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MINIREVIEWS

Regulators of liver cancer stem cells

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Abstract

Hepatocellular carcinoma (HCC) is a leading cause of cancer deaths. It is often detected at a stage when there are few therapeutic options. Liver cancer stem cells (LCSCs) are highly tumorigenic and resistant to chemotherapy and radiation therapy. Their presence in HCC is a major reason why HCC is difficult to treat. The development of LCSCs is regulated by a variety of factors. This review summarizes recent advances on the factors that regulate the development of LCSCs. Due to the importance of LCSCs in the development of HCC, a better understanding of how LCSCs are regulated will help to improve the treatments for HCC patients.

Key Words: Hepatocellular carcinoma; Liver cancer stem cells; Pluripotency transcription factors; Stem cell signaling; Genetic regulators; Epigenetic regulators

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Core Tip: Liver cancer stem cells (LCSCs) play a crucial role in the development of hepatocellular carcinoma (HCC), and their tumorigenicity and chemoresistance are major reasons as to why HCC is difficult to treat. LCSCs are regulated by a variety of factors. These factors include the network of pluripotency transcription factors, signaling pathways that regulate stem cells, genetic alterations that may be caused by mutations or viral factors, and epigenetic regulations including DNA methylation, histone methylation and microRNAs. These factors together induce the cancer stem cell properties of hepatocytes and play a critical role in the development of LCSCs and HCC.

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INTRODUCTION

Hepatocellular carcinoma (HCC) is the major histological subtype of liver cancer, accounting for approximately 75%-85% of primary liver cancer cases [1]. Most HCC cases are diagnosed at an advanced stage when the surgical resection opportunity is lost and there are few other therapeutic options[1]. The aggressiveness of HCC is partially due to the presence of cancer stem cells (CSCs), also known as tumorinitiating cells. CSCs are highly tumorigenic and resistant to chemotherapy and radiotherapy^[2]. They are a small subset of cancer cells with stem cell properties and capable of self-renewal. CSCs can initiate tumorigenesis and proliferate, leading to the production of a heterogeneous population of cancer cells. They may originate from transformed normal stem cells or from differentiated cells[2]. Like CSCs of other tumor types, liver CSCs (LCSCs) may also be derived from transformed liver progenitor cells or from the reprogramming of hepatocytes and biliary cells. LCSCs are also regulated by their own factors, such as pluripotency transcription factors (pTFs), CSC-related signaling pathways, genetic elements, epigenetic elements, and microenvironmental factors (i.e., the tumor microenvironment). In this review, we will focus on the factors that regulate the stemness properties of LCSCs, as the crosstalk between the tumor microenvironment and LCSCs had been discussed extensively by others[3,4].

ROLE OF PTFS IN THE REGULATION OF LCSCS

Normal somatic cells can be reprogrammed into induced pluripotent stem cells using four pTFs known as Yamanaka factors, which are OCT4, SOX2, c-Myc, and KLF4[5]. These factors play critical roles in the regulation of cell stemness. The effect of pTFs on CSCs, including LCSCs, has also received a lot of attention in recent years. In addition to Yamanaka factors, pTFs such as NANOG, SALL4, and SOX9 also promote the stemness of LCSCs[6,7]. The expressions of these pTFs are required for LCSCs to maintain their self-renewing ability and pluripotency, as well as to prevent differentiation[6,7]. The induction of one or more pTFs promotes the conversion of HCC cells into tumor cells with CSC properties [7,8]. However, the silencing of only one pTF such as NANOG cannot completely impair the tumorigenic ability of LCSCs[9], indicating that the loss of a limited number of pTFs is insufficient to destroy the properties of LCSCs. This may be explained by the fact that pTFs form a regulatory network, in which they regulate each other to direct the self-renewal of stem cells and to maintain their stemness properties [10,11]. In the pTF network, for example, OCT4 and SOX2, can promote each other's expression and also form a heterodimeric protein complex to transactivate NANOG expression. NANOG can also induce the expression of OCT4 and SOX2 to create a positive feedback regulation^[12]. Through amplifying leukemia inhibitory factor (LIF) signaling, NANOG also indirectly induces the expressions of other pTFs such as TBX3, KLF4, and TFCP2I1, which in turn induce the expression of NANOG, OCT4, and SOX2[12,13]. Therefore, the induction of only one pTF is sufficient to activate the whole network whereas the silencing of one pTF is insufficient to destroy the functions of the entire network [10]. It should be noted that the functions of pTFs mentioned above are uncovered mostly using established HCC cell lines. Whether and how these pTFs affect hepatocarcinogenesis *in vivo* were rarely reported and thus remain largely unclear. Moreover, although the main function of the established pTF network is to maintain the pluripotency of CSCs, the precise mechanisms involved in the establishment of this network during hepatocarcinogenesis and the tumorigenesis of other tumor types are still unknown. The understanding of the molecular details of these questions will help to identify novel targets for the prevention and treatment of HCC.

LCSC-RELATED SIGNALING PATHWAYS

Several CSC-related signaling pathways are involved in the induction of LCSCs and



the maintenance of their properties during hepatocarcinogenesis. Here, we will mainly discuss the effects of Wnt/Catenin, IL-6/STAT3, Hippo, Notch, and MAPK signaling pathways on the induction and maintenance of LCSCs. Among these pathways, Wnt/Catenin and IL-6/STAT3 are two well-known signaling pathways that support the stemness of HCC cells. Somatic mutations of genes in the Wnt pathway are commonly observed in HCC, which will be discussed in detail in the genetic elements section below. STAT3 is often activated in HCC and can transactivate the expression of many genes including NANOG, SOX2, and OCT4, which are pTFs that promote the development of LCSCs[14]. The cytokine interleukin (IL)-6, an upstream regulator of STAT3, is not only produced by immune cells but can also be produced by hepatocytes and hepatoma cells[15,16]. The production of IL-6 by hepatocytes to exert an autocrine effect to activate STAT3 in hepatocytes is noteworthy, as this effect may reprogram hepatocytes into stem-like cells and induce hepatocarcinogenesis in an immune cellindependent manner, even although the development of HCC is often associated with chronic liver inflammation.

Hippo signaling is a tumor-suppressive pathway that inhibits the proliferation of adult stem cells and progenitor cells[17]. The activation of Hippo signaling leads to the phosphorylation of transcriptional coactivators Yes-associated protein 1 (YAP) and transcriptional coactivator with PDZ-binding motif (TAZ), thereby sequestering them in the cytoplasm. This prevents YAP and TAZ from interacting with transcription factors of the TEA domain (TEAD) family in the nucleus to activate gene expression. The inhibition of TAZ expression was found to suppress the proliferation of HCC cells. However, it was also found to induce the compensatory expression of YAP, increase chemoresistance of tumor cells, and upregulate the expression of CD90, an HCCspecific CSC marker. Thus, when targeting the Hippo signaling pathway, it is necessary to simultaneously suppress both YAP and TAZ, as these two transcriptional coactivators are functionally distinct[18].

NOTCH signaling in CSCs promotes cancer progression and requires the cleavage of NOTCH by a disintegrin and metalloprotease (ADAM). The inducible nitric oxide synthase (iNOS) activates the NOTCH signaling by activating ADAM17/TACE, which leads to the production of activated NOTCH intracellular domain (NICD)[19]. iNOS also activates NOTCH signaling by inducing rhomboid protein 2, which promotes ADAM17/TACE-mediated NICD production[19]. Indeed, the expression of iNOS is associated with the expression of CD24 and CD133 stem cell markers in the tumor tissues of HCC patients and poor disease outcomes. Curiously, in a separate study it was found that the activation of the NOTCH pathway reduced HCC growth when the expression of RB, p107 and p130, three members of the retinoblastoma (RB) family, in the liver was abolished. This observation indicates that the role of NOTCH in hepatocarcinogenesis is complicated and may be context-dependent[20].

HCC often develops in association with liver cirrhosis, and activated fibroblasts (*i.e.*, hepatic stellate cells) play a key role in the development of liver cirrhosis. Cancerassociated fibroblasts isolated from HCC secret hepatocyte growth factor (HGF), which upon binding to its receptor c-MET on hepatoma cells can stimulate MAPK signaling to activate the transcription factor FRA1 (FOS-related antigen 1) to induce the expression of HEY1 (transcription of hairy/enhancer-of-split related with YRPW motif 1). HEY1 is also a downstream target of the NOTCH pathway and can increase the population of LCSCs and chemoresistance of hepatoma cells^[21].

Protein arginine methyltransferase 6 (PRMT6) methylates and binds to arginine-100 (R100) of c-Raf to disrupt its interaction with Ras to inhibit MAPK signaling. The silencing of PRMT6 activates MAPK signaling and enhances the stemness properties of CD133+ LCSCs[22].

GENETIC ELEMENTS THAT REGULATE LCSCS

The accumulation of oncogenic mutations can serve as the driver for the initiation of hepatocarcinogenesis. p53, the best-known tumor suppressor, plays a crucial role in preventing the malignant transformation of differentiated hepatocytes. It accomplishes this through multiple mechanisms, including surveillance of genome stability, arrest of the cell cycle, induction of apoptosis and suppression of cell stemness[16,23]. The p53 dysfunction by itself is insufficient to initiate hepatocarcinogenesis, and p53 by itself is often also sufficient to prevent hepatocytes from oncogenic transformation and acquisition of CSC properties^[24]. When hepatocytes with mutations override p53mediated tumor suppression, they are converted to HCC progenitor cells (HcPCs) with stemness properties and tumorigenic potentials. HcPCs produce IL-6 to stimulate

autocrine IL-6 signaling, cell growth and malignant progression[16,23]. Our recent studies also demonstrated that p53 could suppress the production of CD133⁺ LCSCs by preventing the OCT4-SOX2 complex from binding to the NANOG promoter and hence inhibiting the expression of NANOG[25].

The mutations of canonical Wnt/ β -cantenin signaling-related genes are often observed in HCC. Activating mutations in β -catenin directly activate the Wnt signaling pathway, and inactivating mutations in AXIN 1 and glycogen synthase kinase 3 (GSK- 3β), the protein subunits of the functional destruction complex for β -catenin, indirectly activate this pathway[26,27].

The infection of hepatocytes by hepatitis B virus (HBV) or hepatitis C virus (HCV) can also reprogram hepatocytes into stem-like cells to promote hepatocarcinogenesis. The HBV X protein (HBx) reprograms hepatocytes by activating STAT3 signaling and increasing the expression of farnesoid X receptor (FXR) and NANOG[28,29]. In addition, HBV infection induces autophagy to enhance its replication[30]. The selective removal of mitochondria by autophagy, known as mitophagy, can cause degradation of mitochondria-associated p53 to positively regulate CD133⁺ LCSCs[25]. Thus, HBVinduced autophagy and mitophagy can also contributes to HCC stemness[31]. HCV can also induce autophagy and mitophagy and thus may also induce LCSCs via a similar mechanism[32,33]. The expression of the HCV NS5B polymerase can induce the expression of many stemness-related genes in HCC[34]. The HCV NS5A protein also induces the expression of toll-like receptor 4 (TLR4) in hepatocytes/hepatoblasts. The activation of TLR4 by circulating endotoxin can induce the expression of NANOG, leading to the generation of chemoresistant CD133⁺ LCSCs[35]. As alcohol increases the circulating endotoxin levels and HCV induces the expression of TLR4, these observations provide an explanation as to why alcohol and HCV synergistically activate LCSCs and induce HCC[36].

EPIGENETIC ELEMENTS THAT REGULATE LCSCS

Epigenetic regulators include DNA methylation, histone methylation and gene silencing by microRNAs. Epigenetic changes can predispose hepatocytes to oncogenic mutations and transformation into LCSCs[37,38]. For example, the treatment of primary human cancer cells and liver cancer cell lines with zebularine, a potent DNA methyltransferase-1 inhibitor, induced global DNA hypomethylation and dramatically increased the frequency of cells with CSC properties, as judged by their self-renewal and superior tumor-initiating abilities[39]. Lysine-specific demethylase 1 (LSD1) suppresses the expression of Prickle 1 and APC, two suppressors of β -catenin, by inhibiting mono- and di-methylation of histone H3K4 at the promoters of these two genes. This leads to the activation of Wnt/β -catenin and enhances the stemness and chemoresistance of Lgr5⁺ LCSCs[40]. Some microRNAs can also regulate LCSCs. For examples, microRNA-130b suppresses the expression of tumor protein p53-induced nuclear protein 1 (TP53INP1) to promote CD133+ LCSC growth and self-renewal[41], and microRNA-1246 suppresses AXIN2 and GSK-3ß to activate Wnt signaling to induce CD133⁺ LCSCs[42]. In contrast, microRNA-142-3p directly targets the mRNA of CD133, a surface marker of CSCs that plays an important role in the maintenance of CSC properties by regulating neurotensin, IL-8, CXCL1, and MAPK signaling, resulting in the reduction of CD133⁺ LCSCs[43,44]. LncRNAs also affect the generation of LCSCs. For examples, lncRNA HOTAIR promotes the malignant growth of LCSCs through the downregulation of SETD2, a histone lysine methyltransferase[45], lncRNA MALAT1 promotes LCSC properties by upregulating YAP1 expression via microRNA-375 sponging[46], and lncRNA lncTCF7 promotes self-renewal of LCSCs through activation of Wnt signaling[47].

CONCLUSION

LCSCs are regulated by multiple factors including pTFs, signaling pathways, genetic elements, and epigenetic elements. A summary of these factors is illustrated in Figure 1. These factors affect one another and form an integrated network to regulate the self-renewal and other properties of LCSCs. In-depth understanding of how LCSCs are regulated will be important for the identification of novel therapeutic targets for improving the treatments for HCC.

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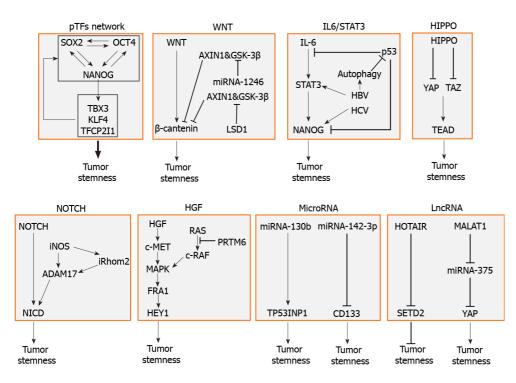


Figure 1 Molecular factors that regulate the stemness of liver cancer stem cells. The factors that regulate liver cancer stem cells (LCSCs) include the pluripotency transcription factor (pTF) network, in which pTFs affect the expression of one another to promote the development of LCSCs, the WNT signaling pathway, which is often activated by gene mutations, the IL6/STAT3 pathway, which can also be stimulated by hepatitis B virus or hepatitis C virus, the tumorsuppressive HIPPO pathway, the NOTCH signaling pathway, and hepatocyte growth factor and its downstream MAPK signaling pathway. Specific microRNAs such as miRNA-130b and miRNA-142-3p, and IncRNAs such as HOTAIR and MALAT1 can also promote the stemness of LCSCs. pTF: Pluripotency transcription factor; IL: Interleukin; HBV: Hepatitis B virus; HCV: Hepatitis C virus; YAP: Yes-associated protein; TEAD: TEA domain; iNOS: Inducible nitric oxide synthase; HGF: Hepatocyte growth factor.

REFERENCES

- 1 Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018; 68: 394-424 [PMID: 30207593 DOI: 10.3322/caac.21492]
- 2 Nio K, Yamashita T, Kaneko S. The evolving concept of liver cancer stem cells. Mol Cancer 2017; 16: 4 [PMID: 28137313 DOI: 10.1186/s12943-016-0572-9]
- Liu YC, Yeh CT, Lin KH. Cancer Stem Cell Functions in Hepatocellular Carcinoma and 3 Comprehensive Therapeutic Strategies. Cells 2020; 9 [PMID: 32466488 DOI: 10.3390/cells9061331]
- Tsui YM, Chan LK, Ng IO. Cancer stemness in hepatocellular carcinoma: mechanisms and 4 translational potential. Br J Cancer 2020; 122: 1428-1440 [PMID: 32231294 DOI: 10.1038/s41416-020-0823-9
- 5 Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. Cell 2006; 126: 663-676 [PMID: 16904174 DOI: 10.1016/j.cell.2006.07.024]
- Zhao W, Li Y, Zhang X. Stemness-Related Markers in Cancer. Cancer Transl Med 2017; 3: 87-95 6 [PMID: 29276782 DOI: 10.4103/ctm.ctm_69_16]
- 7 Liu C, Liu L, Chen X, Cheng J, Zhang H, Shen J, Shan J, Xu Y, Yang Z, Lai M, Qian C. Sox9 regulates self-renewal and tumorigenicity by promoting symmetrical cell division of cancer stem cells in hepatocellular carcinoma. Hepatology 2016; 64: 117-129 [PMID: 26910875 DOI: 10.1002/hep.28509]
- 8 Kim HJ, Jeong J, Park S, Jin YW, Lee SS, Lee SB, Choi D. Establishment of Hepatocellular Cancer Induced Pluripotent Stem Cells Using a Reprogramming Technique. Gut Liver 2017; 11: 261-269 [PMID: 27728962 DOI: 10.5009/gnl15389]
- Liu K, Hao M, Ouyang Y, Zheng J, Chen D. CD133+ cancer stem cells promoted by VEGF accelerate 9 the recurrence of hepatocellular carcinoma. Sci Rep 2017; 7: 41499 [PMID: 28134312 DOI: 10.1038/srep41499
- 10 Boheler KR. Stem cell pluripotency: a cellular trait that depends on transcription factors, chromatin state and a checkpoint deficient cell cycle. J Cell Physiol 2009; 221: 10-17 [PMID: 19562686 DOI: 10.1002/jcp.21866]
- Niwa H. The pluripotency transcription factor network at work in reprogramming. Curr Opin Genet 11 Dev 2014; 28: 25-31 [PMID: 25173150 DOI: 10.1016/j.gde.2014.08.004]
- 12 Papatsenko D, Waghray A, Lemischka IR. Feedback control of pluripotency in embryonic stem cells: Signaling, transcription and epigenetics. Stem Cell Res 2018; 29: 180-188 [PMID: 29727814



DOI: 10.1016/j.scr.2018.02.012]

- Stuart HT, van Oosten AL, Radzisheuskaya A, Martello G, Miller A, Dietmann S, Nichols J, Silva 13 JC. NANOG amplifies STAT3 activation and they synergistically induce the naive pluripotent program. Curr Biol 2014; 24: 340-346 [PMID: 24462001 DOI: 10.1016/j.cub.2013.12.040]
- 14 Yu H, Lee H, Herrmann A, Buettner R, Jove R. Revisiting STAT3 signalling in cancer: new and unexpected biological functions. Nat Rev Cancer 2014; 14: 736-746 [PMID: 25342631 DOI: 10.1038/nrc3818]
- 15 Kumagai N, Tsuchimoto K, Tsunematsu S, Toda K, Takeuchi O, Saito H, Morizane T, Tsuchiya M, Ishii H. Inhibition of growth of human hepatoma cells by dual-function antisense IL-6 oligonucleotides. Hepatol Res 2002; 22: 119-126 [PMID: 11818251 DOI: 10.1016/s1386-6346(01)00128-0]
- He G, Dhar D, Nakagawa H, Font-Burgada J, Ogata H, Jiang Y, Shalapour S, Seki E, Yost SE, Jepsen 16 K, Frazer KA, Harismendy O, Hatziapostolou M, Iliopoulos D, Suetsugu A, Hoffman RM, Tateishi R, Koike K, Karin M. Identification of liver cancer progenitors whose malignant progression depends on autocrine IL-6 signaling. Cell 2013; 155: 384-396 [PMID: 24120137 DOI: 10.1016/j.cell.2013.09.031
- Wang Y, Rattray JB, Thomas SA, Gurney J, Brown SP. In silico bacteria evolve robust cooperaion via complex quorum-sensing strategies. Sci Rep 2020; 10: 8628 [PMID: 32451396 DOI: 10.1038/s41598-020-65076-z
- 18 Hayashi H, Higashi T, Yokoyama N, Kaida T, Sakamoto K, Fukushima Y, Ishimoto T, Kuroki H, Nitta H, Hashimoto D, Chikamoto A, Oki E, Beppu T, Baba H. An Imbalance in TAZ and YAP Expression in Hepatocellular Carcinoma Confers Cancer Stem Cell-like Behaviors Contributing to Disease Progression. Cancer Res 2015; 75: 4985-4997 [PMID: 26420216 DOI: 10.1158/0008-5472.CAN-15-0291
- 19 Wang R, Li Y, Tsung A, Huang H, Du Q, Yang M, Deng M, Xiong S, Wang X, Zhang L, Geller DA, Cheng B, Billiar TR. iNOS promotes CD24⁺CD133⁺ liver cancer stem cell phenotype through a TACE/ADAM17-dependent Notch signaling pathway. Proc Natl Acad Sci USA 2018; 115: E10127-E10136 [PMID: 30297396 DOI: 10.1073/pnas.1722100115]
- Viatour P, Ehmer U, Saddic LA, Dorrell C, Andersen JB, Lin C, Zmoos AF, Mazur PK, Schaffer BE, 20 Ostermeier A, Vogel H, Sylvester KG, Thorgeirsson SS, Grompe M, Sage J. Notch signaling inhibits hepatocellular carcinoma following inactivation of the RB pathway. J Exp Med 2011; 208: 1963-1976 [PMID: 21875955 DOI: 10.1084/jem.20110198]
- 21 Lau EY, Lo J, Cheng BY, Ma MK, Lee JM, Ng JK, Chai S, Lin CH, Tsang SY, Ma S, Ng IO, Lee TK. Cancer-Associated Fibroblasts Regulate Tumor-Initiating Cell Plasticity in Hepatocellular Carcinoma through c-Met/FRA1/HEY1 Signaling. Cell Rep 2016; 15: 1175-1189 [PMID: 27134167 DOI: 10.1016/j.celrep.2016.04.019]
- Chan LH, Zhou L, Ng KY, Wong TL, Lee TK, Sharma R, Loong JH, Ching YP, Yuan YF, Xie D, 22 Lo CM, Man K, Artegiani B, Clevers H, Yan HH, Leung SY, Richard S, Guan XY, Huen MSY, Ma S. PRMT6 Regulates RAS/RAF Binding and MEK/ERK-Mediated Cancer Stemness Activities in Hepatocellular Carcinoma through CRAF Methylation. Cell Rep 2018; 25: 690-701.e8 [PMID: 30332648 DOI: 10.1016/j.celrep.2018.09.053]
- 23 Dhar D, Antonucci L, Nakagawa H, Kim JY, Glitzner E, Caruso S, Shalapour S, Yang L, Valasek MA, Lee S, Minnich K, Seki E, Tuckermann J, Sibilia M, Zucman-Rossi J, Karin M. Liver Cancer Initiation Requires p53 Inhibition by CD44-Enhanced Growth Factor Signaling. Cancer Cell 2018; 33: 1061-1077.e6 [PMID: 29894692 DOI: 10.1016/j.ccell.2018.05.003]
- 24 Zender L, Spector MS, Xue W, Flemming P, Cordon-Cardo C, Silke J, Fan ST, Luk JM, Wigler M, Hannon GJ, Mu D, Lucito R, Powers S, Lowe SW. Identification and validation of oncogenes in liver cancer using an integrative oncogenomic approach. Cell 2006; 125: 1253-1267 [PMID: 16814713 DOI: 10.1016/j.cell.2006.05.030]
- 25 Liu K, Lee J, Kim JY, Wang L, Tian Y, Chan ST, Cho C, Machida K, Chen D, Ou JJ. Mitophagy Controls the Activities of Tumor Suppressor p53 to Regulate Hepatic Cancer Stem Cells. Mol Cell 2017; 68: 281-292.e5 [PMID: 29033320 DOI: 10.1016/j.molcel.2017.09.022]
- 26 Cancer Genome Atlas Research Network. Comprehensive and Integrative Genomic Characterization of Hepatocellular Carcinoma. Cell 2017; 169: 1327-1341.e23 [PMID: 28622513 DOI: 10.1016/j.cell.2017.05.046]
- 27 Ho DWH, Chan LK, Chiu YT, Xu IMJ, Poon RTP, Cheung TT, Tang CN, Tang VWL, Lo ILO, Lam PWY, Yau DTW, Li MX, Wong CM, Ng IOL. TSC1/2 mutations define a molecular subset of HCC with aggressive behaviour and treatment implication. Gut 2017; 66: 1496-1506 [PMID: 27974549 DOI: 10.1136/gutjnl-2016-312734]
- Ng KY, Chai S, Tong M, Guan XY, Lin CH, Ching YP, Xie D, Cheng AS, Ma S. C-terminal truncated hepatitis B virus X protein promotes hepatocellular carcinogenesis through induction of cancer and stem cell-like properties. Oncotarget 2016; 7: 24005-24017 [PMID: 27006468 DOI: 10.18632/oncotarget.8209]
- Ching RHH, Sze KMF, Lau EYT, Chiu YT, Lee JMF, Ng IOL, Lee TKW. C-terminal truncated 29 hepatitis B virus X protein regulates tumorigenicity, self-renewal and drug resistance via STAT3/Nanog signaling pathway. Oncotarget 2017; 8: 23507-23516 [PMID: 28186991 DOI: 10.18632/oncotarget.15183]
- Sir D, Ann DK, Ou JH. Autophagy by hepatitis B virus and for hepatitis B virus. Autophagy 2010; 6: 30 548-549 [PMID: 20305390 DOI: 10.4161/auto.6.4.11669]



- 31 Kim SJ, Khan M, Quan J, Till A, Subramani S, Siddiqui A. Hepatitis B virus disrupts mitochondrial dynamics: induces fission and mitophagy to attenuate apoptosis. PLoS Pathog 2013; 9: e1003722 [PMID: 24339771 DOI: 10.1371/journal.ppat.1003722]
- Chu JYK, Ou JJ. Autophagy in HCV Replication and Protein Trafficking. Int J Mol Sci 2021; 22 32 [PMID: 33499186 DOI: 10.3390/ijms22031089]
- Kim SJ, Syed GH, Siddiqui A. Hepatitis C virus induces the mitochondrial translocation of Parkin 33 and subsequent mitophagy. PLoS Pathog 2013; 9: e1003285 [PMID: 23555273 DOI: 10.1371/journal.ppat.1003285]
- 34 Ali N, Allam H, May R, Sureban SM, Bronze MS, Bader T, Umar S, Anant S, Houchen CW. Hepatitis C virus-induced cancer stem cell-like signatures in cell culture and murine tumor xenografts. J Virol 2011; 85: 12292-12303 [PMID: 21937640 DOI: 10.1128/JVI.05920-11]
- 35 Chen CL, Tsukamoto H, Liu JC, Kashiwabara C, Feldman D, Sher L, Dooley S, French SW, Mishra L, Petrovic L, Jeong JH, Machida K. Reciprocal regulation by TLR4 and TGF- β in tumor-initiating stem-like cells. J Clin Invest 2013; 123: 2832-2849 [PMID: 23921128 DOI: 10.1172/JCI65859]
- Machida K, Tsukamoto H, Mkrtchyan H, Duan L, Dynnyk A, Liu HM, Asahina K, Govindarajan S, 36 Ray R, Ou JH, Seki E, Deshaies R, Miyake K, Lai MM. Toll-like receptor 4 mediates synergism between alcohol and HCV in hepatic oncogenesis involving stem cell marker Nanog. Proc Natl Acad Sci USA 2009; 106: 1548-1553 [PMID: 19171902 DOI: 10.1073/pnas.0807390106]
- Lytle NK, Barber AG, Reya T. Stem cell fate in cancer growth, progression and therapy resistance. 37 Nat Rev Cancer 2018; 18: 669-680 [PMID: 30228301 DOI: 10.1038/s41568-018-0056-x]
- Yang L, Shi P, Zhao G, Xu J, Peng W, Zhang J, Zhang G, Wang X, Dong Z, Chen F, Cui H. 38 Targeting cancer stem cell pathways for cancer therapy. Signal Transduct Target Ther 2020; 5: 8 [PMID: 32296030 DOI: 10.1038/s41392-020-0110-5]
- 39 Marquardt JU, Raggi C, Andersen JB, Seo D, Avital I, Geller D, Lee YH, Kitade M, Holczbauer A, Gillen MC, Conner EA, Factor VM, Thorgeirsson SS. Human hepatic cancer stem cells are characterized by common stemness traits and diverse oncogenic pathways. Hepatology 2011; 54: 1031-1042 [PMID: 21618577 DOI: 10.1002/hep.24454]
- 40 Lei ZJ, Wang J, Xiao HL, Guo Y, Wang T, Li Q, Liu L, Luo X, Fan LL, Lin L, Mao CY, Wang SN, Wei YL, Lan CH, Jiang J, Yang XJ, Liu PD, Chen DF, Wang B. Lysine-specific demethylase 1 promotes the stemness and chemoresistance of Lgr5(+) liver cancer initiating cells by suppressing negative regulators of β-catenin signaling. *Oncogene* 2015; **34**: 3188-3198 [PMID: 25893304 DOI: 10.1038/onc.2015.129
- 41 Ma S, Tang KH, Chan YP, Lee TK, Kwan PS, Castilho A, Ng I, Man K, Wong N, To KF, Zheng BJ, Lai PB, Lo CM, Chan KW, Guan XY. miR-130b Promotes CD133(+) liver tumor-initiating cell growth and self-renewal via tumor protein 53-induced nuclear protein 1. Cell Stem Cell 2010; 7: 694-707 [PMID: 21112564 DOI: 10.1016/j.stem.2010.11.010]
- Chai S, Ng KY, Tong M, Lau EY, Lee TK, Chan KW, Yuan YF, Cheung TT, Cheung ST, Wang XQ, 42 Wong N, Lo CM, Man K, Guan XY, Ma S. Octamer 4/microRNA-1246 signaling axis drives Wnt/βcatenin activation in liver cancer stem cells. *Hepatology* 2016; 64: 2062-2076 [PMID: 27639189 DOI: 10.1002/hep.28821]
- Tang KH, Ma S, Lee TK, Chan YP, Kwan PS, Tong CM, Ng IO, Man K, To KF, Lai PB, Lo CM, 43 Guan XY, Chan KW. CD133(+) liver tumor-initiating cells promote tumor angiogenesis, growth, and self-renewal through neurotensin/interleukin-8/CXCL1 signaling. Hepatology 2012; 55: 807-820 [PMID: 21994122 DOI: 10.1002/hep.24739]
- Chai S, Tong M, Ng KY, Kwan PS, Chan YP, Fung TM, Lee TK, Wong N, Xie D, Yuan YF, Guan 44 XY, Ma S. Regulatory role of miR-142-3p on the functional hepatic cancer stem cell marker CD133. Oncotarget 2014; 5: 5725-5735 [PMID: 25015418 DOI: 10.18632/oncotarget.2167]
- Li H, An J, Wu M, Zheng Q, Gui X, Li T, Pu H, Lu D. LncRNA HOTAIR promotes human liver 45 cancer stem cell malignant growth through downregulation of SETD2. Oncotarget 2015; 6: 27847-27864 [PMID: 26172293 DOI: 10.18632/oncotarget.4443]
- Zhao L, Lou G, Li A, Liu Y. IncRNA MALAT1 modulates cancer stem cell properties of liver cancer 46 cells by regulating YAP1 expression via miR-375 sponging. Mol Med Rep 2020; 22: 1449-1457 [PMID: 32626943 DOI: 10.3892/mmr.2020.11196]
- Wang Y, He L, Du Y, Zhu P, Huang G, Luo J, Yan X, Ye B, Li C, Xia P, Zhang G, Tian Y, Chen R, 47 Fan Z. The long noncoding RNA IncTCF7 promotes self-renewal of human liver cancer stem cells through activation of Wnt signaling. Cell Stem Cell 2015; 16: 413-425 [PMID: 25842979 DOI: 10.1016/j.stem.2015.03.003]





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