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**Goblet cell carcinoids of the appendix: Tumor biology, mutations and management strategies**

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**Abstract**

Malignant neoplasms of the appendix are rare and represent less than 1% of gastrointestinal cancers. Goblet cell carcinoids (GCC) tumors are a distinctive group of heterogeneous appendiceal neoplasm that exhibit unique clinical and pathologic features. This review focuses on the current diagnostic procedures, pathogenesis, possible signaling mechanisms and treatment options for GCC. Perspectives for future research are discussed. The tumor likely arises from pluripotent intestinal epithelial crypt base stem cells. Previous findings of Notch signaling as a tumor suppressor in Neuroendocrine tumors may have a similar role in this tumor too. Loss of Notch signaling may be the driver mutation with other successive downstream mutations likely favors them into progressing and behavior similar to poorly differentiated adenocarcinoma with minimal neuroendocrine differentiation. A multidisciplinary approach is suggested for optimal outcomes. Surgery remains the main treatment modality. Simple appendectomy may be sufficient in early stages while right hemicolectomy is recommended for advanced tumors. Cytoreductive surgery with heated intraperitoneal chemotherapy may improve survival in a select few with metastatic peritoneal disease. These tumors have an unpredictable behavior even in early stages and local recurrence and delayed metastases may be seen. Lifelong surveillance is warranted.

**Key words:** Goblet cell carcinoid of the appendix; Immunomarkers; Intestinal stem cells; Notch-1 signaling; Math-1 signaling; Disease management

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**Core tip:**Goblet cell carcinoids tumors are a distinctive group of heterogeneous appendiceal neoplasm that exhibit unique clinical and pathologic features. The pathogenesis is unclear however the tumor likely arises from pluripotent intestinal epithelial crypt base stem cells. Loss of Notch signaling may be the driver mutation with other successive downstream mutations likely favors them into progressing and behavior similar to poorly differentiated adenocarcinoma with minimal neuroendocrine differentiation. Surgery remains the main treatment modality. We discuss the clinical implications of this cancer focusing on the tumor biology, mutations, signaling mechanisms and management.

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**INTRODUCTION**

Malignant neoplasms of the appendix are rare and represent less than 1% of gastrointestinal cancers. Studies evaluating data for appendiceal malignancies from seer database between 1973-2001 showed the age-adjusted incidence of cancer of appendix was 0.12 cases per 1 million people per year[1,2]. They are further classified into colonic type adenocarcinoma, mucinous tumors, signet ring cell tumors, carcinoids [neuroendocrine tumors (NETs)] and goblet cell carcinoids (GCC). Overall five-year survival is highest for appendiceal carcinoid (83%) and lowest for signet ring cancers (18%)[1,2]. This review focuses on GCC of the appendix. The current diagnostic procedures, pathogenesis, signaling mechanisms and possible mutations are presented. Treatment options for this neoplasm are defined and summarized, although evidence-based data are lacking. Surgery remains the treatment mainstay.

GCC tumors are a distinctive group of heterogeneous appendiceal neoplasm that exhibit unique clinical and pathologic features. These hybrid tumors have both glandular and neuroendocrine morphology and are designated with various terminologies: Adenocarcinoids, crypt cell carcinoma, mixed carcinoid -adenocarcinoma and amphicrine tumors. These various terminologies do not reflect consistent morphology, biologic behavior or accepted criteria for the diagnosis. GCC was first described by Gagné*et al*[3] in 1969, Subbuswamy *et al*[4] subsequently coined the term GCC in 1974. Warner *et al*[5] in 1979 suggested a probable origin from crypt based stem cell. Isaacson in 1981 demonstrated presence of IgA, lysozyme in GCC suggestive of possible role of Paneth cells in this tumor[3-6].

GCC exhibits clear distinction when compared to appendiceal NETs or primary adenocarcinoma in terms of demographics, biology and clinical aggressive behavior. The prognoses of GCC lays intermediate between appendiceal NETs and primary appendiceal adenocarcinoma[1,2].

**CLINICAL PRESENTATION**

GCC are diagnosed in less than 1% of appendectomy specimens[7,8]. Most commonly, patients present with abdominal pain and acute appendicitis (> 50%). They are most often diagnosed incidentally during appendectomy or ileocecal resection and confirmed by the pathologist in post-surgical specimens. About 27% of patients may present with perforated appendicitis[9]. Patients may also present sub-acutely in advanced stages with vague abdominal pain and mass[7,8]. Common in Caucasians, there is equal distribution in male and females with the average age of diagnosis in the fifth decade[1,2]. Up to 50% of patients present with metastatic disease[8,10-12]. Similar to carcinoids a significant number of these patients may harbor a second primary malignancy[12-14].

Morphologically the tumor circumferentially involves the appendiceal wall with transmural extension. Submucosal involvement with mucosal sparing is noted. Most tumors are generally > 2 cm in size. The native appendiceal epithelium may show fibrous obliteration without adenomatous or dysplastic changes[8,15].

GCC display a wide range of histologic patterns both in primary and metastatic sites. Common to all GCC is the presence of mucin containing goblet shaped epithelial cells arranged in clusters in the lamina propria and submucosa. These cells stain positive for mucicarmine, periodic acid–Schiff and alcian blue stains suggestive of goblet cell mucin. Extracellular pools of mucin may also be present. Also seen are cells, which demonstrate focal, inconsistent scattered immunoreactivity for neuroendocrine markers (*i.e.*, chromogranin, synaptophysin)[8,16].

Since GCCs show a submucosal growth pattern, it has a tendency to spread to surrounding bowel. The most common metastatic sites include direct extension into the right colon and ileum, followed by spread to lymph nodes, peritoneum and omentum. The ovaries are common site of metastases in women presenting as Krukenberg tumor. Up to 80 % of women with stage 4 disease present with ovarian metastases[8,12].Solid organ metastases to liver, lung, bones are uncommon[8]. A previous study reports the rate of metastases to lymph nodes increases with the T stage of the tumor T2 (0%), T3 (13%), T4 (60%)[9].

Metastatic lesions from GCC are more aggressive tumors and often show poorly differentiated signet ring cell or undifferentiated adenocarcinoma morphology with minimal neuroendocrine features. They may not share features of primary tumor and carry poorer prognosis. Metastatic tumors usually do not stain for chromogranin A or synaptophysin and stain heavily for mucin, suggestive of degeneration into signet ring cell morphology. The population of endocrine cells and Paneth cells seem to decrease in metastatic lesions. Yan *et al*[17] in their series of 26 patients reported that nine patients (35%) with metastatic GCC failed to stain for neuroendocrine marker. The explanation for this finding remains elusive[17-21].

**CLASSIFICATION**

Currently multiple classification systems exist to describe GCC.

The 2010 World Health Organization (WHO) classification for tumors of the appendix, classifies GCC under the category of neuroendocrine neoplasms based on differentiation and histological grading. Grade refers to the proliferative activity measured with mitotic counts and Ki-67 index. They are further sub-classified as low grade: G1 (< 2 mitosis/10 HPF and ≤ 2% Ki index), intermediate grade: G2 (2-20 mitosis/10 HPF, 3%-20% Ki index) and high grade: G3 (> 20 mitosis/10 HPF, > 20% Ki Index). Differentiation refers to resemblance of tumor cells to the normal neuroendocrine cells. Carcinoids (well differentiated neuroendocrine neoplasm) generally belong to G1 and G2 categories while G3 is considered as a neuroendocrine carcinoma (NEC). Goblet cell tumors are subtyped under mixed adeno-NEC (MANEC). To qualify for this definition at least 30% of tumor should have gland forming epithelial and neuroendocrine components[22].

The 2010 American joint commission on cancer (TNM classification) stages these tumors based on the tumor size, nodal status and metastatic disease into stages (I-IV). Stage I (T1, N0, M0), stage II (T2/T3, N0, M0), stage III (any T/N1, M0), and stage IV (any T /any N/M1)[23].

Tang *et al*[8] in 2008, proposed a system of classification specific for GCC of appendix based on histologic features of the tumor at the primary site. They include the arrangement of the goblet cells, degree of atypia and desmoplasia to label these tumors into three groups. Typical GCC (group A); adenocarcinoma ex GCC, signet ring cell (group B); adenocarcinoma ex GCC, poorly differentiated (group C). Almost all patients in group C presented in advanced stages with wide metastases. This suggests that GCCs display a spectrum of histologic features with the potential to progress to an aggressive adenocarcinoma phenotype[8].

These multiple pathologic definitions and differing terminologies have led to inconsistent reporting and difficult to characterize this disease.

MANEC per the 2010 WHO classification are tumors harboring both epithelial and neuroendocrine components. However based on this definition it requires tumors to have at least 30% representation of each component. In general this is not true for all GCC tumors. Further advanced stages of GCC losses its neuroendocrine differentiation and acquires an aggressive signet ring cell or poorly differentiated morphology.

This tumor may need further investigations to better clarify and define their heterogeneous, molecular profile and classification.

**IMMUNOCHEMISTRY AND MUTATIONAL FINDINGS**

GCC specimens demonstrate focal, inconsistent immunoreactivity for neuroendocrine markers. In contrast diffuse staining is observed in most classic carcinoids of the appendix. Common positive markers are synaptophysin, chromogranin A, serotonin, neuron specific enolase, pancreatic polypeptide. Ultrastructural immuno- histiochemistry staining has shown tumor nests resembling normal crypts in the submucosa. Separate goblet cell and neuroendocrine cells are often located in close proximity to each other[24] (Table 1).

In addition GCC do not exhibit mutations as conventional colorectal adenocarcinoma. These tumors are negative for KRAS, SMAD4 and BRAF mutations[25].They show negative staining for nuclear β-catenin and for P53. MUC2 expression is preserved[8]. Normal colorectal and appendicular epithelium expresses MUC2 only. GCC show strong carcinoembryonic antigen, caudal type homeobox transcription factor 2, cytokeratin 7 (CK7), CK20 expression suggestive of intestinal epithelial origin while these markers remain negative in classic carcinoids[26]. A single study showed allelic loss in chromosomes 11q, 16q, and 18q in GCC similar to ileal carcinoids[25].

The proliferative Ki67 index remains low in typical GCC but rises with advanced stages (Tangs group C). The significance remains unknown as some groups have shown worsening survival rates with rising Ki67 index[19,27] while other have shown no correlation[9,28,29]. Positive staining for p53 and MUC1 with loss of MUC2 expression is suggestive of transformation to adenocarcinoma phenotype similar to colorectal adenocarcinoma[8]. This also correlates with the rising Ki67 index as reported with Tangs *et al*[8]’s classification.

In general patients with GCC do not present with carcinoid syndrome and urinary 5HIAA levels are within normal range[16]. Unlike classic midgut carcinoids, serum chromogranin A levels are normal and have no value in detecting and monitoring GCC. Somatostatin expression is sparse and erratic and therefore functional scans such as 111-Indium pentetreotide scintigraphy (Octreoscan) and Gallium 68-octreotide positron emission tomography (PET) scans are usually normal in patients with GCC, and thus are of limited use[11,19]. Fluorodeoxyglucose PET scan may be useful in advanced disease to detect peritoneal metastatic disease[7,27,30].

GCC also express transcription factor Math-1 and HD5 (Defensins) a known marker for Paneth cells[26]. Math-1 is a basic helix-loop-helix transcription factor essential for development of the pluripotent stem cell towards secretory stem cell lineage and may play a role in pathogenesis of GCC.

**PATHOGENESIS**

The pathogenesis of GCC remains unclear. Unlike adenocarcinomas of the GI tract which arises through an adenoma–carcinoma sequence GCC is thought to arise from pluripotent intestinal epithelial crypt base stem cells[6,26].

An understanding of the embryological origin and signaling pathways associated with development of small bowel and appendix may provide clues and explain the origin and progression of GCC. The epithelial lining of small gut consists of a single layer of columnar cells. This differentiated epithelium arises from the crypts and projects up as villi into the lumen forming the absorptive lining of the gut (Figure 1). Villi begin to form by embryonic day 15 and crypt form by invagination of intervillus pockets at post-natal day 7[31,32]. The four main types of differentiated cells are absorptive enterocytes, goblet cells, neuroendocrine and Paneth cells. The crypt thus forms the proliferative stem cell compartment, where these cells originate, differentiate, amplify and move up into the villi akin to a system of conveyor belt in an assembly line[33]. Paneth cells which also originate from the crypt in an exception and migrates downward into the base of the crypt. These four cell types are the main differentiated cell types found in the epithelial lining of the small intestine.

How is the stem cell niche created and defined? What molecular signaling mechanisms keep the niche intact and regulated? Information on these fundamental questions comes from mouse studies. Advances in development and stem cell biology have also occurred to generate complex three dimensional human intestinal tissues *in vitro* through directed differentiation of human pluripotent stem cells. These human intestinal organoids called mini-gut have expanded our ability to study development, genetics, intestinal pathogens and metabolic disease and cancer[34]. Cheng and Leblond[35,36] in 1974 were first to characterize crypt based columnar cells as intestinal stem cells. Barker et al identified a marker Lgr5/GPR49, a leucine rich orphan G-protein coupled receptor that labels these stem cells[37]. Tritiated radioactive thymidine labelling experiments have confirmed that these Lgr5 cells are the multipotent stem cells. Stem cell division occurs every 24 h and these cells are localized to the crypts. Subsequently cells migrate up from the crypt to the villus in 3-5 d[34,38]. These stem cells by itself are not terminally differentiated and can divide without a limit. The daughter cells have to choose between committing to terminal differentiation or remain as a stem cell. The rapidly dividing groups of cells derived from the crypt stem cells that have committed to differentiation are known as transit amplifying cells. As they migrate further up the crypt they amplify according to their prospective destined fate and differentiate as enterocyte, goblet cell, neuroendocrine and Paneth cell. They cease to divide further once they reach the neck of the crypt at the crypt villous junction[31,33].

Thus this slim, columnar crypt based Lgr5 positive stem cells along with the post mitotic Paneth cells form the stem cell niche through which begins the growth and renewal of all the differentiated cells of the small intestinal epithelium.

There are two major groups of cell signaling mechanisms which govern the crypt-villus axis (Figure 1). The first is an epithelial-epithelial cell communication. The key mediators of these mechanisms are Wnt, Notch, Eph-ephrin and Math 1 signaling pathways. Together and sequentially they are primarily responsible for maintaining the gut stem cells in a proliferative state, differentiation into secretory or absorptive lineage and establish boundaries between these clones of cells. Mutation in these critical pathways has been implicated with excessive uncontrolled, ectopic crypt formation, adenomas, excessive goblet cell or neuroendocrine cells and other risks for colorectal malignancies.

The second major group of cellular signaling comprises of epithelial-mesenchymal (EMT) communications and the mediators in these pathways consist of hedgehog, BMP and PDGAF signaling pathways. The essential function of EMT pathways is to maintain a proper spacing between one crypt and the next. They are negative regulators of crypt formation. Hedgehog signaling increases the expression of BMP in the mesenchyme which further represses the Wnt signaling. Noggin, a BMP inhibitor is expressed in the crypts to maintain unsuppressed Wnt activity in the crypt epithelium(Figure 1). Mutation in these pathways or inhibition of BMP signaling by overexpression of its inhibitors, Noggin or inactivation of its receptor BMPRIA lead to excessive and ectopic crypt formation as seen with juvenile polyposis syndrome due to BMP knock out mutations[39].

We will limit our discussion to the epithelial-epithelial signaling mechanisms which may hold clues to the pathogenesis of GCC.

The initial signaling mechanisms in a crypt-villus axis begin with Wnt and Notch pathways in the crypts (Figure 1). Both the development of small intestine and its homeostasis require canonical Wnt signaling. The Wnt pathways maintain the gut stem cell compartment. The Paneth cell which constitutes part of the stem cell niche generates Wnt signals that act over a short range and keep cells in the crypt in a proliferative state[31,33]. There exists a gradient in Wnt signaling which is highest in the crypt base and diminishes towards the crypt-villus junction (Figure 1). Wnt signaling further drives the expression of Notch pathway. Notch pathway through its ligands such as Delta and jagged and effectors such as Hes and NF-ĸB transcription factors mediate lateral inhibition within the Wnt activated cell population thus driving cells towards different fates. Delta expressing cells escape Notch activation (Wnt+, Notch-) and commit to secretory fate through the downstream Math-1 signaling pathways and exit into a committed fate[32,33,40]. Meanwhile (Wnt+/Notch +) cells continue to migrate up the crypt and divide, generating daughter cells and diversify till they lose Wnt activation as they move up the villus and differentiate as absorptive enterocytes. Inactivation of Notch pathway by deletion of Hes 1 or nonsense mutation of Delta ligands leads to excessive formation of goblet cells and neuroendocrine cells[32,33,40]. Further studies have shown that inhibiting transcription factors in the Notch pathways by deletion of RBP-jk or by use of γ-secretase inhibitors prevents proteolytic cleavage and the release of notch intracellular domain complex. This result in epithelial cells composed exclusive of goblet cells[41]. Conversely experiments have shown that increased Notch activity results in severe reduction of differentiated secretory cells, suggesting a tumor suppressor role of Notch signaling in neuroendocrine tumors[42].

In addition to Wnt-Notch signaling another significant pathway in the crypt-villus axis known to play a role in maintaining cellular boundaries, segregation and establish migratory path are the Eph-ephrin molecules (Figure 1). Expression of Eph B receptors and the Ephrin-B ligands within the intestine is regulated *via* the β-catenin-TCF transcription complex through the Wnt pathway. A definitive gradient exists with proliferative cells in the crypt expressing higher density of Eph-B receptors and progressively decreases at the crypt villus axis. The cells acquiring a differentiated fate switch off the expression of Eph-B and switch on the expression of Ephrin B ligands which progressively increases as they migrate up the axis. Paneth cells express only Eph receptors and therefore remain at the crypt base. In general cells expressing Eph receptors are repelled by contact with cells expressing ephrins on their surface. These mechanisms keep the cells segregated in their respective niches. Dysregulation of the eph-ephrin axis leads to cellular derangement with proliferating cells are not restricted to the bottom of the crypt and abnormally scattered along the crypt-villus axis[33]. Eph B mediated compartmentalization restricts the spreading of Eph B expressing tumor cells into ephrin B1-positive territories *in vitro* and *in vivo*. Loss of EphB-mediated compartmentalization may lead to invasiveness of the tumor cells[43].

All three secretory cell types derive from a precursor expressing Math-1 (mouse atonal homologue 1), also known as Atoh1, Hath-1 (humans) (Figure 2). As explained earlier Delta expressing cells escape Notch activation (Wnt+, Notch-) and commit to secretory fate through the downstream Math-1 signaling pathways[32,33,40].It is a basic helix-loop-helix transcription factor that is required for secretory cell lineage through downstream Neurogenin 3 for neuroendocrine cells, Gfil and Klf4 for goblet cells, β-catenin and sox9 for Paneth cells and subsequent cell cycle exit[31,32,44,45]. Mice deficient in Math1 lack goblet cells and the epithelial cells continue to maintain their proliferative state[32,40]. Overexpression of Math1 results in ectopic secretory cells[46]. The immunohistochemical expression of Math-1 in GCC suggests that this transcription factor is essential for normal development of the pluripotent stem cell towards secretory stem cell lineage and may play a role in its pathogenesis. Possible somewhere along its differentiation a mutation occurs with altered signaling pathways which causes excessive clones of goblet cells and neuroendocrine cells and may explain the hybrid nature of this tumor[18,26].

NETs including GCC appear to be heterogeneous group of tumors with varying signaling mechanisms and gene expressions in different tissue of origin. A number of questions remain to be answered. Is there a Notch signaling dysfunction or inhibition leading to loss of Hes regulated inhibition of Math-1? Studies have confirmed the potential oncogenic role of Notch signaling and its transcription factor in certain solid organ abdominal, lung, breast, and genitourinary, neural and hematological malignancies[47]. However Notch signaling also appears to have a tumor suppressor role in gastrointestinal, thyroid and pulmonary neuroendocrine tumors[42,48,49]. In another recent study of 31 ileal carcinoids, Notch signaling was uniformly absent in ileal neuroendocrine tumors suggestive of loss of tumor suppressive role[50]. Is the loss of Notch signaling, the driving mutation and occurs after the first stem cell division at the level of transit amplifying cells with subsequent progeny showing dysfunction? Could there be a concurrent Eph-ephrin pathway mutation along with loss of notch signaling, leading to loss of compartmentalization of cells and portending invasiveness[43]?

Why does metastatic GCC show minimal neuroendocrine expression and more of signet ring cell and poorly differentiated morphology? Are there further successive mutations downstream in the Math-1 signaling pathway? Are there subsequent epigenetic modifications, chromatin remodeling and inactivation of tumor suppressor genes which further amplify the carcinogenesis?

Further investigations at these levels are needed that may lead to our understanding of the pathogenesis of these tumors and may have therapeutic implications. Targeted therapies to activate Notch signaling with varying concentrations for metastatic GCC may have potential benefits. The origin of goblet cells carcinoid and its transformation from typical GCC and to advanced signet ring cell, poorly differentiated adenocarcinoma could be due to spontaneous, sporadic mutation in the mentioned crypt-villus architecture and or the surrounding mesenchyme and is yet to be successfully identified. Characterizing the levels of expression of Notch pathway components in tumor samples from patients with GCC could serve as a tumor marker. This reinforces the need to further investigate the presence of these mutations in larger cohorts and in institutions treating patients with GCC and appendiceal NETs.

**TREATMENT**

Most patients typically present with acute appendicitis and undergo appendectomy. The dilemma arises after GCC is diagnosed and confirmed, whether simple appendectomy is adequate or further oncologic resection is required (Figure 3). A multi-disciplinary evaluation is recommended for the optimal treatment. Both European and North American Neuroendocrine tumor societies guidelines recommend right hemicolectomy after appendectomy due to the high rate of metastases and its impact on prognosis[30,51]. However other authors have argued against right hemicolectomy in their series[8,13,52,53].In a meta-analysis evaluating 13 studies with 100 patients, the authors concluded no benefits of right hemicolectomy in all patients. Selective criteria were recommended[53]. In another recent retrospective analysis of a larger number of appendiceal NETs, GCC and signet ring cell adenocarcinoma from seer database showed a benefit of right hemicolectomy and statistically improved survivalonly for signet ringcell cancer when compared to appendectomy alone (*P* = 0.01). There was no significant difference in survival for typical NETs (*P* = 0.21) or GCC (*P* = 0.94) based on type of surgery[54]. Based on Tangs classification the histology of the tumor in the appendectomy specimens and not the size of the tumor should determine the extent of oncologic resection[8]. In patients who fulfill all the following criteria: Tumor less than 2 cm localized to appendix with negative margins, pT1 or pT2 tumors, and typical GCC histology group A (Tang *et al*[8] classification) tumors, an appendectomy alone may be sufficient as the definitive treatment[13]. Right hemicolectomy is recommended in tumors greater than two centimeters, locally advanced, positive margins, T3, T4 tumors and histology suggestive of group B, group C (Tangs classification) in the appendectomy specimens[8,11,12,55].

The impact of perforated appendicitis in patients with GCC remains unclear. In a meta-analysis of 18 cases of GCC diagnosed upon perforated appendicitis showed no impact on survival and prognosis[56]. In another retrospective series of 20 GCC patients with perforated appendicitis, a lower rate of peritoneal metastases was observed in the perforated group (15%) compared to the non-perforated group (42%) with no difference in peritoneal relapse between the two groups[9].

A complication of GCC of the appendix is their propensity to spread to the ovaries. GCC of the appendiceal origin express elevated MUC2 and MUC5AC. In contrast mucinous tumors arising from ovarian primaries express only MUC5AC[57]. This could be of benefit in differentiating the origin of these tumors in females with primary ovarian mucinous malignancy[58].In postmenopausal female patients with GCC prophylactic bilateral oophorectomy, although not evidence based should be considered[7,8,12]. In female patients with mucinous ovarian and pelvic malignancies an appendectomy should always be performed in staging laparotomy as these may represent metastatic GCC[12,19,57].

Adjuvant systemic chemotherapy is prescribed for stage III and stage IV diseases and disease recurrence. Due to rarity of GCC a randomized control trial cannot be accomplished. Data is available from scattered anecdotal reports and small series of GCC and therefore guidelines for choice of chemotherapy is lacking. Since metastatic GCC shows clinical and histological resemblance to colorectal adenocarcinoma and not metastatic carcinoids the choice of adjuvant therapy in GCC is similar to colorectal adenocarcinoma. 5-fluorouracil (5-FU) and leucovorin based FOLFOX (5-FU, leucovorin, oxaliplatin) and FOLFIRI (5-FU, folic acid, irinotecan) chemotherapy are standard regimens recommended[11,30].

With locally advanced or recurrent peritoneal disease, cytoreductive surgery with hyperthermic intraperitoneal mitomycin and systemic chemotherapy (CRS+ HIPEC) may improve median survival[11,12,17,59,60]. In a recent study of 45 patients with GCC and peritoneal metastases who received CRS+ HIPEC, the therapy was successfully completed in 71% of patients and 3 years, overall survival (OS) was 63.4 %[60]. Another study on 26 patients report median survival of 51 mo and an overall five-year survival of 43%[17]. However a recent retrospective study on 25 patients who received CRS plus HIPEC therapy reports no reduction in relapse rates or improvement in disease free survival in either stage I and II compared to stage III and IV[9].

The other treatment options generally available for metastatic carcinoids such as interferon, somatostatin analogues (octreotide), targeted agents such as everolimus and sunitinib and radionuclide targeted therapy is not useful for metastatic GCC due to the absence of adequate uptake on Octreoscan or Gallium 68 PET scan and no confirmed mechanistic target of rapamycin or vascular endothelial growth factor pathway dysregulation.

**PROGNOSIS**

The overall disease specific survival for all GCC subtypes is 40%-80% depending on different series[7-9,11,12,54]. The five-year survival for localized, regional and distant metastatic disease based on Tangs classification of group A, B, C are 100%, 36% and 0% respectively. This correlates with the AJCC (TNM) staging system where reported five-year survival with stage I (100%), stage II (76%), stage III (22%), stage IV (14%) respectively.

**CONCLUSION**

GCC are a separate entity from carcinoids and adenocarcinoma. The pathogenesis is unclear however the tumor likely arises from pluripotent intestinal epithelial crypt base stem cells. Successive mutations likely favor them into progressing and behavior similar to poorly differentiated adenocarcinoma with minimal neuroendocrine differentiation. Metastatic lesions differ from the primary appendiceal site in terms of histology and tumor aggressiveness. A multidisciplinary approach is suggested for optimal outcomes. Surgery remains the main treatment modality. Due to its heterogeneity, this tumor should not be classified according to a single system and a combination of size of the tumor (T classification), grade and mitotic index (WHO classification) and arrangement of the goblet cells, degree of atypia and desmoplasia (Tang *et al*[8]’s histopathologic classification) should dictate further definitive therapy. Simple appendectomy may be sufficient in early stages while right hemicolectomy is recommended for advanced tumors. CRC with HIPEC may improve survival in a select few with metastatic peritoneal disease. These tumors have an unpredictable behavior even in early stages and local recurrence and delayed metastases may be frequently seen. Therefore lifelong surveillance is warranted.

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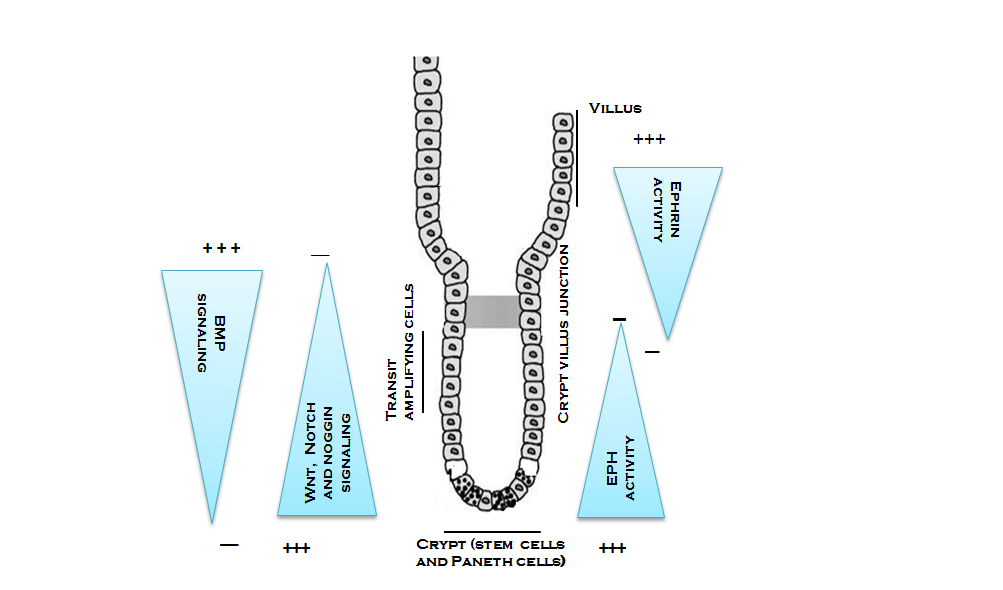
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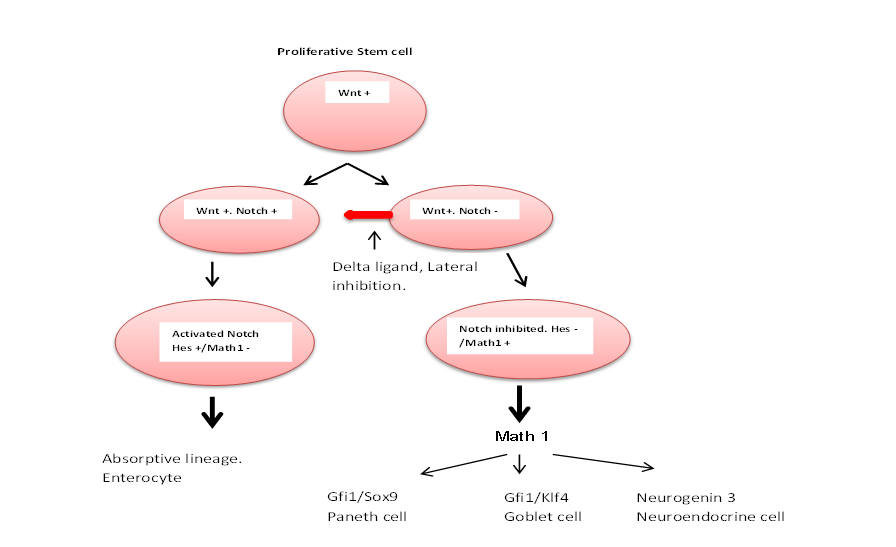
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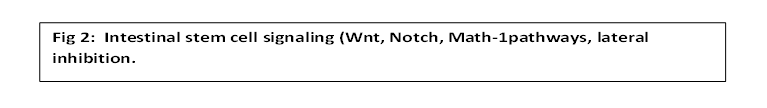
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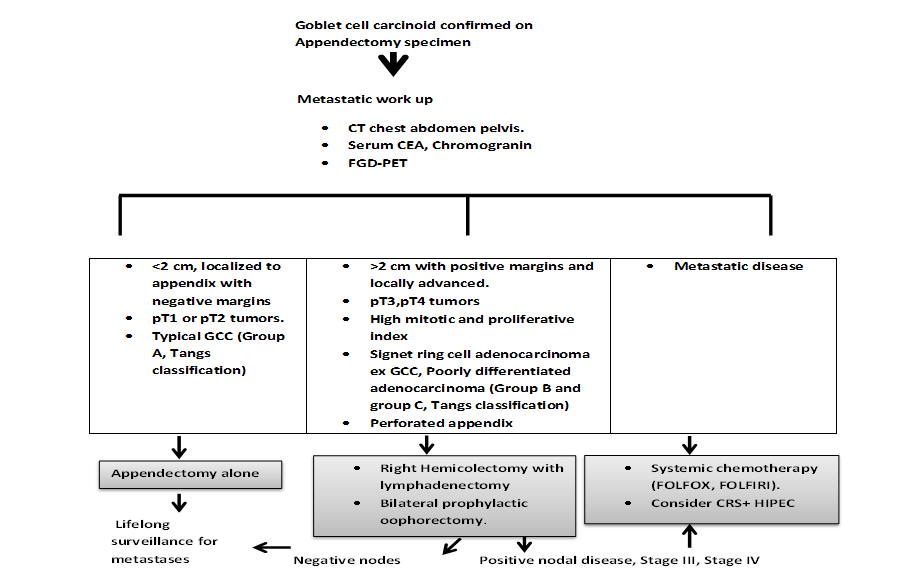


**Figure 1 Schematic diagram of crypt-villus axis.** The initial signaling mechanisms in a crypt-villus axis begin with Wnt and Notch pathways. The Wnt pathways maintain the gut stem cell compartment. The Paneth cell which constitutes part of the stem cell niche generates Wnt signals that act over a short range and keep cells in the crypt in a proliferative state. There exists a gradient in Wnt, notch and noggin signaling which is highest in the crypt base and diminishes towards the crypt-villus junction. BMP signaling is low in the crypt and higher in the crypt villus junction. A definitive gradient exists for Eph-ephrin pathways too. The cells acquiring a differentiated fate switch off the expression of Eph-B and switch on the expression of Ephrin B ligands which progressively increases as they migrate up the axis.





**Figure 2 Intestinal stem cell signaling (Wnt, Notch, Math-1 pathways, lateral inhibition).**



**Figure 3 Treatment algorithm for goblet cell carcinoid.** FDG-PET: Fluorodeoxyglucose positron emission tomography; GCC: Goblet cell carcinoids.

**Table 1 Immunomarkers and mutations for appendiceal goblet cell carcinoids, typical neuroendocrine tumors and adenocarcinoma**

|  |  |  |  |
| --- | --- | --- | --- |
| **Markers** | **Goblet cell carcinoid.** | **Typical carcinoid** | **Adenocarcinoma** |
| CEA  CK7  CK20  CDX2  CD56  CAM5.2  Synaptophysin  Chromogranin A  Β-Catenin (nuclear)  P53  Ki67%  MUC1  MUC2  E-cadherin expression  MATH-1 expression  KRAS mutation  SMAD4 mutation  Notch signaling inhibition  MMR (MSH2, MSH6, MLH1, PMS2) | +  +  +  +  +/-  +  +/-  +/-  -  +/-  +/-  -  ++  N  +  -  -  U  - | -  -  -  -  ++  -  ++  ++  -  -  +/-  -  -  N  +  -  -  +  - | +  +  +  +  -  +  -  -  +  ++  ++  ++  +/-  U  -  +  +  -  +/- |

(+): Present; (-): Absent; (+/-): Present sometimes. CEA: Carcinoembryonic antigen; CK: Cytokeratin; CDX2: Caudal type homeobox transcription factor 2; CD56: Neural cell adhesion molecule; CAM 5.2: Antibody against CK8; P53: Tumor protein 53; Ki-67: Cellular proliferative marker; MUC: Mucin; Math-1: Protein atonal homolog 1 (a [basic helix-loop-helix](https://en.wikipedia.org/wiki/Basic_helix-loop-helix) family of [transcription factors](https://en.wikipedia.org/wiki/Transcription_factor)); kras: V-Ki-ras2 Kirsten rat sarcoma viral oncogene homolog; SMAD4: Mothers against decapentaplegic homolog 4, transcription factors in the TGF pathway; MMR: Mismatch repair genes; N: Normal expression; U: Unknown.