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**Not all arrestins are created equal: Therapeutic implications of the functional diversity of the β arrestins in the heart**

Lymperopoulos A *et al.* Functional differences of cardiac β arrestins

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**Abstract**

The two ubiquitous, outside the retina, G protein-coupled receptor (GPCR) adapter proteins, β arrestin-1 and -2 (also known as arrestin-2 and -3, respectively), have three major functions in cells: GPCR desensitization, *i.e*., receptor decoupling from G-proteins; GPCR internalization *via* clathrin-coated pits; and signal transduction independently of or in parallel to G-proteins. Both β arrestins are expressed in the heart and regulate a large number of cardiac GPCRs. The latter constitute the single most commonly targeted receptor class by Food and Drug Administration-approved cardiovascular drugs, with about one-third of all currently used in the clinic medications affecting GPCR function. Since β arrestin-1 and -2 play important roles in signaling and function of several GPCRs, in particular of adrenergic receptors and angiotensin II type 1 receptors, in cardiac myocytes, they have been a major focus of cardiac biology research in recent years. Perhaps the most significant realization coming out of their studies is that these two GPCR adapter proteins, initially thought of as functionally interchangeable, actually exert diametrically opposite effects in the mammalian myocardium. Specifically, the most abundant of the two β arrestin-1 exerts overall detrimental effects on the heart, such as negative inotropy and promotion of adverse remodeling post-myocardial infarction (MI). In contrast, β arrestin-2 is overall beneficial for the myocardium, as it has anti-apoptotic and anti-inflammatory effects that result in attenuation of post-MI adverse remodeling, while promoting cardiac contractile function. Thus, design of novel cardiac GPCR ligands that preferentially activate β arrestin-2 over β arrestin-1 has the potential of generating novel cardiovascular therapeutics for heart failure and other heart diseases.

**Key words:** Adverse remodeling;β arrestin;Biased signaling; Cardiac myocyte; Cardiac fibroblast; contractility; Functional divergence;G protein-coupled receptor; Heart failure; Hormone; Myocardial infarction; Signal transducer

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**Core tip:** Presumed functionally similar for a long time, we now know that the two β arrestins display significant functional diversity in several organs and tissues, including in the cardiovascular system. Their functional distinction also in the mammalian heart has been clearly documented over the past few years. β arrestin-1, which is far more abundant than β arrestin-2 in almost every tissue including the myocardium, opposes the cyclic adenosine monophosphate (cAMP)-dependent pro-contractile signaling of the β1 adrenergic receptor (β1AR), and promotes cardiac apoptosis, inflammation, and other adverse remodeling-associated processes post-myocardial infarction. Conversely, β arrestin-2 promotes catecholamine-dependent cardiac contractility directly, *via* SERCA2a potentiation, and indirectly, by leaving β1AR’s cAMP-dependent pro-contractile signaling unaffected.

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**INTRODUCTION**

Out of the four mammalian arrestins, only the two ubiquitous (outside the retina) arrestin-2 and -3, also known as β arrestin-1 and -2 respectively, are expressed in the mammalian cardiovascular system. Like in almost every tissue, β arrestin-1 protein is approximately 10-15-fold more abundant than β arrestin-2 in the circulatory system, as well[1]. Bothβ arrestins regulate all non-opsin G protein-coupled receptors (GPCRs), also known as seven transmembrane-spanning receptors (7TMRs), including those responsible for neurohormonal regulation of cardiovascular physiology[2,3]. For instance, cardiac function (contractility) is tightly controlled by the activity of β-adrenergic receptors (βARs) located in the membranes of cardiac myocytes[4-8]. Cardiac structure and morphology are regulated by angiotensin II (AngII) type 1 receptors (AT1Rs) present (mainly) in cardiac fibroblast and endothelial cell membranes[4,7]. Even the production and release of the regulatory hormones per se, whether it be catecholamine and corticosteroid release by the adrenal glands or activation of the renin-angiotensin-aldosterone system by the juxtagomerular apparatus of the kidneys or release of neurotransmitters by central and peripheral neurons innervating cardiovascular organs, is under tight regulation by various GPCRs[1,4,7].

Cardiovascular GPCRs can signal either through G-proteins or β arrestins with the natural, endogenous agonist hormones activating both signal transducers at each receptor fully and equally[1,9]. Several “biased” GPCR ligands have been discovered that (relatively) selectively activate either G proteins or β arrestins[1,9]. This “bias” in terms of the activated signal transducer is always relative but the concept of “biased” signaling and its attainability for therapeutic purposes has been challenged recently. Specifically, recent studies have shown that receptors can activate both G-proteins and β arrestins at the same time[10] or that β arrestins do not even need to bind the agonist-activated receptor to get (“catalytically”) activated[11]. Additionally, it was very recently clearly demonstrated that G-protein activation is absolutely necessary, at least initially upon agonist activation, for β arrestin activation and signaling to follow[12,13]. This sequence of activation of the two signal transducers, *i.e.*, G-proteins being activated first followed by activation of β arrestins, is also corroborated well by the majority of structural studies on mechanisms of GPCR activation done to date. Specifically, the receptor seems to require the interaction with the heterotrimeric G-protein in order to become fully activated by the agonist. In other words, in the absence of a G-protein, agonist binding per se is simply insufficient for the receptor to break the huge energy barrier that prevents it from reaching the active state[2,14]. Taken together, G-protein activation and signaling appears to be a prerequisite for β arrestin signaling by GPCRs and thus, discrimination between these two families of signal transducers for any given GPCR ligand, which represents the foundation of the “biased signaling” concept for GPCRs, is essentially unfeasible. However, whereas the selective stimulation of G-protein vs. β arrestin signaling for therapeutic purposes is most likely impossible, selective stimulation of β arrestin-1 *vs* β arrestin-2 might be feasible, similarly to the selective stimulation (or inhibition) of various G subunits, which is pharmacologically achievable and currently exploited therapeutically. The first hint at signaling and functional differences between the two β arrestins came over a decade ago with the realization that β arrestin-1, but not β arrestin-2 which has a nuclear export signal sequence (NES), can translocate to the nucleus where it regulates gene transcription[15]. Since then, the experimental evidence supporting functional divergence between the signaling properties of the two β arrestins both *in vitro* and in several tissues and organs *in vivo*, including in the heart, has been mounting at an accelerating pace. Thus, β arrestin isoform-selective targeting may have a place in the design and development of novel drugs. Below, we review this evidence known so far for the cardiac β arrestins and discuss what it could signify for heart failure drug development. Given that almost all of the *in vivo* studies on cardiac β arrestins done so far are in relation to the effects of these two proteins on βAR and AT1R signaling in the heart, the evidence for cardiac β arrestins’ functional diversity reviewed below pertains exclusively to cardiac βARs and AT1Rs.

**FUNCTIONAL DIFFERENCES BETWEEN THE TWO Β ARRESTINS IN CARDIAC β AR SIGNALING**

The β1AR is by far the predominant βAR subtype in human adult cardiac myocytes, representing 75%-80% of total βAR density, followed by the β2AR, which comprises about 15-18% of total cardiomyocyte βARs and the remaining 2%-3% is β3ARs[4,7,16]. β1AR stimulation by catecholamines results in the dissociation of the stimulatory G protein alpha subunit (Gs) from Gβγ. Gs stimulates adenylyl cyclase (AC) to produce cyclic adenosine monophosphate (cAMP), which, in turn, activates protein kinase A (PKA) and regulates different intracellular, sarcolemmal and myofibrillar substrates[4,5,7]. Thus, cAMP-dependent signaling in cardiomyocytes mediates the cellular effects of β1AR activation on stimulation of cardiac chronotropy, inotropy, dromotropy, and lusitropy (Figure 1)[4,5,7]. As co-factors of GPCR-kinases (GRKs) in βAR desensitization/downregulation, β arrestins normally diminish the inotropic and β-adrenergic reserves of the failing heart and their inhibition should theoretically be beneficial in acute decompensated heart failure (ADHF)[4,7]. Indeed, genetic deletion of β arrestin-1 in the heart results in several desirable therapeutic effects in heart failure, such as dramatic improvements in both cardiac β-adrenergic and inotropic reserves, amelioration of adverse remodeling and increased survival post-myocardial infarction (MI) (Figure 1)[17,18]. In contrast however, cardiac β arrestin-2 has been shown to be cardio-protective, as it inhibits cardiac apoptosis, inflammation, and significantly attenuates overall adverse remodeling post-MI (Figure 1)[19]. One of the underlying mechanisms for the anti-inflammatory effects of cardiac β arrestin-2 is nuclear factor-kappaB (NFĸB) inhibition in cardiac myocytes, which, again, appears to be mediated only by β arrestin-2 and not by β arrestin-1 in the heart (Figure 1)[19,20]. Importantly, β1AR-stimulatedβ arrestin-2 was also recently documented to increase cardiac contractility both directly and indirectly (Figure 1)[20]. Directly, by interacting with Sarco/Endoplasmic Reticulum Ca2+-ATPase (SERCA)-2a leading to enhanced Small Ubiquitin-related MOdifier (SUMO)-ylation of the latter[20]. This process, deficient in human heart failure, is known to directly stimulate SERCA2a activity, thereby increasing cardiac contractility[21]. β arrestin-2 also increases cardiac function indirectly by leaving the β1AR-stimulated cAMP-dependent pro-contractile signaling intact (*i.e.*, not desensitizing it) in cardiac myocytes *in vitro* and in post-MI heart failure mice *in vivo* (Figure 1)[20]. Importantly, these effects are not shared by the vastly more abundant in the human heart β arrestin-1[22].

One of the salient mechanisms for the anti-apoptotic effects of cardiac β arrestin-2 is transactivation of the epidermal growth factor receptor (EGFR) by the cardiac β1AR (Figure 1)[18,23]. β arrestin-1 seems again unable to stimulate this and instead, promotes cardiac apoptosis post-MI (Figure 1)[18].Older studies had reported that mice expressing a mutant β1AR that cannot undergo GRK-dependent desensitization or activateβ arrestins in their hearts lack cardiac EGFR transactivation and suffer from massive cardiac apoptosis and left ventricular dilatation compared to wild type controls[23]. Interestingly, the β-blocker drug carvedilol, an inverse agonist towards G-protein activation[16], is a weak β arrestin-biased agonist that activates ERKs (Extracellular signal-Regulated Kinases) *via* EGFR transactivation also[24,25]. It should be pointed out though that carvedilol`s “biased” β arrestin agonism has been demonstrated only in heterologous recombinant cell systems without cardiovascular (or any other physiological) relevance (mostly, transfected HEK293 cells). However, if this holds true in actual cardiomyocytes *in vivo*, it might be part of the mechanism for this β-blocker`s cardio-protective effects. However, several studies do not lend support to this notion; sustained β1AR activation by catecholamines, markedly more potent activators of β arrestin-dependent ERKs in the heart than carvedilol, increases cardiac adverse remodeling even in the absence of cardiac injury[26]. Moreover, carvedilol is also a β2- and 1AR antagonist, which may interfere with its β arrestin-activating properties in the heart[27]. On the other hand, carvedilol`s effects in the heart are virtually exclusively mediated by the β1AR, due to its relative selectivity for the human β1AR over the other human AR subtypes (β2AR and 1AR) and to the vast preponderance of the β1AR over the rest of AR subtypes in the human adult myocardium[16]. Last but not least, recent studies have been unable to directly detect β arrestin interactions with either β1AR or β2AR bound to carvedilol, including a study reporting the intriguing finding that carvedilol requires Gi protein recruitment to the β1AR in order to induce EGFR transactivation *via* β arrestins[28-31]. Nevertheless, carvedilol has been shown to selectively recruit β arrestin-2 to the hyperfunctional human polymorphic Arg389 β1AR in cardiomyocytes *in vitro*[32]. Therefore, the more robust, compared to its Gly389 counterpart, pro-contractile signaling of this β1AR variant[33] might be, at least partly, due to its unique β arrestin-2-interacting tropism. In this vein, very recent data from our lab indicate that carvedilol-bound β1AR uniquely stimulates β arrestin-2-dependent SERCA2a activity and fractional shortening of cardiomyocytes *in vitro*[34].

**FUNCTIONAL DIFFERENCES BETWEEN THE TWO β ARRESTINS IN CARDIAC AT1R SIGNALING**

Despite its very low abundance in adult human myocardium (density ratio of AT1R/β1AR in the non-failing human heart: approximately 1:15)[16], the AT1R plays important regulatory roles in the heart, but mainly *via* actions in cardiac fibroblasts and endothelial cells, rather than in cardiac myocytes[4,7,35-37]. The AngII peptide analog SII ([Sar1-Ile4-Ile8]-AngII) does not elicit Gq protein signaling when bound to the AT1R[9,38]. When it was discovered to induce β arrestin signaling from the AT1R, the concept of biased signaling was introduced and ushered in a completely new field in cardiovascular drug development with companies designing novel “biased” AT1R ligands that only activate β arrestins while retaining G-protein-blocking properties. SII has now been documented to not be completely β arrestin-“biased”, as it can activate some G-protein types (*e.g.*, Gs, Gi) from the AT1R[9]. Nevertheless, studies have shown that AT1R-elicited β arrestin-dependent signaling in cardiac myocytes leads to cardiomyocyte proliferation without hypertrophy, which is Gq/11 protein signaling-dependent, and may even result in positive inotropy and lusitropy depending on which GRK isoform is involved (the so-called receptor “bar-coding” by GRKs). Interestingly, which β arrestin is engaged is also crucial[39,40]. Specifically, GRK2-dependent phosphorylation followed by β arrestin-1 binding seems to reduce, whereas GRK6-dependent phosphorylation followed by β arrestin-2 interaction seems to promote AT1R-induced contractility in primary murine adult cardiomyocytes (Figure 1)[39]. However, AT1R-activated β arrestins have no effect on contractility in isolated Langendorff-perfused cardiac preparations[41]. Furthermore, a recombinant AT1AR capable of only signaling through β arrestins inhibits myocardial apoptosis and fibrosis, and enhances cardiomyocyte hypertrophy, upon transgenic overexpression in mouse hearts[42]. Interestingly, a β arrestin-2-“biased” ligand at the AT1R is very beneficial in mice suffering from dilated cardiomyopathy as it prevents maladaptive signaling and improves myofilament Ca2+ sensitivity[43]. Thus, cardiac AT1R promotes hypertrophy and cardiomyocyte proliferation *via* the classic Gq protein/phospholipase C-β signaling pathway, which is inhibited by the β arrestins (classic AT1R desensitization), and, at the same time, β arrestin-2 (but not β arrestin-1) can increase cardiac function *via* its cardiac AT1R-dependent signaling (Figure 1).

Based on the above studies, several SII-derivative peptides have been synthesized and tested in animal models of ADHF with promising initial results[44,45]. Unfortunately, these compounds failed in large phase III clinical trials (BLAST-AHF, ClinicalTrials.gov number, NCT01966601). There are probably several reasons for this. First, findings in animal models do not always translate into humans. Second, the compounds might have not been completely β arrestin-“biased” (*i.e.*, maybe they had some weak, residual activity towards certain G-proteins). One intriguing possibility is that, due to the significantly lower abundance of the cardioprotective β arrestin-2, compared to the cardio-toxic β arrestin-1, in human cardiomyocytes[22], these β arrestin-“biased” compounds stimulated, in reality, β arrestin-1 instead of β arrestin-2 in the patients` hearts and that’s why their clinical outcomes were not the desired ones. Finally, it is very likely that these compounds stimulated the AT1R only in cardiac fibroblasts, which would preclude any clinical benefit for ADHF patients. In fact, both β arrestins have been shown to regulate human cardiac fibroblast differentiation and to mediate collagen synthesis in human failing left ventricle-derived fibroblasts, thereby promoting the adverse remodeling process of cardiac fibrosis[46].

Notably, β arrestins have been reported to mediate signaling by the mechanical stretch-activated (unliganded) cardiac AT1R[47], which has been suggested to underlie one of the fundamental laws of cardiac physiology, the Frank-Starling mechanism of cardiac contractility[48]. Although intriguing, this finding is very difficult to interpret, given that the Frank-Starling mechanism is hemodynamically/biomechanically governed rather than dependent on hormonal receptors/effects. Besides, if it was mediated by a cardiac GPCR, then that receptor would definitely be the β1AR, the by far most abundant GPCR (and at least 15-fold more abundant than the AT1R) in cardiomyocytes[16].

Finally, β arrestin-mediated signaling by the AT1R that can regulate cardiac function occurs in the adrenal cortex, as well. Specifically, the AT1R promotes the adrenocortical production of aldosterone, a cardio-toxic hormone elevated in chronic human heart failure, *via* both Gq/11-proteins and β arrestin-1[35,49-51]. In fact, adrenal β arrestin-1 is essential for aldosterone production, since, in mice lacking the *β arrestin-1* gene, circulating aldosterone levels do not increase even in the presence of MI[18]. Interestingly, the prototypic AT1R antagonist (ARB, angiotensin receptor blocker) losartan is a relatively G protein-“biased” antagonist, which means it cannot suppress β arrestin-1-dependent aldosterone production[51-54]. In contrast, candesartan and valsartan are potent β arrestin-inverse agonists at the adrenal AT1R and very effective at suppressing aldosterone *in vitro* and *in vivo*[52,53]. These differences among ARBs, which are all orthosteric antagonists, in their potency at blocking AT1R-activated β arrestin signaling may reflect their differential abilities to suppress β arrestin signaling by the unliganded (*i.e.*, constitutively active) AT1R[47,55]. In other words, the ARBs seem to be inverse agonists not only for G-proteins but also for β arrestins at the AT1R.

THERAPEUTIC IMPLICATIONS IN HEART FAILURE

It becomes clear from the above that the signaling effects of the two β arrestins in the heart are not just different but actually diametrically opposite. This is true for several other mammalian organ systems and tissue types[56-58] and is completely corroborated by molecular, biophysical, crystallographic, and proteomic studies[59-62]. It also makes sense from the evolutionary point of view, as functional redundancy of proteins is usually not favored by natural selection. Only during embryonic development might the two β arrestins be able to compensate for each other, since single β arrestin-knockout mice (either β arrestin-1- or β arrestin-2-knockouts) are viable and breed normally but the double β arrestin-knockout mouse is embryonic-lethal[63].

Regarding the myocardium, all the functional studies on β arrestins done so far are in relation to either βARs or the AT1R. Studies on β arrestin signaling from other cardiac GPCRs in cardiac cells are lacking. Based on the available data for β1AR, β2AR, or AT1R signaling through β arrestins in cardiac cells, it can be safely concluded that β arrestin-1 is the arrestin responsible for the classic desensitization of the cAMP-dependent pro-contractile signaling of the β1AR. This quite simply means that β arrestin-1 exerts overall negative inotropy and is responsible (together with the elevated in human heart failure cardiac GRK2) for the diminished inotropic and adrenergic reserves of the failing human heart. In addition, β arrestin-1 promotes cardiac apoptosis, inflammation, and accelerates cardiac adverse remodeling post-MI. In direct contrast, β arrestin-2 promotes β1AR-mediated cardiac contractility both directly and indirectly. Directly, *via* augmentation of SERCA2a activity, and indirectly, by leaving the β1AR’s cAMP-dependent pro-contractile signaling intact (*i.e*., no desensitization). On the other hand, it inhibits apoptosis and inflammation, and overall attenuates cardiac adverse remodeling post-MI, *via* stimulation of a variety of molecular pathways, such as EGFR transactivation and NFB inhibition, which β arrestin-1 does not activate. Induction of ERK and of other mitogenic/proliferative molecular signaling pathways in cardiomyocytes play auxiliary roles in β arrestin-2’s reverse remodeling effects, as well. Of note, the same functional distinction between the two cardiac β arrestins (*i.e*., β arrestin-1 being detrimental, β arrestin-2 being beneficial) applies to cardiac AT1R signaling, too. Regardless of how small the contribution of this GPCR to the overall contractile function of the cardiac myocyte is, β arrestin-2 again appears to promote contractility and cardiomyocyte survival in response to AT1R activation. Conversely, β arrestin-1 (again in conjunction with GRK2) opposes the AT1R-dependent pro-contractile signaling in cardiac myocytes. However, β arrestin-1 might exert an indirect beneficial effect in the hypertrophic heart by desensitizing (reducing) the pro-hypertrophic signaling of the cardiac AT1R through Gq/11-proteins. In conclusion, based on their observed effects on the signaling of both β1ARs and AT1Rs in cardiac myocytes, documented either directly (in β arrestin-knockout mice) or indirectly (with the use of “biased” receptor ligands), cardiac β arrestin-2 stimulation and/or β arrestin-1 inhibition might be valid therapeutic strategies in human heart failure. By the way, it is interesting to note here that probably the exact opposite is the case in the systemic vasculature. In vascular smooth muscle cells, β arrestin-1 appears beneficial in terms of vasodilation and attenuation of hyperplasia and β arrestin-2 seems to promote hypertrophy/hyperplasia[58]. This should not come as a surprise at all, given the different cellular machineries and GPCR types involved in β arrestin signaling between cardiomyocytes (mainly β1AR) and vascular smooth muscle cells (mainly AT1R and, to a lesser extent, β2AR). Besides, this is exactly what happens with the major second messenger cAMP: stimulated by the β1AR, it is pro-contractile in cardiomyocytes, but stimulated by the β2AR, it is pro-dilatory in vascular smooth muscle.

**CONCLUSION**

In the adult myocardium, the actions of β arrestin-1 are detrimental, since they result in depletion of the functional and adrenergic reserves of the heart. In contrast, β arrestin-2 is beneficial, since it can increase both of these cardiac reserves or at least preserve them in the face of a cardiac insult/damage, such as an MI. Thus, from the therapeutic standpoint, cardiac β arrestin-1 blockade or β arrestin-2 stimulation should be pursued for heart disease treatment. Of course, there are at least three very important questions awaiting answer in future studies in order to fully validate these strategies as therapeutic options for human heart failure. First, the effects of the two β arrestins on the signaling of other important cardiac GPCRs in the heart, *i.e*., beyond the βARs and the AT1R, need to be elucidated. The second issue to resolve is what the effects of the two β arrestins in other tissues/components of the cardiovascular system are, *e.g*., vasculature, platelets, adrenals, *etc*. This is particularly important if the pharmacological targeting of the β arrestins with systemically administered agents is being explored. For instance, β arrestin-2 is beneficial in the heart, in platelets, and in vascular endothelium but might be detrimental in vascular smooth muscle[58]. Its exact effects in all of these tissues need to be thoroughly investigated and determined, if a drug that stimulates this β arrestin isoform is to be designed and developed. Finally, the third and therapeutically salient unanswered question pertains to the expression levels of the cardiac β arrestins in the failing human heart. Although β arrestin-2 protein is significantly under-expressed, compared to β arrestin-1, in the non-failing human adult myocardium[22,64], which makes cardiac-specific β arrestin-2 gene transfer an enticing approach for heart failure treatment, its protein levels (and if they change) in human heart failure remain unknown. However, given that it is significantly expressed at the mRNA level, and in fact at levels comparable to those of β arrestin-1 mRNA, in the failing human heart[64], it is quite plausible that it might be upregulated at the protein level, similarly to GRK2, as a homeostatic mechanism of the failing human myocardium to confer cardioprotection. The only study done to date investigating β arrestin expression in failing human hearts is quite old (was published almost 25 years ago) and could not detect any changes in the protein levels of either β arrestin[64]. Nevertheless, although β arrestin-1 protein probably does not change in human heart failure, because it is already highly expressed in normal, healthy human hearts, that study failed to detect any β arrestin-2 protein at all, even in normal healthy human hearts, probably due to technical deficiencies of the antibody it used[64]. Therefore, it is rather inconclusive with regard to cardiac β arrestin-2 protein expression in humans and whether cardiac β arrestin-2 protein changes in human heart failure remains an open question. In fact, a much more recent study done in human cardiac fibroblasts isolated from left ventricles of heart failure patients hinted at β arrestin-2 protein being upregulated in human failing hearts[46]. In any case, more studies are certainly warranted, especially in human cardiac specimens, to unravel the full spectrum of physiological (and pathophysiological) actions of the two cardiac β arrestins, beginning with the answering of the three outstanding questions mentioned above. Only then will the true potential of individual cardiac β arrestin isoform targeting for heart failure therapy be revealed, so that the pharmaceutical industry can begin its realization.

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**Figure 1 The functional distinction between** β **arrestin-1 and** β **arrestin-2 in cardiac myocytes.** ATP: Adenosine triphosphate; P: Phosphorylation; SR: Sarcoplasmic reticulum; SUMO1: Small ubiquitin-like modifier protein-1; PLN: Phospholamban; PIP2: Phosphatidyl-inositol 4,5-bisphosphate; IP3: Inositol 1,4,5-trisphosphate; DAG: 1,2-Diacylglycerol; EGFR: Epidermal growth factor receptor; AR: Adrenergic receptor.