 update (3rd JSH-HCC Guidelines). *Hepatol Res* 2015; **45** [PMID: 25625806 DOI: 10.1111/hepr.12464]
- 25 Makuuchi M, Kokudo N, Arii S, Futagawa S, Kaneko S, Kawasaki S, Matsuyama Y, Okazaki M, Okita K, Omata M, Saida Y, Takayama T, Yamaoka Y. Development of evidence-based clinical guidelines for the diagnosis and treatment of hepatocellular carcinoma in Japan. *Hepatol Res* 2008; **38**: 37-51 [PMID: 18039202 DOI: 10.1111/j.1872-034X.2007.00216.x]
- 26 Vlaanderen J, Moore LE, Smith MT, Lan Q, Zhang L, Skibola CF, Rothman N, Vermeulen R. Application of OMICS technologies in occupational and environmental health research; current status and projections. *Occup Environ Med* 2010; **67**: 136-143 [PMID: 19933307 DOI: 10.1136/oem.2008.042788]
- 27 Pesce F, Pathan S, Schena FP. From -omics to personalized

- medicine in nephrology: integration is the key. *Nephrol Dial Transplant* 2013; **28**: 24-28 [PMID: 23229923 DOI: 10.1093/ndt/gfs483]
- 28 **Seregni E**, Botti C, Bombardieri E. Biochemical characteristics and clinical applications of alpha-fetoprotein isoforms. *Anticancer Res* 1995; **15**: 1491-1499 [PMID: 7544570]
- 29 **Gabant P**, Forrester L, Nichols J, Van Reeth T, De Mees C, Pajack B, Watt A, Smitz J, Alexandre H, Szpirer C, Szpirer J. Alpha-fetoprotein, the major fetal serum protein, is not essential for embryonic development but is required for female fertility. *Proc Natl Acad Sci USA* 2002; **99**: 12865-12870 [PMID: 12297623 DOI: 10.1073/pnas.202215399]
- 30 **Seo SI**, Kim SS, Choi BY, Lee SH, Kim SJ, Park HW, Kim HS, Shin WG, Kim KH, Lee JH, Kim HY, Jang MK. Clinical significance of elevated serum alpha-fetoprotein (AFP) level in acute viral hepatitis A (AHA). *Hepatology* 2013; **60**: 1592-1596 [PMID: 24634927]
- 31 **Guo YL**, Zhang YL, Zhu JQ. Primary yolk sac tumor of the retroperitoneum: A case report and review of the literature. *Oncol Lett* 2014; **8**: 556-560 [PMID: 25009643 DOI: 10.3892/ol.2014.2162]
- 32 **Yao M**, Zhao J, Lu F. Alpha-fetoprotein still is a valuable diagnostic and prognosis predicting biomarker in hepatitis B virus infection-related hepatocellular carcinoma. *Oncotarget* 2016; **7**: 3702-3708 [PMID: 26784252 DOI: 10.18632/oncotarget.6913]
- 33 **Bergstrand CG**, Czar B. Demonstration of a new protein fraction in serum from the human fetus. *Scand J Clin Lab Invest* 1956; **8**: 174 [PMID: 13351554 DOI: 10.3109/00365515609049266]
- 34 **Tonami N**, Aburano T, Hisada K. Comparison of alpha1 fetoprotein radioimmunoassay method and liver scanning for detecting primary hepatic cell carcinoma. *Cancer* 1975; **36**: 466-470 [PMID: 50872]
- 35 **Akhmeteli MA**, Linnik AB, Cernov KS. Hepatocarcinogenesis and the appearance of serum alpha-fetoprotein in mice treated with extracts of barley grain infected with *Fusarium sporotrichioides*. *Bull World Health Organ* 1972; **47**: 663-664 [PMID: 4121669]
- 36 **Waidely E**, Al-Yuobi AR, Bashammakh AS, El-Shahawi MS, Leblanc RM. Serum protein biomarkers relevant to hepatocellular carcinoma and their detection. *Analyst* 2016; **141**: 36-44 [PMID: 26606739 DOI: 10.1039/c5an01884f]
- 37 **Lok AS**, Lai CL. alpha-Fetoprotein monitoring in Chinese patients with chronic hepatitis B virus infection: role in the early detection of hepatocellular carcinoma. *Hepatology* 1989; **9**: 110-115 [PMID: 2461890]
- 38 **Maringhini A**, Cottone M, Sciarrino E, Marcenó MP, La Seta F, Fusco G, Rinaldi F, Pagliaro L. Ultrasonography and alpha-fetoprotein in diagnosis of hepatocellular carcinoma in cirrhosis. *Dig Dis Sci* 1988; **33**: 47-51 [PMID: 2448095]
- 39 **He X**, Wang Y, Zhang W, Li H, Luo R, Zhou Y, Liao CL, Huang H, Lv X, Xie Z, He M. Screening differential expression of serum proteins in AFP-negative HBV-related hepatocellular carcinoma using iTRAQ-MALDI-MS/MS. *Neoplasma* 2014; **61**: 17-26 [PMID: 24195504]
- 40 **Sato Y**, Nakata K, Kato Y, Shima M, Ishii N, Koji T, Taketa K, Endo Y, Nagataki S. Early recognition of hepatocellular carcinoma based on altered profiles of alpha-fetoprotein. *N Engl J Med* 1993; **328**: 1802-1806 [PMID: 7684823 DOI: 10.1056/nejm199306243282502]
- 41 **Song P**, Feng X, Inagaki Y, Song T, Zhang K, Wang Z, Zheng S, Ma K, Li Q, Kong D, Wu Q, Zhang T, Zhao X, Hasegawa K, Sugawara Y, Kokudo N, Tang W. Clinical utility of simultaneous measurement of alpha-fetoprotein and des-gamma-carboxy prothrombin for diagnosis of patients with hepatocellular carcinoma in China: A multi-center case-controlled study of 1,153 subjects. *Biosci Trends* 2014; **8**: 266-273 [PMID: 25382443]
- 42 **Kim GA**, Seock CH, Park JW, An J, Lee KS, Yang JE, Lim YS, Kim KM, Shim JH, Lee D, Lee HC. Reappraisal of serum alpha-fetoprotein as a surveillance test for hepatocellular carcinoma during entecavir treatment. *Liver Int* 2015; **35**: 232-239 [PMID: 24576055 DOI: 10.1111/liv.12516]
- 43 **Hosaka T**, Suzuki F, Kobayashi M, Seko Y, Kawamura Y, Sezaki H, Akuta N, Suzuki Y, Saitoh S, Arase Y, Ikeda K, Kobayashi M, Kumada H. Long-term entecavir treatment reduces hepatocellular carcinoma incidence in patients with hepatitis B virus infection. *Hepatology* 2013; **58**: 98-107 [PMID: 23213040 DOI: 10.1002/hep.26180]
- 44 **Sanai FM**, Sobki S, Bzeizi KI, Shaikh SA, Alswat K, Al-Hamoudi W, Almadi M, Al Saif F, Abdo AA. Assessment of alpha-fetoprotein in the diagnosis of hepatocellular carcinoma in Middle Eastern patients. *Dig Dis Sci* 2010; **55**: 3568-3575 [PMID: 20397051 DOI: 10.1007/s10620-010-1201-x]
- 45 **Sherman M**, Peltekian KM, Lee C. Screening for hepatocellular carcinoma in chronic carriers of hepatitis B virus: incidence and prevalence of hepatocellular carcinoma in a North American urban population. *Hepatology* 1995; **22**: 432-438 [PMID: 7543434]
- 46 **Li D**, Mallory T, Satomura S. AFP-L3: a new generation of tumor marker for hepatocellular carcinoma. *Clin Chim Acta* 2001; **313**: 15-19 [PMID: 11694234]
- 47 **Leerapun A**, Suravarapu SV, Bida JP, Clark RJ, Sanders EL, Mettler TA, Stadheim LM, Aderca I, Moser CD, Nagorney DM, LaRusso NF, de Groen PC, Menon KV, Lazaridis KN, Gores GJ, Charlton MR, Roberts RO, Therneau TM, Katzmann JA, Roberts LR. The utility of Lens culinaris agglutinin-reactive alpha-fetoprotein in the diagnosis of hepatocellular carcinoma: evaluation in a United States referral population. *Clin Gastroenterol Hepatol* 2007; **5**: 394-402; quiz 267 [PMID: 17368240 DOI: 10.1016/j.cgh.2006.12.005]
- 48 **Kusaba T**. Relationship between Lens culinaris agglutinin reactive alpha-fetoprotein and biological features of hepatocellular carcinoma. *Kurume Med J* 1998; **45**: 113-120 [PMID: 9658760]
- 49 **Toyoda H**, Kumada T, Tada T, Kaneoka Y, Maeda A, Kanke F, Satomura S. Clinical utility of highly sensitive Lens culinaris agglutinin-reactive alpha-fetoprotein in hepatocellular carcinoma patients with alpha-fetoprotein ≤ 20 ng/mL. *Cancer Sci* 2011; **102**: 1025-1031 [PMID: 21244578 DOI: 10.1111/j.1349-7006.2011.01875.x]
- 50 **Yamamoto K**, Imamura H, Matsuyama Y, Kume Y, Ikeda H, Norman GL, Shums Z, Aoki T, Hasegawa K, Beck Y, Sugawara Y, Kokudo N. AFP, AFP-L3, DCP, and GP73 as markers for monitoring treatment response and recurrence and as surrogate markers of clinicopathological variables of HCC. *J Gastroenterol* 2010; **45**: 1272-1282 [PMID: 20625772 DOI: 10.1007/s00535-010-0278-5]
- 51 **Tanwandee T**, Setthasin S, Charatcharoenwitthaya P, Chainuvati S, Leelakusolvong S, Pausawasdi N, Srikureja W, Pongprasobchai S, Manatsathit S, Kachintorn U, Ekpo P, Senawong S. Clinical utility of lens culinaris agglutinin-reactive alpha-fetoprotein in the diagnosis of hepatocellular carcinoma: evaluation in a Thai referral population. *J Med Assoc Thai* 2009; **92** Suppl 2: S49-S56 [PMID: 19562986]
- 52 **Hu B**, Tian X, Sun J, Meng X. Evaluation of individual and combined applications of serum biomarkers for diagnosis of hepatocellular carcinoma: a meta-analysis. *Int J Mol Sci* 2013; **14**: 23559-23580 [PMID: 24317431 DOI: 10.3390/ijms141223559]
- 53 **Marrero JA**, Feng Z, Wang Y, Nguyen MH, Befeler AS, Roberts LR, Reddy KR, Harnois D, Llovet JM, Normolle D, Dalhgren J, Chia D, Lok AS, Wagner PD, Srivastava S, Schwartz M. Alpha-fetoprotein, des-gamma-carboxyprothrombin, and lectin-bound alpha-fetoprotein in early hepatocellular carcinoma. *Gastroenterology* 2009; **137**: 110-118 [PMID: 19362088 DOI: 10.1053/j.gastro.2009.04.005]
- 54 **Oka H**, Saito A, Ito K, Kumada T, Satomura S, Kasugai H, Osaki Y, Seki T, Kudo M, Tanaka M. Multicenter prospective analysis of newly diagnosed hepatocellular carcinoma with respect to the percentage of Lens culinaris agglutinin-reactive alpha-fetoprotein. *J Gastroenterol Hepatol* 2001; **16**: 1378-1383 [PMID: 11851836]
- 55 **Xu WJ**, Guo BL, Han YG, Shi L, Ma WS. Diagnostic value of alpha-fetoprotein-L3 and Golgi protein 73 in hepatocellular carcinomas with low AFP levels. *Tumour Biol* 2014; **35**: 12069-12074 [PMID: 25209179 DOI: 10.1007/

- s13277-014-2506-8]
- 56 **Lefrere JJ**, Gozin D. Use of des-gamma-carboxyprothrombin in retrospective diagnosis of hidden intoxication of anticoagulants. *J Clin Pathol* 1987; **40**: 589 [PMID: 3584512]
 - 57 **Fujiyama S**, Morishita T, Hashiguchi O, Sato T. Plasma abnormal prothrombin (des-gamma-carboxy prothrombin) as a marker of hepatocellular carcinoma. *Cancer* 1988; **61**: 1621-1628 [PMID: 2450634]
 - 58 **Liebman HA**, Furie BC, Tong MJ, Blanchard RA, Lo KJ, Lee SD, Coleman MS, Furie B. Des-gamma-carboxy (abnormal) prothrombin as a serum marker of primary hepatocellular carcinoma. *N Engl J Med* 1984; **310**: 1427-1431 [PMID: 6201741 DOI: 10.1056/nejm198405313102204]
 - 59 **Yu R**, Ding S, Tan W, Tan S, Tan Z, Xiang S, Zhou Y, Mao Q, Deng G. Performance of Protein Induced by Vitamin K Absence or Antagonist-II (PIVKA-II) for Hepatocellular Carcinoma Screening in Chinese Population. *Hepat Mon* 2015; **15**: e28806 [PMID: 26300931 DOI: 10.5812/hepatmon.28806v2]
 - 60 **Huang TS**, Shyu YC, Turner R, Chen HY, Chen PJ. Diagnostic performance of alpha-fetoprotein, lens culinaris agglutinin-reactive alpha-fetoprotein, des-gamma carboxyprothrombin, and glypican-3 for the detection of hepatocellular carcinoma: a systematic review and meta-analysis protocol. *Syst Rev* 2013; **2**: 37 [PMID: 23738605 DOI: 10.1186/2046-4053-2-37]
 - 61 **Poté N**, Cauchy F, Albuquerque M, Voitot H, Belghiti J, Castera L, Puy H, Bedossa P, Paradis V. Performance of PIVKA-II for early hepatocellular carcinoma diagnosis and prediction of microvascular invasion. *J Hepatol* 2015; **62**: 848-854 [PMID: 25450201 DOI: 10.1016/j.jhep.2014.11.005]
 - 62 **Ertle JM**, Heider D, Wichert M, Keller B, Kueper R, Hilgard P, Gerken G, Schlaak JF. A combination of α -fetoprotein and des- γ -carboxy prothrombin is superior in detection of hepatocellular carcinoma. *Digestion* 2013; **87**: 121-131 [PMID: 23406785 DOI: 10.1159/000346080]
 - 63 **Zhu R**, Yang J, Xu L, Dai W, Wang F, Shen M, Zhang Y, Zhang H, Chen K, Cheng P, Wang C, Zheng Y, Li J, Lu J, Zhou Y, Wu D, Guo C. Diagnostic Performance of Des- γ -carboxy Prothrombin for Hepatocellular Carcinoma: A Meta-Analysis. *Gastroenterol Res Pract* 2014; **2014**: 529314 [PMID: 25165471 DOI: 10.1155/2014/529314]
 - 64 **Sodek J**, Ganss B, McKee MD. Osteopontin. *Crit Rev Oral Biol Med* 2000; **11**: 279-303 [PMID: 11021631]
 - 65 **Wan HG**, Xu H, Gu YM, Wang H, Xu W, Zu MH. Comparison osteopontin vs AFP for the diagnosis of HCC: a meta-analysis. *Clin Res Hepatol Gastroenterol* 2014; **38**: 706-714 [PMID: 25034355 DOI: 10.1016/j.clinre.2014.06.008]
 - 66 **Khalil A**, Elgedawy J, Faramawi MF, Elfert A, Salama I, Abbass A, Elsaid H, Elsebaai H. Plasma osteopontin level as a diagnostic marker of hepatocellular carcinoma in patients with radiological evidence of focal hepatic lesions. *Tumori* 2013; **99**: 100-107 [PMID: 23549008 DOI: 10.1700/1248.13796]
 - 67 **Ba MC**, Long H, Tang YQ, Cui SZ. GP73 expression and its significance in the diagnosis of hepatocellular carcinoma: a review. *Int J Clin Exp Pathol* 2012; **5**: 874-881 [PMID: 23119104]
 - 68 **Kladney RD**, Bulla GA, Guo L, Mason AL, Tollefson AE, Simon DJ, Koutoubi Z, Fimmel CJ. GP73, a novel Golgi-localized protein upregulated by viral infection. *Gene* 2000; **249**: 53-65 [PMID: 10831838]
 - 69 **Kladney RD**, Cui X, Bulla GA, Brunt EM, Fimmel CJ. Expression of GP73, a resident Golgi membrane protein, in viral and nonviral liver disease. *Hepatology* 2002; **35**: 1431-1440 [PMID: 12029628 DOI: 10.1053/jhep.2002.32525]
 - 70 **Marrero JA**, Romano PR, Nikolaeva O, Steel L, Mehta A, Fimmel CJ, Comunale MA, D'Amelio A, Lok AS, Block TM. GP73, a resident Golgi glycoprotein, is a novel serum marker for hepatocellular carcinoma. *J Hepatol* 2005; **43**: 1007-1012 [PMID: 16137783 DOI: 10.1016/j.jhep.2005.05.028]
 - 71 **Xu Z**, Liu L, Pan X, Wei K, Wei M, Liu L, Yang H, Liu Q. Serum Golgi protein 73 (GP73) is a diagnostic and prognostic marker of chronic HBV liver disease. *Medicine* (Baltimore) 2015; **94**: e659 [PMID: 25816035 DOI: 10.1097/md.0000000000000659]
 - 72 **Dai M**, Chen X, Liu X, Peng Z, Meng J, Dai S. Diagnostic Value of the Combination of Golgi Protein 73 and Alpha-Fetoprotein in Hepatocellular Carcinoma: A Meta-Analysis. *PLoS One* 2015; **10**: e0140067 [PMID: 26441340 DOI: 10.1371/journal.pone.0140067]
 - 73 **Witjes CD**, van Aalten SM, Steyerberg EW, Borsboom GJ, de Man RA, Verhoef C, Ijzermans JN. Recently introduced biomarkers for screening of hepatocellular carcinoma: a systematic review and meta-analysis. *Hepatol Int* 2013; **7**: 59-64 [PMID: 23519638 DOI: 10.1007/s12072-012-9374-3]
 - 74 **Jakubovic BD**, Jothy S. Glypican-3: from the mutations of Simpson-Golabi-Behmel genetic syndrome to a tumor marker for hepatocellular carcinoma. *Exp Mol Pathol* 2007; **82**: 184-189 [PMID: 17258707 DOI: 10.1016/j.yexmp.2006.10.010]
 - 75 **Capurro M**, Wanless IR, Sherman M, Deboer G, Shi W, Miyoshi E, Filmus J. Glypican-3: a novel serum and histochemical marker for hepatocellular carcinoma. *Gastroenterology* 2003; **125**: 89-97 [PMID: 12851874]
 - 76 **Peters MG**, Farias E, Colombo L, Filmus J, Puricelli L, Bal de Kier Joffé E. Inhibition of invasion and metastasis by glypican-3 in a syngeneic breast cancer model. *Breast Cancer Res Treat* 2003; **80**: 221-232 [PMID: 12908826 DOI: 10.1023/a:1024549729256]
 - 77 **Capurro MI**, Xiang YY, Lobe C, Filmus J. Glypican-3 promotes the growth of hepatocellular carcinoma by stimulating canonical Wnt signaling. *Cancer Res* 2005; **65**: 6245-6254 [PMID: 16024626 DOI: 10.1158/0008-5472.can-04-4244]
 - 78 **Liu JW**, Zuo XL, Wang S. Diagnosis accuracy of serum Glypican-3 level in patients with hepatocellular carcinoma and liver cirrhosis: a meta-analysis. *Eur Rev Med Pharmacol Sci* 2015; **19**: 3655-3673 [PMID: 26502856]
 - 79 **Pontisso P**, Calabrese F, Benvegnù L, Lise M, Belluco C, Ruvolletto MG, Marino M, Valente M, Nitti D, Gatta A, Fassina G. Overexpression of squamous cell carcinoma antigen variants in hepatocellular carcinoma. *Br J Cancer* 2004; **90**: 833-837 [PMID: 14970861 DOI: 10.1038/sj.bjc.6601543]
 - 80 **Suminami Y**, Nagashima S, Murakami A, Nawata S, Gondo T, Hirakawa H, Numa F, Silverman GA, Kato H. Suppression of a squamous cell carcinoma (SCC)-related serpin, SCC antigen, inhibits tumor growth with increased intratumor infiltration of natural killer cells. *Cancer Res* 2001; **61**: 1776-1780 [PMID: 11280721]
 - 81 **Zhang J**, Shao C, Zhou Q, Zhu Y, Zhu J, Tu C. Diagnostic accuracy of serum squamous cell carcinoma antigen and squamous cell carcinoma antigen-immunoglobulin M for hepatocellular carcinoma: A meta-analysis. *Mol Clin Oncol* 2015; **3**: 1165-1171 [PMID: 26623071 DOI: 10.3892/mco.2015.600]
 - 82 **Fatima S**, Lee NP, Luk JM. Dickkopf-1 and Wnt/ β -catenin signalling in liver cancer. *World J Clin Oncol* 2011; **2**: 311-325 [PMID: 21876852 DOI: 10.5306/wjco.v2.i8.311]
 - 83 **Rachner TD**, Thiele S, Göbel A, Browne A, Fuessel S, Erdmann K, Wirth MP, Fröhner M, Todenhöfer T, Muders MH, Kieslinger M, Rauner M, Hofbauer LC. High serum levels of Dickkopf-1 are associated with a poor prognosis in prostate cancer patients. *BMC Cancer* 2014; **14**: 649 [PMID: 25182503 DOI: 10.1186/1471-2407-14-649]
 - 84 **Gavriatopoulou M**, Dimopoulos MA, Christoulas D, Migkou M, Iakovaki M, Gkatzamanidou M, Terpos E. Dickkopf-1: a suitable target for the management of myeloma bone disease. *Expert Opin Ther Targets* 2009; **13**: 839-848 [PMID: 19530987 DOI: 10.1517/14728220903025770]
 - 85 **Wang J**, Shou J, Chen X. Dickkopf-1, an inhibitor of the Wnt signaling pathway, is induced by p53. *Oncogene* 2000; **19**: 1843-1848 [PMID: 10777218 DOI: 10.1038/sj.onc.1203503]
 - 86 **Zhang J**, Zhao Y, Yang Q. Sensitivity and specificity of Dickkopf-1 protein in serum for diagnosing hepatocellular carcinoma: a meta-analysis. *Int J Biol Markers* 2014; **29**: e403-e410 [PMID: 24980448 DOI: 10.5301/ijbm.5000101]
 - 87 **Kim SU**, Park JH, Kim HS, Lee JM, Lee HG, Kim H, Choi SH, Baek S, Kim BK, Park JY, Kim do Y, Ahn SH, Lee JD, Han KH. Serum Dickkopf-1 as a Biomarker for the Diagnosis of

- Hepatocellular Carcinoma. *Yonsei Med J* 2015; **56**: 1296-1306 [PMID: 26256972 DOI: 10.3349/ymj.2015.56.5.1296]
- 88 **Shen Q**, Fan J, Yang XR, Tan Y, Zhao W, Xu Y, Wang N, Niu Y, Wu Z, Zhou J, Qiu SJ, Shi YH, Yu B, Tang N, Chu W, Wang M, Wu J, Zhang Z, Yang S, Gu J, Wang H, Qin W. Serum DKK1 as a protein biomarker for the diagnosis of hepatocellular carcinoma: a large-scale, multicentre study. *Lancet Oncol* 2012; **13**: 817-826 [PMID: 22738799 DOI: 10.1016/s1470-2045(12)70233-4]
- 89 **Bartel DP**. MicroRNAs: target recognition and regulatory functions. *Cell* 2009; **136**: 215-233 [PMID: 19167326 DOI: 10.1016/j.cell.2009.01.002]
- 90 **Hyun KA**, Kim J, Gwak H, Jung HI. Isolation and enrichment of circulating biomarkers for cancer screening, detection, and diagnostics. *Analyst* 2016; **141**: 382-392 [PMID: 26588824 DOI: 10.1039/c5an01762a]
- 91 **Huang JT**, Liu SM, Ma H, Yang Y, Zhang X, Sun H, Zhang X, Xu J, Wang J. Systematic Review and Meta-Analysis: Circulating miRNAs for Diagnosis of Hepatocellular Carcinoma. *J Cell Physiol* 2016; **231**: 328-335 [PMID: 26291451 DOI: 10.1002/jcp.25135]
- 92 **Tu Y**, Wan L, Fan Y, Wang K, Bu L, Huang T, Cheng Z, Shen B. Ischemic postconditioning-mediated miRNA-21 protects against cardiac ischemia/reperfusion injury via PTEN/Akt pathway. *PLoS One* 2013; **8**: e75872 [PMID: 24098402 DOI: 10.1371/journal.pone.0075872]
- 93 **Asangani IA**, Rasheed SA, Nikolova DA, Leupold JH, Colburn NH, Post S, Allgayer H. MicroRNA-21 (miR-21) post-transcriptionally downregulates tumor suppressor Pcdcd4 and stimulates invasion, intravasation and metastasis in colorectal cancer. *Oncogene* 2008; **27**: 2128-2136 [PMID: 17968323 DOI: 10.1038/sj.onc.1210856]
- 94 **Huang CS**, Yu W, Cui H, Wang YJ, Zhang L, Han F, Huang T. Increased expression of miR-21 predicts poor prognosis in patients with hepatocellular carcinoma. *Int J Clin Exp Pathol* 2015; **8**: 7234-7238 [PMID: 26261620]
- 95 **Liu C**, Yu J, Yu S, Lavker RM, Cai L, Liu W, Yang K, He X, Chen S. MicroRNA-21 acts as an oncomir through multiple targets in human hepatocellular carcinoma. *J Hepatol* 2010; **53**: 98-107 [PMID: 20447717 DOI: 10.1016/j.jhep.2010.02.021]
- 96 **Nakao K**, Miyaaki H, Ichikawa T. Antitumor function of microRNA-122 against hepatocellular carcinoma. *J Gastroenterol* 2014; **49**: 589-593 [PMID: 24531873 DOI: 10.1007/s00535-014-0932-4]
- 97 **Wu K**, Li L, Li S. Circulating microRNA-21 as a biomarker for the detection of various carcinomas: an updated meta-analysis based on 36 studies. *Tumour Biol* 2015; **36**: 1973-1981 [PMID: 25527152 DOI: 10.1007/s13277-014-2803-2]
- 98 **Si ML**, Zhu S, Wu H, Lu Z, Wu F, Mo YY. miR-21-mediated tumor growth. *Oncogene* 2007; **26**: 2799-2803 [PMID: 17072344 DOI: 10.1038/sj.onc.1210083]
- 99 **Arataki K**, Hayes CN, Akamatsu S, Akiyama R, Abe H, Tsuge M, Miki D, Ochi H, Hiraga N, Imamura M, Takahashi S, Aikata H, Kawaoka T, Kawakami H, Ohishi W, Chayama K. Circulating microRNA-22 correlates with microRNA-122 and represents viral replication and liver injury in patients with chronic hepatitis B. *J Med Virol* 2013; **85**: 789-798 [PMID: 23508904 DOI: 10.1002/jmv.23540]
- 100 **Li L**, Gu X, Fang M, Ji J, Yi C, Gao C. The diagnostic value of serum fucosylated fetuin A in hepatitis B virus-related liver diseases. *Clin Chem Lab Med* 2016; **54**: 693-701 [PMID: 26035113 DOI: 10.1515/cclm-2015-0307]
- 101 **Noh CK**, Kim SS, Kim DK, Lee HY, Cho HJ, Yoon SY, Lee GH, Hyun SA, Kim YJ, Kim HJ, Hwang JA, Ahn SJ, Shin SJ, Lee KM, Yoo BM, Cho SW, Cheong JY. Inter-alpha-trypsin inhibitor heavy chain H4 as a diagnostic and prognostic indicator in patients with hepatitis B virus-associated hepatocellular carcinoma. *Clin Biochem* 2014; **47**: 1257-1261 [PMID: 24836184 DOI: 10.1016/j.clinbiochem.2014.05.002]
- 102 **Sun QK**, Zhu JY, Wang W, Lv Y, Zhou HC, Yu JH, Xu GL, Ma JL, Zhong W, Jia WD. Diagnostic and prognostic significance of peroxiredoxin 1 expression in human hepatocellular carcinoma. *Med Oncol* 2014; **31**: 786 [PMID: 24297309 DOI: 10.1007/s12032-013-0786-2]
- 103 **Qiao B**, Wang J, Xie J, Niu Y, Ye S, Wan Q, Ye Q. Detection and identification of peroxiredoxin 3 as a biomarker in hepatocellular carcinoma by a proteomic approach. *Int J Mol Med* 2012; **29**: 832-840 [PMID: 22344546 DOI: 10.3892/ijmm.2012.916]
- 104 **Nafee AM**, Pasha HF, Abd El Aal SM, Mostafa NA. Clinical significance of serum clusterin as a biomarker for evaluating diagnosis and metastasis potential of viral-related hepatocellular carcinoma. *Clin Biochem* 2012; **45**: 1070-1074 [PMID: 22580393 DOI: 10.1016/j.clinbiochem.2012.04.024]
- 105 **Wang Y**, Liu YH, Mai SJ, He LJ, Liao YJ, Deng HX, Guan XY, Zeng YX, Kung HF, Xie D. Evaluation of serum clusterin as a surveillance tool for human hepatocellular carcinoma with hepatitis B virus related cirrhosis. *J Gastroenterol Hepatol* 2010; **25**: 1123-1128 [PMID: 20594228 DOI: 10.1111/j.1440-1746.2009.06205.x]
- 106 **Zheng W**, Yao M, Sai W, Qian Q, Pan L, Qiu L, Huang J, Wu W, Yao D. Diagnostic and prognostic significance of secretory clusterin expression in patients with hepatocellular carcinoma. *Tumour Biol* 2016; **37**: 999-1008 [PMID: 26264614 DOI: 10.1007/s13277-015-3875-3]
- 107 **Xie H**, Ma H, Zhou D. Plasma HULC as a promising novel biomarker for the detection of hepatocellular carcinoma. *Biomed Res Int* 2013; **2013**: 136106 [PMID: 23762823 DOI: 10.1155/2013/136106]
- 108 **Reichl P**, Fang M, Starlinger P, Stauer K, Nenutil R, Muller P, Greplova K, Valik D, Dooley S, Brostjan C, Gruenberger T, Shen J, Man K, Trauner M, Yu J, Gao CF, Mikulits W. Multicenter analysis of soluble Axl reveals diagnostic value for very early stage hepatocellular carcinoma. *Int J Cancer* 2015; **137**: 385-394 [PMID: 25529751 DOI: 10.1002/ijc.29394]
- 109 **Fan JL**, Yang YF, Yuan CH, Chen H, Wang FB. Circulating Tumor Cells for Predicting the Prognostic of Patients with Hepatocellular Carcinoma: A Meta Analysis. *Cell Physiol Biochem* 2015; **37**: 629-640 [PMID: 26344495 DOI: 10.1159/000430382]
- 110 **Kelley RK**, Magbanua MJ, Butler TM, Collisson EA, Hwang J, Sidiropoulos N, Evason K, McWhirter RM, Hameed B, Wayne EM, Yao FY, Venook AP, Park JW. Circulating tumor cells in hepatocellular carcinoma: a pilot study of detection, enumeration, and next-generation sequencing in cases and controls. *BMC Cancer* 2015; **15**: 206 [PMID: 25884197 DOI: 10.1186/s12885-015-1195-z]
- 111 **Yan J**, Fan Z, Wu X, Xu M, Jiang J, Tan C, Wu W, Wei X, Zhou J. Circulating tumor cells are correlated with disease progression and treatment response in an orthotopic hepatocellular carcinoma model. *Cytometry A* 2015; **87**: 1020-1028 [PMID: 26355643 DOI: 10.1002/cyto.a.22782]
- 112 **Papathodoridis G**, Dalekos G, Sypsa V, Yurdaydin C, Buti M, Goulis J, Calleja JL, Chi H, Manolakopoulos S, Mangia G, Gatselis N, Keskin O, Savvidou S, de la Revilla J, Hansen BE, Vlachogiannakos I, Galanis K, Idilman R, Colombo M, Esteban R, Janssen HL, Lampertico P. PAGE-B predicts the risk of developing hepatocellular carcinoma in Caucasians with chronic hepatitis B on 5-year antiviral therapy. *J Hepatol* 2016; **64**: 800-806 [PMID: 26678008 DOI: 10.1016/j.jhep.2015.11.035]
- 113 **Wong VW**, Janssen HL. Can we use HCC risk scores to individualize surveillance in chronic hepatitis B infection? *J Hepatol* 2015; **63**: 722-732 [PMID: 26026875 DOI: 10.1016/j.jhep.2015.05.019]
- 114 **Yang HI**, Lu SN, Liaw YF, You SL, Sun CA, Wang LY, Hsiao CK, Chen PJ, Chen DS, Chen CJ. Hepatitis B e antigen and the risk of hepatocellular carcinoma. *N Engl J Med* 2002; **347**: 168-174 [PMID: 12124405 DOI: 10.1056/NEJMoa013215]
- 115 **Chen CJ**, Yang HI, Su J, Jen CL, You SL, Lu SN, Huang GT, Iloeje UH. Risk of hepatocellular carcinoma across a biological gradient of serum hepatitis B virus DNA level. *JAMA* 2006; **295**: 65-73 [PMID: 16391218 DOI: 10.1001/jama.295.1.65]
- 116 **Chan HL**, Tse CH, Mo F, Koh J, Wong VW, Wong GL, Lam Chan

- S, Yeo W, Sung JJ, Mok TS. High viral load and hepatitis B virus subgenotype cc are associated with increased risk of hepatocellular carcinoma. *J Clin Oncol* 2008; **26**: 177-182 [PMID: 18182659 DOI: 10.1200/jco.2007.13.2043]
- 117 **Yang Y**, Gao J, Li HL, Zheng W, Yang G, Zhang W, Ma X, Tan YT, Rothman N, Gao YT, Chow WH, Shu XO, Xiang YB. Dose-response association between hepatitis B surface antigen levels and liver cancer risk in Chinese men and women. *Int J Cancer* 2016; **139**: 355-362 [PMID: 26990915 DOI: 10.1002/ijc.30086]
- 118 **Tseng TC**, Liu CJ, Yang HC, Su TH, Wang CC, Chen CL, Hsu CA, Kuo SF, Liu CH, Chen PJ, Chen DS, Kao JH. Serum hepatitis B surface antigen levels help predict disease progression in patients with low hepatitis B virus loads. *Hepatology* 2013; **57**: 441-450 [PMID: 22941922 DOI: 10.1002/hep.26041]
- 119 **Tseng TC**, Liu CJ, Yang HC, Su TH, Wang CC, Chen CL, Kuo SF, Liu CH, Chen PJ, Chen DS, Kao JH. High levels of hepatitis B surface antigen increase risk of hepatocellular carcinoma in patients with low HBV load. *Gastroenterology* 2012; **142**: 1140-1149.e3; quiz e13-14 [PMID: 22333950 DOI: 10.1053/j.gastro.2012.02.007]
- 120 **Lao X**, Ren S, Lu Y, Yang D, Qin X, Li S. Genetic polymorphisms of C-reactive protein increase susceptibility to HBV-related hepatocellular carcinoma in a Guangxi male population. *Int J Clin Exp Pathol* 2015; **8**: 16055-16063 [PMID: 26884882]
- 121 **Chanthra N**, Payungporn S, Chuaypen N, Pinjaroen N, Poovorawan Y, Tangkijvanich P. Association of Single Nucleotide Polymorphism rs1053004 in Signal Transducer and Activator of Transcription 3 (STAT3) with Susceptibility to Hepatocellular Carcinoma in Thai Patients with Chronic Hepatitis B. *Asian Pac J Cancer Prev* 2015; **16**: 5069-5073 [PMID: 26163643]
- 122 **Chanthra N**, Payungporn S, Chuaypen N, Piratanantatavorn K, Pinjaroen N, Poovorawan Y, Tangkijvanich P. Single Nucleotide Polymorphisms in STAT3 and STAT4 and Risk of Hepatocellular Carcinoma in Thai Patients with Chronic Hepatitis B. *Asian Pac J Cancer Prev* 2015; **16**: 8405-8410 [PMID: 26745093]
- 123 **Yao JY**, Chao K, Li MR, Wu YQ, Zhong BH. Interleukin-21 gene polymorphisms and chronic hepatitis B infection in a Chinese population. *World J Gastroenterol* 2015; **21**: 4232-4239 [PMID: 25892873 DOI: 10.3748/wjg.v21.i14.4232]
- 124 **Tan A**, Gao Y, Yao Z, Su S, Jiang Y, Xie Y, Xian X, Mo Z. Genetic variants in IL12 influence both hepatitis B virus clearance and HBV-related hepatocellular carcinoma development in a Chinese male population. *Tumour Biol* 2016; **37**: 6343-6348 [PMID: 26631030 DOI: 10.1007/s13277-015-4520-x]
- 125 **Zhang L**, Xu K, Liu C, Chen J. Meta-analysis reveals an association of STAT4 polymorphism with hepatocellular carcinoma risk. *Hepatol Res* 2016; Epub ahead of print [PMID: 27126090 DOI: 10.1111/hepr.12733]
- 126 **Jin F**, Xiong WJ, Jing JC, Feng Z, Qu LS, Shen XZ. Evaluation of the association studies of single nucleotide polymorphisms and hepatocellular carcinoma: a systematic review. *J Cancer Res Clin Oncol* 2011; **137**: 1095-1104 [PMID: 21240526 DOI: 10.1007/s00432-010-0970-0]
- 127 **Yao S**, Johnson C, Hu Q, Yan L, Liu B, Ambrosone CB, Wang J, Liu S. Differences in somatic mutation landscape of hepatocellular carcinoma in Asian American and European American populations. *Oncotarget* 2016; Epub ahead of print [PMID: 27246981 DOI: 10.18632/oncotarget.9636]
- 128 **Xu K**, Meng XY, Wu JW, Shen B, Shi YC, Wei Q. Diagnostic value of serum gamma-glutamyl transferase isoenzyme for hepatocellular carcinoma: a 10-year study. *Am J Gastroenterol* 1992; **87**: 991-995 [PMID: 1353662]
- 129 **Lin YJ**, Lee MH, Yang HI, Jen CL, You SL, Wang LY, Lu SN, Liu J, Chen CJ. Predictability of liver-related seromarkers for the risk of hepatocellular carcinoma in chronic hepatitis B patients. *PLoS One* 2013; **8**: e61448 [PMID: 23613855 DOI: 10.1371/journal.pone.0061448]
- 130 **Tong MJ**, Hsien C, Song JJ, Kao JH, Sun HE, Hsu L, Han SH, Durazo FA, Saab S, Blatt LM. Factors associated with progression to hepatocellular carcinoma and to death from liver complications in patients with HBsAg-positive cirrhosis. *Dig Dis Sci* 2009; **54**: 1337-1346 [PMID: 19242792 DOI: 10.1007/s10620-009-0747-y]
- 131 **Oldberg A**, Antonsson P, Lindblom K, Heinegård D. COMP (cartilage oligomeric matrix protein) is structurally related to the thrombospondins. *J Biol Chem* 1992; **267**: 22346-22350 [PMID: 1429587]
- 132 **Saxne T**, Heinegård D. Cartilage oligomeric matrix protein: a novel marker of cartilage turnover detectable in synovial fluid and blood. *Br J Rheumatol* 1992; **31**: 583-591 [PMID: 1381980]
- 133 **Xiao Y**, Kleeff J, Guo J, Gazdhar A, Liao Q, Di Cesare PE, Büchler MW, Friess H. Cartilage oligomeric matrix protein expression in hepatocellular carcinoma and the cirrhotic liver. *J Gastroenterol Hepatol* 2004; **19**: 296-302 [PMID: 14748877]
- 134 **Norman GL**, Gatselis NK, Shums Z, Liaskos C, Bogdanos DP, Koukoulis GK, Dalekos GN. Cartilage oligomeric matrix protein: A novel non-invasive marker for assessing cirrhosis and risk of hepatocellular carcinoma. *World J Hepatol* 2015; **7**: 1875-1883 [PMID: 26207169 DOI: 10.4254/wjh.v7.i14.1875]
- 135 **Wong VW**, Yu J, Cheng AS, Wong GL, Chan HY, Chu ES, Ng EK, Chan FK, Sung JJ, Chan HL. High serum interleukin-6 level predicts future hepatocellular carcinoma development in patients with chronic hepatitis B. *Int J Cancer* 2009; **124**: 2766-2770 [PMID: 19267406 DOI: 10.1002/ijc.24281]
- 136 **Jang JW**, Oh BS, Kwon JH, You CR, Chung KW, Kay CS, Jung HS. Serum interleukin-6 and C-reactive protein as a prognostic indicator in hepatocellular carcinoma. *Cytokine* 2012; **60**: 686-693 [PMID: 22906998 DOI: 10.1016/j.cyto.2012.07.017]
- 137 **Ohishi W**, Cologne JB, Fujiwara S, Suzuki G, Hayashi T, Niwa Y, Akahoshi M, Ueda K, Tsuge M, Chayama K. Serum interleukin-6 associated with hepatocellular carcinoma risk: a nested case-control study. *Int J Cancer* 2014; **134**: 154-163 [PMID: 23784949 DOI: 10.1002/ijc.28337]
- 138 **Cheng HY**, Kang PJ, Chuang YH, Wang YH, Jan MC, Wu CF, Lin CL, Liu CJ, Liaw YF, Lin SM, Chen PJ, Lee SD, Yu MW. Circulating programmed death-1 as a marker for sustained high hepatitis B viral load and risk of hepatocellular carcinoma. *PLoS One* 2014; **9**: e95870 [PMID: 25427199 DOI: 10.1371/journal.pone.0095870]
- 139 **Rose-John S**, Heinrich PC. Soluble receptors for cytokines and growth factors: generation and biological function. *Biochem J* 1994; **300** (Pt 2): 281-290 [PMID: 8002928]
- 140 **Wang M**, Block TM, Marrero J, Di Bisceglie AM, Devarajan K, Mehta A. Improved biomarker performance for the detection of hepatocellular carcinoma by inclusion of clinical parameters. *Proceedings (IEEE Int Conf Bioinformatics Biomed)* 2012; **2012**: [PMID: 24307972 DOI: 10.1109/bibm.2012.6392612]
- 141 **Shimauchi Y**, Tanaka M, Kuromatsu R, Ogata R, Tateishi Y, Itano S, Ono N, Yutani S, Nagamatsu H, Matsugaki S, Yamasaki S, Tanikawa K, Sata M. A simultaneous monitoring of Lens culinaris agglutinin A-reactive alpha-fetoprotein and des-gamma-carboxy prothrombin as an early diagnosis of hepatocellular carcinoma in the follow-up of cirrhotic patients. *Oncol Rep* 2000; **7**: 249-256 [PMID: 10671666]
- 142 **Choi JY**, Jung SW, Kim HY, Kim M, Kim Y, Kim DG, Oh EJ. Diagnostic value of AFP-L3 and PIVKA-II in hepatocellular carcinoma according to total-AFP. *World J Gastroenterol* 2013; **19**: 339-346 [PMID: 23372355 DOI: 10.3748/wjg.v19.i3.339]
- 143 **Pepe MS**, Etzioni R, Feng Z, Potter JD, Thompson ML, Thornquist M, Winget M, Yasui Y. Phases of biomarker development for early detection of cancer. *J Natl Cancer Inst* 2001; **93**: 1054-1061 [PMID: 11459866]
- 144 **da Costa AN**, Plymoth A, Santos-Silva D, Ortiz-Cuaran S, Camey S, Guilloreau P, Sangrajang S, Khuhaprema T, Mendy M, Lesi OA, Chang HK, Oh JK, Lee DH, Shin HR, Kirk GD, Merle P, Beretta L, Hainaut P. Osteopontin and latent-TGF β binding-protein 2 as potential diagnostic markers for HBV-related hepatocellular carcinoma. *Int J Cancer* 2015; **136**: 172-181 [PMID: 24803312 DOI: 10.1002/ijc.28953]
- 145 **Yang JD**, Gyedu A, Afihene MY, Duduyemi BM, Micah E, Kingham TP, Nyirenda M, Nkansah AA, Bandoh S, Duguru MJ, Okeke EN, Kouakou-Lohoues MJ, Abdo A, Awuku YA, Ajayi

- AO, Omonisi AE, Ocamá P, Malu AO, Mustapha S, Okonkwo U, Kooffreh-Ada M, Debes JD, Onyekwere C, Ekere F, Rufina I, Roberts LR. Hepatocellular Carcinoma Occurs at an Earlier Age in Africans, Particularly in Association With Chronic Hepatitis B. *Am J Gastroenterol* 2015; **110**: 1629-1631 [PMID: 26618430 DOI: 10.1038/ajg.2015.289]
- 146 **Park JW**, Chen M, Colombo M, Roberts LR, Schwartz M, Chen PJ, Kudo M, Johnson P, Wagner S, Orsini LS, Sherman M. Global patterns of hepatocellular carcinoma management from diagnosis to death: the BRIDGE Study. *Liver Int* 2015; **35**: 2155-2166 [PMID: 25752327 DOI: 10.1111/liv.12818]
- 147 **He QY**, Zhu R, Lei T, Ng MY, Luk JM, Sham P, Lau GK, Chiu JF. Toward the proteomic identification of biomarkers for the prediction of HBV related hepatocellular carcinoma. *J Cell Biochem* 2008; **103**: 740-752 [PMID: 17557278 DOI: 10.1002/jcb.21443]
- 148 **Feng JT**, Liu YK, Song HY, Dai Z, Qin LX, Almofti MR, Fang CY, Lu HJ, Yang PY, Tang ZY. Heat-shock protein 27: a potential biomarker for hepatocellular carcinoma identified by serum proteome analysis. *Proteomics* 2005; **5**: 4581-4588 [PMID: 16240287 DOI: 10.1002/pmic.200401309]
- 149 **King LY**, Canasto-Chibuque C, Johnson KB, Yip S, Chen X, Kojima K, Deshmukh M, Venkatesh A, Tan PS, Sun X, Villanueva A, Sangiovanni A, Nair V, Mahajan M, Kobayashi M, Kumada H, Iavarone M, Colombo M, Fiel MI, Friedman SL, Llovet JM, Chung RT, Hoshida Y. A genomic and clinical prognostic index for hepatitis C-related early-stage cirrhosis that predicts clinical deterioration. *Gut* 2015; **64**: 1296-1302 [PMID: 25143343 DOI: 10.1136/gutjnl-2014-307862]
- 150 **Hoshida Y**, Villanueva A, Kobayashi M, Peix J, Chiang DY, Camargo A, Gupta S, Moore J, Wrobel MJ, Lerner J, Reich M, Chan JA, Glickman JN, Ikeda K, Hashimoto M, Watanabe G, Daidone MG, Roayaie S, Schwartz M, Thung S, Salvesen HB, Gabriel S, Mazzaferro V, Bruix J, Friedman SL, Kumada H, Llovet JM, Golub TR. Gene expression in fixed tissues and outcome in hepatocellular carcinoma. *N Engl J Med* 2008; **359**: 1995-2004 [PMID: 18923165 DOI: 10.1056/NEJMoa0804525]

P- Reviewer: Gao YT, Wan SS, Zhou XH **S- Editor:** Yu J
L- Editor: A **E- Editor:** Wang CH





Published by **Baishideng Publishing Group Inc**
8226 Regency Drive, Pleasanton, CA 94588, USA
Telephone: +1-925-223-8242
Fax: +1-925-223-8243
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>
<http://www.wjgnet.com>



ISSN 1007-9327

