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***Retrospective Study***

**diagnostic and prognostic value of secreted protein acidic and rich in cysteine in the diffuse large B-cell lymphoma**

Pan PJ *et al*. SPARC as a biomarker for DLBCL

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**Abstract**

BACKGROUND

Secreted protein acidic and rich in cysteine (SPARC) is an extracellular matrix-associated protein. Studies have revealed that SPARC is involved in the cell interaction and function including proliferation, differentiation, and apoptosis. However, the role of SPARC in cancer is controversial, as it was reported as the promoter or suppressor in different cancers. Further, the role of SPARC in lymphoma is unclear.

AIM

to identify the expression and significance of SPARC in lymphoma, especially in diffuse large B-cell lymphoma (DLBCL).

METHODS

The expression analysis of SPARC in different cancers was evaluated with Oncomine. The Brune, Eckerle, Piccaluga, Basso, Compagno, Alizadeh, and Rosenwald datasets were included to evaluate the mRNA expression of SPARC in lymphoma. The Cancer Genome Atlas (TCGA)-DLBCL was used to analyze the diagnostic value of SPARC in DLBCL. The Compagno and Brune DLBCL datasets were used for validation. Then, the diagnostic value was evaluated with the receiver operating characteristic (ROC) curve. The Kaplan-Meier plot was conducted with TCGA-DLBCL, and the ROC analysis was performed based on the survival time. Further, the overall survival analysis based on the level of SPARC expression was performed with the GSE4475 and E-TABM-346. The Gene Set Enrichment Analyses (GSEA) was performed to make the underlying mechanism-regulatory networks.

RESULTS

The pan-cancer analysis of SPARC showed that SPARC was highly expressed in the brain and central nervous system, breast, colon, esophagus, stomach, head and neck, pancreas, and sarcoma, especially in lymphoma. The overexpression of SPARC in lymphoma, especially DLBCL, was confirmed in several datasets. The ROC analysis revealed that SPARC was a valuable diagnostic biomarker. More importantly, compared with DLBCL patients with low SPARC expression, those with higher SPARC expression represented a higher overall survival rate. The ROC analysis showed that SPARC was a favorable prognostic biomarker for DLBCL. Results of the GSEA confirmed that the high expression of SPARC was closely associated with focal adhesion, extracellular matrix receptor interaction, and leukocyte transendothelial migration, which suggested that SPARC may be involved in the regulation of epithelial-mesenchymal transition, KRAS, and myogenesis in DLBCL.

CONCLUSION

SPARC was highly expressed in DLBCL, and the overexpression of SPARC showed sound diagnostic value. More interestingly, the overexpression of SPARC might be a favorable prognostic biomarker for DLBCL, suggesting that SPARC might be an inducible factor in the development of DLBCL, and inducible SPARC was negative in some oncogenic pathways. All the evidence suggested that inducible SPARC might be a good diagnostic and prognostic biomarker for DLBCL.

**Key Words:** Secreted protein acidic and rich in cysteine; Diffuse large B-cell lymphoma; Inducible expression; Diagnosis; Prognosis; Clinical application

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**Core Tip:** In this study, the expression and significance of secreted protein acidic and rich in cysteine (SPARC) in the diffuse large B-cell lymphoma (DLBCL) were evaluated. The overexpression of SPARC can be an efficient diagnostic and prognostic biomarker for DLBCL, suggesting that SPARC has a potential value in future clinical application.

**INTRODUCTION**

Secreted protein acidic and rich in cysteine (SPARC) is a secreted protein, which regulates various biological activities, including proliferation, migration, adhesion, differentiation, and apoptosis[1,2]. Recently, it was reported that SPARC could control the extracellular matrix (ECM) turnover, which plays a significant role in the regulation of cell interactions[3,4].

The expression of SPARC in different cancers is ragged. According to the latest reports, the expression of SPARC could be controlled by many environmental factors. Studies showed that SPARC expression can be induced remarkably with the development of lung, esophageal, pancreatic, and prostate cancer, suggesting that SPARC could be a tumor promoter[5-9]. In detail, mechanism studies revealed that SPARC regulated the tumor growth factor β (TGF-β) signaling and then promoted the epithelial-mesenchymal transition (EMT), which participates in the cancer metastasis[5,10,11]. SPARC was a tumor promoter to activate the phosphatidylinositol 3-phosphate kinase (PI3K)/AKT pro-oncogenic pathway[12,13]. Additionally, SPARC was an anti-apoptotic factor, inhibiting the caspase activity[14].

However, other reports revealed SPARC was a tumor suppressor. In the colon, prostate, ovarian, and cervical cancers, it was reported that the expression of SPARC was closely associated with negative regulation, indicating that SPARC was a favorable prognostic factor[15]. Sailaja *et al*[16] reported that SPARC could increase the PTEN expression, which was a negative molecule of AKT suppressor. Additionally, SPARC was reported to induce endoplasmic reticulum (ER) stress. Moreover, SPARC inhibited cell proliferation by inducing the G2/M cell cycle arrest[17]. Taken together, the above evidence suggested that SPARC might play a dual role in different cancers. However, the role of SPARC is still unclear in some cancers, including diffuse large B-cell lymphoma (DLBCL), the most common clinical lymphoma. Thus, this study aimed to evaluate the significance of SPARC in the DLBCL.

In this study, we evaluated the pan-cancer expression of SPARC, and then confirmed the overexpression of SPARC in DLBCL. The receiver operating characteristic (ROC) analysis revealed a good diagnostic value of SPARC for DLBCL. Thus, we hypothesized that inducible SPARC was a favorable prognostic factor, as DLBCL patients with higher SPARC expression showed a higher survival rate. Furthermore, to validate the potential mechanism of SPARC in the DLBCL, the enrichment analysis confirmed that the overexpression of SPARC was negatively correlated with some pro-oncogenic pathways. This study confirmed inducible SPARC as a potential diagnostic and prognostic biomarker, which might function as the tumor suppressor in the DLBCL. Further, this study clarified the significance of SPARC in DLBCL and provided a reference for the future clinical transformation.

**MATERIALS AND METHODS**

***The Oncomine gene expression analysis***

The pan-cancer expression of SPARC was confirmed in the Oncomine database[18]. The threshold for *P* value was set below 0.05, the fold change was over 2 times, and the top 1% gene rank was included in the results. The included datasets were shown in Table 1[19-26], including the detailed sample number and the reporter platform.

***ROC evaluation based on different DLBCL datasets***

The TCGA-DLBCL database was evaluated as the training dataset. Firstly, the difference of SPARC expression was confirmed, and the ROC analysis was conducted based on the expression results. The area under curve (AUC) was calculated to evaluate the diagnostic value, and an AUC value from 0.5 to 1 was considered to be statistically significant. The closer the AUC value was to 1, the better diagnostic effect[27,28]. The Compagno and Brune DLBCL datasets were confirmed as validation datasets.

***The survival analysis and the prognostic evaluation***

The patients from TCGA-DLBCL with information on survival status were included in the overall survival analysis for the training test. The log-rank method was used to analyze the difference between SPARC high- and low-expression groups, and hazard ratio (HR) with 95% confidence interval (CI) was set to evaluate the prognostic value of SPARC. The value of HR was below 1.0, indicating that the high-expression group predicted a favorable prognosis[29,30]. The ROC analysis based on the survival rate was conducted in 12, 36, and 60 mo, respectively. The AUC value was calculated to evaluate the statistical difference, with an AUC value over 0.5 considered significant in the prognostic prediction. The GSE4475[31] and E-TABM-346[32] datasets were included for validation tests to evaluate the prognoses of DLBCL patients. In the GSE4475 and E-TABM-346 datasets, the median expression value of SPARC was set as the cut-off, and the survival time of censored data was the time from the start event to the truncation point.

***Gene Set Enrichment Analysis***

The TCGA-DLBCL data was subjected to the Gene Set Enrichment Analysis (GSEA), and the KEGG and HALLMARK modules were adopted in the SangerBox tool (http://sangerbox.com/Index). The detailed setting was as follows: weighted manner enrichment statistic; Signal2Noise was subjected to the metric for ranking genes; the number of permutations was 1000; the number of markers was above 100; the minimum gene number sets was over 15.

***Statistical analysis***

Results were shown in mean ± SEM, and the data was processed with Graphpad Prism 8.0 software. The difference between two groups was analyzed with Student’s *t* test. The AUC value of ROC over 0.50 was considered statistically significant. The survival analysis was conducted with log-rank and Cox methods with CI, respectively. *P* < 0.05 was considered statistically significant.

**RESULTS**

***SPARC is highly expressed in lymphoma***

Previous studies showed that SPARC might play a dual role in some cancers[33]. To further confirm the expression of SPARC in cancers, the Oncomine database with different expression datasets was used. As is shown in Figure 1A, the differences of SPARC expression between tumors and normal tissues were confirmed, and the results showed that SPARC was highly expressed in the brain and central nervous system cancer, breast cancer, cervical cancer, colon cancer, esophageal cancer, stomach cancer, head and neck cancer, pancreatic cancer, lymphoma, and sarcoma. The most significant overexpressing cancer was the lymphoma. Considering the expression of SPARC in lymphoma was controversial, this study focused on the potential role of SPARC in lymphoma. To fully understand the role of SPARC in the lymphoma, more lymphoma datasets were included to study the expression manners. As is shown in Figure 1B-D, three independent datasets revealed that SPARC was overexpressed in lymphoma cells compared with normal control cells. Further study is warranted to understand the significance of SPARC in lymphoma.

***Specific overexpression of SPARC in DLBCL***

Figure 1 shows the overexpression of SPARC in lymphoma. Due to the significant effect of tumor heterogeneity and molecular typing on the development of lymphoma, this study analyzed the SPARC expression in different types of lymphoma. As is shown in Figure 2A, there was no significant difference in the SPARC expression in Burkitt lymphoma and primary effusion lymphoma. However, we found a remarkable difference in DLBCL. Leukemia was included to test the SPARC expression as a blood tumor; however, no significant difference in the SPARC expression was found. These results suggest that SPARC was highly expressed in the lymphoma, mainly in DLBCL. The overexpression of SPARC in the DLBCL was confirmed with three independent datasets, including the Compagno dataset, Alizadeh dataset, and Rosenwald dataset (Figure 2B-D). Together, these data indicate the overexpression of SPARC in lymphoma was primarily in DLBCL, suggesting that SPARC might be closely associated with certain molecular types of lymphoma.

***Diagnostic value of SPARC for DLBCL***

Because SPARC was specifically overexpressed in DLBCL, we hypothesized that the overexpression of SPARC had diagnostic potential for DLBCL. To validate the hypothesis, tumor and normal control cells from DLBCL patients and healthy donors were included for the ROC analysis. Firstly, the overexpression of SPARC was reconfirmed in the TCGA-DLBCL database (Figure 3A). As is shown in Figure 3B, the ROC results of TCGA-DLBCL suggested high diagnostic potential, with the AUC value of up to 0.961 (*P* < 0.001). Furthermore, similar results were validated in Compagno and Brune DLBCL datasets, with the AUC value close to 1.0 (*P* < 0.001). These results indicate that SPARC is a diagnostic marker with a high true positive rate (TPR).

***SPARC as a prognostic biomarker for the DLBCL***

To elucidate the question of whether the overexpression of SPARC could be used as a potential diagnostic biomarker for DLBCL, the overall survival analysis was conducted. As is shown in Figure 4A, DLBCL patients with higher SPARC expression displayed longer survival time (median, 17.6 years), compared with lower SPARC levels (median, 3.4 years). The prognostic evaluation analysis was included with ROC analysis, which revealed that SPARC expression could act as a prognostic biomarker for the DLBCL patients, with the AUC value of 0.626, 0.569, and 0.577 for survival rate analysis at 1, 3, and 5 years, respectively (Figure 4B).

***Validation of SPARC as a prognostic biomarker for DLBCL***

Due to the small samples of DLBCL patients with detailed clinical information, other DLBCL datasets were included to validate the prognostic value of SPARC. The GSE4475 and E-TABM-346 datasets were included, and the detailed information is presented in Table 2. The overall survival analysis was conducted based on the SPARC expression. As is shown in Figure 5A, DLBCL patients with higher SPARC expression presented a higher survival rate than those with low SPARC levels [*P* = 0.0009, HR = 0.6 (0.45-0.8)]. Similar results were confirmed in the E-TABM-346 dataset [*P* = 0.001, HR = 0.54 (0.27-1.08), Figure 5B]. Furthermore, the Cox analysis was conducted in Table 2, and the Cox analysis of SPARC was confirmed in the GSE4475 as a prognostic biomarker. However, due to the small samples, the Cox analysis of the E-TABM-346 dataset showed no statistical significance (*P* = 0.08).

***Potential pathways mediated by SPARC in DLBCL***

The overexpression of SPARC might act as a favorable prognostic biomarker for DLBCL. To investigate the potential pathways mediated by SPARC in DLBCL, the GSEA analysis was performed and revealed that SPARC was implicated in focal adhesion (*P* = 0, FDR = 0, NES = -2.7), ECM receptor interaction (*P* = 0, FDR = 0, NES = -2.6), and leukocyte transendothelial migration (*P* = 0, FDR = 0, NES = -2.5). Furthermore, as is shown in Figure 6B, SPARC in DLBCL was enriched in EMT (*P* = 0, FDR = 0, NES = -2.7), KRAS signaling (*P* = 0, FDR = 0, NES = -2.6), and myogenesis (*P* = 0, FDR = 0, NES = -2.5). These negative correlation results with some pro-oncogenic pathways, combined with the prognostic analysis, suggest that SPARC might work as an onco-promoter in DLBCL.

**DISCUSSION**

DLBCL is the most common type of lymphoma. In clinical practice, the pathological characteristics of DLBCL were in a highly heterogeneous status[34,35]. Thus, there was great variation in response to the treatment and prognoses of DLBCL patients. With the development of the next-generation sequencing, the discovery of novel prognosis biomarkers has achieved great progress. C-myc, Bcl-2, Ki-67, and CD5 were identified as independent prognostic markers for DLBCL, the overexpression of which predicts the poor prognosis of DLBCL[34,36]. However, many factors can regulate the expressions of these genes, and the prognostic value is inaccurate[37,38]. Thus, it is important to screen new biomarkers for DLBCL prognosis.

Interestingly, this study revealed that SPARC was specifically overexpressed in DLBCL, rather than other types of lymphomas (Figures 1 and 2). Previous studies showed that SPARC was overexpressed in some cancers, including melanoma, breast cancer, pancreatic cancer, lung cancer, and liver cancer, and the overexpression of SPARC might act as a tumor promoter[3,33]. In glioma cells, SPARC could activate the PI3K/AKT pathway, and inhibit the apoptotic pathway[13]. However, the molecular functions of SPARC in other cancers are completely the opposite. In neuroblastoma, SPARC induces ER stress and suppresses the AKT activity, suggesting that SPARC might play a dual role in different cancers[16]. Additionally, SPARC was reported to be downregulated in T-cell lymphoma, and acted as a tumor suppressor through the inhibition of cell proliferation and metastasis[39]. This study confirmed the overexpression and the diagnostic potential of SPARC in the DLBCL. The results showed an excellent TPR of SPARC as a diagnostic biomarker for the DLBCL (Figure 3). Considering the complex functions of SPARC in cancers, this study evaluated the significance of SPARC overexpression in the DLBCL. This study revealed that the overexpression of SPARC might represent a favorable prognosis for DLBCL patients (Figures 4 and 5), suggesting that SPARC might function as a tumor suppressor in DLBCL. Meyer *et al*[40] reported the positive SPARC in stromal cells in DLBCL displayed a higher survival rate than those with negative SPARC. Our study further confirmed the clinical significance of SPARC in DLBCL. The survival rate analysis and the potential mechanism network were included in this study. The negative results were associated with the biological processes, including focal adhesion, ECM receptor interaction, and leukocyte transendothelial migration (Figure 6A). Focal adhesion and ECM receptor interaction were fully studied in the migration of cancer cells[41], and leukocyte transendothelial migration had a significant effect on the endothelial barrier function[42]. The three biological processes were closely associated with the tumor development, and the negative association between SPARC and focal adhesion, ECM receptor interaction, and leukocyte transendothelial migration indicated the potential suppressing role of SPARC in DLBCL. Moreover, SPARC-associated pathways were enriched in EMT, KRAS, and myogenesis, which were implicated as tumor promoters in many cancers[43]. Thus, the analysis of SPARC-associated pathways confirmed the negative regulation of SPARC on some pro-oncogenic pathways.

**CONCLUSION**

This study confirmed the overexpression of SPARC in lymphoma, and its overexpression is specific for DLBCL. More importantly, the overexpression of SPARC can be a diagnostic and prognostic biomarker for the DLBCL with high clinical potential.

**ARTICLE HIGHLIGHTS**

***Research background***

Secreted protein acidic and rich in cysteine (SPARC), is a protein related to the extracellular matrix. Studies have shown that SPARC regulate cell interactions and display multi-functions, including proliferation, differentiation and apoptosis. However, the role of SPARC in cancer is controversial because it is reported to be a promoter or inhibitor in different cancers. In addition, the role of SPARC in lymphoma is unclear.

***Research motivation***

The role of SPARC in different cancers is controversial, and the expression and clinical application of SPARC in lymphoma is unclear.

***Research objectives***

This study aimed to explore the expression and clinical value of SPARC in lymphoma, especially in the diffuse large B-cell lymphoma (DLBCL).

***Research methods***

The expression of SPARC in pan-cancer was conducted in Oncomine database. The Gene Expression Omnibus including Brune, Eckerle, Piccaluga, Basso, Compagno, Alizadeh, and Rosenwald datasets were subjected to confirm the expression of SPARC. The diagnostic value of SPARC was conducted in the Cancer Genome Atlas (TCGA)-DLBCL. The validated datasets were included with Compagno and Brune DLBCL datasets. Receiver operating characteristic (ROC) curve was applied to test the diagnostic value. The survival rate was conducted with Kaplan-Meier plot in TCGA-DLBCL database. The effect of SPARC on the overall survival was also confirmed in GSE4475 and E-TABM-346. The potential signaling pathways of SPARC in DLBCL was conducted with The Gene Set Enrichment Analyses (GSEA) software.

***Research results***

SPARC was highly expressed in pan-cancers, including brain and central nervous system cancer, breast cancer, colorectal cancer, esophageal cancer, gastric cancer, head and neck cancer, pancreatic cancer and sarcoma, most significantly in lymphoma. The overexpression of SPARC in lymphoma was confirmed by validated datasets. This study also identified that the overexpression of SPARC occurred significantly in DLBCL. And the overexpression of SPARC in DLBCL was tested in TCGA-DLBCL, and the ROC result showed a significant value of SPARC as a biomarker for DLBCL. Furthermore, the validation datasets including Compagno and Brune datasets confirmed the excellent diagnostic value of SPARC for DLBCL. In further prognostic analysis, DLBCL patients with high SPARC expression represented a favorable survival rate, and the ROC analysis of SPARC also demonstrated that SPARC as a favorable prognostic biomarker. The results of GSEA also revealed that SPARC was closely associated with focal adhesion, extracellular matrix receptor interaction and leukocyte transendothelial migration, which was involved in the regulation of epithelial-mesenchymal transition, KRAS and myogenesis signaling pathways in DLBCL.

***Research conclusions***

SPARC was overexpressed in DLBCL, showing an excellent diagnostic value. Furthermore, the overexpression of SPARC could be a favorable prognostic biomarker. The inducible SPARC was also negatively correlated with some oncogenic pathways. Overall, the inducible SPARC could serve as a good diagnostic and prognostic biomarker for DLBCL.

***Research perspectives***

This study identified that SPARC as a novel biomarker for the diagnosis and prognosis of DLBCL. Also, the inducible SPARC might be negatively correlated with some oncogenic pathways, suggesting that the inducible SPARC in the development of DLBCL could guide the clinical practice of DLBCL. However, this study was based on expression level, SPARC as a secreted protein, the serum level in DLBCL patients could be included in the further study.

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**Footnotes**

**Institutional review board statement:** This study was approved by the Ethics Committee of the Yongchuan Hospital of Chongqing Medical University.

**Informed consent statement:** Patients were not required to give informed consent to this study because the analysis used anonymous data from a database.

**Conflict-of-interest statement:** All authors have no conflict of interests to declare.

**Data sharing statement:** No additional data are available.

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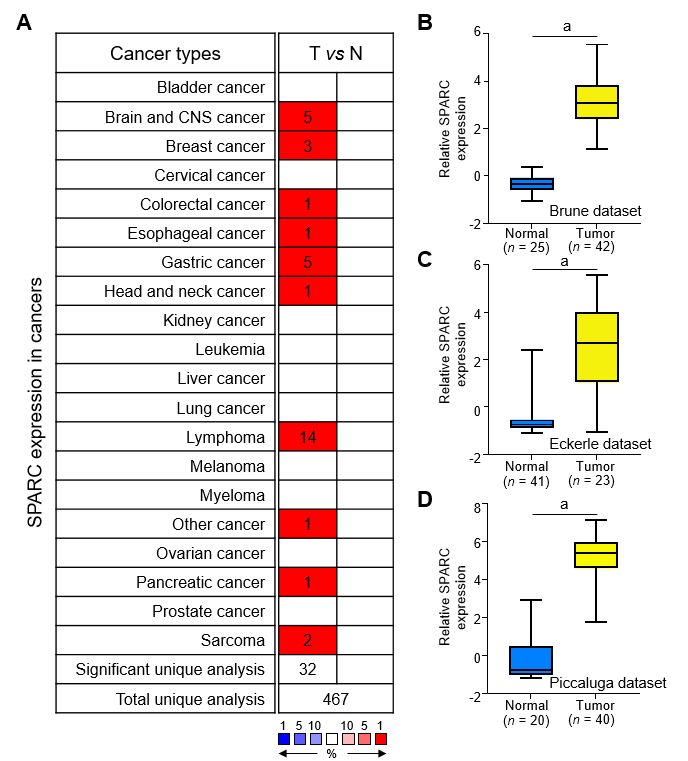
Grade C (Good): 0

Grade D (Fair): 0

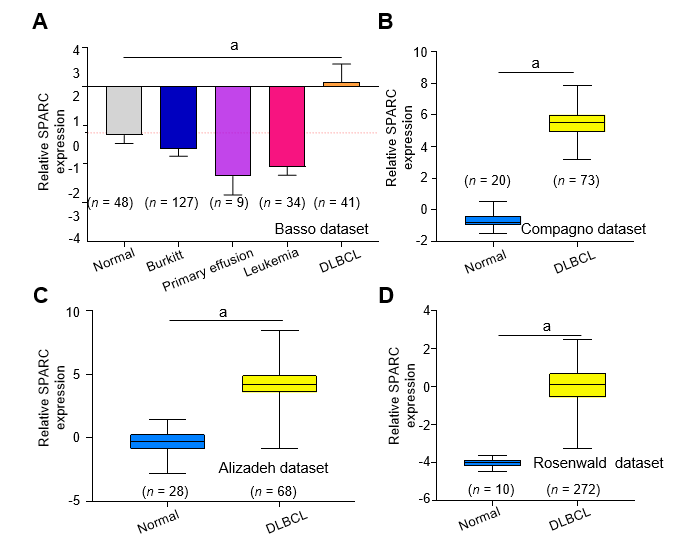
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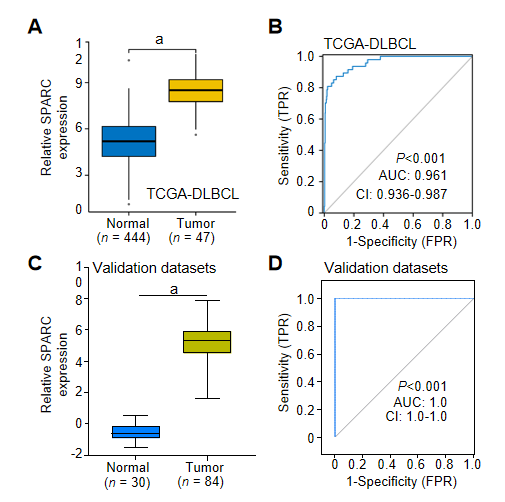
**Figure Legends**



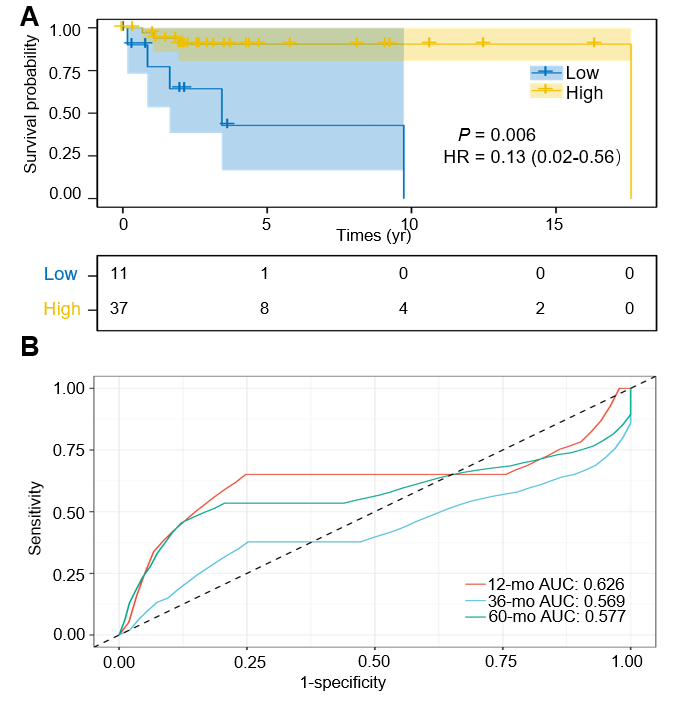
**Figure 1 Significant expression of secreted protein acidic and rich in cysteine in diffuse large B-cell lymphoma.** A: A total of 467 unique datasets were included to evaluate the expression difference of secreted protein acidic and rich in cysteine (SPARC) in pan-cancers. The top 1% ranking datasets were shown, and the fold change was set as over twice; B: The Brune dataset including different types of lymphoma was included to analyze the SPARC expression (a*P* < 0.001 *vs* control group); C: The SPARC mRNA expression was evaluated in the Eckerle dataset of lymphoma (a*P* < 0.001 *vs* control group); D: The SPARC mRNA level was confirmed with 200665\_s\_at in the lymphoma (a*P* < 0.001 *vs* control group). a*P* < 0.001 was considered statistical difference. T *vs* N: Tumor *vs* Normal.



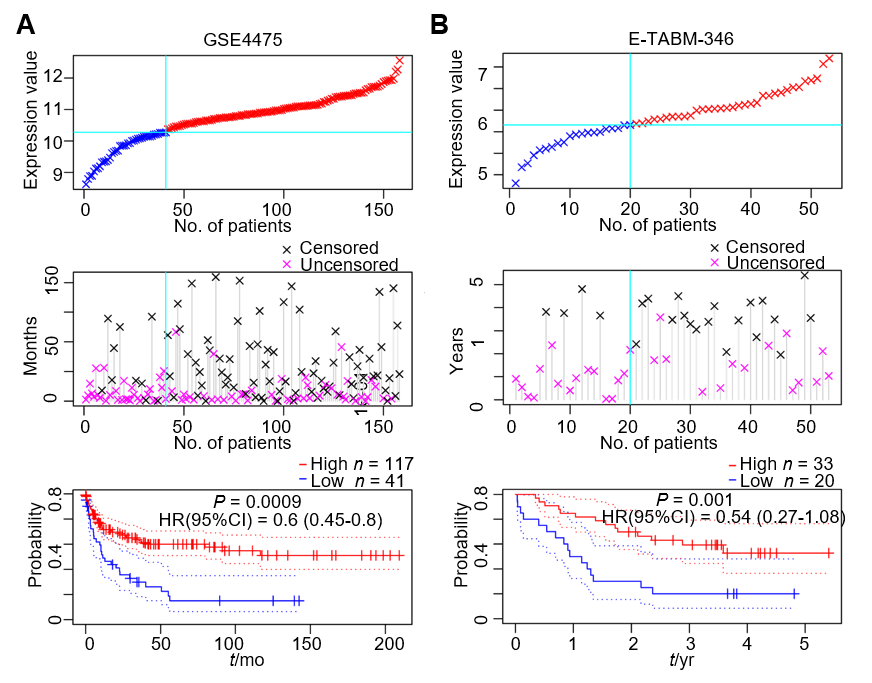
**Figure 2 Specific overexpression of secreted protein acidic and rich in cysteine in diffuse large B-cell lymphoma.** A: The Basso dataset including different types of lymphoma was included to evaluate the secreted protein acidic and rich in cysteine (SPARC) mRNA expression (a*P* < 0.001 *vs* control group); B: The mRNA expression of SPARC in the diffuse large B-cell lymphoma (DLBCL) was confirmed in the Compagno lymphoma dataset (a*P* < 0.001 *vs* control group); C: Alizadeh lymphoma dataset (a*P* < 0.001 *vs* control group); D: Rosenwald dataset (a*P* < 0.001 *vs* control group), and the normal lymphocytes were set as the normal control. a*P* < 0.001 was considered statistical difference.



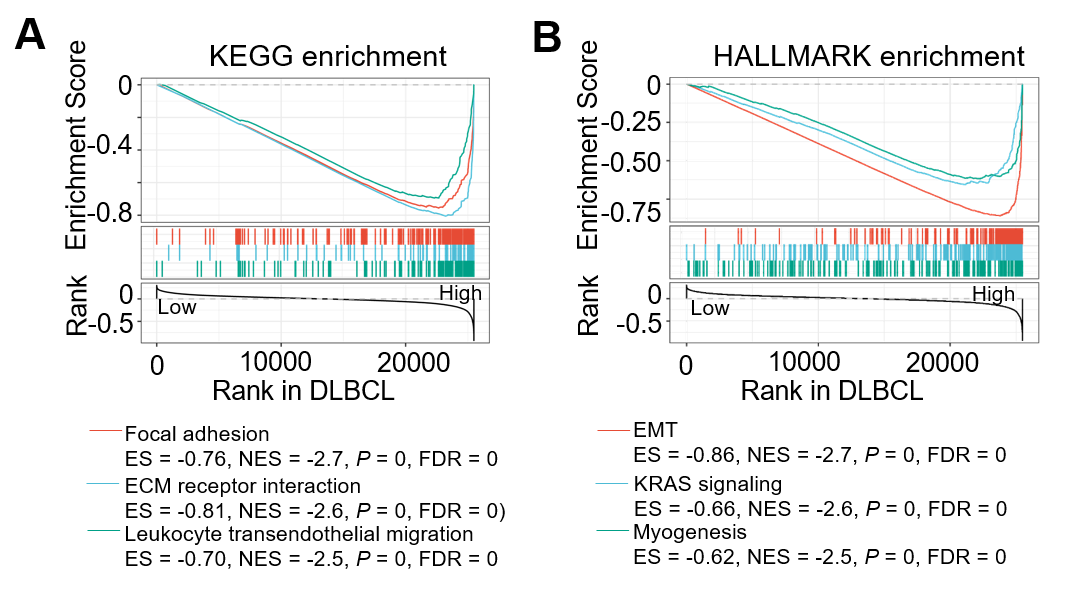
**Figure 3 Diagnostic value of secreted protein acidic and rich in cysteine in diffuse large B-cell lymphoma.** A: The mRNA level of secreted protein acidic and rich in cysteine (SPARC) was confirmed with The Cancer Genome Atlas (TCGA)-diffuse large B-cell lymphoma (DLBCL) database (a*P* < 0.001 *vs* control group); B: The receiver operating characteristic (ROC) analysis was conducted with the TCGA-DLBCL database; C: The validation datasets including the Compagno and Brune DLBCL datasets were adopted to validate the SPARC expression (a*P* < 0.001 *vs* control group); D: The ROC analysis was performed to evaluate the diagnostic value of SPARC for the DLBCL. a*P* < 0.001 was considered statistical difference. AUC: Area under curve; CI: Confidence interval; TPR: True positive rate.



**Figure 4 High expression of secreted protein acidic and rich in cysteine predicted a favorable prognostic biomarker for the diffuse large B-cell lymphoma.** A: Patients from The Cancer Genome Atlas-diffuse large B-cell lymphoma (DLBCL) were included to evaluate the overall survival based on the secreted protein acidic and rich in cysteine (SPARC) expression; B: The receiver operating characteristic analysis of SPARC as a prognostic marker for the DLBCL in different time of follow-up. AUC: Area under curve; HR: Hazard ratio.



**Figure 5 Validation of secreted protein acidic and rich in cysteine as a favorable prognostic biomarker for diffuse large B-cell lymphoma.** A: The GSE4475; B: E-TABM-346 datasets were adopted to draw the overall survival curve, and high or low groups were defined based on the expression value of secreted protein acidic and rich in cysteine; the survival status is shown in the middle panel. The overall survival cure was drawn based on the expression value and survival status. CI: Confidence interval; HR: Hazard ratio.



**Figure 6 Gene Set Enrichment Analysis enrichment analysis based on secreted protein acidic and rich in cysteine of the diffuse large B-cell lymphoma.** A, B: The Kyoto Encyclopedia of Genes and Genomes (KEGG) (A) and HALLMARK (B) enrichment modules were adopted to evaluate the enriched network. DLBCL: diffuse large B-cell lymphoma; ES: Enrichment score; FDR: False discovery rate; NES: Normalized enrichment score.

**Table 1 Lymphoma datasets for secreted protein acidic and rich in cysteine analysis**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Dataset** | **Sample size** | | **Reporter** | **Ref.** |
| **Normal** | **Tumor** |
| Rosenwald | 10 | 272 | AA045463 | [19] |
| Compagno | 20 | 111 | 200665\_s\_at | [20] |
| Alizadeh | 28 | 77 | IMAGE:324597 | [21] |
| Rosenwald Multi- | 14 | 46 | AA046533 | [22] |
| Piccaluga | 20 | 40 | 200665\_s\_at | [23] |
| Basso | 48 | 244 | 671\_at | [24] |
| Brune | 25 | 42 | 200665\_s\_at | [25] |
| Eckerle | 41 | 23 | 200665\_s\_at | [26] |

**Table 2 Statistical prognostic analysis of secreted protein acidic and rich in cysteine in different datasets**

|  |  |  |
| --- | --- | --- |
| **Features** | **GSE4475** | **E-TABM-346** |
| Number | 158 | 53 |
| *P* value | 0.000024 | 0.001367 |
| Adj. *P* value | 0.000984 | 0.033088 |
| Cox *P* value | 0.000532 | 0.082877 |
| HR (95%CI) | 0.60 (0.45-0.80) | 0.54 (0.27-1.08) |
| Sex, *n* (%) |  |  |
| female | 68 (43.0) | 26 (49.1) |
| male | 90 (57.0) | 27 (50.9) |
| Event, *n* (%) |  |  |
| 0 | 83 (52.5) | 23 (43.4) |
| 1 | 75 (47.5) | 30 (56.6) |

CI: Confidence interval; HR: Hazard ratio.