

Observational Study

Hypolactasia is associated with insulin resistance in nonalcoholic steatohepatitis

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Abstract

AIM

To assess lactase gene (*LCT*)-13910C>T polymorphisms in Brazilian non-alcoholic fatty liver disease (NAFLD) and nonalcoholic steatohepatitis (NASH) patients in comparison with healthy controls.

METHODS

This was a transverse observational clinical study with NAFLD patients who were followed at the Hepatology Outpatient Unit of the Hospital das Clínicas, São Paulo, Brazil. The polymorphism of lactase non-persistence/lactase persistence (*LCT*-13910C>T) was examined by PCR-restriction fragment length polymorphism technique in 102 liver biopsy-proven NAFLD patients (steatosis in 9 and NASH in 93) and compared to those of 501 unrelated healthy volunteers. Anthropometric, clinical, biochemical and liver histology data were analyzed. Continuous variables were compared using the *t* or Mann-Whitney tests, and categorical data were compared with the Fisher's exact test. Univariate logistic regression and

multivariate logistic regression adjusted for gender and age were performed.

RESULTS

No differences in the *LCT*-13910 genotype frequencies were noted between the NAFLD patients (66.67% of the patients with steatosis were CC, 33.33% were CT, and none were TT; 55.91% of the patients with NASH were CC, 39.78% were CT, and 4.3% were TT; $P = 0.941$) and the healthy controls (59.12% were CC, 35.67% were CT, and 5.21% were TT) or between the steatosis and NASH patients. That is, the distribution of the lactase non-persistence/lactase persistence polymorphism (*LCT*-13910C>T) in the patients with NAFLD was equal to that in the general population. In the NASH patients, the univariate analysis revealed that the lactase non-persistence (low lactase activity or hypolactasia) phenotype was associated with higher insulin levels ($23.47 \pm 15.94 \mu\text{U/mL}$ vs $15.8 \pm 8.33 \mu\text{U/mL}$, $P = 0.027$) and a higher frequency of insulin resistance (91.84% vs 72.22%, $P = 0.02$) compared with the lactase persistence phenotype. There were no associations between the *LCT* genotypes and diabetes ($P = 0.651$), dyslipidaemia ($P = 0.328$), hypertension ($P = 0.507$) or liver histology in these patients. Moreover, in the NASH patients, hypolactasia was an independent risk factor for insulin resistance even after adjusting for gender and age [OR = 5.0 (95%CI: 1.35-20; $P = 0.017$)].

CONCLUSION

The *LCT*-13910 genotype distribution in Brazilian NAFLD patients was the same as that of the general population, but hypolactasia increased the risk of insulin resistance in the NASH patients.

Key words: Lactose intolerance; Genetic polymorphism; Insulin resistance; Non-alcoholic fatty liver disease; Nonalcoholic steatohepatitis

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Core tip: Non-alcoholic fatty liver disease (NAFLD) exhibits a close relationship with metabolic syndrome (MetS), but the associations of the lactase non-persistence/lactase persistence genotypes with MetS components are controversial. Therefore, we assessed hypolactasia (*LCT*-13910CC) and lactase persistence genotypes in 102 Brazilian NAFLD patients in comparison with 501 healthy controls, the associations of these polymorphisms were verified with the results of biochemical tests, MetS and severity of liver histology in nonalcoholic steatohepatitis (NASH) patients. No differences in the *LCT*-13910C>T polymorphisms were noted between the NAFLD and controls, but hypolactasia increased the risk of insulin resistance in the NASH patients.

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INTRODUCTION

Non-alcoholic fatty liver disease (NAFLD) encompasses a wide spectrum of liver damage that ranges from steatosis to nonalcoholic steatohepatitis (NASH) and advanced fibrosis/cirrhosis in persons without significant alcohol consumption^[1,2]. NAFLD is currently considered the most common liver disease and is associated to metabolic syndrome (MetS) components, such as obesity and diabetes^[3-5]. Several studies have correlated the severity of liver injury with increased frequencies of such components, thus making these components important targets in the management of this condition^[1,6-8]. However, while specific pharmacological therapy are still far from solving all of the issues related to fatty liver disease, the pursuit of high-risk individuals can be a strategy for concentrating efforts on its diagnosis and management.

Milk is the primary energy source for newborns and is rich in lactose. Lactase phlorizin hydrolase in the microvillus membrane of the small intestinal cells digests lactose. However, after 2-12 years of age, a physiological genetically programmed reduction in lactase activity occurs, hypolactasia or lactase non-persistence, which, when accompanied by symptoms, defines lactose intolerance^[9]. In contrast some populations mainly from Northern Europe present lactase persistence during adulthood^[10]. The most interesting report published in 2002^[11] found that the polymorphisms in intron 13 [lactase gene (*LCT*)-13910C>T] and in intron 9 (*LCT*-22018G>A) of the *MCM6* gene conferred lactase persistence in several populations^[9,12-14]. These genotypes render a person a lactose digester. The lactase-persistence phenotype has a prevalence of 43.4% in Caucasian Brazilians, and there is no difference between genders^[12].

Recent studies have raised concerns regarding the possible associations of lactase persistence with the components of MetS. In Europeans those with hypolactasia genotype (*LCT*-13910CC) had lower body mass indices and waist circumferences than those with lactase persistence genotypes^[15,16]. Likewise, in the Canary Islands, those with lactase persistence genotypes exhibit higher odds ratios for MetS than do subjects with the *LCT*-13910CC genotype^[17].

However, other studies have demonstrated that dairy food consumption showed lower susceptibility to type 2 diabetes or worsening of glucose homeostasis indices^[18-20]. Nicklas *et al*^[21] applied a questionnaire to a sample of 3452 American adults and reported that diagnosis of diabetes and hypertension were higher in individuals that considered themselves lactose intolerant with lower ingestion of calcium from dairies. Additionally, Samara *et al*^[22] assessed a French population and

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reported that better metabolic profiles in men was associated with more dairies intake.

As noted, the role of milk in MetS is not clearly defined at this moment, and the literature is controversial^[23]. Moreover, publications regarding the *LCT-13910C>T* polymorphism in patients with NAFLD are scarce. Therefore, the purpose was to assess expression profiles of the *LCT-13910* genotypes in Brazilians with NAFLD compared to those of healthy individuals to investigate whether the *LCT-13910C>T* variant could be a predictor of NASH. An additional goal was to analyze the associations of the lactase-persistence genotype with biochemical tests, components of MetS and the severity of liver histology in NASH patients.

MATERIALS AND METHODS

Ethical considerations

The Ethics Committee of the Hospital das Clínicas (number 448520) approved this study that was conducted following the ethical guidelines of the 1975 Declaration of Helsinki.

Patients and clinical design

This was a transverse study with NAFLD patients who were followed at the Hepatology Outpatient Unit of the Hospital das Clínicas, São Paulo, Brazil. *LCT-13910C>T* polymorphism was investigated in 102 liver biopsy-proven NAFLD patients and 501 unrelated healthy volunteers. All NAFLD patients were previously evaluated for other liver diseases, being excluded viral hepatitis, autoimmune hepatitis, hemochromatosis, Wilson disease and alpha 1-antitrypsin deficiency. MetS components identification followed the recommendations of the Adult Treatment Panel III Report as follows: Triglycerides ≥ 150 mg/dL, high-density lipoproteins (HDL) < 40 mg/dL in men and < 50 mg/dL in women, fasting glucose ≥ 110 mg/dL, ≥ 130 mmHg systolic or ≥ 85 mmHg diastolic pressure, and abdominal obesity^[24]. The study inclusion criteria were patients 18-75 years old with NAFLD diagnoses based on liver histology. Exclusion criteria were any other liver disease, significant alcohol intake (> 100 g/wk), previous exposure to drugs associated with liver steatosis or not accepting to participate in the study.

Liver histology were scored according to the macro- and micro-vacuolar steatosis, the inflammation and the hepatocyte ballooning. Fibrosis pattern and zonal distributions of the analysed variables were also recorded. The slides were classified according to the NASH Clinical Research Network^[25]. The biochemical investigations included the following: Fasting glucose, plasma insulin, total cholesterol and fractions, triglycerides, alanine aminotransferase (ALT), aspartate aminotransferase (AST), and gamma glutamyl transferase (GGT), which were collected after a 12-h overnight fast and evaluated at the time of the liver biopsy. Homeostatic Model of Assessment (HOMA-IR) was used to evaluate insulin resistance [$22.5 \times$ fasting insulin (mU/mL) \times glucose (mmol/L)]^[26]. A HOMA-IR ≥ 2.5 was used as the cutoff

point to define insulin resistance^[27,28]. Retrospective information regarding co-morbidities was also collected.

Genotyping

Leukocytes were used for genomic DNA extraction (Miller *et al.*^[29] 1988). The technique for *LCT-13910* genotyping was described elsewhere^[11,30-32].

Statistical analysis

The continuous variables are presented as the means \pm the standard deviations and were compared using the *t* test (the assumption of normality was verified using the Anderson-Darling test). When appropriate, the Mann-Whitney test was used. The categorical variables are expressed as the percentages (frequencies) of affected individuals and were compared using Fisher's exact test. Univariate logistic regression was performed to evaluate the odds ratios with the respective 95% CIs. Multivariate logistic regression adjusted for gender and age was performed. The best predictive cut-offs for the continuous variables were determined using conditional trees when the traditional cut-offs did not provide interesting information^[33]. *P* values below 0.05 were considered statistically significant. The R Project for Statistical Computing ver. 3.1.1 (R Core Team, Vienna, Austria, 2014) software package was used for the statistical analyses^[34]. A statistical review of the study was performed by a biomedical statistician (Márcio Augusto Diniz).

RESULTS

The anthropometric, clinical, and biochemical characteristics of the patients are provided in Table 1. We evaluated 102 NAFLD patients, including 9 steatosis and 93 with NASH. All of the steatosis patients were women, whereas in the NASH group, 32 patients (34.41%) were men ($P = 0.04$). The NASH patients had higher fasting glucose levels than did the patients with steatosis only (123.14 ± 48.28 vs 91.71 ± 9.2 , respectively, $P = 0.033$). There were no differences between the groups in terms of age, MetS components, BMI, insulin, HOMA-IR values ≥ 2.5 , AST, ALT, GGT, total cholesterol, HDL, LDL or triglycerides (Table 1).

The distributions of alleles and genotypes are presented in Table 2. No differences in *LCT-13910* genotype frequencies were noted between the NAFLD patients (66.67% patients with steatosis were CC, 33.33% were CT and none were TT; 55.91% of those with NASH were CC, 39.78% were CT and 4.3% were TT; $P = 0.941$) and the healthy controls (59.12% were CC, 35.67% CT, 5.21% TT). Likewise, no differences in the *LCT-13910C>T* allele frequencies were noted between the groups (76.95% of the controls, 83.33% of those with steatosis and 75.81% of the NASH patients had the *LCT-13910C* allele; $P = 0.764$). That is, the distribution of the *LCT-13910C>T* polymorphism in the patients with NAFLD was equal to that in the general population.

Table 1 Demographic, clinical and biochemical characteristics of the non-alcoholic fatty liver disease patients

	Steatosis (<i>n</i> = 9)	NASH (<i>n</i> = 93)	<i>P</i> value
Age	55.11 ± 10.3	56.51 ± 10.13	0.692
Men/women (<i>n</i>)	0% (0)/100% (9)	34.41% (32)/65.59% (61)	0.04 ^a
Type 2 diabetes (<i>n</i>)	33.33% (2)	60.67% (54)	0.224
Dyslipidaemia (<i>n</i>)	83.33% (5)	79.78% (71)	1
High-blood pressure (<i>n</i>)	66.67% (4)	64.04% (89)	1
BMI	31.28 ± 5.79	31.25 ± 5.93	0.969
Fasting glucose (mg/dL)	91.71 ± 9.2	123.14 ± 48.28	0.033 ^a
Insulin (μU/mL)	12.44 ± 4.2	19.92 ± 13.29	0.102
HOMA-IR value ≥ 2.5	57.14%	83.53%	0.115
AST (U/L)	25.14 ± 6.89	38.8 ± 37.99	0.159
ALT (U/L)	40 ± 16.74	50.65 ± 54.99	0.934
GGT (U/L)	56.57 ± 59.9	87.36 ± 96.33	0.185
Total cholesterol (mg/dL)	203.29 ± 54.39	195.31 ± 45.71	0.863
HDL (mg/dL)	53 ± 6.58	46.15 ± 13.42	0.067
LDL (mg/dL)	124.14 ± 51.87	114.89 ± 39.74	0.72
Triglycerides (mg/dL)	130.43 ± 59.19	167.25 ± 82.02	0.258

^a*P* value < 0.05. ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; BMI: Body mass index; HDL: High-density lipoprotein; HOMA-IR: Homeostatic Model of Assessment; LDL: Low-density lipoprotein; GGT: Gamma glutamyl transferase; NASH: Nonalcoholic steatohepatitis.

Analysis *via* simple logistic regressions of the associations of the LCT-13910C>T polymorphisms with the results of the biochemical tests, components of MetS and severity of liver histology in the NAFLD patients (steatosis and NASH groups) did not reveal any associations (data not shown). Subsequently, we evaluated the patients with NASH (Table 3). In this group (*n* = 93), univariate analysis revealed that the hypolactasia phenotype was associated with higher insulin levels (*P* = 0.027) and greater insulin resistance (*P* = 0.02). No associations were noted between the liver histology parameters (*i.e.*, steatosis, inflammation and fibrosis) and the LCT-13910 genotype or phenotype. Moreover, no associations were found between the components of MetS or MetS diagnosis (*P* = 1.0) and the LCT-13910 genotype or phenotype.

Table 4 illustrates the logistic regression analysis that was adjusted for gender and age and assessed the independent associations of the LCT-13910C>T polymorphism with HOMA-IR, BMI ≥ 30, insulin value and MetS in the NASH patients. Hypolactasia phenotype was associated with a 5-fold increase in insulin resistance (95%CI: 1.35-20; *P* = 0.017). The LCT-13910CT genotype conferred a 6.25-fold decrease in insulin resistance (95%CI: 0.04-0.64; *P* = 0.009). In this multivariate regression analysis, we no longer observed an association between hypolactasia and insulin level (even when using the cut-off of > 29.8 μU/mL, *P* = 0.197) after adjusting for gender and age. Similarly, the MetS diagnosis and a BMI ≥ 30 were not associated with the LCT-13910C>T polymorphism.

DISCUSSION

Key findings

In this transverse clinical study, we were unable to find any differences in the LCT-13910C>T polymorphism

expression profile between Brazilian NAFLD patients and healthy controls (*P* = 0.941). Moreover, the presence of the T allele was not able to differentiate steatosis from NASH in NAFLD patients (*P* = 0.764). However, in NASH patients, the hypolactasia phenotype (*i.e.*, the LCT-13910CC genotype) was associated with insulin resistance, and conversely, the LCT-13910CT genotype conferred protection against its occurrence.

The LCT-13910C>T polymorphism prevalence varies among different populations across the globe. The lactase-persistence phenotype (*i.e.*, the LCT-13910-CT and LCT-13910-TT genotypes) can occur at rates as high as 72% and 73.7% in New Zealand and Sweden, respectively^[13,35]. In Hungary, the prevalence is 35.9%, and in Caucasian Brazilians, the prevalence is 43.4%^[12,36]. In contrast, in Chinese and Japanese Brazilians, the lactase-persistence phenotype was not found at all in some published studies^[12,37]. The LCT genotype distribution was also the same in NAFLD patients regardless of the presence of NASH or steatosis only.

In a recent European meta-analysis with 31720 individuals, Kettunen *et al.*^[16] found that the LCT-13910CC genotype was associated with a decreased body mass index (BMI), when compared to LCT-13910CT/TT. In an analysis of 17374 Finns, it was observed that when the lactase persistent allele was present, BMI was 0.3 kg/m² higher, which corresponds to approximately 1 kg^[16]. These findings were reproduced by Corella *et al.*^[15] in a Mediterranean population in which LCT-13910CC individuals exhibited a lower risk of obesity, lower body weights, lower BMIs and smaller waist circumferences than LCT-13910T-allele carriers. Although the association between the LCT-13910C>T genotypes and the diagnosis of full-blown MetS was not significant in the overall analysis in the study, a subgroup analysis revealed a significant association in the subjects with a lactose intake higher than 8 g/d^[15]. In a cross-sectional

Table 2 Allele and genotype frequencies of the lactase-13910C>T polymorphisms

		Allele frequency % (n) ^a		Total (%)	Genotype frequency % (n) ^b			Total (%)
		C	T		CC	CT	TT	
<i>LCT</i> -13910	Control (n = 501)	76.95 (768)	23.05 (230)	100	59.12 (295)	35.67 (178)	5.21 (26)	100
	Steatosis (n = 9)	83.33 (15)	16.67 (3)	100	66.67 (6)	33.33 (3)	0 (0)	100
	NASH (n = 93)	75.81 (141)	24.19 (43)	100	55.91 (52)	39.78 (37)	4.3 (3)	100

^aP = 0.764; ^bP = 0.941. NASH: Nonalcoholic steatohepatitis; *LCT*: Lactase gene.

Table 3 Associations of the lactase-13910 phenotype in nonalcoholic steatohepatitis patients (n = 93)

	Hypolactasia	Lactase persistence	P value
Age	55.96 ± 10.91	57.61 ± 9.33	0.443
Gender: Female % (n)	70.83 (34)	60.98 (25)	0.51
Type 2 diabetes % (n)	66.67 (30)	57.5 (23)	0.664
Dyslipidaemia % (n)	82.22 (37)	77.5 (31)	0.792
High-blood pressure % (n)	68.89 (31)	57.5 (23)	0.273
BMI	31.39 ± 6.55	31.28 ± 5.37	0.714
BMI ≥ 30 % (n)	58.14 (25)	65 (26)	0.388
Fasting glucose (mg/dL)	122.61 ± 50.14	123.83 ± 46.43	0.892
Insulin (μU/mL)	23.47 ± 15.94	15.8 ± 8.33	0.027 ^a
HOMA-IR value ≥ 2.5 (n)	91.84 (45)	72.22 (26)	0.02 ^a
AST (U/L)	38.94 ± 37.66	42.67 ± 40.15	0.121
ALT (U/L)	51.47 ± 69.44	52.12 ± 35.02	0.072
GGT (U/L)	97.49 ± 118.27	80.51 ± 65.6	0.427
Total cholesterol (mg/dL)	196.62 ± 47.13	195.55 ± 44.06	0.965
HDL (mg/dL)	46.38 ± 12.26	46.05 ± 15.06	0.698
LDL (mg/dL)	117.19 ± 39.47	114.25 ± 39.9	0.893
Triglycerides (mg/dL)	169.11 ± 82.97	162.3 ± 83.11	0.477
Steatosis			
1	21.28 (10)	24.39 (10)	0.453
2	51.06 (24)	39.02 (16)	
3	27.66 (13)	36.59 (15)	
Inflammation			
0	2.13 (1)	7.32 (3)	0.133
1	61.7 (29)	46.34 (19)	
2	23.4 (11)	39.02 (16)	
3	12.77 (6)	7.32 (3)	
Fibrosis			
0	18.75 (9)	14.63 (6)	0.804
1	39.58 (19)	43.9 (18)	
2	16.67 (8)	17.07 (7)	
3	20.83 (10)	19.51 (8)	
4	4.17 (2)	4.88 (2)	
MetS	51.92 (27)	53.66 (19)	1

^aP value < 0.05. ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; BMI: Body mass index; HDL: High-density lipoprotein; HOMA-IR: Homeostatic Model of Assessment; LDL: Low-density lipoprotein; GGT: Gamma glutamyl transferase; MetS: Metabolic syndrome.

work conducted in the Canary Islands, Almon *et al.*^[17] demonstrated that subjects with the *LCT*-13910CT and *LCT*-13910TT genotypes exhibited higher odds ratio for MetS than subjects with the *LCT*-13910CC genotype. The authors concluded that the T allele might constitute a nutrigenetic factor that increases the susceptibility to MetS development, and this susceptibility was particularly noted in women^[17].

Despite the aforementioned studies that have demonstrated correlations of the CC genotype with decreased BMI, a lower risk of obesity, a lower body weight, and smaller waist circumference compared with the CT and TT genotypes^[15,16] and the even further increased

higher odds ratio for MetS in individuals with the T allele^[17], we could not corroborate these findings in our NAFLD population. Studying only the NASH patients in the univariate analysis, we did not find associations between the *LCT*-13910C>T polymorphism and BMI or MetS diagnoses even after adjusting for gender and age in the multivariate analysis. In fact, the patients with NASH and a genetic profile of persistent lactase activity exhibited less insulin resistance than the patients with hypolactasia. These divergences in our findings could be related to differences in the studied populations and possible positive effects of dairy ingestion on the metabolic profiles of these individuals.

Table 4 Multivariate logistic regression analysis in non-alcoholic steatohepatitis patients

Factor	OR	95%CI	P value
HOMA-IR value ≥ 2.5			
Hypolactasia phenotype	5	1.35-20	0.017 ^a
CT genotype	0.16	0.04-0.64	0.009 ^a
TT genotype	-	-	0.994
BMI ≥ 30			
Hypolactasia phenotype	0.49	0.13-1.81	0.285
CT genotype	1.73	0.69-4.35	0.244
TT genotype	1.01	0.12-8.39	0.991
Insulin > 29.8			
Hypolactasia phenotype	2.04	0.68-6.25	0.197
CT genotype	0.52	0.17-1.56	0.25
TT genotype	-	-	0.991
MetS			
Hypolactasia phenotype	0.94	0.47-2.42	0.89
CT genotype	1.07	0.46-2.49	0.866
TT genotype	0.91	0.11-7.3	0.929

^aP value < 0.05. HOMA-IR: Homeostatic Model of Assessment; OR: Odds ratio; BMI: Body mass index; MetS: Metabolic syndrome.

Our studied population consisted only of NAFLD patients, among which the prevalences of MetS components are expected to be higher than those of the overall population. Therefore, firm direct comparisons are precluded. However, a recently published Brazilian study demonstrated that in the general population, the lactase non-persistence genotype subjects exhibit higher prevalences of hypertension ($P = 0.032$) and MetS ($P = 0.01$) than lactase-persistence genotype individuals based on univariate analysis^[38]. Furthermore, multivariate analyses revealed that lactase persistence was associated with a lower risk for MetS after adjusting for gender, age, BMI and physical activity (OR = 0.462; $P = 0.009$). These data are in line with our findings that demonstrated a favourable profile of MetS components and glucose homeostasis in the NASH patients with lactase persistence. Moreover, in a longitudinal French study encompassing 3575 subjects, Lamri *et al.*^[39] demonstrated that the C allele was associated with a higher frequency of impaired fasting glycaemia and type 2 diabetes. However, Enattah *et al.*^[40] were unable to demonstrate that lactase persistence polymorphisms were risk factors for type 1 or type 2 diabetes in the Finnish study. Similar to NASH, polycystic ovary syndrome is also frequently associated with metabolic disturbances, including dyslipidaemia, insulin resistance and central obesity, and NASH often coexists in these patients^[41]. Lerchbaum *et al.*^[42] demonstrated a significantly higher prevalence of hypolactasia in polycystic ovary syndrome women, which also corroborates our findings.

Ultimately, we believe that dairy consumption appears to modulate the metabolic profiles of these different populations because of the strong association of the LCT-13910 genotype with dairies intake and lactose malabsorption^[11,31,39,43]. Several studies have highlighted the benefits of dairy and dairy components on MetS components^[18-22,44-46] and cardiovascular health^[47]. The

benefits of dairy products may be mediated through several mechanisms, including the following^[23,48]: The insulinotropic role of whey and its beneficial effect on body weight and fat; the favorable effects of amino acids, medium chain fatty acids, calcium and other minerals found in milk and its derivatives; improvements in insulin sensitivity due to medium chain fatty acids; reductions in the absorption of cholesterol and other fats from fermented products; the probiotic bacteria present in these foods and the associated proteins and peptides; and improvements in weight control, blood pressure and plasma lipids due to lactose, citrate, proteins and peptides. Specifically addressing glucose homeostasis, a hypothetical explanation is that milk and dairy consumption may be associated with an enhanced insulinaemic response, decreased glycemic fluctuations, and increased secretion of glucagon-like peptide 1 and glucose-dependent insulinotropic polypeptide^[49].

Experimental models also provide some mechanistic explanations that link dairy consumption with lower incidences of insulin resistance and diabetes^[50]. Milk components such as rumenic acid, vaccenic acid, phytanic acid and its derivative pristanic acid have been demonstrated to improve insulin resistance through PPAR signalling activation in different rat models^[51-54]. These findings suggest that dairy consumption could have a role in insulin resistance and NASH management.

However, in our study, there was no association between LCT-13910 genotype and the severity of liver histology in the NASH patients. The reason for this finding may be that the pathogenesis of NASH involves a complex multiple parallel hits process in which a number of different events may contribute to liver injury^[55]. Lifestyle and genetic predisposition remain relevant disease determinants. The consumption of high-calorie diets rich in lipids results in weight gain, obesity and insulin resistance. Moreover, a diet high in carbohydrates (mainly fructose) and saturated fatty acids contributes to the production of excess free fatty acids, whose safe disposal is impaired, which results in oxidative stress and NASH^[56]. Recent data have also demonstrated a potential role of the microbiota in the induction of insulin resistance and the development of NAFLD/NASH^[57-59]. The major components of the gut microbiota at the phylum level are *Bacteroidetes* and *Firmicutes*^[60]. It has been demonstrated that *Firmicutes* levels are elevated in obesity and related diseases, whereas *Bacteroidetes* levels are decreased, which leads to an increase in the *Firmicutes/Bacteroidetes* ratio^[61,62]. Interestingly, it has been shown that lysozyme-rich milk consumption results in a decline in *Firmicutes* levels (mainly *Clostridia* spp.) and in an increase in *Bacteroidetes* levels over time^[63,64]. Despite the absence of high levels of lysozyme in the milk of dairy animals, these studies highlighted the potential role of milk and its components in the composition of the microbiome in health and disease.

The main limitations of our study are the lack of alimentary reports from the NAFLD patients to quantify the dairy intakes and the absence of ethnic data because

the prevalence of LCT-13910C>T polymorphisms may vary widely, as has been previously demonstrated^[12].

In conclusion, we demonstrate that hypolactasia (*i.e.*, the LCT-13910CC genotype) is associated with a higher insulin resistance frequency in NASH patients. However, further studies that include dairy ingestion reports are needed to elucidate the associations of the lactase-persistence phenotype with MetS and NAFLD/NASH in different populations.

COMMENTS

Background

Non-alcoholic fatty liver disease (NAFLD) encompasses a wide spectrum of liver damage that ranges from steatosis to nonalcoholic steatohepatitis (NASH) and cirrhosis in persons without significant alcohol consumption and has a close relationship with metabolic syndrome (MetS). The lactase gene (LCT)-13910C>T polymorphism located upstream of the LCT is tightly associated with lactase persistence. The LCT-13910CT and LCT-13910TT genotypes are associated with the lactase-persistence phenotype, *i.e.*, they render a person a lactose digester, whereas the LCT-13910CC genotype is associated with lactose malabsorption.

Research frontiers

The role of milk in MetS is not currently clearly defined, and the literature is controversial. Moreover, to our knowledge, there are no published data regarding the LCT-13910C>T polymorphism in patients with NAFLD. Therefore, the authors assessed the expression profile of LCT-13910 genotypes in Brazilian patients with NAFLD in comparison with those of healthy controls to investigate whether the LCT-13910C>T variant could be a predictor of NASH. Furthermore, in NASH patients, the authors analyzed the associations of the lactase-persistence genotype with the results of biochemical tests, components of MetS and the severity of liver histology.

Innovations and breakthroughs

The authors were unable to find any differences in the LCT-13910C>T polymorphism expression profiles between Brazilian NAFLD patients and healthy controls. Moreover, the presence of the T allele was not able to discriminate steatosis from NASH in NAFLD patients. However, in NASH patients, the hypolactasia phenotype (*i.e.*, the LCT-13910CC genotype) was associated with insulin resistance, and conversely, the LCT-13910CT genotype conferred protection against its occurrence.

Applications

Specific pharmacological therapy for NASH is still lacking, so the pursuit of high-risk individuals can be a strategy for concentrating efforts on its diagnosis and management. Dairy consumption appears to modulate the metabolic profile because hypolactasia was found to be an independent risk factor for insulin resistance in NASH patients. Further studies that include dairy ingestion reports are needed to elucidate the associations of the lactase-persistence phenotype with MetS and NAFLD/NASH in different populations.

Terminology

NAFLD: Non-alcoholic fatty liver disease, which encompasses a wide spectrum of liver damage that ranges from steatosis to NASH and cirrhosis in persons without significant alcohol consumption. The MetS components include the following: Fasting glucose ≥ 110 mg/dL, triglyceride ≥ 150 mg/dL, high-density lipoprotein < 40 mg/dL in men or < 50 mg/dL in women, ≥ 130 mmHg systolic or ≥ 85 mmHg diastolic pressure and abdominal obesity. The LCT-13910CT and LCT-13910TT genotypes are associated with the lactase-persistence phenotype, *i.e.*, these genotypes render a person a lactose digester, whereas the LCT-13910CC genotype is associated with hypolactasia, *i.e.*, lactose malabsorption.

Peer-review

The paper indicated that among nonalcoholic steatohepatitis patients, hypo-

lactasia is associated with insulin resistance in Brazil. It is a very interesting and well-written paper.

REFERENCES

- 1 **Ratziu V**, Bellentani S, Cortez-Pinto H, Day C, Marchesini G. A position statement on NAFLD/NASH based on the EASL 2009 special conference. *J Hepatol* 2010; **53**: 372-384 [PMID: 20494470 DOI: 10.1016/j.jhep.2010.04.008]
- 2 **Farrell GC**, Larter CZ. Nonalcoholic fatty liver disease: from steatosis to cirrhosis. *Hepatology* 2006; **43**: S99-S112 [PMID: 16447287 DOI: 10.1002/hep.20973]
- 3 **Younossi ZM**, Stepanova M, Afendy M, Fang Y, Younossi Y, Mir H, Srisord M. Changes in the prevalence of the most common causes of chronic liver diseases in the United States from 1988 to 2008. *Clin Gastroenterol Hepatol* 2011; **9**: 524-530.e1; quiz e60 [PMID: 21440669 DOI: 10.1016/j.cgh.2011.03.020]
- 4 **Gaggini M**, Morelli M, Buzzigoli E, DeFronzo RA, Bugianesi E, Gastaldelli A. Non-alcoholic fatty liver disease (NAFLD) and its connection with insulin resistance, dyslipidemia, atherosclerosis and coronary heart disease. *Nutrients* 2013; **5**: 1544-1560 [PMID: 23666091 DOI: 10.3390/nu5051544]
- 5 **Wong RJ**, Ahmed A. Obesity and non-alcoholic fatty liver disease: Disparate associations among Asian populations. *World J Hepatol* 2014; **6**: 263-273 [PMID: 24868320 DOI: 10.4254/wjh.v6.i5.263]
- 6 **Bettermann K**, Hohensee T, Haybaeck J. Steatosis and steatohepatitis: complex disorders. *Int J Mol Sci* 2014; **15**: 9924-9944 [PMID: 24897026 DOI: 10.3390/ijms15069924]
- 7 **Chalasani N**, Younossi Z, Lavine JE, Diehl AM, Brunt EM, Cusi K, Charlton M, Sanyal AJ. The diagnosis and management of non-alcoholic fatty liver disease: practice Guideline by the American Association for the Study of Liver Diseases, American College of Gastroenterology, and the American Gastroenterological Association. *Hepatology* 2012; **55**: 2005-2023 [PMID: 22488764 DOI: 10.1002/hep.25762]
- 8 **Nascimbeni F**, Pais R, Bellentani S, Day CP, Ratziu V, Loria P, Lonardo A. From NAFLD in clinical practice to answers from guidelines. *J Hepatol* 2013; **59**: 859-871 [PMID: 23751754 DOI: 10.1016/j.jhep.2013.05.044]
- 9 **Mattar R**, de Campos Mazo DF, Carrilho FJ. Lactose intolerance: diagnosis, genetic, and clinical factors. *Clin Exp Gastroenterol* 2012; **5**: 113-121 [PMID: 22826639 DOI: 10.2147/CEG.S32368]
- 10 **Ingram CJ**, Mulcare CA, Itan Y, Thomas MG, Swallow DM. Lactose digestion and the evolutionary genetics of lactase persistence. *Hum Genet* 2009; **124**: 579-591 [PMID: 19034520 DOI: 10.1007/s00439-008-0593-6]
- 11 **Enattah NS**, Sahi T, Savilahti E, Terwilliger JD, Peltonen L, Järvelä I. Identification of a variant associated with adult-type hypolactasia. *Nat Genet* 2002; **30**: 233-237 [PMID: 11788828 DOI: 10.1038/ng826]
- 12 **Mattar R**, Monteiro MS, Villares CA, Santos AF, Silva JM, Carrilho FJ. Frequency of LCT -13910C & gt; T single nucleotide polymorphism associated with adult-type hypolactasia/lactase persistence among Brazilians of different ethnic groups. *Nutr J* 2009; **8**: 46 [PMID: 19799794 DOI: 10.1186/1475-2891-8-46]
- 13 **Almon R**, Engfeldt P, Tysk C, Sjöström M, Nilsson TK. Prevalence and trends in adult-type hypolactasia in different age cohorts in Central Sweden diagnosed by genotyping for the adult-type hypolactasia-linked LCT -13910C & gt; T mutation. *Scand J Gastroenterol* 2007; **42**: 165-170 [PMID: 17327935 DOI: 10.1080/00365520600825257]
- 14 **Khabarova Y**, Torniaainen ST, Nurmi HA, Järvelä IE, Isokoski MK, Mattila KJ. Prevalence of lactase persistent/non-persistent genotypes and milk consumption in a young population in north-west Russia. *World J Gastroenterol* 2009; **15**: 1849-1853 [PMID: 19370782 DOI: 10.3748/wjg.15.1849]
- 15 **Corella D**, Arregui M, Coltell O, Portolés O, Guillem-Sáiz P, Carrasco P, Sorlí JV, Ortega-Azorín C, González JI, Ordovás JM. Association of the LCT-13910C & gt; T polymorphism with obesity and its modulation by dairy products in a Mediterranean population. *Obesity* (Silver Spring) 2011; **19**: 1707-1714 [PMID: 21193851]

- DOI: 10.1038/oby.2010.320]
- 16 **Kettunen J**, Silander K, Saarela O, Amin N, Müller M, Timpson N, Surakka I, Ripatti S, Laitinen J, Hartikainen AL, Pouta A, Lahermo P, Anttila V, Männistö S, Jula A, Virtamo J, Salomaa V, Lehtimäki T, Raitakari O, Gieger C, Wichmann EH, Van Duijn CM, Smith GD, McCarthy MI, Järvelin MR, Perola M, Peltonen L. European lactase persistence genotype shows evidence of association with increase in body mass index. *Hum Mol Genet* 2010; **19**: 1129-1136 [PMID: 20015952 DOI: 10.1093/hmg/ddp561]
 - 17 **Almon R**, Alvarez-Leon EE, Engfeldt P, Serra-Majem L, Magnuson A, Nilsson TK. Associations between lactase persistence and the metabolic syndrome in a cross-sectional study in the Canary Islands. *Eur J Nutr* 2010; **49**: 141-146 [PMID: 19844753 DOI: 10.1007/s00394-009-0058-2]
 - 18 **Aune D**, Norat T, Romundstad P, Vatten LJ. Dairy products and the risk of type 2 diabetes: a systematic review and dose-response meta-analysis of cohort studies. *Am J Clin Nutr* 2013; **98**: 1066-1083 [PMID: 23945722 DOI: 10.3945/ajcn.113.059030]
 - 19 **Kalergis M**, Leung Yinko SS, Nedelcu R. Dairy products and prevention of type 2 diabetes: implications for research and practice. *Front Endocrinol (Lausanne)* 2013; **4**: 90 [PMID: 23888154 DOI: 10.3389/fendo.2013.00090]
 - 20 **Hirahatake KM**, Slavin JL, Maki KC, Adams SH. Associations between dairy foods, diabetes, and metabolic health: potential mechanisms and future directions. *Metabolism* 2014; **63**: 618-627 [PMID: 24636056 DOI: 10.1016/j.metabol.2014.02.009]
 - 21 **Nicklas TA**, Qu H, Hughes SO, He M, Wagner SE, Foushee HR, Shewchuk RM. Self-perceived lactose intolerance results in lower intakes of calcium and dairy foods and is associated with hypertension and diabetes in adults. *Am J Clin Nutr* 2011; **94**: 191-198 [PMID: 21525197 DOI: 10.3945/ajcn.110.009860]
 - 22 **Samara A**, Herbeth B, Ndiaye NC, Fumeron F, Billod S, Siest G, Visvikis-Siest S. Dairy product consumption, calcium intakes, and metabolic syndrome-related factors over 5 years in the STANISLAS study. *Nutrition* 2013; **29**: 519-524 [PMID: 23274089 DOI: 10.1016/j.nut.2012.08.013]
 - 23 **Pfeuffer M**, Schrezenmeier J. Milk and the metabolic syndrome. *Obes Rev* 2007; **8**: 109-118 [PMID: 17300277 DOI: 10.1111/j.1467-789X.2006.00265.x]
 - 24 **Grundy SM**, Brewer HB, Cleeman JI, Smith SC, Lenfant C. Definition of metabolic syndrome: Report of the National Heart, Lung, and Blood Institute/American Heart Association conference on scientific issues related to definition. *Circulation* 2004; **109**: 433-438 [PMID: 14744958 DOI: 10.1161/01.CIR.0000111245.75752.C6]
 - 25 **Kleiner DE**, Brunt EM, Van Natta M, Behling C, Contos MJ, Cummings OW, Ferrell LD, Liu YC, Torbenson MS, Unalp-Arida A, Yeh M, McCullough AJ, Sanyal AJ. Design and validation of a histological scoring system for nonalcoholic fatty liver disease. *Hepatology* 2005; **41**: 1313-1321 [PMID: 15915461 DOI: 10.1002/hep.20701]
 - 26 **Matthews DR**, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 1985; **28**: 412-419 [PMID: 3899825]
 - 27 **Vasques AC**, Rosado LE, Cássia GAlfenas Rd, Geloneze B. [Critical analysis on the use of the homeostasis model assessment (HOMA) indexes in the evaluation of the insulin resistance and the pancreatic beta cells functional capacity]. *Arq Bras Endocrinol Metabol* 2008; **52**: 32-39 [PMID: 18345394 DOI: 10.1590/S0004-27302008000100006]
 - 28 **Madeira IR**, Carvalho CN, Gazolla FM, de Matos HJ, Borges MA, Bordallo MA. [Cut-off point for Homeostatic Model Assessment for Insulin Resistance (HOMA-IR) index established from Receiver Operating Characteristic (ROC) curve in the detection of metabolic syndrome in overweight pre-pubertal children]. *Arq Bras Endocrinol Metabol* 2008; **52**: 1466-1473 [PMID: 19197455 DOI: 10.1590/S0004-27302008000900010]
 - 29 **Miller SA**, Dykes DD, Polesky HF. A simple salting out procedure for extracting DNA from human nucleated cells. *Nucleic Acids Res* 1988; **16**: 1215 [PMID: 3344216]
 - 30 **Mulcare CA**, Weale ME, Jones AL, Connell B, Zeitlyn D, Tarekegn A, Swallow DM, Bradman N, Thomas MG. The T allele of a single-nucleotide polymorphism 13.9 kb upstream of the lactase gene (LCT) (C-13.9kbT) does not predict or cause the lactase-persistence phenotype in Africans. *Am J Hum Genet* 2004; **74**: 1102-1110 [PMID: 15106124 DOI: 10.1086/421050]
 - 31 **Mattar R**, Monteiro Mdo S, Villares CA, dos Santos AF, Carrilho FJ. Single nucleotide polymorphism C/T(-13910), located upstream of the lactase gene, associated with adult-type hypolactasia: validation for clinical practice. *Clin Biochem* 2008; **41**: 628-630 [PMID: 18237552 DOI: 10.1016/j.clinbiochem.2008.01.006]
 - 32 **Büning C**, Genschel J, Jurga J, Fiedler T, Voderholzer W, Fiedler EM, Worm M, Weltrich R, Lochs H, Schmidt H, Ockenga J. Introducing genetic testing for adult-type hypolactasia. *Digestion* 2005; **71**: 245-250 [PMID: 16024930 DOI: 10.1159/000087050]
 - 33 **Hothorn T**, Hornik K, Zeileis A. Unbiased Recursive Partitioning: A Conditional Inference Framework. *J Comput Graph Stat* 2006; **15**: 651-674 [DOI: 10.1198/106186006X133933]
 - 34 **R Core Team**. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. 2014. Available from: URL: <http://www.R-project.org/>
 - 35 **Upton J**, George P. The prevalence of lactose intolerance (adult hypolactasia) in a randomly selected New Zealand population. *N Z Med J* 2010; **123**: 123 [PMID: 20173814]
 - 36 **Nagy D**, Tömöry G, Csányi B, Bogácsi-Szabó E, Czibula Á, Priskin K, Bede O, Bartosiewicz L, Downes CS, Raskó I. Comparison of lactase persistence polymorphism in ancient and present-day Hungarian populations. *Am J Phys Anthropol* 2011; **145**: 262-269 [PMID: 21365615 DOI: 10.1002/ajpa.21490]
 - 37 **Enattah NS**, Trudeau A, Pimenoff V, Maiuri L, Auricchio S, Greco L, Rossi M, Lentze M, Seo JK, Rahgozar S, Khalil I, Alifrangis M, Natah S, Groop L, Shaat N, Kozlov A, Verschubskaya G, Comas D, Bulayeva K, Mehdi SQ, Terwilliger JD, Sahi T, Savilahti E, Perola M, Sajantila A, Järvelä I, Peltonen L. Evidence of still-ongoing convergence evolution of the lactase persistence T-13910 alleles in humans. *Am J Hum Genet* 2007; **81**: 615-625 [PMID: 17701907 DOI: 10.1086/520705]
 - 38 **Friedrich DC**, de Andrade FM, Fiegenbaum M, de Almeida S, Mattevi VS, Callegari-Jacques SM, Hutz MH. The lactase persistence genotype is a protective factor for the metabolic syndrome. *Genet Mol Biol* 2014; **37**: 611-615 [PMID: 25505833 DOI: 10.1590/S1415-4752014005000012]
 - 39 **Lamri A**, Poli A, Emery N, Bellili N, Velho G, Lantieri O, Balkau B, Marre M, Fumeron F. The lactase persistence genotype is associated with body mass index and dairy consumption in the D.E.S.I.R. study. *Metabolism* 2013; **62**: 1323-1329 [PMID: 23647908 DOI: 10.1016/j.metabol.2013.04.006]
 - 40 **Enattah NS**, Forsblom C, Rasinperä H, Tuomi T, Groop PH, Järvelä I. The genetic variant of lactase persistence C (-13910) T as a risk factor for type I and II diabetes in the Finnish population. *Eur J Clin Nutr* 2004; **58**: 1319-1322 [PMID: 15054412 DOI: 10.1038/sj.ejcn.1601971]
 - 41 **Vassilatou E**. Nonalcoholic fatty liver disease and polycystic ovary syndrome. *World J Gastroenterol* 2014; **20**: 8351-8363 [PMID: 25024594 DOI: 10.3748/wjg.v20.i26.8351]
 - 42 **Lerchbaum E**, Giuliani A, Gruber HJ, Pieber TR, Obermayer-Pietsch B. Adult-type hypolactasia and calcium intake in polycystic ovary syndrome. *Clin Endocrinol (Oxf)* 2012; **77**: 834-843 [PMID: 22233423 DOI: 10.1111/j.1365-2265.2012.04334.x]
 - 43 **Högenauer C**, Hammer HF, Mellitzer K, Renner W, Krejs GJ, Toplak H. Evaluation of a new DNA test compared with the lactose hydrogen breath test for the diagnosis of lactase non-persistence. *Eur J Gastroenterol Hepatol* 2005; **17**: 371-376 [PMID: 15716664]
 - 44 **Calton EK**, James AP, Pannu PK, Soares MJ. Certain dietary patterns are beneficial for the metabolic syndrome: reviewing the evidence. *Nutr Res* 2014; **34**: 559-568 [PMID: 25150114 DOI: 10.1016/j.nutres.2014.06.012]
 - 45 **Shin H**, Yoon YS, Lee Y, Kim CI, Oh SW. Dairy product intake is inversely associated with metabolic syndrome in Korean adults:

- Anseong and Ansan cohort of the Korean Genome and Epidemiology Study. *J Korean Med Sci* 2013; **28**: 1482-1488 [PMID: 24133353 DOI: 10.3346/jkms.2013.28.10.1482]
- 46 **Martins ML**, Kac G, Silva RA, Bettiol H, Barbieri MA, Cardoso VC, Silva AA. Dairy consumption is associated with a lower prevalence of metabolic syndrome among young adults from Ribeirão Preto, Brazil. *Nutrition* 2015; **31**: 716-721 [PMID: 25837218 DOI: 10.1016/j.nut.2014.12.017]
- 47 **Crichton GE**, Alkerwi A. Dairy food intake is positively associated with cardiovascular health: findings from Observation of Cardiovascular Risk Factors in Luxembourg study. *Nutr Res* 2014; **34**: 1036-1044 [PMID: 25476191 DOI: 10.1016/j.nutres.2014.04.002]
- 48 **Da Silva MS**, Rudkowska I. Dairy products on metabolic health: current research and clinical implications. *Maturitas* 2014; **77**: 221-228 [PMID: 24445013 DOI: 10.1016/j.maturitas.2013.12.007]
- 49 **Visioli F**, Strata A. Milk, dairy products, and their functional effects in humans: a narrative review of recent evidence. *Adv Nutr* 2014; **5**: 131-143 [PMID: 24618755 DOI: 10.3945/an.113.005025]
- 50 **Parodi PW**. Cooperative action of bioactive components in milk fat with PPARs may explain its anti-diabetogenic properties. *Med Hypotheses* 2016; **89**: 1-7 [PMID: 26968898 DOI: 10.1016/j.mehy.2015.12.028]
- 51 **Belury MA**, Moya-Camarena SY, Lu M, Shi L, Leesnitzer LA, Blanchard SG. Conjugated linoleic acid is an activator and ligand for peroxisome proliferator-activated receptor-gamma (PPAR γ). *Nutr Res* 2002; **2002**: 817-824 [DOI: 10.1016/S0271-5317(02)00393-7]
- 52 **Moya-Camarena SY**, Van den Heuvel JP, Belury MA. Conjugated linoleic acid activates peroxisome proliferator-activated receptor alpha and beta subtypes but does not induce hepatic peroxisome proliferation in Sprague-Dawley rats. *Biochim Biophys Acta* 1999; **1436**: 331-342 [PMID: 9989264]
- 53 **Wang Y**, Jacome-Sosa MM, Ruth MR, Lu Y, Shen J, Reaney MJ, Scott SL, Dugan ME, Anderson HD, Field CJ, Proctor SD, Vine DF. The intestinal bioavailability of vaccenic acid and activation of peroxisome proliferator-activated receptor- α and - γ in a rodent model of dyslipidemia and the metabolic syndrome. *Mol Nutr Food Res* 2012; **56**: 1234-1246 [PMID: 22714958 DOI: 10.1002/mnfr.201100517]
- 54 **Jacome-Sosa MM**, Borthwick F, Mangat R, Uwiera R, Reaney MJ, Shen J, Quiroga AD, Jacobs RL, Lehner R, Proctor SD, Nelson RC. Diets enriched in trans-11 vaccenic acid alleviate ectopic lipid accumulation in a rat model of NAFLD and metabolic syndrome. *J Nutr Biochem* 2014; **25**: 692-701 [PMID: 24775093 DOI: 10.1016/j.jnutbio.2014.02.011]
- 55 **Tilg H**, Moschen AR. Evolution of inflammation in nonalcoholic fatty liver disease: the multiple parallel hits hypothesis. *Hepatology* 2010; **52**: 1836-1846 [PMID: 21038418 DOI: 10.1002/hep.24001]
- 56 **Peverill W**, Powell LW, Skoien R. Evolving concepts in the pathogenesis of NASH: beyond steatosis and inflammation. *Int J Mol Sci* 2014; **15**: 8591-8638 [PMID: 24830559 DOI: 10.3390/ijms15058591]
- 57 **Paolella G**, Mandato C, Pierri L, Poeta M, Di Stasi M, Vajro P. Gut-liver axis and probiotics: their role in non-alcoholic fatty liver disease. *World J Gastroenterol* 2014; **20**: 15518-15531 [PMID: 25400436 DOI: 10.3748/wjg.v20.i42.15518]
- 58 **Miura K**, Ohnishi H. Role of gut microbiota and Toll-like receptors in nonalcoholic fatty liver disease. *World J Gastroenterol* 2014; **20**: 7381-7391 [PMID: 24966608 DOI: 10.3748/wjg.v20.i23.7381]
- 59 **Imajo K**, Yoneda M, Ogawa Y, Wada K, Nakajima A. Microbiota and nonalcoholic steatohepatitis. *Semin Immunopathol* 2014; **36**: 115-132 [PMID: 24337650 DOI: 10.1007/s00281-013-0404-6]
- 60 **Bäckhed F**, Ley RE, Sonnenburg JL, Peterson DA, Gordon JI. Host-bacterial mutualism in the human intestine. *Science* 2005; **307**: 1915-1920 [PMID: 15790844 DOI: 10.1126/science.1104816]
- 61 **Mouzaki M**, Comelli EM, Arendt BM, Bonengel J, Fung SK, Fischer SE, McGilvray ID, Allard JP. Intestinal microbiota in patients with nonalcoholic fatty liver disease. *Hepatology* 2013; **58**: 120-127 [PMID: 23401313 DOI: 10.1002/hep.26319]
- 62 **Turnbaugh PJ**, Hamady M, Yatsunenko T, Cantarel BL, Duncan A, Ley RE, Sogin ML, Jones WJ, Roe BA, Affourtit JP, Egholm M, Henrissat B, Heath AC, Knight R, Gordon JI. A core gut microbiome in obese and lean twins. *Nature* 2009; **457**: 480-484 [PMID: 19043404 DOI: 10.1038/nature07540]
- 63 **Maga EA**, Desai PT, Weimer BC, Dao N, Kültz D, Murray JD. Consumption of lysozyme-rich milk can alter microbial fecal populations. *Appl Environ Microbiol* 2012; **78**: 6153-6160 [PMID: 22752159 DOI: 10.1128/AEM.00956-12]
- 64 **Donovan SM**, Wang M, Li M, Friedberg I, Schwartz SL, Chapkin RS. Host-microbe interactions in the neonatal intestine: role of human milk oligosaccharides. *Adv Nutr* 2012; **3**: 450S-455S [PMID: 22585924 DOI: 10.3945/an.112.001859]

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