

Artificial Intelligence in *Cancer*

Artif Intell Cancer 2020 June 28; 1(1): 1-38



A**I****C**

Artificial Intelligence in Cancer

Contents**Bimonthly Volume 1 Number 1 June 28, 2020****EDITORIAL**

- 1 Artificial intelligence and omics in cancer
Coulouarn C

EVIDENCE REVIEW

- 8 Management of cancer patients during the COVID-19 pandemic: A comprehensive review
Cassell III AK, Cassell LT, Bague AH

MINIREVIEWS

- 19 Application of artificial intelligence in clinical non-small cell lung cancer
Liu Y

ORIGINAL ARTICLE**Basic Study**

- 31 Impact of blurs on machine-learning aided digital pathology image analysis
Ogura M, Kiyuna T, Yoshida H

ABOUT COVER

Editor-in-Chief of *Artificial Intelligence in Cancer*, Dr. Cedric Coulouarn has a long-standing expertise and track record in liver cancer with focus on TGF-beta signaling, non-coding RNA and functional genomics, including a 5-year experience at the National Cancer Institute. He currently heads a team at Inserm in France focused on studying the role of TGF-beta signaling in liver carcinogenesis. He is an active member of the French and European associations for the Study of the Liver (AFEF and EASL), International Liver Cancer Association, European Network for the Study of Cholangiocarcinoma. Dr. Coulouarn is also acting as a referee in scientific committees for evaluation of French and international Grants. He is teaching at University Paris-Diderot and University of Rennes 1, France.

AIMS AND SCOPE

The primary aim of *Artificial Intelligence in Cancer (AIC, Artif Intell Cancer)* is to provide scholars and readers from various fields of artificial intelligence in cancer with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

AIC mainly publishes articles reporting research results obtained in the field of artificial intelligence in cancer and covering a wide range of topics, including artificial intelligence in bone oncology, breast cancer, gastrointestinal cancer, genitourinary cancer, gynecological cancer, head and neck cancer, hematologic malignancy, lung cancer, lymphoma and myeloma, pediatric oncology, and urologic oncology.

INDEXING/ABSTRACTING

There is currently no indexing.

RESPONSIBLE EDITORS FOR THIS ISSUE

Electronic Editor: *Ji-Hong Lin*, Production Department Director: *Xiang Li*, Editorial Office Director: *Jin-Lai Wang*.

NAME OF JOURNAL

Artificial Intelligence in Cancer

ISSN

ISSN 2644-3228 (online)

LAUNCH DATE

June 28, 2020

FREQUENCY

Bimonthly

EDITORS-IN-CHIEF

Mujib Ullah, Cedric Coulouarn

EDITORIAL BOARD MEMBERS

<https://www.wjgnet.com/2644-3228/editorialboard.htm>

PUBLICATION DATE

June 28, 2020

COPYRIGHT

© 2020 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

<https://www.wjgnet.com/bpg/gerinfo/204>

GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjgnet.com/bpg/GerInfo/287>

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjgnet.com/bpg/gerinfo/240>

PUBLICATION ETHICS

<https://www.wjgnet.com/bpg/GerInfo/288>

PUBLICATION MISCONDUCT

<https://www.wjgnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

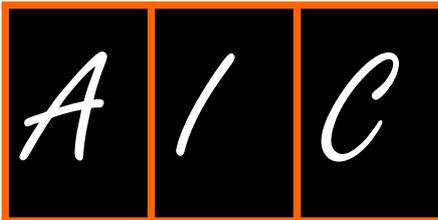
<https://www.wjgnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjgnet.com/bpg/GerInfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>



Basic Study

Impact of blurs on machine-learning aided digital pathology image analysis

Maki Ogura, Tomoharu Kiyuna, Hiroshi Yoshida

ORCID number: Maki Ogura 0000-0002-0380-9396; Tomoharu Kiyuna 0000-0003-3050-6718; Hiroshi Yoshida 0000-0002-7569-7813.

Author contributions: Ogura M, Kiyuna T, and Yoshida H drafted and revised the manuscript and prepared the figures; Ogura M collected the pathological data; Kiyuna T performed all the image analysis; all the authors have read and approved the final manuscript.

Institutional review board statement: The study was conducted in accordance with the Declaration of Helsinki and with the approval of the Institutional Review Board of the National Cancer Center, Tokyo, Japan.

Conflict-of-interest statement: All authors have no competing interests to be declared.

Data sharing statement: No additional data are available.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build

Maki Ogura, Tomoharu Kiyuna, Digital Healthcare Business Development Office, NEC Corporation, Tokyo 108-8001, Japan

Hiroshi Yoshida, Department of Diagnostic Pathology, National Cancer Center Hospital, Tokyo 104-0045, Japan

Corresponding author: Hiroshi Yoshida, MD, PhD, Staff Physician, Department of Diagnostic Pathology, National Cancer Center Hospital, 5-1-1 Tsukiji, Chuo-ku, Tokyo 104-0045, Japan. hiroyosh@ncc.go.jp

Abstract

BACKGROUND

Digital pathology image (DPI) analysis has been developed by machine learning (ML) techniques. However, little attention has been paid to the reproducibility of ML-based histological classification in heterochronously obtained DPIs of the same hematoxylin and eosin (HE) slide.

AIM

To elucidate the frequency and preventable causes of discordant classification results of DPI analysis using ML for the heterochronously obtained DPIs.

METHODS

We created paired DPIs by scanning 298 HE stained slides containing 584 tissues twice with a virtual slide scanner. The paired DPIs were analyzed by our ML-aided classification model. We defined non-flipped and flipped groups as the paired DPIs with concordant and discordant classification results, respectively. We compared differences in color and blur between the non-flipped and flipped groups by L1-norm and a blur index, respectively.

RESULTS

We observed discordant classification results in 23.1% of the paired DPIs obtained by two independent scans of the same microscope slide. We detected no significant difference in the L1-norm of each color channel between the two groups; however, the flipped group showed a significantly higher blur index than the non-flipped group.

CONCLUSION

upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Received: March 21, 2020

Peer-review started: March 21, 2020

First decision: April 22, 2020

Revised: May 2, 2020

Accepted: June 7, 2020

Article in press: June 7, 2020

Published online: June 28, 2020

P-Reviewer: Youness RA, Zhang K

S-Editor: Wang JL

L-Editor: A

E-Editor: Liu JH



Our results suggest that differences in the blur - not the color - of the paired DPIs may cause discordant classification results. An ML-aided classification model for DPI should be tested for this potential cause of the reduced reproducibility of the model. In a future study, a slide scanner and/or a preprocessing method of minimizing DPI blur should be developed.

Key words: Machine learning; Digital pathology image; Automated image analysis; Blur; Color; Reproducibility

©The Author(s) 2020. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Little attention has been paid to the reproducibility of machine learning (ML)-based histological classification in heterochronously obtained Digital pathology images (DPIs) of the same hematoxylin and eosin slide. This study elucidated the frequency and preventable causes of discordant classification results of DPI analysis using ML for the heterochronously obtained DPIs. We observed discordant classification results in 23.1% of the paired DPIs obtained by two independent scans of the same microscope slide. The group with discordant classification results showed a significantly higher blur index than the other group. Our results suggest that differences in the blur of the paired DPIs may cause discordant classification results.

Citation: Ogura M, Kiyuna T, Yoshida H. Impact of blurs on machine-learning aided digital pathology image analysis. *Artif Intell Cancer* 2020; 1(1): 31-38

URL: <https://www.wjgnet.com/2644-3228/full/v1/i1/31.htm>

DOI: <https://dx.doi.org/10.35713/aic.v1.i1.31>

INTRODUCTION

Recent developments in medical image analysis empowered by machine learning (ML) have expanded to digital pathology image (DPI) analysis^[1-3]. For over ten years, NEC Corporation has researched and developed image analysis software that can detect carcinomas in tissue in the digital images of hematoxylin and eosin (HE) stained slides. DPI analysis is generally performed for digital images obtained with special devices such as microscopic cameras or slide scanners. These devices cannot make completely identical digital images or data matrices even when the same microscope slide is repeatedly shot with the same camera or scanned by the same scanner.

In general, image analysis by ML can provide different classification results if an object has multiple images showing different features. Therefore, slight differences in a DPI made by imaging devices can also cause different classification results. Each digital image will have different characteristics even when the same microscope slide of a patient is repeatedly digitized by the same slide scanner. Similarly, the same microscope slide of a patient can be digitized at a local hospital and then at a referral hospital. The resulting differences in image features of the same microscope slide can provide discordant classification results of DPI analysis, confusing both patients and medical professionals. However, only a few reports have mentioned this issue.

The aim of this study is to elucidate the frequency and preventable cause of discordant classification results of DPI analysis using ML in the aforementioned situation. We compared the classification results between paired DPIs of the same microscope slide obtained from two independent scans using the same slide scanner (Figure 1).

MATERIALS AND METHODS

Tissue sample

We conducted the study in accordance with the Declaration of Helsinki and with the approval of the Institutional Review Board of the National Cancer Center, Tokyo, Japan. We consecutively collected 3062 gastric biopsy specimens between January 19-April 30, 2015 at the National Cancer Center (Tsukiji and Kashiwa campuses). The

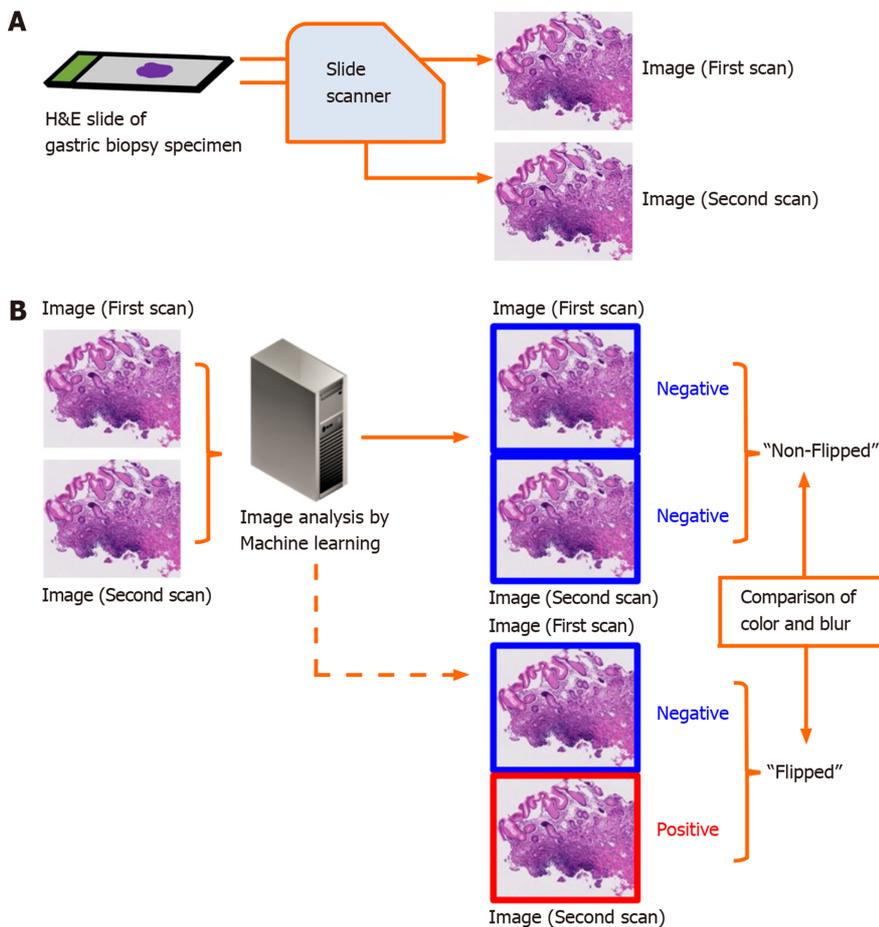


Figure 1 The schema of this study. A: Hematoxylin and eosin stained slides of gastric biopsy specimens were scanned twice by the same slide scanner, then the paired digital images were created; B: The paired images were independently analyzed and classified by our machine-learning model. If concordant classification results were obtained, the case is "Non-flipped"; if discordant classification results were obtained, the case is "Flipped." Then, color and blur differences were compared between the "Non-flipped" and "Flipped" groups.

specimens were placed in 10% buffered formalin and embedded in paraffin. Each block was sliced into 4- μ m thick sections. Routine HE staining was performed for each slide using an automated staining system.

Digital image acquisition and automated image analysis

During the image collection and analysis procedure, the researchers were blind to all of the diagnoses of the human pathologists. We developed an ML model to analyze the DPIs using a multi-instance learning framework^[4]. The results of the concordance between pathological diagnosis by human pathologists and classification by an ML model was previously reported^[5]. In our study, we randomly selected 584 images of the 3062 specimens to use for the present analysis.

We scanned 298 HE stained slides containing 584 tissues twice using the NanoZoomer (Hamamatsu Photonics K. K., Shizuoka, Japan) virtual slide scanner, creating the paired DPIs. The paired DPIs were analyzed by our ML-aided classification model^[4]. Our ML-aided classification model classified the results of each tissue as "Positive" or "Negative". "Positive" denoted neoplastic lesions or suspicion of neoplastic lesions and "Negative" denoted the absence of neoplastic lesions. The procedure for classification of a cancerous areas in a given whole-slide image is as follows: (1) Identify the tissue regions at 1.25 \times ; (2) The tissue area was then divided into several rectangular regions of interest (ROIs); (3) From each ROI, the structural and nuclear features are extracted at different magnification (10 \times and 20 \times); (4) After the feature extraction, all ROIs were classified as positive or negative using a pre-trained classifier (support vector machine, SVM); and (5) The SVM-based classifier assigns a real number t to each ROI, where t takes value in the range (-1.0, 1.0). A value of 1.0 indicates a positive ROI and a value of -1.0 indicates a negative ROI^[5]. In this experiment, we interpreted the value of $t \geq 0.4$ indicates a positive ROI.

We defined the group without discordant classification results between the paired

DPIs as the “non-flipped group” and the group with discordant classification results as the “flipped group”.

For reference, we repeated analysis of the identical DPIs that had identical data matrices twice, then compared their results.

Color analysis

We separated tissue images into tissue regions and non-tissue regions. To examine the differences in tissue color in the first and second scanned images, we measured the L1-norm distance between color distributions of images in each color channel; *i.e.*, red (R), green (G), and blue (B). The L1-norm distance between normalized histograms p and q were defined as **Formula 1**:

Where p_i and q_i are the normalized frequencies at the i -th bin of histograms p and q , respectively.

Quantification of the degree of image blurring

We quantified the degree of image blurring using the variance of wavelet coefficients of an image^[6]. The degree of image blurring is calculated and normalized as follows: (1) 2D convolution by neighboring filter; (2) Local variance of a 5×5 area; and (3) Captures local phase variations after convolution with wavelet filters, normalized by a sigmoid function to (0, 1) range. The degree of blurring was then normalized to between 0 and 255 and we calculated its distribution (normalized histogram). We defined the blur index using the 98th percentile of the above distribution of the variance of wavelet coefficients.

Statistical analysis

We used the Mann-Whitney test to evaluate the significant differences in the blur index between the non-flipped and flipped groups.

RESULTS

Classification results of the paired DPIs

The analysis results did not change in 449 tissues; however, the results changed in 135 tissues (23.1%), either from positive to negative or from negative to positive (**Table 1**). Therefore, 135 tissues were in the flipped group.

On the other hand, 100% (584/584) of the concordance rate was observed between the classification results of the first analysis and the second analysis of the identical DPIs by our ML-aided classification model.

Comparison of the DPI color

We compared the medians of the L1-norm in the non-flipped and flipped groups and found no significant difference (**Table 2**).

Comparison of the blur index of the DPIs

Next, we calculated the blur index of the paired DPIs and compared it between the non-flipped group and the flipped group. The flipped group showed a significantly higher blur index than the non-flipped group (**Figure 2**). **Figure 3** shows a representative case of the flipped group's results.

DISCUSSION

We observed 23.1% of discordant classification results between the paired DPIs obtained from two independent scans of the same microscope slide. Furthermore, we detected differences in blur (not color) of the paired DPIs as a potential cause of different classification results.

Differences in the colors of DPIs did not correlate with discordant classification results in this study. Since differences in the colors of digitized images reportedly result in different features of digitized images and different data matrices^[7], we expected the difference in color to reduce reproducibility in our ML-aided classification model. However, the distribution of RGB value did not differ significantly between the paired DPIs and did not seem to cause discordant classification results. Nevertheless, color differences should be a concern because the

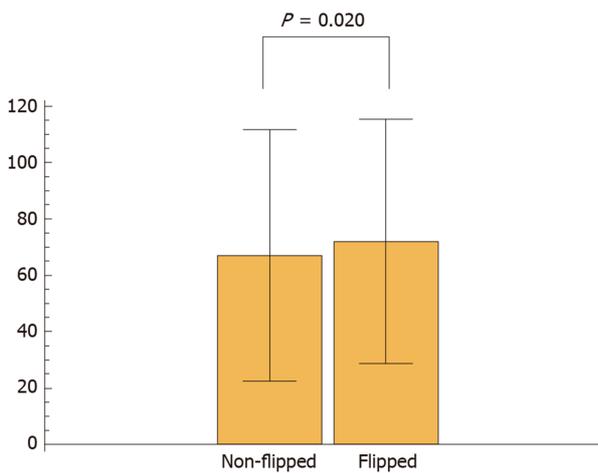
Table 1 Concordance of classification results between the paired digital pathology images

		The second scan		
		Positive	Negative	Unclassifiable
The first scan	Positive	248	66	0
	Negative	69	197	2
	Unclassifiable	1	0	4

Table 2 Comparison of pair-wise L1-norm between non-flipped and flipped groups

Color channel	Median of the non-flipped group	Median of the flipped group	P value
R	0.0350 ± 0.0220	0.0347 ± 0.0217	0.900
G	0.0319 ± 0.0197	0.0313 ± 0.0205	0.931
B	0.0266 ± 0.0148	0.0250 ± 0.0190	0.255

$$D_{L1}(p, q) = \sum_{i=1}^{255} |p_i - q_i|$$

**Figure 2** Differences in the blur index between the “Non-flipped” and “Flipped” groups.

color of HE stained slides obviously differed between different pathological laboratories. In such cases, a discordant classification result was observed in the same specimen with an identical pathological diagnosis (unpublished data). Therefore, even DPIs taken from the same microscope slide might show discordant classification results from obvious color changes due to the miscalibration of an imaging device.

Although qualitative changes in the blurs of the paired DPIs were macroscopically recognizable, their qualitative assessment was difficult. However, we developed a blur index that provided a quantitative comparison and detected the significant differences in blurs between the DPIs of the non-flipped group and those of the flipped group. Reportedly, blur can potentially influence the stability of features of a digitized image^[7]; so, first, our study demonstrated that quantifying blurs revealed their impact on classification results.

A significant portion of cases showed discordant classification results; however, our ML-aided classification model worked efficiently for our intended purpose. 80.7% of all the flipped cases was non-tumor tissue, and 6.5% was carcinoma tissue. Our ML-aided classification model set a lower threshold than the best one (*i.e.*, the threshold that yields a minimum error rate) because we made our model minimize false negative results, classifying carcinoma as non-tumor tissue. This lower threshold caused more frequent flipped cases in non-tumor tissue. In other words, the larger the percentage of

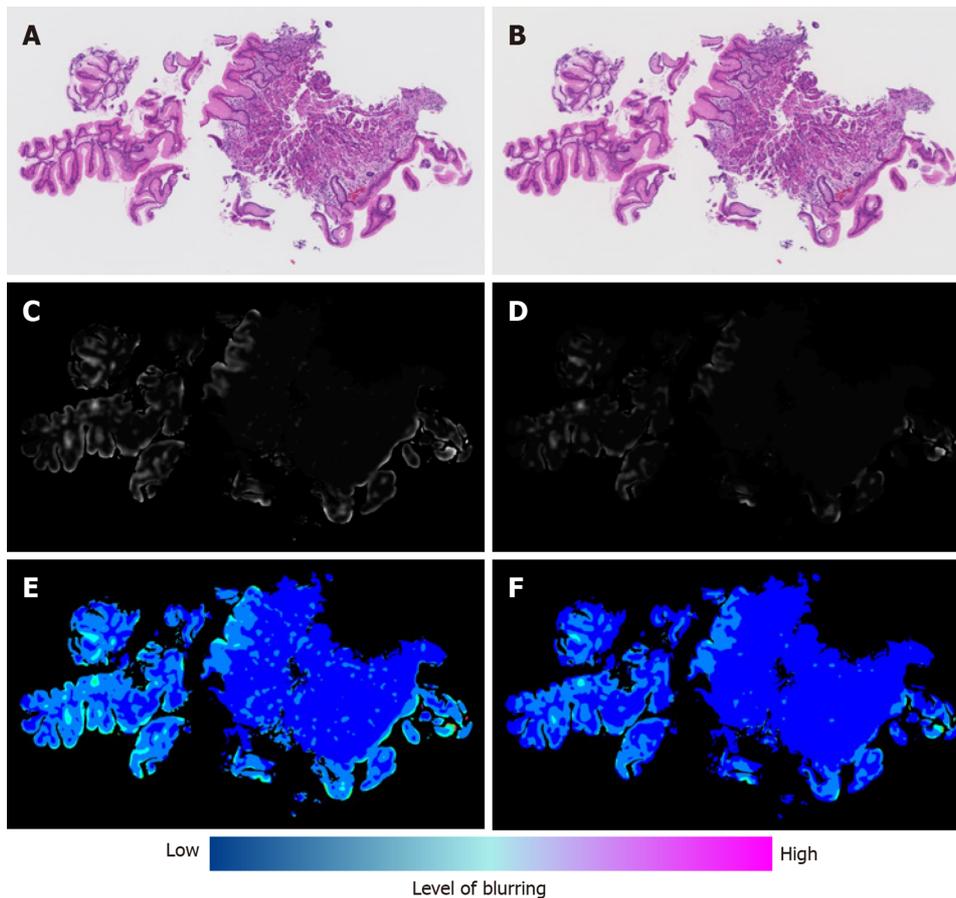


Figure 3 Typical examples of differences in the blurring level. A: Whole-slide image at the first scan; B: Whole-slide image at the second scan; C: The blurring level at the first scan (blur index = 115); D: The blurring level at the second scan (blur index = 78); E: A heat map representation of the blurring level at the first scan; F: A heat map representation of the blurring level at the second scan.

non-tumor tissue included in the dataset, the greater the total number of flipped cases. Our dataset contained non-tumor tissue images 4.4 times more than cancerous tissue images, so the total number of flipped cases increased. Slide scanners have been broadly used to obtain DPIs for ML-aided image analysis, so the issue of blurring should be mentioned more in the implementation of DPI analysis and in the development of more robust ML-aided classification models.

This study had some limitations. First, the robustness of a classification model for DPIs differs depending on the objects being analyzed, the method of machine-learning, and the quality and quantity of the dataset for learning. Therefore, the issue mentioned above should not be overgeneralized. However, a classification model for medical images (including DPI) should be tested to find if image blur might reduce reproducibility of the classification model. Second, we only investigated differences in color and blur in this study, while there may be another potential cause of discordant classification.

In conclusion, our findings suggest that differences in the blur in paired DPIs from the same microscope slide could cause different classification results by an ML-aided classification model. If an ML model has sufficient robustness, these slight differences in DPI might not cause a different classification result. However, an ML-aided classification model for DPI should be tested for this potential cause of the reduced reproducibility of the model. Since our method provides a quantitative measure for the degree of blurring, it is possible to avoid discordance through excluding these disqualified slides using this measure. However, further experiments are required to establish more reliable measure together with other factors, for instance, such as tissue area size and nuclear densities. In a future study, we will develop a slide scanner and/or a preprocessing method that will minimize DPI blur.

ARTICLE HIGHLIGHTS

Research background

Little attention has been paid to the frequency and preventable causes of discordant classification results of digital pathological image (DPI) analysis using machine learning (ML) for the heterochronously obtained DPIs.

Research motivation

Authors compared the classification results between paired DPIs of the same microscope slide obtained from two independent scans using the same slide scanner.

Research objectives

In this study, the authors elucidated the frequency and preventable causes of discordant classification results of DPI analysis using ML for the heterochronously obtained DPIs.

Research methods

Authors created paired DPIs by scanning 298 hematoxylin and eosin stained slides containing 584 tissues twice with a virtual slide scanner. The paired DPIs were analyzed by our ML-aided classification model. Differences in color and blur between the non-flipped and flipped groups were compared by L1-norm and a blur index.

Research results

Discordant classification results in 23.1% of the paired DPIs obtained by two independent scans of the same microscope slide were observed. No significant difference in the L1-norm of each color channel between the two groups; however, the flipped group showed a significantly higher blur index than the non-flipped group.

Research conclusions

The results suggest that differences in the blur - not the color - of the paired DPIs may cause discordant classification results.

Research perspectives

An ML-aided classification model for DPI should be tested for this potential cause of the reduced reproducibility of the model. In a future study, a slide scanner and/or a preprocessing method of minimizing DPI blur should be developed.

ACKNOWLEDGEMENTS

Authors (Ogura M and Kiyuna T) would like to thank Dr. Yukako Yagi, Memorial Sloan Kettering Cancer Center, and Professor Masahiro Yamaguchi, Tokyo Institute of Technology, for their helpful comments and suggestions.

REFERENCES

- 1 **Al-Janabi S**, Huisman A, Van Diest PJ. Digital pathology: current status and future perspectives. *Histopathology* 2012; **61**: 1-9 [PMID: 21477260 DOI: 10.1111/j.1365-2559.2011.03814.x]
- 2 **Park S**, Parwani AV, Aller RD, Banach L, Becich MJ, Borkenfeld S, Carter AB, Friedman BA, Rojo MG, Georgiou A, Kayser G, Kayser K, Legg M, Naugler C, Sawai T, Weiner H, Winsten D, Pantanowitz L. The history of pathology informatics: A global perspective. *J Pathol Inform* 2013; **4**: 7 [PMID: 23869286 DOI: 10.4103/2153-3539.112689]
- 3 **Coudray N**, Ocampo PS, Sakellaropoulos T, Narula N, Snuderl M, Fenyö D, Moreira AL, Razavian N, Tsirigos A. Classification and mutation prediction from non-small cell lung cancer histopathology images using deep learning. *Nat Med* 2018; **24**: 1559-1567 [PMID: 30224757 DOI: 10.1038/s41591-018-0177-5]
- 4 **Cosatto E**, Laquerre PF, Malon C, Graf HP, Saito A, Kiyuna T, Marugame A, Kamijo K. Automated gastric cancer diagnosis on H&E-stained sections; Itraining a classifier on a large scale with multiple instance machine learning. Proceedings of SPIE 8676, Medical Imaging 2013: Digital Pathology, 867605; 2013 Mar 29; Florida, USA [DOI: 10.1117/12.2007047]
- 5 **Yoshida H**, Shimazu T, Kiyuna T, Marugame A, Yamashita Y, Cosatto E, Taniguchi H, Sekine S, Ochiai A. Automated histological classification of whole-slide images of gastric biopsy specimens. *Gastric Cancer* 2018; **21**: 249-257 [PMID: 28577229 DOI: 10.1007/s10120-017-0731-8]
- 6 **Yang G**, Nelson BJ. Wavelet-based autofocusing and unsupervised segmentation of microscopic images. Proceedings. Proceedings 2003 IEEE/RSJ International Conference on Intelligent Robots and Systems

(IROS 2003) (Cat. No.03CH37453); 2003 Oct 27-31; Las Vegas, USA. IEEE, 2003: 2143-2148 [DOI: [10.1109/IROS.2003.1249188](https://doi.org/10.1109/IROS.2003.1249188)]

- 7 **Aziz MA**, Nakamura T, Yamaguchi M, Kiyuna T, Yamashita Y, Abe T, Hashiguchi A, Sakamoto M. Effectiveness of color correction on the quantitative analysis of histopathological images acquired by different whole-slide scanners. *Artif Life Robotics* 2019; **24**: 28-37 [DOI: [10.1007/s10015-018-0451-0](https://doi.org/10.1007/s10015-018-0451-0)]



Published by **Baishideng Publishing Group Inc**
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: <https://www.f6publishing.com/helpdesk>

<https://www.wjgnet.com>

