

World Journal of *Gastrointestinal Oncology*

World J Gastrointest Oncol 2021 September 15; 13(9): 980-1212



REVIEW

- 980** Involvement of integrin-activating peptides derived from tenascin-C in colon cancer progression
Fujita M, Suzuki H, Fukai F
- 995** MicroRNA expression in inflammatory bowel disease-associated colorectal cancer
Grillo TG, Quaglio AEV, Beraldo RF, Lima TB, Baima JP, Di Stasi LC, Sasaki LY
- 1017** Association between intestinal neoplasms and celiac disease: A review
Wang M, Yu M, Kong WJ, Cui M, Gao F
- 1029** Real-time fluorescence image-guided gastrointestinal oncologic surgery: Towards a new era
Martínez-López E, Martínez-Pérez A, Navarro-Martínez S, Sebastián-Tomás JC, de'Angelis N, García-Granero E
- 1043** Neoadjuvant chemotherapy for colorectal liver metastases: A contemporary review of the literature
Guo M, Jin N, Pawlik T, Cloyd JM

MINIREVIEWS

- 1062** Review of incomplete macroscopic resections (R2) in rectal cancer: Treatment, prognosis and future perspectives
Pérez Lara FJ, Hebrero Jimenez ML, Moya Donoso FJ, Hernández Gonzalez JM, Pitarch Martinez M, Prieto-Puga Arjona T
- 1073** Potential utility of liquid biopsies in the management of patients with biliary tract cancers: A review
Shotton R, Lamarca A, Valle J, McNamara MG
- 1086** Conservative management of malignant gastric outlet obstruction syndrome-evidence based evaluation of endoscopic ultrasound-guided gastroentero-anastomosis
Cominardi A, Tamanini G, Brighi N, Fusaroli P, Lisotti A
- 1099** Overgrowth of *Lactobacillus* in gastric cancer
Li ZP, Liu JX, Lu LL, Wang LL, Xu L, Guo ZH, Dong QJ
- 1109** Evidence based tools to improve efficiency of currently administered oncotherapies for tumors of the hepatopancreatobiliary system
Herold Z, Szasz AM, Dank M
- 1121** Screening strategy for gastrointestinal and hepatopancreatobiliary cancers in cystic fibrosis
Hoskins B, Wasuwanich P, Scheimann AO, Karnsakul W
- 1132** Immune aspects of hepatocellular carcinoma: From immune markers for early detection to immunotherapy
Mattos ÁZ, Debes JD, Boonstra A, Vogel A, Mattos AA

- 1144 Characterization of metabolic landscape in hepatocellular carcinoma

Wu J, Xue R, Jiang RT, Meng QH

- 1157 Effect of oncometabolic surgery on gastric cancer: The remission of hypertension, type 2 diabetes mellitus, and beyond

Cheng YX, Peng D, Tao W, Zhang W

ORIGINAL ARTICLE

Basic Study

- 1164 Scoparone inhibits pancreatic cancer through PI3K/Akt signaling pathway

Li N, Yang F, Liu DY, Guo JT, Ge N, Sun SY

Retrospective Study

- 1184 Prognostic value of modified Lauren classification in gastric cancer

Ning FL, Zhang NN, Wang J, Jin YF, Quan HG, Pei JP, Zhao Y, Zeng XT, Abe M, Zhang CD

META-ANALYSIS

- 1196 Neoadjuvant chemotherapy without radiation as a potential alternative treatment for locally advanced rectal cancer: A meta-analysis

Wu P, Xu HM, Zhu Z

LETTER TO THE EDITOR

- 1210 Use of liquid biopsies in gastrointestinal cancers

Khachfe HH

ABOUT COVER

Editorial Board Member of *World Journal of Gastrointestinal Oncology*, Rossana Berardi, MD, PhD, Director, Full Professor, Medical Oncology, Università Politecnica delle Marche, Ancona 60126, Italy. r.berardi@staff.univpm.it

AIMS AND SCOPE

The primary aim of *World Journal of Gastrointestinal Oncology* (WJGO, *World J Gastrointest Oncol*) is to provide scholars and readers from various fields of gastrointestinal oncology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJGO mainly publishes articles reporting research results and findings obtained in the field of gastrointestinal oncology and covering a wide range of topics including liver cell adenoma, gastric neoplasms, appendiceal neoplasms, biliary tract neoplasms, hepatocellular carcinoma, pancreatic carcinoma, cecal neoplasms, colonic neoplasms, colorectal neoplasms, duodenal neoplasms, esophageal neoplasms, gallbladder neoplasms, etc.

INDEXING/ABSTRACTING

The WJGO is now indexed in Science Citation Index Expanded (also known as SciSearch®), PubMed, PubMed Central, and Scopus. The 2021 edition of Journal Citation Reports® cites the 2020 impact factor (IF) for WJGO as 3.393; IF without journal self cites: 3.333; 5-year IF: 3.519; Journal Citation Indicator: 0.5; Ranking: 163 among 242 journals in oncology; Quartile category: Q3; Ranking: 60 among 92 journals in gastroenterology and hepatology; and Quartile category: Q3. The WJGO's CiteScore for 2020 is 3.3 and Scopus CiteScore rank 2020: Gastroenterology is 70/136.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: *Ying-Yi Yuan*, Production Department Director: *Xiang Li*, Editorial Office Director: *Ya-Juan Ma*.

NAME OF JOURNAL

World Journal of Gastrointestinal Oncology

ISSN

ISSN 1948-5204 (online)

LAUNCH DATE

February 15, 2009

FREQUENCY

Monthly

EDITORS-IN-CHIEF

Rosa M Jimenez Rodriguez, Pashtoon Kasi, Monjur Ahmed, Florin Burada

EDITORIAL BOARD MEMBERS

<https://www.wjgnet.com/1948-5204/editorialboard.htm>

PUBLICATION DATE

September 15, 2021

COPYRIGHT

© 2021 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

<https://www.wjgnet.com/bpg/gerinfo/204>

GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjgnet.com/bpg/GerInfo/287>

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjgnet.com/bpg/gerinfo/240>

PUBLICATION ETHICS

<https://www.wjgnet.com/bpg/GerInfo/288>

PUBLICATION MISCONDUCT

<https://www.wjgnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

<https://www.wjgnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjgnet.com/bpg/GerInfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>



Involvement of integrin-activating peptides derived from tenascin-C in colon cancer progression

Motomichi Fujita, Hideo Suzuki, Fumio Fukai

ORCID number: Motomichi Fujita 0000-0001-7502-9797; Hideo Suzuki 0000-0002-1469-7449; Fumio Fukai 0000-0002-5646-167X.

Author contributions: All authors contributed equally to the conception of this study, drafting, critical revision, and editing of the manuscript, and final approval of the submission.

Conflict-of-interest statement: The authors declare no conflicts of interests in regards to this article.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Specialty type: Oncology

Country/Territory of origin: Japan

Motomichi Fujita, Fumio Fukai, Department of Molecular Patho-Physiology, Tokyo University of Science, Noda 278-8510, Chiba, Japan

Hideo Suzuki, Department of Gastroenterology, University of Tsukuba, Tsukuba 305-8575, Ibaraki, Japan

Corresponding author: Fumio Fukai, PhD, Professor, Department of Molecular Patho-Physiology, Tokyo University of Science, 2641 Yamazaki, Noda 278-8510, Chiba, Japan. fukai@rs.noda.tus.ac.jp

Abstract

Tenascin-C (TNC) is an adhesion modulatory protein present in the extracellular matrix that is highly expressed in several malignancies, including colon cancer. Although TNC is considered a negative prognostic factor for cancer patients, the substantial role of the TNC molecule in colorectal carcinogenesis and its malignant progression is poorly understood. We previously found that TNC has a cryptic functional site and that a TNC peptide containing this site, termed TNIIIA2, can potently and persistently activate beta1-integrins. In contrast, the peptide FNIII14, which contains a cryptic bioactive site within the fibronectin molecule, can inactivate beta1-integrins. This review presents the role of TNC in the development of colitis-associated colorectal cancer and in the malignant progression of colon cancer, particularly the major involvement of its cryptic functional site TNIIIA2. We propose new possible prophylactic and therapeutic strategies based on inhibition of the TNIIIA2-induced beta1-integrin activation by peptide FNIII14.

Key Words: Tenascin-C; TNIIIA2; Beta1-integrin; Integrin activation; Colitis-associated colorectal cancer; Colon cancer

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Exposure of the cryptic functional site TNIIIA2 from the Tenascin-C (TNC) molecule and its potent and sustained activation of beta1-integrins appear to be associated with the development of colon cancer and its malignant progression. Inhibition of the biological function of TNIIIA2 derived from TNC molecule may be a

Peer-review report's scientific quality classification

Grade A (Excellent): A

Grade B (Very good): 0

Grade C (Good): C

Grade D (Fair): 0

Grade E (Poor): 0

Received: February 21, 2021**Peer-review started:** February 21, 2021**First decision:** May 8, 2021**Revised:** June 3, 2021**Accepted:** August 11, 2021**Article in press:** August 11, 2021**Published online:** September 15, 2021**P-Reviewer:** El-Nakeep S, Shivaji UN**S-Editor:** Fan JR**L-Editor:** A**P-Editor:** Li X

promising strategy for the prevention and treatment of colon cancer.

Citation: Fujita M, Suzuki H, Fukai F. Involvement of integrin-activating peptides derived from tenascin-C in colon cancer progression. *World J Gastrointest Oncol* 2021; 13(9): 980-994**URL:** <https://www.wjgnet.com/1948-5204/full/v13/i9/980.htm>**DOI:** <https://dx.doi.org/10.4251/wjgo.v13.i9.980>

INTRODUCTION

Extracellular matrix (ECM) proteins such as fibronectin (FN), collagen, and laminin provide a scaffold for cell adhesion and subsequently influence various physiological cellular processes, including cell differentiation, survival/proliferation, and migration. As one of the major components of the tumor microenvironment, the ECM affects the behavior of cells in the cancer microenvironment, such as cancer-associated fibroblasts (CAFs) and immune cells, resulting in cancer development[1]. It therefore plays major roles in carcinogenesis and the malignant progression of cancer.

Integrins are a family of heterodimeric transmembrane glycoproteins composed of alpha- and beta-subunits that directly interact with components of the ECM. These integrins primarily mediate cell adhesion, migration, survival, proliferation, and differentiation. In contrast to membrane receptors for humoral factors such as cytokines and chemokines, integrins are unique in their ability to alter the binding affinity for ECM ligands. Integrins exist mainly in two different structural states, an inactive conformation lacking ligand-binding affinity and an active one with high affinity[2]. On the other hand, integrin signaling contributes to the malignant progression of many cancers. For example, integrin alpha5beta1, a major FN receptor, is highly expressed in glioma/glioblastoma, with its expression levels reported to be associated with poor survival in glioma/glioblastoma patients[3]. Alpha5-integrin promotes cell proliferation and the dissemination of glioblastoma cells[4], modulates angiogenesis[5], and contributes to temozolomide chemoresistance[6]. Thus, the integrin alpha5beta1-mediated adhesive interaction of glioma cells may be associated with the acquisition of a highly aggressive phenotype in glioma/glioblastoma. Therefore, inhibition of integrin functions might be a promising therapeutic approach for cancer.

Tenascin-C (TNC) is a hexameric, multimodular ECM glycoprotein. It is poorly expressed in normal adult tissues but highly expressed in both inflammatory lesions and the tumor microenvironment[3,7-10]. TNC is an endogenous activator of toll-like receptor 4, which triggers and amplifies inflammatory responses[11]. In addition, TNC binds to integrin alphavbeta3 and alpha9beta1 to drive inflammatory responses by inducing the synthesis of proinflammatory cytokines, including interleukin (IL)-6, IL-1beta, and tumor necrosis factor-alpha[12]. TNC is highly expressed and is thought to act as a major driving regulator of acute and chronic inflammatory diseases, including cardiac disease[13], arthritis[14], nephritis[15], sepsis[16], stroke[17], asthma[18], chronic obstructive pulmonary disease[19], and viral infections[20]. Therefore, TNC may be a promising biomarker of disease activity and a therapeutic target in these inflammatory diseases.

Furthermore, the expression levels of TNC are associated with poor prognosis in patients with malignant tumors, such as glioma and breast and colon cancers[3,8,10]. Accumulating evidence indicates a relationship between TNC and tumor progression. For example, TNC plays key roles in several processes of tumor progression related to proliferation[21,22], migration, invasion[23-25], angiogenesis[26,27], immunosuppression[28,29], cancer stemness[30,31], and apoptosis resistance[32], supporting the belief that TNC contributes to cancer progression and aggression. In addition, TNC has been linked to carcinogenesis[33-36]. Analysis of the Rip1-Tag2 model of pancreatic beta-cell carcinogenesis, which drives a multistage carcinogenesis process, revealed that TNC contributes to multiple steps linked to carcinogenesis[34,35]. Moreover, Li *et al*[36] revealed that the expression levels of TNC are higher in adenomatous colon polyps and colon carcinoma *in situ* than in non-neoplastic colonic mucosa and are also correlated with TMN stages of colon cancer, further indicating that TNC might contribute to carcinogenesis and progression[36].

TNC contains several characteristic domains, such as a central domain, heptad repeats, epidermal growth factor (EGF)-like repeats, FN type III repeats (FN-III repeats), and a fibrinogen globe (Figure 1), which can interact with ECM proteins, soluble factors, and cell receptors and express various functions of TNC. In addition, human TNC contains nine alternative splicing sites in FN-III repeats, and 511 possible splice variants can theoretically be generated through alternative splicing[37]. This alternative splicing could control the versatile biological functions of TNC by modulating its interaction with specific binding partners, as well as by exposing post-translational sites and proteolytic cleavage sites[37]. However, the substantial role of the TNC molecule in colorectal carcinogenesis and its malignant progression has remained elusive.

This review presents the role of TNC in the malignant progression of colon cancer and the development of colitis-associated colorectal cancer (CAC), with a particular focus on the major involvement of TNIIIA2, the cryptic functional site of TNC. We propose new possibilities for prophylactic and therapeutic strategies based on peptide FNIII14-mediated inhibition of the TNIIIA2-induced beta1-integrin activation.

PATHOLOGICAL SIGNIFICANCE OF ELEVATED TNC EXPRESSION IN MALIGNANT TUMORS

Most ECM proteins harbor functionally cryptic functional sites that are buried within their molecular structures. These cryptic sites, called matricryptic sites, are revealed *via* structural/conformational changes triggered by interactions with adjacent cells or other ECM components and by remodeling/processing by ECM-degrading proteinases, including matrix metalloproteinases (MMPs) and cathepsins. The proteinases capable of degrading ECM proteins are highly upregulated in a wide variety of cancers[38-40]. ECM degradation often occurs in malignant tumors, and ECM protein fragments with biological functions are released through cleavage by inflammatory proteinases[41,42]. ECM fragments with functional matricryptins show unique biological functions that are not detected in their parental ECM proteins[43]. ECM proteins such as TNC are proteolytically cleaved by several inflammatory proteinases, including MMPs and cathepsins[39,40,42-44]. Proteolytic degradation of TNC has been detected in lung and colon cancer, and early-stage non-small cell lung cancer patients with TNC degradation show significantly worse prognosis and higher recurrence than those without TNC degradation[39,45,46]. Increased MMP-2 activity has been observed in patients with degraded TNC[39], indicating that exposure of the TNC functional cryptic site by several inflammatory proteinases may be associated with the malignant progression of cancer. Saito *et al*[47] previously found that TNC harbors a cryptic and functional site comprising the amino acid residues in the sequence YTITIRGV within the FN type III repeat A2[47]. A 22-mer peptide containing this functional sequence of TNC, termed peptide TNIIIA2, can potentially activate beta1-integrins, a state that is sustained for a long period of time[48] (Figure 1).

The mode of beta1-integrin activation induced by TNIIIA2 is entirely distinct from that induced by “inside-out” signaling, which is the commonly considered mode of integrin activation. Saito *et al*[47] have found that syndecan-4, one of the transmembrane heparin sulfate proteoglycans, serves as a membrane receptor for TNIIIA2 and that engagement with TNIIIA2 induces a lateral association with beta1-integrins, resulting in stabilization of the active conformation of beta1-integrin[47]. Based on this unique mechanism of integrin activation, this TNIIIA2-induced integrin activation is more potent and persistent than other known integrin activators, such as the various cytokines and chemokines that stimulate the “inside-out” signaling pathway[48]. Because TNC variants containing the alternatively spliced domain type III-A2 are highly expressed in malignant tumors[49], the activation of beta1-integrin induced by TNIIIA2 may be related to some forms of cancer pathogenesis. We previously found that TNIIIA2 contributes to the ability of glioblastoma to acquire aggressive properties such as excessive survival/proliferation, disseminative migration, and anoikis resistance through activation of beta1-integrin[50-52]. More recently, we reported that TNIIIA2 establishes inflammatory environments *via* the NOD-like receptor family pyrin domain-containing 3/caspase-1/IL-1beta pathway[53]. These findings suggest that the pathological significance of high TNC expression in inflammation and cancer may lie in activating beta1-integrins based on TNIIIA2 function.

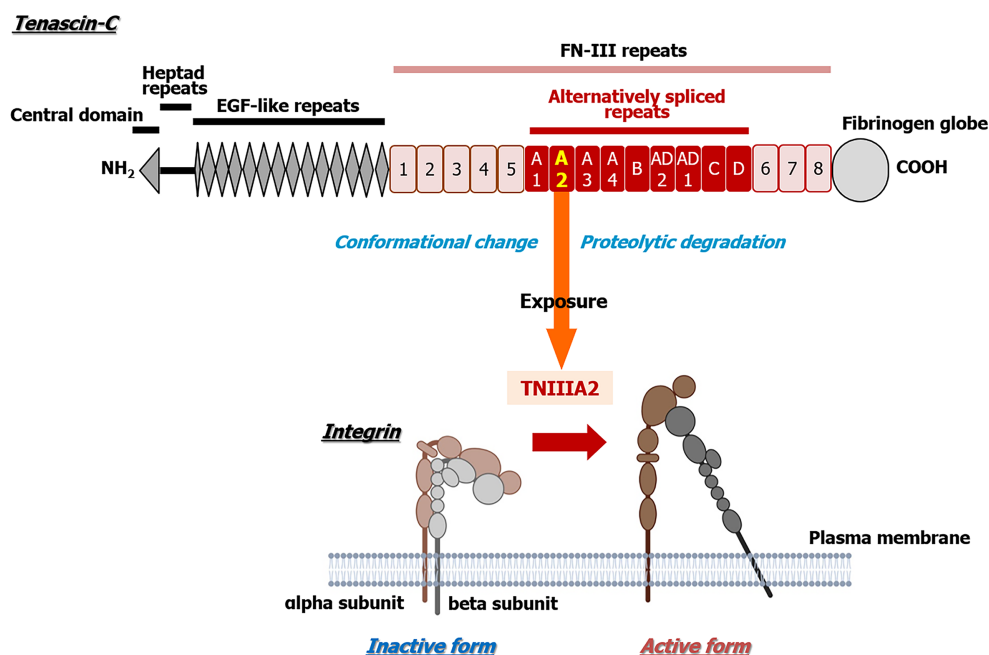


Figure 1 Schematic illustration of Tenascin-C and the amino acid sequence of peptide TNIIIA2. Conformational change in integrin activation via the lateral interaction of integrin with syndecan-4 at peptide TNIIIA2. Created with BioRender.com.

INVOLVEMENT OF TNC IN COLON CANCER

Colorectal cancer is the third most common type of gastrointestinal tract tumor worldwide and the third leading cause of death among men and women[54]. Because of recent substantial progress in diagnostic methods and advances in primary and adjuvant treatments, including standard chemotherapy and targeted treatments, the incidence and mortality of colorectal cancer has been improving[55,56], with a 5-year overall survival rate for colorectal cancer of about 60%[57,58]. However, patients with metastatic colorectal cancer, which comprise 20% of patients with new colorectal cancer diagnoses, show a high mortality rate, with a 5-year overall survival rate of approximately 20%[59-62]. Recently, systemic therapy involving molecular targeted drugs as well as cytotoxic drugs has been adopted for unresectable colorectal cancer. Combination with molecular targeted drugs such as bevacizumab, cetuximab, or panitumumab is recommended, depending on the RAS status[63]. However, although these drugs are effective, they have various problems, including certain adverse events, and eventually become ineffective. Therefore, further investigation is still necessary to develop novel strategies for colorectal cancer, and it is important to elucidate the molecular mechanisms that enable colorectal cancer to acquire malignant properties.

TNC is highly expressed in colon cancer, and high expression levels of TNC in tissue specimens are correlated with distant metastasis, tumor recurrence, advanced TNM stage, and poor prognosis[10,36]. Moreover, colon cancer cells highly expressing TNC show high metastatic potential and are associated with lymph nodes with metastasis[36]. In addition, serum TNC levels, particularly those of large-spliced variants, are higher in patients with colon cancer compared with controls[64]. Such levels are also correlated with tumor depth, lymph node metastasis, and disease progression[64]. Therefore, the levels of TNC in tissue and serum may be a diagnostic or prognostic biomarker in colon cancer. Furthermore, the Wnt/beta-catenin signaling pathway plays a central role in carcinogenesis, and its mutation and activation are found in almost all patients with colon cancers[65]. Because TNC is a Wnt/beta-catenin target gene in human colon tumors[66], the deregulation of Wnt/beta-catenin signaling might lead to the overexpression of TNC in colon cancer. Experimental observations indicated that TNC secreted by myofibroblasts might act as a proinvasive factor for colon cancer cells[67]. Furthermore, TNC promotes proliferation, migration, and invasion and also upregulates cancer stem cell markers *via* the Hedgehog signaling pathway[31]. However, the biochemical functions of TNC in the malignant progression of colon cancer have not yet been established.

MMP-2 is highly expressed in colon cancer tissues and its expression levels increase with an increase in the tumor stage[68]. Furthermore, the expression levels of MMP-2 are correlated with lymph vessel invasion and disease progression in colon cancer[69]. MMP-7 is another Wnt/beta-catenin target gene[70] and both MMP-2 and MMP-7 can degrade TNC[71]. Furthermore, TNC variants containing the alternatively spliced domain types III-A1, -2, and -4 are highly expressed in colon cancer[49]. It is presumed that the functional cryptic site TNIIIA2 of TNC may be released into the tumor microenvironment of colon cancer and contribute to its pathogenesis. Supporting this hypothesis, peptide TNIIIA2 has been shown to act directly on colon cancer cells to enhance their *in vitro* invasive potential by inducing MMP secretion[72]; peptide TNIIIA2 or TNC promotes colon cancer cell invasion by upregulating MMPs[72]. The cell invasion induced by peptide TNIIIA2 or TNC is completely suppressed by anti-TNIIIA2 antibody or MMP-2 inhibitor[72]. Moreover, an *in vivo* observation involving a spontaneous metastasis mouse model mimicking hematogenous metastasis exhibited that peptide TNIIIA2 boosted the metastasis of colon cancer cells to the lung[72]. Taken together, the activation of beta1-integrin by peptide TNIIIA2 (one of the biochemical functions of TNC) may help to promote colon cancer cell metastasis *via* induction of MMP (Figure 2).

Alterations in the density, distribution, and composition of the ECM are common in malignancies. This process creates the tumor microenvironment that helps to confer cancer cells with malignant properties such as tumorigenesis and metastasis[1]. These alterations increase stiffness in the tumor microenvironment, which promotes pro-tumorigenic mechanosignaling. The increased ECM stiffness of colon cancer has been associated with cancer progression[73]. Through analysis of clinical specimens, a gradient of increasing ECM stiffness was observed from healthy to perilesional and colon cancer areas, which might predispose invasion[74]. Furthermore, the expression levels of lysyl oxidase (LOX), which catalyzes the covalent cross-linking of collagens and elastin, are closely correlated with the progression of colon cancer[75]. Compared with control cells or cells expressing a catalytically inactive LOX, colon cancer cells expressing LOX exhibit increased mechanosignaling, ECM stiffness, metastasis, and tumor burden in *in vivo* models *via* activation of beta1-integrin and the focal adhesion kinase-SRC signaling pathway[76], indicating that beta1-integrin activation might be associated with malignant progression *via* increased ECM stiffness in colon cancer. In a recent insightful study on the role of TNC in ECM stiffness in the tumor microenvironment, Barnes *et al*[77] demonstrated that the glycocalyx/ECM-integrin loop induces glioblastoma aggression in a tissue tension-dependent manner, with human recurrent glioblastomas showing an increase in TNC-enriched stiffened ECM and enhanced integrin mechanosignaling[77]. It has also been pointed out that glioblastoma cells expressing a V737N beta1-integrin autoclustering mutant exhibit increased mechanosignaling and ECM stiffness and facilitate tumor growth[77]. It is unlikely that at least the antiadhesive effect of TNC, which has been considered a major biochemical function of this protein, is responsible for the ECM stiffening and consequent enhanced integrin signaling. However, it remains unclear whether proadhesive activity (a biochemical function of TNC) is directly associated with ECM stiffness in the tumor microenvironment of colon cancer. Further investigations are required to determine whether activation of beta1-integrin by peptide TNIIIA2 could actually increase ECM stiffness in colon cancer.

Beta1-integrin is also highly expressed in colon cancer compared with normal mucosa. High expression levels of beta1-integrin have been associated with poor prognosis, and increased expression of beta1-integrin is independently correlated with decreased overall survival and disease-free survival in colon cancer patients[78]. In addition, alpha5-integrin, which is coupled with beta1-integrin, also shows upregulated expression in colon cancer and is expressed mainly in the tumor stroma of clinical samples[79]. Moreover, alpha5beta1-integrin expression is considered a significant independent prognostic factor. Experimental evidence indicates that overexpression of alpha5-integrin accelerates proliferation and suppresses apoptosis in colon cancer cells, with colon cancer cells overexpressing alpha5-integrin found to promote tumor growth in a murine xenograft tumor model[80]. In addition, blockade of alpha5-integrin inhibits cell attachment and induces apoptosis in colon cancer cells *via* Akt suppression[81]. Integrin alpha5beta1 also confers anoikis resistance in colon cancer cells *via* association with EGF receptor and the subsequent activation of ERK and Akt as well as suppression of the caspase signaling pathway[82]. Furthermore, depletion of alpha5-integrin expression in fibroblasts suppresses the tumorigenic activity of colon cancer in *in vivo* experiments, as determined by the co-injection of human colon cancer cells and human normal colonic fibroblast cells into immunocompromised mice[79]. This result indicated that CAFs expressing alpha5-integrin have a

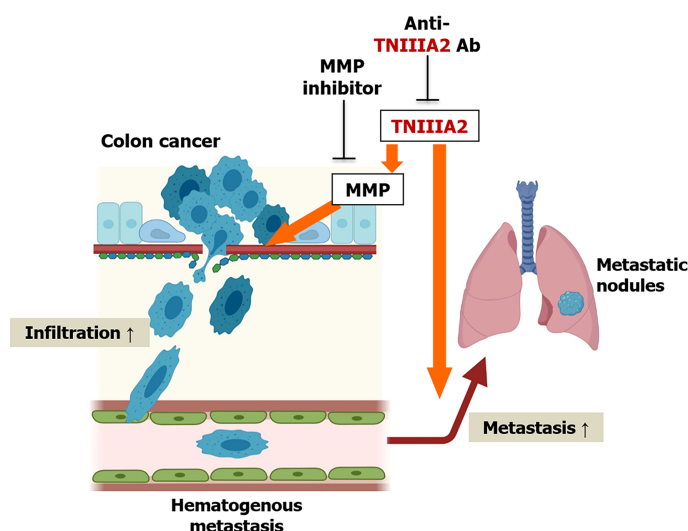


Figure 2 Schematic model of the effect of TNIIIA2 on the metastasis of colon cancer cells. Peptide TNIIIA2 boosts the infiltration of colon cancer cells via matrix metalloproteinases production *in vitro* and promotes pulmonary metastasis in a spontaneous metastasis mouse model. Created with BioRender.com. MMP: Matrix metalloproteinases.

tumor-promoting effect in colon cancer. Pharmacological experiments have demonstrated that the non-peptidic $\alpha 5 \beta 1$ integrin antagonist K34c suppresses the clonogenic survival of colon cancer cells[83]. In addition, ATN-161, a peptidic antagonist of integrin $\alpha 5 \beta 1$ and $\alpha v \beta 3$, reduced tumor vascularization, and combination therapy of ATN-161 and fluorouracil suppressed liver metastases in a murine model of colon cancer[84]. Therefore, integrin $\alpha 5 \beta 1$ might be a promising target for cancer therapeutics.

INVOLVEMENT OF TNC IN CAC

A link between chronic inflammation and the pathogenesis of many malignancies has been well documented. Examples include *Helicobacter pylori* infection-associated gastric cancer and hepatitis virus infection-associated hepatocellular carcinoma[85,86]. In particular, inflammatory bowel disease (IBD) patients, including Crohn's disease and ulcerative colitis, have an increased risk of developing CAC[87-89], which is a subtype of colorectal cancer[90]. Although the incidence of CAC seems to have decreased in recent years because of more frequent surveillance, improved surveillance techniques, and more effective IBD drugs for controlling inflammation, patients with IBD still have higher rates of death from colon cancer[91]. Indeed, a Scandinavian population-based study recently showed that patients with IBD and colorectal cancer had an increased risk of mortality compared with those with sporadic colorectal cancer[92,93]. Therefore, there is still an unmet medical need for the prevention and treatment of CAC. Unlike sporadic colorectal cancer, CAC onset does not show an adenoma-carcinoma sequence, but rather an inflammation-dysplasia-carcinoma sequence[94,95]. Nonetheless, the molecular basis of CAC onset is largely unknown. Thus, research into the molecular mechanisms underlying CAC onset is urgently needed for the development of novel therapeutics.

Several studies have reported that TNC is associated with ulcerative colitis and Crohn's disease[96-99]. A genome-wide association study of African Americans found that single-nucleotide polymorphisms within the TNC gene are associated with IBD risk[100]. Ning *et al*[101] reported that TNC is highly expressed in the inflamed stromal area of the intestinal mucosa of IBD patients[101]. They also showed particularly high levels of serum TNC in patients with severe IBD compared with those with mild or moderate IBD[101]. Riedl *et al*[96] determined that the serum levels of TNC are correlated with clinical and histological parameters of disease activity in IBD patients[96]. Moreover, high levels of TNC mRNA in the mucosa of ulcerative colitis have been associated with a poor response to infliximab therapy, an effective treatment for moderate-to-severe IBD, indicating that TNC may contribute to therapeutic resistance against IBD. Therapy resistance may participate in the malignant progression of IBD due to a lack of inflammatory control, resulting in an increased risk of CAC onset.

Indeed, TNC derived from intestinal myofibroblasts promotes the onset of CAC in an azoxymethane (AOM)/dextran sulfate sodium (DSS) model *via* angiogenesis[102]. Thus, TNC might contribute to the development and/or malignant progression of CAC. Identification of the biological functions of the TNC responsible for the development of CAC would enable the design of agents with prophylactic and therapeutic potential for these diseases. However, the biochemical functions of TNC in CAC onset have not yet been established.

ECM remodeling is often augmented in these pathological lesions, and proteolytic cleavage of ECM proteins is performed by several inflammatory proteinases, including MMPs and cathepsins. Indeed, increased expression levels of several MMPs have been observed in IBD and are associated with disease activity in IBD, indicating that degradation of the ECM, including TNC, might occur at high levels in IBD and during CAC onset[103]. Therefore, it is conceivable that the functional cryptic site TNIIIA2 might be exposed by the high levels of TNC molecules in the lesion and act as a specific pathogenic factor in the development of CAC. Supporting this assumption, our recent work demonstrated the presence of TNC and peptide TNIIIA2 in the stromal area of dysplastic lesions in AOM/DSS mice[104]. Assuming that peptide TNIIIA2 acts mainly on preneoplastic epithelial cells and fibroblasts, which are abundant in the stromal area of dysplastic lesions, our *in vitro* experiments focused on the effects of beta1-integrin activation on both preneoplastic epithelial cells and fibroblasts. Interestingly, although beta1-integrin activation by peptide TNIIIA2 promoted cell adhesion, it had no direct effect on the growth of preneoplastic epithelial cells[104]. Similarly, peptide TNIIIA2 had no direct effect on the growth of fibroblasts, but fibroblasts stimulated by peptide TNIIIA2 released humoral factors, or possibly factors, that drove the malignant transformation of premalignant epithelial cells in a paracrine manner, as judged by anchorage-independent cell growth and focus formation[104]. These factors secreted from peptide TNIIIA2-activated fibroblasts are also able to promote the survival/proliferation of colon cancer cells [104]. Furthermore, peptide FNIII14, a peptidic factor that induces a conformational change in beta1-integrin from the active to the inactive state[105], suppressed not only the TNIIIA2-induced dysregulated survival/proliferation of preneoplastic epithelial cells *in vitro*, but also polyp development in an AOM/DSS mouse model[104]. These results suggest that beta1-integrin activation by peptide TNIIIA2 in fibroblasts may be an important target for the prevention of CAC (Figure 3).

Several studies have demonstrated that cells in the tumor microenvironment, such as CAFs and immune cells, influence tumor progression. Among them, CAFs are key determinants of cancer development and progression[106-108]. Sasaki *et al*[109] demonstrated that CAC incidence is abrogated in CC chemokine ligand 3- or CC chemokine receptor 5-knockout mice treated with AOM/DSS and coincides with lower accumulation of fibroblasts in dysplastic lesions compared with wild-type mice [109]. These fibroblasts express heparin-binding EGF-like growth factor to stimulate the proliferation of tumor cells in CAC in mice[109]. In addition, epiregulin derived from fibroblast promotes the proliferation of intestinal epithelial cells through activation of the ERK signaling pathway, augmenting CAC growth[110]. These studies indicate that CAFs might be responsible for CAC development and progression. However, there is increasing evidence that TNC is upregulated in CAFs and that a high TNC expression as a CAF marker in tumor stroma is correlated with worse prognosis in several malignancies, such as breast ductal carcinoma[7], esophageal squamous cell carcinoma[9], colorectal cancer[10], and prostate cancer[111]. Taken together with our results, the evidence indicates that fibroblasts produce TNC in the tumor microenvironment and that this TNC might activate CAFs to promote tumor onset and progression.

Risk factors for CAC development include pancolitis, a younger age of IBD onset, a long disease duration, chronic cholestatic liver disease, family history[112], and stricture formation[113]. Intestinal fibrosis is a common complication in IBD, particularly Crohn's disease, and the resulting clinically relevant strictures have been observed in about one-third of patients[114]. Intestinal fibrosis is likely to involve increased ECM stiffness, and this stiffness could perpetuate fibrogenesis[114], leading to the development of fibrotic strictures. More recently, accumulating evidence has linked increased ECM stiffness to several malignancies, with recent studies showing that cancer progression and aggression are correlated with the stiffness of a TNC-enriched ECM[115] (please see the previous section). In IBD, increased ECM stiffness has been observed in strictures, and the increased ECM stiffness enhances adhesive properties, such as the formation of focal adhesion and actin stress fibers of colonic fibroblasts[116]. Moreover, increased expression levels of TNC have been reported in lesions of ulceration in ulcerative colitis[98]. Erdem *et al*[117] reported the possible

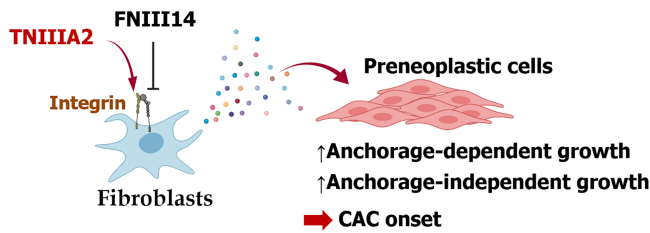


Figure 3 Schematic model of the effect of TNIIIA2 on colitis-associated colorectal cancer onset. Peptide TNIIIA2 stimulates fibroblasts to secrete humoral factors, promoting anchorage-dependent and -independent growth in preneoplastic cells. Created with BioRender.com. CAC: Colitis-associated colorectal cancer.

involvement of increased expression levels of TNC in the development of ulcerative colitis-related strictures[117]. Given that peptide TNIIIA2 can induce potent and persistent activation of beta1-integrin as well as its clustering[47,48], peptide TNIIIA2 in stromal lesions might contribute to the development of colitis-related strictures through increased ECM stiffness, leading to increased risk of CAC onset. Although further research is required to determine whether beta1-integrin activation by peptide TNIIIA2 actually increases ECM stiffness, TNIIIA2-targeting agents such as an anti-TNIIIA2 antibody might be a promising strategy for the prophylaxis or treatment of CAC development and malignant progression.

Several studies have suggested that integrin inactivation could be a promising strategy for controlling CAC development and progression. ATN-161, a peptidic antagonist of integrin alpha5beta1 and alphavbeta3, suppressed disease activity by blocking angiogenesis in IL-10-deficient mice that develop spontaneous Crohn's disease-like colitis[118] as well as in a CD4⁺CD45RB^{high} T-cell transfer model that induced chronic pancolitis[119]. Furthermore, ATN-161 also inhibits CAC development *via* inhibition of integrin alphavbeta3-mediated angiogenesis in a chemically induced AOM/DSS mouse model of intestinal and colon carcinogenesis[102], although no recent development status of ATN-161 is available. More recently, Terasaki *et al* [120] showed that fucoxanthin induces anoikis in colonic adenocarcinoma through attenuation of beta1-integrin signaling, which blocks CAC development in AOM/DSS mice[120]. Taken together, beta1-integrin activation might become a promising target for preventing and treating CAC, and inactivation of beta1-integrin by peptide FNIII14, which can neutralize the detrimental effects of peptide TNIIIA2 on beta1-integrin activation, might be a novel and promising strategy for the management of CAC development and malignant progression.

BETA1-INTEGRINS AS POTENTIAL THERAPEUTIC TARGETS IN COLON CANCER

Several antagonists of integrin alpha5beta1 and alphavbeta3 were well tolerated in clinical testing[121,122] but failed to show therapeutic benefits in patients with malignancies. Although inhibition of integrin alpha5beta1 and alphavbeta3 might be a safe therapeutic strategy, alternative approaches should be considered, including the application of integrin inhibitors as anti-cancer drugs (reviewed in Ref.[123]). Regarding other therapeutic modalities, OS2966, a humanized monoclonal antibody targeting human beta1-integrins, is undergoing testing in a phase I clinical trial for the treatment of recurrent/progressive glioma[124]. In addition, one possible strategy may be to develop drugs with modes of inhibition other than competitive inhibition of integrin. Unlike integrin antagonists, peptide FNIII14 – which has the ability to induce a conformational change in beta1-integrin from the active to the inactive state[105] – has shown therapeutic efficacy against several malignancies in animal models, including CAC, glioblastoma, neuroblastoma, and acute myelogenous leukemia[105]. Although further research is needed regarding its effect on the malignant progression of colon cancer, peptide FNIII14 may possess promising therapeutic properties.

CONCLUSION

Although TNC is considered a negative prognostic factor in several malignancies, the

substantial role of TNC molecule in the development of colorectal cancer and its malignant progression has remained elusive. We suggest that one of the pathological roles of TNC, which is highly expressed in colon cancer, may be in activating beta1-integrins through TNIIIA2 function. This hypothesis and the previous findings open the door to prophylactic and therapeutic strategies for colon cancer that involve inhibition of TNIIIA2-induced beta1-integrin activation by peptide FNIIII14.

REFERENCES

- 1 **Winkler J**, Abisoye-Ogunniyan A, Metcalf KJ, Werb Z. Concepts of extracellular matrix remodelling in tumour progression and metastasis. *Nat Commun* 2020; **11**: 5120 [PMID: [33037194](#) DOI: [10.1038/s41467-020-18794-x](#)]
- 2 **Shattil SJ**, Kim C, Ginsberg MH. The final steps of integrin activation: the end game. *Nat Rev Mol Cell Biol* 2010; **11**: 288-300 [PMID: [20308986](#) DOI: [10.1038/nrm2871](#)]
- 3 **Midwood KS**, Hussenet T, Langlois B, Orend G. Advances in tenascin-C biology. *Cell Mol Life Sci* 2011; **68**: 3175-3199 [PMID: [21818551](#) DOI: [10.1007/s00018-011-0783-6](#)]
- 4 **Blandin AF**, Noulet F, Renner G, Mercier MC, Choulier L, Vauchelles R, Ronde P, Carreiras F, Etienne-Selloum N, Vereb G, Lelong-Rebel I, Martin S, Dontenwill M, Lehmann M. Glioma cell dispersion is driven by $\alpha 5$ integrin-mediated cell-matrix and cell-cell interactions. *Cancer Lett* 2016; **376**: 328-338 [PMID: [27063097](#) DOI: [10.1016/j.canlet.2016.04.007](#)]
- 5 **Dudvarski Stanković N**, Bicker F, Keller S, Jones DT, Harter PN, Kienzle A, Gillmann C, Arnold P, Golebiewska A, Keunen O, Giese A, von Deimling A, Bäuerle T, Niclou SP, Mittelbronn M, Ye W, Pfister SM, Schmidt MHH. EGFL7 enhances surface expression of integrin $\alpha_5\beta_1$ to promote angiogenesis in malignant brain tumors. *EMBO Mol Med* 2018; **10** [PMID: [30065025](#) DOI: [10.15252/emmm.201708420](#)]
- 6 **Janouskova H**, Maglott A, Leger DY, Bossert C, Noulet F, Guerin E, Guenot D, Pinel S, Chastagner P, Plenat F, Entz-Werle N, Lehmann-Che J, Godet J, Martin S, Teisinger J, Dontenwill M. Integrin $\alpha 5\beta 1$ plays a critical role in resistance to temozolomide by interfering with the p53 pathway in high-grade glioma. *Cancer Res* 2012; **72**: 3463-3470 [PMID: [22593187](#) DOI: [10.1158/0008-5472.CAN-11-4199](#)]
- 7 **Yang Z**, Ni W, Cui C, Fang L, Xuan Y. Tenascin C is a prognostic determinant and potential cancer-associated fibroblasts marker for breast ductal carcinoma. *Exp Mol Pathol* 2017; **102**: 262-267 [PMID: [28223108](#) DOI: [10.1016/j.yexmp.2017.02.012](#)]
- 8 **Ishihara A**, Yoshida T, Tamaki H, Sakakura T. Tenascin expression in cancer cells and stroma of human breast cancer and its prognostic significance. *Clin Cancer Res* 1995; **1**: 1035-1041 [PMID: [9816077](#)]
- 9 **Yang ZT**, Yeo SY, Yin YX, Lin ZH, Lee HM, Xuan YH, Cui Y, Kim SH. Tenascin-C, a Prognostic Determinant of Esophageal Squamous Cell Carcinoma. *PLoS One* 2016; **11**: e0145807 [PMID: [26731558](#) DOI: [10.1371/journal.pone.0145807](#)]
- 10 **Yang Z**, Zhang C, Qi W, Cui C, Cui Y, Xuan Y. Tenascin-C as a prognostic determinant of colorectal cancer through induction of epithelial-to-mesenchymal transition and proliferation. *Exp Mol Pathol* 2018; **105**: 216-222 [PMID: [30170017](#) DOI: [10.1016/j.yexmp.2018.08.009](#)]
- 11 **Midwood K**, Sacre S, Piccinini AM, Inglis J, Trebaul A, Chan E, Drexler S, Sofat N, Kashiwagi M, Orend G, Brennan F, Foxwell B. Tenascin-C is an endogenous activator of Toll-like receptor 4 that is essential for maintaining inflammation in arthritic joint disease. *Nat Med* 2009; **15**: 774-780 [PMID: [19561617](#) DOI: [10.1038/nm.1987](#)]
- 12 **Marzeda AM**, Midwood KS. Internal Affairs: Tenascin-C as a Clinically Relevant, Endogenous Driver of Innate Immunity. *J Histochem Cytochem* 2018; **66**: 289-304 [PMID: [29385356](#) DOI: [10.1369/0022155418757443](#)]
- 13 **Imanaka-Yoshida K**, Tawara I, Yoshida T. Tenascin-C in cardiac disease: a sophisticated controller of inflammation, repair, and fibrosis. *Am J Physiol Cell Physiol* 2020; **319**: C781-C796 [PMID: [32845719](#) DOI: [10.1152/ajpcell.00353.2020](#)]
- 14 **Hasegawa M**, Yoshida T, Sudo A. Tenascin-C in Osteoarthritis and Rheumatoid Arthritis. *Front Immunol* 2020; **11**: 577015 [PMID: [33101302](#) DOI: [10.3389/fimmu.2020.577015](#)]
- 15 **Izumi K**, Miyazaki N, Okada H, Tsujimoto A, Matsumoto-Miyazaki J, Naito J, Yoshida G, Murata I, Nagashima K, Ohno M, Imanaka-Yoshida K, Okura H, Ohashi H, Takemura G. Tenascin-C expression in renal biopsies from patients with tubulointerstitial nephritis and its relation to disease activity and prognosis. *Int J Clin Exp Pathol* 2020; **13**: 1842-1852 [PMID: [32782713](#)]
- 16 **Yuan W**, Zhang W, Yang X, Zhou L, Hanghua Z, Xu K. Clinical significance and prognosis of serum tenascin-C in patients with sepsis. *BMC Anesthesiol* 2018; **18**: 170 [PMID: [30442110](#) DOI: [10.1186/s12871-018-0634-1](#)]
- 17 **Okada T**, Suzuki H. The Role of Tenascin-C in Tissue Injury and Repair After Stroke. *Front Immunol* 2020; **11**: 607587 [PMID: [33552066](#) DOI: [10.3389/fimmu.2020.607587](#)]
- 18 **Yasuda M**, Harada N, Harada S, Ishimori A, Itoigawa Y, Matsuno K, Makino F, Ito J, Ono J, Tobino K, Akiba H, Atsuta R, Izuhara K, Takahashi K. Characterization of tenascin-C as a novel biomarker for asthma: utility of tenascin-C in combination with periostin or immunoglobulin E. *Allergy Asthma Clin Immunol* 2018; **14**: 72 [PMID: [30473714](#) DOI: [10.1186/s13223-018-0300-7](#)]

- 19 **Löfdahl M**, Kaarteenaho R, Lappi-Blanco E, Tornling G, Sköld MC. Tenascin-C and alpha-smooth muscle actin positive cells are increased in the large airways in patients with COPD. *Respir Res* 2011; **12**: 48 [PMID: [21496259](#) DOI: [10.1186/1465-9921-12-48](#)]
- 20 **Mills JT**, Schwenzer A, Marsh EK, Edwards MR, Sabroe I, Midwood KS, Parker LC. Airway Epithelial Cells Generate Pro-inflammatory Tenascin-C and Small Extracellular Vesicles in Response to TLR3 Stimuli and Rhinovirus Infection. *Front Immunol* 2019; **10**: 1987 [PMID: [31497021](#) DOI: [10.3389/fimmu.2019.01987](#)]
- 21 **Cai J**, Lu W, Du S, Guo Z, Wang H, Wei W, Shen X. Tenascin-C Modulates Cell Cycle Progression to Enhance Tumour Cell Proliferation through AKT/FOXO1 Signalling in Pancreatic Cancer. *J Cancer* 2018; **9**: 4449-4462 [PMID: [30519351](#) DOI: [10.7150/jca.25926](#)]
- 22 **Sarkar S**, Mirzaei R, Zemp FJ, Wei W, Senger DL, Robbins SM, Yong VW. Activation of NOTCH Signaling by Tenascin-C Promotes Growth of Human Brain Tumor-Initiating Cells. *Cancer Res* 2017; **77**: 3231-3243 [PMID: [28416488](#) DOI: [10.1158/0008-5472.CAN-16-2171](#)]
- 23 **Sun Z**, Schwenzer A, Rupp T, Murdamoothoo D, Vegliante R, Lefebvre O, Klein A, Hussenet T, Orend G. Tenascin-C Promotes Tumor Cell Migration and Metastasis through Integrin $\alpha\beta 1$ -Mediated YAP Inhibition. *Cancer Res* 2018; **78**: 950-961 [PMID: [29259017](#) DOI: [10.1158/0008-5472.CAN-17-1597](#)]
- 24 **Cai J**, Du S, Wang H, Xin B, Wang J, Shen W, Wei W, Guo Z, Shen X. Tenascin-C induces migration and invasion through JNK/c-Jun signalling in pancreatic cancer. *Oncotarget* 2017; **8**: 74406-74422 [PMID: [29088796](#) DOI: [10.18632/oncotarget.20160](#)]
- 25 **Sun Z**, Velázquez-Quesada I, Murdamoothoo D, Ahowesso C, Yilmaz A, Spenlé C, Averous G, Erne W, Oberndorfer F, Oszward A, Kain R, Bourdon C, Mangin P, Deligne C, Midwood K, Abou-Faycal C, Lefebvre O, Klein A, van der Heyden M, Chenard MP, Christofori G, Mathelin C, Loustau T, Hussenet T, Orend G. Tenascin-C increases lung metastasis by impacting blood vessel invasions. *Matrix Biol* 2019; **83**: 26-47 [PMID: [31288084](#) DOI: [10.1016/j.matbio.2019.07.001](#)]
- 26 **Cai HP**, Wang J, Xi SY, Ni XR, Chen YS, Yu YJ, Cen ZW, Yu ZH, Chen FR, Guo CC, Zhang J, Ke C, Chen ZP. Tenascin-mediated vasculogenic mimicry formation via regulation of MMP2/MMP9 in glioma. *Cell Death Dis* 2019; **10**: 879 [PMID: [31754182](#) DOI: [10.1038/s41419-019-2102-3](#)]
- 27 **Rupp T**, Langlois B, Koczorowska MM, Radwanska A, Sun Z, Hussenet T, Lefebvre O, Murdamoothoo D, Arnold C, Klein A, Biniossek ML, Hyenne V, Naudin E, Velázquez-Quesada I, Schilling O, Van Obberghen-Schilling E, Orend G. Tenascin-C Orchestrates Glioblastoma Angiogenesis by Modulation of Pro- and Anti-angiogenic Signaling. *Cell Rep* 2016; **17**: 2607-2619 [PMID: [27926865](#) DOI: [10.1016/j.celrep.2016.11.012](#)]
- 28 **Mirzaei R**, Sarkar S, Dzikowski L, Rawji KS, Khan L, Faissner A, Bose P, Yong VW. Brain tumor-initiating cells export tenascin-C associated with exosomes to suppress T cell activity. *Oncoimmunology* 2018; **7**: e1478647 [PMID: [30288344](#) DOI: [10.1080/2162402X.2018.1478647](#)]
- 29 **Jachetti E**, Caputo S, Mazzoleni S, Brambillasca CS, Parigi SM, Grioni M, Piras IS, Restuccia U, Calcinotto A, Freschi M, Bachi A, Galli R, Bellone M. Tenascin-C Protects Cancer Stem-like Cells from Immune Surveillance by Arresting T-cell Activation. *Cancer Res* 2015; **75**: 2095-2108 [PMID: [25808872](#) DOI: [10.1158/0008-5472.CAN-14-2346](#)]
- 30 **Yang Z**, Zhang C, Feng Y, Qi W, Cui Y, Xuan Y. Tenascin-C is involved in promotion of cancer stemness via the Akt/HIF1 α axis in esophageal squamous cell carcinoma. *Exp Mol Pathol* 2019; **109**: 104239 [PMID: [30904401](#) DOI: [10.1016/j.yexmp.2019.03.007](#)]
- 31 **Yang Z**, Zhang C, Feng Y, Quan M, Cui Y, Xuan Y. Tenascin-C predicts poor outcomes for patients with colorectal cancer and drives cancer stemness via Hedgehog signaling pathway. *Cancer Cell Int* 2020; **20**: 122 [PMID: [32322169](#) DOI: [10.1186/s12935-020-01188-w](#)]
- 32 **Shi M**, He X, Wei W, Wang J, Zhang T, Shen X. Tenascin-C induces resistance to apoptosis in pancreatic cancer cell through activation of ERK/NF- κ B pathway. *Apoptosis* 2015; **20**: 843-857 [PMID: [25690319](#) DOI: [10.1007/s10495-015-1106-4](#)]
- 33 **Esposito I**, Penzel R, Chaib-Harrireche M, Barcena U, Bergmann F, Riedl S, Kaye H, Giese N, Kleeff J, Friess H, Schirmacher P. Tenascin C and annexin II expression in the process of pancreatic carcinogenesis. *J Pathol* 2006; **208**: 673-685 [PMID: [16450333](#) DOI: [10.1002/path.1935](#)]
- 34 **Saupe F**, Schwenzer A, Jia Y, Gasser I, Spenlé C, Langlois B, Kammerer M, Lefebvre O, Hlushchuk R, Rupp T, Marko M, van der Heyden M, Cremel G, Arnold C, Klein A, Simon-Assmann P, Djonov V, Neuville-Méchine A, Esposito I, Slotta-Huspenina J, Janssen KP, de Wever O, Christofori G, Hussenet T, Orend G. Tenascin-C downregulates wnt inhibitor dickkopf-1, promoting tumorigenesis in a neuroendocrine tumor model. *Cell Rep* 2013; **5**: 482-492 [PMID: [24139798](#) DOI: [10.1016/j.celrep.2013.09.014](#)]
- 35 **Spenlé C**, Gasser I, Saupe F, Janssen KP, Arnold C, Klein A, van der Heyden M, Mutterer J, Neuville-Méchine A, Chenard MP, Guenot D, Esposito I, Slotta-Huspenina J, Ambartsumian N, Simon-Assmann P, Orend G. Spatial organization of the tenascin-C microenvironment in experimental and human cancer. *Cell Adh Migr* 2015; **9**: 4-13 [PMID: [25611571](#) DOI: [10.1080/19336918.2015.1005452](#)]
- 36 **Li M**, Peng F, Li G, Fu Y, Huang Y, Chen Z, Chen Y. Proteomic analysis of stromal proteins in different stages of colorectal cancer establishes Tenascin-C as a stromal biomarker for colorectal cancer metastasis. *Oncotarget* 2016; **7**: 37226-37237 [PMID: [27191989](#) DOI: [10.18632/oncotarget.9362](#)]
- 37 **Giblin SP**, Midwood KS. Tenascin-C: Form vs function. *Cell Adh Migr* 2015; **9**: 48-82 [DOI: [10.4161/19336918.2014.987587](#)]

- 38 **Passlick B**, Sienel W, Seen-Hibler R, Wöckel W, Thetter O, Mutschler W, Pantel K. Overexpression of matrix metalloproteinase 2 predicts unfavorable outcome in early-stage non-small cell lung cancer. *Clin Cancer Res* 2000; **6**: 3944-3948 [PMID: [11051242](#)]
- 39 **Cai M**, Onoda K, Takao M, Kyoko IY, Shimpō H, Yoshida T, Yada I. Degradation of tenascin-C and activity of matrix metalloproteinase-2 are associated with tumor recurrence in early stage non-small cell lung cancer. *Clin Cancer Res* 2002; **8**: 1152-1156 [PMID: [11948127](#)]
- 40 **Mai J**, Sameni M, Mikkelsen T, Sloane BF. Degradation of extracellular matrix protein tenascin-C by cathepsin B: an interaction involved in the progression of gliomas. *Biol Chem* 2002; **383**: 1407-1413 [PMID: [12437133](#) DOI: [10.1515/BC.2002.159](#)]
- 41 **Mohan V**, Das A, Sagi I. Emerging roles of ECM remodeling processes in cancer. *Semin Cancer Biol* 2020; **62**: 192-200 [PMID: [31518697](#) DOI: [10.1016/j.semcancer.2019.09.004](#)]
- 42 **Stamenkovic I**. Extracellular matrix remodelling: the role of matrix metalloproteinases. *J Pathol* 2003; **200**: 448-464 [PMID: [12845612](#) DOI: [10.1002/path.1400](#)]
- 43 **Davis GE**, Bayless KJ, Davis MJ, Meininger GA. Regulation of tissue injury responses by the exposure of matricryptic sites within extracellular matrix molecules. *Am J Pathol* 2000; **156**: 1489-1498 [PMID: [10793060](#) DOI: [10.1016/S0002-9440\(10\)65020-1](#)]
- 44 **Shimshoni E**, Yablecovitch D, Baram L, Dotan I, Sagi I. ECM remodelling in IBD: innocent bystander or partner in crime? *Gut* 2015; **64**: 367-372 [PMID: [25416065](#) DOI: [10.1136/gutjnl-2014-308048](#)]
- 45 **Sakai T**, Kawakatsu H, Hirota N, Yokoyama T, Sakakura T, Saito M. Specific expression of tenascin in human colonic neoplasms. *Br J Cancer* 1993; **67**: 1058-1064 [PMID: [7684238](#) DOI: [10.1038/bjc.1993.194](#)]
- 46 **Kusagawa H**, Onoda K, Namikawa S, Yada I, Okada A, Yoshida T, Sakakura T. Expression and degeneration of tenascin-C in human lung cancers. *Br J Cancer* 1998; **77**: 98-102 [PMID: [9459152](#) DOI: [10.1038/bjc.1998.15](#)]
- 47 **Saito Y**, Imazeki H, Miura S, Yoshimura T, Okutsu H, Harada Y, Ohwaki T, Nagao O, Kamiya S, Hayashi R, Kodama H, Handa H, Yoshida T, Fukai F. A peptide derived from tenascin-C induces beta1 integrin activation through syndecan-4. *J Biol Chem* 2007; **282**: 34929-34937 [PMID: [17901052](#) DOI: [10.1074/jbc.M705608200](#)]
- 48 **Tanaka R**, Seki Y, Saito Y, Kamiya S, Fujita M, Okutsu H, Iyoda T, Takai T, Owaki T, Yajima H, Taira J, Hayashi R, Kodama H, Matsunaga T, Fukai F. Tenascin-C-derived peptide TNIIIA2 highly enhances cell survival and platelet-derived growth factor (PDGF)-dependent cell proliferation through potentiated and sustained activation of integrin $\alpha 5\beta 1$. *J Biol Chem* 2014; **289**: 17699-17708 [PMID: [24808173](#) DOI: [10.1074/jbc.M113.546622](#)]
- 49 **Dueck M**, Riedl S, Hinz U, Tandara A, Möller P, Herfarth C, Faissner A. Detection of tenascin-C isoforms in colorectal mucosa, ulcerative colitis, carcinomas and liver metastases. *Int J Cancer* 1999; **82**: 477-483 [PMID: [10404058](#) DOI: [10.1002/\(sici\)1097-0215\(19990812\)82:4<477::aid-ijc2>3.0.co;2-5](#)]
- 50 **Fujita M**, Yamamoto T, Iyoda T, Fujisawa T, Nagai R, Kudo C, Sasada M, Kodama H, Fukai F. Autocrine Production of PDGF Stimulated by the Tenascin-C-Derived Peptide TNIIIA2 Induces Hyper-Proliferation in Glioblastoma Cells. *Int J Mol Sci* 2019; **20** [PMID: [31261783](#) DOI: [10.3390/ijms20133183](#)]
- 51 **Fujita M**, Yamamoto T, Iyoda T, Fujisawa T, Sasada M, Nagai R, Kudo C, Otsuka K, Kamiya S, Kodama H, Fukai F. Aggressive Progression in Glioblastoma Cells through Potentiated Activation of Integrin $\alpha 5\beta 1$ by the Tenascin-C-Derived Peptide TNIIIA2. *Mol Cancer Ther* 2019; **18**: 1649-1658 [PMID: [31189613](#) DOI: [10.1158/1535-7163.MCT-18-1251](#)]
- 52 **Fujita M**, Sasada M, Iyoda T, Nagai R, Kudo C, Yamamoto T, Osada S, Kodama H, Fukai F. Anoikis resistance conferred by tenascin-C-derived peptide TNIIIA2 and its disruption by integrin inactivation. *Biochem Biophys Res Commun* 2021; **536**: 14-19 [PMID: [33360093](#) DOI: [10.1016/j.bbrc.2020.12.050](#)]
- 53 **Iyoda T**, Fujita M, Fukai F. Biologically Active TNIIIA2 Region in Tenascin-C Molecule: A Major Contributor to Elicit Aggressive Malignant Phenotypes From Tumors/Tumor Stroma. *Front Immunol* 2020; **11**: 610096 [PMID: [33362799](#) DOI: [10.3389/fimmu.2020.610096](#)]
- 54 **Siegel RL**, Miller KD, Fuchs HE, Jemal A. Cancer Statistics, 2021. *CA Cancer J Clin* 2021; **71**: 7-33 [PMID: [33433946](#) DOI: [10.3322/caac.21654](#)]
- 55 **Levin TR**, Corley DA, Jensen CD, Schottinger JE, Quinn VP, Zauber AG, Lee JK, Zhao WK, Udaltsova N, Ghai NR, Lee AT, Quesenberry CP, Fireman BH, Doubeni CA. Effects of Organized Colorectal Cancer Screening on Cancer Incidence and Mortality in a Large Community-Based Population. *Gastroenterology* 2018; **155**: 1383-1391.e5 [PMID: [30031768](#) DOI: [10.1053/j.gastro.2018.07.017](#)]
- 56 **Chibaudel B**, Tournigand C, Bonnetain F, Richa H, Benetkiewicz M, André T, de Gramont A. Therapeutic strategy in unresectable metastatic colorectal cancer: an updated review. *Ther Adv Med Oncol* 2015; **7**: 153-169 [PMID: [26673925](#) DOI: [10.1177/1758834015572343](#)]
- 57 **Ji K**, Zhang M, Chu Q, Gan Y, Ren H, Zhang L, Wang L, Li X, Wang W. The Role of p-STAT3 as a Prognostic and Clinicopathological Marker in Colorectal Cancer: A Systematic Review and Meta-Analysis. *PLoS One* 2016; **11**: e0160125 [PMID: [27504822](#) DOI: [10.1371/journal.pone.0160125](#)]
- 58 **Dyson JK**, Rutter MD. Colorectal cancer in inflammatory bowel disease: what is the real magnitude of the risk? *World J Gastroenterol* 2012; **18**: 3839-3848 [PMID: [22876036](#) DOI: [10.3748/wjg.v18.i29.3839](#)]

- 59 **Maida M**, Macaluso FS, Ianiro G, Mangiola F, Sinagra E, Hold G, Maida C, Cammarota G, Gasbarrini A, Scarpulla G. Screening of colorectal cancer: present and future. *Expert Rev Anticancer Ther* 2017; **17**: 1131-1146 [PMID: [29022408](#) DOI: [10.1080/14737140.2017.1392243](#)]
- 60 **Issa IA**, Noureddine M. Colorectal cancer screening: An updated review of the available options. *World J Gastroenterol* 2017; **23**: 5086-5096 [PMID: [28811705](#) DOI: [10.3748/wjg.v23.i28.5086](#)]
- 61 **Loupakis F**, Cremolini C, Masi G, Lonardi S, Zagonel V, Salvatore L, Cortesi E, Tomasello G, Ronzoni M, Spadi R, Zaniboni A, Tonini G, Buonadonna A, Amoroso D, Chiara S, Carlomagno C, Boni C, Allegrini G, Boni L, Falcone A. Initial therapy with FOLFOXIRI and bevacizumab for metastatic colorectal cancer. *N Engl J Med* 2014; **371**: 1609-1618 [PMID: [25337750](#) DOI: [10.1056/NEJMoa1403108](#)]
- 62 **Biller LH**, Schrag D. Diagnosis and Treatment of Metastatic Colorectal Cancer: A Review. *JAMA* 2021; **325**: 669-685 [PMID: [33591350](#) DOI: [10.1001/jama.2021.0106](#)]
- 63 **Taniguchi H**, Yamazaki K, Yoshino T, Muro K, Yatabe Y, Watanabe T, Ebi H, Ochiai A, Baba E, Tsuchihara K; Japanese Society of Medical Oncology. Japanese Society of Medical Oncology Clinical Guidelines: RAS (KRAS/NRAS) mutation testing in colorectal cancer patients. *Cancer Sci* 2015; **106**: 324-327 [PMID: [25800101](#) DOI: [10.1111/cas.12595](#)]
- 64 **Takeda A**, Otani Y, Iseki H, Takeuchi H, Aikawa K, Tabuchi S, Shinozuka N, Saeki T, Okazaki Y, Koyama I. Clinical significance of large tenascin-C spliced variant as a potential biomarker for colorectal cancer. *World J Surg* 2007; **31**: 388-394 [PMID: [17219282](#) DOI: [10.1007/s00268-006-0328-6](#)]
- 65 **Wanitsuwan W**, Kanngurn S, Boonpipattanapong T, Sangthong R, Sangkhatth S. Overall expression of beta-catenin outperforms its nuclear accumulation in predicting outcomes of colorectal cancers. *World J Gastroenterol* 2008; **14**: 6052-6059 [PMID: [18932285](#) DOI: [10.3748/wjg.14.6052](#)]
- 66 **Beiter K**, Hiendlmeyer E, Brabletz T, Hlubek F, Haynl A, Knoll C, Kirchner T, Jung A. beta-Catenin regulates the expression of tenascin-C in human colorectal tumors. *Oncogene* 2005; **24**: 8200-8204 [PMID: [16091738](#) DOI: [10.1038/sj.onc.1208960](#)]
- 67 **De Wever O**, Nguyen QD, Van Hoorde L, Bracke M, Bruyneel E, Gespach C, Mareel M. Tenascin-C and SF/HGF produced by myofibroblasts in vitro provide convergent pro-invasive signals to human colon cancer cells through RhoA and Rac. *FASEB J* 2004; **18**: 1016-1018 [PMID: [15059978](#) DOI: [10.1096/fj.03-1110fje](#)]
- 68 **Li ZL**, Wang ZJ, Wei GH, Yang Y, Wang XW. Changes in extracellular matrix in different stages of colorectal cancer and their effects on proliferation of cancer cells. *World J Gastrointest Oncol* 2020; **12**: 267-275 [PMID: [32206177](#) DOI: [10.4251/wjgo.v12.i3.267](#)]
- 69 **Sis B**, Sağol O, Küpelioğlu A, Sokmen S, Terzi C, Fuzun M, Ozer E, Bishop P. Prognostic significance of matrix metalloproteinase-2, cathepsin D, and tenascin-C expression in colorectal carcinoma. *Pathol Res Pract* 2004; **200**: 379-387 [PMID: [15239346](#) DOI: [10.1016/j.prp.2004.02.012](#)]
- 70 **Buchert M**, Rohde F, Eissmann M, Tebbutt N, Williams B, Tan CW, Owen A, Hirokawa Y, Gnann A, Orend G, Orner G, Dashwood RH, Heath JK, Ernst M, Janssen KP. A hypermorphic epithelial β -catenin mutation facilitates intestinal tumorigenesis in mice in response to compounding WNT-pathway mutations. *Dis Model Mech* 2015; **8**: 1361-1373 [PMID: [26398937](#) DOI: [10.1242/dmm.019844](#)]
- 71 **Siri A**, Knäuper V, Veirana N, Caocci F, Murphy G, Zardi L. Different susceptibility of small and large human tenascin-C isoforms to degradation by matrix metalloproteinases. *J Biol Chem* 1995; **270**: 8650-8654 [PMID: [7536739](#) DOI: [10.1074/jbc.270.15.8650](#)]
- 72 **Suzuki H**, Sasada M, Kamiya S, Ito Y, Watanabe H, Okada Y, Ishibashi K, Iyoda T, Yanaka A, Fukai F. The Promoting Effect of the Extracellular Matrix Peptide TNIIIA2 Derived from Tenascin-C in Colon Cancer Cell Infiltration. *Int J Mol Sci* 2017; **18** [PMID: [28106752](#) DOI: [10.3390/ijms18010181](#)]
- 73 **Liu C**, Pei H, Tan F. Matrix Stiffness and Colorectal Cancer. *Onco Targets Ther* 2020; **13**: 2747-2755 [PMID: [32280247](#) DOI: [10.2147/OTT.S231010](#)]
- 74 **Nebuloni M**, Albarello L, Andolfo A, Magagnotti C, Genovese L, Locatelli I, Tonon G, Longhi E, Zerbi P, Allevi R, Podestà A, Puricelli L, Milani P, Soldarini A, Salonia A, Alfano M. Insight On Colorectal Carcinoma Infiltration by Studying Perilesional Extracellular Matrix. *Sci Rep* 2016; **6**: 22522 [PMID: [26940881](#) DOI: [10.1038/srep22522](#)]
- 75 **Wei B**, Zhou X, Liang C, Zheng X, Lei P, Fang J, Han X, Wang L, Qi C, Wei H. Human colorectal cancer progression correlates with LOX-induced ECM stiffening. *Int J Biol Sci* 2017; **13**: 1450-1457 [PMID: [29209148](#) DOI: [10.7150/ijbs.21230](#)]
- 76 **Baker AM**, Bird D, Lang G, Cox TR, Erler JT. Lysyl oxidase enzymatic function increases stiffness to drive colorectal cancer progression through FAK. *Oncogene* 2013; **32**: 1863-1868 [PMID: [22641216](#) DOI: [10.1038/onc.2012.202](#)]
- 77 **Barnes JM**, Kaushik S, Bainer RO, Sa JK, Woods EC, Kai F, Przybyla L, Lee M, Lee HW, Tung JC, Maller O, Barrett AS, Lu KV, Lakins JN, Hansen KC, Obernier K, Alvarez-Buylla A, Bergers G, Phillips JJ, Nam DH, Bertozzi CR, Weaver VM. A tension-mediated glycocalyx-integrin feedback loop promotes mesenchymal-like glioblastoma. *Nat Cell Biol* 2018; **20**: 1203-1214 [PMID: [30202050](#) DOI: [10.1038/s41556-018-0183-3](#)]
- 78 **Liu QZ**, Gao XH, Chang WJ, Gong HF, Fu CG, Zhang W, Cao GW. Expression of ITGB1 predicts prognosis in colorectal cancer: a large prospective study based on tissue microarray. *Int J Clin Exp Pathol* 2015; **8**: 12802-12810 [PMID: [26722470](#)]

- 79 **Lu L**, Xie R, Wei R, Cai C, Bi D, Yin D, Liu H, Zheng J, Zhang Y, Song F, Gao Y, Tan L, Wei Q, Qin H. Integrin $\alpha 5$ subunit is required for the tumor supportive role of fibroblasts in colorectal adenocarcinoma and serves as a potential stroma prognostic marker. *Mol Oncol* 2019; **13**: 2697-2714 [PMID: [31600854](#) DOI: [10.1002/1878-0261.12583](#)]
- 80 **Yu M**, Chu S, Fei B, Fang X, Liu Z. O-GlcNAcylation of ITGA5 facilitates the occurrence and development of colorectal cancer. *Exp Cell Res* 2019; **382**: 111464 [PMID: [31202709](#) DOI: [10.1016/j.yexcr.2019.06.009](#)]
- 81 **Murillo CA**, Rychahou PG, Evers BM. Inhibition of alpha5 integrin decreases PI3K activation and cell adhesion of human colon cancers. *Surgery* 2004; **136**: 143-149 [PMID: [15300173](#) DOI: [10.1016/j.surg.2004.04.006](#)]
- 82 **Guha D**, Saha T, Bose S, Chakraborty S, Dhar S, Khan P, Adhikary A, Das T, Sa G. Integrin-EGFR interaction regulates anoikis resistance in colon cancer cells. *Apoptosis* 2019; **24**: 958-971 [PMID: [31641961](#) DOI: [10.1007/s10495-019-01573-5](#)]
- 83 **Janouskova H**, Ray AM, Noulet F, Lelong-Rebel I, Choulier L, Schaffner F, Lehmann M, Martin S, Teisinger J, Dontenwill M. Activation of p53 pathway by Nutlin-3a inhibits the expression of the therapeutic target $\alpha 5$ integrin in colon cancer cells. *Cancer Lett* 2013; **336**: 307-318 [PMID: [23523610](#) DOI: [10.1016/j.canlet.2013.03.018](#)]
- 84 **Stoeltzing O**, Liu W, Reinmuth N, Fan F, Parry GC, Parikh AA, McCarty MF, Bucana CD, Mazar AP, Ellis LM. Inhibition of integrin $\alpha 5 \beta 1$ function with a small peptide (ATN-161) plus continuous 5-FU infusion reduces colorectal liver metastases and improves survival in mice. *Int J Cancer* 2003; **104**: 496-503 [PMID: [12584749](#) DOI: [10.1002/ijc.10958](#)]
- 85 **Uemura N**, Okamoto S, Yamamoto S, Matsumura N, Yamaguchi S, Yamakido M, Taniyama K, Sasaki N, Schlemper RJ. Helicobacter pylori infection and the development of gastric cancer. *N Engl J Med* 2001; **345**: 784-789 [PMID: [11556297](#) DOI: [10.1056/NEJMoa001999](#)]
- 86 **Saito I**, Miyamura T, Ohbayashi A, Harada H, Katayama T, Kikuchi S, Watanabe Y, Koi S, Onji M, Ohta Y. Hepatitis C virus infection is associated with the development of hepatocellular carcinoma. *Proc Natl Acad Sci USA* 1990; **87**: 6547-6549 [PMID: [2168552](#) DOI: [10.1073/pnas.87.17.6547](#)]
- 87 **Ekbom A**, Helmick C, Zack M, Adami HO. Ulcerative colitis and colorectal cancer. A population-based study. *N Engl J Med* 1990; **323**: 1228-1233 [PMID: [2215606](#) DOI: [10.1056/NEJM199011013231802](#)]
- 88 **Jess T**, Loftus EV Jr, Velayos FS, Harmsen WS, Zinsmeister AR, Smyrk TC, Schleck CD, Tremaine WJ, Melton LJ 3rd, Munkholm P, Sandborn WJ. Risk of intestinal cancer in inflammatory bowel disease: a population-based study from olmsted county, Minnesota. *Gastroenterology* 2006; **130**: 1039-1046 [PMID: [16618397](#) DOI: [10.1053/j.gastro.2005.12.037](#)]
- 89 **Eaden JA**, Abrams KR, Mayberry JF. The risk of colorectal cancer in ulcerative colitis: a meta-analysis. *Gut* 2001; **48**: 526-535 [PMID: [11247898](#) DOI: [10.1136/gut.48.4.526](#)]
- 90 **Feagins LA**, Souza RF, Spechler SJ. Carcinogenesis in IBD: potential targets for the prevention of colorectal cancer. *Nat Rev Gastroenterol Hepatol* 2009; **6**: 297-305 [PMID: [19404270](#) DOI: [10.1038/nrgastro.2009.44](#)]
- 91 **Bewtra M**, Kaiser LM, TenHave T, Lewis JD. Crohn's disease and ulcerative colitis are associated with elevated standardized mortality ratios: a meta-analysis. *Inflamm Bowel Dis* 2013; **19**: 599-613 [PMID: [23388544](#) DOI: [10.1097/MIB.0b013e31827f27ae](#)]
- 92 **Olén O**, Erichsen R, Sachs MC, Pedersen L, Halfvarson J, Askling J, Ekbom A, Sørensen HT, Ludvigsson JF. Colorectal cancer in Crohn's disease: a Scandinavian population-based cohort study. *Lancet Gastroenterol Hepatol* 2020; **5**: 475-484 [PMID: [32066530](#) DOI: [10.1016/S2468-1253\(20\)30005-4](#)]
- 93 **Olén O**, Erichsen R, Sachs MC, Pedersen L, Halfvarson J, Askling J, Ekbom A, Sørensen HT, Ludvigsson JF. Colorectal cancer in ulcerative colitis: a Scandinavian population-based cohort study. *Lancet* 2020; **395**: 123-131 [PMID: [31929014](#) DOI: [10.1016/S0140-6736\(19\)32545-0](#)]
- 94 **Ullman TA**, Itzkowitz SH. Intestinal inflammation and cancer. *Gastroenterology* 2011; **140**: 1807-1816 [PMID: [21530747](#) DOI: [10.1053/j.gastro.2011.01.057](#)]
- 95 **Zisman TL**, Rubin DT. Colorectal cancer and dysplasia in inflammatory bowel disease. *World J Gastroenterol* 2008; **14**: 2662-2669 [PMID: [18461651](#) DOI: [10.3748/wjg.14.2662](#)]
- 96 **Riedl S**, Tandara A, Reinshagen M, Hinz U, Faissner A, Bodenmüller H, Buhr HJ, Herfarth C, Möller P. Serum tenascin-C is an indicator of inflammatory bowel disease activity. *Int J Colorectal Dis* 2001; **16**: 285-291 [PMID: [11686525](#) DOI: [10.1007/s003840100312](#)]
- 97 **Spénlé C**, Lefebvre O, Lacroute J, Méchine-Neuville A, Barreau F, Blottiére HM, Duclos B, Arnold C, Hussenet T, Hemmerlé J, Gullberg D, Kedingier M, Sorokin L, Orend G, Simon-Assmann P. The laminin response in inflammatory bowel disease: protection or malignancy? *PLoS One* 2014; **9**: e111336 [PMID: [25347196](#) DOI: [10.1371/journal.pone.0111336](#)]
- 98 **Geboes K**, El-Zine MY, Dalle I, El-Haddad S, Rutgeerts P, Van Eyken P. Tenascin and strictures in inflammatory bowel disease: an immunohistochemical study. *Int J Surg Pathol* 2001; **9**: 281-286 [PMID: [12574843](#) DOI: [10.1177/106689690100900404](#)]
- 99 **Riedl S**, Kadmon M, Tandara A, Hinz U, Möller P, Herfarth C, Faissner A. Mucosal tenascin C content in inflammatory and neoplastic diseases of the large bowel. *Dis Colon Rectum* 1998; **41**: 86-92 [PMID: [9510316](#) DOI: [10.1007/BF02236901](#)]
- 100 **Brant SR**, Okou DT, Simpson CL, Cutler DJ, Haritunians T, Bradfield JP, Chopra P, Prince J, Begum F, Kumar A, Huang C, Venkateswaran S, Datta LW, Wei Z, Thomas K, Herrinton LJ, Klapproth JA, Quiros AJ, Seminerio J, Liu Z, Alexander JS, Baldassano RN, Dudley-Brown S,

- Cross RK, Dassopoulos T, Denson LA, Dhere TA, Dryden GW, Hanson JS, Hou JK, Hussain SZ, Hyams JS, Isaacs KL, Kader H, Kappelman MD, Katz J, Kellermayer R, Kirschner BS, Kuemmerle JF, Kwon JH, Lazarev M, Li E, Mack D, Mannon P, Moulton DE, Newberry RD, Osuntokun BO, Patel AS, Saeed SA, Targan SR, Valentine JF, Wang MH, Zonca M, Rioux JD, Duerr RH, Silverberg MS, Cho JH, Hakonarson H, Zwick ME, McGovern DP, Kugathasan S. Genome-Wide Association Study Identifies African-Specific Susceptibility Loci in African Americans With Inflammatory Bowel Disease. *Gastroenterology* 2017; **152**: 206-217.e2 [PMID: 27693347 DOI: 10.1053/j.gastro.2016.09.032]
- 101 **Ning L**, Li S, Gao J, Ding L, Wang C, Chen W, Shan G, Zhang F, Yu J, Xu G. Tenascin-C Is Increased in Inflammatory Bowel Disease and Is Associated with response to Infliximab Therapy. *Biomed Res Int* 2019; **2019**: 1475705 [PMID: 31886172 DOI: 10.1155/2019/1475705]
- 102 **Kawamura T**, Yamamoto M, Suzuki K, Suzuki Y, Kamishima M, Sakata M, Kurachi K, Setoh M, Konno H, Takeuchi H. Tenascin-C Produced by Intestinal Myofibroblasts Promotes Colitis-associated Cancer Development Through Angiogenesis. *Inflamm Bowel Dis* 2019; **25**: 732-741 [PMID: 30517646 DOI: 10.1093/ibd/izy368]
- 103 **Kofla-Dlubacz A**, Matusiewicz M, Krzystek-Korpacka M, Iwanczak B. Correlation of MMP-3 and MMP-9 with Crohn's disease activity in children. *Dig Dis Sci* 2012; **57**: 706-712 [PMID: 21997756 DOI: 10.1007/s10620-011-1936-z]
- 104 **Fujita M**, Ito-Fujita Y, Iyoda T, Sasada M, Okada Y, Ishibashi K, Osawa T, Kodama H, Fukai F, Suzuki H. Peptide TNIIIA2 Derived from Tenascin-C Contributes to Malignant Progression in Colitis-Associated Colorectal Cancer via β 1-Integrin Activation in Fibroblasts. *Int J Mol Sci* 2019; **20** [PMID: 31195598 DOI: 10.3390/ijms20112752]
- 105 **Fujita M**, Sasada M, Iyoda T, Fukai F. Involvement of Integrin-Activating Peptides Derived from Tenascin-C in Cancer Aggression and New Anticancer Strategy Using the Fibronectin-Derived Integrin-Inactivating Peptide. *Molecules* 2020; **25** [PMID: 32708610 DOI: 10.3390/molecules25143239]
- 106 **Erez N**, Truitt M, Olson P, Arron ST, Hanahan D. Cancer-Associated Fibroblasts Are Activated in Incipient Neoplasia to Orchestrate Tumor-Promoting Inflammation in an NF-kappaB-Dependent Manner. *Cancer Cell* 2010; **17**: 135-147 [PMID: 20138012 DOI: 10.1016/j.ccr.2009.12.041]
- 107 **Mukaida N**, Sasaki S. Fibroblasts, an inconspicuous but essential player in colon cancer development and progression. *World J Gastroenterol* 2016; **22**: 5301-5316 [PMID: 27340347 DOI: 10.3748/wjg.v22.i23.5301]
- 108 **Sahai E**, Astsaturov I, Cukierman E, DeNardo DG, Egeblad M, Evans RM, Fearon D, Greten FR, Hingorani SR, Hunter T, Hynes RO, Jain RK, Janowitz T, Jorgensen C, Kimmelman AC, Kolonin MG, Maki RG, Powers RS, Puré E, Ramirez DC, Scherz-Shouval R, Sherman MH, Stewart S, Tlsty TD, Tuveson DA, Watt FM, Weaver V, Weeraratna AT, Werb Z. A framework for advancing our understanding of cancer-associated fibroblasts. *Nat Rev Cancer* 2020; **20**: 174-186 [PMID: 31980749 DOI: 10.1038/s41568-019-0238-1]
- 109 **Sasaki S**, Baba T, Shinagawa K, Matsushima K, Mukaida N. Crucial involvement of the CCL3-CCR5 axis-mediated fibroblast accumulation in colitis-associated carcinogenesis in mice. *Int J Cancer* 2014; **135**: 1297-1306 [PMID: 24510316 DOI: 10.1002/ijc.28779]
- 110 **Neufert C**, Becker C, Türeci Ö, Waldner MJ, Backert I, Floh K, Atreya I, Leppkes M, Jefremow A, Vieth M, Schneider-Stock R, Klinger P, Greten FR, Threadgill DW, Sahin U, Neurath MF. Tumor fibroblast-derived epiregulin promotes growth of colitis-associated neoplasms through ERK. *J Clin Invest* 2013; **123**: 1428-1443 [PMID: 23549083 DOI: 10.1172/JCI63748]
- 111 **Ni WD**, Yang ZT, Cui CA, Cui Y, Fang LY, Xuan YH. Tenascin-C is a potential cancer-associated fibroblasts marker and predicts poor prognosis in prostate cancer. *Biochem Biophys Res Commun* 2017; **486**: 607-612 [PMID: 28341124 DOI: 10.1016/j.bbrc.2017.03.021]
- 112 **Yashiro M**. Ulcerative colitis-associated colorectal cancer. *World J Gastroenterol* 2014; **20**: 16389-16397 [PMID: 25469007 DOI: 10.3748/wjg.v20.i44.16389]
- 113 **Rutter MD**, Saunders BP, Wilkinson KH, Rumbles S, Schofield G, Kamm MA, Williams CB, Price AB, Talbot IC, Forbes A. Cancer surveillance in longstanding ulcerative colitis: endoscopic appearances help predict cancer risk. *Gut* 2004; **53**: 1813-1816 [PMID: 15542520 DOI: 10.1136/gut.2003.038505]
- 114 **Rieder F**, Fiocchi C, Rogler G. Mechanisms, Management, and Treatment of Fibrosis in Patients With Inflammatory Bowel Diseases. *Gastroenterology* 2017; **152**: 340-350.e6 [PMID: 27720839 DOI: 10.1053/j.gastro.2016.09.047]
- 115 **Miroshnikova YA**, Mouw JK, Barnes JM, Pickup MW, Lakins JN, Kim Y, Lobo K, Persson AI, Reis GF, McKnight TR, Holland EC, Phillips JJ, Weaver VM. Tissue mechanics promote IDH1-dependent HIF1 α -tenascin C feedback to regulate glioblastoma aggression. *Nat Cell Biol* 2016; **18**: 1336-1345 [PMID: 27820599 DOI: 10.1038/ncb3429]
- 116 **Johnson LA**, Rodansky ES, Sauder KL, Horowitz JC, Mih JD, Tschumperlin DJ, Higgins PD. Matrix stiffness corresponding to strictured bowel induces a fibrogenic response in human colonic fibroblasts. *Inflamm Bowel Dis* 2013; **19**: 891-903 [PMID: 23502354 DOI: 10.1097/MIB.0b013e3182813297]
- 117 **Erdem E**, Kochan K, Paker N, Gokden Y, Degirmenci AS, Kocak F, Gonen C. The correlation between tenascin-C expression, and formation of intestinal stricture. *North Clin Istanbul* 2014; **1**: 127-131 [PMID: 28058317 DOI: 10.14744/nci.2014.13008]
- 118 **Danese S**, Sans M, Spencer DM, Beck I, Doñate F, Plunkett ML, de la Motte C, Redline R, Shaw

- DE, Levine AD, Mazar AP, Fiocchi C. Angiogenesis blockade as a new therapeutic approach to experimental colitis. *Gut* 2007; **56**: 855-862 [PMID: 17170016 DOI: 10.1136/gut.2006.114314]
- 119 **Chidlow JH Jr**, Langston W, Greer JJ, Ostanin D, Abdelbaqi M, Houghton J, Senthikumar A, Shukla D, Mazar AP, Grisham MB, Kevil CG. Differential angiogenic regulation of experimental colitis. *Am J Pathol* 2006; **169**: 2014-2030 [PMID: 17148665 DOI: 10.2353/ajpath.2006.051021]
- 120 **Terasaki M**, Ikuta M, Kojima H, Tanaka T, Maeda H, Miyashita K, Mutoh M. Dietary Fucoxanthin Induces Anoikis in Colorectal Adenocarcinoma by Suppressing Integrin Signaling in a Murine Colorectal Cancer Model. *J Clin Med* 2019; **9** [PMID: 31905803 DOI: 10.3390/jcm9010090]
- 121 **Cianfrocca ME**, Kimmel KA, Gallo J, Cardoso T, Brown MM, Hudes G, Lewis N, Weiner L, Lam GN, Brown SC, Shaw DE, Mazar AP, Cohen RB. Phase 1 trial of the antiangiogenic peptide ATN-161 (Ac-PHSCN-NH(2)), a beta integrin antagonist, in patients with solid tumours. *Br J Cancer* 2006; **94**: 1621-1626 [PMID: 16705310 DOI: 10.1038/sj.bjc.6603171]
- 122 **Malric L**, Monferran S, Gilhodes J, Boyrie S, Dahan P, Skuli N, Sesen J, Filleron T, Kowalski-Chauvel A, Cohen-Jonathan Moyal E, Toulas C, Lemarié A. Interest of integrins targeting in glioblastoma according to tumor heterogeneity and cancer stem cell paradigm: an update. *Oncotarget* 2017; **8**: 86947-86968 [PMID: 29156849 DOI: 10.18632/oncotarget.20372]
- 123 **Alday-Parejo B**, Stupp R, Rüegg C. Are Integrins Still Practicable Targets for Anti-Cancer Therapy? *Cancers (Basel)* 2019; **11** [PMID: 31336983 DOI: 10.3390/cancers11070978]
- 124 **Nwagwu CD**, Immidisetti AV, Bukanowska G, Vogelbaum MA, Carbonell AM. Convection-Enhanced Delivery of a First-in-Class Anti-β1 Integrin Antibody for the Treatment of High-Grade Glioma Utilizing Real-Time Imaging. *Pharmaceutics* 2020; **13** [PMID: 33396712 DOI: 10.3390/pharmaceutics13010040]



Published by **Baishideng Publishing Group Inc**
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: <https://www.f6publishing.com/helpdesk>

<https://www.wjgnet.com>

